



***MORPHOLOGICAL CHANGES IN LIVER CELLS OF
HYPERCHOLESTEROLAEMIC-INDUCED RATS AND HEPG2 CELL LINES
ASSOCIATED WITH EDIBLE BIRD'S NEST SUPPLEMENTATION***

MOHD AKMAL BIN MOHD NOOR

FPV 2020 8



**MORPHOLOGICAL CHANGES IN LIVER CELLS OF
HYPERCHOLESTEROLAEMIC-INDUCED RATS AND HEPG2 CELL
LINES ASSOCIATED WITH EDIBLE BIRD'S NEST SUPPLEMENTATION**

By

MOHD AKMAL BIN MOHD NOOR

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia
in the Fulfilment of Requirements for the degree of Doctor of Philosophy**

February 2020

COPYRIGHT

All material contained within the thesis, including without limitation texts, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purpose from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

**MORPHOLOGICAL CHANGES IN LIVER CELLS OF
HYPERCHOLESTEROLAEMIC-INDUCED RATS AND HEPG2 CELL
LINES ASSOCIATED WITH EDIBLE BIRD'S NEST SUPPLEMENTATION**

By

MOHD AKMAL BIN MOHD NOOR

February 2020

Chairman : Assoc. Prof. Intan Shameha binti Abdul Razak, DVM, PhD
Faculty : Veterinary Medicine

Edible bird's nest (EBN) is an animal-based natural product that has high interest in Chinese committee. Since in the Tang Dynasty (618-907 CE), it had been consumed frequently by the royal families for well-being purposes. This polymerized swiftlet's salivary secretion is mainly composed of protein that presence in the form of glycoprotein. It has been anecdotally claimed to have broad medicinal benefits, including metabolic stimulant. Nonetheless, the effect of EBN supplementation on the cholesterol metabolism with scientific evidence is poorly elucidated. Therefore, we hypothesised the EBN supplementation is able to improve cholesterol metabolism in HepG2 cell lines and hypercholesterolaemic-induced rats. This study focuses on evaluating the effect of EBN supplementation on the cholesterol metabolism via assessing the relevant genes expression, quantifying hepatic cholesterol concentration, measuring blood lipid profiling, localizing important structure and receptor in cholesterol metabolism, and assessing hepatic and extra-hepatic microscopic changes in the HepG2 and hypercholesterolaemic-induced rats. Cellular toxicity assessment of the edible bird's nest extract (EBNE) at different concentrations (0.2, 0.5, 0.8, 1.0 and 1.5 mg/mL) was done and revealed the cell viability remains as high as 71% even at the highest concentration (1.5 mg/mL) after 24 hours incubation. Thus, in the subsequent assay the Hep-G2 was supplemented with EBNE at three different concentrations (0.5, 1.0 and 1.5 mg/mL) in high lipid media [exogenous lipid (1:500) and cholesterol (1:250)] for 24 hours. Besides that, there were three control groups including baseline control (BC) that cultured in AMEM media only, negative control (NC) in high lipid media, and positive control (PC) in high lipid media with Simvastatin (4.60 µg/mL) as an anti-cholesterol drug. Quantification of gene expression for both low-density lipoprotein receptor (LDLR) and acyl Coenzyme A: cholesterol acyltransferase 2 (ACAT2) were significantly up regulated in a dose-dependent manner. However, 3-hydroxyl-3-metylglutaryl coenzyme A reductase (HMGCR) was observed significantly to down-regulate this enzyme at the highest dose of EBNE (1.5 mg/mL). Quantitatively, distribution of immunofluorescence intensity against LDLR protein and lipid droplets (LDs) was increased as the EBNE concentration increased. Total cholesterol storage was showing the amount of cholesterol stored in the LDs increased as the concentration of EBNE increased. In

the 12 weeks *in-vivo* study, a total of 30 male Sprague Dawley rats aged 10-week old were randomly assigned into five different groups (n=6); BC (normal rat diet), NC [hypercholesterolaemia induced with high-fat diet (HFD) and Triton-X 100 (150 mg/kg SQ q42d)], PC (hypercholesterolaemia with Simvastatin 10 mg/kg PO SID), EBNE [hypercholesterolaemia with EBNE (6.5 mg/kg PO SID)] and EBNS [hypercholesterolaemia with EBNS (843.2 mg/kg PO SID)]. Post-treatment blood lipid profiling was showing the EBNS group significantly reduced the triglycerides (TAG), total cholesterol (TC), low-density lipoprotein (LDL) and very low-density lipoprotein (VLDL), compared to the NC group and but only TAG and LDL significantly reduced up to BC group. Meanwhile, the EBNE group showed significant reduction for TC and LDL only compared to NC but not reduced up to the BC group. Statistically, both EBNS and EBNE group were showing significant elevation of high-density lipoprotein (HDL) compared to the NC and BC group but not as high as in the PC group. Nonetheless, the HDL of EBNS group had two-fold significant increment compared to EBNE group. Remarkably, cardiogenic indices that predicting cardiac diseases and atherosclerosis occurrence showed a significant protective effect in the EBNS group only, which comparable with PC's indices. The hepatic cholesterol concentrations in the EBNS and EBNE were significantly reduced as compared to the NC. Gene expression revealed EBNS significantly down-regulated HMGCR and proprotein convertase subtilisin/kexin 9 (PCSK9) which stimulating and maintaining the upregulation of LDLR via cleavage of sterol regulatory element-binding protein 2 (SREBP2). Concomitantly, EBNS up-regulated also the cytochrome P450 family 7 subfamily A member 1 (CYP7a1) for bile production. On the other hand, EBNE was observed to up-regulate the expression of LDLR via cleavage of SREBP2 only. Grossly, liver of rat fed on EBNS appeared mild yellowish discolouration, meanwhile the EBNE group was observed moderate yellowish discolouration. Histological examination of the liver revealed mild hepatic steatosis in the EBNS group and mild non-alcoholic steatohepatitis (NASH) in EBNE group. This finding was consistent with the ultrastructure finding (TEM), whereby the mitochondria of EBNS were demonstrated enlarged with intact mitochondrial membrane, meanwhile the mitochondria in the EBNE group showed swollen with loss of cristae and translucent matrix indicating liver injury. The hepatic tissue sections were also immunologically-labelled with fluorescence against the LDLR and demonstrated high intensity expression of LDLR in the EBNS (5.65 ± 0.12), which statistically equivalent to the PC (5.53 ± 0.17) group. Meanwhile, EBNE (3.54 ± 0.04) group appeared lower LDLR distribution compared to EBNS group, but ameliorated than NC (1.81 ± 0.06). Quantitative SEM revealed the cranial thoracic aorta of NC and EBNE were significantly occluded the aortic lumen up to 31% and 30%, respectively; compared to the BC (0%). Meanwhile, EBNS documented insignificant aortic lumen occlusion (<1%). These outcomes were consistent with degree of immunoreactivity in the CD40 immunoperoxidase assay, in which detecting atherosclerosis biomarker. Renal tissue demonstrated absence of glomerulonephritis in the EBNS group, but prominently observed in the EBNE with thickening of the glomerular basement membrane. These findings were in line with detection of the renal inflammasome (NLRP3) in the tubules of EBNE group which covered up to 61% of the tissue section, compared to the EBNS which less than 17% immunoreactivity. Concurrently, inflammatory cells in the renal interstitial space was showing mild and moderate infiltration in respective group. As a conclusion, EBNE shows significant cholesterol improvement in the *in-vitro* study not in the *in-vivo* model. Nonetheless, in the hypercholesterolaemic-induced rats, EBNS was

profoundly exhibited a significant effect in improving cholesterol metabolism and slowing progression of non-alcoholic fatty liver disease (NAFLD), atherosclerosis and chronic kidney disease (CKD).



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PERUBAHAN MORFOLOGIKAL PADA SEL HATI TIKUS TERINDUKSI-
HIPERKOLESTEROLEMIA DAN TISU KULTUR HEPG2 YANG
DITAMBAHAN DENGAN SARANG BURUNG WALET**

Oleh

MOHD AKMAL BIN MOHD NOOR

Februari 2020

Pengerusi : Prof. Madya Intan Shameha binti Abdul Razak, DVM, PhD
Fakulti : Perubatan Veterinar

Sarang burung walet (EBN) merupakan produk semulajadi berasaskan haiwan yang mempunyai nilai tersendiri dalam kalangan masyarakat Cina. Sejak zaman pemerintahan Dinasti Tang (618-907 SM), ia menjadi makanan kesihatan alternatif yang lazim bagi golongan bangsawan diraja. Produk ini merupakan cecair liur burung walit terpolimer yang terdiri daripada protein sebagai kandungan utama dan wujud di dalam bentuk. Secara tradisionalnya, sarang burung walit dipercayai mempunyai kebaikan perubatan seperti agen penggalak metabolisme. Namun demikian, kesan pengambilan EBN terhadap metabolisme kolesterol masih kurang dibincangkan secara saintifik. Oleh itu, kajian ini menghipotesiskan pengambilan EBN mampu meningkatkan metabolisme kolesterol di dalam pengkulturan HepG2 dan tikus terinduksi hiperkolesterolemia. Penyelidikan ini memfokuskan kesan pengambilan EBN terhadap metabolisme kolesterol melalui penilaian pengekspresian gen-gen berkaitan, pengkuantitian kepekatan kolesterol di dalam hati, analisis profil lemak darah, pengesanan struktur dan reseptor penting di dalam metabolisme kolesterol, dan pemeriksaan perubahan mikroskopik hepatik dan ekstra-hepatik melalui penggunaan tisu kultur HepG2 dan tikus terinduksi hiperkolesterolaemia. Penilaian toksisiti selular EBNE (0.2, 0.5, 0.8, 1.0 dan 1.5 mg/mL) telah menunjukkan sel mampu untuk terus hidup pada kadar 71%, walaupun ditambah pada dos EBNE yang tertinggi (1.5 mg/mL) selama 24 jam. Lanjutan daripada itu, tisu kultur HepG2 telah ditambah dengan EBNE (0.5, 1.0 dan 1.5 mg/mL) di dalam media tinggi lemak [lemak eksogenus (1:500) dan kolesterol (1:250)] selama 24 jam. Selain daripada itu, terdapat tiga kumpulan kawalan yang terdiri daripada kawalan asas (BC) yang dikultur di dalam larutan AMEM sahaja, kawalan negative (NC) di dalam media tinggi lemak dan kawalan positif (PC) di dalam media tinggi lemak bersama Simvastatin (4.60 µg/mL) sebagai anti-kolesterol. Kuantifikasi pengekspresian gen bagi kedua-dua LDLR dan ACAT2 telah menunjukkan kesan positif yang berkadar terus dengan peningkatan dos. Walau bagaimanapun, gen HMGCR telah menunjukkan perencatan yang signifikan hanya pada dos EBNE yang tinggi (1.5 mg/mL). Secara kuantitatif, taburan penandaan imunofluoresen terhadap LDLR protein dan titisan lemak (LDs) semakin meningkat apabila dos EBNE meningkat. Jumlah kolesterol tersimpan di dalam HepG2 turut menunjukkan peningkatan apabila dos EBNE semakin meningkat.

Di dalam 12 minggu kajian in-vivo, sejumlah 30 ekor tikus Sprague Dawley yang berusia 10 minggu telah diagihkan secara rawak kepada lima kumpulan (n=6) yang terdiri daripada BC (diet tikus biasa), NC [tikus hiperkolesterolemia diaruh melalui pemberian makanan diet tinggi lemak (HFD) dan Triton-X 100 (150 mg/kg SQ q42d)], PC (tikus hiperkolesterolemia dengan Simvastatin 10 mg/kg PO SID), EBNE [tikus hiperkolesterolemia dengan EBNE (6.5 mg/kg PO SID)] dan EBNS [tikus hiperkolesterolemia dengan EBNS (843.2 mg/kg)]. Profil lemak darah bagi kumpulan EBNS selepas eksperimen telah menunjukkan penurunan trigliserida (TAG), jumlah kolesterol (TC), lipoprotein ketumpatan rendah (LDL) dan lipoprotein ketumpatan sangat rendah (VLDL) yang signifikan berbanding kumpulan NC, tetapi hanya TAG dan LDL sahaja yang menunjukkan penurunan signifikan yang setara dengan kumpulan BC. Manakala, kumpulan EBNE hanya mencatatkan penurunan TC and LDL yang signifikan berbanding kumpulan NC, tetapi tidak setara dengan kumpulan BC. Secara statistik, kedua-dua kumpulan EBNE dan EBNS telah menunjukan peningkatan signifikan bagi lipoprotein ketumpatan tinggi (HDL) berbanding kumpulan BC dan NC, tetapi tidak setinggi yang direkodkan dalam kumpulan PC. Secara perbandingan, bacaan HDL di dalam kumpulan EBNS adalah dua kali lebih tinggi berbanding kumpulan EBNE. Berdasarkan data ini, hanya kumpulan EBNS telah menunjukkan bacaan perlindungan indeks jangkaan penyakit jantung dan aterosklerosis yang signifikan berbanding kumpulan-kumpulan lain dan setara dengan indeks yang dicatatkan dalam kumpulan PC. Selain itu, jumlah kepekatan kolesterol hepatic di dalam kumpulan EBNE dan EBNS telah merekodkan penurunan yang signifikan berbanding kumpulan NC. Ekspresi gen hati di dalam kumpulan EBNS menunjukkan perencatan dwi-gen HMGCR dan PCSK9 yang signifikan dan menyumbang kepada ransangan dan pengekalatan peningkatan gen LDLR melalui pengaktifan gen SREBP2. Pada masa yang sama, kumpulan EBNS telah mencatatkan peningkatan ekspresi gen CYP7a1 yang penting di dalam penghasilan jus hempedu. Manakala, EBNE telah merekodkan peningkatan ekspresi gen LDLR melalui pengaktifan tunggal gen SREBP2 sahaja. Secara kasar, hati kumpulan EBNS menunjukkan perubahan warna yang sedikit kekuningan, berbanding hati kumpulan EBNE yang kelihatan sederhana kekuningan. Pemeriksaan histologi mendapati hati EBNS dan EBNE masing-masing adalah steatosis hepatic sederhana dan steatohepatitis bukan-alkoholik (NASH) sederhana. Dapatan ini adalah selari dengan penemuan ultra-struktur di dalam TEM, yang mana mitokondria hati kumpulan EBNS kelihatan membesar dengan struktur membran yang masih utuh, berbanding mitokondria kumpulan EBNE yang membengkak dengan kehilangan kristae dan matriks yang kosong. Apabila LDLR protein pada tisu hati ditanda dengan imunofluoresen, kumpulan EBNS (5.65 ± 0.12) menunjukkan taburan protein yang banyak berbanding kumpulan EBNE, dan ini adalah setara dengan penemuan di kumpulan PC (5.53 ± 0.17). Walau bagaimanapun, tisu hati EBNE (3.54 ± 0.04) menunjukkan taburan yang lebih banyak berbanding kumpulan NC (1.81 ± 0.06). Kuantitatif SEM pada aorta torasik kranial, menunjukkan lumen aortik kumpulan NC dan EBNE adalah tersumbat sehingga 31% dan 30% berbanding kumpulan BC (0%). Manakala, kumpulan EBNS menunjukkan peratusan lumen aortik tersumbat kurang daripada 1% dan setara dengan kumpulan PC. Kuputusan ini adalah selari dengan dapatan di dalam imunoperoxidase bio-penandaan aterosklerosis (CD40). Tisu ginjal pula menunjukkan EBNS mampu mencegah glomerulonefritis, berbanding EBNE yang menunjukkan penebalan membran dasar glomerular atau glomerulonefritis. Penemuan ini adalah selari dengan pengesanan inflamasi (NLRP3) pada tubul renal kumpulan EBNE yang meliputi 61% potongan tisu, berbanding kumpulan EBNS yang

menunjukkan imunoreaktiviti pada kurang daripada 17%. Pada masa yang sama, masing-masing telah menunjukkan kehadiran sel inflamatori di dalam ruang interstitial renal pada kadar yang sedikit dan serdahana bagi kedua-dua kumpulan tersebut. Secara keseluruhannya, EBNE telah menunjukkan peningkatan metabolisme kolesterol di dalam tisu kultur HepG2 tetapi tidak pada tikus terinduksi hiperkolesterolemia. Manakala, EBNS telah menunjukkan kesan yang signifikan pada peningkatan metabolisme kolesterol dan memperlahankan pembentukan penyakit hati berlemak bukan-alkoholik (NAFLD), aterosklerosis dan penyakit ginjal kronik (CKD).



ACKNOWLEDGEMENTS

Praise to Allah the all mighty, the most gracious and giving of glad tidings. I would like to thank my main supervisor Assoc. Prof. Dr. Intan Shameha Abdul Razak for giving me this opportunity to participate as her Doctor of Philosophy's student. She was always open for any inquiries or trouble that I would have faced during my research time. My deepest gratitude for her endless efforts for giving the best to his 'son'.

I would like to acknowledge Dr. Mokrish Md. Ajat for his time and positive feedbacks in my research methodology, research progression until my final draft written. I am gratefully indebted to his very valuable comments on this thesis and thoughts. Thanks for being my 'brother' in the faculty.

I would also like to acknowledge Prof. Dr. Md. Zuki Abu Bakar@Zakaria for all his great remarks on technical and valuable ideas throughout the entire study. Without his comments and suggestions, this study could have taken a turn to the complicated way.

I would also like to thank my best friends at the faculty, Dr. Diyana Mohd Tahir for her morals supports, time and patience while I was using her laboratory for writing this thesis. Thank you again for guiding me and helping me out to understand the fundamental of being patience and thankful with whatever I had. Thanks to my bestie, Dr. Izdihar Ishak, Dr. Faizal Hahlan, Dr. Deva Darshini and Dr. Hafizah to lend yours ears and bear with me throughout this journey.

I would also like to express my deepest gratitude for my laboratory mates in Biochemistry Laboratory of the Faculty of Veterinary Medicine for the times we had together; during hardship and struggle, with all the valuable comments and suggestions, those were priceless moments I will remember. To Danish, Fadzly, Zakiah, Kak Mizah, Shidah and Qayyum, thank you for helping me out in troubleshooting my research. Not to be forgotten, all the laboratory staffs that were struggling to guide me in conducting my research and provided everything that I need.

Finally, I must express my very profound gratitude to my parents and my family for providing me unfailing support and continuous encouragement throughout these years of my study and through the process of research and writing this thesis. This accomplishment would not have been possible without them. Only Allah could repay them with His surplus kindness and blesses.

Thank you very much!

I certify that a Thesis Examination Committee has met on 24 February 2020 to conduct the final examination of Mohd Akmal bin Mohd Noor on his thesis entitled “Morphological Changes in Liver Cells of Hypercholesterolaemic-Induced Rats and HepG2 Cell Lines Associated with Edible Bird’s Nest Supplementation” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Goh Yong Meng, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Jalila binti Abu, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Hazilawati binti Hamzah, PhD

Associated Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Dewi Apri Astuti, PhD

Professor
Faculty of Animal Science
Bogor Agricultural University
Indonesia
(External Examiner)

ZURIATI AHMAD ZUKARNAIN, PhD

Professor Ts and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 07 AUGUST 2020

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Intan Shameha binti Abdul Razak, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Mohd Mokrish bin Md. Ajat, PhD

Senior Lecturer
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Md. Zuki bin Abu Bakar@Zakaria, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 13 AUGUST 2020

Declaration by Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from the supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/ fabrication in the thesis. And scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rule 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____

Date: 1st September 2020

Name and Matric No.: Mohd Akmal bin Mohd Noor (GS45622)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibility as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of
Chairman of
Supervisory
Committee: Assoc. Prof. Dr. Intan Shameha binti Abdul Razak

Signature: _____

Name of
Member of
Supervisory
Committee: Dr. Mohd Mokrish bin Md. Ajat

Signature: _____

Name of
Member of
Supervisory
Committee: Prof. Dr. Md. Zuki bin Abu Bakar@Zakaria

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iv
ACKNOWLEDGEMENTS	vii
APPROVAL	viii
DECLARATION	x
LIST OF TABLES	xv
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xvii
CHAPTER	
1 INTRODUCTION	1
1.1 Problem Statement	2
1.2 Hypothesis	3
1.3 General Objectives	4
2 LITERATURE REVIEW	5
2.1 Nutritional Analysis of EBN	5
2.2 Medicinal Properties of EBN	8
2.2.1 Supplement for Cardiometabolic Diseases	8
2.2.2 Prophylactic and Alternative Antiviral Agent	11
2.2.3 Post-Menopausal Supplement	13
2.2.4 Chondroprotective Agent	14
2.3 Cholesterol Metabolism and its Significance	15
2.4 Atherosclerosis, Non-Alcoholic Fatty Liver Disease (NAFLD) and Chronic Kidney Disease (CKD)	20
2.5 Dyslipidaemia Approaches in Natural Product	26
3 HIGH-DOSE SUPPLEMENTATION OF EDIBLE BIRD'S NEST EXTRACT (EBNE) IMPROVES CHOLESTEROL METABOLISM IN HIGH-LIPID MEDIA OF HEPG2	31
3.1 Introduction	31
3.2 Materials and Methods	33
3.2.1 EBN Extract Preparation	33
3.2.2 HepG2 Cell Maintenance and Treatment	34
3.2.3 HepG2 Cell Viability Assay	35
3.2.4 Cholesterol Gene Expression	36
3.2.5 HepG2 Immunofluorescence (IF) Staining	37
3.2.6 Intrahepatocellular Cholesterol Determination	38
3.2.7 Statistical Analysis	39
3.3 Results	39
3.3.1 EBN Extract (EBNE) Preparation	39
3.3.2 HepG2 Cell Viability	40
3.3.3 Qualitative Gene Expression in HepG2	41
3.3.4 HepG2 Immunofluorescence Staining	43
3.3.5 Intrahepatocellular Cholesterol Concentration	45

3.4	Discussion	46
3.4.1	HepG2 Cell Viability	46
3.4.2	Quantitative Gene Expression in HepG2	47
3.4.3	HepG2 Immunofluorescence Staining	49
3.4.4	Intrahepatocellular Cholesterol Storage in HepG2	49
3.5	Conclusion	50
4	EDIBLE BIRD'S NEST SOUP (EBNS) AMELIORATES BLOOD LIPID PROFILE VIA SUPPRESSION OF HEPATIC HMGCR AND PCSK9 GENES IN HYPERCHOLESTEROLAEMIC-INDUCED RATS	51
4.1	Introduction	51
4.2	Materials and Methods	53
4.2.1	Animal Study and Experimental Design	53
4.2.2	EBNS and EBNE Preparation	55
4.2.3	Quantitative Hepatic Gene Expression	56
4.2.4	Blood Lipid Profiling and Atherogenic Indices	58
4.2.5	Hepatic Cholesterol Determination	59
4.3	Results	60
4.3.1	Quantitative Hepatic Gene Expression	60
4.3.2	Blood Lipid Profiles and Atherogenic Indices	62
4.3.3	Hepatic Cholesterol Concentration	65
4.4	Discussions	66
4.4.1	<i>In-vivo</i> Study of Edible Bird's Nest (EBN)	66
4.4.2	Quantitative Hepatic Cholesterol Gene Expression	67
4.4.3	Blood Lipid Profiling and Cardiogenic Indices	71
4.4.4	Hepatic Cholesterol Storage Determination	74
4.5	Conclusion	75
5	EDIBLE BIRD'S NEST SOUP (EBNS) MODULATES PROGRESSION OF ATHEROSCLEROSIS, NAFLD AND CKD IN HYPERCHOLESTEROLAEMIC-INDUCED RATS	76
5.1	Introduction	76
5.2	Materials and Methods	78
5.2.1	Experimental Design and EBNE and EBNS Preparation	78
5.2.2	Detection of Atherosclerosis Biomarker (CD40)	78
5.2.3	Quantitative SEM of Aortic Lumen Patency	79
5.2.4	Gross and Histological Hepatic Evaluation in H&E and Masson's Trichrome	80
5.2.5	TEM of Hepatic Mitochondrial	81
5.2.6	Immunofluorescence Staining of LDLR	82
5.2.7	Renal Staining and NLRP3 Inflammasome Detection	82
5.3	Results	83
5.3.1	Distribution of CD40 in the Cranial Thoracic Aorta.	83
5.3.2	Quantitative SEM of Aorta	85
5.3.3	Gross and Histological Evaluations of Hepatic Parenchyma	87
5.3.4	TEM of Hepatic Mitochondria	94
5.3.5	Localization of Fluorescence Hepatic LDLR	96
5.3.6	Glomerulonephritis and NLRP3 Immunohistochemistry	98

5.4	Discussions	102
5.4.1	Localization of CD40 and Quantitative SEM of the Cranial Thoracic Aorta	102
5.4.2	TEM and Histological Evaluations of the Liver	104
5.4.3	Distribution of Fluorescence LDLR in the Liver	106
5.4.4	Renal Protectant Effect of the EBN	107
5.5	Conclusion	109
6	GENERAL DISCUSSION	111
7	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATION	114
	REFERENCES	116
	APPENDICES	137
	BIODATA OF STUDENT	162
	LIST OF PUBLICATIONS	163

LIST OF TABLES

Table	Page	
2.1	Identified proteins in EBN via LC-MS/MS technique	6
2.2	Identified metabolites in EBN via GC-MS and LC-MS techniques	8
3.1	Gene description in <i>in-vitro</i> study	37
3.2	Protein concentration of EBN extract for respective assays	40
4.1	Gene description for <i>in-vivo</i> study	57
4.2	Pre, peri and post treatment blood lipid profiles & cardiogenic indices	64
5.1	Rodent hepatic scoring system	81
5.2	Quantitative of aortic CD40 immunoreactivity	83
5.3	Percentage of aortic lumen patency	85
5.4	Hepatic scoring	93
5.5	Histomorphology of hepatic mitochondria	96
5.6	Quantitative of tubular NLRP3 immunoreactivity	100

LIST OF FIGURES

Figure	Page
2.1 Photographs of swiftlets and raw EBN	5
2.2 Chronological of chronic hypercholesterolaemia	26
2.3 Simplified cholesterol metabolism	30
3.1 Raw EBN and processed EBN granule	31
3.2 EBN extract (EBNE)	34
3.3 The cell viability of HepG2	40
3.4 Gene expression of <i>in-vitro</i> study	42
3.5 Micrograph of immunofluorescence (IF) staining	44
3.6 Mean fluorescence intensity of LDLR and LDs	45
3.7 Intrahepatocellular cholesterol concentration	46
3.8 Proposed schematic pathway for EBNE in HepG2	48
4.1 Graphical grouping of <i>in-vivo</i> study	54
4.2 EBN soup (EBNS)	55
4.3 Gene expression of <i>in-vivo</i> study	61
4.4 Hepatic cholesterol concentration	65
4.5 Proposed schematic pathway for EBNS in hypercholesterolaemic rats	70
4.6 Proposed schematic pathway for EBNE in hypercholesterolaemic rats	71
5.1 Micrographs of CD40 detection in the cranial thoracic aorta wall	84
5.2 Ultramicrographs of cranial thoracic aorta	86
5.3 Closed up view of arterial plaque	87
5.4 Gross morphology of hepatic lobes	88
5.5 Macrovesicular steatosis	89
5.6 Routine H&E staining of hepatic parenchyma	90
5.7 Microvesicular steatosis	91
5.8 Masson's trichrome staining of collagen fibres formation	92
5.9 Masson's trichrome staining of hepatic parenchyma	93
5.10 Ultramicrographs of hepatic mitochondria	95
5.11 Immunofluorescence staining of LDLR on the hepatic tissue	97
5.12 Mean intensity of LDLR fluorescence	98
5.13 Glomerular PAS staining of renal section	99
5.14 PAS staining of protein plasma insudation in the Bowman's space	100
5.15 Micrographs of IHC against NLRP3 in the kidney	101
5.16 Micrographs of inflammatory cells infiltration in the renal tubules	103

LIST OF ABBREVIATIONS

6-OHDA	Neurotoxin 6-Hydroxydopamine
ACAT	Acyl Coenzyme A: Cholesterol Acyltransferase
ACE	Angiotensin I Converting Enzyme
ACP	Aggrecan Core Protein
ACS	Apoptosis-associated Speck-like Protein Containing A Caspase Activation and Recruitment Domain
AGEs	Advanced Glycation End-Products
AKI	Acute Kidney Injury
AMEM	Advanced Minimum Essential Media
APOA5	Apolipoprotein-A5
ApoB-100	Apolipoprotein B-100
APP	Amyloid Precursor Protein
APTT	Activated Partial Thromboplastin Time
ATCC	American Type Culture Collection
ATP	Adenosine Triphosphate
BC	Baseline Control
bHLH-Zip	Basic Helix-Loop-Helix Leucine Zipper
CAT	Catalase
CCL2	Chemokine (C-C Motif) Ligand 2
CD40	Cluster of Differentiation 40
CD40L	Cluster of Differentiation 40 Ligand
CE	Cholesteryl Ester
CETP	Cholesteryl Ester Transport Protein
CKD	Chronic Kidney Disease
CLS	Crown-like Structure
CMS	Cardiometabolic Syndrome
CNS	Central Nervous System
Co-A	Coenzyme-A
COL	Collagen
Co-Q	Coenzyme-Q
COX-2	Cyclooxygenase-2
CRP	C-Reactive Protein
CSPCP	Cartilage-Specific Proteoglycan Core Protein
CVD	Cardiovascular Disease
DAMP	Damage-Associated Molecular Pattern
DM	Diabetes Mellitus
DN	Diabetic Nephropathy
E ₂	Estradiol
EAA	Essential Amino Acid
EBN	Edible's Bird Nest
EBNE	Edible's Bird Nest Extract
EBNS	Edible's Birds Nest Soup
ECM	Extracellular Matrix
EGF-A	Epidermal Growth Factor-A
eLDs	End Lipid Droplets
ESRD	End Stage Renal Disease
ETC	Electron Transport Chain
FADH ₂	Flavin Adenine Dinucleotide

FATP1	Fatty Acid Transport Protein 1
FC	Free Cholesterol
FFA	Free Fatty Acid
Fox03a	Forkhead Box 03a
FPE	Fermented Fine Needle Extract
FSH	Follicular Stimulating Hormone
GAG	Glycosaminoglycan
GALNT2	Polypeptide N-Acetylgalactosaminyltransferase 2
GAPDH	Glyceraldehyde-3-Phosphate Dehydrogenase
GBM	Glomerular Basement Membrane
GCK	Glucokinase
GC-MS	Gas Chromatography-Mass Spectrometry
GEEnCs	Glomerular Endothelial Cells
GI	Gastrointestinal
GIT	Gastrointestinal Tract
GN	Glomerulonephritis
Gpx	Glutathione Peroxidase
GRF	Glomerular Rate Filtration
Gsr	Glutathione-Disulphide Reductase
HA	Hemagglutinin
HCD	High-Cholesterol Diet
HDL	High-Density Lipoprotein
HFD	High-Fat Diet
HMGCR	3-Hydroxy-3-Methylglutaryl-CoA Reductase
HNF-1 α	Hepatic Nuclear Factor 1 Alpha
HOMA-IR	Homeostatic Model Assessment of Insulin Resistance
HPLC	High-Performance Liquid Chromatography
HRT	Hormone Replacement Therapy
HSPs	Heat-Shock Proteins
IAV	Influenza A Virus
IFN- γ	Interferon-Gamma
IHC	Intrahepatocellular Concentration
IHC	Immunohistochemistry
IL	Interleukin
IL-18R β	Interleukin-18 Receptor Beta
IL-1R	Interleukin 1 Receptor
IL-1 β	Interleukin 1 Beta
iLDs	Initial Lipid Droplets
iNOS	Inducible Nitric Oxide Synthase
INSR	Insulin Receptor Precursor
IRS2	Insulin Receptor Substrate 2
JAK2	Janus-Activated Kinase 2
KCNJ11	Potassium Voltage-Gated Channel Subfamily J Member 11
LAL	Lysosomal Acid Lipase
LC3-II	Autophagosome Membrane Protein
LC-MS/MS	Liquid Chromatography-Mass Spectrometry/Mass Spectrometry
LDL	Low-Density Lipoprotein
LDLR	Low-Density Lipoprotein Receptor
LDs	Lipid Droplets
LF	Lactoferrin

LH	Luteinizing Hormone
LIPC	Lipase C
LOX-1	Oxidised Low-Density Lipoprotein Receptor 1
LPL	Lipoprotein Lipase
MCH-I	Major Histocompatibility Complex Class I
MCH-II	Major Histocompatibility Complex Class II
MCP-1	Monocyte Chemoattractant Protein 1
MDA	Malondialdehyde
MetS	Metabolic Syndrome
MGN	Membranous Glomerulonephritis
MLXIPL	MLX Interacting Protein Like
MMPs	Matrix Metalloproteinases
MPGN	Membranoproliferative Glomerulonephritis
MTT	3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide
NA	Neuraminidase
NADH	Nicotinamide Adenine Dinucleotide
NAFLD	Non-Alcoholic Fatty Liver Disease
NASH	Non-Alcoholic Steatohepatitis
NC	Negative Control
nCEH	Neutral Cholesteryl Ester Hydrolase
NF- κ β	Nuclear Factor Kappa Beta
NLRP3	NOD-like receptor family, pyrin containing domain 3
NO	Nitric Oxide
NOD	Nucleotide-binding Oligomerization Domain
OA	Osteoarthritis
OVF	Ovotransferrin
Ox-LDL	Oxidised Low-Density Lipoprotein
PC	Positive Control
PCSK9	Proprotein Convertase Subtilisin/Kexin Type 9
PD	Parkinson's Disease
PE	Pine Needle Extract
PI3K	Phosphoinositide-3-Kinase
PK	Pyruvate Kinase
PON3	Serum Paraoxonase/Lactonase 3
PRR	Pattern Recognition Receptor
PSEN	Presenilin
PT	Prothrombin Time
ROS	Reactive Oxygen Species
S1P	Site-1 Protease
S2P	Site-2 Protease
SCAP	SREBP Cleavage-Activating Protein
SEM	Scanning Electron Microscopy
SOD	Superoxide Dismutase
SRA	Scavenger Receptor A
SRE	Sterol Regulatory Element
SREBP	Sterol Regulatory Element-Binding Protein
TAG	Triglycerides
TCM	Traditional Chinese Medicine
TECs	Tubular Epithelium Cells
TEM	Transmission Electron Microscopy

TGF- β 1	Transforming Growth Factor Beta 1
TLR	Toll-like Receptor
TNF- α	Tumour Necrosis Factor Alpha
TRAIL	Tumour Necrosis Factor-related Apoptosis Inducing Legend
TRX	Triton X-100
VLDL	Very Low-Density Lipoprotein
VRF	Vanillin Rich Fraction
vRNP	Viral Ribonucleoprotein Complex
VSMCs	Vascular Smooth Muscle Cells



CHAPTER 1

INTRODUCTION

The edible bird's nest (EBN) is an animal-derived natural product that serves as the Chinese delicacy for thousands of years. The history of this natural product was documented back in the Tang Dynasty (618-907 CE) and frequently consumed by the Chinese royal family due to their extensive medicinal properties and well-being effects (Marcone, 2005). In worldwide, there are two type of EBN that are available in the market including house-EBN and cave-EBN (Looi and Rahman, 2016). In general, EBN is mainly composed by conjugated glycoprotein such as sialic acid, glucosamine and galactosamine (Ma and Liu, 2012b). Most of the protein is highly solubilized amino acids. Thus, most of the extraction methods in recent studies were adopting water-based extraction, which is a traditional way in which the Chinese community prepared the EBN soup (Marcone, 2005; Utomo *et al.*, 2014; Chua *et al.*, 2014a and Zulkefli *et al.*, 2017). Besides that, it also contained minerals, vitamins and some hormones (Yu-Qin *et al.*, 2000; Ma and Liu, 2012a). This product is derived from the secretion of swiftlet's sublingual salivary glands that is produced all year round, but predominantly between Septembers to December during the breeding season. Both sexes are involved in the nest building but mainly built by the male swiftlets (Looi and Omar, 2016). Microscopically, the EBN composed of polymerized salivary secretion with varies degree of the feather and plumage composition that will support the nestling and the mother up to 40 days (Langham, 1980; Ma and Liu, 2012b). In general, swiftlets (*Aerodramus fuciphagus*) is an aerial insectivore that mainly inhabits in limestone cave situated more than 1300 meters above the sea level to avoid interspecies competition (Lim *et al.*, 2002). This amazing bird is anatomically different from other common swift, as they have shorter metatarsal bones and digits together with underdeveloped (smaller and thinner) caudoproximal of pelvic muscles bundle (biceps femoris, semitendinosus, semimembranosus and gastrocnemius), resulting inability of the swiftlets to perch, walk and stand (Zuki *et al.*, 2012).

In recent years, extensive preclinical studies including *in-vitro* and *in-vivo* experiments were discovered several potential medicinal effects of the EBN. Based on those studies, EBN able to improve cognitive function, insulin resistance, regulate coagulation pathway, serves as therapeutic and prophylactic agents including anti-hypertensive, antioxidant, anti-inflammatory, antiviral, natural hormonal supplement and chondroprotective agent. In the ancient Traditional Medicine (TCM) books, EBN had been claimed to improve metabolism. Therefore, we were postulating the EBN might have a potential effect in regulating cholesterol metabolism as well. Cholesterol metabolism is a well-established biochemical pathway in the body and alteration of this metabolism can lead to the development of cardiovascular disease (CVD) (Charlton-Menys and Durrington, 2007). A major risk factor of CVD is the atherogenic dyslipidaemia which characterized with the elevation of triglycerides (TAG) and low-density lipoprotein (LDL), and reduction of high-density lipoprotein (HDL) in the blood (Miller *et al.*, 2011). When there is a dysregulation of lipoprotein metabolism, particularly in LDL, which commonly known as bad cholesterol, it can increase the risk of an individual to succumb with CVD (Chong *et al.*, 2011). As this LDL is readily infiltrated to the sub-endothelial space of the arteries wall due to light

molecular weight, these particles are rapidly phagocytized by arterial wall macrophages in the tunica intima, which eventually transforms into the foam cells (Zilvermit, 1979; Myasoedova *et al.*, 2017). Prolong accumulation of foam cells on the arterial wall will trigger an inflammatory reaction that will lead to the formation of a solidified substance, known as a plaque (Laderis-Lopes *et al.*, 2015). As the arterial plaque is growing, important atherosclerosis biomarker, which is CD40 will start to express at any stage of the development (Leivens *et al.*, 2009; Lusis, 2000).

Formation of the plaques will result in high blood pressure and eventually, multiple diseases can be observed; such as stroke, acute pulmonary, liver failure or renal failure (WHO, 2014). As the dyslipidaemia advanced, the hepatic manifestation of the metabolic disease will be triggered to progress with the maturation of key cholesterol metabolism genes; including 3-hydroxy-3-methylglutaryl-CoA reductase (HMGCR) and sterol regulatory element-binding protein 2 (SREBP2) (Mi *et al.*, 2012). These two genes will induce accumulation of free cholesterol that contributed to the development of non-alcoholic fatty liver diseases (NAFLD), despite accumulation of other lipid components (TAG and cholesteryl ester) (Polimeni *et al.*, 2015; Ioannou *et al.*, 2017); which can be ranged from simple hepatic steatosis to hepatocellular carcinoma (Tacer and Rozman, 2011; Polimeni *et al.*, 2015). NAFLD is characterized with an elevation of lipid load in the hepatic parenchyma which causing histological and physiological alteration that can lead to a diseased liver (Jayakumar *et al.*, 2011; Tacer and Rozman, 2011; Polimeni *et al.*, 2015; Tsutsumi *et al.*, 2017). As the liver is undergoing pathological changes, an abundance of damaged cellular components that accompanied with high blood cholesterol remain to circulate in the bloodstream. All these components will be served as damage-associated molecular pattern (DAMP) which will activate the NLRP3 inflammasome in the kidney and leading to the wide range of glomerulonephritis (GN) and tubular nephritis (Hutton *et al.*, 2016; Anders and Schaefer, 2014; Fioretta and Mauer, 2007; Chun *et al.*, 2016)

Initiated with alteration of lipid metabolism, or particularly dysregulation of cholesterol metabolism, the clinical manifestation of lesions in the several internal organs is widely described in the previous pre-clinical studies and series of case reports. Therefore, the significance of cholesterol metabolism in diseases development currently is the main highlight in the biomedical research, which aims to improve cholesterol metabolism. In line with the claim that has been anecdotally documented in the TCM books, undiscovered and novel potential anti-cholesterol properties of the EBN is incompletely described. Thus, the present study was conducted to evaluate the effect of the EBN(s) in the *in-vitro* (HepG2 cell lines) and *in-vivo* (hypercholesterolaemic-induced rat) setting, particularly elucidating the biochemical regulation in the liver, and histological alteration in the cranial thoracic aorta, liver and kidney, which can be observed upon supplementation of EBN.

1.1 Problem Statement

In line with the emerging of metabolic diseases or collectively known as metabolic syndrome (MetS); the occurrence of this problem was withdrawing interest among the medical practitioner worldwide; as it was causing among the highest mortality rate in

the human population for the past few decades (Polimeni *et al.*, 2015). Incidence of this syndrome is highly associated with prolonging consumption of high-calories diet, sedentary physical activities and metabolic gene polymorphism (Chan, Barrett and Watts, 2014). Association of MetS with metabolism impairment was epidemiologically explained, and one of the approaches to solve this problem is consumption of chemically derived medications to alter such abnormalities (Charlton-Menys and Durrington, 2007). As the flourishing of holistic approaches in this century, discovering of potential natural product was withdrawing researchers' attention, and it was eccentrically studied from the ethnic's origin, beliefs and religion (Calixto, 2000).

One of the natural products that has been extensively studied in the previous five years, is the edible bird's nest (EBN). In the early Traditional Chinese Medicine (TCM) books such as "Shen Non Ben Cao Jing" (1695), "Ben Cao Cong Xin" (1759) and "Ben Cao Qiu Zhen" (1769), they have been claimed that the EBN able to maintain general health wellness, including several medicinal properties such as antioxidant, antiviral, immune booster and metabolism stimulant (Haghani *et al.*, 2016). Nonetheless, for the past few decades, there were insufficient scientific evidence to prove such claims (Ma and Liu, 2012). In the experimental setting, EBN was supplemented in two forms, either EBN extract (EBNE) and/or EBN soup (EBNS), depended on the study design. EBNE was predominantly tested in the *in-vitro* studies, as the EBNS will be causing mortality in the cell culture. Meanwhile, in *in-vivo* studies, both forms of EBNS can be supplemented via oral-gavage for hypothetical testing purposes. According to TCM books, the EBN was traditionally claimed to stimulate and boost body metabolism. However, the terminology of 'metabolism' itself was not clearly stated, and which metabolism the EBN able to trigger upon the consumption? Thus, the definition of metabolism was literally elaborated to get a better explanation. Metabolism is a sum-up sequential chemical reactions in the living organism, that is crucial for life-sustaining; which initially processed at the cellular level and mediated by all the nutritional metabolites including carbohydrates, protein, lipid, minerals and vitamins (Matthews *et al.*, 2013). As the component of the metabolism is well-known and broadly explained above, the current study was postulating the EBN might have the specific effect on the sub-type of lipid component which cholesterol, that plays a major risk in modulating metabolic syndrome; particularly hypercholesterolaemia. Finally, the effect of EBN on cholesterol metabolism is poorly been described, and this research will be the first and novel study to describe it.

1.2 Hypothesis

Supplementation of the edible bird's nest (EBN) can up-regulate cholesterol metabolism in the HepG2 cell lines and hypercholesterolaemic-induced rats, which consequently ameliorates blood lipid profiling, hepatic lipid concentration and histological changes of liver, kidney and cranial thoracic aorta.

1.3 General Objectives

The main objective of this study is to evaluate the effect of edible bird's nest (EBN) supplementation on the cholesterol metabolism in HepG2 cell lines and hypercholesterolaemic-induced rats.

1.3.1 Specific Objectives

1. To quantify relevant gene expression on cholesterol metabolism in in the HepG2 cell lines and hypercholesterolaemic-induced rats supplemented with EBNs.
2. To localize hepatic distribution of low-density lipoprotein receptor (LDLR) and lipid droplets (LDs) in the HepG2 and hypercholesterolaemic-induced rats supplemented with EBNs.
3. To determine hepatic lipid concentration in the HepG2 cell lines and hypercholesterolaemic-induced rats supplemented with EBNs.
4. To assess blood lipid profiling in hypercholesterolaemic-induced rats supplemented with EBNs.
5. To evaluate histological and ultrastructure changes in the liver, kidney and cranial thoracic aorta of hypercholesterolaemic-induced rats supplemented with EBNs.

REFERENCES

- Abais, J. M., Zhang, C., Xia, M. *et al.* (2013). NADPH oxidase-mediated triggering of inflammasome activation in mouse podocytes and glomeruli during hyperhomocysteinemia. *Antioxidant and Redox Signalling*. 18: 1537–1548.
- Abifadel, M., Varret, M., Rebes, J. P., Allard, D., Ouguerram, K., Devillers, M. and Cruaud, C. (2003). Mutation in PCSK9 cause autosomal dominant hypercholesterolaemia. *Nature Genetic*. 34(2): 154-156
- Al-Naqeb, G., Ismail, M., Bagalkotkar, G. and Adamu, H. A. (2010). Vanillin rich fraction regulates LDLR and HMGCR gene expression in HepG2 cells. *Food Research International*. 43: 2437-2443
- Alberti, K. G., Eckel, R. H., Grundy, S. M., Zimmet, P. Z., Cleeman, J. I., Donato, K. A., Fruchart, J. C., James, W. P., Loria, C. M. & Smith Jr. S. C. (2009). Harmonizing the metabolic syndrome: a joint interim statement of International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung and Blood Institute; American Heart Association, World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. *Circulation*. 120(16), 1640-1645.
- Aldridge, J. R., Moseley, C. E., Boltz, D. A., Negovetich, N. J., Reynolds, C., Franks, J., Brown, S. A., Doherty, P. C., Webster, R. G. & Thomas P. G. (2009). From the cover: TNF/iNOS-producing dendritic cells are the necessary evil of lethal influenza virus infection. *Proceeding of the National Academy of Sciences*. 5306-5311.
- Alger, H. M., Brown, J. M., Sawyer, J. K., Kelley, K. L., Shah, R., Wilson, M. D., Willingham, M. C. & Rudel, L. L. (2010). Inhibition of ACAT2 prevents dietary cholesterol-associated steatosis by enhancing TAG mobilization. *Journal of Biological Chemistry*. 285(19): 14267-14274.
- Aoun, M., Fouret, G., Michel, F., Bonafos, B., Ramos, J. *et al.*, (2012). Dietary fatty acids modulate liver mitochondrial cardiolipin content and its fatty acid composition in rats with non-alcoholic fatty liver disease, *Journal of Bioenergetics and Biomembrane*. 44: 439–452.
- Aowphol, A., Voris, H. K., Feldheim, K. A., Harnyuttanakorn, P. & Thirakhupt, K. (2008). Genetic homogeneity among colonies of the white-nest swiftlet (*Aerodramus fuciphagus*) in Thailand. *Zoological Sciences*. 25(4), 372-380.
- Anders, H. J. & Muruve, D. A. (2011). The inflammasomes in kidney disease. *Journal of the American Society of Nephrology*. 22: 1007–1018.
- Anders, H. J. and Schaefer, L. (2014). Beyond tissue injury-damage-associated molecular patterns, toll-like receptors, and inflammasomes also drive regeneration and fibrosis. *Journal of the American Society of Nephrology*. 25: 1387–1400.

- Angelico, F., Del Ben, M., Conti, R., Francioso, S., Feole, K., Fiorello, S., Cavallo, M. G., Zalunardo, B., Lirussi, F., Alessandri, C. and Violi, F. (2005). Insulin resistance, the metabolic syndrome, and non-alcoholic fatty liver disease. *Journal of Clinical Endocrinology and Metabolism*.
- Arab, J. B., Arrese, M. and Trauner, M. (2018). Recent insights into the pathogenesis of non-alcoholic fatty liver disease. *Annual Review of Pathology: Mechanism of Disease*. 13: 321-350.
- Ashton, M. J., Brown, T. J., Fenton, G., Halley, F., Harper, M. F., Lockey, P. M., Porter, B., Roach, A. G., Shuttle, K. A. J., Vicker, N. and Walsh, R. J. A. (1996). New low-density lipoprotein receptor up-regulators acting via a novel mechanism. *Journal of Medicinal Chemistry*. 39: 3343-3356.
- Aswir, A. R. and Wan Nazaimoon, W. M. (2011). Effect of edible bird's nest on cell proliferation and tumour necrosis factor-alpha release in vitro. *International Food Research Journal*. 18(3), 1123-1127.
- Au, R. Y., Al-Talib, T. K., Au, A. Y., Phan, P. V. & Frondoza, C. G. (2007). Avocado soybean unsaponifiables (ASU) suppress TNF- α , IL-1 β , COX-2, iNos gene expression, and prostaglandin E2 and oxide production in articular chondrocytes and monocytes/macrophages. *Osteoarthritis and Cartilage*. 15, 1249-1255.
- Bakovic, M. P., Sukhorukov, V., Zakiev, E., Orekhov, A., Lauc, G. & Kontush, A. (2017). Sialylation of human plasma lipoprotein as a key determinant of biological functions. *Atherosclerosis*. 263: e91-e92.
- Barter, P. J., Brandrup-Wognseh, G., Palmer, M. K. & Nicholls, S. J. (2010). Effect of statins on HDL-C: a complex process unrelated to changes in LDL-C: analysis of the VOYAGER Database. *Journal of Lipid Research*. 51(6): 1546-1553.
- Barik, A., Mishra, B., Kunwar, A., *et al.* (2007). Comparative study of copper(II)-curcumin complexes as superoxide dismutase mimics and free radical scavengers. *European Journal of Medical Chemistry*. 42: 431-439.
- Benz, K. & Amann, K. (2011). Endothelin in diabetic renal disease. *Contributions to Nephrology*. 172: 139-148.
- Berger, S., Raman, G., Vishwanathan, R., Jacques, P. F. and Johnson, E. J. (2015). Dietary cholesterol and cardiovascular disease: a systemic review and meta-analysis. *American Journal of Clinical Nutrition*. 102(2): 276-294.
- Bligh, E. G. and Dryer, W. J. (1959). A rapid method of total lipid extraction and purification. *Canadian Journal of Biochemistry and Physiology*. 37(8): 911-917.
- Blount, K. J. & Hagspiel, K. D. (2009). Aortic diameter, true lumen and false lumen growth rates in chronic type B aortic dissection. *American Journal of Roentgenology*. 192(5): W222-229.
- Bodary, P. F. & Eitzman, D. T. (2009). Animal model of thrombosis. *Current Opinion in Haematology*. 16(5): 342-346.

- Brooke, R. K. (1970). Taxonomic and evolutionary notes on the subfamilies, tribes, genera and subgenera of Swifts (Aves: Apodidae). *Durban Museum Novitates*. 9, 13-24.
- Brooke, R. K. (1972). Generic limits in old world Apodidae and Hirundinidae. *Bulletin of the British Ornithologists' Club*. 92, 53-57.
- Brown, M. S. and Goldstein, J. L. (1997). The SREBP pathway: regulation of cholesterol metabolism by proteolysis of a membrane-bound transcription factor. *Cell*. 89: 331-340.
- Brown, M. S. and Goldstein, J. L. (1986). A receptor mediated pathway for cholesterol homeostasis. *Science*. 23: 34-47.
- Browning, J. D. & Horton, J. D. (2004). Molecular mediators of hepatic steatosis and liver injury. *Journal of Clinical Investigation*. 114: 147-152.
- Bugianesi, E., Moscatiello, S., Ciaravella, M. F. & Marchesini, G. (2010). Insulin resistance in non-alcoholic fatty liver disease. *Current Pharmacology of Design*. 16(17): 1941-1951.
- Buhaescu, I and Izzedine, H. (2007). Mevalonate pathway: a review of clinical and therapeutical implications. *Clinical Biochemistry*. 40(9): 575-584.
- Buhman, K. F., Accad, M. and Farese Jr, R. V. (2000). Mammalian acyl-CoA: cholesterol acyltransferase. *Biochimica et Biophysica Acta*. 1529: 142-154.
- Calixto, B. J. (2000). Efficacy, safety, quality, control, marketing and regulatory guidelines for herbal medicines (phytotherapeutic agents). *Brazilian Journal of Medical and Biological Research*. 33, 179-189.
- Cardile, V., Panico, A., Gentile, B., Borrelli, F. & Russo, A. (2003). Effects of propolis on human cartilage and chondrocytes. *Life Science*. 73, 1027-1035.
- Careskey, H. E., Davis, R. A., Alborn, W. E., Troutt, J. S., Cao, G., & Konrad, R. J. (2008). Atorvastatin increases human serum levels of proprotein convertase subtilisin/kexin type 9. *Journal of Lipid Research*. 49: 394-398.
- Chalasani, N. (2005). Statin and hepatotoxicity: focus on patients with fatty liver. *Hepatology*. 41(4): 690-695.
- Chan, D. C., Barrett, P. H. R. & Watts, G. F. (2014). The metabolic and pharmacological bases for treating atherogenic dyslipidaemia. *Best Practice & Research Clinical Endocrinology & Metabolism*. 28, 369-385.
- Chan, W. K., Tan, A. T. B., Vethakkan, S. R., Tah, P. C., Vijayanathan, A. & Goh, K. L. (2013). NAFLD in diabetic- prevalence and predictive factors in a multiracial hospital clinic population in Malaysia. *Journal of Gastroenterology and Hepatology*. 28: 1375-1383.

- Chang, T. Y., Li, B. L., Chang, C. C. and Urano, Y. (2009). Acyl-coenzyme A: cholesterol acyl transferase. *American Journal of Physiology- Endocrinology and Metabolism*. 297: E1-E9
- Charlton-Menys, V. and Durrington, P. N. (2007). Human cholesterol metabolism and therapeutic molecules. *Experimental Physiology*. 93(1): 27-42.
- Chaudhary, R., Garg, J., Shah, N. & Summer, A. (2016). PCSK9 inhibitors: a new era of lipid lowering therapy. *World Journal of Cardiology*. 9(2): 76-91.
- Chen, S., Wassenhove-McCarthy, D. J., Yamaguchi, Y., Holzman, L. B., van Kuppevelt, T. H., Jenniskens, G. J., Wijnhoven, T. J., Woods, A. C. & McCarthy, K. J. (2008). Loss of heparan sulfate glycosaminoglycan assembly in podocytes does not lead to proteinuria. *Kidney International*. 74: 289-299.
- Chen, S. C., Hung, C. C., Kuo, M. C., Lee, J. J., Chiu, Y. W., Chnag, J. M., Hwang, S. J. and Chen, H. C. (2013). Association of dyslipidaemia with renal outcomes in CKD. *PLoS One*. 8(2): e55643.
- Chiang, J. Y. L. (2017). Targeting bile acids and lipotoxicity for NASH treatment. *Hepatology Communications*. 1(10): 1002-1004.
- Cholesterol Treatment Trialists' (CTT) Collaboration (2010). Efficacy and safety of more intensive lowering of LDL cholesterol: a meta-analysis of data from 170 000 participants in 26 randomised trials. *Lancet*. 376: 1670-1681.
- Chong, S. C., Dollah, M. A., Chong, P. P. & Maha, A. (2011). *Phaleria macrocarpa* (Scheff.) Boerl fruit aqueous extract enhances LDL receptors and PCSK9 expression in-vivo and in-vitro. *Journal of Ethnopharmacology*. 137, 313-327.
- Chong, T., Naples, M., Federico, L., Taylor, D., Smith, G. J., Cheung, R. C and Adeli, K. (2006). Effect of rosuvastatin on hepatic production of apolipoprotein B-containing lipoproteins in an animal model of insulin resistance and metabolic dyslipidaemia. *Atherosclerosis*. 185: 21-31
- Chua, K. H., Lee, T. H., Nagandran, K., Yahya, N. H. M., Lee, C. T., Tijh, E. T. T. & Aziz, R. A. (2013). Edible bird's nest extract as a chondro-protective agent for human chondrocytes isolated from osteoarthritic knee: invitro study. *BMC Complementary and Alternatives Medicine*. 13, 19-27.
- Chua, Y. G., Bloodworth, B. C. Leong, L. P. & Li, S. F. Y. (2014a). Metabolite profiling od edible bird's nest using gas chromatography-mass spectrometry and liquid chromatography-mass spectrometry. *Rapid Communication in Mass Spectrometry*. 28, 1387-1400.
- Chua, Y. G., Chan, S. H., Bloodworth, B. C., Li, S. F. Y. & Leong, L. P. (2014b). Identification of edible bird's nest with amino acid and monosaccharide analysis. *Journal of Agricultural and Food Chemistry*. 63, 279-289.
- Chun, J., Chung, H., Wang, X., Barry, R., Taheri, Z. M., Platnich, J. M., Ahmed, S. B. *et al.*, (2016). NLRP3 localizes to the tubular epithelium in human kidney and correlates with outcome in IgA nephropathy. *Scientific Reports*. 6, 24667.

- Coker, L. H., Espeland, M. A., Rapp, S. R., Legault, C., Resnick, S. M., Hogan, P., Gaussoin, S., Dailey, M. & Shumaker, S. A. (2010). Postmenopausal hormone therapy and cognitive outcomes: The Women's Health Initiative Memory Study (WHIMS). *Journal of Steroid Biochemistry and Molecular Biology*. 118, 304-310.
- Colombo, J. P., Garcia-Rodenas, C., Guesry, P. R. & Ray, J. (2003). Potential effect of supplementation with amino acids, choline or sialic acid on cognitive development in young infants. *Acta Paediatrica*. 92 (442), 42-46.
- Del Ben, M., Baratta, F., Polimeni, L. and Angelico, F. (2012). Non-alcoholic fatty liver disease and cardiovascular disease: epidemiological, clinical and pathophysiological evidences. *Internal and Emergence Medicine*. 7(3): S291-S296.
- Denis, M., Marcinkiewicz, J., Zaid, A., Gauthier, D., Poirier, S., Lazure, C., Seidah, N. G. & Prat, A. (2012). Gene inactivation of proprotein convertase subtilisin/kexin type 9 reduces atherosclerosis in mice. *Circulation*. 125: 894-901.
- Despres, J. P. and Lemieux, I. (2006). Abdominal obesity and metabolic syndrome. *Nature*. 444: 881-887.
- Ding, Y., Xiao, C., Wu, Q., Xie, Y., Li, X., Hu, H. & Li, L. (2016). The mechanism underlying the hypolipidaemic effects of *Grifola frondosa* in the liver of rats. *Frontiers in Microbiology*. 7: 1186.
- Draganov, D. I., Stetson, P. L., Watson, D. E., Billecke, S. S. & La Dun, B. N. (2000). Rabbit serum paraoxonase 3 (PONS3) is a high density lipoprotein-associated lactonase and protects low-density lipoprotein against oxidation. *Journal of Biology and Chemistry*. 275 (43): 33435-33442.
- Duff, C. J., Scott, M. J., Kirby, I. T., Hutchinson, S. E., Martin, S. L. & Hooper, N. M. (2009). Antibody-mediated disruption of the interaction between PCSK9 and the low-density lipoprotein receptor. *Biochemical Journal*. 419: 577-584.
- Durrington, P. N. (2007). *Hyperlipidaemia: Diagnosis and Management*. 3rd edition (London: Hodder Arnold). 258-293
- Edinger, T. O., Pohl, M. O. & Stertz, S. (2014). Entry of influenza A virus: host factors and antiviral targets. *Journal of General Virology*. 95 (2), 263-277.
- Elliot, W. J. & Meyer, P. M. (2007). Incident diabetes in clinical trials of antihypertensive drugs: a network meta-analysis. *Lancet*. 369, 201-207.
- Endo, A. (1992). The discovery and development of HMG-CoA reductase inhibitors. *Journal of Lipid Research*. 33: 1569-1582.
- Esterbauer, H., Schaur, R. J. & Zollner, H. (1991). Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radical Biology Medicine*. 11: 81-128

- Ferramosca, A., Giacomo, M. D. & Zara, V. (2017). Antioxidant dietary approach in treatment of fatty liver: new insights and updates. *World Journal of Gastroenterology*. 23(23): 4146-4157.
- Fioretto, P. and Mauer, M. (2007). Histopathology of diabetic nephropathy. *Seminars in Nephrology*. 27 (2): 195-207.
- Flanagan-Cato, C. M. (2000). Estrogen-induced remodeling of hypothalamic neural circuitry. *Frontiers in Neuroendocrinology*. 21, 309-329.
- Franko, A., von Kleist-Retzow, J. C., Bose, M., Sanchez-Lasheras, C., Brodesser, S. *et al.*, (2012). Complete failure of insulin-transmitted signalling, but not obesity-induced insulin resistance, impairs respiratory chain function in muscle, *Journal of Molecular Medicine*. 90: 1145–1160.
- Fukuyama, S. & Kawaoka, Y. (2011). The pathogenesis of influenza virus infection: the contribution of virus and host factors. *Current Opinion in Immunology*. 23(4), 481-486.
- Furuichi, K., Wada, T., Iwata, Y. *et al.* (2006). Interleukin-1-dependent sequential chemokine expression and inflammatory cell infiltration in ischemia–reperfusion injury. *Critical Care Medicine*. 34: 2447–2455
- Furukawa, S., Fujita, T., Shimabukuro, M. *et al.* (2004). Increased oxidative stress in obesity and its impact on metabolic syndrome. *Journal of Clinical Investigation*. 114: 1752–1761.
- Gabay, C., Lamacchia, C. and Palmer, G. (2010). IL-1 pathways in inflammation and human diseases. *National Review of Rheumatology*. 6: 232–241.
- Galkina, E. & Ley, K. (2009). Immune and inflammatory mechanism of atherosclerosis. *Annual Review of Immunology*. 27, 165-197.
- Galloway, C. A., Lee, H., Brookes, P. S. and Yoon, Y. (2014). Decreasing mitochondrial fission alleviates hepatic steatosis in a murine model of non-alcoholic fatty liver disease. *American Journal of Physiology-Gastrointestinal and Liver Physiology*. 307: G632–G641.
- Gansevoort, R. T., Correa-Rotter, R., Hemmelgarn, B. R., Jafar, T. H., Heerspink, H. J. L., Mann, J. F., Matsushita, K. & Wen, C. P. (2013). Chronic kidney disease and cardiovascular risk: epidemiology, mechanisms, and prevention. *Lancet*. 382: 339-352.
- Gartner, L. P. 2016. Urinary system. In *Textbook of Histology*. United State of America: Elsevier.
- Gauer, S., Sichler, O., Obermuller, N. *et al.*, (2007). IL-18 is expressed in the intercalated cell of human kidney. *Kidney International*. 72: 1081–1087.
- Ghassem, M., Arihara, K., Mohammadi, S., Sani, N. A and Babji, A. S. 2017. Identification of two novel antioxidant peptides from EBN protein hydrolysate. *Food and Function*. 8(5): 2046-2052.

- Goh, D. L. M., Chua, K. Y., Chew, F. T., Liang, R. C. M. Y, Seow, T. K., Ou, K. L., Fong, C. Y. & Lee, B. W. (2001). Immunochemical characterization of edible bird's nest allergens. *Journal of Allergy and Clinical Immunology*. 107(6), 1082-1088.
- Gong, X., Li, J., Shao, W., Wu, J., Qian, H., Ren, R, Espenshade, P. and Yan, N. (2015). Structure of the WD40 domain of SCAP from fission yeast reveals the molecular basis for SREBP recognition. *Cell Research*. 25: 401-411.
- Gradinaru, D., Borsa, C., Ionescu, C. and Prada, G. I. (2015). Oxidized LDL and NO synthesis-biomarkers of endothelial dysfunction and aging. *Mechanism of Ageing and Development*. 151: 101-113.
- Gray, G. R. (1841). *A list of the genera of birds, with an indication of the typical species of each genus*. Oxford: Oxford University Press.
- Graziottin, A, & Serafini, A. (2009). Depression and the menopause: why antidepressants are not enough. *Menopause International Journal*. 15, 76-81.
- Greene, J. A., Portillo, J. C., Carcino, Y. L. & Subauste, C. S. (2015). CD40-TRAF signalling upregulates CX3CL1 and THF- α in human aortic endothelial cells. *PLoS One*. 10(2): e0144133.
- Guo, C. T., Takahashi, T., Bukawa, W., Takahashi, N., Yagi, H., Kato, K., Miyamoto, D., Suzuki, D. & Suzuki, Y (2006). Edible bird's nest extract inhibits influenza virus infection. *Antiviral Research*. 70, 140-146.
- Haghani, A., Mehrbod, P., Safi, N., Aminuddin, N. A., Bahadoran, A., Omar, A. R. & Ideris, A. (2016). In-vitro and in-vivo mechanism of immunomodulatory and anti-viral activity of Edible Bird's Nest (EBN) against Influenza A virus (IAV) infection. *Journal of Ethnopharmacology*. 185, 327-340.
- Haghani, A., Mehrbod, P., Safi, N., Kadir, F. A. A., Omar, A. R. & Ideris, A. (2017). Edible bird's nest modulates intracellular molecular pathways of influenza A virus infected cells. *BMC Complementary and Alternative Medicine*. 17, 22-34.
- Hall, J. E. (2011). Gastrointestinal Physiology. In *Guyton and Hall Textbook of Medical Physiology* 12th edition (pp. 753-803). Saunders Elsevier, Philadelphia.
- Hansson, G. K. & Libby, P. (2006). The immune response in atherosclerosis: a double-edged sword. *National Review of Immunology*. 6: 508-519.
- Heringdorf, D. M. & Jakobs, K. H. (2007). Lysophospholipid receptors: signaling, pharmacology and regulation by lysophospholipid metabolism. *Biochimica et Biophysica Acta*. 1768, 923-940.
- Herold, S., Steinmueller, M., von Wulffen, W., Cakarova, L., Pinto, R., Pleschka, S., Mack, M., Kuziel, W. A., Corazza, N., Brunner, T., Seeger, W. & Lohmeyer, J. (2008). Lung epithelial apoptosis in influenza virus pneumonia: the role of macrophage-expressed TNF-related apoptosis-inducing ligand. *The Journal of Experimental Medicine*. 205, 3065-3077.

- Hobbs, J. J. (2004). Problem in the harvest of edible bird's nest in Sarawak and Sabah, Malaysian Borneo. *Biodiversity and Conservation*. 13(12): 2209-2226.
- Horton, J. D., Goldstein, J. L. and Brown, M. S. (2002). SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver. *Journal of clinical Investigation*. 109(9): 1125-1131.
- Hou, Z., Imam, M. U., Ismail, M., Azmi, N. H., Ismail, N., Ideris, A & Mahmud, R. (2015). Lactoferrin and ovotransferrin contribute toward antioxidative effects of edible bird's nest against hydrogen peroxide-induced oxidative stress in human SH-SY5Y cells. *Bioscience, Biotechnology and Biochemistry*. 79(10), 1570-1578.
- Hutton, H. L., Ooi, J. D., Holdsworth, S. R. and Kitching, A. R. (2016). The NLRPE inflammasome in kidney disease and autoimmunity. *Nephrology*. 21: 736-744.
- Ibrahim, M. M., Zakaria, Z. A. B., Amin, F. M. and Omar, A. R. (2017). Comparative histological evaluation of the sublingual salivary glands of EBN swiftlets in man-made and natural caves. *Pertanika Journal of Tropical Agricultural Science*. 40(1): 19-34.
- Im, S. S., Yousef, L., Blaschitz, C., Liu, J. Z., Edwards, R. A., Young, S. G., Raffatellu, M. and Osborne, T. F. (2011). Linking lipid metabolism to the innate immune response in macrophages through SREBP-1a. *Cell Metabolism*. 13(5): 540-549.
- Infante, J. P. & Huszagh, V. A. (2000). Secondary carnitine deficiency and impaired docosahexaenoic (22: 6n-3) acid synthesis: a common denominator in the pathophysiology of diseases of oxidative phosphorylation and beta-oxidation. *FEBS Letters*. 468: 1-5.
- Institute of Public Health (IPH) (2015). National Health and Morbidity Survey 2015 (NHMS 2015) Vol. II non-communicable diseases. Kuala Lumpur: Ministry of Health, Malaysia.
- Ioannou, G. N., Subramanian, S., Chait, A., Haigh, W. G., Yeh, M. M., Farrell, G. C., Lee, S. P. & Savard, C. (2017). Cholesterol crystallization within hepatocyte lipid droplets and its role in murine NASH. *Journal of Lipid Research*. 58: 1067-1079.
- Isa, K. M. (2016). Edible-Birdnest: Nature and future, presented at 3rd Edible-Birdnest Industry Conference (EBNIC 2016)-Empowering the Essence of Edible-Birdnest. Marriott Hotel, Putrajaya, 11-12 October 2016. Putrajaya. Centre of Excellence for Swiftlets.
- Ismail, M. D., Jalalonmuhali, M., Azhari, Z., Mariapun, J., Lee, Z. V., Abidin, I. M., Wan Ahmad, W. A. & Zuhdi, A. S. M. (2018). Outcomes of STEMI patients with CKD treated with PIC: the Malaysian National CVD Database- Percutaneous Coronary Intervention (NCVD-PCI) registry data from 2007-2014. *BMC Cardiovascular Disorder*. 18:184.

- Jacobs, D. R., Gross, M. D. and Tapsell, L. C. (2009) Food synergy: an operational concept for understanding nutrition. *American Journal of Clinical Nutrition*. 89: 1543S-1548S
- James, A. M., Collins, Y., Logan, A. and Murphy, M. P. (2012). Mitochondrial oxidative stress and the metabolic syndrome. *Trends in Endocrinology and Metabolism*. 23: 429-434.
- James, S. J., Rose, S., Melnyk, S., *et al.* (2009). Cellular and mitochondrial glutathione redox imbalance in lymphoblastoid cells derived from children with autism. *FASEB Journal*. 23: 2374-2383.
- Jayakumar, S., Guillot, S., Argo, C., Redick, J. and Caldwell, S. (2011). Ultrastructure findings in human non-alcoholic steatohepatitis. *Expert Review of Gastroenterology and Hepatology*. 5(2): 141-145.
- Jehle, G., Yackel-Adams, A. A., Savidge, J. A. & Skagen, S. K. (2004). Nest survival estimation: A review of alternatives to the Mayfield estimator. *The Condor*. 106(3), 472-482.
- Jerret, S. A. and Goodge, W. R. (1973). Evidence for amylase in avian salivary glands. *Journal of Morphology*. 137(1): 27-35.
- Jose, J. (2016). Statins and its hepatic effects: newer data, implications and changing recommendations. *Journal of Pharmacy and BioAllied Sciences*. 8(1): 23-28.
- Kahn, S. E., Hull, R. L. & Utzschneider, K. M. (2006). Mechanism linking obesity to insulin resistance and type-2 diabetes. *Nature*. 444 (7121), 840-846.
- Kanis, J. A. (1996). Estrogens, the menopause and osteoporosis. *Bone*. 19, 185-190.
- Karahalil, B., Hare, E., Koc, G., Usli, I., Senturk, K. & Ozkan, Y. (2017). Hepatotoxicity associated with statins. *Achieves of Industrial and Hygiene Toxicology*. 68: 254-260.
- Karalliedde, J. & Gnudi, L. (2011). Endothelial factors and diabetic nephropathy. *Diabetes Care*. 34(2): S291-S296.
- Karimi, I. (2012). Animal model as tools for translation research: focus on atherosclerosis, metabolic syndrome and type-2 diabetes mellitus. *In Tech*. 21: 509-532.
- Kato, Y., Tsuda, T., Hosaka, Y., Akahashi, T., Shirakawa, K., Furusato, S., *et al.* (2001). Effects of trapidil on effector functions of monocytes related to atherosclerotic plaque. *European Journal of Pharmacology*. 12: 371-379.
- Kaur, G. & Meena, C. (2013). Evaluation of anti-hyperlipidaemic potential of combinatorial extract of curcumin, piperine and quercetin in triton-induced hyperlipidaemia in rats. *Science International*. 1: 57-63.

- Kaur, C. & Kapoor, H. C. (2001). Antioxidants in fruits and vegetables-the millennium's health. *International Journal of Food Sciences and Technology*. 36: 703-725.
- Kechrid, Z. & Bouzerna, N. (2004). Effect of zinc deficiency on zinc and carbohydrate metabolism in genetically diabetic C57BL/Ksj Db+/Db+ and non-diabetic original strain (C57BL/Ksj) mice. *Turkish Journal of Medical Sciences*. 34, 367-373.
- Kikuchi, K. & Matahira, Y. (2002). Oral N-acetylglucosamine supplementation improves skin condition of female volunteers: clinical evaluation by a microscopic three-dimensional skin surface analyzer. *Journal of Applied Cosmetology*. 20, 143-152
- Klima, H. (2012). Transport of lipid in plasma. In *Biochemical Pathways: An Atlas of Biochemistry and Molecular Biology 2nd Edition*, eds G. Michal and D. Schomburg (pp. 279-281). New York: Wiley.
- Knight, C. J. (2003). Antiplatelet treatment in stable coronary artery disease. *Heart*. 89(10): 1273-1278.
- Koller, M. M., Cowman, R. A., Humphreys-Beher, M. G. and Scarpace, P. J. (2000). An analysis of submandibular salivary glands function with desipramine and age of female NIA Fischer 344 rats. *Mechanism and Ageing Development*. 119(3): 131-147.
- Kontush, A. & Chapman, M. J. (2006). Functional defective high-density lipoprotein: a new therapeutic target at the crossroads of dyslipidemia, inflammation and atherosclerosis. *Pharmacological Reviews*. 58: 352-374.
- Korff, T., Aufgebauer, K. & Hecker, M. (2007). Cyclic stretch controls the expression of CD40 in endothelial cells by changing their transforming growth factor- β 1 response. *Circulation*. 116: 2288-2297.
- Koutts, J. (1985). Clinching the diagnosis: assessment of homeostatic function. *Pathology*. 17(4): 643-647.
- Kromer, A. & Moosmann, B. (2009). Statin-induced liver injury involves cross-talk between cholesterol and selenoprotein biosynthetic pathways. *Molecular Pharmacology*. 75(6): 1421-1429.
- Ladeiras-Lopes, R., Agewall, S., Tawakol, A., Staels, B., Stein, E., Mentz, R. J., Leite-Moreira, A., Zannad, F. and Koenig, W. (2015). Atherosclerosis: recent trials, new targets and future directions. *International Journal of Cardiology*. 192: 72-81.
- Lamkanfi, M. and Dixit, V. M. (2014). Mechanisms and functions of inflammasomes. *Cell*. 157: 1013-22.

- Lan, H. Y., Nikolic-Paterson, D. J., Zarama, M., Vannice, J. L. and Atkins, R. C. (1993). Suppression of experimental crescentic glomerulonephritis by the interleukin-1 receptor antagonist. *Kidney International*. 43: 479–485.
- Langham, N. (1980). Breeding biology of the edible-nest swiftlet *Aerodramus fuciphagus*. *The International Journal of Avian Science*. 122(4), 447-461.
- Last, A. R., Ference, J. D. & Falleroni, J. (2011). Pharmacologic treatment of hyperlipidaemia. *American Family Physician*. 84(5): 551-558.
- Lee, N., Wong, C. K., Chan, P. K., Lindegardh, N., White, N. J., Hayden, F. G., Wong, E. H., Wonf, K. S., Cockram, C. S., Sung, J. J. & Hui, D. S. (2010). Acute encephalopathy associated with influenza A infection in adult. *Emerging Infectious Disease Journal*. 16, 139-142.
- Lee, P. L. M., Clayton, D. H., Griffiths, R. & Page, R. D. M. (1996). Does behavior reflect phylogeny in swiftlets (Aves: Apodidae)? A test using cytochrome b mitochondrial DNA sequences. *Proceeding of the National Academy of Sciences*. 93 (14): 7091-7096.
- Lenzen, S. (2014). A fresh view of glycolysis and glucokinase regulation: history and current status. *Journal of Biological Chemistry*. 289(18): 12189-12194.
- Levin, B. E. & Dunn-Meynell, A. A. (2006). Differential effects of exercise on body weight gain and adiposity in obesity-prone and –resistance rats. *International Journal of Obese*. 30: 722-727.
- Levin, B. E., Triscari, J. & Sullivan, A. C. (1983). Relationship between sympathetic activity and diet-induced obesity in two rat strains. *American Journal of Physiology*. 259: 1103-1110.
- Lionetti, L., Mollica, M. P., Donizzetti, I., Gifuni, G., Sica, R. *et al.*, (2014). High-lard and high-fish-oil diets differ in their effects on function and dynamic behaviour of rat hepatic mitochondria. *PLoS One*. 9: e92753.
- Levy, E., Spahis, S., Sinnett, D., Peretti, N., Maupus-Schwalm, F., Delvin, E., Lambert, M. and Lavoie, M. A. (2007). Intestinal cholesterol transport protein: an update and beyond. *Current Opinion Lipidology*. 18: 310-318.
- Li, T & Apte, U. (2015). Bile acid metabolism and signalling in cholestasis, inflammation and cancer. *Advanced Pharmacology*. 74: 263-302.
- Liang, H., Bao, X., Dong, X., Tan, R., Zhang, C., Lu, Q. & Cheng, Y. (2007). Antibacterial thymol derivatives isolated from *Centipeda minima*. *Molecules*. 12(8): 1606-1613.
- Liang, W., Menke, A. L., Driessen, A., Koek, G. H., Lindeman, J. H., Stoop, R., Havekes, L. M., Kleeman, R. & Hoek, A. M. (2014). Establishment of general NAFLD scoring system for rodent models and comparison to human liver pathology. *PLoS One*. 9(12): e115922.

- Lievens, D., Eijgelaar, W. J., Biessen, E. A., Daemen, M. J. A. P. and Lutgens, E. (2009). The multi-functionality of CD40L and its receptor CD40 in atherosclerosis. *Thrombosis and Haemostasis*. 102: 206-214.
- Lifton, R. P., Gharavi, A. G. & Geller, D. S. (2001). Molecular mechanism of human hypertension. *Cell*. 104: 545-556.
- Lim, C. K., Cranbrook, G. G. H. & Zoologist, G. B. (2002). *Swiftlets of Borneo: Builders of edible nest*. Borneo: Natural History Publications
- Lin, J. R., Zhou, H., Lai, X. P., Hou, Y., Zian, X. M., Chen, J. N., Wang, P. X., Zhou, L. & Dong, Y. (2009). Genetic identification of edible bird's nest based on mitochondrial DNA sequences. *Food Research International*. 42(8): 1053-1061.
- Lindbohm, N., Gylling, H. & Miettinen, T. A. (2000). Sialic acid content of LDL and its relation to lipid concentration and metabolism of LDL and cholesterol. *Journal of Lipid Research*. 41: 1110-1117.
- Lips, P., Duong, T., Oleksik, A., Black, D., Cummings, S., Cox, D. & Nickelsen, T. (2001). A global study of vitamin D status and parathyroid function in postmenopausal women with osteoporosis: baseline data from multiple data raloxifene evaluation clinical trial. *The Journal of Clinical Endocrinology and Metabolism*. 86(3), 1212-1221.
- Liu, X., Lai, X., Zhang, S., Huang, X., lan, Q., Li, Y., Li, B. C., Zhang, Q., Hong, D. & Yang, G. (2012). Proteomic profile of edible bird's nest. *Journal of Agriculture and Food Chemistry*. 60, 12477-12481.
- Looi, Q. H. and Omar, A. R. (2016). Swiftlet and edible bird's nest industry in Asia. *Pertanika Journal of Scholarly Research Review*. 2(1): 32-48.
- Lu, J. M., Lin, P. H., Yao, Q. & Chen, C. (2010). Chemical and molecular mechanism of antioxidants; experimental approaches and model systems. *Journal of Cellular and Molecular Medicine*. 14(4): 840-860.
- Lumeng, C. N. & Saltiel, A. R. (2011). Inflammatory link between obesity and metabolic disease. *Journal of Clinical Investigation*. 121(6), 2111-2117.
- Lusis, A. J. (2000). Atherosclerosis. *Nature*. 407: 233-241.
- Ma, F. and Liu, D. (2012a). Extraction and determination of hormones in the edible bird's nest. *Asian Journal Chemistry*. 24(1), 117-120.
- Ma, F. and Liu, D. (2012b). Sketch of edible bird's nest and its important bioactivities. *Food Research International*. 48, 559-567.
- Marcone, M. F. (2005). Characterization of edible bird's nest the 'Caviar of the East'. *Food Research International*. 38, 1125-1134.

- Marshall, C. B. (2016). Rethinking glomerular basement membrane thickening in diabetic nephropathy: adaptive or pathogenic? *American Journal of Physiology – Renal Physiology*. 311: F831-843.
- Martin, S. and Parton, R. G. (2006). Lipid droplets: a unified view of a dynamic organelle. *Nature Review Molecular and Cell Biology*. 7: 373-378.
- Martinez-Gonzalez, D., Bonilla-Jaime, H., Morales-Otal, S., Henriksen, J., Velazquez-Moctezuma, O. & Prospero-Gracia, O. (2004). Oleamide and anandamide effect on food intake and sexual behavior of rats. *Neuroscience Letters*. 364(1), 1-6.
- Mathews, C. K., Holde, K. E. V., Appling, D. R. and Anthony-Cahill, S. J. (2013). Lipid metabolism: fatty acids, triacylglycerols, and lipoprotein. In *Biochemistry* 4th edition (pp. 708-752). Pearson Education, New York.
- Matsuda, H., Hakamata, H., Miyazaki, A., Sakai, M., Chang, C. C. Y., Chang, T. Y., Kobori, S., Shichiri, M. and Hiriuchi, S. (1999). Activation of acyl-coenzyme A: cholesterol acyltransferase activity by cholesterol is not due to altered mRNA levels in HepG2 cells. *Biochimica et Biophysica Acta*. 1301: 76-84.
- Matsukawa, N., Matsumoto, M., Bukawa, W., Chiji, H., Nakayama, K., Hara, H. & Tsukahara, T. (2011). Improvement of bone strength and dermal thickness due to dietary edible bird's nest extract in ovariectomized rats. *Bioscience, Biotechnology and Biochemistry*. 75 (3), 590-592.
- Mayne, J., Dewpura, T., Raymond, A., Cousines, M., Chaplin, A., Lahey, K., Lahaye, S. A., Mbikay, M., Ooi, T. C. & Chrétien, M. (2008). Plasma PCSK9 levels are significantly modified by statins and fibrates in humans. *Lipids Health Disease*. 7: 22.
- Mehrbod, P., Omar, A. R., Hair-Bejo, M., Haghani, M. & Ideris, A. (2014). Mechanism of action and efficacy of stains against influenza. *BioMed Research International*. 11, 1125-1134.
- Milan, J., Pinto, X., Munoz, A., Zuniga, M., Rubies-Part, J., Pallardo, L. F., Masana, L. et al., (2009). Lipoprotein ratios: physiological significance and clinical usefulness in cardiovascular prevention. *Vascular Health and Risk Management*. 5: 757-765.
- Millar, J. S. (2001). The sialylation of plasma lipoproteins. *Atherosclerosis*. 154: 1-13.
- Mills, D. C. (2012). M1 and M2 macrophages: oracles of health and disease. *Critical Reviews in Immunology*. 32(6): 463-488.
- Min, H. K., Kapoor, A., Guchs, M., Mirshani, F., Zhou, H., Maher, J., Kellum, J., Warnick, R., Contos, M. J. & Sanyal, A. J. 2012. Increased hepatic synthesis and dysregulation of cholesterol metabolism is associated with severity of NAFLD. *Cell Metabolism*. 15: 665-674.

- Ministry of Health (MOH) (2017). 5th edition of clinical practice guidelines in management of dyslipidaemia. Putrajaya: Ministry of Health, Malaysia.
- Mural, R. J., Adam, M. D., Myers, E. W., Smith, H. O., Miklos, G. L., Wides, R., Halpern, R. et.al. (2004). A comparison of whole-genome shotgun-derived mouse chromosome 16 and the human genome. *Science*. 296(5573): 1661-1671.
- Myasoedova, V. A., Ivashinnikova, G. A., Sobinen, I. A., Ivanova, E. A. and Orekhov E. N. (2017). Blood serum atherogenicity: cellular test for development of anti-sclerotic therapy. *Current Pharmaceutical Design*. 23: 1-12.
- Nagashima, H., Aoka, Y., Sakomura, Y., Uto, K., Sakuta, A., Aomi, S., Kurosawa, H., et al., (2004). MMP-2 is suppressed by trapidil in human abdominal aortic aneurysm wall. *Journal of Vascular Surgery*. 29(2): 447-453.
- Naples, M., Federico, L., Xu, E., Nelken, J. & Adeli, K (2008). Effects of rosuvastatin on insulin sensitivity in an animal model of insulin resistance: Evidence for statin-induced hepatic insulin sensitization. *Atherosclerosis*. 198: 94-103.
- Naoki, I., Toshihisa, K. & Robin, P. (2002) Mechanism of cartilage destruction in osteoarthritis. *Nogoya Journal of Medical Science*. 65, 73-84.
- Nguyen, P., Leray, V., Diez, M., Seisier, S., Le Bloc'h, J., Siliart, B. and Dumon, H. (2008). Liver lipid metabolism. *Journal of Animal Physiology and Nutrition*. 92: 272-283.
- Nguyen, Q. P., Quang, Y. V. & Voisin, J. F. (2002). The white-nest swiftlet and the black-nest swiftlets: A monograph. Paris: Societe Nouvelle Des Editions Boubee.
- Nilsson, S., Makela, S., Treuter, E., Tujague, M., Thomsen, J., Andersson, G., Enmark, E., Pettersson, K., Warner, M. & Gustafsson, J. (2001). Mechanisms of estrogen action. *Physiological Reviews*. 81(4), 1535-1565.
- Novick, D., Kim, S., Kaplanski, G. and Dinarello, C. A. (2013). Interleukin-18, more than a Th1 cytokine. *Seminars in Immunology*. 25: 439-448.
- Nurfatin, M. H., Ety Sharmila, I. K., Nur 'Aliah, D., Zalifah, M. K., Babji, A. S. & Ayob, M. K. (2016). Effect of enzymatic hydrolysis on angiotensin converting enzyme (ACE) inhibitory activity in swiftlet saliva. *International Food Research Journal*. 23(1), 141-146.
- Octava, M., Shkedy, Z., Talloen, W., Verheyen, G. R. and Kasim, A. (2015). Identification of *n-vitro* and *in-vivo* disconnects using transcriptomic data. *BMC Genomic*. 16(1): 615.
- Ohlrogge, J. B. and Jaworski, J. G. (1997). Regulation of fatty acid synthesis. *Annual Review Plant Physiology and Plant Molecular Biology*. 48: 109-136.
- Owens, A. P., Byrnes, J. R. & Mackmann, N. (2014). Hyperlipidemia, tissue factor, coagulation, and simvastatin. *Trends in Cardiovascular Medicine*. 24(3), 95-98

- Park, E. Y., Choi, H., Yoon, J. Y., Lee, I. Y., Seo, Y., Moon, H. S., Hwang, J. H. & Jun, H. S. (2015). Polyphenol-rich fraction of *Ecklonia cava* improves NAFLD in HFD-fed Mice. *Marine Drugs*. 13(11): 6866-6883.
- Parmar, K., Suthar, B., Prajapati, S. & Suthar, A. (2010). Synthesis and biological activity of novel 1,3,5-trisubstituted 1,2,4-triazole derivatives. *Journal of Heterocyclic Chemistry*. 47, 156-161.
- Polimeni, L., Ben, M. D., Baratta, F., Perri, L., Albanese, F., Pastori, D., Violi, F. & Angelico, F. (2015). Oxidative stress: New insights on the association of NAFLD and atherosclerosis. *World Journal of Hepatology*. 7(10): 1325-1336
- Poole, A. R. (1999). An introduction to the pathophysiology of osteoarthritis. *Frontier in Biosciences*. 4(1), 662-670
- Priore, P., Siculella, L. & Gnoni, G. V. (2014). Extra virgin oil phenols down regulate lipid synthesis in primary-cultured rat-hepatocytes. *Journal of Nutritional Biochemistry*. 25, 683-691.
- Proks, S., Girard, C. & Ashcroft, F. M. (2005). Functional effect of KCNJ11 mutations causing neonatal diabetes: enhanced activation by MgATP. *Human Molecular Genetics*. 14(18), 2717-2726.
- Reis, A., Rudnitskaya, A., Blackburn, G. J., Fauzi, N. M., Pitt, A. R. and Spickett, C. M. (2013). A comparison of five lipid extraction solvent systems for lipidomic studies of human LDL. *Journal of Lipid Research*. 54: 1812-1824.
- Rinninger, F., Heine, M., Singaraja, R., Hayden, M., Brundert, M., Ramakrishnan, R. & Heeren, J. (2014). HDL metabolism in LDLR-deficient mice. *Journal of Lipid Research*. 55: 1914-1924.
- Rogers, M. A., Liu, J., Song, B. L., Li, B. L., Chang, C. C. Y. and Chang, T. Y. (2015). Acyl-CoA: cholesterol acyltransferase (ACATs/SOATs): enzymes with multiple sterol substrate and as activators. *The Journal of Steroid Biochemistry and Molecular Biology*. 151: 102-107.
- Roglans, N., Verd, J. C., Peris, C., Alegret, M., Vazquez, M., Adzet, T., Diaz, C., Hernandez, G., Laguna, J. C. & Sanchez, R. M. (2002). High doses of atorvastatin and simvastatin induce key enzymes involved in VLDL production. *Lipids*. 37: 445-454
- Rosen, S. D. (2004). Ligands for L-selectin: homing, inflammation and beyond. *Annual Review Immunology*. 22: 129-156.
- Russo, M. W., Hoofnagle, J. H., Gu, J., Fontana, R. J., Barnhart, H., Kleiner, D. E., Chalasani, N. & Bonkovsky, H. L. (2014). The spectrum of statin hepatotoxicity: Experience of the drug induced liver injury network. *Hepatology*. 60(2): 679-686
- Santos, L., Davel, A. P., Almeida, T. I. R., Almeida, M. R., Soares, E. A., Fernandes, G. J. M. *et al.* (2017). Soy milk versus simvastatin for preventing atherosclerosis

- and left ventricles remodelling in LDL receptor knockout mice. *Brazilian Journal of Medical and Biological Research*. 50(3): e5854.
- Schaeffner, E. S., Kurth, T., Curhan, G. C., Glynn, R. J., Rexrode, K. M., *et al.* (2003). Cholesterol and the risk of renal dysfunction in apparently healthy men. *Journal of American Society in Nephrology*. 14: 2084–2091.
- Schaffer, J. & Fantl, J. A. (2006). 4 urogenital effects of the menopause. *Bailliere's Clinical Obstetrics and Gynaecology*. 10(3), 401-417.
- Schönbeck, U. & Libby, P. (2001). CD40 signalling and plaque instability. *Circulation Research*. 89: 1092-1103.
- Semba, R. D., Najjar, S., Sun, K., Lakatta, E. & Ferrucci, L. (2009). Serum carboxymethyl-lysine, and advanced glycation end product, is associated with increased aortic pulse wave velocity in adults. *American Journal of Hypertension*. 22, 74-79.
- Shah, S. W. and Aziz, N. A. (2014). Morphology of the lingual apparatus of the of the Swiftlet, *Aerodramus faciphagus* (Aves, Apodiformes, Apodidae). *Journal of Microscopy and Ultrastructure*. 2, 100-103.
- Shahzad, K., Bock, F., Dong, W. *et al.* (2015). NLRP3-inflammasome activation in non-myeloid-derived cells aggravates diabetic nephropathy. *Kidney International*. 87: 74–84.
- Sharman, A., Fish, B. L., Moulder, J. E., Medhora, M., Baker, J. E., Mader, M. & Cohen, E. P. (2014). Safety blood sample volume and quality of a refined retro-orbital bleeding technique in rats using a lateral approach. *Laboratory Animal*. 43(2): 63-66.
- Shimano, H and Sato, R. (2017). SREBP-regulated lipid metabolism: convergent physiology- divergent pathophysiology. *Nature Reviews Endocrinology*. 1-21.
- Shi, Y., Guo, R., Wang, X., Yuan, D., Zhang, S., Wang, J., Yan, X. & Wang, C. (2014). The regulation of alfalfa Saponin extract on the key genes involved in hepatic cholesterol metabolism in hyperlipidemic rats. *PLoS One*. 9(2): e88282.
- Simpson, D. M. and Beynon, R. J. 2010. Acetone precipitation of proteins and the modification of peptides. *Journal of Proteome Research*. 9(1): 444-450
- Sirtori, C. R., Lovati, M. R., Manzoni, C., Castiglioni, S., Duranti, M., Magni, C., Morandi, S., D'Agostina, A., and Arnoldi, A. (2004). Proteins of white lupin seed, a naturally isoflavone-poor legume, reduce cholesterolemia in rats and increase LDLReceptor activity in HepG2 cells. *Journal of Nutrition*. 134(1),18-23.
- Soliman, G. A. (2018). Dietary cholesterol and the lack of evidence in cardiovascular diseases. *Nutrients*. 10(6): 780.
- Soran, H., Hama, S., Yadav, R. & Durrington, P. N. (2012). HDL functionality. *Current Opinion in Lipidology*. 23(4): 353-366.

- Snider, M. D., McGarry, J. D. and Hanson, R. W. (2011). Lipid Metabolism: Synthesis, Storage, and Utilization of Fatty Acids and Triacylglycerols. In *Textbook of Biochemistry with Clinical Correlations* 7th edition, ed. T. M. Devlin (673-706). Philadelphia: John Wiley and Sons.
- Snyder, C. H. & Adler, C. H. (2007). The patient with Parkinson's disease: part I-treating the motor symptoms; part II-treating the non-motor symptoms. *Journal of the American Association of Nurse Practitioners*. 19(4), 179-197.
- Song, Z., Jin, R., Yu, S., Nanda, A., Granger, D. L. & Li, G. (2012) Crucial role of CD40 signaling in vascular wall cells in neointima formation and vascular remodeling after vascular interventions. *Atherosclerosis, Thrombosis and Vascular Biology*. 32: 50-64.
- Stancu, C. & Sima, A. (2001). Statins: mechanism of action and effects. *Journal of Cellular and Molecular Medicine*. 5: 378-387
- Stein, E. A., Mellis, S., Yancopoulos, G. D., Stahl, N., Logan, D., Smith, W. B., Lisbon, E., Gutierrez, M., Webb, C., Wu, R., Du, Y., Kranz, T., Gasparino, E. & Swergold, G. D. (2012). Effect of a monoclonal antibody to PCSK9 on LDL cholesterol. *The New England Journal of Medicine*. 366: 1108-1118.
- Steinberg, D. (2006). The thematic review series: the pathogenesis of atherosclerosis. An interpretive history of the cholesterol controversy, part V: the discovery of the statins and the end of the controversy. *Journal of Lipid Research*. 47: 1339-1351.
- Stone, N. J., Robinson, J. G., Lichtenstein, A.H., *et al.* (2013). ACC/AHA guideline on the treatment of blood cholesterol to reduce atherosclerotic cardiovascular risk in adults: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines. *Circulation*. 129(2): S1-S45.
- Sun, J., Dodd, H., Moser, E. K., Sharma, R. & Braciale, T. J. (2014). CD4⁺ T cell help and innate-derived IL-27 induce Blimp-1-dependent IL-10 production by antiviral CTLs. *Nature Immunology*. 12, 327-334.
- Swardfager, W., Lanctot, K., Rothenburg, L., Wong, A., Cappell, J. & Herrmann, N. (2010). A meta-analysis of cytokines in Alzheimer's disease. *Biological Psychiatry*. 68 (10), 930-941.
- Takahashi, Y. and Fukosato, T. (2014). Histopathology of non-alcoholic fatty liver disease/ non-alcoholic steatohepatitis. *World Journal of Gastroenterology*. 20(42): 1539-15548.
- Takuya, Y., Manami, N., Makoto, S., Jun, I. and Ryuichiro, S. (2012). Resveratrol increases the expression and activity of the low-density lipoprotein receptor in hepatocytes by the proteolytic activation of the sterol regulatory element-binding proteins. *Atherosclerosis*. 220, 369-374
- Taylor, F., Huffman, M. D., Macedo, A. F., Moore, T. H., Burke, M., Davey Smith, G., Ward, K. & Ebrahim, S. (2013). Statins for primary prevention of

- cardiovascular diseases. *Cochrane database Systematic Review*. 31(1): CD004816.
- Thapar, M., Russo, M. W. & Bonkovsky, H. L. (2013). Statins and liver injury. *Gastroenterology and Hepatology*. 9(9): 605-606.
- Thevissen, K., Hillaert, U., Meert, E. M. K., Chow, K. K., Cammune, B. P. A., Van Calenbergh, S., & Francois, I. E. J. A. (2008). Fungicidal activity of truncated analogue of dihydrosphingosine. *Bioorganic and Medicinal Chemistry Letters*. 18(13), 3728-3730.
- Thomassen, H. A., Den Tex, R. J., De Bakker, M. A. & Povel, G. D. E. (2005). Phylogenetic relationships amongst swift and swiftlets: a multi locus approach. *Molecular Phylogenetics and Evolution*. 37(1), 264-277.
- Thomassen, H. A., Wiersema, A. T., De Bakker, M. A., Knijff, P., Hetebrij, E. & Povel, G. D. E. (2003). A new phylogeny of swiftlets (Aves: Apodidae) based on cytochrome-b DNA. *Molecular Phylogenetics and Evolution*. 29(1), 86-93.
- Trapani, L., Segatto, M. & Pallottini, V. (2012). Regulation and dysregulation of cholesterol homeostasis: the liver as a metabolic 'power station'. *World Journal of Hepatology*. 4(6): 184-190.
- Trevisan, R., Dodesini, A. R. & Lepore, G. (2006). Lipid and renal disease. *Journal of American Society in Nephrology*. 17: S145-S147.
- Tsutsumi, V., Nakamura, T., Ueno, T., Torimura, T. and Aguirre-Garcia, J. (2017). Structure and ultrastructure of the normal and diseased liver. In *Liver Pathophysiology* 1st edition (23-44). Elsevier, Mexico.
- Reaven, G. M. (2011). Insulin resistance: the link between obesity and cardiovascular disease. *Medical Clinics of North America*. 95(5), 875-892.
- Ruilope, L. M., Sierra, A. D. L., Segura, J. & Garcia-Donaire, J. (2007). The meaning of cardiometabolic risk in hypertensive patients. *US Endocrinology*. 1, 60-63.
- Uchida, K., Nomura, Y., Kadowaki, M., Takse, H., Takano, K. & Takeuchi, N. (1978). Age-related changes in cholesterol and bile acid metabolism in rats. *Journal of Lipid Research*. 19: 544-552.
- Utomo, B., Rosyidi, D., Radiati, L. E., Puspaningsih, N. N. T. & Proborini, W. D. (2014). Protein characterization of extracted water from three kinds of edible bird's nest using SD-PAGE, CBB staining and SDS-PAGE glycoprotein staining, and LC-MS/MS analyses. *IOSR Journal of Agriculture Veterinary Science*. 7, 33-38.
- van der Sluijs, K. F., van Elden, I. J., Nijhuis, M., Schuurman, R., Pater, J. M., Florquin, S., Goldman, M., Jansen, H. M., Lutter, R. & van der Poll, T. (2004). IL-10 is an important mediator of the enhanced susceptibility to pneumococcal pneumonia after influenza infection. *Journal of Immunology*. 172, 7603-7609.

- van der Velde, M., Matsushita, K., Coresh, J., Astor, B. C., Woodward, M., Levey, A. S., Paul Jong, D. & Gansevoort, R. (2011). Prognosis CKD. Lower estimated glomerular filtration rate and higher albuminuria are associated with all-cause and cardiovascular mortality. A collaborative meta-analysis of high-risk population cohorts. *Kidney International*. 79: 1341-1352.
- van der Wulp, M. Y., Verkade, H. J. & Groen, A. K. (2013). Regulation of cholesterol homeostasis. *Molecular and Cellular Endocrinology*. 368(1-2): 1-16.
- van Raalte, D. H. & Diamant, M. (2011). Glucolipotoxicity and beta cells in type 2 diabetes mellitus: target for durable therapy? *Diabetes Research and Clinical Practice*. 93(1), 37-46.
- Vaziri, N. D. (2006) Dyslipidemia of chronic renal failure: The nature, mechanisms, and potential consequences. *American Journal of Physiology-Renal Physiology*. 290: F262–272.
- Wakabayashi, T. (2002). Megamitochondria formation- physiology and pathology. *Journal of Cellular and Molecular Medicine*. 6(4): 497-538.
- Wakamatsu, K., Masaki, T., Itoh, F., Kondo, K. Sudo, K., (1990). Isolation fatty acids amide as an angiogenic principle from bovine mesentery. *Biochemical and Biophysical Research Communications*. 168(2), 423-429.
- Walter, T. C., Chung, J. and Farese Jr., R. V. (2017). Lipid droplet biogenesis. *Annual Review of Cell and Development Biology*. 33: 491-510.
- Watts, G. F. & Karpe, F. (2011). Triglyceride and atherogenic dyslipidemia: extending treatment beyond statins in the high-risk cardiovascular patient. *Heart*. 97, 350-356.
- Wernette-Hammond, M. E., Lauer, S. J., Corsini, A., Walker, D., Taylor, J. M. & Rall, S. C. (1989). Glycosylation of human apolipoprotein. *European Journal of Biology Chemistry*. 264: 9094–9191.
- White, P. J. and Marette, A. (2014). Potential role of omega-3-derived resolution mediators in metabolic inflammation. *Immunology and Cell Biology*. 92: 324–330.
- Wheeler, D. C. & Chana, R. S. (1993). Interaction between lipoproteins, glomerular cells and matrix. *Mineral and Electrolyte Metabolism*. 19: 149-164, 1993
- WHO (2013, March 23). Diabetes. In *World Health Organization*. Retrieved from <https://web.archive.org/web/20130826174444/http://www.who.int/mediacentre/factsheets/fs312/en/>.
- Wong, S. W., Ting, Y. W. & Chan, W. K. (2018). Epidemiology of NAFLD-related hepatocellular carcinoma and its implications. *Journal of Gastroenterology and Hepatology*. 2(5): 235-241.
- Wong, Z. C. F., Chan, G. K. L., Wu, L., Lam, H. H. N., Yao, P., Dong, T. T. X. & Tsim, K. W. K. (2018). A comprehensive proteomics study on edible bird's nest

- using new monoclonal antibody approach and application in quality control. *Journal of Food Composition and Analysis*. 66: 145-151.
- Xioasheng, S. (2011). Qing dynasty 'the new materia medica' and modern herbal health. *New Chinese Medicine*. 43, 154-153.
- Xu, J., Zhu, J., Shi, C., Guo, K. & Yew, D. T. (2007). Effects of genistein on hippocampal neurodegeneration of ovariectomized rats. *Journal of Molecular Neuroscience*. 31, 101-112.
- Yagi, R., McBurney, D. & Laverty, D. (2005). Intra joint comparisons of gene expression patterns in human osteoarthritis suggest a change in chondrocytes phenotype. *Journal of Orthopaedic Research*. 23, 1128-1138.
- Yashiro, T., Nanmoku, M., Shimizu, M., Inoue, J. & Sato, R. (2012). Resveratrol increases the expression and activity of LDLR in hepatocytes by the proteolytic activation of SREBPs. *Atherosclerosis*. 220, 369-374.
- Yew, M. Y., Koh, R. Y., Chye, S. M., Othman, I. & Ng, K. Y. (2015). Edible bird's nest ameliorates oxidative stress-induced apoptosis in SH-SY5Y human neuroblastoma cells. *BMC Complementary and Alternative Medicine*. 14, 391-402.
- Yida, Z., Imam, M. U. & Ismail, M. (2014). In-vitro bioaccessibility and antioxidant properties of edible bird's nest following simulated human gastrointestinal digestion. *BMC Complementary and Alternative Medicine*. 14, 468-474.
- Yida, Z., Imam, M. U., Ismail, M., Der Jiun, O., Sarega, N., Azmi, N. H., Ismail, N., Chan, K. W., Hou, Z. & Yusuf, N. (2015a). Edible bird's nest prevents high fat diet-induced insulin resistance in rats. *Journal of Diabetes Research*. 2015, 760535.
- Yida, Z., Imam, M. U., Ismail, M., Ismail, N. & Hou, Z. (2015b). Edible bird's nest attenuates procoagulation effects of high-fat diet in rats. *Drug Design, Development and Therapy*. 9, 3951-3959.
- Yida, Z., Imam, M. U., Ismail, M., Hou, Z., Abdullah, M. A., Ideris, A & Ismail, N (2015c). Edible bird's nest attenuates high fat diet-induced oxidative stress and inflammation via regulation of hepatic antioxidant and inflammatory genes. *BMC Complementary and Alternatives Medicine*. 15, 310-316.
- Young, B. and Heath, J. W. (2000) *Wheater's Functional Histology* 4th edition (London: Churchill Livingstone). 274-285
- Yu, C. L., Sun, K. H., Shei, S. C., Tsai, C. Y., Tsai, S. T., Wang, J. C., Liao, T. S., Lin, W. M., Chen, H. L., Yu, H. S. & Han, S. H. (1994). IL-8 modulates IL-1 β , IL-6 and TNF- α release from normal human mononuclear cells. *Immunopharmacology*. 27, 207-214.
- Yu-Qin, Y., Liang, X., Hua, W., Hui-Xing, Z., Xin-Fang, Z. & Bu-Sen, L. (2000). Determination of edible bird's nest and its products by gas chromatography. *Journal of Chromatography Science*. 38(1), 27-32.

- Zaheer, M., Chrysostomou, P. & Papademetriou, V. (2016). Hypertension and atherosclerosis: pathophysiology, mechanisms and benefits of BP control. In E. Andreadis (Eds), *Hypertension and Cardiovascular Disease*. Springer, Cham.
- Zhang, C., Boini, K. M., Xia, M. *et al.* (2012). Activation of Nod-like receptor protein 3 inflammasomes turns on podocyte injury and glomerular sclerosis in hyperhomocysteinemia. *Hypertension*. 60: 154–162.
- Zhang, R., Chi, X., Wang, S., Qi B, Yu, X. & Chen, J. L. (2014a). The regulation of autophagy by influenza virus A. *BioMed Research International*. 2014 (7), 498083.
- Zhang, Y., Liu, J., Li, S., Xu, R. X., Sun, J., Tang, Y. & Li, J. J. (2014b). PCSK9 expression is transiently up-regulated in the acute period of myocardial infarction in rat. *BMC Cardiovascular Disorders*. 14: 192.
- Zhipping, H., Imam, M. U., Ismail, M., Ismail., N., Yida, Z., Ideris, A., Sarega, N. & Mahmud, R. (2015). Effect of edible bird's nest on hippocampal and cortical neurodegeneration in ovariectomized rats. *Food and Function*. 6, 1701-1711.
- Zhirnov, O. P. & Klenk, H. D. (2013). Influenza A virus protein NS1 and hemagglutinin along with M2 are involved in stimulation of autophagy in infected cells. *Autophagy*. 5(3), 321-328.
- Zhou, R., Tardivel, A., Thorens, B., Choi, I. and Tschopp, J. (2010). Thioredoxin interacting protein links oxidative stress to inflammasome activation. *National Immunology*. 11: 136-140.
- Zilversmit, D. B. (1979). Atherogenesis: post-prandial phenomenon. *Circulation*. 60: 473-485.
- Zucchetto, A., Serraino, D., Polesel, J., Negri, E., De Paoli, A., Dal Maso, L., Montella, M., La Vecchia, C., Franceschi, S. & Talamini, R. (2009). Hormone-related factors and gynaecological conditions in relation to endometrial cancer risk. *European Journal of Cancer Prevention*. 18, 316-321.
- Zulkefli, S. N., Chua, L. S. & Rahmat, Z. (2017). Protein extraction and identification by gel electrophoresis and mass spectrometry from edible bird's nest samples. *Food Analytical Methods*. 10, 387-398.
- Zulkifli, D. 2019. Nutritional, proteomic and metabolomics analysis of EBN from different regions of Malaysia, MSc Thesis, Universiti Putra Malaysia.