



**EFFECTS OF CRUDE METHANOLIC LEAF EXTRACT OF  
*Clinacanthus nutans* (BURM.F.) LINDAU ON HIGH-FAT DIET-INDUCED  
OBESE MICE**

**SAMIAA JAMIL ABDULWAHID**

**FPV 2020 14**



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By

**SAMIAA JAMIL ABDULWAHID**

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

September 2019

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of  
the requirement for the degree of Doctor of Philosophy

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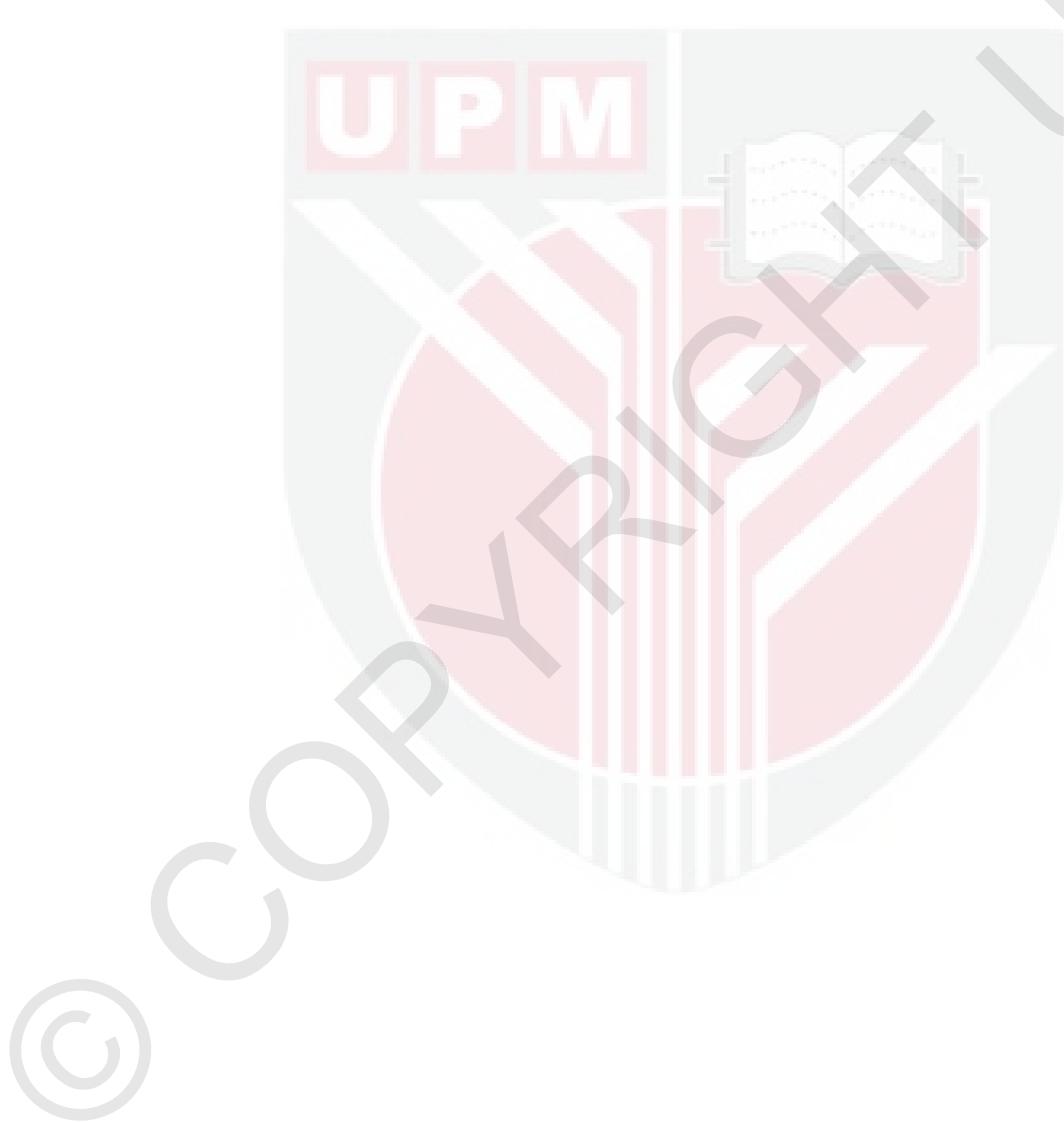
**SAMIAA JAMIL ABDULWAHID**

September 2019

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**Faculty : Veterinary Medicine**

Obesity is a major health concern that has reached epidemic proportions globally. Malaysia has the highest obesity rate at 14% in the Southeast Asia region. The cost and side effects of synthetic anti-obesity drugs necessitate the finding of suitable herbal alternatives. The current study investigated the antiobesity effects of *Clinacanthus nutans* crude leaf extract in 80 % methanol (MECN). The sub-acute oral toxicity of MECN was evaluated in 6-week-old ICR mice (21 males, 21 females). The mice were randomly divided into six treatment groups of seven animals each, comprising of untreated control, mice treated with 1000 and 2000 mg/kg MECN, for males and females. Animals were gavaged with the treatment agents once daily for 28 days. Despite the incidental lesions noted for the livers and kidneys, there no difference ( $P>0.05$ ) between the histopathological changes seen among mice treated with MECN, and that of the untreated controls in both sexes. No significant changes were also noted for the physical, hematological and serum biochemical parameters between the control and treatment groups for both sexes. However, the serum sodium level in mice treated with 2000 mg/kg MECN was lower than the controls ( $P<0.05$ ). In the ensuing experiment, MECN was used at 2000 mg/kg as a potential anti-obesity agent in obese mice. Fifty (4-weeks-old) male mice were randomly assigned into 2 groups: (1) a normal diet group (NC, n=10); and (2) treatment group (fed a high-fat diet for 16 weeks, n=40). At 20 weeks of age, the mice fed high-fat diet were randomly assigned into 4 sub-groups comprising of, high-fat diet only (HFDC); MECN at 500 mg/kg (HFD+CN500); 1000 mg/kg (HFD+CN1000) and 1500 mg/kg (HFD+CN1500). All mice were then subjected to 21 days of treatment. The current study showed that MECN treatment at 1000 and 1500 mg/kg reduced body weight, relative visceral fat, serum lipid profile, malondialdehyde (MDA) levels in muscle, cholesterol and saturated fatty acid composition ( $P<0.05$ ). Visceral fat among MECN-treated mice showed significant decrease ( $P<0.05$ ) in hypertrophic adipocyte cell size compared to the HFDC group after treatment. The PPAR $\gamma$  and SCD1 genes expression

were down-regulated, especially in mice fed with 1000 and 1500 mg/kg of MECN compared to the HFDC group. Mice treated with MECN at 1500 mg/kg showed a decreased PPAR $\alpha$  expression, and an increased expression of HSL mRNA. In terms of adipocytokines, mice treated with MECN at 1000 and 1500 mg/kg showed a significantly ( $P<0.05$ ) elevated level of adiponectin, and reduced levels ( $P<0.05$ ) of leptin, interleukin-6 (IL-6), and tumor necrosis factor- $\alpha$  (TNF $\alpha$ ) compared to the HFDC. In summary, the results suggested that MECN could ameliorate diet-induced obesity via the regulation of gene expressions and adipocytokines involved in lipid metabolism. However, future clinical trials are necessary to ascertain its clinical efficacy.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KESAN EKSTRAK MENTAH METANOL DAUN *Clinacanthus nutans*  
(BURM.F.) LINDAU KE ATAS MENCIT OBES YANG DIARUH OLEH  
DIET**

Oleh

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Obesiti adalah masalah kesihatan utama di seluruh dunia. Malaysia mempunyai prevalens obesiti tertinggi di Asia Tenggara pada 14%. Kos dan kesan sampingan ubatan anti-obesiti sintetik mendesak keperluan untuk alternatif herba yang sesuai. Kajian ini menyelidik kesan ekstrak mentah daun *Clinacanthus nutans* dalam 80 % metanol, (MECN) terhadap obesiti. Ketoksykan sub akut MECN dikaji pada mencit ICR berumur enam minggu (21 jantan dan 21 betina). Mencit dibahagikan secara rawak kepada enam kumpulan yang mengandungi tujuh individu setiap satu, yang terdiri daripada, kumpulan kawalan yang tidak dirawat, kumpulan mencit yang dirawat dengan MECN 1000 mg/kg; dan mencit dirawat dengan MECN 2000 mg/kg, untuk kedua-dua jantina. Dos rawatan diberikan secara gavaj setiap hari selama 28 hari. Walaupun terdapat lesi insidental pada hati dan ginjal, tiada perbezaan ( $P>0.05$ ) perubahan histopatologi di antara mencit kawalan dengan mencit yang dirawat dengan MECN untuk kedua-dua jantina. Tiada perubahan juga dicerap pada parameter fizikal, hematologi dan biokimia pada kedua-dua jantina ( $P>0.05$ ). Walaubagaimanapun, tahap sodium serum pada mencit yang dirawat dengan MECN pada 2000 mg/kg, adalah lebih rendah berbanding kawalan ( $P<0.05$ ). Dalam eksperimen berikutnya, MECN digunakan pada 2000 mg/kg untuk merawat mencit yang telah diaruh menjadi obes menggunakan diet. Lima puluh ekor mencit jantan ICR berumur empat minggu dibahagikan ke dalam dua kumpulan secara rawak: kumpulan diet biasa (NC, n=10), dan kumpulan mencit yang diberi makan diet tinggi lemak (60% tenaga makanan dari lemak selama 16 minggu, n=40). Pada usia 20 minggu, mencit yang diberi makan diet lemak tinggi diasinkan kepada 4 sub-kumpulan secara rawak, yang terdiri daripada: mencit diet lemak tinggi (HFDC); mencit menerima rawatan MECN pada 500 mg/kg (HFD + CN500); mencit menerima MECN pada 1000 mg/kg/ (HFD+CN1000); dan mencit yang dirawat dengan MECN pada 1500 mg/kg (HFD+CN1500). Semua mencit telah dirawat selama 21 hari. Kajian ini menunjukkan bahawa rawatan MECN pada 1000 dan 1500 mg/kg mengurangkan

berat badan, lemak visera relatif, profil lipid serum, malondialdehid (MDA) dalam otot, kolesterol dan komposisi asid lemak tenu ( $P<0.05$ ). Histologi lemak visera menunjukkan penurunan saiz sel adiposit hipertrofi selepas rawatan dengan MECN berbanding dengan HFDC ( $P<0.05$ ). Ekspresi gen PPAR $\gamma$  hepatic dan SCD1 mRNA telah berkurang, terutamanya pada mencit yang diberi makan 1000 dan 1500 mg/kg MECN berbanding dengan kumpulan HFDC. Mencit yang dirawat dengan MECN pada 1500 mg/kg menunjukkan penurunan ekspresi PPAR $\alpha$ , dan peningkatan ekspresi mRNA HSL. Perbandingan adipositokin menunjukkan mencit yang dirawat dengan MECN pada 1000 dan 1500 mg/kg mempamerkan paras adiponektin yang meningkat ( $P<0.05$ ), dan penurunan paras ( $P<0.05$ ) leptin, IL-6, dan TNF $\alpha$  yang ketara berbanding dengan HFDC. Kesimpulannya, keputusan menunjukkan bahawa MECN berupaya mengurangkan obesiti yang disebabkan oleh diet, melalui proses kawalatur ke atas ekspresi gen dan adipositokin yang terlibat dalam metabolisme lipid. Walaubagaimanpun, kajian klinikal yang selanjutnya adalah perlu untuk mengesahkan efikasi klinikalnya.

## **ACKNOWLEDGEMENTS**

All praise is due to Allah and may peace and blessings be upon Mohammed and all his family members and companions. I will be forever grateful to the people in my life who believed in me and who stood by me when all the doors were closed. First, I would like to thank Prof. Dr. Goh Yong Meng, the chairman of my supervisory committee. Also, I have never met a more honest and positive man than Dr. Goh Yong Meng. It was a privilege for me to have Dr. Goh Yong-Meng, Prof. Zailina binti Hashim and Dr. Mahdi Ebrahimi as my co-supervisors and thank them for their advice and suggestions. Without their perceptive supervision and encouragement, this work would not have been accomplished. I would like to express my deepest gratitude to Dr. Mahdi Ebrahimi for his valuable insights, his intellectual, moral support, wisdom and sustenance, as well as giving me the opportunity to continue my PhD study. Acknowledgement also goes to the Ministry of Higher Education and Scientific Research, Kurdistan Regional Government, and the President, Dean and staff of Soran University and Faculty of Education for giving me a study leave. I also wish to express my sincere gratitude to the Dean of the Faculty of Veterinary Medicine and Medicine and Health Science, Universiti Putra Malaysia for their facilities and the unlimited assistance from their staff. I would also like to thank many people who stood beside me, whose names may not be mentioned here. I will never forget them. I am indebted to my parents, sisters and brothers for their unlimited support and patience during my study.

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## LIST OF ABBREVIATIONS

ABST	2-2'-Azinobis 3-ethyl-benzothiazoline -6-sulfonic acid
AchE	Acetylcholinesterase
AGT	Angiotensinogen
ALP	Alkaline phosphate NF-kBROS
Aq.	Gly Aquaglycoporin channels
AST	Aspartate aminotransferase
AT	Adipose tissue
ATGL	Adipose triglyceride lipase
BHA	butylated hydroxyanisole
BHT	butylated hydroxytoluene
BAT	Brown adipose tissue
BM1	Body mass index
<i>C. nutans</i>	<i>Clinacanthus nutans</i>
cAMP	Cyclic adenosine monophosphate
CETP	Cholesterol ester transfer protein
COMT	Catechol methyl transferase
cDNA	Complementary DNA
CO <sub>x</sub>	Cyclooxygenase-2
CRP	C-reactive protein
CT	Cycle threshold
DPPH	2, 2-Diphenyl-2 picrylhydrazyl hydrate
DG	Diacylglycerol
ELISA	Enzyme-linked immunosorbent assay
ER	Endoplasmic reticulum

FA	Fatty acids
FBR	Foundation for Biomedical Research
G	Gram
G-3-P	Glycerol-3-phosphate
HFD+CN500	High fat diet group treated with 500 mg/kg MECN
HFD+CN1000	High fat diet group treated with MECN at 1000 mg/kg
HFD+CN1500	High fat diet group treated with MECN at 1500 mg/kg
HFD+Orlistat	High fat diet group treated with orlistat
HFDC	High-fat diet control group
HSL	Hormone sensitive lipase
IACUC	Institutional Animal Care and Use Committee
ICAM-1	Intracellular adhesion molecule 1
I.P	Intraperitoneally
I.V	Intravenously
IL-6	Interleukin-6
LDL	Low-density lipoprotein
LDLC	Low density lipoprotein cholesterol
LPL	Lipoprotein lipase.
MCP-1	Monocyte chemoattractant protein 1
MDA	Malondialdehyde
MECN	Methanolic leaf extracts of <i>Clinacanthus nutans</i>
MG	Monoacylglycerol
mg/kg	Milligram/ kilogram
mRNA	Messenger ribonucleic acid
MUFA	Monounsaturated fatty acids
NC	Normal diet control

NE	Nor epinephrine
NHMS	National Health and Morbidity Survey
NMDAR	N-methyl-D aspartate receptors
nm	Nanometre
NF- $\kappa$ B	Nuclear factor kappa-light-chain-enhancer of activated B cells
NSAIDs	non-steroidal anti-inflammatory drugs
PAI	Plasminogen activator inhibitor
PCV	Packed cell volume
PPAR $\gamma$	Peroxisome proliferator-activated receptors $\gamma$
RBC	Red blood cell
ROS	Reactive oxygen species
RT-qPCR	Real-time quantitative polymerase chain reaction
SFA	Saturated fatty acid
SDS	Sodium dodecyl sulfate
SCD	Stearoyl-Coenzyme A-Desaturase
SEM	Standard error of mean
SNS	Sympathetic nervous system
SOCS <sup>3</sup>	Suppressor of cytokine signaling
SREBP	Sterol regulatory element-binding protein
T2DM	Type 2 diabetes mellitus
TBA	Thiobarbituric acid
TBARS	Thiobarbituric acid reactive substances
TG	Triglyceride
TNF- $\alpha$	Tumor necrosis factor alpha
UCP1	Uncoupling protein 1

UK	United Kingdom
VLDL	Very-low-density lipoproteins
WAT	White adipose tissue
WBC	White blood cell
WHO	World Health Organization



# CHAPTER 1

## INTRODUCTION

Obesity occurs when the adipose tissue and adipose mass grow dramatically through two mechanisms namely, adipocyte hyperplasia and hypertrophy (Sun *et al.*, 2011). Obesity is caused by several factors including diet, inadequate physical activity, dysregulation of the genes and adipocytokines involved in the lipid metabolism. However, the over consumption of fat and decreased physical activity are the most common causes of obesity (Hsu *et al.*, 2014). According to the World Health Organization (WHO) obesity is regarded as a disease (Ahmad and Imam, 2016). It is usually accompanied with many serious medical complications, such as hypertension, coronary heart, renal diseases, type 2 diabetes and several forms of cancer such as colon, breast and gastrointestinal cancers (WHO, 2000a; Renehan *et al.*, 2008). Overweight and obesity have been recently found to be the fifth leading cause of deaths worldwide (Amano, 2013). Specifically, their complications account for 100,000 to 400,000 annually (Zawawi, 2011). If no serious actions are taken, obesity may reach the pandemic level in 2040 (Amano, 2013). The development of fat tissues produce different bioactive substances, known as adipocytokines or adipokines, which could cause the development of different metabolic-illnesses that result from the adjusted glucose, lipid homeostasis and inflammatory responses (Dandona *et al.*, 2004). Undeniably, adipose tissue has emerged as an endocrine organ which plays a vital role in metabolic regulation and production of adipocytes derivatives involved in both the pro-inflammatory and anti-inflammatory adipocytokines (Wang and Huang, 2015). Obese subjects have been implicated with high levels of pro-inflammatory adipocytokines which are interleukin-6 (IL-6) and tumor necrosis factor (TNF $\alpha$ ) and leptin along with decreased production of anti-inflammatory adipocytokines (adiponectin) (Makki *et al.*, 2013). The accumulated fat may induce the release of free fatty acids from adipocytes into the circulation, which could be an important factor in the regulation of insulin sensitivity (Dandona *et al.*, 2004). The body systems use energy for the maintenance of essential physiological functions (basal metabolic rate) necessary for performing physical activities and for adaptive thermogenesis. Moreover, the body responds to the temperature of the environment, quantity and types of nutrient consumed (Pandey *et al.*, 2016). Factors such as gender, growth, age, physical bodily function and body composition affect energy expenditure (Pandey *et al.*, 2016). Increased energy consumption combined with diet modification reduces both stored and circulating fat in the body (Rocandio *et al.*, 2001). Weight loss through pharmacotherapy treatment can also reduce the hazard to morbidity and mortality (Rudelle *et al.*, 2007). Therefore, treating obese people through lifestyle interventions and/or pharmacological therapies is a continued research effort (Rudelle *et al.*, 2007). Among the most common pharmacological drugs concurrently used as anti-obesity agent is orlistat (Ferraz *et al.* 2004). It inhibits the gastric and pancreatic lipases and consequently reduces the lipid absorption from the gut and sibutramine that acts as an oral anorexiant (Rudelle *et al.*, 2007). The obvious side effects of anti-obesity medicines, such as sibutramine which was associated with myocardial ischemia, have been the reason for the withdrawals of this product from the market in several countries (Mohamed *et al.*, 2014). The side effects of synthetic anti-obesity drugs

necessitate the finding of a suitable natural/herbal alternative. The natural products extracted from medicinal plants have been practiced as a folk medicine for a long time (Newman and Cragg, 2012). Herbal drugs have active ingredients in their crude or processed condition, in addition to certain excipients like solvents, diluents or preservatives (Calixto, 2000). Evaluations suggest that some medicinal plants and their extracts can be utilized to prevent diet-induced obesity and thus may reduce weight (Ranjbar *et al.*, 2010). A previous work has suggested the effectiveness of *Clinacanthus nutans* Lindau, a plant from the family of Acanthaceae. It is commonly known as Sabah snake grass in Malaysia (Roosita *et al.*, 2008). *Clinacanthus nutans* is one of the endemic plants that have proved to have medicinal properties as was evidenced through the traditional use. *Clinacanthus nutans* has important constituents, like phenolics, flavonoids, stigmasterol,  $\beta$ -sitosterol, lupeol, betulin, chlorophyll derivatives, protocatechuic acid, C-glycosyl flavones, vitexin, isovitexin, shaftoside, isomollupentin, 7-O- $\beta$ -gluco pyranoside, orientin, isoorientin, cerebrosides, steroids, triterpenoids, glycerides, monoacylmonogalactosylglycerol and sulfur-containing glucosides (Tu *et al.*, 2014; Mustapa *et al.*, 2015; Sarega *et al.*, 2016a; Alam *et al.*, 2016). The phytochemical compounds namely, phenols, tannins, alkaloids, steroids, protocatechuic acid and terpenes, may have the ability to exert hypolipidemic activity (Panmei *et al.*, 2007). Different plant-based polyphenols have been known to quench free radicals and exhibit anti-inflammatory properties, as well as anti-hyperglycemic and anti-hyperlipidemic properties (Nakazato *et al.*, 2006; Atanassova *et al.*, 2011; Tresserra *et al.*, 2014). Despite all known biological activities in earlier work, as well as its increased general acceptance, this plant is believed to be have the potential to reduce weight and lower blood cholesterol. Nonetheless, such assumptions still required further empirical evidence. The present study aims to investigate this plant by using doses of methanolic leaf extract of *Clinacanthus nutans* (MECN) similar to previous research (Zakaria *et al.*, 2016). Recent evidence has shown that MECN improves lipid profiles in rats (Sarega *et al.*, 2016a) and can increase the acetylcholinesterase (AchE) activity in the liver and heart (Lau *et al.*, 2014), in addition to its powerful anti-inflammatory property in inhibiting the neutrophil responsiveness (Wanikiat *et al.*, 2008). Polyphenols have been known to regulate lipid metabolism by inducing metabolic gene expression, or trigger the transcription factors that play a significant part in energy metabolism (Ali *et al.*, 2014). There are numerous mechanisms for the reduction of adipogenesis, such as suppression of growth and differentiation of adipocytes by inhibiting the peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ) and sterol regulatory element-binding protein (SREBP2) gene expression (Mayoral *et al.*, 2015; Moseti *et al.*, 2016). Besides, the down regulation of stearoyl-CoA desaturase (SCD) gene can stimulate the insulin sensitivity action, and thus produce effective energy utilization (Karahashi *et al.*, 2013). However, even though the presence of polyphenols in *Clinacanthus nutans* has been determined, there is still little evidence about the anti-obesity properties and the mechanism by which *Clinacanthus nutans* could exert potential anti-obesity effects.

## **1.2 Problem Statement**

Obesity is a severe public health challenges in the 21<sup>st</sup> century, in Southeast Asia and elsewhere in the world. Obesity is considered one of the primary risk factors for chronic and non-transmissible diseases (Dans *et al.*, 2011). There are recent reports which suggest that Malaysia has the highest obesity 13.3% and overweight 38.5% cases in Southeast Asia (AROFIIN, 2017). More specifically, around 60% of the Malaysian aged  $\geq 18$  has body mass index (BMI)  $>25 \text{ kg/m}^2$  (Ng *et al.*, 2014). Overweight which leads to higher health and economic burdens is today a global challenge (Tuan and Nicklas, 2009). Obese people have been observed to incur about 30% of higher medical costs in comparison with normal weight individuals (Withrow and Alter, 2011). The Asia Roundtable on Food Innovation for Improved Nutrition has reported that obesity accounts for 10 to 19% of the overall healthcare costs in Malaysia, totaling RM 4.26 to 8.53 billion (AROFIIN, 2017). In the USA, similar reports suggest that every American adult will be either overweight or obese by 2048. This will increase the expenditure attributed to obesity, costing USD 860.7 to 956.9 billion or 16 to 18% of the overall US health care expenditure by 2030 (Wang *et al.*, 2008). Attempts at treatments via anti-obesity drugs are hampered by their side-effects. In this regard, coming up with a dietary supplementary that could potentially reduce the weight gain is the main concern of the research in the field (Mohamed *et al.*, 2014). The increased prevalence of overweight and obesity, in several non-Western countries including Malaysia, has made the demands for alternative approaches to body weight management very necessary. More research endeavors are being investigated for their anti-obesity properties (Rudelle *et al.*, 2007). The present study aimed to provide better insights on the toxicity profile of methanolic leaf extract of *Clinacanthus nutans* and the beneficial effects of methanolic leaf extract of *Clinacanthus nutans* against overweight/obesity.

## **1.3 Significance of the Study**

The *Clinacanthus nutans* has been used as a traditional medicine to treat diseases like skin rashes, scorpion and insect bites, diabetes mellitus and many more other diseases in Southeast Asia, particularly in Malaysia, Indonesia and Thailand (Tuntiwachwuttikul *et al.*, 2003; Sakdarat *et al.*, 2009; Alam *et al.*, 2016). Many recent investigations have revealed that this plant holds great promise for health and disease prevention. This has brought a lot of attention on how this plant would contribute in health management. The findings of this research are hoped to provide insights into the effect of *Clinacanthus nutans* on physiological and biochemical changes in obese mice. The outcome of this work can be exploited to produce a new supplement from natural plant to treat or prevent obesity phenotype and its complications in future.

## **1.4 Research Objectives**

### **1.4.1 General Objective**

The main objective of the study was to investigate the potential anti-obesity effects of crude methanolic leaf extract of *Clinacanthus nutans* (MECN) on high-fat diet-induced obese mice.

### **1.4.2 Specific Objectives**

The specific objectives of this study were:

- 1- To identify the total phenolic and flavonoid contents, and antioxidant activities of methanolic leaf extract of *Clinacanthus nutans* (MECN).
- 2- To determine the potential toxic effects of MECN treatment in mouse model with respect to the hematological, biochemical and histopathological parameters after oral administration for 28 days.
- 3- To investigate the effects of MECN treatment on body weight, adipose cellularity, biochemical parameters and lipid profile in obese mice fed a high-fat diet.
- 4- To examine the effects of MECN treatment on gene expressions of PPAR $\alpha$ , PPAR $\gamma$ , HSL, SCD and SREBP mRNA, as well as adipocytokines including adiponectin, leptin, IL-6 and TNF regulation in obese mice fed a high-fat diet.

### **1.4.3 Hypothesis**

It was expected that: sub-acute oral toxicity study of MECN at 1500 mg/kg or lower is safe in mice. MECN would affect adipose cellularity, lipid profiles, biochemical parameters, mRNA expression levels of the different genes and adipocytokines on a high-fat diet-induced obese mice.

## REFERENCES

- Abbaspour, N., Hurrell, R., and Kelishadi, R. (2014). Review on iron and its importance for human health. *Journal of Research in Medical Sciences: The Official Journal of Isfahan University of Medical Sciences*, 19(2), 164.
- Abdelaal, M., le Roux, C. W., and Docherty, N. G. (2017). Morbidity and mortality associated with obesity. *Annals of Translational Medicine*, 5(7).
- Abugassa, I., Bashir, A., Doubali, K., Etwir, R., Abu-Enawel, M., and Abugassa, S. (2008). Characterization of trace elements in medicinal herbs by instrumental neutron activation analysis. *Journal of Radioanalytical and Nuclear Chemistry*, 278(3), 559-563.
- Adedapo, A. A., Abatan, M. O., and Olorunsogo, O. O. (2004). Toxic effects of some plants in the genus *Euphorbia* on haematological and biochemical parameters of rats. *Veterinarski Arhiv*, 74(1), 53-62.
- Adrogue, H. J., and Madias, N. E. (2000). Hyponatremia. *New England Journal of Medicine*, 342(21), 1581-1589.
- Aggarwal, B. B. (2010). Targeting inflammation-induced obesity and metabolic diseases by curcumin and other nutraceuticals. *Annual Review of Nutrition*, 30: 173.
- Agouni, A., Lagrue-Lak-Hal, A., Mostefai, H. A., Tesse, A., Mulder, P., and Rouet, P. (2009). Red wine polyphenols prevent metabolic and cardiovascular alterations associated with obesity in Zucker fatty rats (Fa/Fa). *PLoS ONE* 4:e5557.doi:10.1371/journal.pone.0005557.
- Aherne, S. A., and O'Brien, N. M. (2002). Dietary flavonols: chemistry, food content, and metabolism. *Nutrition*, 18(1), 75-81.
- Ahmad, S. I., and Imam, S. K. (2016). *Obesity: A Practical Guide*. Switzerland: Springer.
- Ahn, J., Lee, H., Kim, S., and Ha, T. (2010). Curcumin-induced suppression of adipogenic differentiation is accompanied by activation of Wnt/  $\beta$ -catenin signaling. *American Journal of Physiology-Cell Physiology*, 298(6), 1510-1516.
- Alam, A., Ferdosh, S., Ghafoor, K., Hakim, A., Juraimi, A. S., Khatib, A., and Sarker, Z. I. (2016). *Clinacanthus nutans*: A review of the medicinal uses, pharmacology and phytochemistry. *Asian Pacific Journal of Tropical Medicine*, 9(4), 402-409.

- Al-Awwadi, N. A., Araiz, C., Bornet, A., Delbosc, S., Cristol, J. P., and Linck, N (2005). Extractsenrichedindifferentpolyphenolicfamiliesnormalizeincreased cardiac NADPH oxidase expression while having differential effects on insulin resistance, hypertension, and cardiac hypertrophy in high-fructose-fed rats. *Journal of Agricultur Food Chem*.53,151–157.doi:10.1021/jf048919f
- Al-Awwadi, N. A., Bornet, A., Alempijevic, T., and Kovacevic, N. (2007). Right liver lobe diameter: albumin ratio: a new non-invasive parameter for prediction of oesophageal varices in patients with liver cirrhosis (preliminary report). *Gut*, 56(8), 1166-1167.
- Alfaia, C. M., Ribeiro, V. S., Lourenc, M. R., Auaresma, M. A., Martins, S. I., Portugal, A. P., Fontes, C. M., Bessa, R. J., Castro, M. L., and Prater, J. A. (2006). Fatty acid composition, conjugated linoleic acid isomers and cholesterol in beef from crossbred bullocks intensively produced and from Alentejana purebred bullocks reared according to Carnalentejana- PDO specifications. *Meat Science*, 72, 425-436.
- Ali, A. (2009). Proximate and mineral composition of the marchubeh (*Asparagus officinalis*). *World Dairy and Food Science*, 4(2), 142-149
- Ali, F., Ismail, A., and Kersten, S. (2014). Molecular mechanisms underlying the potential antiobesity-related diseases effect of cocoa polyphenols. *Molecular Nutrition and Food Research*, 58(1), 33-48.
- Al-Rejaie, S. S. (2013). Thymoquinone treatment alleviate ovariectomy-induced hepatic oxidative damage in rats. *Journal of Applied Pharmaceutical Science*, 3(6), 126-131. <http://imsear.hellis.org/handle/123456789/151665>
- Amano, S. U. (2013). *Local Macrophage Proliferation in Adipose Tissue is A Characteristic of Obesity-Associated Inflammation*. PhD Thesis. Universiti of Massachusetts Medical School, USA.
- An, S., Han, J. I., Kim, M. J., Park, J. S., Han, J. M., Baek, N. I., and Jeong, T. S. (2010). Ethanolic extracts of *Brassica campestris* spp. *rapa* roots prevent high-fat diet-induced obesity via 3-adrenergic regulation of white adipocyte lipolytic activity. *Journal of Medicinal Food*, 13(2), 406-414.
- Anandh Babu, P. V., Sabitha, K. E., and Shyamaladevi, C. S. (2006). Green tea extract impedes dyslipidaemia and development of cardiac dysfunction in streptozotocin-diabetic rats. *Clinical and Experimental Pharmacology and Physiology*, 33(12), 1184-1189.
- Andersen, R. E., Crespo, C. J., Bartlett, S. J., Cheskin, L. J., and Pratt, M. (1998). Relationship of physical activity and television watching with body weight and level of fatness among children: results from the third national health and nutrition examination survey. *Jama*, 279(12), 938-942.

- Andersson, A., Nalsen, C., Tengblad, S., and Vessby, B. (2002). Fatty acid composition of skeletal muscle reflects dietary fat composition in humans. *The American Journal of Clinical Nutrition*, 76(6), 1222-1229.
- Anstee, Q. M., and Goldin, R. D. (2006). Mouse models in non-alcoholic fatty liver disease and steatohepatitis research. *International Journal of Experimental Pathology*, 87(1), 1-16.
- Anthony, A., and Parsons, J. (1957). Variation in normal sodium, potassium, and calcium levels in Wistar albino rats. *Science*, 125(3253), 881-883.
- AOAC (1997). *Official Methods of Analysis* (16th Ed.). Washington D.C.: Association of Official Analytical Chemist
- AOAC (2005). *Official Methods of Analysis of AOAC International*. Association of Official Analysis Chemists International. Retrieved from <http://www.eoma.aoac.org/>
- Aprikian, O., Duclos, V., Guyot, S., Besson, C., Manach, C., Bernalier, A., and Demigni, C. (2003). Apple pectin and a polyphenol-rich apple concentrate are more effective together than separately on cecal fermentations and plasma lipids in rats. *The Journal of Nutrition*, 133(6), 1860-1865.
- Araujo, J., and Martel, F. (2012). Sibutramine effects on central mechanisms regulating energy homeostasis. *Current Neuropharmacology*, 10(1), 49-52.
- Ardevol, A., Blade, C., Salvado, M. J., and Arola, L. (2000). Changes in lipolysis and hormone-sensitive lipase expression caused by procyanidins in 3T3-L1 adipocytes. *International Journal of Obesity*, 24(3), 319.
- Armstrong, D., and Al-Awadi, F. (1991). Lipid peroxidation and retinopathy in streptozotocin-induced diabetes. *Free Radical Biology and Medicine*, 11(4), 433-436.
- Aronne, L. J. (1998). Modern medical management of obesity: the role of pharmaceutical intervention. *Journal of the American Dietetic Association*, 98(10), S23-S26.
- Arullappan, S., Rajamanickam, P., Thevar, N., and Kodimani, C. C. (2014). In Vitro screening of cytotoxic, antimicrobial and antioxidant activities of *Clinacanthus nutans* (Acanthaceae) leaf extracts. *Tropical Journal of Pharmaceutical Research*, 13(9), 1455-1461.
- Asaolu, S. S., Adefemi, O. S., Oyakilome, I. G., Ajibulu, K. E., and Asaolu, M. F. (2012). Proximate and mineral composition of Nigerian leafy vegetables. *Journal of Food Research*, 1(3), 214.
- Asarian, L., and Geary, N. (2013). Sex differences in the physiology of eating. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 305(11), R1215-R1267.

- Aseervatham, G. S. B., Sivasudha, T., Jeyadevi, R., and Ananth, D. A. (2013). Environmental factors and unhealthy lifestyle influence oxidative stress in humans-an overview. *Environmental Science and Pollution Research*, 20(7), 4356-4369.
- Asia Roundtable on Food Innovation for Improved Nutrition (2017). *Obesity in Asian, a call to action; AROFIIN*. Retrieved from <http://www.arofiin.org/News/tid/57/Obesity-in-ASEAN-A-Call-to-Action>, [Published on 10 Jul 2017].
- Asilmaz, E., Cohen, P., Miyazaki, M., Dobrzyn, P., Ueki, K., and Fayzikhodjaeva, G. (2004). Site and mechanism of leptin action in a rodent form of congenital lipodystrophy. *Journal of Clinical Investigation*, 113(3), 414-424.
- Aslam, M. S., Ahmad, M. S., and Mamat, A. S. (2015). A review on phytochemical constituents and pharmacological activities of *Clinacanthus nutans*. *World*, 2, 4.
- Aslam, M. S., Ahmad, M. S., Mamat, A. S., Ahmad, M. Z., and Salam, F. (2016). Antioxidant and wound healing activity of polyherbal fractions of *Clinacanthus nutans* and *Elephantopus scaber*. *Evidence-Based Complementary and Alternative Medicine*, 2016.
- Asyura, S. Hamzah, H., R. Shaari, R., Sithambaram, S., and Mustapha, N. (2016). Blood profiles and histopathological changes of liver and kidney tissues from male Sprague Dawley rats treated with ethanol extracts of *Clinacanthus nutans* leaf. *Journal of Clinical Toxicology*, 6(6). DOI: 10.4172/2161-0495.1000329.
- Atanasov, A. G., Waltenberger, B., Pferschy-Wenzig, E. M., Linder, T., Wawrosch, C., Uhrin, P., and Rollinger, J. M. (2015). Discovery and resupply of pharmacologically active plant-derived natural products: a review. *Biotechnology Advances*, 33(8), 1582-1614.
- Atanassova, M., Georgieva, S., and Ivancheva, K. (2011). Total phenolic and total flavonoid contents, antioxidant capacity and biological contaminants in medicinal herbs. *Journal of the University of Chemical Technology and Metallurgy*, 46(1), 81-88.
- Austin, M. B., and Noel, J. P. (2003). The chalcone synthase superfamily of type III polyketide synthases. *Natural Product Reports*, 20(1), 79-110. Retrieved from DOI 10.1007/978-3-319-19821-7-21.
- Azay, J., Araiz, C., Delbosc, S., Cristol, J. P. (2004). Red wine polyphenols alone or in association with ethanol prevent hypertension, cardiac hypertrophy, and production of reactive oxygen species in the insulin-resistant fructose-fed rat. *Journal Agriculture Food Chemistry*. 52, 5593–5597.

- Baba, S., Natsume, M., Yasuda, A., Nakamura, Y., Tamura, T., Osakabe, N., and Kondo, K (2007). Plasma LDL and HDL cholesterol and oxidized LDL concentrations are altered in normo-and hypercholesterolemic humans after intake of different levels of cocoa powder. *The Journal of Nutrition*, 137(6), 1436-1441.
- Bagnis, C. I., Deray, G., Baumelou, A., Le Quintrec, M., and Vanherweghem, J. L. (2004). Herbs and the kidney. *American Journal of Kidney Diseases*, 44(1), 1-11.
- Bailey, S. A., Zidell, R. H., and Perry, R. W. (2004). Relationships between organ weight and body/brain weight in the rat: what is the best analytical endpoint? *Toxicologic Pathology*, 32(4), 448-466.
- Bais, S., Singh, G. S., and Sharma, R. (2014). Anti-obesity and hypolipidemic activity of *Moringa oleifera* leaves against high-fat diet-induced obesity in rats. *Advances in Biology*, 2014, 1-9.
- Baker, M. E. (1998). Albumin's role in steroid hormone action and the origins of vertebrates: is albumin an essential protein? *FEBS letters*, 439(1-2), 9-12.
- Balijepalli, M. K., Suppaiah, V., Chin, A. M., Buru, A. S., Sagineedu, S. R., and Pichika, M. R. (2015). Acute oral toxicity studies of *Swietenia macrophylla* seeds in Sprague Dawley rats. *Pharmacognosy Research*, 7(1), 38.
- Barnea, M., Shamay, A., Stark, A. H., and Madar, Z. (2006). A high-fat diet has a tissue-specific effect on adiponectin and related enzyme expression. *Obesity*, 14: 2145–2153.
- Barle, E. L., Looser, R., Cerne, M., and Bechter, R. (2012). The value of acute toxicity testing of pharmaceuticals for estimation of human response. *Regulatory Toxicology and Pharmacology*, 62(3), 412-418.
- Basgel, S., and Erdemoglu, S. B. (2006). Determination of mineral and trace elements in some medicinal herbs and their infusions consumed in Turkey. *Science of the Total Environment*, 35 9(1-3), 82-89.
- Basu, A., Du, M., Sanchez, K., Leyva, M. J., Betts, N. M., Blevins, S., and Lyons, T. J. (2011). Green tea minimally affects biomarkers of inflammation in obese subjects with metabolic syndrome. *Nutrition*, 27(2), 206-213.
- Bell, D. D., and Zucker, I. (1971). Sex differences in body weight and eating: organization and activation by gonadal hormones in the rat. *Physiology and Behavior*, 7(1), 27-34.
- Bell, J. M., and Erfle, J. D. (1958). The requirement for potassium in the diet of the growing mouse. *Canadian Journal of Animal Science*, 38(2), 145-147.

- Ben-Jonathan, N., Hugo, E. R., and Brandebourg, T. D. (2009). Effects of bisphenol A on adipokine release from human adipose tissue: Implications for the metabolic syndrome. *Molecular and Cellular Endocrinology*, 304: 49–54.
- Bhaskar, R. K. (2016). Side effects and biochemical changes in rat liver on drug treatment. *J. Technological Advances and Scientific Resources*, 5(2), 1881-1886.
- Bilal, S., Khan, A. L., Waqas, M., Shahzad, R., Kim, I. D., Lee, I. J., and Shin, D. H. (2016). Biochemical constituents and *in vitro* antioxidant and anticholinesterase potential of seeds from native Korean persimmon genotypes. *Molecules*, 21(7), 893.
- Bin, C., Xiaoru, W., and Lee, F. S. (2001). Pyrolysis coupled with atomic absorption spectrometry for the determination of mercury in Chinese medicinal materials. *Analytica Chimica Acta*, 447(1-2), 161-169.
- Birari, R. B., and Bhutani, K. K. (2007). Pancreatic lipase inhibitors from natural sources: unexplored potential. *Drug Discovery Today*, 12(19), 879-889.
- Black, C. L. (2010). *A Role for Central Leptin Signaling in Maintaining Myocardial Fatty Acid Oxidation*. (Doctoral dissertation, University of Utah), USA.
- Boeing, J.S., Barizão, E.O., Silva, B.C., Montanher, P.F., Almeida, V.C. and Visentainer, J.V. (2014). Evaluation of solvent effect on the extraction of phenolic compounds and antioxidant capacities from the berries: application of principal component analysis. *Chemistry Central Journal*, 8(48), 1-9.
- Bonthuis, P. J., Cox, K. H., Searcy, B. T., Kumar, P., Tobet, S., and Rissman, E. F. (2010). Of mice and rats: key species variations in the sexual differentiation of brain and behavior. *Frontiers in Neuroendocrinology*, 31(3), 341-358.
- Boque, N., Iglesia, R., Garza, A. L., Milagro, F. I., Olivares, M., Banuelos, and Campion, J. (2013). Prevention of diet-induced obesity by apple polyphenols in Wistar rats through regulation of adipocyte gene expression and DNA methylation patterns. *Molecular Nutrition and Food Research*, 57(8), 1473-1478.
- Botsoglou, N. A., Christaki, E., Fletouris, D. J., Florou-Paneri, P., and Spais, A. B. (2002). The effect of dietary oregano essential oil on lipid oxidation in raw and cooked chicken during refrigerated storage. *Meat Science*, 62(2), 259-265.
- Bouayed, J., and Bohn, T. (2010). Exogenous antioxidants-double-edged swords in cellular redox state: health beneficial effects at physiologic doses versus deleterious effects at high doses. *Oxidative Medicine and Cellular Longevity*, 3(4), 228-237.

- Box, R. J., and Spielmann, H. (2005). Use of the dog as non-rodent test species in the safety testing schedule associated with the registration of crop and plant protection products (pesticides): present status. *Archives of Toxicology*, 79(11), 615-626.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1-2), 248-254.
- Bray, G. A. (2000). A concise review on the therapeutics of obesity. *Nutrition*, 16(10), 953-960.
- Brayton, C. F. (1986). Dimethyl sulfoxide (DMSO): a review. *The Cornell Veterinarian*, 76(1), 61-90.
- Brodie, B. B., Butler, W. M., Horning, M. G., Maickel, R. P., and Maling, H. M. (1961). Alcohol-induced triglyceride deposition in liver through derangement of fat transport. *The American Journal of Clinical Nutrition*, 9(4), 432–435.
- Bruunsgaard, H. (2005). Physical activity and modulation of systemic low-level inflammation. *Journal of Leukocyte Biology*, 78(4), 819-835.
- Burton, R. F. (2010). Waist circumference as an indicator of adiposity and the relevance of body height. *Medical Hypotheses*, 75(1), 115-119.
- Buysschaert, B., Aydin, S., Morelle, J., Hermans, M. P., Jadoul, M., and Demoulin, N. (2016). Weight loss at a high cost: orlistat-induced late-onset severe kidney disease. *Diabetes and Metabolism*, 42(1), 62-64.
- Calder, P. C., Ahluwalia, N., Brouns, F., Buetler, T., Clement, K., Cunningham, K., and Lansink, M. (2011). Dietary factors and low-grade inflammation in relation to overweight and obesity. *British Journal of Nutrition*, 106(S3), S1-S78.
- Calixto, J. B. (2000). Efficacy, safety, quality control, marketing and regulatory guidelines for herbal medicines (phytotherapeutic agents). *Brazilian Journal of Medical and Biological Research*, 33(2), 179-189.
- Candiracci, M., Justo, M. L., Castao, A., Rodriguez-Rodriguez, R., and Herrera, M. (2014). Rice bran enzymatic extract supplemented diets modulate adipose tissue inflammation markers in Zucker rats. *Nutrition*, 30(4), 466-472.
- Cao, H., Gerhold, K., Mayers, J.R., Wiest, M.M., Watkins, S.M., and Hotamisligil, G.S. (2008). Identification of a lipokine, a lipid hormone linking adipose tissue to systemic metabolism. *Cell*, 134, 933-944.
- Castelhano-Carlos, M. J., and Baumans, V. (2009). The impact of light, noise, cage cleaning and in-house transport on welfare and stress of laboratory rats. *Laboratory Animals*, 43(4), 311-327.

- Cavaletti, G., Oggioni, N., Sala, F., Pezzoni, G., Cavalletti, E., Marmiroli, P., and Tredici, G. (2000). Effect on the peripheral nervous system of systemically administered dimethylsulfoxide in the rat: a neurophysiological and pathological study. *Toxicology letters*, 118(1-2), 103-107.
- Cazzola, R., Rondanelli, M., Russo-Volpe, S., Ferrari, E., and Cestaro, B. (2004). Decreased membrane fluidity and altered susceptibility to peroxidation and lipid composition in overweight and obese female erythrocytes. *Journal of Lipid Research*, 45(10), 1846-185.
- Chaiyasit, W., Elias, R. J., McClements, D. J., and Decker, E. A. (2007). Role of physical structures in bulk oils on lipid oxidation. *Critical Reviews in Food Science and Nutrition*, 47(3), 299-317.
- Chan, P. T., Fong, W. P., Cheung, Y. L., Huang, Y., Ho, W. K. K., and Chen, Z.Y. (1999). Jasmine green tea epicatechins are hypolipidemic in hamsters (*Mesocricetus auratus*) fed a high-fat diet. *The Journal of Nutrition*, 129(6), 1094–1101.
- Chelyn, J. L., Omar, M. H., Mohd Yousof, N. S. A., Ranggasamy, R., Wasiman, M. I., and Ismail, Z. (2014). Analysis of flavone C-glycosides in the leaves of *Clinacanthus nutans* (Burm. f.) Lindau by HPTLC and HPLC-UV/DAD. *The Scientific World Journal*, 2014. <http://dx.doi.org/10.1155/2014/724267>.
- Chen, C. Y., Chang, K. K., Chow, N. H., Leow, T. C., Chou, T. C., and Lin, X. Z. (1995). Toxic effects of cholelitholytic solvents on gallbladder and liver. *Digestive Diseases and Sciences*, 40(2), 419-426.
- Chen, X., Matthews, J., Zhou, L., Pelton, P., Liang, Y., Xu, J., and Demarest, K. (2008). Improvement of dyslipidemia, insulin sensitivity, and energy balance by a peroxisome proliferator-activated receptor  $\alpha$  agonist. *Metabolism*, 57: 1516–1525.
- Chevassus, H., Gaillard, J. B., Farret, A., Costa, F., Gabillaud, I., Mas, E., and Galtier, F. (2010). A fenugreek seed extract selectively reduces spontaneous fat intake in overweight subjects. *European Journal of Clinical Pharmacology*, 66(5), 449-455.
- Ching, S., Zakaria, Z., Paimin, F., and Jalalian, M. (2013). Complementary alternative medicine use among patients with type 2 diabetes mellitus in the primary care setting: a cross-sectional study in Malaysia. *BMC Complementary and Alternative Medicine*, 13 (1), (148), 1-7.
- Cho, A. S., Jeon, S. M., Kim, M. J., Yeo, J., Seo, K. I., Choi, M. S., and Lee, M. K. (2010). Chlorogenic acid exhibits anti-obesity property and improves lipid metabolism in high-fat diet-induced-obese mice. *Food and Chemical Toxicology*, 48: 937–943. Retrieved from <http://dx.doi.org/10.1016/j.fct.2010.01.003>

- Choa, J. B. D., Lu, R. V., and Nombrado, M. A. D. (2016). Anti-obesity Activity of *Broussonetia luzonicus* (Moraceae) leaves. *Journal of Chemical and Pharmaceutical Research*, 8(2), 783-788.
- Choi, K. M., Ryu, O. H., Lee, K. W., Kim, H. Y., Seo, J. A., Kim, S. G., and Baik, S. H. (2007). Serum adiponectin, interleukin-10 levels and inflammatory markers in the metabolic syndrome. *Diabetes Research and Clinical Practice*, 75: 235–240.
- Choi, B.K., Park, S.B., Lee, D.R., Lee, H.J., Jin, Y.Y., Yang, S.H. and Suh, J.W. (2016). Green coffee bean extract improves obesity by decreasing body fat in high-fat diet-induced obese mice. *Asian Pacific Journal of Tropical Medicine*, 9(7), 635–643.
- Christenson, L. K., Osborne, T. F., McAllister, J. M., and Strauss, J. F. (2001). Conditional response of the human steroidogenic acute regulatory protein gene promoter to sterol regulatory element binding protein-1a 1. *Endocrinology*, 142(1), 28–36.
- Christou, G. A., and Kiortsis, D. N. (2013). Adiponectin and lipoprotein metabolism. *Obesity Reviews*, 14(12), 939-949.
- Chu, W. S., Furusato, B., Wong, K., Sesterhenn, I. A., Mostofi, F. K., Wei, M. Q., and Liang, Q. (2005). Ultrasound-accelerated formalin fixation of tissue improves morphology, antigen and mRNA preservation. *Modern Pathology*, 18(6), 850-863.
- Chua, K. (2007). *Studies on Andrographis Paniculata (Burm. f.) Nees (HDM 15) a Medicinal Native Plant of Brunei Darussalam*. PhD Thesis. Universiti of Bradford, UK.
- Chuang, C. C., and McIntosh, M. K. (2011). Potential mechanisms by which polyphenol-rich grapes prevent obesity-mediated inflammation and metabolic diseases. *Annual Review of Nutrition*, 31, 155–176.
- Cohen, P., and Friedman, J. M. (2004). Leptin and the control of metabolism: role for stearoyl-CoA desaturase-1 (SCD-1). *The Journal of Nutrition*, 134(9), 2455S-2463S.
- Cohen, P., Miyazaki, M., Socci, N. D., Hagge-Greenberg, A., Liedtke, W., Soukas, A. A., and Friedman, J. M. (2002). Role for stearoyl-CoA desaturase-1 in leptin-mediated weight loss. *Science*, 297(5579), 240-243.
- Considine, R. V., Sinha, M. K., Heiman, M. L., Kriauciunas, A., Stephens, T. W., Nyce, M. R., and Caro, J. F. (1996). Serum immune-reactive-leptin concentrations in normal-weight and obese humans. *New England Journal of Medicine*, 334 (5), 292-295.

- Cook, K. S., Min, H.Y., Johnson, D., Chaplinsky, R. J., Flier, J.S., Hunt, C. R., and Spiegelman, B.M. (1987). Adipsin: a circulating serine protease homolog secreted by adipose tissue and sciatic nerve. *Science*, 237(4813), 402-405.
- Cos, P., Vlietinck, A. J., Berghe, D. V., and Maes, L. (2006). Anti-infective potential of natural products: how to develop a stronger in vitro ‘proof-of-concept’. *Journal of ethnopharmacology*, 106(3), 290-302.
- Costa, D. C., Costa, H. S., Albuquerque, T. G., Ramos, F., Castilho, M. C., and Sanches-Silva, A. (2015). Advances in phenolic compounds analysis of aromatic plants and their potential applications. *Trends in Food Science and Technology*, , 45(2), 336-354. <https://doi.org/10.1016/j.tifs.2015.06.009>.
- Costacou, T., and Mayer-Davis, E. J. (2003). Nutrition and prevention of type 2 diabetes. *Annual Review of Nutrition*, 23(1), 147-170.
- Courts, F. L., and Williamson, G. (2015). The occurrence, fate and biological activities of C-glycosyl flavonoids in the human diet. *Critical Reviews in Food Science and Nutrition*, 55(10), 1352-1367.
- Cui, D., Daley, W. P., Fratkin, J. D., Haines, D. E., Lynch, J. C., Naftel, J. P., and Yang, G. (2011). *Atlas of histology: with functional and clinical correlations*. Wolters Kluwer/Lippincott Williams & Wilkins.
- Culp, S. J. (2004). *NTP Technical Report on the Toxicity Studies of Malachite Green Chloride and Leucomalachite Green*. National Toxicology Program; Toxicity Report Series N71: NIH publication No. 04-4416.
- Cunha, C. A., Lira, F. S., Rosa Neto, J. C., Pimentel, G. D., Souza, G. I. H., da Silva, C. M.G., and Oller do Nascimento, C. M. (2013). Green tea extract supplementation induces the lipolytic pathway, attenuates obesity, and reduces low-grade inflammation in mice fed a high-fat diet. *Mediators of Inflammation*, 2013. <http://dx.doi.org/10.1155/2013/635470>
- Currie-Elolf, L. M. (2011). *The Association between Obesity, Cognition, and Visual Function*. PhD Thesis. University of North Texas Health Science Center at Fort Worth, USA.
- Czaja, J. A. (1984). Sex differences in the activational effects of gonadal hormones on food intake and body weight. *Physiology and Behavior*, 33(4), 553-558.
- Dabbagh, A., and Rajaei, S. (2013). The role of anesthetic drugs in liver apoptosis. *Hepatitis Monthly*, 13(8), e13162.
- Dai J. L, Zhu Y. G, Zhang M, Huang Y. Z. (2004). Selecting iodine-enriched vegetables and the residual effect of iodate application to soil. *Biological Trace Element Research*, 101(3), 265–276.

- Dandona, P., Aljada, A., and Bandyopadhyay, A. (2004). Inflammation: the link between insulin resistance, obesity and diabetes. *Trends in Immunology*, 25(1), 4-7.
- Dans, A., Ng, N., Varghese, C., Tai, E. S., Firestone, R., and Bonita, R. (2011). The rise of chronic non-communicable diseases in Southeast Asia: time for action. *The Lancet*, 377(9766), 680-689.
- Darmady, E. M., and Stranack, F. (1957). Micro dissection of the nephron in disease. *British Medical Bulletin*, 13(1), 21-25.
- Das, U. N. (2007). Acetylcholinesterase and butyrylcholinesterase as possible markers of low-grade systemic inflammation. *Medical Science Monitor*, 13(12), RA214-RA221.
- Davidson, M. H., Hauptman, J., DiGirolamo, M., Foreyt, J. P., Halsted, C. H., Heber, D., and Heymsfield, S. B. (1999). Weight control and risk factor reduction in obese subjects treated for 2 years with orlistat: a randomized controlled trial. *Jama*, 281(3), 235-242.
- de Boer, H. J., Kool, A., Broberg, A., Mziray, W. R., Hedberg, I., and Levenfors, J. J. (2005). Anti-fungal and anti-bacterial activity of some herbal remedies from Tanzania. *Journal of Ethnopharmacology*, 96(3), 461-469.
- De Caterina, R. (2011). N-3 fatty acids in cardiovascular disease. *New England Journal of Medicine*, 364(25), 2439-2450.
- De Matteis, F., and Smith, L. L. (1995). *Molecular and Cellular Mechanisms of Toxicity*. Florida, USA: CRC Press. p. 167-171.
- de Melo, C. L., Queiroz, M. G. R., Fonseca, S. G. C., Bizerra, A. M. C., Lemos, T. L. G., Melo, T. S., and Rao, V. S. (2010). Oleanolic acid, a natural triterpenoid improves blood glucose tolerance in normal mice and ameliorates visceral obesity in mice fed a high-fat diet. *Chemico-Biological Interactions*, 185(1), 59-65.
- de Nigris, F., Balestrieri, M. L., Williams-Ignarro, S., Darmiento, F. P., Fiorito, C., Ignarro, L. J., and Napoli, C. (2007). The influence of pomegranate fruit extract in comparison to regular pomegranate juice and seed oil on nitric oxide and arterial function in obese Zucker rats. *Nitric Oxide*, 17(1), 50-54.
- Decker, E. A., Warner, K., Richards, M. P., and Shahidi, F. (2005). Measuring antioxidant effectiveness in food. *Journal of Agricultural and Food Chemistry*, 53(10), 4303-4310.
- Delany, J. P., Blohm, F., Truett, A. A., Scimeca, J. A., and West, D. B. (1999). Conjugated linoleic acid rapidly reduces body fat content in mice without affecting energy intake. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 276(4), R1172-R1179.

- Dennis, J., and Witting, P. (2017). Protective role for antioxidants in acute kidney disease. *Nutrients*, 9(7), 718.
- De Gasquet, P., and Pequignot, E. (1972). Lipoprotein lipase activities in adipose tissues, heart and diaphragm of the genetically obese mouse (ob-ob). *Biochemical Journal*, 127(2), 445.
- Di Carlo, G., Mascolo, N., Izzo, A. A., and Capasso, F. (1999). Flavonoids: old and new aspects of a class of natural therapeutic drugs. *Life Sciences*, 65(4), 337-353.
- Diet, C. B., Diet, U., and Diet, N. P. (1977). Report of the American Institute of Nutrition ad hoc committee on standards for nutritional studies. *Journal of Nutrition*, 107, 1340.
- Ding, L., Li, J., Song, B., Xiao, X., Zhang, B., Qi, M., Huang, W., Yang, Li., and Wang, Z. (2016). Curcumin rescues high-fat diet induced obesity and insulin sensitivity in mice through regulating SREBP pathway. *Toxicology and Applied Pharmacology*, 304, 99-109.
- Dinkova-Kostova, A. T., and Talalay, P. (2008). Direct and indirect antioxidant properties of inducers of cytoprotective proteins. *Molecular Nutrition and Food Research*, 52(S1), S128-S138.
- Dixit, M., Doan, T., Kirschner, R., and Dixit, N. (2010). Significant acute kidney injury due to non-steroidal anti-inflammatory drugs: inpatient setting. *Pharmaceuticals*, 3(4), 1279-1285.
- Dong, Z., Wu, T., Qin, W., An, C., Wang, Z., Zhang, M., and An, F. (2011). Serum amyloid A directly accelerates the progression of atherosclerosis in apolipoprotein E-deficient mice. *Molecular Medicine*, 17(11-12), 1357-1364.
- Droge, W. (2002). Free radicals in the physiological control of cell function. *Physiological Reviews*, 82(1), 47-95.
- Duthie, G. G., Gardner, P. T., and Kyle, J. A. M. (2003). Plant polyphenols: are they the new magic bullet? *Proceedings of the Nutrition Society*, 62(3), 599-603.
- Ebrahimi, M., Rajion, M. A., Goh, Y. M., Sazili, A. Q., and Schonewille, J. T. (2013). Effect of linseed oil dietary supplementation on fatty acid composition and gene expression in adipose tissue of growing goats. *BioMed Research International*, 2013. Retrieved from <http://dx.doi.org/10.1155/2013/194625>.
- Ebrahimi, M., Rajion, M. A., Meng, G. Y., and Soleimani, A. (2014). Omega-3 fatty acid enriched chevon (goat meat) lowers plasma cholesterol levels and alters gene expressions in rats. *BioMed Research International*. Retrieved from <http://dx.doi.org/10.1155/2014/749341>.
- Eckel, R. H. (2008). Nonsurgical management of obesity in adults. *New England Journal of Medicine*, 358(18), 1941-1950.

- Eckhert, C. D., Lockwood, M. K., and Shen, B. (1993). Influence of selenium on the microvasculature of the retina. *Microvascular Research*, 45(1), 74-82.
- Elkhoury, D., and Anderson, G. H. (2013). Recent advances in dietary proteins and lipid metabolism. *Current Opinion in Lipidology*, 24(3), 207-213.
- Ellman, G. L., Courtney, K. D., Andres, V., and Featherstone, R. M. (1961). A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochemical Pharmacology*, 7(2), 88-95.
- Everds, N. E., Snyder, P. W., Bailey, K. L., Bolon, B., Creasy, D. M., Foley, G. L., and Sellers, T. (2013). Interpreting stress responses during routine toxicity studies: a review of the biology, impact, and assessment. *Toxicologic Pathology*, 41(4), 560-614.
- Ezejiofor, A. N., Orish, C. N., and Orisakwe, O. E. (2013). Effect of aqueous leaves extract of *Costus afer Ker Gawl* (Zingiberaceae) on the liver and kidney of male albino Wistar rat. *Ancient Science of Life*, 33(1), 4.
- Fabricatore, A. N., and Wadden, T. A. (2003). Psychological functioning of obese individuals. *Diabetes Spectrum*, 16(4), 245-252.
- Fain, J. N. (2010). Release of inflammatory mediators by human adipose tissue is enhanced in obesity and primarily by the nonfat cells: a review *Mediators of Inflammation*, 2010. Retrieved from <http://dx.doi.org/10.1155/2010/513948>.
- Farmer, S. R. (2006). Transcriptional control of adipocyte formation. *Cell Metabolism*, 4(4), 263-273.
- Farsi, E., Majid, A. S., and Majid, A. M. S. (2016a). *Clinacanthus nutans*, yesterday's practice, and future's drug: a comprehensive. Review. *American Journal of Phytomedicine and Clinical Therapeutics*, 4(04), 113-126
- Farsi, E., Esmailli, K., Shafaei, A., Moradi Khaniabadi, P., Al Hindi, B., Khadeer Ahamed, MB., Sandai, D., Abdul Sattar, M., Ismail, Z., and Abdul Majid, AMS. (2016b). Mutagenicity and preclinical safety assessment of the aqueous extract of *Clinacanthus nutans* leaves. *Drug and Chemical Toxicology*, 39(4), 461-473
- Feige, J. N., Gelman, L., Michalik, L., Desvergne, B., and Wahli, W. (2006). From molecular action to physiological outputs: peroxisome proliferator-activated receptors are nuclear receptors at the crossroads of key cellular functions. *Progress in Lipid Research*, 45(2), 120-159.
- Felhi, S., Daoud, A., Hajlaoui, H., Mnafgui, K., Gharsallah, N., and Kadri, A. (2017). Solvent extraction effects on phytochemical constituents profiles, antioxidant and antimicrobial activities and functional group analysis of Ecballium elaterium seeds and peels fruits. *Food Science and Technology*, 37(3), 483-492.

- Fernandes, E. F. A., Meloni, F., Borella, J. C., and Lopes, N. P. (2013). Effect of fertilisation and harvest period on polar metabolites of *Calendula officinalis*. *Revista Brasileira de Farmacognosia*, 23(5), 731-735.
- Fernandez-Mar, M. I., Mateos, R., Garcia-Parrilla, M. C., Puertas, B., and Cantos-Villar, E. (2012). Bioactive compounds in wine: resveratrol, hydroxytyrosol and melatonin: a review. *Food Chemistry*, 130(4), 797-813.
- Fernandez-Riejos, P., Najib, S., Santos-Alvarez, J., Martin-Romero, C., Perez-Perez, A., Gonzalez-Yanes, C., and Sanchez-Margalef, V. (2010). Role of leptin in the activation of immune cells. *Mediators of Inflammation*, 2010.
- Ferraz, R. R. N., Tiselius, H. G., and Heiberg, I. P. (2004). Fat malabsorption induced by gastrointestinal lipase inhibitor leads to an increase in urinary oxalate excretion. *Kidney International*, 66(2), 676-682.
- Ferre, P. (2004). The biology of peroxisome proliferator-activated receptors: relationship with lipid metabolism and insulin sensitivity. *Diabetes*, 53(suppl1), S43-S50. Retrieved from <https://doi.org/10.2337/diabetes.53.2007.S43>.
- Ferrer, F., Silva, B. M., Andrade, P. B., Seabra, R. M., and Ferreira, M. A. (2003). Approach to the study of C-glycosyl flavones by ion trap HPLC-PAD-ESI/MS/MS: application to seeds of quince (*Cydonia oblonga*). *Phytochemical Analysis: An International Journal of Plant Chemical and Biochemical Techniques*, 14(6), 352-359.
- Fidrianny, I., Permatasari, L. and Wirasutisna, K.R. (2013). Antioxidant activities frm various bulbs extracts of three kinds allium using DPPH, ABTS assays and correlation with total phenolic, flavonoid, carotenoid content. *International Journal of Research and Pharmaceutical Sciences*, 4(3), 438-444.
- Finer, N., James, W. P. T., Kopelman, P. G., Lean, M. E. J., and Williams, G. (2000). One-year treatment of obesity: a randomized, double-blind, placebo-controlled, multicentre study of orlistat, a gastrointestinal lipase inhibitor. *International Journal of Obesity*, 24(3), 306.
- Finkelstein, E. A., Khavjou, O. A., Thompson, H., Trogdon, J. G., Pan, L., Sherry, B., and Dietz, W. (2012). Obesity and severe obesity forecasts through 2030. *American Journal of Preventive Medicine*, 42(6), 563-570.
- Flegal, K. M., Kit, B. K., Orpana, H., and Graubard, B. I. (2013). Association of all-cause mortality with overweight and obesity using standard body mass index categories: a systematic review and meta-analysis. *JAMA*, 309(1), 71-82.
- Flier, J. S. (1995). The adipocyte: storage depot or node on the energy information superhighway. *Cell*, 80(1), 15-18.

- Floegel, A., Kim, D. O., Chung, S. J., Koo, S. I., and Chun, O. K. (2011). Comparison of ABTS/DPPH assays to measure antioxidant capacity in popular antioxidant-rich US foods. *Journal of Food Composition and Analysis*, 24(7), 1043-1048.
- Flowers, J.B., Rabaglia, M.E., Schueler, K.L., Flowers, M.T., Lan, H., and Keller, M.P. (2007). Loss of stearoyl-CoA desaturase improves insulin sensitivity in lean mice but worsens diabetes in leptin-deficient obese mice. *Diabetes*, 56, 1228-1239.
- Folch, J., Lees, M., and Sloane Stanley, G. (1957). A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biology Chemistry*, 226(1), 497-509.
- Fong, S. Y. (2015). *Genetic, Phytochemical and Bioactivity Studies of Clinacanthus nutans (Burm. f.) Lindau (Acanthaceae)*. PhD Thesis. Applied Science, Royal Melbourne Institute of Technology Universiti, Australia.
- Formanek, Z., Kerry, J. P., Higgins, F. M., Buckley, D. J., Morrissey, P. A., and Farkas, J. (2001). Addition of synthetic and natural antioxidants to  $\alpha$ -tocopheryl acetate supplemented beef patties: effects of antioxidants and packaging on lipid oxidation. *Meat Science*, 58(4), 337-341.
- Foster, M. T., and Pagliassotti, M. J. (2012). Metabolic alterations following visceral fat removal and expansion: beyond anatomic location. *Adipocyte*, 1(4), 192-199.
- Foufelle, F., and Ferre, P. (2002). New perspectives in the regulation of hepatic glycolytic and lipogenic genes by insulin and glucose: a role for the transcription factor sterol regulatory element binding protein-1c. *Biochemical Journal*, 366(2), 377-391.
- Fraga, C. G., Galleano, M., Verstraeten, S. V., and Oteiza, P. I. (2010). Basic biochemical mechanisms behind the health benefits of polyphenols. *Molecular Aspects of Medicine*, 31(6), 435-445.
- Francis, F.J. (2000). *Wiley Encyclopedia of Food Science and Technology*. (2 nd edition). (PP. 2352-2356). New York: A Wiley- Interscience publication John Wiley & Sons, Inc,
- Fried, S. K., Ricci, M. R., Russell, C. D., and LaFerrere, B. (2000). Regulation of leptin production in humans. *The Journal of Nutrition*, 130(12), 3127S-3131S. <https://doi.org/10.1093/jn/130.12.3127S>.
- Fritz, R.S., Hochwender, C.G., Lewkiewicz, D.A., Bothwell, S., and Orians, C.M. (2001). Seedling herbivory by slugs in a willow hybrid system: Developmental changes in damage, chemical defense, and plant performance. *Oecologia*, 129(1), 87-97.

- Frossard, E., Bucher, M., Mächler, F., Mozafar, A., and Hurrell, R. (2000). Potential for increasing the content and bioavailability of Fe, Zn and Ca in plants for human nutrition. *Journal of the Science of Food and Agriculture*, 80(7), 861-879.
- Fuhrman, B., Buch, S., Vaya, J., Belinky, P. A., Coleman, R., Hayek, T., and Aviram, M. (1997). Licorice extract and its major polyphenol glabridin protect low-density lipoprotein against lipid peroxidation: in vitro and ex vivo studies in humans and in atherosclerotic apolipoprotein E-deficient mice. *The American Journal of Clinical Nutrition*, 66(2), 267-275.
- Funabashi, T., Hagiwara, H., Mogi, K., Mitsushima, D., Shinohara, K., and Kimura, F. (2009). Sex differences in the responses of orexin neurons in the lateral hypothalamic area and feeding behavior to fasting. *Neuroscience Letters*, 463(1), 31-34.
- Furuyashiki, T., Nagayasu, H., Aoki, Y., Bessho, H., Hashimoto, T., Kanazawa, K., and Ashida, H. (2004). Tea catechin suppresses adipocyte differentiation accompanied by down-regulation of PPAR $\gamma$ 2 and C/EBP $\alpha$  in 3T3-L1 cells. *Bioscience, Biotechnology, and Biochemistry*, 68(11), 2353–2359.
- Gastaldelli, A., Kozakova, M., Højlund, K., Flyvbjerg, A., Favuzzi, A., Mitrakou, A., and Balkau, B. (2009). Fatty liver is associated with insulin resistance, risk of coronary heart disease, and early atherosclerosis in a large European population. *Hepatology*, 49(5), 1537-1544.
- Geloen, A. L. A. I. N., Roy, P. E., and Bukowiecki, L. J. (1989). Regression of white adipose tissue in diabetic rats. *American Journal of Physiology-Endocrinology and Metabolism*, 257(4), E547-E553.
- Gervois, P., Fruchart, J. C., and Staels, B. (2007). Drug Insight: mechanisms of action and therapeutic applications for agonists of peroxisome proliferator-activated receptors. *Nature Clinical Practice Endocrinology and Metabolism*, 3(2), 145-156.
- Ghasemzadeh, A. (2011). *Effect of different light intensities and CO<sub>2</sub> enrichment on yield and pharmaceutical quality of young ginger (Zingiber officinale roscoe)*. Doctoral dissertation. PhD Thesis, Universiti Putra, Malaysia.
- Ghasemzadeh, A., and Ghasemzadeh, N. (2011). Flavonoids and phenolic acids: Role and biochemical activity in plants and human. *Journal of Medicinal Plants Research*, 5(31), 6697-6703.
- Ghasemzadeh, A., Azarifar, M., Soroodi, O., and Jaafar, H. Z. (2012). Flavonoid compounds and their antioxidant activity in extract of some tropical plants. *Journal of Medicinal Plants Research*, 6(13), 2639-2643.

- Ghasemzadeh, A., Nasiri, A., Jaafar, H., Baghdadi, A., and Ahmad, I. (2014). Changes in phytochemical synthesis, chalcone synthase activity and pharmaceutical qualities of Sabah snake grass (*Clinacanthus nutans* L.) in relation to plant age. *Molecules*, 19(11), 17632-17648.
- Giannini, E. G., Testa, R., and Savarino, V. (2005). Liver enzyme alteration: a guide for clinicians. *Canadian Medical Association Journal*, 172(3), 367-379.
- Gibellini, L., Pinti, M., Nasi, M., Montagna, J. P., De Biasi, S., Roat, E., and Cossarizza, A. (2011). Quercetin and cancer chemoprevention. *Evidence-Based Complementary and Alternative Medicine*, 2011.
- Goh, L. Y., and Goh, K. L. (2013). Obesity: an epidemiological perspective from Asia and its relationship to gastrointestinal and liver cancers. *Journal of Gastroenterology and Hepatology*, 28(S4), 54-58.
- Gomes, C., Lourenço, E. L. B., Liuti, É. B., Duque, A. O., Nihi, F., Lourenço, A. C., and Dalsenter, P. R. (2012). Evaluation of subchronic toxicity of the hydroethanolic extract of *Tropaeolum majus* in Wistar rats. *Journal of Ethnopharmacology*, 142(2), 481-487.
- Gonzalez-Castejon, M., and Rodriguez-Casado, A. (2011). Dietary phytochemicals and their potential effects on obesity: A review. *Pharmacological Research*, 64(5), 438-455. Retrieved from <https://doi.org/10.1016/j.phrs.2011.07.004>
- Gotes, J., Kasian, K., Jacobs, H., Cheng, Z. Q., and Mink, S. N. (2012). Benefits of ethyl gallate versus norepinephrine in the treatment of cardiovascular collapse in *Pseudomonas aeruginosa* septic shock in dogs. *Critical Care Medicine*, 40(2), 560-572.
- Graf, D., Seifert, S., Jaudszus, A., Bub, A., and Watzl, B. (2013). Anthocyanin-rich juice lowers serum cholesterol, leptin, and resistin and improves plasma fatty acid composition in -fischer rats. *Plos One*, 8(6), e66690.
- Greaves, P. (2011). *Histopathology of Preclinical Toxicity Studies: Interpretation and Relevance in Drug Safety Evaluation*. 4<sup>th</sup> Ed. Oxford, UK: Academic Press. p. 291-303,
- Green, B., and Duffull, S. B. (2004). What is the best size descriptor to use for pharmacokinetic studies in the obese?. *British journal of clinical pharmacology*, 58(2), 119-133.
- Gross, P. (2001). Treatment of severe hyponatremia. *Kidney International*, 60(6), 2417-2427.
- Grygiel-Górniak, B. (2014). Peroxisome proliferator-activated receptors and their ligands: nutritional and clinical implications-a review. *Nutrition Journal*, 13(1), 17.

- Guerrero-Berroa, E., Schmeidler, J., and Beeri, M. S. (2014). Neuropathology of type 2 diabetes: a short review on insulin-related mechanisms. *European Neuropsychopharmacology*, 24(12), 1961-1966
- Guicciardi, M. E., Malhi, H., Mott, J. L., and Gores, G. J. (2013). Apoptosis and necrosis in the liver. *Comprehensive Physiology*, 3(2), 977-1010.
- Guilherme, A., Virbasius, J. V., Puri, V., and Czech, M. P. (2008). Adipocyte dysfunctions linking obesity to insulin resistance and type 2 diabetes. *Nature Reviews Molecular Cell Biology*, 9(5), 367-377.
- Gurib-Fakim, A. (2006). Medicinal plants: traditions of yesterday and drugs of tomorrow. *Molecular Aspects of Medicine*, 27(1), 1-93.
- Gustafson, B., Hammarstedt, A., Andersson, C. X., and Smith, U. (2007). Inflamed adipose tissue a culprit underlying the metabolic syndrome and atherosclerosis. *Arteriosclerosis, Thrombosis and Vascular Biology*, 27(11), 2276-2283
- Gustavsson, C., Yassin, K., Wahlström, E., Cheung, L., Lindberg, J., Brisimar, K., and Tollet-Egnell, P. (2010). Sex-different hepatic glycogen content and glucose output in rats. *BMC Biochemistry*, 11(1), 38.
- Guyton, A., and Hall, J. (2006). Textbook of medical physiology, 11<sup>th</sup> Ed. Philadelphia, Pennsylvania 19103 ISBN 0-7216-0240-1
- Guzik, T., Korbut, R., and Adamek-Guzik, T. (2003). Nitric oxide and superoxide in inflammation. *Journal of Physiology and Pharmacology*, 54(4), 469-487
- Hafez, E. M., Sahar, I. Y., and Safaa, A. M. (2015). Parenchymatous toxicity of tramadol: histopathological and biochemical study. *J Alcohol Drug Depend*, 3(5), 225.
- Hagerstrand, I. (1975). Distribution of alkaline phosphatase activity in healthy and diseased human liver tissue. *Acta Pathologica Microbiologica Scandinavica Section A Pathology*, 83(5), 519-526.
- Hajiboland, R., and Amjad, L. (2007). Does antioxidant capacity of leaves play a role in growth response to selenium at different sulfur nutritional status? *Plant Soil and Environment*, 53(5), 207.
- Haley, M. (2014). *Mechanisms of Interaction between Obesity and Ischaemic Stroke*. PhD Thesis. Manchester University, UK.
- Hall, P. and Cash, J. (2012). What is the real function of the Liver 'function' tests? *The Ulster Medical Journal*, 81(1), 30.
- Hamza, R. Z., and Alharbi, M. S. (2015). Amelioration of paracetamol hepatotoxicity and oxidative stress on mice liver with *Silymarin* and *Nigella sativa* extract supplements. *Asian Pacific Journal of Tropical Biomedicine*, 5(7), 521-531.

- Handjieva-Darlenska, T., and Boyadjieva, N. (2009). The effect of high-fat diet on plasma ghrelin and leptin levels in rats. *Journal of Physiology and Biochemistry*, 65(2), 157-164.
- Hanhineva, K., Törrönen, R., Bondia-Pons, I., Pekkinen, J., Kolehmainen, M., Mykkänen, H., and Poutanen, K. (2010). Impact of dietary polyphenols on carbohydrate metabolism. *International Journal of Molecular Sciences*, 11(4), 1365-1402.
- Hanl, L. K., Kimura, Y., and Okuda, H. (2005). Anti-obesity effects of natural products. *Studies in Natural Products Chemistry*, 30, 79–110.
- Harada, M., Hanada, S., Toivola, D. M., Ghori, N., and Omary, M. B. (2008). Autophagy activation by rapamycin eliminates mouse Mallory-Denk bodies and blocks their proteasome inhibitor-mediated formation. *Hepatology*, 47(6), 2026-2035.
- Harborne, J. B., and Simmonds, N. W. (1964). *The Natural Distribution of the Phenolic Aglycones. Biochemistry of Phenolic Compounds*. London and New York: Academic Press.
- Harizal, S.N., Mansor S. M., Hasnan, J., Tharakan, J. K., and Abdullah, J. (2010). Acute toxicity study of standardized methanolic extract of *Mitragyna speciose* Korth in rodent. *Journal Enthapharmacol*, 131(2), 404-409.
- Hartikainen, H., Xue, T., and Piironen, V. (2000). Selenium as an anti-oxidant and pro-oxidant in ryegrass. *Plant and Soil*, 225(1-2), 193-200.
- Harvey, J., and Withers, D. J. (2008). *Neurobiology of Obesity*. New York, USA: Cambridge University Press.
- Hawrylak-Nowak, B. (2013). Comparative effects of selenite and selenate on growth and selenium accumulation in lettuce plants under hydroponic conditions. *Plant Growth Regulation*, 70(2), 149-157.
- Hayashi, T., Boyko, E. J., McNeely, M. J., Leonetti, D. L., Kahn, S. E., and Fujimoto, W. Y. (2008). Visceral adiposity, not abdominal subcutaneous fat area, is associated with an increase in future insulin resistance in Japanese Americans. *Diabetes*, 57(5), 1269-1275.
- He, R. R., Chen, L., Lin, B. H., Matsui, Y., Yao, X. S., and Kurihara, H. (2009). Beneficial effects of oolong tea consumption on diet-induced overweight and obese subjects. *Chinese Journal of Integrative Medicine*, 15(1), 34-41.
- Heal, D. J., Aspley, S., Prow, M. R., Jackson, H. C., Martin, K. F., and Cheetham, S. C. (1998). Sibutramine: a novel anti-obesity drug. A review of the pharmacological evidence to differentiate it from d-amphetamine and d-fenfluramine. *International Journal of Obesity*, 22(1), S18-S28.

- Heal, D. J., Gosden, J., and Smith, S. L. (2012). What is the prognosis for new centrally-acting anti-obesity drugs? *Neuropharmacology*, 63(1), 132-146.
- Heinonen, S., Saarinen, L., Naukkarinen, J., Rodriguez, A., Fruhbeck, G., Hakkarainen, A., and Moilanen, E. (2014). Adipocyte morphology and implications for metabolic derangements in acquired obesity. *International Journal of Obesity*, 38(11), 1423-1431.
- Hofbauer, K. G., Nicholson, J. R., and Boss, O. (2007). The obesity epidemic: current and future pharmacological treatments. *Annual Review Pharmacology Toxicology*, 47, 565-592.
- Holben, D. H., and Smith, A. M. (1999). The diverse role of selenium within selenoproteins: a review. *Journal of the American Dietetic Association*, 99(7), 836-843.
- Holm, C., Kirchgessner, T. G., Svenson, K. L., Fredrikson, G., Nilsson, S., Miller, C. G., and Mohandas, T. (1988). Hormone-sensitive lipase: sequence, expression, and chromosomal localization to 19 cent-q13. 3. *Science*, 241(4872), 1503-1506.
- Hong, J., Stubbins, R. E., Smith, R. R., Harvey, A. E., and Nunez, N. P. (2009). Differential susceptibility to obesity between male, female and ovariectomized female mice. *Nutrition Journal*, 8(1), 11.
- Hor, S. Y., Ahmad, M., Farsi, E., Lim, C. P., Asmawi, M. Z., and Yam, M. F. (2011). Acute and subchronic oral toxicity of *Coriolus versicolor* standardized water extract in Sprague-Dawley rats. *Journal of Ethnopharmacology*, 137(3), 1067-1076.
- Horie, T., Ono, K., Horiguchi, M., Nishi, H., Nakamura, T., Nagao, K., and Hasegawa, K. (2010). MicroRNA-33 encoded by an intron of sterol regulatory element-binding protein 2 (Srebp2) regulates HDL in vivo. *Proceedings of the National Academy of Sciences*, 107(40), 17321-17326.
- Horton, J. D., Goldstein, J. L., and Brown, M. S. (2002). SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver. *The Journal of Clinical Investigation*, 109(9), 1125-1131.
- Hossain, M. A., AL-Raqmi, K. A. S., AL-Mijizy, Z. H., Weli, A. M., and Al-Riyami, Q. (2013). Study of total phenol, flavonoids contents and phytochemical screening of various leaves crude extracts of locally grown Thymus vulgaris. *Asian Pacific Journal of Tropical Biomedicine*, 3(9), 705-710.
- Householder, L. A. (2013). *Examining the Impact of Growth Hormone on the Collagen Content of Adipose Tissue in Transgenic Mice*. (Doctoral dissertation, Ohio University), USA.

- Hsu, H.M., Chen, W.Y., Pan, P.H., and Mao, F. C. (2014). *Vitis thunbergii* supplementation demonstrates an anti-obesity effect in developing obese mice. *European Journal of Integrative Medicine*, 6(5), 581-587.
- Hsu, Y. W., Tsai, C. F., Chen, W. K., Huang, C. F., and Yen, C. C. (2011). A subacute toxicity evaluation of green tea (*Camellia sinensis*) extract in mice. *Food and Chemical Toxicology*, 49(10), 2624-2630.
- Hu, C.C., Qing, K. and Chen, Y. (2004). Diet-induced changes in stearoyl-CoA desaturase 1 expression in obesity-prone and -resistant mice. *Obesity Research*, 21(8), 1264–1270.
- Huang, J., Wang, Y., Xie, Z., Zhou, Y., Zhang, Y., and Wan, X. (2014). The anti-obesity effects of green tea in human intervention and basic molecular studies. *European Journal of Clinical Nutrition*, 68(10), 1075-1087.
- Huang, W., Dedousis, N., Bandi, A., Lopaschuk, G. D., and O'Doherty, R. M. (2006). Liver triglyceride secretion and lipid oxidative metabolism are rapidly altered by leptin in vivo. *Endocrinology*, 147(3), 1480-1487.
- Huerta-Leidenz N. O., Cross H. R., Savell J. W., Lunt D. K., Baker J. F., and Smith S. B. (1996). Fatty acid composition of subcutaneous adipose tissue from male calves at different stages of growth. *Journal of Animal Sciences*, 74(6), 1256-1264
- Humasti, S., and Hotamisligil, G. S. (2010). Endoplasmic reticulum stress and inflammation in obesity and diabetes. *Circulation Research*, 107(5), 579–591.
- Hursel, R., and Westerterp-Plantenga, M. S. (2010). Thermogenic ingredients and body weight regulation. *International Journal of Obesity*, 34(4), 659-669.
- Hursel, R., and Westerterp-Plantenga, M. S. (2013). Catechin-and caffeine-rich teas for control of body weight in humans. *The American Journal of Clinical Nutrition*, 98(6), 1682S-1693S.
- Hwang, C. S., Loftus, T. M., Mandrup, S., and Lane, M. D. (1997). Adipocyte differentiation and leptin expression. *Annual Review of Cell and Developmental Biology*, 13(1), 231-259.
- Hynie, S., and Klenerova, V. (1980). Effects of dimethyl sulfoxide and other dipolar aprotic solvents on rat hepatic adenylate cyclase. Potentiating effects on glucagon and guanylylimidodiphosphate stimulation. *Naunyn-Schmiedeberg's Archives of Pharmacology*, 310(3), 231-236.
- Ibrahim, M. H., Jaafar, H. Z., Rahmat, A., and Rahman, Z. A. (2011). The relationship between phenolics and flavonoids production with total non-structural carbohydrate and photosynthetic rate in *Labisia pumila* Benth under high CO<sub>2</sub> and nitrogen fertilization. *Molecules*, 16(1), 162-174.

- Ibrahim, K., Al-Mutary, M., Bakhet, A., & Khan, H. (2018). Histopathology of the liver, kidney, and spleen of mice exposed to gold nanoparticles. *Molecules*, 23(8), 1848.
- Ichimura, T., Hung, C. C., Yang, S. A., Stevens, J. L., and Bonventre, J. V. (2004). Kidney injury molecule-1: a tissue and urinary biomarker for nephrotoxicant-induced renal injury. *American Journal of Physiology-Renal Physiology*, 286(3), F552-F563.
- Ide, Y., Waki, M. Hayasaka, T., Nishio, T., Morita, Y., Tanaka, H., and Hosokawa, Y. (2013). Human breast cancer tissues contain abundant phosphatidylcholine (36:1) with high stearoyl-CoA desaturase-1 expression. *Plos One*, 8(4), e61204.
- Igal, R. A. (2010). Stearoyl-CoA desaturase-1: a novel key player in the mechanisms of cell proliferation, programmed cell death and transformation to cancer. *Carcinogenesis*, 31(9), 1509–1515. <https://doi.org/10.1093/carcin/bgq131>
- Ikeda, I., Imasato, Y., Sasaki, E., Nakayama, M., Nagao, H., Takeo, T., and Sugano, M. (1992). Tea catechins decrease micellar solubility and intestinal absorption of cholesterol in rats. *Biochimica et Biophysica Acta (BBA)-Lipids and Lipid Metabolism*, 1127(2), 141-146.
- Im Choi, S., Jeong, C. S., Cho, S. Y., and Lee, Y. S. (2007). Mechanism of apoptosis induced by apigenin in HepG2 human hepatoma cells: involvement of reactive oxygen species generated by NADPH oxidase. *Archives of Pharmacal Research*, 30(10), 1328-1335.
- Imam, M. U., Musa, S. N. A., Azmi, N. H., and Ismail, M. (2012). Effects of white rice, brown rice and germinated brown rice on antioxidant status of type 2 diabetic rats. *International Journal of Molecular Sciences*, 13(10), 12952-12969.
- Inadera, H. (2008). The usefulness of circulating adipokine levels for the assessment of obesity-related health problems. *International Journal of Medical Sciences*, 5(5), 248-262
- Iqbal, S., Bhanger, M. I., and Anwar, F. (2005). Antioxidant properties and components of some commercially available varieties of rice bran in Pakistan. *Food Chemistry*, 93(2), 265-272.
- Iqbal, S., and Bhanger, M. I. (2006). Effect of season and production location on antioxidant activity of *Moringa oleifera* leaves grown in Pakistan. *Journal of Food Composition and Analysis*, 19(6-7), 544-551.
- Irshad, M., Zafaryab, M., Singh, M., and Rizvi, M. (2012). Comparative analysis of the antioxidant activity of *Cassia fistula* extracts. *International Journal of Medicinal Chemistry*, 2012.

- Ismail, N. Z., Arsal, H., Samian, M. R., and Hamdan, M. R. (2017). Determination of phenolic and flavonoid contents, antioxidant activities and GC-MS analysis of *Clinacanthus nutans* (Acanthaceae) in different locations. *AGRIVITA, Journal of Agricultural Science*, 39(3), 335-344.
- Iwasa, T., Matsuzaki, T., Yiliyasi, M., Yano, K., and Irahara, M. (2017). The effects of chronic testosterone administration on body weight, food intake, and fat weight were age-dependent. *Steroids*, 127, 18-23.
- Iwatani, M., Ikegami, K., Kremenska, Y., Hattori, N., Tanaka, S., Yagi, S., and Shiota, K. (2006). Dimethyl sulfoxide has an impact on epigenetic profile in mouse embryoid body. *Stem Cells*, 24(11), 2549-2556.
- James, W. P. T., Astrup, A., Finer, N., Hilsted, J., Kopelman, P., Rössner, S., and Storm Study Group. (2000). Effect of sibutramine on weight maintenance after weight loss: a randomised trial. *The Lancet*, 356(9248), 2119-2125.
- Jang, E. M., Choi, M. S., Jung, U. J., Kim, M. J., Kim, H. J., Jeon, S. M., and Lee, M. K. (2008). Beneficial effects of curcumin on hyperlipidemia and insulin resistance in high-fat fed hamsters. *Metabolism*, 57(11), 1576-1583.
- Jang, W.S., and Choung, S.Y. (2013). Antioesity effects of the ethanol extract of *laminaria japonica* areshoung in high-fat-diet-induced obese rat. *Evidence-Based Complementary and Alternative Medicine*, 492807, 1-17.
- Jeffcoat, R. (2007). Obesity- perspective based on the biochemical interrelationship of lipids and carbohydrates. *Medical Hypotheses*, 68(5), 1159-1171.
- Jensen, E. N., Buch-Andersen, T., Ravn-Haren, G., and Dragsted, L. O. (2009). Mini-review: The effects of apples on plasma cholesterol levels and cardiovascular risk—a review of the evidence. *The Journal of Horticultural Science and Biotechnology*, 84(6), 34-41.
- Jensen, T. L., Kiersgaard, M. K., Sørensen, D. B., and Mikkelsen, L. F. (2013). Fasting of mice: a review. *Laboratory Animals*, 47(4), 225-240.
- Jeong, Y.J., Sohm, E.H., Jung, Y.H., Yoon, W.J., Cho, Y.M., Kim, I., Lee, S.R., and Kang, S.C. (2016). Anti-obesity effect of *Crinum asiaticum* var. *Japonicum* Baker extract in high-fat diet-induced and monogenic obese mice. *Biomedicine and Pharmacotherapy*, 82, 35-43. <https://doi.org/10.1016/j.biopha.2016.04.067>
- Jesch, E. D., and Carr, T. P. (2017). Food ingredients that inhibit cholesterol absorption. *Preventive Nutrition and Food Science*, 22(2), 67-80.
- Jia, Y., Kim, S., Kim, J., Kim, B., Wu, C., Lee, J. H., and Lee, S. J. (2015). Ursolic acid improves lipid and glucose metabolism in high-fat-fed C57BL/6J mice by activating peroxisome proliferator-activated receptor alpha and hepatic autophagy. *Molecular Nutrition and Food Research*, 59(2), 344-354.

- Jiang, G., Li, Z., Liu, F., Ellsworth, K., Dallas-Yang, Q., and Wu, M. (2005). Prevention of obesity in mice by antisense oligonucleotide inhibitors of stearoyl-CoA desaturase-1. *Journal of Clinical Investigation*, 115(4), 1030-1038.
- Jie, Z.Y., Yue-qin, L., Shan-yan, C., and Jie, S. (2012). Effects of dietary energy level on the expression of the HSL gene in different tissues of sheep. *Journal of Integrative Agriculture*, 11(7), 1167-1172.
- Jobgen, W., Meininger, C. J., Jobgen, S. C., Li, P., Lee, M. J., Smith, S. B., and Wu, G. (2008). Dietary L-arginine supplementation reduces white fat gain and enhances skeletal muscle and brown fat masses in diet-induced obese rats. *The Journal of Nutrition*, 139(2), 230-237.
- Joo, S. Y., Song, Y. A., Park, Y. L., Myung, E., Chung, C. Y., Park, K. J., and Kim, N. S. (2012). Epigallocatechin-3-gallate inhibits LPS-induced NF- $\kappa$ B and MAPK signaling pathways in bone marrow-derived macrophages. *Gut and Liver*, 6(2), 188.
- Jothy, S. L., Zakaria, Z., Chen, Y., Lau, Y. L., Latha, L., and Sasidharan, S. (2011). Acute oral toxicity of methanolic seed extract of *Cassia fistula* in mice. *Molecules*, 16(6), 5268-5282.
- Jung, U. J., and Choi, M. S. (2014). Obesity and its metabolic complications: the role of adipokines and the relationship between obesity, inflammation, insulin resistance, dyslipidemia and nonalcoholic fatty liver disease. *International Journal of Molecular Sciences*, 15(4), 6184-6223.
- Juhel, C., Armand, M., Pafumi, Y., Rosier, C., Vandermander, J., and Lairon, D. (2000). Green tea extract (AR25®) inhibits lipolysis of triglycerides in gastric and duodenal medium in vitro. *The Journal of Nutritional Biochemistry*, 11(1), 45-51.
- Kadir, N.A., Rahmat, A., and Jaafat, H. Z. (2015). Protective effects of tamarillo (*Cyphomandra betacea*) extract against high-fat diet-induced obesity in Sprague-Dawley rats. *Journal of Obesity*, 2015. <http://dx.doi.org/10.1155/2015/846041>
- Kadowaki, T., Yamauchi, T., Kubota, N., Hara, K., Ueki, K., and Tobe, K. (2006). Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. *The Journal of Clinical Investigation*, 116(7), 1784-1792.
- Kamarudin, M. N. A., Sarker, M. M. R., Kadir, H. A., and Ming, L. C. (2017). Applications of *Clinacanthus nutans* (Burm. f.) Lindau: a comprehensive review. *Journal of Ethnopharmacology*, 206, 245-266.

- Kandhare, A. D., Bodhankar, S. L., Mohan, V., and Thakurdesai, P. A. (2016). Acute and repeated doses (28 days) oral toxicity study of Vicenin-1, a flavonoid glycoside isolated from fenugreek seeds in laboratory mice. *Regulatory Toxicology and Pharmacology*, 81, 522-531. <https://doi.org/10.1016/j.yrtph.2015.05.003>
- Kang, N. E., Ha, A. W., Woo, H. W., and Kim, W. K. (2014). Peanut sprouts extract (*Arachis hypogaea* L.) has anti-obesity effects by controlling the protein expressions of PPAR $\gamma$  and adiponectin of adipose tissue in rats fed high-fat diet. *Nutrition Research and Practice*, 8(2), 158-164.
- Karahashi, M., Ishii, F., Yamazaki, T., Imai, K., Mitsumoto, A., Kawashima, Y., and Kudo, N. (2013). Up-regulation of stearoyl-CoA desaturase 1 increases liver MUFA content in obese Zucker but not Goto-Kakizaki rats. *Lipids*, 48(5), 457-467.
- Kaur, C., and Kapoor, H. C. (2001). Antioxidants in fruits and vegetables—the millennium's health. *International Journal of Food Science and Technology*, 36(7), 703-725.
- Kazemipoor, M., Cordell, G. A., Sarker, M. M. R., Radzi, C. W. J. B. W. M., Hajifaraji, M., and En Kiat, P. (2015). Alternative treatments for weight loss: safety/risks and effectiveness of anti-obesity medicinal plants. *International Journal of Food Properties*, 18(9), 1942-1963.
- Keaney, J. F., Larson, M. G., Vasan, R. S., Wilson, P. W. F., Lipinska, I., Corey, D., and Benjamin, E. J. (2003). Obesity and systemic oxidative stress clinical correlates of oxidative stress in the Framingham study. *Arteriosclerosis, Thrombosis and Vascular Biology*, 23(3), 434-439.
- Kelava, T., Čavar, I., and Čulo, F. (2011). Biological actions of drug solvents. *Periodicum Biologorum*, 113(3), 311-320.
- Kershaw, E. E., and Flier, J. S. (2004). Adipose tissue as an endocrine organ. *The Journal of Clinical Endocrinology and Metabolism*, 89(6), 2548-2556.
- Kersten, S. (2001). Mechanisms of nutritional and hormonal regulation of lipogenesis. *EMBO Reports*, 2(4), 282-286.
- Kersten, S., Desvergne, B., and Wahli, W. (2000). Roles of PPARs in health and disease. *Nature*, 405(6785), 421-424.
- Khan, S., Khan, A., Khattak, F. S., and Naseem, A. (2012). An accurate and cost effective approach to blood cell count. *International Journal of Computer Applications*, 50(1), 18-24.
- Khanam, U. K. S., Oba, S., Yanase, E., and Murakami, Y. (2012). Phenolic acids, flavonoids and total antioxidant capacity of selected leafy vegetables. *Journal of Functional Foods*, 4(4), 979-987.

- Khoo, L., Foong Kow, A., Maulidiani, M., Lee, M., Tan, C., Shaari, K., and Abas, F. (2018). Haematological, biochemical, histopathological and 1H-NMR metabolomics application in acute toxicity evaluation of *Clinacanthus nutans* water leaf extract. *Molecules*, 23(9), 2172; <https://doi.org/10.3390/molecules23092172>.
- Kim, D. O., Lee, K. W., Lee, H. J., and Lee, C. Y. (2002). Vitamin C equivalent antioxidant capacity (VCEAC) of phenolic phytochemicals. *Journal of Agricultural and Food Chemistry*, 50(13), 3713-3717.
- Kim, S., Shin, H. J., Kim, S. Y., Kim, J. H., Lee, Y. S., Kim, D. H., and Lee, M. O. (2004). Genistein enhances expression of genes involved in fatty acid catabolism through activation of PPAR $\alpha$ . *Molecular and Cellular Endocrinology*, 220(1), 51-58.
- Kim, Y. J., and Son, D. Y. (2011). Antioxidant effects of solvent extracts from the dried jujube (*Zizyphus jujube*) sarcocarp, seed, and leaf via sonication. *Food Science and Biotechnology*, 20(1), 167-173.
- Kitada, H., Miyata, M., Nakamura, T., Tozawa, A., Honma, W., Shimada, M., and Yamazoe, Y. (2003). Protective role of hydroxysteroid sulfotransferase in lithocholic acid-induced liver toxicity. *Journal of Biological Chemistry*, 278(20), 17838-17844.
- Kn, B. P., Gopalan, V., Lee, S. S., and Velan, S. S. (2014). Quantification of abdominal fat depots in rats and mice during obesity and weight loss interventions. *Plos One*, 9(10), e108979.
- Kohnke, R., Lindqvist, A., Goransson, N., Emek, S. C., Albertsson, P., Rehfeld, J. F., and Erlanson-Albertsson, C. (2009). Thylakoids suppress appetite by increasing cholecystokinin resulting in lower food intake and body weight in high-fat fed mice. *Phytotherapy Research*, 23(12), 1778-1783.
- Kong, H. S., and Abdullah Sani, N. (2017). Nutritional values and amino acid profiles of *Clinacanthus nutans* (Belalai Gajah/Sabah Snake Grass) from 2 farms in Negeri Sembilan, Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 40(4).
- Kono, Y., Kobayashi, K., Tagawa, S., Adachi, K., Ueda, A., Sawa, Y., and Shibata, H. (1997). Antioxidant activity of polyphenolics in diets: rate constants of reactions of chlorogenic acid and caffeic acid with reactive species of oxygen and nitrogen. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1335(3), 335-342.
- Kopaei, M.R., Setorki, M., Doudi, M., Baradaran, A., and Nasri, H. (2014). Atherosclerosis: process, indicators, risk factors and new hopes. *International Journal of Preventive Medicine*, 5(8), 927-946.
- Kopelman, P. G. (2000). Obesity as a medical problem. *Nature*, 404(6778), 635-643.

- Kraemer, F. B., and Shen, W. J. (2002). Hormone-sensitive lipase control of intracellular tri-(di-) acylglycerol and cholesteryl ester hydrolysis. *Journal of Lipid Research*, 43(10), 1585-1594.
- Kraemer, F. B., Patel, S., Saedi, M. S., and Sztalryd, C. (1993). Detection of hormone-sensitive lipase in various tissues. I. Expression of an HSL/bacterial fusion protein and generation of anti-HSL antibodies. *Journal of Lipid Research*, 34(4), 663-671.
- Kranendonk, M.E., Herwaarden, J.A., Stupkova, T., Jager, W.D., Vink, A., Moll, F.L., Kalkhoven, E., and Visseren F.L.J. (2015). Inflammatory characteristics of distinct abdominal adipose tissue depots relate differently to metabolic risk factors for cardiovascular disease. Distinct fat depots and vascular risk factors. *Atherosclerosis*, 239(2), 419-427.
- Krishnamoorthy, D., Frechette, D. M., Adler, B. J., Green, D. E., Chan, M. E., and Rubin, C. T. (2015). Marrow adipogenesis and bone loss that parallels estrogen deficiency is slowed by low-intensity mechanical signals. *Osteoporosis International*, 27(2), 747-756.
- Kubota, K., Sumi, S., Tojo, H., and Sumi-Inoue, Y. (2011). Improvements of mean body mass index and body weight in pre-obese and overweight Japanese adults with black Chinese tea (Pu-Erh) water extract. *Nutritional Research*, 31(6), 421-428
- Kucukbay, F., and Kuyumcu, E. (2010). Determination of trace element contents of Thymus species from Turkey. *Turkish Journal of Chemistry*, 34(6), 911-920.
- Kulkarni, Y. A., and Veeranjaneyulu, A. (2012). Toxicological evaluation of the methanol extract of *Gmelina arborea* Roxb bark in mice and rats. *Toxicology International*, 19(2), 125-131.
- Kumar, S., Alagawadi, K. R., and Rao, M. R. (2011). Effect of *Argyreia speciosa* root extract on cafeteria diet-induced obesity in rats. *Indian Journal of Pharmacology*, 43(2), 163-167.
- Kuppusamy, U. R., and Das, N. P. (1994). Potentiation of  $\beta$ -adrenoceptor agonist-mediated lipolysis by quercetin and fisetin in isolated rat adipocytes. *Biochemical Pharmacology*, 47(3), 521-529.
- Lago, F., Gomez, R., Gomez-Reino, J. J., Dieguez, C., and Gualillo, O. (2009). Adipokines as novel modulators of lipid metabolism. *Trends in Biochemical Sciences*, 34(10), 500–510
- Lai, N., Sims, J. K., Jeon, N. L., and Lee, K. (2012). Adipocyte induction of preadipocyte differentiation in a gradient chamber. *Tissue Engineering Part C: Methods*, 18(12), 958-967.

- Lakmichi, H., Bakhtaoui, F. Z., Gadhi, C. A., Ezoubeiri, A., El Jahiri, Y., El Mansouri, A., and Loutfi, K. (2011). Toxicity profile of the aqueous ethanol root extract of *Corrigiola telephifolia pour* (Caryophyllaceae) in rodents. *Evidence-Based Complementary and Alternative Medicine*, 2011.
- Lalisan, A., Nuñez Olga, M., and Uy Mylene, M. (2014). Brine shrimp (*Artemia salina*) bioassay of the medicinal plant *Pseudelephantopus spicatus* from Iligan City, Philippines.
- Lang, A., and Froelicher, E. S. (2006). Management of overweight and obesity in adults: behavioral intervention for long-term weight loss and maintenance. *European Journal of Cardiovascular Nursing*, 5(2), 102-114.
- Latha, R. C. R., and Daisy, P. (2011). Insulin-secretagogue, antihyperlipidemic and other protective effects of gallic acid isolated from *Terminalia bellerica* Roxb, in streptozotocin-induced diabetic rats. *Chemico-Biological Interactions*, 189(1-2), 112-118.
- Lau, K. W., Lee, S. K., and Chin, J. H. (2014). Effect of the methanol leaves extract of *Clinacanthus nutans* on the activity of acetylcholinesterase in male mice. *Journal of Acute Disease*, 3(1), 22-25.
- Leathem, J. H. (1964). Some aspects of hormone and protein metabolic interrelationships. In *Mammalian Protein Metabolism*. Academic Press, pp. 343-380.
- Lecumberri, E., Goya, L., Mateos, R., Alia, M., Ramos, S., Izquierdo-Pulido, M., and Bravo, L. (2007). A diet rich in dietary fiber from cocoa improves lipid profile and reduces malondialdehyde in hypercholesterolemic rats. *Nutrition*, 23(4), 332-341.
- Lee, M. S., Kim, C. T., Kim, I. H., and Kim, Y. (2009a). Inhibitory effects of green tea catechin on the lipid accumulation in 3T3-L1 adipocytes. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 23(8), 1088-1091.
- Lee, M. S., Kim, C.T., and Kim, Y. (2009b). Green tea (-)-epigallocatechin-3-gallate reduces body weight with regulation of multiple genes expression in adipose tissue of diet-induced obese mice. *Annals of Nutrition and Metabolism*, 54: 151–157.
- Lee, D.R., Lee, Y.S., Choi, B.K., Lee, H.J., Park, S.B., Kim, T.M., Oh, H.J., Yang, S.H., and Suh, J.W. (2015a). Roots extracts of *Adenophora triphylla* var. japonica improve obesity in 3T3-L1 adipocytes and high-fat diet-induced obese mice. *Asian Pacific Journal of Tropical Medicine*, 8(11), 898-906.
- Lee, J., Han, S. I., Yun, J. H., and Kim, J. H. (2015b). Quercetin 3-O-glucoside suppresses epidermal growth factor-induced migration by inhibiting EGFR signaling in pancreatic cancer cells. *Tumor Biology*, 36(12), 9385-9393.

- Lende, A. B., Kshirsagar, A. D., Deshpande, A. D., Muley, M. M., Patil, R. R., Bafna, P. A., and Naik, S. R. (2011). Anti-inflammatory and analgesic activity of protocatechuic acid in rats and mice. *Inflammopharmacology*, 19(5), 255.
- Lenon, G. B., Chang, Y. H., Yang, A. W., Da Costa, C., Li, C. G., and Xue, C. C. (2012). Efficacy and safety of a Chinese herbal medicine formula (RCM-104) in the management of simple obesity: a randomized, placebo-controlled clinical trial. *Evidence-Based Complementary and Alternative Medicine*, 2012.
- Levin, B. E., Dunn-Meynell, A. A., and Routh, V. H. (1999). Brain glucose sensing and body energy homeostasis: role in obesity and diabetes. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 276(5), R1223–R1231.
- Li, Z., Yang, L., Wang, J., Shi, W., Pawar, R.A., Liu, Y., Xu, C., Cong, W., Hua, Q., Lu, T., Xia, F., Guo, W., Zhao, M., and Zhang, Y. (2010). B-actin is a useful internal control for tissue-specific gene expression studies using quantitative real-time PCR in the half-smooth tongue sole *Cynoglossus semilaevis* challenged with LPS or *Vibrio anguillarum*. *Fish and Shellfish Immunology*, 29(1), 89-93.
- Liang, X., and Fang, W. S. (Eds.). (2006). *Medicinal Chemistry of Bioactive Natural Products* (p. 480). New Jersey: Wiley.
- Lin, H. Z., Yang, S. Q., Chuckaree, C., Kuhajda, F., Ronnet, G., and Diehl, A. M. (2000). Metformin reverses fatty liver disease in obese, leptin-deficient mice. *Nature Medicine*, 6(9), 998-1003.
- Lin, L., and Wong, H. (2017). Predicting oral drug absorption: mini review on physiologically-based pharmacokinetic models. *Pharmaceutics*, 9(4), 41.
- Liu, S. F., and Malik, A. B. (2006). NF-κB activation as a pathological mechanism of septic shock and inflammation. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 290(4), 622-645.
- Lobo, V., Patil, A., Phatak, A., and Chandra, N. (2010). Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacognosy Reviews*, 4(8), 118.
- Lohr, J. W., Willsky, L. R., and Acara, M. A. (1998). Renal drug metabolism. *Pharmacological Reviews*, 50(1), 107-142.
- Lonnqvist, F., Nordfors, L., Jansson, M., Thörne, A., Schalling, M., and Arner, P. (1997). Leptin secretion from adipose tissue in women. Relationship to plasma levels and gene expression. *Journal of Clinical Investigation*, 99(10), 2398-2404.
- Łozak, A., Sołyk, K., Ostapczuk, P., and Fijałek, Z. (2002). Determination of selected trace elements in herbs and their infusions. *Science of the Total Environment*, 289(1-3), 33-40.

- Lumeng, C.N. and Saltiel, A.R. (2011). Inflammatory links between obesity and metabolic disease. *Journal of Clinical Investigations*, 121(6), 2111-2117.
- Lustig, R. H., Mietus-Snyder, M. L., Bacchetti, P., Lazar, A. A., Velasquez-Meyer, P. A. and Christensen, M. L. (2006). Insulin dynamics predict body mass index and z-score response to insulin suppression or sensitization pharmacotherapy in obese children. *The Journal of Pediatrics*, 148(1), 23-29.
- Lynch, S. M., and Frei, B. (1993). Mechanisms of copper-and iron-dependent oxidative modification of human low density lipoprotein. *Journal of Lipid Research*, 34(10), 1745-1753.
- Macedo, L. F. L., Rogero, M. M., Guimaraes, J. P., Granato, D., Lobato, L. P., and Castro, I. A. (2013). Effect of red wines with different *in vitro* antioxidant activity on oxidative stress of high-fat diet rats. *Food Chemistry*, 137(1), 122-129.
- Magnusson, Y. K., Friberg, P., Sjavall, P., Malm, J., and Chen, Y. (2008). TOF-SIMS analysis of lipid accumulation in the skeletal muscle of ob/ob mice. *Obesity*, 16(12), 2745-2753.
- Mai, C. W., Yap, K. S., Kho, M. T., Ismail, N. H., Yusoff, K., Shaari, K., and Lim, E. S. (2016). Mechanisms underlying the anti-inflammatory effects of *Clinacanthus nutans* Lindau extracts: inhibition of cytokine production and toll-like receptor-4 activation. *Frontiers in Pharmacology*, 7, 7.
- Makki, K., Froguel, P., and Wolowczuk, I. (2013). Adipose tissue in obesity-related inflammation and insulin resistance: cells, cytokines, and chemokines. *International Scholarly Research Network Inflammation*, 2013, 1-12 <http://dx.doi.org/10.1155/2013/139239>.
- Malaguarnera, G., Cataudella, E., Giordano, M., Nunnari, G., Chisari, G., and Malaguarnera, M. (2012). Toxic hepatitis in occupational exposure to solvents. *World Journal of Gastroenterology*, 18(22), 2756.
- Mancuso, G., Midiri, A., Beninati, C., Piraino, G., Valenti, A., Nicocia, G., and Teti, G. (2002). Mitogen-activated protein kinases and NF- $\kappa$ B are involved in TNF- $\alpha$  responses to group B streptococci. *The Journal of Immunology*, 169(3), 1401-1409.
- Martignoni, M., Groothuis, G. M., and de Kanter, R. (2006). Species differences between mouse, rat, dog, monkey and human CYP-mediated drug metabolism, inhibition and induction. *Expert Opinion on Drug Metabolism and Toxicology*, 2(6), 875-894.
- Martin, D. A., and Bolling, B. W. (2015). A review of the efficacy of dietary polyphenols in experimental models of inflammatory bowel diseases. *Food and Function*, 6(6), 1773-1786.

- Martins, F., Noso, T. M., Porto, V. B., Curiel, A., Gambero, A., Bastos, D. H., and Carvalho, P. D. O. (2010). Maté tea inhibits in vitro pancreatic lipase activity and has hypolipidemic effect on high-fat diet-induced obese mice. *Obesity*, 18(1), 42-47.
- Marquez-Martin, A., De La Puerta, R., Fernandez-Arche, A., Ruiz-Gutierrez, V., and Yaqoob, P. (2006). Modulation of cytokine secretion by pentacyclic triterpenes from olive pomace oil in human mononuclear cells. *Cytokine*, 36(5), 211–217.
- Mashmoul, M., Azlan, A., Yusof, B. N. M., Khaza'ai, H., Mohtarrudin, N., and Boroushaki, M. T. (2014). Effects of saffron extract and crocin on anthropometrical, nutritional and lipid profile parameters of rats fed a high-fat diet. *Journal of Functional Foods*, 8, 180-187.
- Mayer, J. (1952). The glucostatic theory of regulation of food intake and the problem of obesity. *Bulletin. New England Medical Center*, 14(2), 43.
- Mayoral, R., Osborn, O., McNelis, J., Johnson, A. M., Izquierdo, C. L., Chung, H., and Ofrecio, J. M. (2015). Adipocyte SIRT1 knockout promotes PPAR $\gamma$  activity, adipogenesis and insulin sensitivity in chronic-HFD and obesity. *Molecular Metabolism*, 4(5), 378-391.
- McDonald, S., Prenzler, P. D., Antolovich, M., and Robards, K. (2001). Phenolic content and antioxidant activity of olive extracts. *Food Chemistry*, 73(1), 73-84.
- Mehenni, C., Atmani-Kilani, D., Dumarçay, S., Perrin, D., Gérardin, P., and Atmani, D. (2016). Hepatoprotective and anti-diabetic effects of *Pistacia lentiscus* leaf and fruit extracts. *Journal of Food and Drug Analysis*, 24(3), 653-669.
- Mensah, J. K., Okoli, R. I., Ohaju-Obodo, J. O., and Eifediyi, K. (2008). Phytochemical, nutritional and medical properties of some leafy vegetables consumed by Edo people of Nigeria. *African Journal of Biotechnology*, 7(14).
- Mensah, J. K., Okoli, R.I., Turay, A.A., and Ogie-Odia, E.A. (2009). Phytochemical analysis of medicinal plants used for the management of hypertension by Esan people of Edo state, Nigeria. *Ethnobotanical Leaflets*, 13, 1273–1287.
- Mercier, Y., Gatellier, P., Viau, M., Remignon, H., and Renerre, M. (1998). Effect of dietary fat and vitamin E on color stability and on lipid and protein oxidation in Turkey meat during storage. *Meat Science*, 48(3-4), 301-318.
- Mermel, V. L. (2004). Old paths new directions: the use of functional foods in the treatment of obesity. *Trends in Food Science and Technology*, 15(11), 532–540.
- Meydani, M., and Hasan, S. T. (2010). Dietary polyphenols and obesity. *Nutrients*, 2(7), 737–751.

- Mezei, O., Li, Y., Mullen, E., Ross-Viola, J. S., and Shay, N. F. (2006). Dietary isoflavone supplementation modulates lipid metabolism via PPARalpha-dependent and-independent mechanisms. *Physiological Genomics*, 26(1), 8-14.
- Mielnik, M. B., Aaby, K., and Skrede, G. (2003). Commercial antioxidants control lipid oxidation in mechanically deboned turkey meat. *Meat Science*, 65(3), 1147-1155.
- Minokoshi, Y., Kim, Y. B., Peroni, O. D., Fryer, L. G., Müller, C., Carling, D., and Kahn, B. B. (2002). Leptin stimulates fatty-acid oxidation by activating AMP-activated protein kinase. *Nature*, 415(6869), 339-343.
- Misawa, K., Horiba, T., Arimura, N., Hirano, Y., Inoue, J., Emoto, N., and Sato, R. (2003). Sterol regulatory element-binding protein-2 interacts with hepatocyte nuclear factor-4 to enhance sterol isomerase gene expression in hepatocytes. *Journal of Biological Chemistry*, 278(38), 36176-36182.
- Miyazaki, M., Kim, Y. C., Gray-Keller, M. P., Attie, A. D., and Ntambi, J. M. (2000). The biosynthesis of hepatic cholesterol esters and triglycerides is impaired in mice with a disruption of the gene for stearoyl-CoA desaturase 1. *Journal of Biological Chemistry*, 275(39), 30132-30138.
- Miyazaki, M., Kim, Y.C., and Ntambi, J. M. (2001). A lipogenic diet in mice with a disruption of the stearoyl-CoA desaturase 1 gene reveals a stringent requirement of endogenous monounsaturated fatty acids for triglyceride synthesis. *Journal of Lipid Research*, 42(7), 1018-1024.
- Miyazaki, Y., Pipek, R., Mandarino, L.J. and DeFronzo, R.A. (2003). Tumor necrosis factor  $\alpha$  and insulin resistance in obese type 2 diabetic patients. *International Journal of Obesity and Related Metabolic Disorder*, 27(1), 88-94.
- Mohamed, G. A., Ibrahim, S. R., Elkhayat, E. S., and El Dine, R. S. (2014). Natural anti-obesity agents. *Bulletin of Faculty of Pharmacy, Cairo University*, 52(2), 269-284.
- Mohd-Radzman, N. B., Ismail, W.I.W., Adam, Z., Jaapar, S.S. and Adam A. (2013). Potential roles of *Stevia rebaudiana* bertoni in abrogating insulin resistance and diabetes: A review. *Evidence-Based Complementary Medicine*, 718049, 1-10.
- Montelius, C., Erlandsson, D., Vitija, E., Stenblom, E., Egecioglu, E., and Erlanson-Albertsson, C. (2014). Body weight loss, reduced urge for palatable food and increased release of GLP-1 through daily supplementation with green-plant membranes for 3 months in overweight women. *Appetite*, 81, 295-304.
- Morris, D., and Rui, L. (2009). Recent advances in understanding leptin signaling and leptin resistance. *American Journal of Physiology-Endocrinology and Metabolism*, 297(6), 1247- 1259.

- Morrison, M., van der Heijden, R., Heeringa, P., Kaijzel, E., Verschuren, L., Blomhoff, R., and Kleemann, R. (2014). Epicatechin attenuates atherosclerosis and exerts anti-inflammatory effects on diet-induced human-CRP and NF B in vivo. *Atherosclerosis*, 233(1), 149-156.
- Morselli, E., Mariño, G., Bennetzen, M.V., Eisenberg, T., Megalou, E., and Schroeder, S. (2011). Spermidine and resveratrol induce autophagy by distinct pathways converging on the acetylproteome. *Journal of Cell Biology*, 192(4), 615-629.
- Moses, L. B., Aziz, Z. A., Mamat, H., and Bakar, M. F. A. (2015). Nutritional composition and trace elements contents of unfermented and fermented *Clinacanthus nutans* L. herbal tea. *Journal of Tropical Resources Sustain. Sciences*, 3, 16-29.
- Moseti, D., Regassa, A., and Kin, W.K. (2016). Molecular regulation of adipogenesis and potential anti-adipogenic bioactive molecules. *International Journal of Molecular Science*, 17(124), 1-24.
- Moyo, B., Oyedemi, S., Masika, P. J., and Muchenje, V. (2012). Polyphenolic content and antioxidant properties of *Moringa oleifera* leaf extracts and enzymatic activity of liver from goats supplemented with *Moringa oleifera* leaves/sunflower seed cake. *Meat Science*, 91(4), 441-447.
- Muls, E., Kolanowski, J., and Scheen, A. (2001). The effects of orlistat on weight and on serum lipids in obese patients with hypercholesterolemia: a randomized, double-blind, placebo-controlled, multicentre study. *International Journal of Obesity*, 25(11), 1713.
- Mulvihill, E.E. and Huff, M.W. (2010). Antiartherogenic properties of flavonoids: implication Must, A., Spadano, J., Coakley, E. H., Field, A. E., Colditz, G., and Dietz, W. H. (1999). The disease burden associated with overweight and obesity. *Jama*, 282(16), 1523-1529.
- Mustapa, A. N., Martin, A., Mato, R. B., and Cocero, M. J. (2015). Extraction of phytocompounds from the medicinal plant *Clinacanthus nutans* Lindau by microwave-assisted extraction and supercritical carbon dioxide extraction. *Industrial Crops and Products*, 74, 83-94.
- Myhre, M. J. (2000). Herbal remedies, nephropathies, and renal disease. *Nephrology Nursing Journal*, 27(5), 473.
- Nakai, M., Fukui, Y., Asami, S., Toyoda-Ono, Y., Iwashita, T., Shibata, H., and Kiso, Y. (2005). Inhibitory effects of oolong tea polyphenols on pancreatic lipase in vitro. *Journal of Agricultural and Food Chemistry*, 53(11), 4593-4598.
- Nakazato, K., Song, H., and Waga, T. (2006). Effects of dietary apple polyphenol on adipose tissues weights in Wistar rats. *Experimental Animals*, 55(4), 383-389.
- Narayanaswamy, R., and Ismail, I. S. (2015). Cosmetic potential of Southeast Asian herbs: an overview. *Phytochemistry Reviews*, 14(3), 419-428.

- Nasution, R., and Marzuki, I. (2016). Isolation Compound Anti-obesity from the Bark Ara (*Ficus Racemosa*) of Aceh. *Oriental Journal of Chemistry*, 32(5), 2693-2699.
- Newman, D. J., and Cragg, G. M. (2012). Natural products as sources of new drugs over the 30 years from 1981 to 2010. *Journal of Natural Products*, 75(3), 311-335.
- Ng, M., Fleming, T., Robinson, M., Thomson, B., Graetz, N., Margono, C., and Abraham, J. P. (2014). Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: a systematic analysis for the global burden of disease study 2013. *The lancet*, 384(9945), 766-781.
- Ngondi, J. L., Etoundi, B. C., Nyamgono, C.B., Mbofung, C. M.., and Oben, J.E. (2009). IGOB131, a novel seed extract of the West African plant *irvingia gabonensis* significantly reduces body weight and improves metabolic parameters in overweight humans in a randomized double-blind placebo controlled investigation. *Lipids Health and Disease*, 8, 1-7.
- Nichenametla, S. N., Taruscio, T. G., Barney, D. L., and Exon, J. H. (2006). A review of the effects and mechanisms of polyphenolics in cancer. *Critical Reviews in Food Science and Nutrition*, 46(2), 161-183.
- Nicolas-Vazquez, I., Torres, J. H., Borbolla, J. C., Ruvalcaba, R. M., and Aceves-Hernández, J. M. (2014). Orlistat interaction with sibutramine and carnitine. A physicochemical and theoretical study. *Journal of Molecular Structure*, 1062, 1-12. <https://doi.org/10.1016/j.molstruc.2013.12.072>
- Nimse, S. B., and Pal, D. (2015). Free radicals, natural antioxidants, and their reaction mechanisms. *RSC Adv.*, 5(35), 27986–28006. <https://doi.org/10.1039/C4RA13315C>.
- Nishikawa, S., Yasoshima, A., Doi, K., Nakayama, H., and Uetsuka, K. (2007). Involvement of sex, strain and age factors in high-fat diet-induced obesity in C57BL/6J and BALB/cA mice. *Experimental Animals*, 56(4), 263-272.
- Nisoli, E., and Carruba, M. O. (2000). An assessment of the safety and efficacy of sibutramine, an anti-obesity drug with a novel mechanism of action. *Obesity Reviews*, 1(2), 127-139.
- Nobili, S., Lippi, D., Witort, E., Donnini, M. and Bausi, L. (2009). Natural compounds for cancer treatment and prevention. *Pharmacological Research*, 59(6), 365-378.
- Nohturfft, A., Brown, M. S., and Goldstein, J. L. (1998). Sterols regulate processing of carbohydrate chains of wild-type SREBP cleavage-activating protein (SCAP), but not sterol-resistant mutants Y298C or D443N. *Proceedings of the National Academy of Sciences*, 95(22), 12848-12853.

- Norata, G. D., Pirillo, A., and Catapano, A. (2006). Modified HDL: biological and physiopathological consequences. *Nutrition, Metabolism and Cardiovascular Diseases*, 16(5), 371-386.
- Ntambi, J. M., and Miyazaki, M. (2004). Regulation of stearoyl-CoA desaturases and role in metabolism. *Progress in Lipid Research*, 43(2), 91-104.
- Ntambi, J. M., Miyazaki, M., Stoehr, J.P., Lan, H., Kendziorski, C.M., and Yandell, B.S. (2002). Loss of stearoyl-CoA desaturase-1 function protects mice against adiposity. *Proceedings of the National Academy of Sciences of the United States of America*, 99(17), 11482-11486.
- Nurul, S. A. S., Hazilawati, H., Mohd, R. S., Mohd, F. H. R., Noordin, M. M., and Norhaizan, M. E. (2018). Subacute oral toxicity assessment of ethanol extract of *mariposa christia vespertilionis* leaves in male Sprague Dawley rats. *Toxicological Research*, 34(2), 85-95.
- Nworgu, Z.A.M., Onwukaeme, D.N., Afolayan A.J., Ameachine, F.C., and Ayinde, B.A. (2008). Preliminary studies of blood pressure lowering effect of *Nauclea latifolia* in rats. *African Journal of Pharmacy and Pharmacology*, 2(2), 37-41.
- The Organization of Economic Co-operation and Development (OECD)(2008). *Guideline for the Testing of Chemicals, Repeated Dose 28-Day: Oral Toxicity Study in Rodent*. Paris: OECD. No.407. Retrieved October 3, 2008 from <https://ntp.niehs.nih.gov/iccvam/suppdocs/feddocs/oecd/oecdtg407-2008>
- Odchimar, N. M. O., Nuñez, O. M., Uy, M. M., and Senarath, W. T. P. S. K. (2016). Antioxidant activity, total phenolic content, and GC-MS analysis of the root of Kawilan (*Embeliaphilippinensis* A. DC.). *Bulletin of Environment, Pharmacology and Life Sciences*, 5(5), 42-47.
- Ohta, Y., Sami, M., Kanda, T., Saito, K., Osada, K., and Kato, H. (2006). Gene expression analysis of the anti-obesity effect by apple polyphenols in rats fed a high-fat diet or a normal diet. *Journal of Oleo Science*, 55(6), 305-314.
- Okigbo, R. N., Eme, U. E., and Ogbogu, S. (2008). Biodiversity and conservation of medicinal and aromatic plants in Africa. *Biotechnology and Molecular Biology Review* 3(6), 127-134.
- Okokon, J. E., Nwafor, P. A., and Ekpo, M. D. (2010). Subchronic toxicity studies of the ethanolic root extract of *Croton zambesicus*. *Pakistan Journal of Pharmaceutical Sciences*, 23(2), 160-169.
- Ondrejovicova, I., Muchova, J., Mislanova, C., Nagyova, Z., and Durackova, Z. (2010). Hypercholesterolemia, oxidative stress and gender dependence in children. *Prague Medical Report*, 111(4), 300-312.
- Ooi, D. J., Iqbal, S., and Ismail, M. (2012). Proximate composition, nutritional attributes and mineral composition of *Peperomia pellucida L* (Ketumpangan Air) grown in Malaysia. *Molecules*, 17(9), 11139-11145.

- Osuga, J., Ishibashi, S., Oka, T., Yagyu, H., Tozawa, R., Fujimoto, A., and Tsutsumi, O. (2000). Targeted disruption of hormone-sensitive lipase results in male sterility and adipocyte hypertrophy, but not in obesity. *Proceedings of the National Academy of Sciences*, 97(2), 787-792.
- Ostapowicz, G., Fontana R.J., Schiodt F.V., Larson A., Davron J.T., Steven H.B., Timothy M., Reish J. (2002). Results of a prospective study of acute liver failure at 17 tertiary care centers in the United States. *Annal Internet Medicine*, 137(12), 947–954.
- Otieno, D. (2016). *Effects of Beetroot Extract in Brown Adipose Tissue from Diet-induced Obese Mice*. PhD Thesis. North Carolina Agricultural and Technical State University, USA.
- Ouchi, N., Ohashii, K., Shibata, R. and Murohara, T. (2012). Adipocytokines and obesity-linked disorders. *Nagoya Journal of Medicine*, 74(1-2), 19-30.
- Ozer, J., Ratner, M., Shaw, M., Bailey, W., and Schomaker, S. (2008). The current state of serum biomarkers of hepatotoxicity. *Toxicology*, 245(3), 194-205.
- Padwal, R.S., and Majumdar, S.R. (2007). Drug treatments for obesity: orlistat, sibutramine, and rimonabant. *Lancet*, 369(9555), 71-77.
- Pandey, D., Suneetha, E., Senussi, F., Al-Fakhry, R., and Al-Agouri, W. (2016). Diet quality assessment of Benghazi medical students using the healthy eating index. In Salam, A., Pandey, D. and Alshekteria, A. (Eds.). *Public Health Research in Libya Innovations and Methodologies* (pp.159-204). Deutschland, Germany: Scholar's Press.
- Pandit, A., Sachdeva, T., and Bafna, P. (2012). Drug-induced hepatotoxicity: a review. *Journal of Applied Pharmacology and Sciences* 2(5), 233-243.
- Pang, J., Choi, Y., and Park, T. (2008). *Ilex paraguariensis* extract ameliorates obesity induced by high-fat diet: potential role of AMPK in the visceral adipose tissue. *Archives of Biochemistry and Biophysics*, 476(2), 178-185.
- Panmei, C., Singh, P.K., Gautam, S., Variyar, P.S., Shantibala Devi, G.A., and Sharma, A. (2007). Phenolic acids in Albizia bark used as a starter for rice fermentation in Zou preparation. *Journal of Food, Agricultural and Environment*, 5(3), 147-150.
- Park, H. J., DiNatale, D. A., Chung, M.-Y., Park, Y.-K., Lee, J.-Y., Koo, S. I., and Bruno, R. S. (2011). Green tea extract attenuates hepatic steatosis by decreasing adipose lipogenesis and enhancing hepatic antioxidant defenses in ob/ob mice. *The Journal of Nutritional Biochemistry*, 22: 393–400
- Partap, U., Young, E. H., Allotey, P., Sandhu, M. S., and Reidpath, D. D. (2017). The use of different international references to assess child anthropometric status in a Malaysian population. *The Journal of Pediatrics*, 190, 63-68.

- Parola, M. and Marra, F. (2011). Adipokines and redox signaling: Impact on fatty liver disease. *Antioxidants and Redox Signalling*, 15(2), 461-483.
- Patterson, H. B. W. (1989). *Handling and storage of oilseeds, oils, fats and meal* (No. 633.85 P38 1989.). London: Elsevier Applied Science.
- Paur, I., Balstad, T., Kolberg, M., Pedersen, M., Austenaa, L., Jacobs, D., and Blomhoff, R (2010). Extract of oregano, coffee, thyme, clove, and walnuts inhibits NF-kappa B in monocytes and in transgenic reporter mice. *Cancer Prevention Research*. 3(5), 653–63.
- Perrone, R. D., Madias, N. E., and Levey, A. S. (1992). Serum creatinine as an index of renal function: new insights into old concepts. *Clinical Chemistry*, 38(10), 1933-1953.
- Pfortmueller, C. A., Uehlinger, D., von Haehling, S., and Schefold, J. C. (2018). Serum chloride levels in critical illness-the hidden story. *Intensive Care Medicine Experimental*, 6(10), 1-14.
- Phua, Q. Y., Subramaniam, S., Lim, V., and Chew, B. L. (2018). The establishment of cell suspension culture of Sabah snake grass (*Clinacanthus nutans* (Burm. F.) Lindau). *In Vitro Cellular and Developmental Biology-Plant*, 54(4), 413-422.
- Pierleoni, C., Verdenelli, F., Castellucci, M., and Cinti, S. (1998). Fibronectins and basal lamina molecules expression in human subcutaneous white adipose tissue. *European Journal of Histochemistry*, 42(3), 183-188.
- Piersma, T., Koolhaas, A., Dekkinga, A., and Gwinner, E. (2000). Red blood cell and white blood cell counts in sandpipers (*Philomachus pugnax*, *Calidris canutus*): effects of captivity, season, nutritional status, and frequent bleedings. *Canadian Journal of Zoology*, 78(8), 1349-1355.
- Pinent, M., Blade, M. C., Salvadó, M. J., Arola, L., and Ardévol, A. (2005). Intracellular mediators of procyanidin-induced lipolysis in 3T3-L1 adipocytes. *Journal of Agricultural and Food Chemistry*, 53(2), 262-266.
- Piotrowska, K., Tarnowski, M., Zgutka, K., and Pawlik, A. (2016). Gender differences in response to prolonged every-other-day feeding on the proliferation and apoptosis of hepatocytes in mice. *Nutrients*, 8(3), 176.
- P'ng, X. W., Akowuah, G. A., and Chin, J. H. (2012). Acute oral toxicity study of *Clinacanthus nutans* in mice. *International Journal of Pharmaceutical Sciences and Research*, 3(11), 4202-4205.
- P'ng, X. W., Akowuah, G. A., and Chin, J. H. (2013). Evaluation of the sub-acute oral toxic effect of methanol extract of *Clinacanthus nutans* leaves in rats. *Journal of Acute Disease*, 2(1), 29-32.

- Popeijus, H. E., Saris, W. H., and Mensink, R. P. (2008). Role of stearoyl-CoA desaturases in obesity and the metabolic syndrome. *International Journal of Obesity*, 32(7), 1076–1082.
- Popkin, B. M., Adair, L. S., and Ng, S. W. (2012). Global nutrition transition and the pandemic of obesity in developing countries. *Nutrition Reviews*, 70(1), 3-21.
- Poston, W. S. C., and Foreyt, J. P. (1999). Obesity is an environmental issue. *Atherosclerosis*, 146(2), 201-209.
- Pounis, G., Bonaccio, M., Di Castelnuovo, A., Costanzo, S., De Curtis, A., Persichillo, M., and Iacoviello, L. (2016). Polyphenol intake is associated with low-grade inflammation, using a novel data analysis from the Moli-sani study. *Thrombosis and Haemostasis*, 116(02), 344-352. <https://doi.org/10.1160/TH15-06-0487>.
- Pradhan, A. D., Manson, J. E., Rifai, N., Buring, J. E., and Ridker, P. M. (2001). C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. *Jama*, 286(3), 327-334.
- Pratoomsoot, C., Sruamsiri, R., Dilokthornsakul, P., and Chaiyakunapruk, N. (2015). Quality of reporting of randomised controlled trials of herbal interventions in Asean plus six countries: a systematic review. *Plos One*, 10(1), e108681.
- Prieto-Hontoria, P. L., Perez-Matute, P., Fernandez-Galilea, M., Bustos, M., Martinez, J. A., and Moreno-Aliaga, M. J. (2011). Role of obesity-associated dysfunctional adipose tissue in cancer: a molecular nutrition approach. *Biochimica et Biophysica Acta-Bioenergetics*, 1807(6), 664-678.
- Prins, J. B., and O'rahilly, S. (1997). Regulation of adipose cell number in man. *Clinical Science*, 92(1), 3-11.
- Prior, R. L., E. Wilkes, S., R. Rogers, T., Khanal, R. C., Wu, X., and Howard, L. R. (2010). Purified blueberry anthocyanins and blueberry juice alter development of obesity in mice fed an obesogenic high-fat diet. *Journal of Agricultural and Food Chemistry*, 58(7), 3970-3976.
- Pryor, W. A. (1991). Can vitamin E protect humans against the pathological effects of ozone in smog? *The American Journal of Clinical Nutrition*, 53(3), 702-722. <https://doi.org/10.1093/ajcn/53.3.702>.
- Qatanani, M., and Lazar, M. A. (2007). Mechanisms of obesity-associated insulin resistance: many choices on the menu. *Genes and Development*, 21, 1443-1455.
- Qiao, L., Kinney, B., Schaack, J., and Shao, J. (2011). Adiponectin inhibits lipolysis in mouse adipocytes. *Diabetes*, 60(5), 1519-1527.

- Qing-hua, Y., Qing, W., and Xiao-qin, M. (2012). Determination of major and trace elements in six herbal drugs for relieving heat and toxicity by ICP-AES with microwave digestion. *Journal of Saudi Chemical Society*, 16(3), 287-290.
- Rahman, M. M., Sayeed, M. S. B., Haque, M. A., Hassan, M. M., and Islam, S. A. (2012). Phytochemical screening, antioxidant, anti-Alzheimer and anti-diabetic activities of *Centella asiatica*. *Journal Natural Product Plant Resources*, 2(4), 504-511.
- Rains, T. M., Agarwal, S., and Maki, K. C. (2011). Antiobesity effects of green tea catechins: a mechanistic review. *The Journal of Nutritional Biochemistry*, 22(1), 1-7.
- Rajala, M. W., and Scherer, P. E. (2003). Mini review: the adipocyte-at the crossroads of energy homeostasis, inflammation and atherosclerosis. *Endocrinology*, 144(9), 3765-3773.
- Rajeh, M. A. B., Kwan, Y. P., Zakaria, Z., Latha, L. Y., Jothy, S. L., and Sasidharan, S. (2012). Acute toxicity impacts of *Euphorbia hirta* L extract on behavior, organs body weight index and histopathology of organs of the mice and *Artemia salina*. *Pharmacognosy Research*, 4(3), 170-177.
- Rajendra, P., Nandakumar, N., Rengarajan, T., Palaniswami, R., Gnanadhas, E.N., Lakshminarasiah, U., Gopas, J. and Nishigaki, I. (2014). Antioxidants and human diseases. *Clinica Chimica Acta*, 436, 332-347.
- Ramirez, J. E., Zambrano, R., Sepúlveda, B., and Simirgiotis, M. J. (2014). Antioxidant properties and hyphenated HPLC-PDA-MS profiling of chilean pica mango fruits (*Mangifera indica* L. Cv. piqueño). *Molecules*, 19(1), 438–458. <https://doi.org/10.3390/molecules19010438>.
- Ramos, S. J., Faquin, V., Guilherme, L. R. G., Castro, E. M., Ávila, F. W., Carvalho, G. S., and Oliveira, C. (2010). Selenium biofortification and antioxidant activity in lettuce plants fed with selenate and selenite. *Plant, Soil and Environment*, 56(12), 584-588.
- Ranjbar, S., Nayebi, N., Larijani, B., and Abdollahi, M. (2009). A systematic review of the efficacy and safety of herbal medicines used in the treatment of obesity. *World Journal of Gastroenterology*, 15(25), 3073-3085.
- Ranjbar, S., Nayebi, N., Moradi, L., Mehri, A., Larijani, B. and Abdollahi, M. (2010). The efficacy and safety of herbal medicines used in the treatment of hyperlipidemia; a systematic review. *Current Pharmaceutical Design*, 16(26), 2935-2947.
- Rasekh, H. R., Nazari, P., Kamli-Nejad, M., and Hosseinzadeh, L. (2008). Acute and subchronic oral toxicity of *Galega officinalis* in rats. *Journal of Ethnopharmacology*, 116 (1), 21-26.

- Rashid, T.S. (2016) Antibacterial activity of *R. Rhus coriaria* crude extracts against selected bacterial pathogens, PhD Thesis, UPM.
- Raya, K. B., Ahmad, S. H., Farhana, S. F., Mohammad, M., Tajidin, N. E., and Parvez, A. (2015). Changes in phytochemical contents in different parts of *Clinacanthus nutans* (Burm. f.) lindau due to storage duration. *Bragantia*, 44(4), 445-452. <http://dx.doi.org/10.1590/1678-4499.0469>.
- Rayalam, S., Della-Fera, M. A. and Baile, C. A. (2008). Phytochemicals and regulation of the adipocyte life cycle. *The Journal of Nutritional Biochemistry*, 19(11), 717-726.
- Raza, M., Al-shabanah, O., Elhadiyah, T., and Almajed, A. (2002). Effect of prolonged vigabatrin treatment on haematological and biochemical parameters in plasma, liver and kidney of Swiss albino mice. *Scientia Pharmaceutica*, 70(2), 135-145.
- Reeves, P. G., Nielsen, F. H., and Fahey Jr, G. C. (1993). AIN-93 purified diets for laboratory rodents: final report of the American Institute of Nutrition ad hoc writing committee on the reformulation of the AIN-76A rodent diet.
- Reddy, J. K. (2004). Peroxisome proliferators and peroxisome proliferator-activated receptor  $\alpha$ : biotic and xenobiotic sensing. *The American Journal of Pathology*, 164(6), 2305-2321.
- Rehman, Z. U. (2003). Evaluation of antioxidant activity of methanolic extract from peanut hulls in fried potato chips. *Plant Foods for Human Nutrition*, 58(1), 75-83.
- Renehan, A. G., Tyson, M., Egger, M., Heller, R. F. and Zwahlen, M. (2008). Body-mass index and incidence of cancer: a systematic review and meta-analysis of prospective observational studies. *The Lancet*, 371(9612), 569-578.
- Ríos, J. J., Blasco, B., Cervilla, L. M., Rosales, M. A., Sanchez-Rodriguez, E., Romero, L., and Ruiz, J. M. (2009). Production and detoxification of  $H_2O_2$  in lettuce plants exposed to selenium. *Annals of Applied Biology*, 154(1), 107-116. <https://doi.org/10.1111/j.1744-7348.2008.00276.x>
- Rocandio, A. M., Ansotegui, L. and Arroyo, M. (2001). Comparison of dietary intake among overweight and non-overweight schoolchildren. *International Journal of Obesity and Related Metabolic Disorders*, 25(11).
- Rodushkin, I., Ruth, T., and Huhtasaari, Å. (1999). Comparison of two digestion methods for elemental determinations in plant material by ICP techniques. *Analytica Chimica Acta*, 378(1-3), 191-200.
- Roeslan, M. O., Ayudhya, T. D. N., and Kootongkaew, S. (2012). Characteristics of *Clinacanthus nutans* extraction from Thailand and Indonesia (preliminary study). In *Conference on Graduate Student Network of Thailand*.

- Rogers, C. Q., Ajmo, J. M., and You, M. (2008). Adiponectin and alcoholic fatty liver disease. *IUBMB Life*, 60: 790–797.
- Rohner-Jeanrenaud, F., and Jeanrenaud, B. (1996). The discovery of leptin and its impact in the understanding of obesity. *European Journal of Endocrinology*, 135(6), 649-650.
- Roosita, K., Kusharto, C. M., Sekiyama, M., Fachrerozi, Y. and Ohtsuka, R. (2008). Medicinal plants used by the villagers of a Sundanese community in West Java, Indonesia. *Journal of Ethno-pharmacology*, 115(1), 72-81.
- Rosini, T., Silva A., and Moraes, C. (2012). Diet-induced obesity: rodent model for the study of obesity-related disorders. *Revista da Associação Médica Brasileira*, 58(3), 383-387.
- Rubin, L. F. (1983). Toxicologic update of dimethyl sulfoxide. *Annals of the New York Academy of Sciences*, 411(1), 6-10.
- Rudel, L., and Morris, M. (1973). Determination of cholesterol using O-phthalaldehyde. *Journal of Lipid Research*, 14(3), 364-366.
- Rudelle, S., Ferruzzi, M., Cristiani, I., Moulin, J., Mace, K., Acheson, K., and Tappy, L. (2007). Effect of a thermogenic beverage on 24 hour energy metabolism in humans. *Obesity*, 15(2), 349-355.
- Russell, E. S., and Bernstein, S. E. (1966). Blood and blood formation. *Biology of the Laboratory Mouse*, 2, 351-372.
- Russell, A., Gastaldi, G., Bobbioni-Harsch, E., Arboit, P., Gobelet, C., Dériaz, O., and Giacobino, J. P. (2003). Lipid peroxidation in skeletal muscle of obese as compared to endurance-trained humans: a case of good vs. bad lipids? *FEBS Letters*, 551(1-3), 104-106.
- Russo, G. L., Russo, M., and Spagnuolo, C. (2014). The pleiotropic flavonoid quercetin: from its metabolism to the inhibition of protein kinases in chronic lymphocytic leukemia. *Food and Function*, 5(10), 2393-2401.
- Saganuwan, S. A., Aondoaver, A. D., and Ronan, I. T. (2014). Reassessment of acute and chronic toxicity effects of aqueous leaf extract of *Morinda lucida* in *Rattus norvegicus*. *Journal of Hematology Research*, 1, 36-46.
- Sakdarat, S., Shuyprom, A., Pientong, C., Ekalaksananan, T., and Thongchai, S. (2009). Bioactive constituents from the leaves of *Clinacanthus nutans* Lindau. *Bioorganic and Medicinal Chemistry*, 17(5), 1857–1860.
- Sano, M., Takenaka, Y., Kojima, R., Saito, S. I., Tomita, I., Katou, M. and Shibuya, S. (1986). Effects of pu-erh tea on lipid metabolism in rats. *Chemical and Pharmaceutical Bulletin*, 34(1), 221-228.

- Santos, E. W., Oliveira, D. C. D., Hastreiter, A., Silva, G. B. D., Beltran, J. S. D. O., Tsujita, M., and Borelli, P. (2016). Hematological and biochemical reference values for C57BL/6, Swiss Webster and BALB/c mice. *Brazilian Journal of Veterinary Research and Animal Sciences*, 53(2), 138-145.
- Sarega, N., Imam, M., Ooi, D., Chan, K., Md Esa, N., Zawawi, N., and Ismail, M. (2016a). Phenolic rich extract from *Clinacanthus nutans* attenuates hyperlipidemia-associated oxidative stress in rats. *Oxidative Medicine and Cellular Longevity*, 2016. <http://dx.doi.org/10.1155/2016/4137908>.
- Sarega, N., Imam, M. U., Esa, N. M., Zawawi, N., and Ismail, M. (2016b). Effects of phenolic-rich extracts of *Clinacanthus nutans* on high-fat and high cholesterol diet-induced insulin resistance. *BMC Complementary and Alternative Medicine*, 16(1), 88.
- Sarma, D. N., Barrett, M. L., Chavez, M. L., Gardiner, P., Ko, R., Mahady, G. B., and Dog, T. L. (2008). Safety of green tea extracts. *Drug Safety*, 31(6), 469-484.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M., and Latha, L. Y. (2011). Extraction, isolation and characterization of bioactive compounds from plants' extracts. *African Journal of Traditional, Complementary and Alternative Medicines*, 8(1).
- Sayyad, M., Tiang, N., Kumari, Y., Goh, B. H., Jaiswal, Y., Rosli, R. and Shaikh, M. F. (2017). Acute toxicity profiling of the ethyl acetate fraction of *Swietenia macrophylla* seeds and in-vitro neuroprotection studies. *Saudi Pharmaceutical Journal*, 25(2), 196-205.
- Scazzocchio, B., Vari, R., Filesi, C., D'archivio, M., Santangelo, C., Giovannini, C., and Masella, R. (2011). Cyanidin-3-O- $\beta$ -glucoside and protocatechuic acid exert insulin-like effects by upregulating PPAR $\gamma$  activity in human omental adipocytes. *Diabetes*, 60(9), 2234-2244.
- Scheen, A. J. and Lefebvre, P. J. (1999). Pharmacological treatment of obesity: present status. *International Journal of Obesity and Related Metabolic Disorders*, 199(3), 47-53.
- Scherer, P. E., Williams, S., Fogliano, M., Baldini, G., and Lodish, H. F. (1995). A novel serum protein similar to C1q, produced exclusively in adipocytes. *Journal of Biological chemistry*, 270(45), 26746-26749.
- Schindhelm, R. K., Diamant, M., and Heine, R. J. (2007). Nonalcoholic fatty liver disease and cardiovascular disease risk. *Current Diabetes Reports*, 7(3), 181-187.
- Schindhelm, R. K., Diamant, M., Dekker, J. M., Tushuizen, M. E., Teerlink, T., and Heine, R. J. (2006). Alanine aminotransferase as a marker of non-alcoholic fatty liver disease in relation to type 2 diabetes mellitus and cardiovascular disease. *Diabetes/Metabolism Research and Reviews*, 22(6), 437-443.

- Sekar, M. and Rashid, N. (2016). Formulation, evaluation and antibacterial properties of herbal Ointment containing methanolic extract of *Clinacanthus nutans* leaves. *International Journal of Pharmaceutical and Clinical Research*, 8(8), 1170-1174.
- Sekiya, M., Osuga, J. I., Okazaki, H., Yahagi, N., Harada, K., Shen, W. J. and Okazaki, M. (2004). Absence of hormone-sensitive lipase inhibits obesity and adipogenesis in Lep ob/ob mice. *Journal of Biological Chemistry*, 279(15), 15084-15090.
- Seo, M., Lee, Y., Hwang, J., Kim, K., and Lee, B. (2015). The inhibitory effects of quercetin on obesity and obesity-induced inflammation by regulation of MAPK signaling. *The Journal of Nutritional Biochemistry*, 26(11), 1308–1316. Retrieved from <http://dx.doi.org/10.1016/j.jnutbio.2015.06.005>.
- Sen, A., Mishra, S., Ghosh, A., Bhattacharjee, B., De, S., Ghosh, A., and Bandyopadhyay, D. (2017). Aqueous leaf extract of Tulsi (*Ocimum sanctum*) protects against high-fat diet-induced injury to rat liver through antioxidant mechanisms: a dose-and time-dependent study. *Journal of Pharmacy Research*, 11(4), 334-351.
- Serfilippi, L. M., Stackhouse Pallman, D. R., Russell, B., and Spainhour, C. B. (2003). Serum clinical chemistry and hematology reference values in outbred stocks of albino mice from three commonly used vendors and two inbred strains of albino mice. *Journal of the American Association for Laboratory Animal Science*, 42(3), 46-52.
- Sethi, J. K., and Vidal-Puig, A. J. (2007). Thematic review series: adipocyte biology. Adipose tissue function and plasticity orchestrate nutritional adaptation. *Journal of Lipid Research*, 48(6), 1253-1262.
- Shah, S., Shah, S. M. M., Ahmad, Z., Yaseen, M., Shah, R., Sadiq, A. and Khan, B. (2015). Phytochemicals, *in vitro* antioxidant, total phenolic contents and phytotoxic activity of *Cornus macrophylla* wall bark collected from the North-West of Pakistan. *Pakistan Journal Pharmacology Science*, 28(1), 23-28.
- Shahidi, F. and Naczk, M. (2003). *Phenolics in food and nutraceuticals*. Boca Raton, Florida: CRC press.
- Shimano, H., Shimomura, I., Hammer, R. E., Herz, J., Goldstein, J. L., Brown, M. S. and Horton, J. D. (1997). Elevated levels of SREBP-2 and cholesterol synthesis in livers of mice homozygous for a targeted disruption of the SREBP-1 gene. *Journal of Clinical Investigation*, 100(8), 2115.
- Shen, B., Chen, H., Shen, C., Xu, P., Li, J., Shen, G., Yuan, H. and Han, Jin. (2015). Hepatoprotective effects of lignans extract from *Herpetospermum caudigerum* against CCl<sub>4</sub>-induced acute liver injury in mice. *Journal of Ethnopharmacology*, 164, 46-52.

- Shimomura, I., Bashmakov, Y., and Horton, J. D. (1999a). Increased levels of nuclear SREBP-1c associated with fatty livers in two mouse models of diabetes mellitus. *Journal of Biological Chemistry*, 274(42), 30028-30032.
- Shimomura, I., Hammer, R. E., Ikemoto, S., Brown, M. S., and Goldstein, J. L. (1999b). Leptin reverses insulin resistance and diabetes mellitus in mice with congenital lipodystrophy. *Nature*, 401(6748), 73-76.
- Shoelson, S. E., Lee, J., and Goldfine, A. B. (2006). Inflammation and insulin resistance. *The Journal of Clinical Investigation*, 116 (7), 1793-1801.
- Shrestha, S., Ehlers, S. J., Lee, J.-Y., Fernandez, M.-L., and Koo, S. I. (2009). Dietary green tea extract lowers plasma and hepatic triglycerides and decreases the expression of sterol regulatory element-binding protein-1c mRNA and its responsive genes in fructose-fed, ovariectomized rats. *The Journal of Nutrition*, 139 (4), 640-645.
- Shuke, N., Aburano, T., Okizaki, A., Zhao, C., Nakajima, K., Yokoyama, K., and Tonami, N. (2003). Estimation of fractional liver uptake and blood retention of 99mTc-DTPA-galactosyl human serum albumin: an application of a simple graphical method to dynamic SPECT. *Nuclear Medicine Communications*, 24(5), 503-511.
- Siersbæk, R., Nielsen, R., and Mandrup, S. (2010). PPAR $\gamma$  in adipocyte differentiation and metabolism—novel insights from genome-wide studies. *FEBS Letters*, 584(15), 3242-3249.
- Sikora, E., and Bodziarczyk, I. (2012). Composition and antioxidant activity of kale (*Brassica oleracea* L. var. acephala) raw and cooked. *Acta scientiarum polonorum. Technologia alimentaria*, 11(3), 239-248.
- Skinner, A.C., and Skelton, J.A. (2014). Prevalence and trends in obesity and severe obesity among children in the United States, 1999-2012. *The Journal of American Medical Association Pediatrics*, 168(6), 561-566.
- Slavin, J. L. (2005). Dietary fiber and body weight. *Nutrition*, 21(3), 411-418.
- Slawik, M., and Vidal-Puig, A. (2006). Lipotoxicity, overnutrition and energy metabolism in aging. *Ageing Research Reviews*, 5(2), 144-164.
- Sova, M. (2012). Antioxidant and antimicrobial activities of cinnamic acid derivatives. *Mini Reviews in Medicinal Chemistry*, 12(8), 749-767.
- Spiegelman, B. M., and Flier, J. S. (2001). Obesity and the regulation of energy balance. *Cell*, 104(4), 531-543.
- Starmans, D., and Nijhuis, H. (1996). Extraction of secondary metabolites from plant material: a review. *Trends Food Science and Technology*, 7(6):191-197.

- Strable, M. S., and Ntambi, J. M. (2010). Genetic control of de novo lipogenesis: role in diet-induced obesity. *Critical reviews in biochemistry and molecular biology*, 45(3), 199-214.
- Stern, J. S., Hirsch, J., Blair, S. N., Foreyt, J. P., Frank, A., Kumanyika, S. K., and Stunkard, A. J. (1995). Weighing the options: criteria for evaluating weight-management programs the committee to develop criteria for evaluating the outcomes of approaches to prevent and treat obesity. *Obesity Research*, 3(6), 591-604.
- Stein, F. L. P., Schmidt, B., Furlong, E. B., Soares, L. A. S., Soares, M. C. F., Vaz, M. R. C., and Baisch, A. L. M. (2005). Vascular responses to extractable fractions of *Ilex paraguariensis* in rats fed standard and high-cholesterol diets. *Biological Research for Nursing*, 7(2), 146-156.
- Sulaiman, I. S. C., Basri, M., Chan, K. W., Ashari, S. E., Masoumi, H. R. F., and Ismail, M. (2015). In vitro antioxidant, cytotoxic and phytochemical studies of *Clinacanthus nutans* Lindau leaf extracts. *African Journal of Pharmacy and Pharmacology*, 9(34), 861-874.
- Sulaiman, I. S. C., Basri, M., Masoumi, H. R. F., Chee, W. J., Ashari, S. E., and Ismail, M. (2017). Effects of temperature, time, and solvent ratio on the extraction of phenolic compounds and the anti-radical activity of *Clinacanthus nutans* Lindau leaves by response surface methodology. *Chemistry Central Journal*, 11(54), 1-11.
- Sun, K., Kusminski, C., and Scherer, P. E. (2011). Adipose tissue remodeling and obesity. *The Journal of Clinical Investigation*, 121(6), 2094-2101.
- Sung, Y.Y., Yoon, T., Yang, W. K., Kim, S. J., Kim, D. S., and Kim, H. K. (2013). The anti-obesity effect of *Polygonum aviculare* L. ethanol extract in high-fat diet-induced obese mice. *Evidence-Based Complementary and Alternative Medicine*, 2013, 1-11. <http://dx.doi.org/10.1155/2013/626397>.
- Szkudelska, K., Szkudelski, T., and Nogowski, L. (2002). Daidzein, coumestrol and zearalenone affect lipogenesis and lipolysis in rat adipocytes. *Phytomedicine*, 9(4), 338-345.
- Taib, I. S., Budin, S. B., Ain, S. M. S. N., Mohamed, J., Louis, S. R., Das, S., and Hidayatulfathi, O. (2009). Toxic effects of *Litsea elliptica* Blume essential oil on red blood cells of Sprague-Dawley rats. *Journal of Zhejiang University Science B*, 10(11), 813.
- Tammen, S., Friso, S. and Choi, S. (2013). Epigenetics: the link between nature and nurture. *Molecular Aspects of Medicine*, 34(4), 753-764.
- Tamori, Y., Masugi, J., Nishino, N., and Kasuga, M. (2002). Role of peroxisome proliferator-activated receptor-Y in maintenance of the characteristics of mature 3T3-L1 adipocytes. *Diabetes*, 51(7), 2045-2055.

- Tanizawa, T., Yamaguchi, A., Uchiyama, Y., Miyaura, C., Ikeda, T., Ejiri, S., and Nakamura, T. (2000). Reduction in bone formation and elevated bone resorption in ovariectomized rats with special reference to acute inflammation. *Bone*, 26(1), 43-53.
- Tao, R., Xiong, X., DePinho, R.A., Deng, C.X., and Dong, X.C. (2013). Hepatic SREBP-2 and cholesterol biosynthesis are regulated by FoxO3 and Sirt6. *Journal of Lipid Research*, 54(10), 2745-2753.
- Targher, G., and Arcaro, G. (2007). Non-alcoholic fatty liver disease and increased risk of cardiovascular disease. *Atherosclerosis*, 191(2), 235-240.
- Teh, C. C., Khoo, Z. Y., Khursiah, F., Rao, N. K., and Chin, J. H. (2010). Acetyl cholinesterase inhibitory activity of star fruit and its effect on serum lipid profiles in rats. *International Food Resources Journal*, 17, 987-994.
- Teixeira, S. (2002). Bioflavonoids: proanthocyanidins and quercetin and their potential roles in treating musculoskeletal conditions. *Journal of Orthopaedic and Sports Physical Therapy*, 32(7), 357-363.
- Teodoro, J. S., Zouhar, P., Flachs, P., Bardova, K., Janovska, P., Gomes, A. P., and Kopecký, J. (2014). Enhancement of brown fat thermogenesis using chenodeoxycholic acid in mice. *International Journal of Obesity*, 38(8), 1027-1034.
- Teshome, K., Gebre-Mariam, T., Asres, K., and Engidawork, E. (2010). Toxicity studies on dermal application of plant extract of *Dodonaea viscosa* used in Ethiopian traditional medicine. *Phytotherapy Research*, 24(1), 60-69.
- Thaipong, K., Boonprakob, U., Crosby, K., Zevallos, L.C. and Byrne, D.H. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis*, 19(7,8), 669–675.
- Thomas, T.R., Pellechia, J., Rector, R.S., Sun, G.Y., Sturek, M.S., and Laughlin, M.H. (2002). Exercise training does not reduce hyperlipidemia in pigs fed a high-fat diet. *Metabolism-Clinical and Experimental*, 51(12), 1587-1595.
- Thompson, A. M., Zhang, Y., Tong, W., Xu, T., Chen, J., Zhao, L., and He, J. (2011). Association of obesity and biomarkers of inflammation and endothelial dysfunction in adults in Inner Mongolia, China. *International Journal of Cardiology*, 150(3), 247-252.
- Tian, Y., Zou, B., Yang, L., Xu, S., Yang, J., Yao, P., and Li, C. (2011). High molecular weight persimmon tannin ameliorates cognition deficits and attenuates oxidative damage in senescent mice induced by D-galactose. *Food and Chemical Toxicology*, 49(8), 1728-1736.

- Tiew, W. P., P'ng, X. W., Chin, J. H., and Akowuah, G. A. (2014). Effect of methanol extract of *Clinacanthus nutans* on serum biochemical parameters in rats. *Journal of Applied Pharmacy*, 6(1), 77-86.
- Tilg, H., and Moschen, A. R. (2006). Adipocytokines: mediators linking adipose tissue, inflammation and immunity. *Nature Reviews Immunology*, 6(10), 772-783.
- Tokalioglu, S. (2012). Determination of trace elements in commonly consumed medicinal herbs by ICP-MS and multivariate analysis. *Food Chemistry*, 134(4), 2504-2508.
- Tormen, L., Torres, D. P., Dittert, I. M., Araújo, R. G., Frescura, V. L., and Curtius, A. J. (2011). Rapid assessment of metal contamination in commercial fruit juices by inductively coupled mass spectrometry after a simple dilution. *Journal of Food Composition and Analysis*, 24(1), 95-102.
- Toyama, T., Kudo, N., Hibino, Y., Mitsumoto, A., Nishikawa, M., and Kawashima, Y. (2007). Effects of pioglitazone on stearoyl-CoA desaturase in obese Zucker fa/fa rats. *Journal of Pharmacological Sciences*, 104(2), 137-145.
- Tresserra, A., Rimm, E.B., Medina-Remón, A., Martínez-González, M.A., López Sabater, M.C., Covas, M.I., Corella, D., Salas-Salvadó, J., Gómez-Gracia, E., Lapetra, J., Arós, F., Fiol, M., Ros, E., Serra-Majem, L., Pintó, X., Muñoz, M.A., Gea, A., Ruiz-Gutiérrez, V., Estruch, R., and Lamuela-Raventós, R.M. (2014). Polyphenol intake and mortality risk: a re-analysis of the predimed trial. *BioMed Central Medicine*, 12(77), 1-11.
- Tsuchida, A., Yamauchi, T., Takekawa, S., Hada, Y., Ito, Y., Maki, T., and Kadokawa, T. (2005). Peroxisome proliferator-activated receptor (PPAR)  $\alpha$  activation increases adiponectin receptors and reduces obesity-related inflammation in adipose tissue: comparison of activation of PPAR $\alpha$ , PPAR $\gamma$ , and their combination. *Diabetes*, 54(12), 3358-3370.
- Tu, S. F., Liu, R. H., Cheng, Y. B., Hsu, Y. M., Du, Y. C., El-Shazly, M., and Chang, F. R. (2014). Chemical constituents and bioactivities of *Clinacanthus nutans* aerial parts. *Molecules*, 19(12), 20382-20390.
- Tuan, N. T., and Nicklas, T. A. (2009). Age, sex and ethnic differences in the prevalence of underweight and overweight, defined by using the CDC and IOTF cut points in Asian children. *European Journal of Clinical Nutrition*, 63(11), 1305-1312.
- Tubesha, Z., Iman, M.U., Mahmud, R., and Ismail, M. (2013). Study on the potential toxicity of thymoquinone-rich fraction nanoemulsion in Sprague Dawley rats. *Molecules*, 18(7), 7460-7472.
- Tuntiwachwuttikul, P., Pootaeng-On, Y., Pansa, P., Srisanpang, T., and Taylor, W. C. (2003). Sulfur-containing compounds from *Clinacanthus siamensis*. *Chemical and Pharmaceutical Bulletin*, 51(12), 1423-1425.

- Turner, P. V., Brabb, T., Pekow, C., and Vasbinder, M. A. (2011). Administration of substances to laboratory animals: routes of administration and factors to consider. *Journal of the American Association for Laboratory Animal Science*, 50(5), 600-613.
- Umar Imam, M., Ismail, M., George, A., Chinnappan, S. M., and Yusof, A. (2019). Aqueous leaf extract of *Clinacanthus nutans* improved metabolic indices and sorbitol-related complications in type II diabetic rats (T2D). *Food Science and Nutrition*, 1482-1493. <https://doi.org/10.1002/fsn3.988>.
- van de Sande-Lee, S., Pereira, F. R. S., Cintra, D. E., Fernandes, P. T., Cardoso, A. R., Garlipp, C. R., and Li, L. M. (2011). Partial reversibility of hypothalamic dysfunction and changes in brain activity after body mass reduction in obese subjects. *Diabetes*, 60(6), 1699-1704.
- van der Heijden, R. A., Morrison, M. C., Sheedfar, F., Mulder, P., Schreurs, M., Hommelberg, P. P. H. and Tietge, U. J. F. (2016). Effects of anthocyanin and flavanol compounds on lipid metabolism and adipose tissue associated systemic inflammation in diet-induced obesity. *Mediators of Inflammation*, 2016. <http://dx.doi.org/10.1155/2016/2042107>
- Vandenbergh, J. G., Drickamer, L. C., and Colby, D. R. (1972). Social and dietary factors in the sexual maturation of female mice. *Reproduction*, 28(3), 397-405.
- Vendrell, J., Broch, M., Vilarrasa, N., Molina, A., Gómez, J. M., Gutiérrez, C and Richart, C. (2004). Resistin, adiponectin, ghrelin, leptin, and proinflammatory cytokines: relationships in obesity. *Obesity Research*, 12(6), 962-971.
- Vermaak, I., Viljoen, A. M., and Hamman, J. H. (2011). Natural products in anti-obesity therapy. *Natural Product Reports*, 28(9), 1493-1533.
- Vidal-Puig, A. J., Considine, R. V., Jimenez-Liñan, M., Werman, A., Pories, W. J., Caro, J. F., and Flier, J. S. (1997). Peroxisome proliferator-activated receptor gene expression in human tissues. Effects of obesity, weight loss, and regulation by insulin and glucocorticoids. *Journal of Clinical Investigation*, 99(10), 2416-2422.
- Visht, S., and Chaturvedi, S. (2012). Isolation of natural products. *Current Pharma Research*, 2(3), 584-599.
- Viuda-Martos, M., Ruiz Navajas, Y., Sánchez Zapata, E., Fernández-López, J., and Pérez-Álvarez, J. A. (2010). Antioxidant activity of essential oils of five spice plants widely used in a Mediterranean diet. *Flavour and Fragrance Journal*, 25(1), 13-19.
- Wadden, T. A. (1993). Treatment of obesity by moderate and severe caloric restriction: results of clinical research trials. *Annals of Internal Medicine*, 119 (7-2), 688-693.

- Walker, J. (2001). The ELISA Guidebook Methods in Molecular Biology". M1 Cytoskeleton Methods and Protocols Gene Knockout Protocols Hmail Kola Joyce C. Salheim. <https://doi.org/10.1007/978-1-60327254-4>
- Wallenius, V., Wallenius, K., Ahren, B., Rudling, M., Carlsten, H., Dickson, S. L., and Jansson, J. O. (2002). Interleukin-6-deficient mice develop mature-onset obesity. *Nature Medicine*, 8(1), 75-79
- Wang, Z., Al-Regaiey, K. A., Masternak, M. M., and Bartke, A. (2006). Adipocytokines and lipid levels in Ames dwarf and calorie-restricted mice. *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, 61(4), 323-331.
- Wang, Y., Beydoun, M. A., Liang, L., Caballero, B., and Kumanyika, S. K. (2008). Will all Americans become overweight or obese? Estimating the progression and cost of the US obesity epidemic. *Obesity*, 16(10), 2323-2330
- Wang, C.Y., and Liao, J.K. (2012). A mouse model of diet-induced obesity and insulin resistance. In: Weichhart T. (Ed.), *mTOR Methods in Molecular Biology 821, Methods and Protocols*, Vol. 821. (pp. 421-433). New York, USA: Humana Press.
- Wang, C., Shi, C., Yang, X., Yang, M., Sun, H., and Wang, C. (2014). Celastrol suppresses obesity process via increasing antioxidant capacity and improving lipid metabolism. *European Journal of Pharmacology*, 744, 52-58.
- Wang, Y. and Huang, F. (2015). N-3 polyunsaturated fatty acids and inflammation in obesity: local effect and systemic benefit. *BioMedical Central Research International*, 2015(2015), 1-16. <http://dx.doi.org/10.1155/2015/581469>
- Wanikiat, P., Panthong, A., Sujayanon, P., Yoosook, C., Rossi, A. G., and Reutrakul, V. (2008). The anti-inflammatory effects and the inhibition of neutrophil responsiveness by *Barleria lupulina* and *Clinacanthus nutan* extracts. *Journal of Ethno-pharmacology*, 116(2), 234-244.
- Warwick, Z. S., and Schiffman, S. S. (1992). Role of dietary fat in calorie intake and weight gain. *Neuroscience and Biobehavioral Reviews*, 16(4), 585-596.
- Waturuocha, C., Fan, L., Kudo, L., Gilbert, W., and Rouch, A. (2017). Sex differences in blood pressure and mRNA expression of renal sodium transporters in mice under normal sodium diet. *The FASEB Journal*, 31(1), 855-2.
- Wei, Y., Chen, K., Adam, I., Whaley-Connell, T., Stump, C.S., Ibdah, J. A. and Sowers, J.R. (2008). Skeletal muscle insulin resistance: role of inflammatory cytokines and reactive oxygen species. *The American Journal of Physiology Regulatory Integrative and Comparative Physiology*, 294(3), 673-680.
- Weinreb, O., Mandel, S., Amit, T., and Youdim, M. B. (2004). Neurological mechanisms of green tea polyphenols in Alzheimer's and Parkinson's diseases. *The Journal of Nutritional Biochemistry*, 15(9), 506-516.

- Weinsier, R. L., Hunter, G. R., Heini, A. F., Goran, M. I., and Sell, S. M. (1998). The etiology of obesity: relative contribution of metabolic factors, diet and physical activity. *The American Journal of Medicine*, 105(2), 145-150.
- Werner, M., Chott, A., Fabiano, A. and Battifora, H. (2000). Effect of formalin tissue fixation and processing on immunohistochemistry. *The American Journal of Surgical Pathology*, 24(7), 1016-1019.
- White, U. A., and Stephens, J. M. (2010). Transcriptional factors that promote formation of white adipose tissue. *Molecular and cellular endocrinology*, 318(1-2), 10-14.
- White, P. J., and Brown, P. H. (2010). Plant nutrition for sustainable development and global health. *Annals of Botany*, 105(7), 1073-1080.
- Wilson, C. H., Nikolic, A., Kentish, S. J., Shalini, S., Hatzinikolas, G., Page, A. J., and Kumar, S. (2016). Sex-specific alterations in glucose homeostasis and metabolic parameters during ageing of caspase-2-deficient mice. *Cell Death Discovery*, 2, 16009.
- Wing, R., and Phelan, S. (2005). Long-term weight loss maintenances. *The American Journal of Clinical Nutrition*, 82(1), 222S-225S.
- Wink, M. (2015). Modes of action of herbal medicines and plant secondary metabolites. *Medicines*, 2(3), 251-286.
- Withrow, D., and Alter, D. A. (2011). The economic burden of obesity worldwide: a systematic review of the direct costs of obesity. *Obesity Reviews*, 12(2), 131-141.
- Wolfram, S., Raederstorff, D., Wang, Y., Teixeira, S. R., Elste, V., and Weber, P. (2005). Teavigotm (epigallocatechin gallate) supplementation prevents obesity in rodents by reducing adipose tissue mass. *Annals of Nutrition and Metabolism*, 49(1), 54–6.
- Wong, H., and Schotz, M. C. (2002). The lipase gene family. *Journal of Lipid Research*, 43(7), 993-999.
- Wong, F. C., Yong, A. L., Ong, H. C., and Chai, T. T. (2013). Evaluation of the antibacterial activities of selected medicinal plants and determination of their phenolic constituents. *Science Asia*, 39(6), 591-595.
- Wong, F. C., Yong, A. L., Ting, E. P. S., Khoo, S. C., Ong, H. C., and Chai, T. T. (2014). Antioxidant, metal chelating, anti-glucosidase activities and phytochemical analysis of selected tropical medicinal plants. *Iranian Journal of Pharmaceutical Research*, 13(4), 1409-1415.
- World Health Organization (2000a). *Obesity Preventing and Managing the Global Epidemic*. WHO Technical Report Series 894. Geneva, Switzerland.

World Health Organization (2000b). *The Asia-Pacific Perspective Redefining Obesity and Its Treatment*: International Obesity Taskforce. Geneva, Switzerland: Inoue, S., and Zimmet, P.

World Health Organization (2017) Obesity and Overweight Factsheet from the WHO: 2017 October 18. Available at:<http://www.who.int/news-room/factsheets/detail/obesity-and-overweight>.

Wu, T., Qi, X., Liu, Y., Guo, J., Zhu, R., W., and Yu, T. (2013). Dietary supplementation with purified mulberry (*Morus australis Poir*) anthocyanins suppresses body weight gain in high-fat diet fed C57BL/6 mice. *Food Chemistry*, 141(1), 482-487.

Xu, H., Barnes, G. T., Yang, Q., Tan, G., Yang, D., Chou, C. J., and Tartaglia, L. A. (2003). Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *The Journal of Clinical Investigation*, 112(12), 1821-1830.

Yamada, C., Saegusa, Y., Nahata, M., Sadakane, C., Hattori, T., and Takeda, H. (2015). Influence of aging and gender differences on feeding behavior and ghrelin-related factors during social isolation in mice. *Close One*, 10 (10), e0140094.

Yamauchi, T., Kamon, J., Ito, Y., Tsuchida, A., Yokomizo, T., Kita, S., and Murakami, K. (2003). Cloning of adiponectin receptors that mediate antidiabetic metabolic effects. *Nature*, 423(6941), 762.

Yang, L., Wen, K. S., Ruan, X., Zhao, Y. X., Wei, F., and Wang, Q. (2018). Response of plant secondary metabolites to environmental factors. *Molecules*, 23(4), 762.

Yang, Z. H., Miyahara, H., Takemura, S., and Hatanaka, A. (2011). Dietary saury oil reduces hyperglycemia and hyperlipidemia in diabetic KKAY mice and in diet-induced obese C57BL/6J mice by altering gene expression. *Lipids*, 46(5), 425-434.

Yin, X. M., Ding, W. X., and Gao, W. (2008). Autophagy in the liver. *Hepatology*, 47(5), 1773-1785.

Yokoyama, M., Se o, T., Park, T., Yagyu, H., Hu, Y., Son, N. H., and Pulawa, L. K. (2007). Effects of lipoprotein lipase and statins on cholesterol uptake into heart and skeletal muscle. *Journal of Lipid Research*, 48(3), 646-655.

Yokozawa, T., and Dong, E. (1997). Influence of green tea and its three major components upon low-density lipoprotein oxidation. *Experimental and Toxicologic Pathology*, 49(5), 329–335.

- Yong, Y. K., Tan, J. J., Teh, S. S., Mah, S. H., Ee, G. C. L., Chiong, H. S., and Ahmad, Z. (2013). Clinacanthus nutans extracts are antioxidant with antiproliferative effect on cultured human cancer cell lines. *Evidence-Based Complementary and Alternative Medicine*, 2013:462751. 14.
- Yu, J., Ahmedna, M. and Goktepe, I. (2005). Effects of processing methods and extraction solvents on concentration and antioxidant activity of peanut skin phenolics. *Food Chemistry*, 90(1-2), 199-206.
- Yu, Q., Duan, Z. H., Duan, W. W., Shang, F. F., and Yang, G. X. (2015, December). Analysis and evaluation of nutrition composition of *Clinacanthus nutans*. In *Resources, Environment and Engineering II: Proceedings of the 2nd Technical Congress on Resources, Environment and Engineering (CREE 2015, Hong Kong, 25-26 September 2015)* (p. 369). CRC Press.
- Yun, J. W. (2010). Possible anti-obesity therapeutics from nature. A review. *Phytochemistry*, 71(14-15), 1625-1641.
- Yusri, N. M., Chan, K. W., Iqbal, S., and Ismail, M. (2012). Phenolic content and antioxidant activity of *Hibiscus cannabinus* L. seed extracts after sequential solvent extraction. *Molecules*, 17(11), 12612-12621.
- Zakaria, Z. A., Rahim, M. H. A., Mohtarrudin, N., Kadir, A. A., Cheema, M. S., Ahmad, Z., and Tohid, S. F. M. (2016). Acute and sub-chronic oral toxicity studies of methanol extract of *Clinacanthus nutans* in mice. *African Journal of Traditional, Complementary and Alternative Medicines*, 13(2), 210-222.
- Zarrouki, B., Pillon, N. J., Kalbacher, E., Soula, H. A., N'Jomen, G. N., Grand, L., and Soulage, C. O. (2010). Cirsimarin, a potent antilipogenic flavonoid, decreases fat deposition in mice intra-abdominal adipose tissue. *International Journal of Obesity*, 34(11), 1566-1575.
- Zawawi, N. (2011). *Effect of Strobilanthes crispus extract and individual polyphenols on lipolysis*. PhD Thesis. University of Nottingham, UK.
- Zhang, L., Lan, G. E., Parimoo, S., Stenn, K., and Prouty, S. M. (1999). Human stearoyl-CoA desaturase: alternative transcripts generated from a single gene by usage of tandem polyadenylation sites. *Biochemical Journal*, 340(1), 255-264.
- Zhang, Y., Xie, M., Xue, J., and Gu, Z. (2007). Osthole improves fat milk-induced fatty liver in rats: modulation of hepatic PPAR-alpha/gamma-mediated lipogenic gene expression. *Planta Medica*, 73(08), 718-724.
- Zhao, J., Sun, X. B., Ye, F., and Tian, W.X. (2011). Suppression of fatty acid synthase, differentiation and lipid accumulation in adipocytes by curcumin. *Molecular and Cellular Biochemistry*, 351(1-2), 19-28.

- Zhou, C. J., Huang, S., Liu, J. Q., Qiu, S. Q., Xie, F. Y., Song, H. P., and Lai, X. P. (2013). Sweet tea leaves extract improves leptin resistance in diet-induced obese rats. *Journal of Ethno-pharmacology*, 145(1), 386-392.
- Zhou, M., Xu, A., Tam, P. K., Lam, K. S., Chan, L., Hoo, R. L., and Wang, Y. (2008). Mitochondrial dysfunction contributes to the increased vulnerabilities of adiponectin knockout mice to liver injury. *Hepatology*, 48(4), 1087-1096.
- Zhou, Y., and Rui, L. (2013). Leptin signaling and leptin resistance. *Frontier Medicine*, 7(2), 207-222.
- Zhu, X.X., Ma, S.T., Eirin, A., Woolard, J.R., Hickson, L.J., Sun, D., Lerman, A. and Lerman, L.O. (2016). Functional plasticity of adipose-derived stromal cells during development of obesity. *Stem Cells Translational Medicine*, 5, 893-900
- Zinman, B., Hanley, A.J., Harris, S.B., Kwan, J. and Fantus, I.G. (1999). Circulating tumor necrosis factor  $\alpha$  concentrations in a native Canadian population with high rates of type 2 diabetes mellitus. *Journal of Clinical Endocrinology and Metabolism*, 84, 272–278.
- Zor, T., and Selinger, Z. (1996). Linearization of the Bradford protein assay increases its sensitivity: theoretical and experimental studies. *Analytical Biochemistry*, 236(2), 302-308.
- Zulkipli, I. N., Rajabalaya, R., Idris, A., Sulaiman, N. A., and David, S. R. (2017). *Clinacanthus nutans*: a review on ethnomedicinal uses, chemical constituents and pharmacological properties. *Pharmaceutical Biology*, 55(1), 1093-1113.