



UNIVERSITI PUTRA MALAYSIA

**IDENTIFICATION AND SCREENING OF ANTIMICROBIAL ACTIVITY OF
LACTIC ACID BACTERIA ISOLATED FROM DAIRY PRODUCTS**

SITI NUR ALIA RAMLI

FBSB 2015 129

PENGESAHAN

Dengan ini adalah disahkan bahawa projek yang bertajuk “Identification and screening of antimicrobial activity of lactic acid bacteria isolated from the dairy products” telah disiapkan serta dikemukakan kepada Jabatan Mikrobiologi oleh SITI NUR ALIA BINTI RAMLI (162568) sebagai syarat untuk kursus BMY 4999 projek.

Disahkan oleh:

.....

Tarikh:

Dr. Wan Zuhainis Saad

Penyelia

Jabatan Mikrobiologi

Fakulti Bioteknologi dan Sains Biomolekul

Universiti Putra Malaysia

.....

Tarikh:

Prof. Madya Dr. Muhajir Hamid

Ketua

Jabatan Mikrobiologi

Fakulti Bioteknologi dan Sains Biomolekul

Universiti Putra Malaysia

ABSTRACT

This study was conducted to screen for the antimicrobial activity of isolated LAB from dairy product and to identify the genera of the LAB. Four isolates of unknown bacteria were obtained from Mycology Laboratory at Biotech 2 with the permission from the supervisor. All the unknown bacteria were isolated from 4 different resources of dairy product. Among four isolates, only one showed the clear zone in the screening of antimicrobial activity against *Escherichia coli* and *Listeria monocytogenes*. The isolate was labelled as sample 1, it showed rod shape under the microscope and was revealed as the gram positive bacteria due its appearance in purple colour on gram staining test. Presumption test was done on sample 1 with a few series of biochemical test and it was identified as *Lactobacillus* sp. The LAB was tested using critical dilution method to determine the quantitatively amount of the antimicrobial activity on *E. coli* and *L. monocytogenes*. Regarding to this finding, it can be further commercialized on fighting the food disease associated by food borne pathogen and and may involve in the food preservative method.

ABSTRAK

*Projek ini telah dilakukan untuk saringan aktiviti antimikrob terhadap bakteria asid laktik daripada produk tenusu dan untuk mengenal pasti jenis bakteria asid laktik. Empat jenis bakteria yang tidak diketahui telah diperolehi dari Makmal Mikologi di Biotech 2 dengan kebenaran daripada penyelia. Semua bakteria tidak diketahui dimaklumkan telah diperolehi daripada 4 sumber tenusu yang berbeza. Antara empat bakteria yang diambil hanya satu bakteria yang tetap zon jelas dalam aktiviti antimikrob terhadap *E. coli* dan *L. monocytogenes*. Bakteria yang telah dilabelkan sebagai sample 1 ini menunjukkan bentuk rod di bawah mikroskop dan dinyatakan sebagai bakteria gram positif kerana kelihatan warna ungu pada ujian gram pewarnaan. Ujian anggapan dilakukan pada sample 1 dengan beberapa siri ujian biokimia dan telah dikenalpasti sebagai *Lactobacillus* sp. LAB telah menjalani kaedah pencairan kritikal untuk menentukan jumlah yang kuantitatif aktiviti antimikrob pada *E.coli* dan *L. monocytogenes*. Berkenaan dengan penemuan ini, ia mungkin boleh dikomersialkan pada masa hadapan untuk melawan penyakit makanan yang berkaitan dengan patogen bawaan dan dalam kaedah pengawetan makanan.*

ACKNOWLEDGEMENT

I would like to express my thanks and gratitude to Dr. Wan Zuhainis Saad, my supervisor, for her guidance, support, initiating the intellectual impetus and absolutely for her unfailing patience throughout the whole course.

I wish to express my appreciation to Miss Laavanya and Mr Nazif for their kind assistance guidance, their willingness to share their knowledge and constructive comments regarding the project and Azri for his helps and ideas.

I would also like to thank all the lectures and general staff of Microbiology department, whose assistance during the course of this study which is extremely valuable.

My heartfelt thanks, gratitude and appreciation to my family especially my parents for their constant support, encouragement and understanding they have given to me.

Finally, I would like to express my deepest thanks to all my friends who have helped me in one way or another. This project is made possible with the help of a lot.

TABLE OF CONTENT

PENGESAHAN	i
ABSTRACT	ii
ABSTRAK	iii
ACKNOWLEDGEMENT	iv
TABLE OF CONTENT	v
LIST OF TABLES	viii
LIST OF FIGURE	ix
ABBREVIATIONS	x
CHAPTER 1	1
INTRODUCTION	1
CHAPTER 2	4
LITERATURE REVIEW	4
2.1 Lactic acid bacteria	4
2.2 Antimicrobial properties of lactic acid bacteria	8
2.2.1 Homofermentative metabolism of hexose	10
2.2.2 Heterofermentative metabolism of penthose	10
2.2.3 Bacteriocins	10
2.2.4 Hydrogen peroxide	11
2.2.5 Carbon dioxide	12
2.3 Products of lactic acid	13
CHAPTER 3	14
MATERIALS AND METHODS	14
3.1 Media preparation	14
3.2 Methods	16

CHAPTER 4	21
RESULTS AND DISCUSSIONS	21
4.1 Identification of lactic acid bacteria	21
4.1.1 Morphology	21
4.1.1.1 Gram stain	22
4.1.2 Biochemical test	23
4.1.2.1 Catalase	23
4.1.2.2 Carbohydrate fermentation	25
4.2 Antimicrobial activity screening test	26
4.3 Determine the antimicrobial activity	29
CHAPTER 5	31
CONCLUSION AND RECOMMENDATIONS	31
REFERENCES	32
APPENDICES	37

LIST OF TABLES

Table	Caption	Page
1	Food borne pathogen	2
2	Phenotypic characteristics for differentiating genus <i>Lactobacillus</i> from the other genera of the LAB	5
3	Low molecular mass antimicrobial metabolites of LAB	9
4	Carbohydrate fermentation of sample 1	25
5	Antimicrobial activity of isolated bacteria labelled as sample 1, sample 2, sample 3, sample 4 against eight pathogens	27
6	Result of critical dilution method for sample 1 against <i>E. coli</i> and <i>L. monocytogenes</i>	29

LIST OF FIGURES

Figures	Caption	Page
1	The phylogenetic tree of LAB base on the 16 rRNA gene sequence.	7
2	The single pure colony of LAB labelled as sample 1	21
3	Sample 1 showed purple in colour of gram stain test (1000× magnifications)	22
4	Non spore formation cell of sample 1 (1000× magnifications).	23
5	Negative result of sample 1 on catalase test	24
6	Positive result of catalase test	24
7	Plate shows the clear zone made by bacteria sample 1 inhibited growth of <i>L. monocytogenes</i>	28
8	Plate shows clear zone made by isolated bacteria labelled as sample 1 inhibited growth of <i>E. coli</i> .	28
9	Plate shows the clear zone on Muller-Hinton agar to determine the antimicrobial activity of sample 1 against <i>E. coli</i>	30
10	Plate shows the clear zone on Muller-Hinton agar to determine the antimicrobial activity of sample 1 against <i>L. monocytogenes</i>	30

ABBREVIATIONS

Abbreviations for the names of units used in the text:

AU	Arbitrary unit
LAB	Lactic acid bacteria
μl	micro litre
g	gram
mg	milligram
μg	microgram
NB	nutrient broth
OD	optical density
nm	nanometre
l	litre
ml	millilitre
EHEC	Enteromorrhagic <i>Escherichia coli</i>
Da	Dalton
Cfu	colony forming unit
$^{\circ}\text{C}$	Degree Celsius
nm	nanometers
GC content	guanine-cytosine content

CHAPTER 1

INTRODUCTION

Food borne disease still linger as one of the greatest public health problems in Malaysia and other developing country. Ingestion of food stuff contaminated with microorganisms or chemical will lead to the outbreak of food born disease such as cholera, typhoid fever, hepatitis A, dysentery and food poisoning. Most of the food borne illnesses is excruciating, meaning they happen suddenly and most of patient recover on their own without treat but sometimes food borne disease may lead to more serious complication such as dehydration and will become worst which can cause chronic health problem and even can cause death. Symptoms of food borne disease are dependent on the type of microorganism or chemical present but most of them share common symptoms including vomiting, diarrhea or bloody diarrhea, abdominal pain, fever, chills and all these symptoms can extent from lenient to grievous and can last a few hours to several days.

E. coli, *Staphylococcus aureus*, *Bacillus cereus* and *Clostridium botulinum* always become the popular microorganisms that can cause the food borne disease (Cappucino and Sherman, 2005). All these pathogen can infect human gastrointestinal tract and some of them can dangerously affect the nervous system which show some symptoms such as headache, tingling, weakness, numbness of skin, blurred vision and dizziness. If we want to prevent the food poisoning from happen, we have to deal with the prevention from getting infected by the pathogen and we have to think about how to kill microorganism or suppressing their growth when if we already got infected. Table 1 shows the type of food borne pathogen,

characteristic and example for each pathogen that always being isolated from the person who got food poisoning.

Table 1 : Food borne pathogen (Sharifa et al., 2013)

Type of pathogens	Characteristics	Example
Bacteria	In environment or animal resevoir and multiply in or on food	<i>Campylobacter</i> and <i>Salmonella</i>
Virus	Reproduce only in within living cell and remain infectious in food	Hepatitis A and Norwalk virus
Parasite	Reproduce within the host cells and cannot multiply on food	<i>Gardia lamblia</i>
Toxin	Produce by some bacteria or chemical	<i>Bacillus cereus</i> , <i>Staphylococcus aureus</i> , <i>E.coli</i> and <i>Clostridium botulinum</i>

Most of parents prefer to give dairy product to their children to prevent them from get infected by the pathogen. They believe the content of the product especially lactic acid bacteria that present in the milk can help to protect their children from the pathogen. From here lactic acid bacteria isolated from dairy product have received attention as a potential microorganism that can help us to protect our health. Japan 1930, Dr. Minoru Shirota, who was conducting research in a Microbiology Lab the Medical Faculty of Kyoto University, became the first person in the world that succeed in culturing a strain of lactic acid bacteria which give beneficial to human health especially to prevent from being infected by food borne pathogen. He wanted

to develop a stronger strain of lactic acid bacteria which would destroy the harmful bacteria living in the intestinal because the pathogen can cause the irritation of gastro intestinal tract.

Lactic acid bacteria are gram positive bacteria that able to ferment glucose primarily into lactic acid. Beside lactic acid, it also produced acetic acid, hydrogen peroxide, carbon dioxide and bacteriocin which are useful in inhibiting food borne pathogen (Senthikumar et al., 2012). The low external pH of the pathogens surrounding caused the acidification of pathogens' cell cytoplasm and inhibit the growth of the pathogens. Normally lactic acid bacteria can be found in the dairy product. Few of the lactic acid bacteria are pathogenic to animals especially some members of the genus *Streptococcus*. Many genera of bacteria produce lactate as a primary or secondary end product of fermentation but term lactic acid bacteria is well known reserved for genera of *Lactobacillus*, which includes *Lactobacillus*, *Leuconostoc*, *Pediococcus*, *Lactococcus* and *Streptococcus* (Fujitoshi et al., 2005). Therefore, the aim of this final year project was to identify lactic acid bacteria from the dairy product in addition to detect the antimicrobial effects of isolated lactic acid bacteria against a few types of food borne pathogen.

Hypothesis

1. Lactic acid bacteria which isolated from dairy product may have antimicrobial activity.

Objectives

1. To identify the lactic acid bacteria that has the antimicrobial activity against pathogenic bacteria.
2. To measure the amount of the antimicrobial activity
3. To identify the species of the lactic acid bacteria that has the antimicrobial activity.

REFERENCES

- Annuk, H., Schopetova, J., Kullisaar, T., Songisepp, E., Zilmer, M., Milelsaar, M. (2003). Characterization of intestinal *Lactobacilli* as putative probiotic candidates. *J. Appl. Microbio.*, 94: 403–412.
- Aslim, B., Yukesakdag, Z. N., Sarikaya, E. and Beyati. Y. (2005). Determination of The Bacteriocin-like Substances Produced by Some Lactic Acid Bacteria Isolated from Turkish Dairy Products. *LWT- Food Science and Technology*, 38:691-694.
- Axelsson, L.T., Chung, T. C., Dbrogosz, W. J., Lindgren, S. E. (1989). Production of a broad spectrum antimicrobial substance by *Lactobacillus reuteri*. *Microbiol Ecol in Health Disease*, 2: 131-136.
- Björkroth, J. and Holzapfel, W. (2003). Genera *Leuconostoc*, *Oenococcus* and *Weisella*. In the prokaryotes : An Evolving Electronic Resources for the Microbiology Community. Retrieved from <http://link.springer-ny.com/link/servic/books/1025/>
- Brashears, M. M., Amezcuita, A. & Jaroni, D. (2005). Lactic acid bacteria and their uses in animal feeding to improve food safety . *Adv. Food Nutr. Res.*, 50: 1-31
- Byczkowski, J. and Gessner, T. (1988). Biological role of superoxide ion- radical. *International Journal of Biochemistry*. 20:569-580.
- Calderon, M., Loiseau, G., Guyot, J. P. (2001). Nutritional improvement and simplified cultivation medium to study the growth and energetic of sourdough lactic acid bacterium *Lactobacillus fermentum* Ogi EI during heterolactic fermentation of starch. *J. Appl. Microbiol*, 90: 1-9.
- Campbell, N. A., Reece, J. B., Urry, L. A., Cain, M. L., Wasserman, S. A., Minorsky, P. V., Jakckson, R. B. (2008). *Biology*. San Francisco, United States of America: Pearson Education, Inc.
- Carminati, D., Giraffa, G. and Bossi, M. G. (1989). Bacteriocin like inhibitor of *Streptococcus lactis* against *Listeria monocytogenes*. *J. Food Prot*, 52: 614-617.
- Caplice, E., Fitzgerald, G. F. (1999). Food fermentation: role of microorganisms in food production and preservation. *Int J Food Microbiol*, 50: 131-149.
- Cappucino, J. G., Sherman, N. (2005). *Microbiology; A Laboratory Manual*. San Francisco, United States of America: Pearson Education, Inc.
- Chen, H., Hoover, D.G. (2003). Bacteriocins and their food applications. *Compr. Rev. Food Sci. Food Saf.*, 2 : 82–100.

- Collins, J. W., La Ragione, R. M. and Woodward, M. J., (2009). L.E.J. Searle: Application of Prebiotics and Probiotics in Livestock. In: Prebiotics and Probiotics Science and Technology, D. Charalampopoulos, R.A. Rastall (Eds.). *Springer Science Business Media B.V., New York, NY, USA* :1123–1192.
- Daeschel, M. A. (1989). Antimicrobial substances from lactic acid bacteria for use as food preservatives. *Food Technol.* 43:164–167.
- Dahiya, R. S and Speck, M. L. (1968). Hydrogen peroxide formation by *Lactobacillus* and its effect on *Staphylococcus aureus*. *J.Dairy. Sci*, 51(10): 1568-1572.
- Deegan, L. H., Cotter, P., Hill, C. and Ross, P. (2006). Bacteriocins: Biological tools for biopreservation and shelf-life extension. *Int. Dairy J.*, 16: 1058-1071.
- Deraz, S. F., Karlsson, E. N., Hedstorm, M., Andersson, M. M and Mattiasson B. (2005). Purification and characterisation of acidocin D20079, a bacteriocin produced by *Lactobacillus acidophilus* DSM 20079. *J.Biotechnol*, 117: 343-354.
- Derek, A. A., Joost, V. D. B., Inge, M. K. M., Jack, T. P., Antonius, J. A. V. M. (2009). Anaerobic homolactate fermentation with *Saccharomyces cerevisiae* results in depletion of ATP and impaired metabolic activity. *FEMS Yeast Research*, 9(3): 349-357.
- Eklund, T. (1984). The effect of carbon dioxide on bacterial growth and on uptake processes in the bacterial membrane vesicles. *International Journal of Food Microbiology*, 1: 179-185.
- Eva Rodriguez, Juan L., Arquos, Manuel Nunez, Pilar Gaya and Margarita Medina.(2005). Combined effect of High- pressure treatment and bacteriocin producing LAB on inactivation of *E. coli* in raw milk cheese. *Applied Environmental Microbiology*, 3399-3404.
- Farber, J. M. (Microbiology aspects of modified atmosphere packaging technology-areview. *Journal of Food Protection*, 54: 58-70.
- Fleming, D. W., Cochi, S. L., Macdonald, K. L., Brondum, J., Hayes, P.S., Plikaytis, B. D. (1985). Pasteurized Milk as a Vehicle of Infection in an Outbreak of Listeriosis. *New England Journal of Medicine*. 312: 404-407.
- Fujitoshi, Y., Yi-sheng, C., Takashi, S. (2005). Isolation and characterization of lactic acid bacteria from solid in vineyard, Institute of Enology and Viticulture, University of Yamashi. *Journal of General Applied Microbiology*, 51: 313-318.
- Garneu, S., Martin, N. I., and Vederas, J. C. (2002). Two-peptide bacteriocins produced by lactic acid bacteria. *Biochimie*, 84: 577-592.

- Gobben, G. J., I. Cain-Joe, V. A. Kitzen., I. C. Boels., F. Boer, J. A. M. De Bont. (1998). Enhancement of exopolysaccharide production by *Lactobacillus delbruekii* subsp. *bulgaricus* NCFB 2772 with a simplified define medium. *Applied. Environ. Microbiol.*, 64 (4): 1333-1337.
- Hammes, W. D and Hertel, C. (2003). The genera *Lactobacillus* and *Carnobacterium*. In the Prokaryotes: An Evolving Electrone Resources for Microbiology Community. Retrived from <http://link.springer-ny.com/link/servic/books/1025/>
- Harrid, L. J., M. A. Daeschel, M. E. Stile, and Klaenhammer, T. R. (1989). Antimicrobial activity of lactic acid bacteria against *Listeria monocytogenes*. *J. Food Prot*, 52: 384-387.
- Jacobsen,. C. N., Rosenfeldt Nielson, V., Hayford, A. E., Moller, R. L., Michaelsen, K. F., Pearregaard, A., Sandstrom, B., Tvede, M., Jakobsen, M. (1999). Screening of Probiotic Activities of Forty-seven Strains of *Lactobacillus* sp. By In Vitro Techniques and Evaluation of The Colonization Ability of Five Selected Strains in Humans. *Applied and Environment Microbiology*, 65: 4949-4956.
- Jay, J. M. (1982). Antimicrobial properties of diacetyl. *Applied and Environmetal Microbiology*, 44: 525-532.
- Jyoti, B., Suresh, A. K. & Venkatesh, K.V. (2003). Diacetyl production and growth of *Lactobacillus rhamnosus* on multiple substrates. *World J. Microbiol. Biotechnol*, 19 : 509–515
- Kashket, E. R. (1987). Bioenergetics of lactic acid bacteria: Cytoplasmic pH and osmotolerance. *FEMS Microbiology Reviews*, 46: 233-244.
- Klaenhammer, T. R. (1988). Genetics of bacteriocins produced by Lactic acid bacteria. *FEMS Microbiology Reviews*, 12: 39 - 85.
- Ko, S. H. & Ahn, C. (2000). Bacteriocin production by *Lactococcus lactis* KCA 2386 isolated from white kimchi. *Food Science and Biotechnology*, 9: 263 - 269.
- Kong, S. and Davison, A. J. (1980). The role of interaction between O₂, H₂, OH⁻, e⁻, and O²⁻ in the free radical damage to biological systems. *Archives of Biochemistry and Biophysics*, 204: 18-29.
- Lin, W., Yu B., Jang, S., Tsen, H. (2007). Different Probiotic Properties of *Lactobacillus fermentum* Strains Isolated from Swine and Poultry. *Anaerobe*. 13: 107-113.
- Lindgren, S. E. and Dobrogosz, W. J. (1990). Antagonistic activities of lactic acid bacteria in food and feed fermentations. *FEMS Microbiology Reviews*, 7:149-163).

- Maithili, S. S., Viveka, S., Kamaraj, P., Ramanathan, G. (2014). Isolation and identification of lactic acid bacteria from fermented products and its spectrum of inhibitory activity. *Int. Res J Pharm. App Sci.*, 4(1): 88-93.
- Mayr-Harting, A., Hedges, A and Berkeley, R.C.W. (1972). Methods for Studying Bacteriocin. In "Method in Microbiology" (T. Bergen and J. R Norris. Eds), Vol. 7A, pp. 315-422. Academic Press, Inc., London.
- Oranusi, S. V., Umoh, V. J., Kwaga, J. K. P. (2003). Hazards and critical control points of Kunu-zaki, a non-alcoholic beverage in Northern Nigeria. *Food Microbiol*, 20: 127-132.
- Piard, J. C and Desmazeaud, M. (1992). Inhibiting factors produced by lactic acid bacteria: 2. Bacteriocin and other antibacterial substances. *Lait*, 72: 113-142.
- Podolok, P. K., Zayas, J. F., Kastner, C. L and Fung, D. Y. C (1996). Inhibition of *Listeria monocytogenes* and *Escherichia coli* 0157:H7 on beef by application of organic acids. *Journal of Food Protection*, 59:370-372.
- Senthikumar, P. K, Retha, D., Ramya, D. and Stella, D. (2012). Antibacterial potential of lactic acid bacteria and its metabolites against food borne pathogens. *International Journal of Pharmaceutical & Biological Archives* 2012.vol 3 no 2: 342-347.
- Schillinger, U., Lucke, F. (1989). Antimicrobial activity of *Lactobacillus sake* isolated from meat. *Applied Environ. Microbiol*, 55: 1901-1906.
- Schleifer, K., Ludwig, W. (1995). Phylogenetic identification and in situ detection of individual microbial cells without cultivation. *Microbiology Review*, 59: 70-78.
- Sharifa, E. W. P., Netty, D., Sangaran, D. (2013). Paper review of factors, surveillance and burden of food borne disease outbreak in Malaysia. *Malaysian Journal of Public Health Medicine* 2013, 13(2): 98-105.
- Siragusa, G. R. and Johnson, M. G. (1989). Inhibition of *L.monocytogenes* growth by the lactoperoxidase-thiocyanate H₂O₂ antimicrobial system. *Appl. Environ. Microbiol*, 55 (11): 2802-2805.
- Snijders, J. M., Van Logtestijn G., Mossel, D. A. A., and Smulders, F. J. M. (1985). Lactic acid as a decon, J. taminat is slaughter and processing procedures. *Veterinary Quarterly*, 7: 277-282.
- Spelhaug, S. R. and Harlander, S. K.. (1989). Inhibition of food borne bacterial pathogens by bacteriocins from *Lactococcus lactis* and *Pediococcus pentosaceus*. *J. Food Prot*, 52: 856-862.

- Suskovic, J., Kos, B., Beganovic, J., Lebos P. A., Habjanic, K. and Matosic, S. (2010). Antimicrobial Activity of Lactic Acid Bacteria. *Food Technol. Biotechnol.*,48 (3): 296–307
- Vanderbergh, P. A. (1993) . Lactic acid bacteria, their metabolic products and interference with microbial growth, *FEMS Microbiol. Rev.*, 12: 221–238.
- Wagner, M. K., and Moberg, L. J. (1989). Present and future use of traditional antimicrobials. *Food Technology*, 43: 143-147.
- Watson, J. A. and Schubert, J. (1969). Action of hydrogen peroxide on growth inhibition of *Salmonella typhimurium*. *J. Gen. Microbiol*, 57(1): 25-34.

