



**UNIVERSITI PUTRA MALAYSIA**

**SYNTHESIS AND *IN VITRO* DELIVERY OF PLASMID-ENCAPSULATED  
CHITOSAN NANOPARTICLES IN HUMAN KIDNEY CANCER CELLS AS  
A VECTOR FOR GENE DELIVERY**

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**FBSB 2015 140**

**Synthesis and *in vitro* delivery of plasmid-encapsulated chitosan nanoparticles in  
human kidney cancer cells as a vector for gene delivery**

By

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Thesis Submitted to the Department of Cell and Molecular Biology, Faculty of  
Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia in Fulfillment of  
the Bachelor of Science (HONS.) Cell and Molecular Biology

**June 2015**

Abstract of thesis presented to the Department of Cell and Molecular Biology in fulfillment of the requirement for the Bachelor of Science (HONS.) Cell and Molecular Biology

**SYNTHESIS AND *IN VITRO* DELIVERY OF PLASMID-ENCAPSULATED CHITOSAN NANOPARTICLES IN HUMAN KIDNEY CANCER CELLS AS A VECTOR FOR GENE DELIVERY**

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**June 2015**

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Chitosan has been extensively adopted in biomedical and pharmaceutical research due to its advantageous properties such as biocompatibility, low-toxicity, and biodegradability. These properties suggest chitosan as a robust material for numerous biomedical applications as gene and drug delivery systems. In this study, the synthesis and *in vitro* delivery of plasmid-encapsulated chitosan nanoparticles in human kidney cancer cells (786-O cell line) were reported. A modified ionic gelation system was utilized to synthesize spherical nanoparticles, and its ability for *in vitro* delivery of a plasmid containing a GFP reporter gene was evaluated. CNPs were characterized using Dynamic

Light Scattering to analyze size distribution and polydispersity index, while Field Emission-Scanning Electron Microscopy was used to analyze surface morphology of the nanoparticles. The 786-O human kidney cancer cell line was then established and treated with CNP-pEGFP formulated at different incubation timepoints. Transfection of 786-O cells were performed by using blank media, CNP and plasmid alone and visualized under both bright-field and fluorescent microscope. FESEM images was found to correlate with DLS data, where the optimum nanoparticle size and PDI value was obtained at a chitosan: TPP ratio of 3:1. Transfection results showed that CNP and cells did not readily fluoresce, and successful GFP expression in cells were only achieved with CNP encapsulation. The results of cell transfection at two different time points showed positive GFP expression, suggesting that CNP was potentially an effective delivery system. However, optimizations are required to further increase efficiency of the gene delivery system for subsequent utilization in biomedical and pharmaceutical fields.

*Keywords:* Nanobiotechnology, CNPs, CNP-pEGFP, gene delivery system

Abstrak tesis yang dikemukakan kepada Jabatan Biology Sel dan Molekul Sebagai memenuhi keperluan untuk ijazah Biologi Sel dan Molekul

**SINTESIS DAN PENYAMPAIAN PLASMID-DIKAPSULKAN  
NANOPARTIKEL KITOSAN *IN VITRO* DALAM SEL-SEL KANSER BUAH  
PINGGANG MANUSIA SEBAGAI VEKTOR UNTUK PENGHANTARAN GEN**

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Kitosan telah banyak diguna pakai dalam penyelidikan bioperubatan dan farmaseutikal kerana ciri berfaedah seperti biocompatibility, ketoksikan rendah, dan biodegradasi.

Ciri-ciri ini mencadangkan kitosan sebagai bahan mantap bagi pelbagai aplikasi

bioperubatan sebagai system penghantaran gen dan ubatan. Dalam kajian ini, sintesis dan penyampaian *in vitro* plasmid-dikapsulkan nanopartikel kitosan dalam sel-sel kanser buah pinggang manusia (sel 786-O) dilaporkan. Sistem penggelan ionic yang diubahsuai telah digunakan untuk mensintesis nanopartikel sfera, dan keupayaannya untuk

penghantaran *in vitro* plasmid yang mengandung gen wartawan GFP telah dinilai. CNPs telah dicirikan dengan menggunakan penyerakan cahaya dinamik untuk menganalisis taburan saiz dan indeks polydispersity, manakala mikroskop elektron pengimbas pancaran medan (field emission scanning electron microscope, FESEM) digunakan untuk menganalisis morfologi permukaan nanopartikel. Pertumbuhan sel kanser buah pinggang manusia (sel 786-O) kemudiannya ditubuhkan dan dirawat dengan CNP-pEGFP yang dirumuskan dengan masa inkubasi yang berbeza. Transfection daripada sel-sel 786-O telah dilakukan dengan menggunakan media kosong, CNP dan plasmid sahaja dan digambarkan di bawah mikroskop medan cahaya and mikroskop pendarfluor. Imej FESEM didapati korelasi dengan data DLS, di mana saiz nanopartikel dan PDI yang optimum telah diperolehi pada nisbah kitosan dan TPP 3:1. Keputusan transfection menunjukkan CNP dan sel-sel tidak berkilau sendiri dan ungkapan GFP dalam sel-sel hanya akan dicapai dengan CNP pengkapsulan. Keputusan transfection sel pada dua titik masa yang berbeza menunjukkan ungkapan GFP positif, menunjukkan bahawa CNP adalah berpotensi dalam sistem penyampaian yang berkesan. Walau bagaimanapun, pengoptimuman dikehendaki meningkatkan lagi kecekapan dalam sistem penyampaian gen untuk penggunaan seterusnya dalam bidang bio-perubatan dan farmaseutikal.

kata kunci: Nanobioteknologi, CNPs, CNP-pEGFP, sistem penyampaian gen

## Approval

This thesis was submitted to the Department of Cell and Molecular Biology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia and has been accepted as fulfillment for the Bachelor of Science (HONS.) Cell and Molecular Biology.

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# DECLARATION

## Declaration by undergraduate student

I hereby confirm that:

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## Acknowledgement

I would like to express my gratitude to my beloved supervisor, Dr. Mas Jaffri Masarudin, whose expertise, understanding and patience that had helped to complete this thesis and research. I appreciate his broad knowledge in nanobiotechnology areas. Under her guidance and motivation, I have been able to successfully complete this research and thesis. Also, thanks to Department of Cell and Molecular Biology, Faculty of Biotechnology and Biomolecular Sciences, UPM that provide this chance for me to do this research study.

Besides that, I wish to express my sincere thanks to all the labmates and coursemates that give advice, guidance and help me throughout this study. Thanks to Miss Ummu Afiqah and Miss Syahzira in assisting and guiding me on how to complete the study patiently. Thanks to Hanis bt. Faudzi and Andrina Tay Chu Huey for all the cooperation in this research. Without their guidance and assistance, the path in completion of this study will not be that smooth and easy.

In addition to that, I am also grateful to my friends especially Christopher Chan Seow Neng and Ng Pui San that had helped me and provide me with the necessary information and support to complete this study. I would also like to express my gratitude and appreciation to my family for the support they provided me through my entire life and especially during my degree life. Without their support and encouragement, I won't be able to make it through my university life.

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in both treatment but higher light intensity was observed in (b).

## LIST OF ABBREVIATIONS AND SYMBOLS

A	-absorbance
B	- Beta
°	-degree
C	-Celcius
CO <sub>2</sub>	-carbon dioxide
CS	-chitosan
CNP/s	-chitosan nanoparticle/s
DLS	-dynamic light scattering
DNA	-deoxyribonucleic acid
EPR	-enhanced permeability effect
FE-SEM	-field emission-scanning electron microscopy
FITC	-fluorescein isothiocyanate
g	-grams
h	-hour(s)
HCl	-hydrochloric acid
l	-litre
m	-metre
mg	-milligram
min	-minute(s)
ml	-mililitre
NaOH	-sodium hydroxide

nm	-nanometer
PBS	-phosphate buffer saline
PDI	-polydispersity index
PEGFP	-plasmid enhanced green fluorescence protein
PSD	-particle size distribution
RNA	- Ribonucleic acid
rpm	-revolutions per minute
RSV	-respiratory syncytial virus
s	-second(s)
siRNA	-small interfering RNA
TPP	-(sodium) triphosphate
UV	-ultraviolet
%	-percentage
μ	-micro

# Chapter 1

## Introduction

### 1.1 Introduction and research background

Over the last decades, radiation, chemotherapy and surgery have become important methods to treat cancer patients. Among these methods, chemotherapy remains the most significant treatment, with anthracyclines being the most potent chemotherapeutic drugs used (Chalasan *et al.*, 2013). The main obstacle faced by current treatment is that the drugs are unable to be delivered efficiently to cancer cells and remain circulated throughout the body. Thus, the treatment often lead to side effects in the patients due to non-specific interaction of the drugs with non-cancerous cells in the body. This has become a constraint for the treatment because the dosage of anthracyclines is limited to a lifetime dose. Due to this reason, the emergence of nanotechnology has arose to enhance the effectiveness of anthracycline drug delivery for the cancer treatment (Masarudin , 2012).

Nanotechnology can be defined as the construction and utilization of devices, materials and systems through the control of matter on the nanometer length scale, which is equal to 1 billionth of a meter. The utilizations of nanotechnology in life sciences is termed as nanobiotechnology (Jain, 2005). Among the application of nanobiotechnology, nanoparticle for drug delivery and biomedical applications are the most significant tools for medical purposes. Since the scale of nanoparticles are relatively small, they can be utilized for the enhanced permeability and retention effect (EPR) of tumor cells to solve

the problem of lack of tumor selectivity for the chemotherapeutic drugs. The nanoparticles are able to exhibit more than 10 to 200 times higher concentrations drugs in tumor cells than that in normal tissues, such as kidney, muscle and heart (Greish, 2010). It means that the nanoparticles help to increase the accumulation of drugs in the tumor tissues.

Basically, nanoparticles can be divided into two categories, which is engineered and nonengineered nanoparticles. Engineered nanoparticles are not naturally existed and invented purposely for application whereas non-engineered nanoparticles are naturally produced (Scalf and West, 2006). Chitosan is one of the appropriate material to be utilized as nanoparticle delivery vector. It is a natural linear polysaccharide encompasses an unbranched chain consisting of  $\beta$ -(1-4)-2-amino-2-deoxy-D-glucopyranose and is formed during the deacetylation of chitin in an alkaline condition. Besides that, this exclusive polysaccharide also been studied because of its biodegradability, biological process, biocompatibility and bioactivity. These characteristics become more popular because do not cause any harmful side-effect to our body as compared with heavy-metal synthesized nanoparticles. In addition, the parameter and payload parameter that we employed will greatly affect the successfulness of the implementation of chitosan nanoparticles. It will be the key for the synthesis of best nanoparticle size. The chitosan nanoparticles can be synthesized by the interaction of amino groups of chitosan backbone with sodium tripolyphosphate (TPP). TPP is a cross-linking agent where this ionic cross-linking with chitosan is beneficial because it is relatively simple and usually made under mild conditions without using organic solvents.

In this thesis, the synthesis and *in vitro* delivery of plasmid-encapsulated chitosan nanoparticles in cells as DNA vector candidates will be studied. The best parameter to

synthesis nanoparticles need to be chosen. Besides that, the ability of delivery of plasmid-encapsulated chitosan nanoparticles in cells as DNA vector candidates is evaluated. Chapter 2 will comprises of the comprehensive literature review of the synthesis of nanoparticles and the delivery system of plasmid-encapsulated nanoparticles in cells. Chapter 3 will includes the methodology for nanoparticles synthesis, the protocol of analysis and delivery of nanoparticles to tumor cells and also the characterization assays. Chapter 4 will covers the results and discussion for the research. Finally, the chapter 5 will conclude the whole research in a nut shell.

## **1.2 Objectives**

The work described in this thesis was designed to achieve the following objectives:

1. To synthesize chitosan nanoparticles by ionic gelation routes using chitosan (CS) and sodium triphosphate, TPP.
2. To encapsulate plasmid DNA into the synthesized chitosan nanoparticles, CNP.
3. To effectively deliver the encapsulated plasmid into human cancer cells.

## References

- Al-Dosari, M. S. & Gao, X. (2009). Nonviral Gene Delivery: Principles, Limitations, and Recent Progress. *American Association of Pharmaceutical Scientist Journal*. 11; 671-681.
- Alyamani, A. & Lemine, O. M. (2012). FE-SEM Characterization of Some Nanomaterial, Scanning Electron Microscopy. 23; 463-472.
- Alyautdin, R. N., Tezikov, E. B., Range, P., Kharkevich, D. A., Begley, D. J. & Kreuter, J. (1998). Significant entry of tubocurarine into the brain of rats by adsorption to polysorbate 80-coated polybutylcyanoacrylate nanoparticles: an in situ brain perfusion study. *J Microencapsul*. 15:67-74.
- Bathool, A., Vishakante, G. D., Khan, M. S., Shivakumar, H. G. (2012). Development and characterization of atorvastatin calcium loaded chitosan nanoparticles for sustain drug delivery. *Adv. Mat. Lett*. 2012, 3(6), 466-470
- Chalasan, S., Prakash, K. V., Pulla, R. P., Tekula, R., Manasa, E., & Umasankar, B. (2013). Development and Validation of Doxorubicin Hcl in Bulk and Its Pharmaceutical Dosage Form by Visible Spectrophotometry. *International Journal of Pharma Sciences*. Vol. 3, No. 3, 216-218.
- Collins, P. L., McIntosh, K. & Chanock, R. M. (1996). *Fields virology*. Philadelphia: Lippincott-Raven. p. 1313.
- Datta, N. (1977). Classification of plasmids as an aid to understanding their epidemiology and evolution. *J. Antimicrob. Chemother*. 3 (Suppl. C), 19-23.
- Davis, M. E. (2002). Non-viral gene delivery systems. *Current Opinion in Biotechnology*. 13: 128-131.
- Davis, S. S. (1997). Biomedical applications of nanotechnology—implications for drug targeting and gene therapy. *Trends Biotechnol*. 15, 217-224.

- Dekhtyar, M., Morin, A. & Sakanyan, V. (2008). Triad pattern algorithm for predicting strong promoter candidates in bacterial genomes. *BMC Bioinformatics* 9, 233.
- Ebersbach, G. & Gerdes, K. (2005). Plasmid segregation mechanisms. *Annu Rev. Genet.* 39, 453–479.
- Ellenberg, J., Lippincott-Schwartz, J., & Presley, J. F. (1998). Two-color green fluorescent protein time-lapse imaging. *Biotechniques* 25, 838–842, 844–846.
- Erbacher, P., Zou, S., Bettinger, T., Ştepan, A. M. & Remy, J. S. (1998). Chitosan-based vector/DNA complexes for gene delivery: biophysical characteristics and transfection ability, *Pharm. Res.* 15 1332-1339.
- Escors, D. & Brecht, K. (2010). Lentiviral vectors in gene therapy: their current status and future potential. *Archivum Immunologiae et Therapia Experimentalis*;58;107–119.
- Evdokimov, Yu.M., Zakharov, M.A., & Skuridin, S.G. (2006) *Vestnik Ross. Akad. Nauk.* vol. 76, pp. 112–120.
- Fakruddin, M., Hossain, Z. & Afroz, H. (2012). Prospects and applications of nanobiotechnology: a medical perspective. *Journal of Nanobiotechnology* 10, 31.
- Feng, S. S., Mu, L. & Win, K. Y. (2004). Nanoparticles of biodegradable polymers for clinical administration of paclitaxel. *Curr Med Chem.* 11, 413–424.
- Gan, Q., Wang, T., Cochrane, C. & McCarron, P. (2005). Modulation of surface charge, particle size and morphological properties of chitosan–TPP nanoparticles intended for gene delivery. *Colloids and Surfaces B: Biointerfaces* 44, 65–73.
- Gao, X., Kim, K. S. & Liu, D. (2007). Nonviral Gene Delivery: What We Know and What Is Next. *American Association of Pharmaceutical Scientist Journal.* 9: 92-104.
- Garcia-Garcia, E., Gil, S., Andrieux, K., Desmaître, D., Nicolas, V., Taran, F., Georgin, D., Andrieux, J. P., Roux, F. & Couvreur P. (2005). A relevant *in vitro* rat model for the evaluation of blood–brain barrier translocation of nanoparticles. *Cell Mol Life Sci.* 62(12). 1400–1408.

Gardlík, R., Pálffy, R., Hodosy, J., Lukács, J., Turňa, J. & Celec, P. (2005). Vectors and delivery systems in gene therapy. *Med Sci Monit.* 11(4). RA 110-121.

Ghosh, S. K., Hajra, S., Paek, A. & Jayaram, M. (2006). Mechanisms for chromosome and plasmid segregation. *Annu. Rev. Biochem.* 75, 211–241.

Gowda, R., Jones, N. R., Banerjee, S. & Robertson, G. P. (2013). Use of Nanotechnology to Develop Multi-Drug Inhibitors For Cancer Therapy. *J Nanomed Nanotechnol* 4, 184.

Greish, K. (2010). Enhanced permeability and retention (EPR) effect for anticancer nanomedicine drug targeting. , *Cancer Nanotechnology, Methods in Molecular Biology* 624, 25-38

Gurunathan, S., Klinman, D. M. & Seder, R. A. (2000). DNA vaccines: immunology, application, and optimization. *Annu Rev Immunol.*18:927–74.

Handy, R. D., von der Kammer, F., Lead, J. R., Hassello v, M., Owen, R. & Crane, M. (2008). The ecotoxicology and chemistry of manufactured nanoparticles. *Ecotoxicology* 17, 287–314.

He, C. H., Tabata, Y. & Gao, J.Q. (2010). Non-viral gene delivery carrier and its three-dimensional transfection system. *International Journal of Pharmaceutics.* 386; 232–242.

Hu, L., Tang, X. & Cui, F. (2004) Solid lipid nanoparticles (SLNs) to improve oral bioavailability of poorly soluble drugs. *J Pharm Pharmacol* 56, 1527–1535.

Illum, L. (1998) Chitosan and its use as a pharmaceutical excipient. *Pharm Res.* 15:1326–31.

Ishii, T., Sato, T. & Okahata, Y. (2001). Mechanism of cell transfection with plasmid/chitosan complexes. *Biochim Biophys Acta.* 1514:51–64.

Jain, K. K. (2005). The role of nanobiotechnology in drug discovery. *DDT*, Vol 10 (21) 1435-1442.

Katare, D.P. & Aeri, V. (2010). Progress in gene therapy: A Review. *I.J.T.P.R.* Vol. 1, No. 2, pp. 33-41

Khatri, K., Amit, K., Goyal, Prem, N., Gupta, Mishra, N. & Suresh, P. Vyas. (2008). *International Journal of Pharmaceutics*. 354, 235–241.

Klaine, S. J., Alvarez, P. J., Batley, G. E., Fernandes, T.F., Handy, R.D., Lyon, D.Y., Mahendra, S., McLaughlin, M.J. & Lead, J.R. (2008). Nanomaterials in the environment: behavior, fate, bioavailability, and effects. *Environ Toxicol Chem*. 31(12), 2893.

Kline, B. C. & Palchaudhuri, S. (1980). Genetic studies of F plasmid maintenance genes. *Plasmid* 4, 281–291.

Komoda, T. & Saito, S. (1972). Experimental resolution limit in the secondary electron mode for a field emission source scanning electron microscope. *Scanning Electron Microscopy*. 129-136 .

LaVan, D. A., Lynn, D. M. & Langer, R. (2002). Timeline: Moving Smaller in drug discovery and delivery. *Nat Rev Drug Discov*. 1,77–84.

Lederberg, J. (1952). Cell genetics and hereditary symbiosis. *Physiological reviews*. 32(4):403-30.

Liu, H. & Gao, C. (2009). Preparation and properties of ionically cross-linked chitosan nanoparticles. *Polym. Adv. Technol*. 20, 613–619.

Masarudin, M. J. (2012). Chitosan nanoparticles as a delivery vehicle for [14c]-doxorubicin and the formaldehyde releasing prodrug an-250. Ph.D. thesis, La Trobe University.

Mazzarino, L., Travelet, C., Ortega-Murillo, S., Otsuka, I., Pignot-Paintrand, I., Lemos-Senna, E. & Borsali, R. (2012). Elaboration of chitosan-coated nanoparticles loaded with curcumin for mucoadhesive applications. *Journal of Colloid and Interface Science*. 370(1), 58-66.

Meyer, W. V., Smart, A. E., Wegdam, G. H., & Brown, R.G.W. (2006). Photon correlation and scattering: introduction to the feature issue. *Appl. Opt*. 45. pp 2149.

Mhashilkar, A., Chada, S., Roth, J. A. & Ramesh, R. (2001). Gene therapy; Therapeutic approaches and implications. *Biotechnology Advances*; 19; 279-297.

Miyazaki, M., Obata, Y., Abe, K., Furusu, A., Koji, T., Tabata, Y. & Kohno, S. (2006). Technological Advances in Peritoneal Dialysis Research: Gene Transfer Using Nonviral Delivery Systems. *Peritoneal Dialysis International*; 26; 633–640.

Mrsny, R. (2005). In Amiji MM (ed). Tissue-and Cell-Specific Targeting for the Delivery of Genetic Information. *Polimeric Gene Delivery*. CRC PRESS, USA. pp: 1-30.

Mumper, R.J., Wang, J., Claspell, J. M. & Rolland, A. P., (1995) Novel polymeric condensing carriers for gene delivery. *Proc Intern Symp Control Rel Bioact Mater* 22:178–179

Nayerossadat, N., Maedeh, T., & Ali, P. A. (2012). Viral and nonviral delivery systems for gene delivery. *Advanced Biomedical Research*, 1, 27.

Noskin, V.A. (1983). Gatchina: Leningrad Inst. of Nuclear Physics.

Park, I. K. (2002) Ph.D. thesis, Seoul National University.

Paulsson, J. & Chattoraj, D.K. (2006). Origin inactivation in bacterial DNA replication control. *Mol. Microbiol.* 61, 9–15.

Pecora, R. (1985). Dynamic Light Scattering: Applications of Photon Correlation Spectroscopy. New York London: Plenum.

Pillai, O. & Panchagnula, R. (2001). Polymers in drug delivery. *Current Opinion in Chemical Biology*; 5: 447–451.

Raghuwanshi, D., Mishra, V., Das, D., Kaur, K., & Suresh, M. R. (2012). Dendritic Cell Targeted Chitosan Nanoparticles for Nasal DNA Immunization against SARS CoV Nucleocapsid Protein. *Mol Pharm.* 2012 April 2; 9(4): 946–956.

Scalf, J. & West, P. (2006). Part I: Introduction to Nanoparticle Characterization with AFM. *Pacific Nanotechnology*.

Shimizu, R., Kuroda, K., Suzuki, T., Nakamura, S., Suganuma, T. & Hashimoto, H. (1973). Field Emission scanning electron Microscope with parallel plate gun electrodes. *Scanning Electron Microscopy*. 74-80

Shimomura, O., Johnson, F.H. & Saiga, Y. (1962) *J. Cell. Comp. Physiol.* 59 223-240.

Somiari, S., Glasspool-Malone, J., Drabick, J. J., Gilbert, R. A., Heller, R., Jaroszeski, M. J., & Malone, R. W. (2000). Theory and in vivo Application of Electroporative Gene Delivery. *Molecular Therapy*: 2; 178-187.

Son, S., Chae, S. Y., Choi, C., Kim, M., Ngugen, V. G., Jang, M. & Nah, J. (2004). Preparation of a Hydrophobized Chitosan Oligosaccharide for Application as an Efficient Gene Carrier. *Macromolecular Research*. 12, (6), 573-580.

Sun, Y., Zhang, S., Peng, X., Gong, Z., Li, X., Yuan, Z., Li, Y., Zhang, D. & Peng, Y. (2012). Preparation, characterization and transfection efficacy of chitosan nanoparticles containing the intestinal trefoil factor gene. *Mol Biol Rep*. 39, 945–952.

Tan, WB., Jiang, S. & Zhang, Y. (2007). Quantum-dot based nanoparticles for targeted silencing of HER2/neu gene via RNA interference. *Biomaterials*, 28(8):1565-1571.

Vasir, J. K., Reddy, M. K. & Labhasetwar, V. (2005). *Curr Nanosci* 1:47.

Velten, J. & Schell, J. (1985). Selection-expression plasmid vectors for use in genetic transformation of higher plants. *Nucleic Acids Res*. 13, 6981–6998.

Wilfinger, W. W., Mackey, K. & Chomczynski, P. (1997). Effect of pH and Ionic Strength on the Spectrophotometric Assessment of Nucleic Acid Purity. *BioTechniques*. 22, 474-481.

Williams, D.B. & Carter, C.B. (2009). Transmission Electron Microscopy: A Textbook for Materials Science. Chapter 1. 3-22

Yao, H. & Kimura, K. (2007). Field Emission Scanning Electron Microscopy for Structural Characterization of 3D Gold Nanoparticle Superlattices. *Modern Research and Educational Topics in Microscopy*.

Yi, Y., Noh, M. J. & Lee, K. H. (2011). Current advances in retroviral gene therapy. *Curr Gene Ther*. 11(3) 218–28.

Yuan, Y. J., Tan, J. Y., Wang, Y. F., Qian, C. W. & Zhang, M. Y. (2009). Chitosan nanoparticles as non-viral gene delivery vehicles based on atomic force microscopy study. *Acta Biochim Biophys Sin*: 41(6). 515–526.

Zhang, J., Yang, P. L. & Gray, N. S. (2009). Targeting cancer with small molecule kinase inhibitors. *Nat Rev Cancer* 9: 28-39.

Zhang, Z., Fang, X., Hao, J., Li, Y. & Sha, X. (2011). Triolein-based polycation lipid nanocarrier for efficient gene delivery: characteristics and mechanism. *International Journal of Nanomedicine*. 6, 2235–2244.

Zverev, V. V., Kuzmin, N. P., Zuyeva, L. A., Burova, E. I., Alexandrov, A. A. & Khmel, I. A. (1984). Regions of homology in small colicinogenic plasmids. *Plasmid* 12, 203–205.

