



UNIVERSITI PUTRA MALAYSIA

**INVASIVE PROPERTY OF AN ATTENUATED GDHA- DERIVATIVE OF
Pasteurella multocida B:2**

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By

FARAHANI BINTI MUHAMMAD AZAM

Thesis Submitted to the Department of Cell and Molecular Biology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Bachelor of Science (Hons.)

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Abstract of thesis presented to the Department of Cell and Molecular Biology in fulfillment of the requirement for the degree of Bachelor of Science (Hons.)

INVASIVE PROPERTY OF AN ATTENUATED GDH⁻ DERIVATIVE OF *Pasteurella multocida* B:2

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Haemorrhagic septicaemia (HS) is a disease of buffaloes and cattle that can cause acute and fatal mortality if contracted. The animals are infected by the bacteria through the intranasal pathway which translocated intracellularly into the vascular system. Live-attenuated vaccine, containing attenuated live bacteria is the alternative vaccine that has been shown to confer longer period of immunity towards the field-animals. The attenuated GdhA⁻ derivative of *Pasteurella multocida* B:2 has been proven to be a suitable candidate as a live-attenuated vaccine towards HS among other live-attenuated strains tested. A further insight into the intracellular viability and pathogenicity of this disease can be discovered by understanding the mechanism of interaction between the host cell and the vaccine candidate. In this study, the GdhA⁻ derivative strain was shown to adhere and to invade bovine aortic endothelial cells (BAECs) *in vitro* alongside another derivative strain, the AroA⁻ derivative and the wild-type of *P. multocida* B:2 strains as a control. Infection of GdhA⁻ derivative towards BAECs had shown no cytotoxicity effect with >80% viability towards the cell lines through trypan blue viability assessment. Adhesion and invasion of the GdhA⁻ derivative was shown to be more efficient with highest adhesion rate (45.75 ± 5.74 number of bacteria/BAEC cell) and invasion rate (1.41 ± 0.27 number of bacteria/BAEC cell) when compared to the AroA⁻ derivative and the wild-type strains.

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keperluan untuk ijazah Sarjana Muda Sains (Kepujian)

PENCIRIAN INVASIF OLEH DERIVATIF GDHA ATENUAT DARIPADA *Pasteurella multocida* B:2

Oleh

FARAHANI BINTI MUHAMMAD AZAM

Sampar berdarah (HS) adalah penyakit berjangkit yang menyerang populasi kerbau dan lembu, menyebabkan maut dengan kadar kematian yang tinggi. Secara amnya, haiwan-haiwan tersebut telah dijangkiti bakteria melalui laluan intranasal yang ditranslokasikan secara intraselular ke dalam sistem vaskular. Vaksin hidup-atenuat, mengandungi bakteria hidup yang teratenuat adalah vaksin alternatif yang telah terbukti memberikan imuniti dalam tempoh masa yang panjang terhadap haiwan tersebut. Di antara strain-strain hidup lain yang dilemahkan dan telah diuji, derivatif GdhA⁻ yang teratenuat daripada *P. multocida* B:2 telah terbukti menjadi calon yang sesuai sebagai strain hidup yang dilemahkan terhadap HS. Penyingkapan terhadap kebolehhidupan intraselular dan patogenisiti penyakit ini boleh difahami dengan menyelidiki mekanisme interaksi antara sel tuan rumah dan calon vaksin. Melalui kajian ini, derivatif GdhA⁻ yang dilemahkan telah menunjukkan dapat melekat kepada dan menakluki sel-sel endothelial aorta lembu (BAECs) secara *in vitro* bersama strain hidup yang lain; strain derivatif AroA⁻ dan strain jenis liar daripada *P. multocida* B:2 sebagai kawalan. Jangkitan daripada derivatif GdhA⁻ terhadap BAECs telah tidak menunjukkan sebarang kesan sitotoksiti dengan viability >80% terhadap barisan sel melalui penaksiran kebolehhidupan “trypan blue”. Pelekatan dan penaklukan derivatif GdhA adalah lebih berkesan dengan nilai tinggi kadar pelekatan (45.75 ± 5.74 nombor bakteria/sel BAEC) dan kadar penaklukan (1.41 ± 0.27 nombor bakteria/sel BAEC) apabila dibandingkan dengan strain derivatif AroA⁻ dan jenis liar.

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APPROVAL

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DECLARATION

Declaration by undergraduate student:

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LIST OF ABBREVIATIONS

BAECs	-	bovine aortic endothelial cells
BHI	-	brain-heart infusion broth
bp	-	basepairs
CO ₂	-	carbon dioxide
CFU	-	colony forming unit
EDTA	-	ethylenediamine tetraacetate
°C	-	degree Celcius
<i>gdhA</i>	-	glutamate dehydrogenase
HS	-	haemorrhagic septicaemia
h	-	hour
IgG	-	immunoglobulin G
kb	-	kilobasepairs
µl	-	microliter
ml	-	mililitre
min	-	minute
MOI	-	multiplicity of infection
rpm	-	rotation per minute
SEM	-	standard error of mean
TEM	-	transmission electron microscopy
UV	-	ultraviolet
V	-	voltage

CHAPTER 1: INTRODUCTION

Hemorrhagic septicemia (HS) is a major disease in cattle and buffaloes which is caused by the infection of *Pasteurella multocida* B:2 (Abubakar and Saad, 2011). In Asia, the serotype of the bacteria responsible for the disease is *P. multocida* B:2 (Tabatabaei et al., 2001). HS when contracted will exhibit symptoms like depression, fever (pyrexia), throat swelling (submandibular oedema), breathing difficulties (dyspnea) and death (Abubakar and Saad, 2011). Transmission occurred from diseased animals or carriers through intranasal and oral routes (Abubakar and Saad, 2011). Invasion of the bacteria through endothelial cells resulted in rapid infiltration of the animals' bloodstream (Galdiero et al., 2001). Vaccine for HS usually was registered prior to rainy season using oil-adjuvant vaccine or alum-precipitated vaccine. Despite both vaccines contained bacterin, only short-termed protection was detected (Othman, 2007). Live attenuated vaccine consisted of live organisms such as the attenuated bacteria with reduced virulence compared to the wild-type (Saad, 2013). GdhA⁻ derivative, known as GDH7, is the attenuated derivative generated through the disruption of the *gdhA* gene by the insertion of kanamycin cassette (Othman, 2007). This resulted in the metabolism interference and hence arrested its pathogenicity. Since currently available vaccines such as alum-precipitated vaccine and oil-adjuvant vaccine were discovered to be less effective, a new alternative is paramount. The aforementioned *gdhA* gene disruption has been found to be a promising manipulation for non-pathogenic *P. multocida* B:2 vaccine development (Othman et al., 2012). However, the invasion mechanism of the derivative bacteria was still unknown. Therefore, in this study, the invasion mechanism of the GDH7 strain of *P. multocida* B:2 will be assessed in order to understand the effectiveness of the vaccine to further develop the vaccine strain as a DNA delivery vehicle.

There were a few problem statements recognized prior to this study. A mutant derived from *P. multocida* B:2, the AroA⁻ attenuated derivative, also known as JRMT12, was non-pathogenic. Tabatabaei et al., (2007) and Dagleish et al., (2007) proved that the JRMT12 vaccine strain when intramuscularly (i.m.) injected had provided strong immunity towards a cattle

population when challenged with the wild-type. This vaccine strain had shown a higher rate of adhesion and invasion towards embryonic bovine lung cells as compared to the parent strain (Othman et al., 2012). In another study, another mutant of *P. multocida* B:2, GDH7 was developed from local isolated strain to be manipulated as a live-attenuated vaccine for HS and this strain showed positive protection efficacy in cattle even during stressful conditions (Rafidah et al., 2013; Othman, 2007). In her study, Rafidah et al., (2013) also proved that through intranasal (i.n.) vaccination of GDH7 strain and allowing the vaccinated cattle to mingle freely with the non-vaccinated cattle in a grazing area. This managed to confer sufficient protection and lasting immunity towards HS in a larger group. This vaccination route (i.n.), was considered as an advantage towards the poor farmers especially in the countries with endemic outbreaks like Malaysia instead of the intramuscular (i.m.) route requiring individual injection of cattle that would be laborious and expensive.

In this study, the adherence and invasive properties of the GDH7 strain are of interest and worthy to be investigated to provide a better insight on the pathogenicity of HS. The investigation of the mechanism in the infection process between the host cell lines and the GDH7 vaccine strain was evaluated based on the methods by Othman et al., (2012). Therefore the objective of this study was to assess the adherence and the invasive properties of the attenuated derivatives of *P. multocida* B:2 and its wild-type strain towards BAEC cell lines.

This study is expected to contribute significant knowledge in the adherence and invasive property of the GDH7 derivative strain compared to other strains of *P. multocida* B:2, to be developed as a vaccine. Hence, through the analysis of this study, this strain can be used as a delivery vehicle of DNA vaccine for HS in the future.

REFERENCE

- Abubakar, M. S., Saad, M. Z. (2011). Clinico-pathological changes in buffalo calves following oral exposure to *Pasteurella multocida* B:2. *Basic and Applied Pathology*, 4: 130-135.
- Ataei, S., Burchmore, R., Hodgson, J. C., Finucane, A., Parton, R. (2009). Identification of immunogenic proteins associated with protection against haemorrhagic septicaemia after vaccination of calves with a live-attenuated *aroA* derivative of *Pasteurella multocida* B:2. *Research in Veterinary Science*, 87: 207-210.
- Badgett, M. R., Auer, A., Carmichael, L. E., Parrish, C. R., Bull, J. J. (2002). Evolutionary Dynamics of Viral Attenuation. *Journal of Virology*, 76(20): 10524-10529.
- Benkirane, A., De Alwis, M. C. L. (2002). Haemorrhagic septicaemia, its significance, prevention and control in Asia. *Veterinary Medicine*, 47(8): 234-240.
- Bosch, M., Tarrago, R., Garrido, E., Campoy, S., Fernandez de Henestrosa, A. R., Perez de Rozas, A. M., Badiola, I., Barbe, J. (2001). Expression of the *Pasteurella multocida ompH* gene is negatively regulated by the Fur protein. *Federal of European Microbiological Societies Microbiology Letters*, 20: 35-40.
- Boyce, J. D., Chung, J. Y., Adler, B. (2000). *Pasteurella multocida* capsule: composition, function, and genetics. *Journal of Biotechnology*, 8: 153-160.
- Dagleish, M. P., Hodgson, J. C., Ataei, S., Finucane, A., Finlayson, J., Sales, J., Parton, R., Coote, J. G. (2007). Safety and Protective Efficacy of Intramuscular Vaccination with a Live *aroA* Derivative of *Pasteurella multocida* B:2 against Experimental Haemorrhagic Septicaemia in Calves. *Infection and Immunity*, 75(12): 5837-5844.
- Fernandez-Rojas, M. A., Vaca, S., Reyes-Lopez, M., Garza, M., Aguilar-Romero, F., Zenteno, E., Soriano-Vargas, E., Negrete-Abascal, E. (2014). Outer membrane vesicles of *Pasteurella multocida* contain virulence factors. *Microbiology Open*, 711-717.
- Galdiero, M., De Martino, L., Pagnini, U., Pisciotta, M. G., Galdiero, E. (2001). Interactions between bovine endothelial cells and *Pasteurella multocida*: association and invasion. *Research in Microbiology*, 152: 57-65.
- Garriado, M. E., Bosch, M., Bigas, A., Badiola, I., Barbe, J., Llagostera, M. (2008). Heterologous protective immunization elicited in mice by *Pasteurella multocida fur ompH*. *International Microbiology*, 11: 17-24.

Hazwani, S. O., Saad, M. Z., Rosfarizan, M., Siti-Khairani, B. (2013). *In vitro* and *in vivo* survivality of *gdhA* derivative *Pasteurella multocida* B:2. *Malaysian Journal of Microbiology*, 10: 63-66.

Horadagoda, N. U., Hodgson, J. C., Moon, G. M., Wijewardana, T. G., Eckersall, P. D. (2001). Role of endotoxin in the pathogenesis of haemorrhagic septicaemia in the buffalo. *Microbial Pathogenesis*, 30: 171-178.

Jamali, H., Rezagholipour, M., Fallah, S., Dadrasnia, A., Chelliah, S., Velappan, R. D., Wei, K. S. C., Ismail, S. (2014). Prevalence, characterization and antibiotic resistance of *Pasteurella multocida* isolated from bovine respiratory infection. *The Veterinary Journal*, 202: 381-383.

Khin, M. W. (2009). Bovine mucosal immune response to intranasal exposure with live *Pasteurella multocida* B:2. PhD Thesis. Faculty of Veterinary Medicine, Universiti Putra Malaysia.

Letourneau, J., Levesque, C., Berthiaume, F., Jacques, M., Mourez, M. (2011). *In Vitro* Assay of Bacterial Adhesion onto Mammalian Epithelial Cells. *Journal of Visualized Experiments*, 1-4.

Othman, S. S. (2007). Construction of an attenuated *Pasteurella multocida* B:2 by mutation in the *gdhA* gene. Master of Science Thesis. Faculty of Veterinary Medicine, Universiti Putra Malaysia.

Othman, S., Parton, R., Coote, J. (2012). Interaction between mammalian cells and *Pasteurella multocida* B:2. Adherence, invasion and intracellular survival. *Microbial Pathogenesis*, 52: 353-358.

Othman, S., Roe, A. J., Parton, R., Coote, J. G. (2013). Use of a Dual Reporter Plasmid to Demonstrate Bactofection with an Attenuated AroA⁻ Derivative of *Pasteurella multocida* B:2. *Plos One*, 8(8): 1-9.

Rafidah, O., Saad, M. Z. (2013). Effect of Dexamethasone on protective efficacy of live *gdhA* derivative *Pasteurella multocida* B:2 vaccine. *Asian Journal of Animal and Veterinary Advances*, 8: 548-554.

Saad, M. Z. (2013). *Haemorrhagic Septicimia of Cattle & Buffaloes in Asia*. Serdang: Universiti Putra Malaysia Press.

Shivachandra, S. B., Kumar A., Mohanty, N. N., Yogisharadhya, R., Chacko, N., Viswas, K. N., Ramakrishnan, M. A. (2014). Homogeneity of *VacJ* outer membrane lipoproteins among *Pasteurella multocida* strains and heterogeneity among members of Pasteurellaceae. *Research in Veterinary Science*, 96: 415-421.

Tabatabaei, M., Liu, Z., Finucane, A., Parton, R., Coote, J. (2002). Protective Immunity Conferred by Attenuated *aroA* Derivative of *Pasteurella multocida* B:2 Strains in a Mouse Model of Haemorrhagic Septicaemia. *Infection and Immunity*, 70(7): 3355-3362.

Tabatabaei, M., Maozzeni Jula, G. R., Jabbari, A. R., Esmailzadeh, M. (2007). Pathogenicity and immunogenicity of native and mutant strains of *Pasteurella multocida*, the causative agents of haemorrhagic septicaemia. *Iranian Journal of Veterinary Research, University of Shiraz*, 8: 40-44.

Townsend, K. M., Boyce, J. D., Chung, J. Y., Frost, A. J., Adler, B. (2001). Genetic Organization of *Pasteurella multocida cap* Loci and Development of a Multiplex Capsular PCR Typing System. *Journal of Clinical Microbiology*, 39(3): 924-929.

Verma, S., Sharma, M., Katoch, S., Verma, L., Kumar, S., Dogra, V., Chahota, R., Dhar, P., Singh, G. (2013). Profiling of virulence associated genes of *Pasteurella multocida* isolated from cattle. *Veterinary Research Communication*, 37: 83-89.