



***EPIDEMIOLOGY, ANTIBIOGRAMS AND MULTILOCUS SEQUENCE
TYPING OF *Salmonella enterica* AMONG Ayam Kampong
(*Gallus gallus domesticus* Linnaeus) FROM SOUTH-CENTRAL
PENINSULAR MALAYSIA***

SALEH MOHAMMED JAJERE

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By

SALEH MOHAMMED JAJERE

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

January 2020

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DEDICATION

This work is dedicated to my late father, Alhaji Mohammed Ahmed Jajere of blessed memory, and to my mother, Hajiya Rukayyatu Abdullahi.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

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January 2020

Chairman : Professor Latiffah binti Hassan, DVM PhD
Faculty : Veterinary Medicine

Salmonella enterica represents an important foodborne pathogen worldwide. In Malaysia, several large foodborne disease outbreaks involving human fatalities have been linked to eating chicken and its products contaminated with *Salmonella*. Village chickens are presumed to be raised in a more 'organic' environment and have not been studied for the prevalence of *Salmonella*. With increasing consumer awareness on food safety issues related to chemical residues and antibiotics used in producing commercial chickens, the demand for village chickens in Malaysia and other parts of the world has increased. Consumers are willing to pay higher price for products raised in organic setups. However, the epidemiology of *Salmonella* infection amongst village chickens in Malaysia remains largely unknown. Thus, the current study investigates the epidemiology, antibiograms and genetic diversity of *Salmonella enterica* amongst village chickens from the South-central Peninsular Malaysia.

Thirty-five village flocks were sampled from Selangor ($n=19$), Melaka ($n=10$), Johor ($n=4$), and Negeri Sembilan ($n=2$). In total, 1042 samples were collected; these included cloacal swabs ($n=675$), eggs ($n=62$), pooled drinking water ($n=175$), pooled feeds ($n=70$), and pooled flies ($n=60$). Isolation of *Salmonella* from these samples was carried out according to the protocols and recommendations of the World Organization for Animal Health (OIE) terrestrial manual. The prevalence of *Salmonella* at an individual bird-level was 2.5% (17/675, 95% CI: 1.6–4.0). All eggs screened were negative. For environmental samples, *Salmonella* was detected in 5.14% (9/175), 7.14% (5/70), and 5.0% (3/60) for water, feed, and flies, respectively. Thirty-four isolates and eight *Salmonella* serotypes were identified. *S. Weltevreden* (20.6%) was the most common, followed by *S. Typhimurium* and *S. Agona* (17.6%),

S. Albany and *S. Enteritidis* (8.8%), *S. Molade* (5.9%), *S. Corvallis* and *S. Schleissheim* (2.9%), and others grouped as *Salmonella* spp. (11.8%). Multivariable logistic regression models revealed that *Salmonella* positivity among flocks could be strongly predicted by storage of feeds (uncovered feeds; OR=10.38; 95% CI: 1.25–86.39; $p=0.030$) and uncovered water tanks (uncovered tank; OR=6.43; 95% CI: 1.02–40.60; $p=0.048$).

Among the isolates, 26.5% (n=9) were susceptible to all antibiotics, while 73.5% (n=25) were resistant to at least one antibiotic tested. Multidrug resistance was displayed by 8 isolates (23.5%). Ciprofloxacin (100%), gentamicin (97.1%), norfloxacin (97.1%), cefotaxime (97.1%) and ceftiofur (97.1%) were very effective against most isolates. The highest level of resistance was observed for tetracycline (35.3%) and streptomycin (35.3%). Eight isolates (23.5%) were MDR to two or more antibiotic agents belonging to ≥ 3 antimicrobial classes. Colistin resistance (Minimum inhibitory concentrations: 4 - 16 mg/L) was detected among 5 (14.7%) isolates comprising *S. Weltevreden*, *S. Albany*, *S. Typhimurium* and *Salmonella* spp. Representative *Salmonella* isolates (n=15) were examined by multilocus sequence typing (MLST) using seven housekeeping genes from the MLST online database. The *Salmonella enterica* serovars were resolved into 7 sequence types (STs). The ST13 (n=3) and ST11 (n=3) were the predominant STs respectively found in serovars Agona and Enteritidis, followed by ST34 (n=2) and ST36 (n=2), ST365 (n=2), ST1541 (n=2) and ST19 (n=1) respectively found in serovars Typhimurium, Weltevreden, Corvallis and Typhimurium. It was found that, with the exception of serotype Typhimurium, the MLST indicated a strong correlation between the STs and serovars. Each serovar of the Enteritidis, Agona, Weltevreden and Corvallis strains was represented unique STs.

In conclusion, it was found that the prevalence of *Salmonella* in village chickens in the study area was lower than that reported from commercial chickens in Malaysia. Uncovered feed and water storage were strongly associated with *Salmonella* positivity among the village chicken flocks. The present study also demonstrated the contamination of local village chickens with MDR non-typhoidal *Salmonella* strains, some of which were resistant to multiple classes of antimicrobials. Generally, the level of resistance was meaningfully lower than those reported from commercial chicken carcasses and its products in Malaysia and elsewhere. However, despite the minimum application of antibiotics in the free-range chicken production and lack of antibiotic growth promotion use, the resistance to colistin was observed. The exposure to natural environment such as wild birds, proximity to commercial chicken farms as well as contaminated water and soils may play complex role in determining antibiotic resistance. Molecularly, all sequence types identified among the isolates tested in the present study has been documented in Malaysia previously, and were also reported in many Asian countries including China, Singapore, Indonesia, Thailand, Vietnam and Myanmar (ST11). ST365 was also reported in Singapore. This finding highlights that these village chicken *Salmonella* strains are not unique and have been circulating in the Asian region in multiple animal species and humans.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**EPIDEMIOLOGI, ANTIBIOGRAM DAN PENJENISAN MULTILOKUS
JUJUKAN *Salmonella enterica* DI KALANGAN AYAM KAMPUNG (*Gallus
gallus domesticus* Linnaeus) DARI BAHAGIAN TENGAH-SELATAN
SEMENANJUNG MALAYSIA**

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Salmonella enterica merupakan patogen makanan yang penting di seluruh dunia. Di Malaysia, beberapa wabak penyakit yang besar mengakibatkan kematian manusia telah dikaitkan dengan daging ayam serta produknya yang telah dicemari dengan *Salmonella*. Ayam kampung dianggap dipelihara dan dibesarkan dalam persekitaran yang lebih 'organik' dan belum dikaji prevalens (kelaziman) terhadap *Salmonella*. Dengan meningkatnya kesedaran pengguna mengenai isu keselamatan makanan yang berkaitan dengan bahan kimia dan penggunaan antibiotik dalam penghasilan ayam komersil, permintaan ayam kampung di Malaysia dan negara-negara lain di dunia telah meningkat. Pengguna sanggup membayar dengan harga yang lebih tinggi untuk produk yang dikeluarkan dalam persekitaran organik. Walaubagaimanapun, epidemiologi jangkitan *Salmonella* di kalangan ayam kampung di Malaysia masih tidak di ketahui. Oleh itu, kajian ini mengkaji epidemiologi, antibiogram dan kepelbagaian genetik *Samonella enterica* di kalangan ayam kampung di selatan-tengah semenanjung Malaysia.

Tiga puluh lima kawanan (*flocks*) ayam kampung dari Selangor (n=19), Melaka (n=10), Johor (n=4), dan Negeri Sembilan (n=2) telah di ambil sampelnya. Secara keseluruhan, 1042 sampel telah dikumpulkan; ini termasuk swab kloaka (n=675), telur (n=62), air minuman yang di satukan (*pooled*) (n=175), makanan yang dikumpulkan (n=70) dan lalat yang dikumpulkan (n=60). Pemencilan *Salmonella* dari sampel-sampel ini dilakukan mengikut protokol dan cadangan dari manual Pertubuhan Kesihatan Haiwan Sedunia (OIE). Prevalens *Salmonella* pada setiap tahap-burung (bird-level) ialah 2.5% (17/675, 95% CI: 1.6-4.0). Semua telur yang telah diperiksa adalah negatif. Untuk sampel persekitaran, *Salmonella* dikesan sebanyak 5.14% (9/175), 7.14% (5/70), dan 5.0% (3/60) masing-masing pada air,

makanan, dan lalat. Tiga puluh empat pencilan dan lapan serotip *Salmonella* telah dikenalpasti. *S. Weltevreden* (20.6%) adalah yang paling banyak, diikuti oleh *S. Typhimurium* dan *S. Agona* (17.6%), *S. Albany* dan *S. Enteritidis* (8.8%), *S. Molade* (5.9%), *S. Corvallis* dan *S. Schleissheim* (2.9%) dan selebihnya sebagai *Salmonella* spp. (11.8%). Model regresi logistik multipemboleh ubah menunjukkan bahawa *Salmonella* adalah positif dalam kawan dapat diramalkan dengan simpanan makanan (makanan yang terdedah; OR = 10.38; 95% CI: 1.25-86.39; p = 0.030) dan tangki air yang terbuka; OR = 6.43; 95% CI: 1.02-40.60; p = 0.048).

Di antara pencilan, 26.5% (n=9) adalah rentan terhadap semua antibiotik, manakala 73.5% (n=25) adalah rintang terhadap sekurang-kurangnya satu antibiotik yang diuji. Kerintangan dadah-pelbagai (MDR) ditunjukkan oleh 8 pencilan (23.5%). Kebanyakan pencilan rentan terhadap *ciprofloxacin* (100%), *gentamicin* (97.1%), *norfloxacin* (97.1%), *cefotaxime* (97.1%) dan *ceftiofur* (97.1%). Tahap rintangan tertinggi dilihat pada *tetracycline* (35.3%) dan *streptomycin* (35.3%). Lapan pencilan (23.5%) adalah MDR kepada dua atau lebih agen antibiotik yang tergolong dalam kelas antimikrob ≥ 3 . Rintang kolistin (kepekatan perencatan minimum: 4-16 mg / L) dikesan dari 5 pencilan (14.7%) yang terdiri dari *S. Weltevreden*, *S. Albany*, *S. Typhimurium* dan *Salmonella* spp.

Perwakilan pencilan *Salmonella* (n=15) telah diperiksa menggunakan penjenisan jujukan multilokus (MLST) dengan menggunakan tujuh gen jaga selia (*housekeeping gene*) dari pangkalan data dalam talian MLST. *Salmonella enterica* serovar telah dirumuskan menjadi 7 jenis jujukan (ST). ST13 (n=3) dan ST11 (n=3) adalah ST yang pradominan masing-masing ditemui dalam serovar *Agona* dan *Enteritidis*, diikuti oleh ST34 (n=2) dan ST36 (n=2), ST365 (n=2), ST1541 (n=2) dan ST19 (n=1) masing-masing ditemui dalam serovar *Typhimurium*, *Weltevreden*, *Corvallis* dan *Typhimurium*. Didapati bahawa, kecuali serotype *Typhimurium*, MLST menunjukkan korelasi yang kuat antara STs dan serovars. Setiap strain serovar daripada *Enteritidis*, *Agona*, *Weltevreden* dan *Corvallis* telah diwakili STs yang unik.

Sebagai kesimpulan, didapati bahawa prevalens *Salmonella* dalam kalangan ayam kampung di kawasan kajian lebih rendah berbanding yang dilaporkan dalam kalangan ayam komersil di Malaysia. Makanan dan air takungan yang tidak ditutup paling berkait dengan positifnya *Salmonella* dalam kawan ayam kampung. Kajian ini juga menunjukkan bahawa kontaminasi (pencemaran) ayam kampung tempatan dengan strain MDR bukan tifoid, sebahagiannya adalah rintang terhadap pelbagai kelas antimikrobia. Secara amnya, tahap kerintangan adalah lebih rendah berbanding yang dilaporkan daripada bangkai (*carcasses*) ayam komersil serta produknya yang terdapat di Malaysia dan di tempat lain. Walaubagaimanapun, aplikasi minimum antibiotik terhadap pengeluaran ayam *free-range* dan kurangnya penggunaan promosi pertumbuhan antibiotik, kerintangan terhadap kolistin diperhatikan. Pendedahan terhadap persekitaran semulajadi seperti burung liar, berdekatan dengan ladang ayam komersil serta air dan tanah yang tercemar mungkin memainkan peranan yang kompleks dalam menentukan rintangan antibiotik. Secara

molekular, semua jenis jujukan yang dikenal pasti daripada pencilan yang diuji dalam kajian ini telah di dokumentasikan di Malaysia sebelum ini, dan juga dilaporkan di banyak negara Asia termasuk China, Singapura, Indonesia, Thailand, Vietnam dan Myanmar (ST11). ST365 juga dilaporkan di Singapura. Penemuan ini memperlihatkan bahawa strain *Salmonella* ayam kampung ini tidak unik dan telah beredar (*circulating*) ke rantau Asia dan melibatkan pelbagai spesies haiwan dan manusia.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

ACSSuT	Ampicillin, Chloramphenicol, Streptomycin, Sulfamethoxazole, Tetracycline
AFLP	Amplified fragment length polymorphism
ASEAN	Association of Southeast Asian Nations
ATCC	American type culture collection
Amc	Amoxicillin-clavulanate
Amp	Ampicillin
BAM	Bacteriological analytical manual
BGA	Brilliant green agar
BPW	Buffered peptone water
CDC	Centres for disease control and prevention
cgMLST	Core genome multilocus sequence typing
CI	Confidence interval
CLSI	Clinical and laboratory standards institute
Cn	Gentamicin
Ct	Chloramphenicol
Ctx	Cefotaxime
DNA	Deoxyribonucleic acid
DVS	Department of veterinary services
ELISA	Enzyme linked immunosorbent assay
ESBLs	Extended- β -lactamases
EUCAST	European committee on antimicrobial susceptibility testing
F	Nitrofurantoin
FAO	Food and agriculture organization of the United Nations
FDA	Food and drug administration
g	Gram
gDNA	Genomic deoxyribonucleic acid

IBM	International business machines
IMR	Institute of medical research
K	Kanamycin
mm	Millimetres
μM	Micromolar
μL	Microliters
mg	Milligram
mL	Millilitres
MAR	Multiple antibiotic resistance
MCR-1	Plasmid-mediated colistin resistance gene
MDR	Multidrug resistant
MEGA	Molecular evolutionary genetics analysis
MIC	Minimum inhibitory concentration
MLEE	Multilocus enzyme electrophoresis
MLST	Multilocus sequence typing
MLVA	Multilocus variable number of tandem repeats analysis
MLVF	Multilocus variable number of tandem repeats fingerprinting
MS	Microsoft
Na	Nalidixic acid
NA	Nutrient agar
NJ	Neighbour joining
NTS	Non-typhoidal <i>Salmonella</i>
OIE	World Organization for animal health
OR	Odds ratio
PCR	Polymerase chain reaction
PFGE	Pulsed field gel electrophoresis
RAPD	Randomly amplified polymorphic deoxyribonucleic acid
rpm	Revolutions per minutes

R/I/S	Resistant, Intermediate, Susceptible
RNA	Ribonucleic acid
RVB	Rappaport vassiliadis broth
S	Streptomycin
S3	Sulfonamides
SE	<i>Salmonella</i> Enteritidis
SEAs	Southeast Asian countries
SG	<i>Salmonella</i> Gallinarum
SLVs	Single locus variants
SNP	Single nucleotide polymorphism
SP	<i>Salmonella</i> Pullorum
SPI	<i>Salmonella</i> Pathogenicity Island
SPSS	Statistical package for social sciences
ST	<i>Salmonella</i> Typhimurium
ST	Sequence Type
TBE	Tris-Borate ethylenediaminetetraacetic acid
Te	Tetracycline
TLVs	Triple locus variants
tRNA	Transfer ribonucleic acid
TSB	Tryptic soy broth
UPM	Universiti Putra Malaysia
VNTR	Variable number of tandem repeats
VRI	Veterinary research institute
W	Trimethoprim
WGS	Whole genome sequencing
XLD	Xylose lysine desoxycholate

CHAPTER 1

INTRODUCTION

1.1 Study background

Village chickens or “*Ayam Kampung*” are hybrids from the natural crossbreeding between the Malay fowl, jungle fowl and mixed exotic breeds brought in by the Europeans (Azahan & Zahari, 1983; Azahan, 1994). They are common in the Southeast Asia particularly Malaysia and Indonesia, and are traditionally raised as backyard chickens where they are let free to roam and scavenge for food and sometimes fed with household leftovers or scraps to supplement their dietary requirements (Azahan et al., 1980; Azahan & Zahari, 1983; Azahan, 1994; Padhi, 2016). Recently, the emerging food safety and animal welfare issues such as antibiotic and chemical drug residues in food animals as well as increased development of antibiotic resistance by foodborne pathogens particularly common in commercial poultry production, has resulted in increased demand for organically grown food animals such as village chickens because consumers believed they are drug - or antibiotic - free, wholesome and medicinal (Hassan et al., 2005; Miao et al., 2005; Rahman & Haziqah, 2015; Suhaila et al., 2015).

Salmonella enterica with more than 2,500 different serotypes identified, has been recognised as a major bacterial foodborne pathogen for humans, and until today has remained a leading cause of foodborne illnesses worldwide (Egualde, 2018; Nair et al., 2018; Shang et al., 2018; Zhang et al., 2018). For instance, in the United States, it is responsible for high mortality rates and increasing cost associated with the treatment (Batz et al., 2012). The World Health Organization (WHO) and the Food Agriculture Organization (FAO) have declared *Salmonella* since 1950s, as the most common and important zoonosis; and this has led to its subsequent inclusion in the terrestrial animal health code of the WHO (Moultotou et al., 2017). Salmonellosis is caused by two species of *Salmonella* namely *S. enterica* and *S. bongori*, the former being the most important aetiological agent for human systemic and diarrheal infections (Saravanan et al., 2015). The *S. enterica* serovars, *S. Typhimurium* (ST), *S. Enteritidis* (SE) and *S. Heidelberg* are the epidemiologically important serotypes among the non – typhoidal *Salmonella* (NTS) that can be found in both domestic and wild animals including birds worldwide (Najwa et al., 2015; Prestinaci et al., 2015; Saravanan et al., 2015). Human infections with NTS is frequently associated with consumption of *Salmonella* contaminated foods or food-producing animals. Although, non-typhoidal *Salmonella* has been isolated from different food-producing animals and sources (Heredia & García, 2018), poultry and poultry products have particularly served as major reservoirs with contaminated poultry products as major vehicles of human infections along the *farm-to-fork* continuum (Andino & Hanning, 2015; Andoh et al., 2016; Foley et al., 2008).

Recently, the similarity of the *Salmonella* isolates including multidrug resistant ST recovered from food-producing animals, with those responsible for human salmonellosis suggests a possible transmission from food animals to humans (Hong et al., 2018; Kagambèga et al., 2018; Li et al., 2013). Poultry and poultry products such as eggs, food products containing eggs, undercooked poultry meat are the most commonly implicated sources and primary vehicles of infection caused by ST and SE (Abdullah et al., 2010; Afshari et al., 2018; Ong et al., 2014; Saravanan et al., 2015; Thung et al., 2018, 2016) . While *Salmonella* and other related foodborne infections in poultry is often asymptomatic, its colonization in chickens may present a significant health risk to humans as *Salmonella* may transmit to humans from the infected birds through ingestion of undercooked retail meat contaminated with *Salmonella* (Thakur et al., 2013).

In chicken farms, several factors play critical role in the spread and perpetuation of *Salmonella*. Of these factors, feed and water contaminated with *Salmonella* remain important sources of infection (Frederick & Huda, 2011). Additionally, critical factors commonly identified as sources of *Salmonella* contamination in farms included the contaminated drinkers, feeders, litters, feedstuffs, air inside chickens houses, soil, bedding and faecal matter (Hoelzer et al., 2011; Hoover et al., 1997; Rodriguez et al., 2006). Transmissions of *Salmonella* among birds are horizontal or vertical. Horizontal transmission occurs through the intermittent shedding of the *Salmonella* by asymptomatic birds thereby contaminating the dust, feathers of other birds, soil, water, litters etc. In contrast, the vertical involves systemic *Salmonella* infection of the parent laying birds in the hatchery and this is passed onto chicks through the infected reproductive tissues of the parent birds (Moultotou et al., 2017).

Antimicrobial resistance by bacterial foodborne pathogens like *Salmonella* has become a major public health threat worldwide due to the potential of these resistant agents to transmit to humans via the food chain (Brown et al., 2017; Nair et al., 2018; Xiong et al., 2018). The emergence of this resistance has been linked with the routine use of antibiotics in animals for prevention and treatment of diseases as well as a growth promoter (Nair et al., 2018; Tang et al., 2017). Furthermore, studies from different countries have demonstrated antibiotic resistance including multidrug resistant (MDR) profiles of *Salmonella* isolates recovered from humans and food-producing animals (Cameron-Veas et al., 2018; Chatham-Stephens et al., 2019; Dang-Xuan et al., 2019; Sharma et al., 2019; Yang et al., 2019). In Malaysia, MDR *Salmonella* has been reported from retail chicken carcasses, beef, vegetables and other animal derived food products (Salleh et al., 2003; Khoo et al., 2015; Kuan et al., 2017; Thung et al., 2018; Yoke-Kqueen et al., 2008). The reported prevalence of *Salmonella* in Malaysia ranged from 10.5% to 44.4% as reported by different authors from retail markets, processing plants, wet markets and other ready-to-eat-foods. These studies highlighted *Salmonella* contamination rates in raw fresh fruits and vegetables (Salleh et al., 2003), poultry litters (Rusul et al., 1996), poultry farms (Rusul et al., 1996), chicken meat portions (Arumugaswamy et al., 1995), liver and gizzard (Arumugaswamy et al., 1995), ready-to-eat cooked meat, prawn and oriental shrimps (Arumugaswamy et al., 1995). Because village chickens are often allowed to roam freely in the house compound, they are at high risks of coming in contact

with pathogens in the environment. However, no research or information could be found regarding the microbiological quality of *Ayam Kampung* and its products in Malaysia. Therefore, exploring the on-farm transmission dynamics of *Salmonella* serotypes and their antibiotic resistance profiles in village chickens will allow the relevant Malaysian government agencies to better implement appropriate control measures and strategies in order to mitigate its prevalence and the threat it posed to human health. Information on antibiotic resistance phenotypes and genotypes of *Salmonella* from food animals and their products is necessary to combat the spread of resistance and public health threat posed by antibiotic resistant *Salmonella*. This information will aid in exploring the epidemiology of antibiotic resistance, identify or trace new emerging resistant pathogens and assist in prudent use of clinically available (last resort) antimicrobials in both humans and animals.

1.2 Problem statement

Malaysia is considered as one of the countries with highest chicken consumers, and according to a recent report by the department of statistics, Malaysia, among the livestock and products, chicken meat had the highest per capita consumption (52kg) per year followed by chicken/duck eggs (22.2kg/year) and pork (16.3kg/year) (Department of Statistics Malaysia, 2017). Chicken meat and eggs are the most popular and cheapest sources of protein in the country, mainly because there exist no restrictions or any religious prohibition of its consumption. Malaysia is self sufficient in terms of egg and even exports appreciable amounts to a number of neighbouring countries (Department of Veterinary Services, 2012). Chicken's meat and eggs are considered the major vehicles harbouring and transmitting foodborne *Salmonella* to humans via the food chain. In Malaysia, several large foodborne disease outbreaks involving human fatalities have been reported in recent time. A substantial proportion of the outbreaks have been linked to eating foods, fruits and vegetables, chickens or eggs contaminated with *Salmonella* (Salleh et al., 2017; Soon et al., 2011).

The increasing concern about the overuse of drugs and chemical in livestock production and improving awareness of food safety and animal welfare resulted in expanding demand for free-range livestock products including chickens. Consumers are willing to pay more to obtain products they consider safer and more wholesome. Therefore, *Ayam Kampung* has gained popularity and demand over the past few years. However, there have been no reports on the microbiological quality of *Ayam Kampung* and their products in Malaysia to substantiate the assumptions of safety. This study will address that gap of knowledge.

1.3 Justification of the study

Majority of published work on chickens has focused on processed chicken meat, gizzards, liver, and other organs of chicken carcasses, most of which were at retail outlets and wet markets. None of these studies explored on the microbiological quality of village chickens and their products despite getting more attention and increased preference among consumers compared to the commercial and other exotic chicken and chicken products in Malaysia. Consumers are paying more for something that is believed to be safer and more wholesome. This study will provide information on the safety aspect of the meat, by focusing on the most common foodborne pathogen *Salmonella* and determine the antibiotic resistance patterns of the isolates as well as their genetic diversity.

1.4 Research Questions

- i. What is the prevalence of *Salmonella* infection among *Ayam Kampung* in the central and southern Peninsular Malaysia?
- ii. What are the epidemiological risk factors that contribute to the occurrence of salmonellae among *Ayam Kampung* in the central and southern Peninsular Malaysia?
- iii. What are the antibiotic resistance profiles/susceptibility patterns of the *Salmonella* isolates?
- iv. What are the genotypes of the *Salmonella* isolates?

1.5 Research Hypothesis

- i. The prevalence of *Salmonella* among *Ayam kampung* in the South-central Peninsular Malaysia is high.
- ii. Certain farm management factors play a vital role in the occurrence and epidemiology of salmonellosis among *Ayam Kampung*
- iii. *Salmonella* isolates from village chickens are multidrug resistant
- iv. *Salmonella* isolates recovered from *Ayam Kampung* are genetically diverse.

1.6 General Objective

To determine the epidemiology, antibiotic resistance level, associated risk factors and genetic diversity of *Salmonella* isolates among *Ayam kampung* in the south-central Peninsular Malaysia.

1.7 Specific Objectives

- i. To determine the prevalence of salmonellosis and epidemiological risk factors associated with its occurrence among *Ayam kampung* in the South-central Peninsular Malaysia.
- ii. To determine the antimicrobial resistance susceptibility patterns of the *Salmonella* isolates recovered.
- iii. To determine the genetic diversity of the *Salmonella* isolates by multilocus sequence typing.



REFERENCES

- Abalaka, G. O. (2013). Rearing methods, seasons of the year and survivability of rural poultry enterprise in Nigeria. *Journal of Agriculture and Sustainability*, 3(1), 27–55.
- Abdel-Maksoud, M., Abdel-Khalek, R., El-Gendy, A., Gamal, R. F., Abdelhady, H. M., & House, B. L. (2015). Genetic characterisation of multidrug-resistant *Salmonella enterica* serotypes isolated from poultry in Cairo, Egypt. *African Journal of Laboratory Medicine*, 4(1).
- Abdul-Mutalib, N. A., Syafinaz, A. N., Sakai, K., & Shirai, Y. (2015). An overview of foodborne illness and food safety in Malaysia. *International Food Research Journal*, 22(3), 896–901.
- Abdullah, K., Orhan, B., Nafiz, K., Belkis, L., Can Polat, E., Omer Faruk, T., Levent, G., Orhan, B., & A. Celal, B. (2010). Analysis of an outbreak of *Salmonella* Enteritidis by repetitive-sequence-based PCR and pulsed-field gel electrophoresis. *Internal Medicine*, 49, 31–36.
- Achtman, M., Wain, J., Weill, F.-X., Nair, S., Zhou, Z., Sangal, V., Krauland, M. G., Hale, J. L., Harbottle, H., Uesbeck, A., Dougan, G., Harrison, L. H., & Brisse, S. (2012). Multilocus sequence typing as a replacement for serotyping in *Salmonella enterica*. *PLoS Pathogens*, 8(6), e1002776.
- Adzitey, F., Rusul, G., & Huda, N. (2012). Prevalence and antibiotic resistance of *Salmonella* serovars in ducks, duck rearing and processing environments in Penang, Malaysia. *Food Research International*, 45(2), 947–952.
- Afema, J. A., & Sischo, W. M. (2016). *Salmonella* in wild birds utilizing protected and human impacted habitats, Uganda. *EcoHealth*, 13(3), 558–569.
- Afshari, A., Baratpour, A., Khanzade, S., & Jamshidi, A. (2018). *Salmonella* Enteritidis and *Salmonella* Typhimorium identification in poultry carcasses. *Iranian Journal of Microbiology*, 10(1), 45–50.
- Ahlstrom, C. A., Bonnedahl, J., Woksepp, H., Hernandez, J., Olsen, B., & Ramey, A. M. (2018). Acquisition and dissemination of cephalosporin-resistant *E. coli* in migratory birds sampled at an Alaska landfill as inferred through genomic analysis. *Scientific Reports*, 8(1), 7361.
- Ahmer, B. M. M., Tran, M., & Heffron, F. (1999). The virulence plasmid of *Salmonella* Typhimurium is self-transmissible. *Journal of Bacteriology*, 181(4), 1364–1368.
- Aini, I. (1990). Indigenous chicken production in South-east Asia. *World's Poultry Science Journal*, 46, 51–57.

- Al-Tawfiq, J. A., Laxminarayan, R., & Mendelson, M. (2017). How should we respond to the emergence of plasmid-mediated colistin resistance in humans and animals? *International Journal of Infectious Diseases*, *54*, 77–84.
- Almakki, A., Jumas-Bilak, E., Marchandin, H., & Licznar-Fajardo, P. (2019). Antibiotic resistance in urban runoff. *Science of The Total Environment*, *667*, 64–76.
- Amalaradjou, M. (2019). Pre-harvest Approaches to Improve Poultry Meat Safety. In K. Venkitanarayanan, S. Thakur, & S. C. Ricke (Eds.), *Food Safety in Poultry Meat Production. Food Microbiology and Food Safety*. Springer international publishing.
- Ambler, R. P., Coulson, A. F. W., Frère, J. M., Ghuysen, J. M., Joris, B., Forsman, M., Levesque, R. C., Tiraby, G., & Waley, S. G. (1991). A standard numbering scheme for the class A β -lactamases. *Biochemical Journal*, *276*(1), 269–270.
- Amoako, D. G., Somboro, A. M., Abia, A. L. K., Molechan, C., Perrett, K., Bester, L. A., & Essack, S. Y. (2019). Antibiotic resistance in *Staphylococcus aureus* from poultry and poultry products in uMgungundlovu district, South Africa, using the “farm to fork” approach. *Microbial Drug Resistance*, *00*(00).
- Andino, A., & Hanning, I. (2015). *Salmonella enterica*: Survival, colonization, and virulence differences among serovars. *Scientific World Journal*, Article ID. 520179.
- Andoh, L. A., Dalsgaard, A., Obiri-Danso, K., Newman, M. J., Barco, L., & Olsen, J. E. (2016). Prevalence and antimicrobial resistance of *Salmonella* serovars isolated from poultry in Ghana. *Epidemiology and Infection*, *144*(15), 3288–3299.
- Andres, V. M., & Davies, R. H. (2015). Biosecurity measures to control *Salmonella* and other infectious agents in pig farms: A review. *Comprehensive Reviews in Food Science and Food Safety*, *14*(4), 317–335.
- Andrés-Barranco, S., Vico, J. P., Garrido, V., Samper, S., Herrera-León, S., de Frutos, C., & Mainar-Jaime, R. C. (2014). Role of wild bird and rodents in the epidemiology of subclinical salmonellosis in finishing pigs. *Foodborne Pathogens and Disease*, *11*(9), 689–697.
- Ansari, F., Pourjafar, H., Bokaie, S., Peighambari, S. M., Mahmoudi, M., Fallah, M. H., Tehrani, F., Rajab, A., Ghafouri, S. A., & Shabani, M. (2017). Association between poultry density and *Salmonella* infection in commercial laying flocks in Iran using a Kernel density. *Pakistan Veterinary Journal*, *37*(3), 299–304.
- Arsenault, J., Letellier, A., Quessy, S., Normand, V., & Boulianne, M. (2007). Prevalence and risk factors for *Salmonella spp.* and *Campylobacter spp.*

caecal colonization in broiler chicken and turkey flocks slaughtered in Quebec, Canada. *Preventive Veterinary Medicine*, 81(4), 250–264.

- Arumugaswamy, R. K., Rusul, G., Abdul Hamid, S. N., & Cheah, C. T. (1995). Prevalence of *Salmonella* in raw and cooked foods in Malaysia. *Food Microbiology*, 12, 3–8.
- Assefa, M., Teklu, a, & Negussie, H. (2011). The prevalence and public health importance of *Salmonella* from chicken table eggs, Ethiopia. *American-Eurasian Journal of Agricultural & Environmental Sciences*, 11(4), 512–518.
- Atterby, C., Ramey, A. M., Hall, G. G., Järhult, J., Börjesson, S., & Bonnedahl, J. (2016). Increased prevalence of antibiotic-resistant *E. coli* in gulls sampled in Southcentral Alaska is associated with urban environments. *Infection Ecology & Epidemiology*, 6(1), 32334.
- Ayachi, A., Bennoune, O., Heleili, N., & Alloui, N. (2015). Minor *Salmonella*: Potential pathogens in eggs in Algeria. *Journal of Infection in Developing Countries*, 9(10), 1156–1160.
- Azahan, E. A. E., Seet, C. P., & Zainab, O. (1980). A comparative study of physiological and productive performance of local (*kampung*) and commercial chickens. *Malaysian Agricultural Research Journal*, 52, 61–70.
- Azahan, E. E. A. (1994). The red and black-red native chickens of Malaysia. *Malaysian Agricultural Research and Development Institute (MARDI) Research Journal*, 22(1), 73–78.
- Azahan, E. E. A., & Zahari, W. M. (1983). Observation on some characteristics of carcass and meat of Malaysian *kampung* chickens. *Mardi Research Bulletin*, 11, 225–232.
- Azam, M., Ehsan, I., Sajjad-ur-Rahman, Saleemi, M. K., Javed, M. R., & Mohsin, M. (2017). Detection of the colistin resistance gene *mcr-1* in avian pathogenic *Escherichia coli* in Pakistan. *Journal of Global Antimicrobial Resistance*, 11, 152–153.
- Bailey, J. S. (1988). Integrated colonization control of *Salmonella* in poultry. *Poultry Science*, 67(6), 928–932.
- Bakowski, M. A., Braun, V., & Brumell, J. H. (2008). *Salmonella*-containing vacuoles: Directing traffic and nesting to grow. *Traffic*, 9(12), 2022–2031.
- Baron, S., Hadjadj, L., Rolain, J. M., & Olaitan, A. O. (2016). Molecular mechanisms of polymyxin resistance: knowns and unknowns. *International Journal of Antimicrobial Agents*, 48(6), 583–591.
- Bartoloni, A., Pallecchi, L., Rodríguez, H., Fernandez, C., Mantella, A., Bartalesi, F., Strohmeyer, M., Kristiansson, C., Gotuzzo, E., Paradisi, F., & Rossolini, G.

- M. (2009). Antibiotic resistance in a very remote Amazonas community. *International Journal of Antimicrobial Agents*, 33(2), 125–129.
- Batz, M. B., Hoffmann, S., & Morris, J. G. (2012). Ranking the disease burden of 14 pathogens in food sources in the United States using attribution data from outbreak investigations and expert elicitation. *Journal of Food Protection*, 75(7), 1278–1291.
- Bäumler, A. J., Tsolis, R. M., Ficht, T. A., & Adams, L. G. (1998). Evolution of host adaptation in *Salmonella enterica*. *Infection and Immunity*, 66(10), 4579–4587.
- Beier, R. C., Pillai, S. D., Phillips, T. D., & Ziprin, R. L. (2004). *Preharvest and Postharvest Food Safety: Contemporary Issues and Future Directions* (R. C. Beier, S. D. Pillai, & T. D. Phillips (eds.)). Blackwell Publishing.
- Benacer, D., Thong, K.-L., Watanabe, H., & Puthucheary, S. D. (2010). Characterization of drug resistant *Salmonella enterica* serotype Typhimurium by antibiograms, plasmids, integrons, resistance genes and PFGE. *Journal of Microbiology and Biotechnology*, 20(6), 1042–1052.
- Bentley, S. D., & Parkhill, J. (2015). Genomic perspectives on the evolution and spread of bacterial pathogens. *Proceedings of the Royal Society B*, 282(1821).
- Bertrand, S., De Lamine De Bex, G., Wildemaue, C., Lunguya, O., Phoba, M. F., Ley, B., Jacobs, J., Vanhoof, R., & Mattheus, W. (2015). Multi-locus variable-number tandem repeat (MLVA) typing tools improved the surveillance of *Salmonella* Enteritidis: A 6 years retrospective study. *PLoS ONE*, 10(2), 1–13.
- Bhan, M. K., Bahl, R., & Bhatnagar, S. (2005). Typhoid and paratyphoid fever. *Lancet*, 366(2), 749–762.
- Bhunja, A. K. (2008). *Foodborne microbial pathogens: Mechanisms and pathogenesis*. Springer Science +Business Media, LLC.
- Biswas, C., Leboveic, A., Burke, K., & Biswas, D. (2019). Post-harvest approaches to improve poultry meat safety. In *Food Safety in Poultry Meat Production* (pp. 123–138). Springer International Publishing.
- Bitrus, A. A., Zunita, Z., Bejo, S. K., Othman, S., & Nadzir, N. A. A. (2017). In vitro transfer of methicillin resistance determinants *mecA* from methicillin resistant *Staphylococcus aureus* (MRSA) to methicillin susceptible *Staphylococcus aureus* (MSSA). *BMC Microbiology*, 17(1), 83.
- Boonkhot, P., Tadee, P., Yamsakul, P., Pocharoen, C., Chokesajjawatee, N., & Patchanee, P. (2015). Class 1 integrons characterization and multilocus sequence typing of *Salmonella spp.* from swine production chains in Chiang

- Mai and Lamphun provinces, Thailand. *Japanese Journal of Veterinary Research*, 63(2), 83–94.
- Borges, K., Furian, T., Souza, S., Salle, C., Moraes, H., & Nascimento, V. (2019). Antimicrobial resistance and molecular characterization of *Salmonella enterica* serotypes isolated from poultry sources in Brazil. *Brazilian Journal of Poultry Science*, 21(1).
- Boxall, A., & Wilkinson, J. (2019). Identifying hotspots of resistance selection from antibiotic exposure in urban environments around the world. *SETAC Europe 29th Annual Meeting, Helsinki, Finland*.
- Brenner, F. W., Villar, R. G., Angulo, F. J., Tauxe, R., & Swaminathan, B. (2000). *Salmonella* nomenclature. *Journal of Clinical Microbiology*, 38(7), 2465–2467.
- Brown, A. C., Grass, J. E., Richardson, L. C., Nisler, A. L., Bicknese, A. S., & Gould, L. H. (2017). Antimicrobial resistance in *Salmonella* that caused foodborne disease outbreaks: United States, 2003–2012. *Epidemiology and Infection*, 145(4), 766–774.
- Bruun, T., Sørensen, G., Forshell, L., Jensen, T., Nygard, K., Kapperud, G., Lindstedt, B., Berglund, T., Wingstrand, A., Petersen, R., Müller, L., Kjelsø, C., Ivarsson, S., Hjertqvist, M., Löfdahl, S., & Ethelberg, S. (2009). An outbreak of *Salmonella* Typhimurium infections in Denmark, Norway and Sweden, 2008. *Eurosurveillance*, 14(10).
- Bryant, E. H. (1977). Morphometric adaptation of the housefly, *Musca domestica* L., in the United States. *Evolution*, 31(3), 580.
- Bush, K. (2003). Beta-lactam antibiotics: Penicillins. In R. G. Finch, D. Greenwood, S. R. Noorby, & R. J. Whitley (Eds.), *Antibiotic and chemotherapy: Anti-infective agents and their use in therapy* (pp. 224–258).
- Bush, K., Jacoby, G., & Medeiros, A. (1995). A functional classification scheme for beta-lactamases and its correlation with molecular structure. *Antimicrobial Agents and Chemotherapy*, 39(6), 1211–1233.
- Butaye, P., Cloeckert, A., & Schwarz, S. (2003). Mobile genes coding for efflux-mediated antimicrobial resistance in Gram-positive and Gram-negative bacteria. *International Journal of Antimicrobial Agents*, 22(3), 205–210.
- Cameron-Veas, K., Fraile, L., Napp, S., Garrido, V., Grilló, M. J., & Migura-Garcia, L. (2018). Multidrug resistant *Salmonella enterica* isolated from conventional pig farms using antimicrobial agents in preventative medicine programmes. *The Veterinary Journal*, 234, 36–42.
- Cardoso, M. O., Ribeiro, A. R., Dos Santos, L. R., Pilotto, F., De Moraes, H. L. S., Salle, C. T. P., Rocha, S. L. D. S., & Do Nascimento, V. P. (2006). Antibiotic

resistance in *Salmonella* Enteritidis isolated from broiler carcasses. *Brazilian Journal of Microbiology*, 37(3), 368–371.

Carrique-Mas, J. J., & Davies, R. H. (2008). Sampling and bacteriological detection of *Salmonella* in poultry and poultry premises: A review. *Revue Scientifique et Technique (International Office of Epizootics)*, 27(3), 665–677.

Centers for Disease Control and Prevention. (2006). Multistate outbreak of *Salmonella* Typhimurium infections associated with eating ground beef—United States, 2004. *Morbidity and Mortality Weekly Report*, 55(7), 180–182.

Chashni, E., Hassanzadeh, S. H., & Fard, B. (2009). Characterization of the *Salmonella* isolates from backyard chickens in North of Iran, by serotyping, multiplex PCR and antibiotic resistance analysis. *Archives of Razi Institute*, 64(2), 77–83.

Chatham-Stephens, K., Medalla, F., Hughes, M., Appiah, G. D., Aubert, R. D., Caidi, H., Angelo, K. M., Walker, A. T., Hatley, N., Masani, S., Nash, J., Belko, J., Ryan, E. T., Mintz, E., & Friedman, C. R. (2019). Emergence of extensively drug-resistant *Salmonella* Typhi infections among travelers to or from Pakistan — United States, 2016–2018. *Morbidity and Mortality Weekly Report*, 68(1), 11–13.

Chen, S., Zhao, S., White, D. G., Schroeder, C. M., Lu, R., Yang, H., McDermott, P. F., Ayers, S., & Meng, J. (2004). Characterization of multiple-antimicrobial-resistant *Salmonella* serovars isolated from retail meats. *Applied and Environmental Microbiology*, 70(1), 1–7.

Chen, Y., Zhang, W., & Knabel, S. J. (2007). Multi-virulence-locus sequence typing identifies single nucleotide polymorphisms which differentiate epidemic clones and outbreak strains of *Listeria monocytogene*. *Journal of Clinical Microbiology*, 45(3), 835–846.

Choo, L. C., Saleha, A. A., Wai, S. S., & Fauziah, N. (2011). Isolation of *Campylobacter* and *Salmonella* from houseflies (*Musca domestica*) in a university campus and a poultry farm in Selangor, Malaysia. *Tropical Biomedicine*, 28(1), 16–20.

Chopra, I., & Roberts, M. (2001). Tetracycline antibiotics: mode of action, applications, molecular biology, and epidemiology of bacterial resistance. *Microbiology and Molecular Biology Reviews*, 65(2), 232–260.

Clinical and Laboratory Standards Institute. (2018). *Performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals*. Clinical and Laboratory Standards Institute. (4th ed.).

Collignon, P. C., Conly, J. M., Andremont, A., McEwen, S. A., & Aidara-Kane, A. (2016). World health organization ranking of antimicrobials according to

their importance in human medicine: A critical step for developing risk management strategies to control antimicrobial resistance from food animal production. *Clinical Infectious Diseases*, 63(8), 1087–1093.

Crump, J. A., Luby, S. P., & Mintz, E. D. (2004). The global burden of typhoid fever. *Bulletin of the World Health Organization*, 82(5), 346–353.

D'Aoust, J. Y. (1989). Salmonella. In M. P. Doyle (Ed.), *Foodborne Bacterial Pathogens* (pp. 327–445). Marcel, Inc.

Dahiya, S., Kapil, A., Kumar, R., Das, B. K., Sood, S., Chaudhry, R., Kabra, S. K., & Lodha, R. K. (2013). Multiple locus sequence typing of *Salmonella* Typhi, isolated in north India - a preliminary study. *The Indian Journal of Medical Research*, 137(5), 957–962.

Dang-Xuan, S., Nguyen-Viet, H., Pham-Duc, P., Unger, F., Tran-Thi, N., Grace, D., & Makita, K. (2019). Risk factors associated with *Salmonella* spp. prevalence along smallholder pig value chains in Vietnam. *International Journal of Food Microbiology*, 290, 105–115.

Davies, P. R., Scott Hurd, H., Funk, J. A., Fedorka-Cray, P. J., & Jones, F. T. (2004). The role of contaminated feed in the epidemiology and control of *Salmonella enterica* in pork production. *Foodborne Pathogens and Disease*, 1(4), 202–215.

Davies, R. H., & Wray, C. (1995). Mice as carriers of *Salmonella* Enteritidis on persistently infected poultry units. *The Veterinary Record*, 137(14), 337–341.

Denagamage, T., Jayarao, B., Patterson, P., Wallner-Pendleton, E., & Kariyawasam, S. (2015). Risk factors associated with *Salmonella* in laying hen farms: Systematic review of observational studies. *Avian Diseases*, 59(2), 291–302.

Department of Statistics Malaysia. (2017). *Department of Statistics Malaysia: Supply and Utilization Accounts Selected Agricultural Commodities, Malaysia, 2013-2017*. <https://www.dosm.gov.my>

Department of Veterinary Services. (2012). *Department of Veterinary Services, Malaysia*. <https://www.dvs.gov.my>

Diwyanto, K., & Iskandar, S. (1999). Kampung chickens: A key part of Indonesia's livestock sector. In *FAO. Central Research Institute for Animal Sciences and Research Institute for Animal Production*.

Economou, V., & Gousia, P. (2015). Agriculture and food animals as a source of antimicrobial-resistant bacteria. *Infection and Drug Resistance*, 8, 49–61.

Ed-Dra, A., Karraouan, B., Allaoui, A. El, Khayatti, M., Ossmani, H. El, Filali, F. R., ElMdaghri, N., & Bouchrif, B. (2018). Antimicrobial resistance and genetic diversity of *Salmonella* Infantis isolated from foods and human

- samples in Morocco. *Journal of Global Antimicrobial Resistance*, *14*, 297–301.
- Eguale, T. (2018). Non-typhoidal *Salmonella* serovars in poultry farms in central Ethiopia: Prevalence and antimicrobial resistance. *BMC Veterinary Research*, *14*(1), 1–8.
- Eiamphungporn, W., Yainoy, S., Jumderm, C., Tan-arsuwongkul, R., Tiengrim, S., & Thamlikitkul, V. (2018). Prevalence of the colistin resistance gene *mcr-1* in colistin-resistant *Escherichia coli* and *Klebsiella pneumoniae* isolated from humans in Thailand. *Journal of Global Antimicrobial Resistance*, *15*, 32–35.
- Elias, S. de O., Decol, L. T., & Tondo, E. C. (2018). Foodborne outbreaks in Brazil associated with fruits and vegetables: 2008 through 2014. *Food Quality and Safety*, *2*(4), 173–181.
- Elkenany, R., Elsayed, M. M., Zakaria, A. I., El-sayed, S. A., & Rizk, M. A. (2019). Antimicrobial resistance profiles and virulence genotyping of *Salmonella enterica* serovars recovered from broiler chickens and chicken carcasses in Egypt. *BMC Veterinary Research*, *15*(1), 124.
- Elkenany, R. M., Eladl, A. H., & El-Shafei, R. A. (2018). Genetic characterisation of class 1 integrons among multidrug-resistant *Salmonella* serotypes in broiler chicken farms. *Journal of Global Antimicrobial Resistance*, *14*, 202–208.
- Eng, S. K., Pusparajah, P., Ab Mutalib, N. S., Ser, H. L., Chan, K. G., & Lee, L. H. (2015). *Salmonella*: A review on pathogenesis, epidemiology and antibiotic resistance. *Frontiers in Life Science*, *8*(3), 284–293.
- European Medicines Agency. (2016). *Updated advice on the use of colistin products in animals within the European Union: development of resistance and possible impact on human and animal health*. EMA/231573/2016.
- Feil, E. J., Li, B. C., Aanensen, D. M., Hanage, W. P., & Spratt, B. G. (2004). eBURST: Inferring patterns of evolutionary descent among clusters of related bacterial genotypes from multilocus sequence typing data. *Journal of Bacteriology*, *186*(5), 1518–1530.
- Foley, S. L., & Lynne, A. M. (2008). Food animal-associated *Salmonella* challenges: Pathogenicity and antimicrobial resistance. *Journal of Animal Science*, *86*, E173–E187.
- Foley, S. L., Lynne, A. M., & Nayak, R. (2008). *Salmonella* challenges: Prevalence in swine and poultry and potential pathogenicity of such isolates. *Journal of Animal Science*, *86*, E149–E162.
- Foley, S. L., Nayak, R., Hanning, I. B., Johnson, T. J., Han, J., & Ricke, S. C. (2011). Population dynamics of *Salmonella enterica* serotypes in commercial egg and poultry production. *Applied and Environmental Microbiology*,

77(13), 4273–4279.

- Food and Drug Administration. (2010). *National Antimicrobial Monitoring System - Enteric bacteria (NARMS): 2007 executive summary report*. Rockville, MD: U.S. Department of health and human services, US Food and Drug Administration.
- Food Safety News. (2018). *Laksa food poisoning caused by Salmonella Weltevreden in Kedah, Malaysia*.
- Founou, L. L., Founou, R. C., & Essack, S. Y. (2016). Antibiotic resistance in the food chain: A developing country-perspective. *Frontiers in Microbiology*, 7, 1–19.
- Francisco, A. P., Bugalho, M., Ramirez, M., & Carriço, J. A. (2009). Global optimal eBURST analysis of multilocus typing data using a graphic matroid approach. *BMC Bioinformatics*, 10(1), 152.
- Francisco, A. P., Vaz, C., Monteiro, P. T., Melo-Cristino, J., Ramirez, M., & Carriço, J. A. (2012). PHYLOViZ: phylogenetic inference and data visualization for sequence based typing methods. *BMC Bioinformatics*, 13(1), 87.
- Frederick, A., & Huda, N. (2011). *Salmonella*, poultry house environments and feeds: A review. *Journal of Animal and Veterinary Advances*, 10(5), 679–685.
- Galanis, E., Lo Fo Wong, D. M. A., Patrick, M. E., Binsztein, N., Cieslik, A., Chalermchaikit, T., Aidara-Kane, A., Ellis, A., Angulo, F. J., & Wegener, H. C. (2006). Web-based surveillance and global *Salmonella* distribution, 2000–2002. *Emerging Infectious Diseases*, 12(3), 381–388.
- Geidam, Y. A., Zakaria, Z., Aziz, S. A., Bejo, S. K., Abu, J., & Omar, S. (2012). High prevalence of multi-drug resistant bacteria in selected poultry farms in Selangor, Malaysia. *Asian Journal of Animal and Veterinary Advances*, 7(9), 891–897.
- Gerlach, R. G., & Hensel, M. (2007). *Salmonella* pathogenicity islands in host specificity, host pathogen-interactions and antibiotics resistance of *Salmonella enterica*. *Berliner Und Munchener Tierarztliche Wochenschrift*, 120(7-8), 317–327.
- Ghaderi, R., Tadayon, K., Khaki, P., & Mosavari, N. (2015). Iranian clonal population of *Salmonella enterica* serovar Enteritidis, characterized by multi-locus sequence typing (MLST) method. *Iranian Journal of Microbiology*, 7(5), 251–259.
- Ghafur, A., Shankar, C., GnanaSoundari, P., Venkatesan, M., Mani, D., Thirunarayanan, M. A., & Veeraraghavan, B. (2019). Detection of

chromosomal and plasmid-mediated mechanisms of colistin resistance in *Escherichia coli* and *Klebsiella pneumoniae* from Indian food samples. *Journal of Global Antimicrobial Resistance*, 16, 48–52.

Ghoddusi, A., Nayeri Fasaee, B., Karimi, V., Ashrafi Tamai, I., Moulana, Z., & Zahraei Salehi, T. (2015). Molecular identification of *Salmonella* Infantis isolated from backyard chickens and detection of their resistance genes by PCR. *Iranian Journal of Veterinary Research*, 16(3), 293–297.

Goering, R. V. (2010). Pulsed field gel electrophoresis: A review of application and interpretation in the molecular epidemiology of infectious disease. *Infection, Genetics and Evolution*, 10(7), 866–875.

Gomes, A., Quinteiro-Filho, W., Ribeiro, A., Ferraz-de-Paula, V., Pinheiro, M., Baskeville, E., Akamine, A., Astolfi-Ferreira, C., Ferreira, A., & Palermo-Neto, J. (2014). Overcrowding stress decreases macrophage activity and increases *Salmonella* Enteritidis invasion in broiler chickens. *Avian Pathology*, 43(1), 82–90.

Gow, A. G., Gow, D. J., Hall, E. J., Langton, D., Clarke, C., & Pappasouliotis, K. (2009). Prevalence of potentially pathogenic enteric organisms in clinically healthy kittens in the UK. *Journal of Feline Medicine and Surgery*, 11(8), 655–662.

Grassl, G. A., & Finlay, B. B. (2008). Pathogenesis of enteric *Salmonella* infections. *Current Opinion in Gastroenterology*, 24(1), 22–26.

Gray, J. T., & Fedorka-Cray, P. J. (2002). *Salmonella*. In D. O. Cliver & H. P. Riemann (Eds.), *Foodborne diseases* (pp. 55–68). Academic Press.

Guerra, B., Fischer, J., & Helmuth, R. (2014). An emerging public health problem: Acquired carbapenemase-producing microorganisms are present in food-producing animals, their environment, companion animals and wild birds. *Veterinary Microbiology*, 171(3-4), 290–297.

Gullan, P., & Cranston, P. (2010). *The Insects: An Outline of Entomology* (4th ed.). Wiley.

Hanes, D. (2003). Nontyphoid *Salmonella*. In O. Henegariu, N. A. Heerema, S. R. Dlouhy, G. H. Vance, & P. H. Vogt (Eds.), *International handbook of foodborne pathogens* (pp. 137–149). Marcel Dekker, Inc.

Hanson, D. L., Loneragan, G. H., Brown, T. R., Nisbet, D. J., Hume, M. E., & Edrington, T. S. (2016). Evidence supporting vertical transmission of *Salmonella* in dairy cattle. *Epidemiology and Infection*, 144(5), 962–967.

Hassan, L., Zaleha Suhaimi, S., & Saleha, A. A. (2005). The detection and comparison of antimicrobial resistance pattern of vancomycin-resistant *Enterococci* and *Salmonella* isolated from eggs of commercial layers and

free-range chickens. *Jurnal Veterinar Malaysia*, 17(1), 7–11.

- Hawkey, J., Le Hello, S., Doublet, B., Granier, S. A., Hendriksen, R. S., Fricke, W. F., Ceyskens, P.-J., Gomart, C., Billman-Jacobe, H., Holt, K. E., & Weill, F.-X. (2019). Global phylogenomics of multidrug-resistant *Salmonella enterica* serotype Kentucky ST198. *Microbial Genomics*, 5(7).
- Hembach, N., Schmid, F., Alexander, J., Hiller, C., Rogall, E. T., & Schwartz, T. (2017). Occurrence of the mcr-1 colistin resistance gene and other clinically relevant antibiotic resistance genes in microbial populations at different municipal wastewater treatment plants in Germany. *Frontiers in Microbiology*, 8, 1282.
- Hendriksen, R. S., Vieira, A. R., Karlslose, S., Lo Fo Wong, D. M. A., Jensen, A. B., Wegener, H. C., & Aarestrup, F. M. (2011). Global monitoring of *Salmonella* serovar distribution from the world health organization global foodborne infections network country data bank: Results of quality assured laboratories from 2001 to 2007. *Foodborne Pathogens and Disease*, 8(8), 887–900.
- Heredia, N., & García, S. (2018). Animals as sources of food-borne pathogens: A review. *Animal Nutrition*, 4(3), 250–255.
- Hewitt, C. G. (2011). *The house-fly, Musca domestica Linn: its structure, habits, development, relation to disease and control*. Cambridge University Press.
- Hoelzer, K., Switt, A. I. M., & Wiedmann, M. (2011). Animal contact as a source of human non-typhoidal salmonellosis. *Veterinary Research*, 42(1).
- Holmes, A. H., Moore, L. S., Sundsfjord, A., Steinbakk, M., Regmi, S., Karley, A., & et al. (2016). Understanding the mechanisms and drivers of antimicrobial resistance. *Lancet*, 387, 176–187.
- Holt, K. E., Thomson, N. R., Wain, J., Phan, M. D., Nair, S., Hasan, R., Bhutta, Z. A., Quail, M. A., Norbertczak, H., Walker, D., Dougan, G., & Parkhill, J. (2007). Multidrug-resistant *Salmonella enterica* serovar Paratyphi A harbors IncHI1 plasmids similar to those found in serovar Typhi. *Journal of Bacteriology*, 189(11), 4257–4264.
- Hölzel, C. S., Tetens, J. L., & Schwaiger, K. (2018). Unraveling the role of vegetables in spreading antimicrobial-resistant bacteria: A need for quantitative risk assessment. *Foodborne Pathogens and Disease*, 15(11), 671–688.
- Hong, Y., Wang, Y., Huang, I., Liao, Y., Kuo, H., Liu, Y., Tu, Y., Chen, B., Liao, Y., & Chiou, C. (2018). Genetic relationships among multidrug-resistant *Salmonella enterica* serovar Typhimurium strains from humans and animals. *Antimicrobial Agents and Chemotherapy*, 62(5), e00213–e00218.

- Hoover, N. J., Kenney, P. B., Amick, J. D., & Hypes, W. A. (1997). Preharvest sources of *Salmonella* colonization in turkey production. *Poultry Science*, 76(9), 1232–1238.
- Huang, X., Yu, L., Chen, X., Zhi, C., Yao, X., Liu, Y., Wu, S., Guo, Z., Yi, L., Zeng, Z., & Liu, J. H. (2017). High prevalence of colistin resistance and mcr-1 gene in *Escherichia coli* isolated from food animals in China. *Frontiers in Microbiology*, 8(APR).
- Huneau-Salaün, A., Chemaly, M., Le Bouquin, S., Lalande, F., Petetin, I., Rouxel, S., Michel, V., Fravalo, P., & Rose, N. (2009). Risk factors for *Salmonella enterica* subsp. *enterica* contamination in 519 French laying hen flocks at the end of the laying period. *Preventive Veterinary Medicine*, 89(1-2), 51–58.
- Huovinen, P., Sundstrom, L., Swedberg, G., & Skold, O. (1995). Trimethoprim and sulfonamide resistance. In *Antimicrobial Agents and Chemotherapy* (Vol. 39, Issue 2, pp. 279–289).
- Hur, J., Jawale, C., & Lee, J. H. (2012). Antimicrobial resistance of *Salmonella* isolated from food animals: A review. *Food Research International*, 45(2), 819–830.
- Ibrahim, G. M., & Morin, P. M. (2018). *Salmonella* serotyping using whole genome sequencing. *Frontiers in Microbiology*, 9.
- Im, M. C., Jeong, S. J., Kwon, Y. K., Jeong, O. M., Kang, M. S., & Lee, Y. J. (2015). Prevalence and characteristics of *Salmonella spp.* isolated from commercial layer farms in Korea. *Poultry Science*, 94(7), 1691–1698.
- Isaacs, D. (2015). Antibiotic resistance in remote tribespeople. *Journal of Paediatrics and Child Health*, 51(9), 940–940.
- Ishida, A., Law, S. H., & Aita, Y. (2003). Changes in food consumption expenditure in Malaysia. *Agribusiness*, 19(1), 61–76.
- Jafari, R. A., Ghorbanpour, M., & Jaideri, A. (2007). An investigation into *Salmonella* infection status in backyard chickens in Iran. *International Journal of Poultry Science*, 6(3), 227–229.
- Jalaludin, S., Sivarajasingam, S., & Oh, B. T. (1985). Some breed characteristics of *kampung* chicken. *Regional Seminar on Future Developments in Poultry Industry*, 43–45.
- Jegathesan, M. (1984). *Salmonella* serotypes isolated from man in Malaysia over the 10-year period 1973-1982. *Journal of Hygiene*, 92(3), 395–399.
- Jones, D. R., Karcher, D. M., Gast, R. K., Guraya, R., & Anderson, K. E. (2017). Frequency and duration of fecal shedding of *Salmonella* Enteritidis by experimentally infected laying hens housed in enriched colony cages at

different stocking densities. *Frontiers in Veterinary Science*, 4.

- Joshi, P. R., Thummeepak, R., Paudel, S., Acharya, M., Pradhan, S., Banjara, M. R., Leungtongkam, U., & Sitthisak, S. (2019). Molecular characterization of colistin-resistant *Escherichia coli* isolated from chickens: First report from Nepal. *Microbial Drug Resistance*.
- Kadhim, K. K., Abu Bakar, M. Z., Mustapha, N. M., Babjee, M. A., & Saad, M. Z. (2014). The enzyme activities of pancreas and small intestinal contents in the Malaysian village chicken and broiler strains. *Pertanika Journal of Tropical Agricultural Science*, 37(2), 203–214.
- Kagambèga, A., Lienemann, T., Frye, J. G., Barro, N., & Haukka, K. (2018). Whole genome sequencing of multidrug-resistant *Salmonella enterica* serovar Typhimurium isolated from humans and poultry in Burkina Faso. *Tropical Medicine and Health*, 46, 4.
- Karim, B. A., Latip, A. L., Shukor, A. S. A., Rashid, N. A., Mohd, W. M. W., & Kamaludin, F. (2017). A large common source outbreak of *Salmonella* Typhimurium linked to Kuala Terengganu night markets, Malaysia, 2014. *Outbreak, Surveillance and Investigation Reports*, 10(2), 1–7.
- Karp, B. E., Tate, H., Plumblee, J. R., Dessai, U., Whichard, J. M., Thacker, E. L., Robertson Hale, K., Wilson, W., Friedman, C. R., Griffin, P. M., & McDermott, P. F. (2017). National antimicrobial resistance monitoring system: Two decades of advancing public health through integrated surveillance of antimicrobial resistance. *Foodborne Pathogens and Disease*, 14(10), 545–557.
- Kauffmann, F. (1941). A typical variant and a new serological variation in the *Salmonella* group. *Journal of Bacteriology*, 41, 127–141.
- Kaur, J., & Jain, S. K. (2012). Role of antigens and virulence factors of *Salmonella enterica* serovar Typhi in its pathogenesis. *Microbiological Research*, 167(4), 199–210.
- Kempf, I., Jouy, E., & Chauvin, C. (2016). Colistin use and colistin resistance in bacteria from animals. *International Journal of Antimicrobial Agents*, 48(6), 598–606.
- Khan, M. R. T., Chamhuri, S., & Farah, H. S. (2015). Green food consumption in Malaysia: A review of consumers' buying motives. *International Food Research Journal*, 22(1), 131–138.
- Khoo, E., Roseliza, R., Khoo, L., Nafizah, M., Saifu Nazri, R., Hasnah, Y., Norazariyah, M., Rosnah, Y., Rosna, D., Siti Nor Hanani, R., & Ramlan, M. (2015). Antimicrobial resistance of *Salmonella enterica* serovar Typhimurium from various meats received in VRI. *Malaysian Journal of Veterinary Research*, 6(1), 61–65.

- Kich, J. D., Mores, N., Piffer, I. A., Coldebella, A., Amaral, A., Ramminger, L., & Cardoso, M. (2005). Factors associated with seroprevalence of *Salmonella* in commercial pig herds. *Ciência Rural*, 35, 398–405.
- Kilic, A., Bedir, O., Kocak, N., Levent, B., Eyigun, C. P., Tekbas, O. F., Gorenk, L., Baylan, O., & Basustaoglu, A. C. (2010). Analysis of an outbreak of *Salmonella* Enteritidis by repetitive-sequence-based PCR and pulsed-field gel electrophoresis. *Internal Medicine*, 49(1), 31–36.
- Ko, K. S., Suh, J. Y., Kwon, K. T., Jung, S. I., Park, K. H., Kang, C. I., Chung, D. R., Peck, K. R., & Song, J. H. (2007). High rates of resistance to colistin and polymyxin B in subgroups of *Acinetobacter baumannii* isolates from Korea. *Journal of Antimicrobial Chemotherapy*, 60(5), 1163–1167.
- Köser, C. U., Ellington, M. J., Cartwright, E. J. P., Gillespie, S. H., Brown, N. M., Farrington, M., Holden, M. T. G., Dougan, G., Bentley, S. D., Parkhill, J., & Peacock, S. J. (2012). Routine use of microbial whole genome sequencing in diagnostic and public health microbiology. *PLoS Pathogens*, 8(8).
- Krumperman, P. H. (1983). Multiple antibiotic resistance indexing of *Escherichia coli* to identify high-risk sources of fecal contamination of foods. *Applied and Environmental Microbiology*, 46(1), 165–170.
- Kuan, C. H., Rukayadi, Y., Ahmad, S. H., Wan Mohamed Radzi, C. W. J., Kuan, C. S., Yeo, S. K., Thung, T. Y., New, C. Y., Chang, W. S., Loo, Y. Y., Tan, C. W., Ramzi, O. S. B., Mohd Fadzil, S. N., Nordin, Y., Kwan, S. Y., & Son, R. (2017). Antimicrobial resistance of *Listeria monocytogenes* and *Salmonella* Enteritidis isolated from vegetable farms and retail markets in Malaysia. *International Food Research Journal*, 24(4).
- Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution*, 33(7), 1870–1874.
- Kumari, N., & Thakur, S. K. (2014). Randomly amplified polymorphic DNA - A brief review. *American Journal of Animal and Veterinary Sciences*, 9(1), 6–13.
- Kusumoto, M., Ogura, Y., Gotoh, Y., Iwata, T., Hayashi, T., & Akiba, M. (2016). Colistin-resistant mcr-1 –positive pathogenic *Escherichia coli* in swine, Japan, 2007–2014. *Emerging Infectious Diseases*, 22(7), 1315–1317.
- Lamas, A., Fernandez-No, I. C., Miranda, J. M., Vázquez, B., Cepeda, A., & Franco, C. M. (2016). Prevalence, molecular characterization and antimicrobial resistance of *Salmonella* serovars isolated from northwestern Spanish broiler flocks (2011–2015). *Poultry Science*, 95(9), 2097–2105.
- Lamas, A., Miranda, J. M., Regal, P., Vázquez, B., Franco, C. M., & Cepeda, A. (2018). A comprehensive review of non-enterica subspecies of *Salmonella*

enterica. *Microbiological Research*, 206, 60–73.

- Langridge, G. C., Wain, J., & Nair, S. (2012). Invasive salmonellosis in humans. *EcoSal Plus*, 5(1).
- Lawal, J. R., Jajere, S. M., Ibrahim, U. I., Geidam, Y. A., Gulani, I. A., Musa, G., & Ibekwe, B. U. (2016). Prevalence of coccidiosis among village and exotic breed of chickens in Maiduguri, Nigeria. *Veterinary World*, 9(6), 653–659.
- Le Minor, L. (1991). The genus *Salmonella*. In A. Balows, H. G. Truper, C. Dworkin, W. Harder, & K. H. Scheilfer (Eds.), *The Prokaryotes* (pp. 2760–2774). Springer-Verlag.
- Lee, J. H., Park, K. S., Jeon, J. H., & Lee, S. H. (2018). Antibiotic resistance in soil. *The Lancet Infectious Diseases*, 18(12), 1306–1307.
- Lee, K.-M., Runyon, M., Herrman, T. J., Phillips, R., & Hsieh, J. (2015). Review of *Salmonella* detection and identification methods: Aspects of rapid emergency response and food safety. *Food Control*, 47, 264–276.
- Leotta, G., Suzuki, K., Alvarez, F. L., Nuñez, L., Silva, M. G., Castro, L., Faccioli, M. L., Zarate, N., Weiler, N., Alvarez, M., & Copes, J. (2010). Prevalence of *Salmonella* spp. in backyard chickens in Paraguay. *International Journal of Poultry Science*, 9(6), 533–536.
- Li, B., Yang, X., Tan, H., Ke, B., He, D., Wang, H., Chen, Q., Ke, C., & Zhang, Y. (2018). Whole genome sequencing analysis of *Salmonella enterica* serovar Weltevreden isolated from human stool and contaminated food samples collected from the Southern coastal area of China. *International Journal of Food Microbiology*, 266, 317–323.
- Li, J., Hulth, A., Nilsson, L. E., Börjesson, S., Chen, B., Bi, Z., Wang, Y., Schwarz, S., & Wu, C. (2018). Occurrence of the mobile colistin resistance gene *mcr-3* in *Escherichia coli* from household pigs in rural areas. *Journal of Antimicrobial Chemotherapy*, 73(6), 1721–1723.
- Li, J., Nation, R. L., Turnidge, J. D., Milne, R. W., Coulthard, K., Rayner, C. R., & Paterson, D. L. (2006). Colistin: the re-emerging antibiotic for multidrug-resistant Gram-negative bacterial infections. *Lancet Infectious Diseases*, 6(9), 589–601.
- Li, R., Lai, J., Wang, Y., Liu, S., Li, Y., Liu, K., Shen, J., & Wu, C. (2013). Prevalence and characterization of *Salmonella* spp. isolated from pigs, ducks and chickens in Sichuan Province, China. *International Journal of Food Microbiology*, 163(1), 14–18.
- Li, X., Bethune, L. A., Jia, Y., Lovell, R. A., Proescholdt, T. A., Benz, S. A., Schell, T. C., Kaplan, G., & McChesney, D. G. (2012). Surveillance of *Salmonella* prevalence in animal feeds and characterization of the *Salmonella* isolates by

serotyping and antimicrobial susceptibility. *Foodborne Pathogens and Disease*, 9(8), 692–698.

Liu, W. bing, Liu, B., Zhu, X. na, Yu, S. jing, & Shi, X. ming. (2011). Diversity of *Salmonella* isolates using serotyping and multilocus sequence typing. *Food Microbiology*, 28(6), 1182–1189.

Liu, Y.-Y., Wang, Y., Walsh, T. R., Yi, L.-X., Zhang, R., Spencer, J., Doi, Y., Tian, G., Dong, B., Huang, X., Yu, L.-F., Gu, D., Ren, H., Chen, X., Lv, L., He, D., Zhou, H., Liang, Z., Liu, J.-H., & Shen, J. (2016). Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *The Lancet Infectious Diseases*, 16(2), 161–168.

Magiorakos, A. P., Srinivasan, A., Carey, R. B., Carmeli, Y., Falagas, M. E., Giske, C. G., Harbarth, S., Hindler, J. F., Kahlmeter, G., Olsson-Liljequist, B., Paterson, D. L., Rice, L. B., Stelling, J., Struelens, M. J., Vatopoulos, A., Weber, J. T., & Monnet, D. L. (2012). Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: An international expert proposal for interim standard definitions for acquired resistance. *Clinical Microbiology and Infection*, 18(3), 268–281.

Majowicz, S. E., Musto, J., Scallan, E., Angulo, F. J., Kirk, M., O'Brien, S. J., Jones, T. F., Fazil, A., & Hoekstra, R. M. (2010). The global burden of non-typhoidal *Salmonella* gastroenteritis. *Clinical Infectious Diseases*, 50(6), 882–889.

Malhotra-Kumar, S., Xavier, B. B., Das, A. J., Lammens, C., Hoang, H. T. T., Pham, N. T., & Goossens, H. (2016). Colistin-resistant *Escherichia coli* harbouring mcr-1 isolated from food animals in Hanoi, Vietnam. *The Lancet Infectious Diseases*, 16(3), 286–287.

Manning, J., Gole, V., & Chousalkar, K. (2015). Screening for *Salmonella* in backyard chickens. *Preventive Veterinary Medicine*, 120(2), 241–245.

Marin, C., Balasch, S., Vega, S., & Lainez, M. (2011). Sources of *Salmonella* contamination during broiler production in Eastern Spain. *Preventive Veterinary Medicine*, 98(1), 39–45.

Marshall, B. M., & Levy, S. B. (2011). Food animals and antimicrobials: Impacts on human health. *Clinical Microbiology Reviews*, 24(4), 718–733.

Matias, C. A. R., Pereira, I. A., Reis, E. M. F. dos, Rodrigues, D. dos P., & Siciliano, S. (2016). Frequency of zoonotic bacteria among illegally traded wild birds in Rio de Janeiro. *Brazilian Journal of Microbiology*, 47(4), 882–888.

Matuschek, E., Åhman, J., Webster, C., & Kahlmeter, G. (2018). Antimicrobial susceptibility testing of colistin – evaluation of seven commercial MIC products against standard broth microdilution for *Escherichia coli* ,

Klebsiella pneumoniae , *Pseudomonas aeruginosa* , and *Acinetobacter spp.*
Clinical Microbiology and Infection, 24(8), 865–870.

- Mba-Jonas, A., Culpepper, W., Hill, T., Cantu, V., Loera, J., Borders, J., Saathoff-Huber, L., Nsubuga, J., Zambrana, I., Dalton, S., Williams, I., & Neil, K. P. (2018). A Multistate outbreak of human *Salmonella* Agona infections associated with consumption of fresh, whole papayas imported from Mexico - United States, 2011. *Clinical Infectious Diseases*, 66(11), 1756–1761.
- McCusker, M. P., Hokamp, K., Buckley, J. F., Wall, P. G., Martins, M., & Fanning, S. (2014). Complete genome sequence of *Salmonella enterica* serovar Agona pulsed-field type SAGOXB.0066, cause of a 2008 pan-European outbreak. *Genome Announcements*, 2(1), e01219–13.
- Meerburg, B. G., & Kijlstra, A. (2007). Role of rodents in transmission of *Salmonella* and *Campylobacter*. *Journal of the Science of Food and Agriculture*, 87(15), 2774–2781.
- Mezal, E. H., Sabol, A., Khan, M. A., Ali, N., Stefanova, R., & Khan, A. A. (2014). Isolation and molecular characterization of *Salmonella enterica* serovar Enteritidis from poultry house and clinical samples during 2010. *Food Microbiology*, 38, 67–74.
- Miao, Z. H., Glatz, P. C., & Ru, Y. J. (2005). Free-range poultry production - A Review. *Asian-Australian Journal of Animal Science*, 18(1), 113–132.
- Mikoleit, M. L. (2014). *Biochemical identification of Salmonella and Shigella using an abbreviated panel of tests* (Laboratory Protocol No. 2010GFNLAB001). World Health Organization, Global Foodborne Infections Network.
- Milligan, R., Paul, M., Richardson, M., & Neuberger, A. (2018). Vaccines for preventing typhoid fever. *Cochrane Database of Systematic Reviews*, 5, Art. No.: CD001261.
- Ministry of health. (2006). *Annual report 2005. The Ministry of Health Malaysia, Kuala Lumpur.*
- Ministry of health. (2009). *Ministry of Health. Malaysia Health facts 2008.* <<http://www.moh.gov>.
- Ministry of health. (2014). *Annual Reports 2004-2013. Planning Division, Health Informatics Centre, Ministry of Health, Malaysia.* www.moh.gov.my
- Ministry of health. (2015). *Health facts 2015. Health Informatics Centre, Planning and Development Division, Ministry of Health Malaysia, Kuala Lumpur.*
- Ministry of health. (2017). Malaysian action plan on antimicrobial resistance (MyAP-AMR) 2017-2021. In *Ministry of Health Malaysia.*

- Modarressi, S., & Thong, K. (2010). Isolation and molecular sub-typing of *Salmonella enterica* from chicken, beef and street foods in Malaysia. *Scientific Research and Essays*, 5(18), 2713–2720.
- Mohsin, M., Raza, S., Roschanski, N., Schaufler, K., & Guenther, S. (2016). First description of plasmid-mediated colistin-resistant extended-spectrum β -lactamase-producing *Escherichia coli* in a wild migratory bird from Asia. *International Journal of Antimicrobial Agents*, 48(4), 463–464.
- Montville, T., & Matthews, K. (2008). *Food microbiology: an introduction* (2nd edition). ASM Press.
- Mouttotou, N., Ahmad, S., Kamran, Z., & Koutoulis, K. C. (2017). Prevalence, risks and antibiotic resistance of *Salmonella* in poultry production Chain. In *Current Topics in Salmonella and Salmonellosis* (pp. 216–234). InTechOpen.
- Murakami, K., Noda, T., Onozuka, D., & Sera, N. (2013). *Salmonella* in liquid eggs and other foods in fukuoka prefecture, Japan. *International Journal of Microbiology*, 2013.
- Murinda, S. E., Nguyen, L. T., Nam, H. M., Almeida, R. A., Headrick, S. J., & Oliver, S. P. (2004). Detection of sorbitol-negative and sorbitol-positive Shiga toxin-producing *Escherichia coli*, *Listeria monocytogenes*, *Campylobacter jejuni*, and *Salmonella* spp. in dairy farm environmental samples. *Foodborne Pathog Dis*, 1(2), 97–104.
- Musa, I. W., Mansur, M. S., Sa'Idu, L., Mohammed, B., & Aliyu, H. B. (2014). Poultry environment and farm practices influencing the isolation rate of multi-drug resistant *Salmonella* from water and poultry feed in Zaria, Nigeria. *Journal of Applied Biology and Biotechnology*, 2(1), 13–16.
- Mustaffa, S. S., Saleha, A. A., & Jalila, A. (2014). Occurrence of antibiotic resistant *Salmonella* and *Campylobacter* in wild birds. *Jurnal Veterinar Malaysia*, 26(1/2), 17–19.
- Nair, D. V. T., & Johny, A. K. (2019). *Salmonella* in poultry meat production. In *Food Safety in Poultry Meat Production* (pp. 1–24). Springer international publishing.
- Nair, D. V. T., Venkitanarayanan, K., & Johny, A. K. (2018). Antibiotic-resistant *Salmonella* in the food supply and the potential role of antibiotic alternatives for control. *Foods*, 7(10), 167.
- Najwa, M. S., Rukayadi, Y., Ubong, A., Loo, Y. Y., Chang, W. S., Lye, Y. L., Thung, T. Y., Aimi, S. A., Malcolm, T. T. H., Goh, S. G., Kuan, C. H., Yoshitsugu, N., Nishibuchi, M., & Son, R. (2015). Quantification and antibiotic susceptibility of *Salmonella* spp., *Salmonella* Enteritidis and *Salmonella* Typhimurium in raw vegetables (ulam). *International Food Research Journal*, 22(5), 1761–1769.

- Nei, M., & Kumar, S. (2000). *Molecular Evolution and Phylogenetics*. Oxford University Press.
- New, C. Y., A., U., Premarathne, J. M. K. J. K., Thung, T. Y., Lee, E., Chang, W. S., Loo, Y. Y., Kwan, S. Y., Tan, C. W., Kuan, C. H., & R., S. (2017). Microbiological food safety in Malaysia from the academician's perspective. *Food Research*, 1(6), 183–202.
- Ni, P., Xu, Q., Yin, Y., Liu, D., Zhang, J., Wu, Q., Tian, P., Shi, X., & Wang, D. (2018). Prevalence and characterization of *Salmonella* serovars isolated from farm products in Shanghai. *Food Control*, 85, 269–275.
- Nidaullah, H., Abirami, N., Shamila-Syuhada, A. K., Chuah, L. O., Huda, N., Tan, T. P., Zainal Abidin, F. W., & Rusul, G. (2017). Prevalence of *Salmonella* in poultry processing environments in wet markets in Penang and Perlis, Malaysia. *Veterinary World*, 10(3), 286–292.
- Noda, T., Murakami, K., Asai, T., Etoh, Y., Ishihara, T., Kuroki, T., Horikawa, K., & Fujimoto, S. (2011). Multi-locus sequence typing of *Salmonella enterica* subsp. *enterica* serovar Enteritidis strains in Japan between 1973 and 2004. *Acta Veterinaria Scandinavica*, 53(1), 38.
- Nor Faiza, S., Saleha, A. A., Jalila, A., & Fauziah, N. (2013). Occurrence of *Campylobacter* and *Salmonella* in ducks and duck eggs in Selangor, Malaysia. *Tropical Biomedicine*, 30(1), 155–158.
- Ohsaki, Y., Hayashi, W., Saito, S., Osaka, S., Taniguchi, Y., Koide, S., Kawamura, K., Nagano, Y., Arakawa, Y., & Nagano, N. (2017). First detection of an *Escherichia coli* strain harboring the mcr-1 gene in retail domestic chicken meat in Japan. *Japanese Journal of Infectious Diseases*, 70(5), 590–592.
- Okoli, I. C., Ndujihe, G. E., & Ogbuwu, I. P. (2006). Frequency of isolation of *Salmonella* from commercial poultry feeds and their anti-microbial resistance profiles, Imo State, Nigeria. *Online Journal of Health and Allied Sciences*, 5, 2.
- Olaitan, A. O., Thongmalayvong, B., Akkhavong, K., Somphavong, S., Paboriboune, P., Khounsy, S., Morand, S., & Rolain, J. M. (2015). Clonal transmission of a colistin-resistant *Escherichia coli* from a domesticated pig to a human in Laos. *The Journal of Antimicrobial Chemotherapy*, 70(12), 3402–3404.
- Ong, L. P., Muniandy, K., How, S. P., Yip, L. S., & Lim, B. K. (2014). *Salmonella* isolation from poultry farms in Malaysia from 2011 to 2013. *Malaysian Journal of Veterinary Research*, 5(June), 1–7.
- Packierisamy, P. R., Haron, R. M. A. R., Mustafa, M., Ahmad Mahir, H. M., Ayob, A., & Balan, V. (2018). Outbreak caused by food-borne *Salmonella enterica* serovar Enteritidis in a residential school in Perak state, Malaysia in April 2016. *International Food Research Journal*, 25(6), 2379–2384.

- Padhi, M. K. (2016). Importance of Indigenous Breeds of chicken for rural economy. *Scientifica*, 2016(Article ID 2604685), 9.
- Pan, H., Zhou, X., Chai, W., Paudyal, N., Li, S., Zhou, X., Zhou, K., Wu, Q., Wu, B., Li, G., Rajkovic, A., Fang, W., Rankin, S. C., Li, Y., Xu, X., Schifferli, D. M., & Yue, M. (2019). Diversified sources for human infections by *Salmonella enterica* serovar Newport. *Transboundary and Emerging Diseases*, 66(2), 1044–1048.
- Partridge, S. R., Pilato, V. Di, Doi, Y., Feldgarden, M., Haft, D. H., Klimke, W., Kumar-Singh, S., Liu, J. H., Malhotra-Kumar, S., Prasad, A., Rossolini, G. M., Schwarz, S., Shen, J., Walsh, T., Wang, Y., & Xavier, B. B. (2018). Proposal for assignment of allele numbers for mobile colistin resistance (*mcr*) genes. *Journal of Antimicrobial Chemotherapy*, 73(10), 2625–2630.
- Patchanee, P., Boonkhot, P., Kittivan, N., Tadee, P., & Chotinun, S. (2015). Dissemination of *Salmonella enterica* sequence types among ASEAN economic community countries. *The Southeast Asian Journal of Tropical Medicine and Public Health*, 46(4), 707–719.
- Perez, F., Endimiani, A., Hujer, K. M., & Bonomo, R. A. (2007). The continuing challenge of ESBLs. *Current Opinion in Pharmacology*, 7(5), 459–469.
- Pérez-Losada, M., Cabezas, P., Castro-Nallar, E., & Crandall, K. A. (2013). Pathogen typing in the genomics era: MLST and the future of molecular epidemiology. *Infection, Genetics and Evolution*, 16, 38–53.
- Peters, T., Bertrand, S., Bjorkman, J. T., Brandal, L. T., Brown, D. J., Erdosi, T., Heck, M., Ibrahim, S., Johansson, K., Kornschöber, C., Kotila, S. M., Le Hello, S., Lienemann, T., Mattheus, W., Nielsen, E. M., Ragimbeau, C., Rumore, J., Sabol, A., Torpdahl, M., Trees, E., Tuohy, A. & de Pinna, E. (2017). Multi-laboratory validation study of multilocus variable number tandem repeat analysis (MLVA) for *Salmonella enterica* serovar Enteritidis, 2015. *Eurosurveillance*, 22(9), 1–8.
- Pethlerdprao, P., Supa-amornkul, S., Panvisavas, N., & Chaturongakul, S. (2017). *Salmonella enterica* multilocus sequence typing and its correlation with serotypes. *Food Biotechnology*, 31(2), 73–79.
- Poirel, L., Jayol, A., & Nordmann, P. (2017). Polymyxins: Antibacterial activity, susceptibility testing, and resistance mechanisms encoded by plasmids or chromosomes. *Clinical Microbiology Reviews*, 30(2), 557–596.
- Poole, K. (2005). Aminoglycoside resistance in *Pseudomonas aeruginosa*. *Antimicrobial Agents and Chemotherapy*, 49(2), 479–487.
- Poonlaphdecha, S., Ribas, A., Saijuntha, W., Prantlová, V., & Agatsuma, T. (2016). Rodents as a source of *Salmonella* contamination in wet markets in Thailand. *Vector-Borne and Zoonotic Diseases*, 16(8), 537–540.

- Popoff, M. Y., Bockemühl, J., & Gheesling, L. L. (2003). Supplement 2001 (no. 45) to the Kauffmann-White scheme. *Research in Microbiology*, 154(3), 173–174.
- Popoff, M. Y., & Le Minor, L. (2005). *Salmonella*. In D. J. Brenner, N. R. Krieg, & J. T. Staley (Eds.), *Bergey's Manual of Systemic Bacteriology*. (2nd ed., pp. 764–799). Springer.
- Poppe, C., Johnson, R. P., Forsberg, C. M., & Irwin, R. J. (1992). *Salmonella* Enteritidis and other *Salmonella* in laying hens and eggs from flocks with *Salmonella* in their environment. *Canadian Journal of Veterinary Research*, 56(3), 226–232.
- Prescott, J., Young, O., O'Neill, L., Yau, N. J. N., & Stevens, R. (2002). Motives for food choice: A comparison of consumers from Japan, Taiwan, Malaysia and New Zealand. *Food Quality and Preference*, 13(7-8), 489–495.
- Prestinaci, F., Pezzotti, P., & Pantosti, A. (2015). Antimicrobial resistance: a global multifaceted phenomenon. *Pathogens and Global Health*, 109(7), 309–318.
- Rahman, W. A., & Haziqah, F. (2015). Ectoparasitic fauna of scavenging chickens (*Gallus domesticus*) from Penang island, Peninsular Malaysia. *Malaysian Journal of Veterinary Research*, 6(1), 33–42.
- Ramey, A. M., Hernandez, J., Tyrlöv, V., Uher-Koch, B. D., Schmutz, J. A., Atterby, C., Järhult, J. D., & Bonnedahl, J. (2018). Antibiotic-resistant *Escherichia coli* in migratory birds inhabiting remote Alaska. *EcoHealth*, 15(1), 72–81.
- Ramlah, A. H. (1999). Production aspects of village chicken in South-east Asian region. *International Network for Family Poultry Development, INFPDE - Conference*, 61–76.
- Randall, L. P., Cooles, S. W., Osborn, M. K., Piddock, L. J. V., & Woodward, M. J. (2004). Antibiotic resistance genes, integrons and multiple antibiotic resistance in thirty-five serotypes of *Salmonella enterica* isolated from humans and animals in the UK. *Journal of Antimicrobial Chemotherapy*, 53(2), 208–216.
- Ranjbar Malidareh, N., Firouzi, S., Ranjbar Malidareh, N., & Habibi, H. (2013). In vitro and in vivo susceptibility of *Salmonella spp.* isolated from broiler chickens. *Comparative Clinical Pathology*, 22(6), 1065–1068.
- Ranjbar, R., Elhaghi, P., & Shokoohizadeh, L. (2017). Multilocus sequence typing of the clinical isolates of *Salmonella enterica* serovar Typhimurium in Tehran hospitals. *Iranian Journal of Medical Sciences*, 42(5), 443–448.
- Ranjbar, R., Karami, A., Farshad, S., Giammanco, G. M., & Mammina, C. (2014). Typing methods used in the molecular epidemiology of microbial pathogens:

a how-to guide. *New Microbiol*, 37(1), 1–15.

- Reeves, M. W., Evins, G. M., Heiba, A. A., Plikaytis, B. D., & Farmer, J. J. (1989). Clonal nature of *Salmonella* Typhi and its genetic relatedness to other *Salmonella* as shown by multilocus enzyme electrophoresis and proposal of *Salmonella bongori* comb. nov. *Journal of Clinical Microbiology*, 27(2), 313–320.
- Rejab, S. B. M., Shariruzi, H., Desa, Z. M., & Abidin, S. A. Z. (2019). Retrospective study on persistent *Salmonella* serotypes in meat samples tested in the veterinary public health section, regional of veterinary laboratory Bukit Tengah, Penang. *Malaysian Journal of Veterinary Research*, 10(1), 34–42.
- Ribas, A., Saijuntha, W., Agatsuma, T., Prantlová, V., & Poonlaphdecha, S. (2016). Rodents as a source of *Salmonella* contamination in Wet Markets in Thailand. *Vector-Borne and Zoonotic Diseases*, 16(8), 537–540.
- Rodriguez, A., Pangloli, P., Richards, H. A., Mount, J. R., & Draughon, F. A. (2006). Prevalence of *Salmonella* in diverse environmental farm samples. *Journal of Food Protection*, 69(11), 2576–2580.
- Rodríguez, I., Barownick, W., Helmuth, R., Mendoza, M. C., Rodicio, M. R., Schroeter, A., & Guerra, B. (2009). Extended-spectrum β -lactamases and AmpC β -lactamases in ceftiofur-resistant *Salmonella enterica* isolates from food and livestock obtained in Germany during 2003-07. *Journal of Antimicrobial Chemotherapy*, 64(2), 301–309.
- Rolain, J.-M., & Olaitan, A. O. (2016). Plasmid-mediated colistin resistance: the final blow to colistin? *International Journal of Antimicrobial Agents*, 47(1), 4–5.
- Rose, N., Beaudreau, F., Drouin, P., Toux, J. Y., Rose, V., & Colin, P. (1999). Risk factors for *Salmonella enterica* subsp. *enterica* contamination in French broiler-chicken flocks at the end of the rearing period. *Preventive Veterinary Medicine*, 39(4), 265–277.
- Roseliza, R., Maswati, M. A., Hasnah, Y., & Ramlan, M. (2011). Identification of *Salmonella* serotypes isolated from meat samples in Malaysia. *Malaysian Journal of Veterinary Research*, 2, 59–64.
- Rossi, F., Girardello, R., Cury, A. P., Di Gioia, T. S. R., Almeida, J. N. de, & Duarte, A. J. da S. (2017). Emergence of colistin resistance in the largest university hospital complex of São Paulo, Brazil, over five years. *Brazilian Journal of Infectious Diseases*, 21(1), 98–101.
- Rotger, R., & Casadesús, J. (1999). The virulence plasmids of *Salmonella*. *International Microbiology*, 2(3), 177–184.

- Russo, E., Biggerstaff, G., Hoekstra, R., Meyer, S., Patel, N., & Miller, B. et al. (2013). A recurrent, multistate outbreak of *Salmonella* serotype agona infections associated with dry, unsweetened cereal consumption, United States, 2008. *J Food Prot*, 76(2), 227–230.
- Rusul, G., Khair, J., Radu, S., Cheah, C. T., & Yassin, R. M. (1996). Prevalence of *Salmonella* in broilers at retail outlets, processing plants and farms in Malaysia. *International Journal of Food Microbiology*, 33(2-3), 183–194.
- Saad, A. M., Almujaali, D. M., Babiker, S. H., Shuaib, M. A. M., Abdelgadir, K. A., & Alfadul, Y. A. (2007). Prevalence of *Salmonella* in broiler chicken carcasses and poultry farms in the central region, K.S.A. *Journal of Animal and Veterinary Advances*, 6(2).
- Sabat, A. J., Budimir, A., Nashev, D., Sá-Leão, R., van Dijl, J. M., Laurent, F., Grundmann, H., Friedrich, A. W., & ESCMID Study Group of Epidemiological Markers (ESGEM). (2013). Overview of molecular typing methods for outbreak detection and epidemiological surveillance. *European Communicable Disease Bulletin*, 18(4), 20380.
- Sabat, A. J., Chlebowicz, M. A., Grundmann, H., Arends, J. P., Kampinga, G., Meessen, N. E. L., Friedrich, A. W., & Van Dijn, J. M. (2012). Microfluidic-chip-based multiple-locus variable-number tandem-repeat fingerprinting with new primer sets for methicillin-resistant *Staphylococcus aureus*. *Journal of Clinical Microbiology*, 50(7), 2255–2262.
- Safman, R. M. (2010). Avian influenza control in Thailand: balancing the interests of different poultry producers. In I. Scoones (Ed.), *Avian influenza: science, policy and politics* (pp. 169–206). Earthscan.
- Sahilah, A. M., Son, R., Rusul, G., Samuel, L., Hassan, Z., Lum, K. Y., & Ahmad, M. A. (2000). Molecular typing of *Salmonella* Weltevreden and *Salmonella* Chincol by pulsed-field gel electrophoresis (PFGE) and enterobacterial repetitive intergenic consensus-polymerase chain reaction (ERIC-PCR). *World Journal of Microbiology and Biotechnology*, 16(7), 621–624.
- Saitou, N., & Nei, M. (1987). The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol*, 4, 406–425.
- Salleh, N. A., Rusul, G., Hassan, Z., Reezal, A., Hajar Isa, S., Nishibuchi, M., & Radu, S. (2003). Incidence of *Salmonella* spp. in raw vegetables in Selangor, Malaysia. *Food Control*, 14(7), 475–479.
- Salleh, W., Lani, M. N., Abdullah, W. Z. W., Chilek, T. Z. T., & Hassan, Z. (2017). A review on incidences of foodborne diseases and interventions for a better national food safety system in Malaysia. *Malaysian Applied Biology*, 46(3), 1–7.

- Samanta, I., Joardar, S. N., Das, P. K., Sar, T. K., Bandyopadhyay, S., Dutta, T. K., & Sarkar, U. (2014). Prevalence and antibiotic resistance profiles of *Salmonella* serotypes isolated from backyard poultry flocks in West Bengal, India. *Journal of Applied Poultry Research*, 23(3), 536–545.
- Sanganyado, E., & Gwenzi, W. (2019). Antibiotic resistance in drinking water systems: Occurrence, removal, and human health risks. *Science of The Total Environment*, 669, 785–797.
- Saravanan, S., Purushothaman, V., Ramasamy, T., Krishna, G., Sukumar, K., Srinivasan, P., Gowthaman, V., Balusamy, M., Atterbury, R., & Kuchipudi, S. V. (2015). Molecular epidemiology of non-typhoidal *Salmonella* in poultry and poultry Products in India : Implications for human health. *Indian Journal of Microbiology*, 55(3), 319–326.
- Sasipreeyajan, J., Jerngklinchan, J., Koowatananukul, C., & Saitanu, K. (1996). Prevalence of *Salmonella* in broiler, layer and breeder flocks in Thailand. *Tropical Animal Health and Production*, 28(2), 174–180.
- Schmidt, H., & Hensel, M. (2004). Pathogenicity islands in bacterial pathogenesis. *Clinical Microbiology Reviews*, 17(1), 14–56.
- Schürch, A. C., & van Soolingen, D. (2012). DNA fingerprinting of Mycobacterium tuberculosis: from phage typing to whole-genome sequencing. *Infection, Genetics and Evolution*, 12(4), 602–609.
- Sefton, M. A. (2002). Mechanisms of antimicrobial resistance: Their clinical relevance in the new millenium. *Drugs*, 62, 557–566.
- Sengupta, S., Chattopadhyay, M. K., & Grossart, H.-P. (2013). The multifaceted roles of antibiotics and antibiotic resistance in nature. *Frontiers in Microbiology*, 4.
- Shah, D. H., Paul, N. C., Sischo, W. C., Crespo, R., & Guard, J. (2016a). Microbiology and food safety: Population dynamics and antimicrobial resistance of the most prevalent poultry-associated *Salmonella* serotypes. *Poultry Science*, 96(3), 687–702.
- Shang, K., Wei, B., & Kang, M. (2018). Distribution and dissemination of antimicrobial-resistant *Salmonella* in broiler farms with or without enrofloxacin use. *BMC Veterinary Research*, 14(1), 257.
- Sharma, J., Kumar, D., Hussain, S., Pathak, A., Shukla, M., Prasanna Kumar, V., Anisha, P. N., Rautela, R., Upadhyay, A. K., & Singh, S. P. (2019). Prevalence, antimicrobial resistance and virulence genes characterization of nontyphoidal *Salmonella* isolated from retail chicken meat shops in Northern India. *Food Control*, 102, 104–111.

- Shobrak, M. Y., & Abo-Amer, A. E. (2014). Role of wild birds as carriers of multi-drug resistant *Escherichia coli* and *Escherichia vulneris*. *Brazilian Journal of Microbiology*, 45(4), 1199–1209.
- Singh, R., Yadav, A. S., Tripathi, V., & Singh, R. P. (2013). Antimicrobial resistance profile of *Salmonella* present in poultry and poultry environment in north India. *Food Control*, 33(2), 545–548.
- Smith, K. F., & Guégan, J.-F. (2010). Changing geographic distributions of human pathogens. *Annual Review of Ecology, Evolution, and Systematics*, 41(1), 231–250.
- Somasundram, C., Razali, Z., & Santhirasegaram, V. (2016). A review on organic food production in Malaysia. *Horticulturae*, 2(3), 12.
- Soon, J. M., Singh, H., & Baines, R. (2011). Foodborne diseases in Malaysia: A review. *Food Control*, 22(6), 823–830.
- St. Amand, J. A., Cassis, R., King, R. K., & Annett Christianson, C. B. (2017). Prevalence of *Salmonella* spp. in environmental samples from table egg barns in Alberta. *Avian Pathology*, 46(6), 594–601.
- Stegniy, B., Gerilovych, A., Arefyev, V., Glebova, K., & Potkonjak, A. (2014). A method for detecting and typing of *Salmonella* by multiplex PCR. *Arhiv Veterinarske Medicine*, 7(2), 47–56.
- Stepan, R. M., Sherwood, J. S., Petermann, S. R., & Logue, C. M. (2011). Molecular and comparative analysis of *Salmonella enterica* Senftenberg from humans and animals using PFGE, MLST and NARMS. *BMC Microbiology*, 11, 153.
- Stoesser, N., Mathers, A. J., Moore, C. E., Day, N. P., & Crook, D. W. (2016). Colistin resistance gene *mcr-1* and pHNSHP45 plasmid in human isolates of *Escherichia coli* and *Klebsiella pneumoniae*. *The Lancet Infectious Diseases*, 16(3), 285–286.
- Ström Hallenberg, G., Börjesson, S., Sokerya, S., Sothyra, T., & Magnusson, U. (2019). Detection of *mcr*-mediated colistin resistance in *Escherichia coli* isolates from Pigs in small-scale farms in Cambodia. *Antimicrobial Agents and Chemotherapy*, 63(3).
- Su, S., Bi, Y., Wong, G., Gray, G. C., Gao, G. F., & Li, S. (2015). Epidemiology, evolution, and recent outbreaks of avian influenza virus in China. *Journal of Virology*, 89(17), 8671–8676.
- Suhaila, A. H., Sabrina, D. L., Nik Ahmad Irwan Izzauidin, N. H., Hamdan, A., & Khadijah, S. (2015). Study of parasites in commercial free-range chickens in northern Peninsular Malaysia. *Malaysian Journal of Veterinary Research*, 6(2), 53–64.

- Tabo, D. adjim, Diguimbaye, C. D., Granier, S. A., Moury, F., Brisabois, A., Elgroud, R., & Millemann, Y. (2013). Prevalence and antimicrobial resistance of non-typhoidal *Salmonella* serotypes isolated from laying hens and broiler chicken farms in N'Djamena, Chad. *Veterinary Microbiology*, 166(1-2), 293–298.
- Taitt, C. R., Shubin, Y. S., Angel, R., & Ligler, F. S. (2004). Detection of *Salmonella enterica* serovar typhimurium by using a rapid, array-based immunosensor. *Applied and Environmental Microbiology*, 70(1), 152–158.
- Tan, T. Y., & Ng, S. Y. (2006). The in-vitro activity of colistin in gram-negative bacteria. *Singapore Medical Journal*, 47(7), 621–624.
- Tang, K. L., Caffrey, N. P., Nóbrega, D. B., Cork, S. C., Ronksley, P. E., Barkema, H. W., Polachek, A. J., Ganshorn, H., Sharma, N., Kellner, J. D., & Ghali, W. A. (2017). Restricting the use of antibiotics in food-producing animals and its associations with antibiotic resistance in food-producing animals and human beings: a systematic review and meta-analysis. *The Lancet Planetary Health*, 1(8), e316–e327.
- Tauxe, R. V., Doyle, M. P., Kuchenmüller, T., Schlundt, J., & Stein, C. E. (2010). Evolving public health approaches to the global challenge of foodborne infections. *International Journal of Food Microbiology*, 139, 16–28.
- Thakur, S., Brake, J., Keelara, S., Zou, M., & Susick, E. (2013). Farm and environmental distribution of *Campylobacter* and *Salmonella* in broiler flocks. *Research in Veterinary Science*, 94(1), 33–42.
- Thanh, L. T., Phan, T. H., Rattanavong, S., Nguyen, T. M., Duong, A. Van, Dacon, C., Hoang, T. N., Nguyen, L. P. H., Tran, C. T. H., Davong, V., Nguyen, C. V. V., Thwaites, G. E., Boni, M. F., Dance, D., Ashton, P. M., & Day, J. N. (2019). Multilocus sequence typing of *Cryptococcus neoformans* var. *grubii* from Laos in a regional and global context. *Medical Mycology*, 57(5), 557–565.
- Thong, K. L., Goh, Y. L., Radu, S., Noorzaleha, S., Yasin, R., Koh, Y. T., Lim, V. K. E., Rusul, G., & Puthuchery, S. D. (2002). Genetic diversity of clinical and environmental strains of *Salmonella enterica* serotype Weltevreden isolated in Malaysia. *Journal of Clinical Microbiology*, 40(7), 2498–2503.
- Thung, T. Y., Mahyudin, N. A., Basri, D. F., Wan Mohamed Radzi, C. W. J., Nakaguchi, Y., Nishibuchi, M., & Radu, S. (2016). Prevalence and antibiotic resistance of *Salmonella* Enteritidis and *Salmonella* Typhimurium in raw chicken meat at retail markets in Malaysia. *Poultry Science*, 95(8), 1888–1893.
- Thung, T. Y., Radu, S., Mahyudin, N. A., Rukayadi, Y., Zakaria, Z., Mazlan, N., Tan, B. H., Lee, E., Yeoh, S. L., Chin, Y. Z., Tan, C. W., Kuan, C. H., Basri, D. F., & Wan Mohamed Radzi, C. W. J. (2018). Prevalence, virulence genes

and antimicrobial resistance profiles of *Salmonella* serovars from retail beef in Selangor, Malaysia. *Frontiers in Microbiology*, 8.

- Trung, N. V., Carrique-Mas, J. J., Nghia, N. H., Tu, L. T. P., Mai, H. H., Tuyen, H. T., Campbell, J., Nhung, N. T., Nhung, H. N., Minh, P. V., Chieu, T. T. B., Hieu, T. Q., Mai, N. T. N., Baker, S., Wagenaar, J. A., Hoa, N. T., & Schultsz, C. (2017). Non-typhoidal *Salmonella* colonization in chickens and humans in the Mekong Delta of Vietnam. *Zoonoses and Public Health*, 64(2), 94–99.
- Tsai, H. J., & Hsiang, P. H. (2004). The Prevalence and antimicrobial susceptibilities of *Salmonella* and *Campylobacter* in ducks in Taiwan. *Journal of Veterinary Medical Science*, 67(1), 7–12.
- Tyrrell, C., Burgess, C. M., Brennan, F. P., & Walsh, F. (2019). Antibiotic resistance in grass and soil. *Biochemical Society Transactions*, 47(1), 477–486.
- United States Department of Agriculture. (2013). *Foodborne illness cost calculator: Salmonella*. <https://www.ers.usda.gov/amber-waves/2013/november/recent-estimates-of-the-cost-of-foodborne-illness-are-in-general-agreement/>
- Van Asten, A. J. A. M., & Van Dijk, J. E. (2005). Distribution of “classic” virulence factors among *Salmonella* spp. *FEMS Immunology and Medical Microbiology*, 44(3), 251–259.
- Van, T. T. H., Nguyen, H. N. K., Smooker, P. M., & Coloe, P. J. (2012). The antibiotic resistance characteristics of non-typhoidal *Salmonella enterica* isolated from food-producing animals, retail meat and humans in South East Asia. *International Journal of Food Microbiology*, 154(3), 98–106.
- Vanselow, B. A., Hornitzky, M. A., Walker, K. H., Eamens, G. J., Bailey, G. D., Gill, P. A., Coates, K., Corney, B., Cronin, J. P., & Renilson, S. (2007). *Salmonella* and on-farm risk factors in healthy slaughter-age cattle and sheep in eastern Australia. *Australian Veterinary Journal*, 85(12), 498–502.
- Voss-Rech, D., Vaz, C. S. L., Alves, L., Coldebella, A., Leao, J. A., Rodrigues, D. P., & Back, A. (2015). A temporal study of *Salmonella enterica* serotypes from broiler farms in Brazil. *Poultry Science*, 94(3), 433–441.
- Walsh, T. R., & Wu, Y. (2016). China bans colistin as a feed additive for animals. *The Lancet Infectious Diseases*, 16(10), 1102–1103.
- Wang, X., Wang, Y., Zhou, Y., Li, J., Yin, W., Wang, S., Zhang, S., Shen, J., Shen, Z., & Wang, Y. (2018). Emergence of a novel mobile colistin resistance gene, mcr-8, in NDM-producing *Klebsiella pneumoniae*. *Emerging Microbes and Infections*, 7(1).
- Wang, X., Wang, Y., Zhou, Y., Wang, Z., Wang, Y., Zhang, S., & Shen, Z. (2019). Emergence of colistin resistance gene mcr-8 and its variant in *Raoultella*

ornithinolytica. *Frontiers in Microbiology*, 10, 228.

- Wang, Y., Yang, B., Wu, Y., Zhang, Z., Meng, X., Xi, M., Wang, X., Xia, X., Shi, X., Wang, D., & Meng, J. (2015). Molecular characterization of *Salmonella enterica* serovar Enteritidis on retail raw poultry in six provinces and two national cities in China. *Food Microbiology*, 46, 74–80.
- Welch, T. J., Fricke, W. F., McDermott, P. F., White, D. G., Rosso, M. L., Rasko, D. A., Mammel, M. K., Eppinger, M., Rosovitz, M. J., Wagner, D., Rahalison, L., Leclerc, J. E., Hinshaw, J. M., Lindler, L. E., Cebula, T. A., Carniel, E., & Ravel, J. (2007). Multiple antimicrobial resistance in plague: an emerging public health risk. *PLoS ONE*, 2, e309.
- Whelan, J., Noel, H., Friesema, I., Hofhuis, A., de Jager, C. M., Heck, M., Heuvelink, A., & van Pelt, W. (2010). National outbreak of *Salmonella* Typhimurium (Dutch) phage-type 132 in the Netherlands, October to December 2009. *Eurosurveillance*, 15(44), 1–5.
- White, P. B. (1925). Med. Res. Council, Gt. Brit. *Spec. Rep. Ser. No. 91*.
- Wilkinson, J., & Boxall, A. (2019). The first global study of pharmaceutical contamination in riverine environments. *SETAC Europe 29th Annual Meeting, Helsinki, Finland*.
- World Health Organization. (2012). *Critically important antimicrobials for human medicine*. WHO, Geneva, Switzerland. http://apps.who.int/iris/bitstream/10665/77376/1/9789241504485_eng.pdf.
- World Health Organization. (2015). *World health statistics 2015*. Geneva: World Health Organization; 2015.
- Xiong, W., Sun, Y., & Zeng, Z. (2018). Antimicrobial use and antimicrobial resistance in food animals. *Environmental Science and Pollution Research*, 25(19), 18377–18384.
- Xu, Y., Tao, S., Hinkle, N., Harrison, M., & Chen, J. (2018). *Salmonella*, including antibiotic-resistant *Salmonella*, from flies captured from cattle farms in Georgia, U.S.A. *Science of the Total Environment*, 616-617, 90–96.
- Yamaguchi, T., Kawahara, R., Harada, K., Teruya, S., Nakayama, T., Motooka, D., Nakamura, S., Do Nguyen, P., Kumeda, Y., Dang, C. Van, Hirata, K., & Yamamoto, Y. (2018). The presence of colistin resistance gene *mcr-1* and *-3* in ESBL producing *Escherichia coli* isolated from food in Ho Chi Minh City, Vietnam. *FEMS Microbiology Letters*, 365(11).
- Yang, J., Gao, S., Chang, Y., Su, M., Xie, Y., & Sun, S. (2019). Occurrence and characterization of *Salmonella* isolated from large-scale breeder farms in Shandong Province, China. *BioMed Research International*, 2019, 1–8.

- Yasin, R. M., Jegathesan, M., & Tiew, C. C. (1997). *Salmonella* serotypes isolated in Malaysia over the ten-year period 1983-1992. *Asia-Pacific Journal of Public Health*, 9(1), 1–5.
- Yasin, R. M., Tiew, C. C., & Jegathesan, M. (1995). Human salmonellosis in Malaysia for the period 1989 - July 1994. *Southeast Asian J Trop Med Public Health*, 26, 457–460.
- Yassin, A. K., Zhang, J., Wang, J., Chen, L., Kelly, P., Butaye, P., Lu, G., Gong, J., Li, M., Wei, L., Wang, Y., Qi, K., Han, X., Price, S., Hathcock, T., & Wang, C. (2017). Identification and characterization of mcr mediated colistin resistance in extraintestinal *Escherichia coli* from poultry and livestock in China. In *FEMS Microbiology Letters* (Vol. 364, Issue 24).
- Yildirim, Y., Gonulalan, Z., Pamuk, S., & Ertas, N. (2011). Incidence and antibiotic resistance of *Salmonella* spp. on raw chicken carcasses. *Food Research International*, 44(3), 725–728.
- Yoke-Kqueen, C., Learn-Han, L., Noorzaleha, A. S., Son, R., Sabrina, S., Jiun-Horng, S., & Chai-Hoon, K. (2008). Characterization of multiple-antimicrobial-resistant *Salmonella enterica* subsp. *enterica* isolated from indigenous vegetables and poultry in Malaysia. *Letters in Applied Microbiology*, 46(3), 318–324.
- Yusoff, S. S. B. M., Arshad, M. M. B., & Woldergiorgis, E. A. (2016). *Antibiotic resistance patterns and genetic relatedness of Salmonella serotypes isolated from chicken carcasses of retail markets and slaughterhouses in Kota Bharu, Kelantan*. (Unpublished MSc. Thesis). Universiti Malaysia Kelantan, Kelantan.
- Zamora-Sanabria, R., & Alvarado, A. M. (2017). Preharvest *Salmonella* Risk Contamination and the Control Strategies. In *Current Topics in Salmonella and Salmonellosis* (pp. 193–213). InTechOpen.
- Zhang, G., Thau, E., Brown, E. W., & Hammack, T. S. (2013). Comparison of a novel strategy for the detection and isolation of *Salmonella* in shell eggs with the food and drug administration bacteriological analytical manual method. *Poultry Science*, 92(12), 3266–3274.
- Zhang, L., Fu, Y., Xiong, Z., Ma, Y., Wei, Y., Qu, X., Zhang, H., Zhang, J., & Liao, M. (2018). Highly prevalent multidrug-resistant *Salmonella* from chicken and pork meat at retail markets in Guangdong, China. *Frontiers in Microbiology*, 9(2018), 2104.
- Zhao, X., Gao, Y., Ye, C., Yang, L., Wang, T., & Chang, W. (2016). Prevalence and Characteristics of *Salmonella* isolated from free-range chickens in Shandong Province, China. *BioMed Research International*, 2016, 1–6.