



UNIVERSITI PUTRA MALAYSIA

**DETERMINATION OF THE GENOMIC STRUCTURE OF
HOMOGENISATE GERANYLGERANYL TRANSFERASE GENE OF OIL
PALM**

MOHD FAUZE HAZIS ABDULLAH

FP 2015 145

**DETERMINATION OF THE GENOMIC STRUCTURE OF HOMOGENTISATE
GERANYLGERANYL TRANSFERASE GENE OF OIL PALM**

The logo of Universiti Putra Malaysia (UPM) is a shield-shaped emblem. It features a red and white color scheme. At the top left, the letters 'UPM' are written in white on a red background. In the center, there is a stylized white tree or plant motif. To the right of the tree, there is an open book with text on its pages. The shield is surrounded by a grey border.

MOHD FAUZE HAZIS BIN ABDULLAH

FACULTY OF AGRICULTURE

UNIVERSITI PUTRA MALAYSIA

SERDANG, SELANGOR DARUL EHSAN

2014/2015

DETERMINATION OF THE GENOMIC STRUCTURE OF HOMOGENTISATE
GERANYLGERANYL TRANSFERASE GENE OF OIL PALM

The logo of Universiti Putra Malaysia (UPM) is a shield-shaped emblem. It features a red and white color scheme. At the top left, the letters 'UPM' are written in white on a red background. In the center, there is a stylized white tree or plant motif. To the right of the tree, there is an open book. The shield is surrounded by a grey border.

MOHD FAUZE HAZIS BIN ABDULLAH

FACULTY OF AGRICULTURE

UNIVERSITI PUTRA MALAYSIA

2014/2015

DETERMINATION OF THE GENOMIC STRUCTURE OF HOMOGENITISATE
GERANYLGERANYL TRANSFERASE GENE OF OIL PALM

BY

MOHD FAUZE HAZIS BIN ABDULLAH

A project submitted to Faculty of Agriculture, Universiti Putra Malaysia, in fulfillment
of the requirement of PRT 4999 (Final Year Project) for the award of the degree of
Bachelor of Agriculture Science

FACULTY OF AGRICULTURE

UNIVERSITI PUTRA MALAYSIA

2014/2015

CERTIFICATION

This project paper entitled **“Determination of The Genomic Structure of Homogentisate Geranylgeranyl Transferase Gene of Oil Palm”** is prepared by Mohd Fauze Hazis Bin Abdullah and submitted to the Faculty of Agriculture in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Agriculture Science.

Student's name:

Student's signature:

Mohd Fauze Hazis Bin Abdullah

Matric No: 168577

Certified by:

.....
(PROF. DATIN DR. SITI NOR AKMAR ABDULLAH)

Project Supervisor,

Department of Agricultural Technology,

Faculty of Agriculture,

Universiti Putra Malaysia.

Date:

ACKNOWLEDGEMENT

Bismillahirrahmanirrahim, in the name of Allah the Most Gracious, and the Most Merciful. Alhamdulillah, praises to Allah S.W.T for giving me the strength, passion and capability in finishing my final year project entitled Determination of The Genomic Structure of Homogentisate Geranylgeranyl Transferase Gene of The Oil Palm on time.

First of all, I would like to thank my beloved supervisor, PROF. DATIN DR. SITI NOR AKMAR ABDULLAH for her guidance, advice, and patience in supervising this research project. Her willingness to encourage me contributed tremendously to my project.

Special appreciation goes to Miss Farah Hanan Binti Abu Hanifah and Mrs. Azzreena Mohamad Azzeme who had always guided and assisted me in my laboratory works. I also would like to thank all the staff of Agrobiotechnology Laboratory for being kind and helpful to me during the time I carried out my work in the laboratory.

Next, my deepest gratitude and appreciation goes to my beloved parents, Abdullah Bin Ismail and Normah Binti Atan and also to all my family members for their endless love, encouragement and prayers. Last but not least, special thanks to my friend, Khairul Anuar Mis Man and to those who directly or indirectly contributed in this research, your kindness is very meaningful. Thank you very much.

TABLE OF CONTENTS

Contents	Page
ACKNOWLEDGEMENT	i
TABLE OF CONTENT	ii
LIST OF TABLES	iv
LIST OF FIGURES	v
ABBREVIATION	vii
ABSTRACT	ix
ABSTRAK	x
CHAPTER 1: INTRODUCTION	1
CHAPTER 2: LITERATURE REVIEW	3
2.1 Vitamin E	3
2.2 Benefits of Tocotrienols	4
2.3 Biosynthesis of Tocopherols and Tocotrienols	6
2.4 Oil Palm Breeding in Malaysia	8
2.5 Germplasm Collections	9
2.6 Molecular Marker	10
2.7 Gene Structure	11

CHAPTER 3: MATERIALS AND METHODS	12
3.1 Leaf Sampling	12
3.2 Genomic DNA Extraction	12
3.3 DNA Concentration and Purity Quantification	14
3.4 Primer Design	14
3.5 Polymerase Chain Reaction (PCR)	15
3.6 Electrophoresis of PCR Product	16
3.7 Gel Extraction	17
3.8 Sequencing	18
CHAPTER 4: RESULTS AND DISCUSSION	19
4.1 PCR Product	20
4.2 Purified PCR product	23
4.3 Alignment of Sequencing Results	25
CHAPTER 5: CONCLUSION	29
REFERENCES	30

LIST OF TABLES

	Page
Table 4.1 List of primer pairs that have been designed. Each primer pair consists of forward and reverse primer.	19
Table 4.2 The sizes of intron by each primer set in the amplified product.	20
Table 4.3 The concentration and nucleic acid purity of each sample obtained from the gel extraction.	22

LIST OF FIGURES

	Page
Figure 2.1: The structures of tocopherols and tocotrienols which differ in the carbon phytyl tail.	4
Figure 2.2: The biosynthesis of tocopherols and tocotrienols	7
Figure 3.1: Software program for designing primer	14
Figure 3.2: Software program for aligning the sequence	18
Figure 4.1: PCR product analysed by using 1 % agarose gel from primer pair 1 at the annealing temperature of 60 °C	21
Figure 4.2: PCR product analysed by using 1 % of agarose gel from primer pair 2 at the annealing temperature of 60 °C	21
Figure 4.3: PCR product analysed by using 1 % of agarose gel from primer pair 3 at the annealing temperature of 58° C	22
Figure 4.4: Gel extraction product of Fsq1 analyzed by agarose gel electrophoresis	23
Figure 4.5: Gel extraction product of Fsq2 analyzed by agarose gel electrophoresis	24
Figure 4.6: Gel extraction product of Fsq3 analyzed by agarose gel electrophoresis	24

Figure 4.7 & 4.8: Percentage of alignment of the sequence from Fs_{q1} when aligned with the sequence of the previous study. 26

Figure 4.9 & 4.10: Percentage of alignment of the sequence from Fs_{q2} when aligned with the sequence of the previous study. 27

Figure 4.11 & 4.12: Percentage of alignment of the sequence from Fs_{q3} when aligned with the sequence of the previous study. 28



ABBREVIATION

DNA	deoxyribonucleic acid
PE	Phycoerythrin
EB	Elution buffer
QG	QIAGEN
UV	Ultraviolet
TAE	Tris-Acetate -EDTA
PCR	Polymerase Chain Reaction
RNA	Ribonucleic acid
TE	Tris-EDTA
PCI	Phenol chloroform isoamyl
CI	Chloroform isoamyl
CTAB	Cetyltrimethyl ammonium bromide
DOA	Department of Agriculture
HMG-CoA	3-hydroxy-3-methylglutaryl-coenzyme A
HGGT	Homogentisate geranylgeranyl transferase
HPT	Homogentisate phytyltransferase

PDP	Phytyl diphosphate
g	gram
ml	milliliter
nm	nanometre
bp	base pair
μ l	microliter
$^{\circ}$ C	degree Celsius
γ	gamma
α	alpha
mg/kg	milligram/kilogram
%	percentage

ABSTRACT

Elaeis guineensis originated from West Africa is the only species that was planted in the Malaysian plantation as a commercial oil palm. Since 1971, Malaysia has been the largest producer and exporter of oil palm. Oil palm is rich in vitamin E that consists of two major molecules which are tocopherols and tocotrienols. Oil palm is the richest source of tocotrienol. Homogentisate geranylgeranyl transferase (HGGT) is the enzyme that initiates tocotrienol synthesis in monocot seeds such as oil palm. The main objective of this study was to compare the genomic structure of HGGT gene of oil palm with other genomic structure of HGGT obtained from the previous study by using polymerase chain reaction (PCR) based method. Young leaves of oil palm were collected from the plantation area at Universiti Putra Malaysia, Serdang, Selangor, Malaysia and the leaves were extracted by using modified cetyltrimethyl ammonium bromide (CTAB). From this study, the genomic structure obtained was not exactly the same with the previous study due to the sequencing errors or the presence of introns in the sequence. From the alignment between genomic and the cDNA sequence, the introns that separate the coding sequence was able to be determined.

ABSTRAK

Elaeis guineensis yang berasal dari Afrika Barat adalah satu-satunya spesies yang ditanam di ladang Malaysia sebagai kelapa sawit komersial. Sejak 1971, Malaysia telah menjadi pengeluar dan pengimport terbesar kelapa sawit. Kelapa sawit yang kaya dengan vitamin E yang terdiri daripada dua molekul utama iaitu tokoferol dan tokotrienol. Kelapa sawit adalah sumber terkaya tokotrienol. *Homogentisate Geranylgeranyl Transferase* (HGGT) adalah enzim yang memulakan sintesis tokotrienol dalam biji monokot seperti kelapa sawit. Objektif utama kajian ini adalah untuk membandingkan struktur genomik gen HGGT kelapa sawit dengan struktur genomik gen HGGT yang diperoleh dari kajian yang lepas dengan menggunakan kaedah berasaskan tindak balas rantaian polymerase (PCR). Daun muda kelapa sawit dikumpulkan dari ladang di Universiti Putra Malaysia, Serdang, Selangor, Malaysia dan daun ini diekstrak dengan menggunakan penyelesaian CTAB yang diubahsuai. Daripada kajian ini, struktur genomik yang diperoleh adalah tidak sama dengan kajian yang lepas disebabkan oleh kesilapan pengurutan atau kehadiran intron dalam jujukan. Daripada penjajaran antara jujukan genom dan cDNA, intron yang memisahkan urutan pengkodan dapat ditentukan.

CHAPTER 1

INTRODUCTION

Oil palm is the most productive oil-bearing crop. Although it is planted on only 5% of the total world vegetable oil acreage, palm oil accounts for 33% of vegetable oil and 45% of edible oil worldwide, but increased cultivation competes with dwindling rainforest reserves. The first evidence of palm oil consumption in human diets dates back as far as 3000 BC, with a long history of use in Western Africa (Kiple and Ornelas., 2000). Oil palm trees were introduced to the West (Brazil, West Indies) in the 15th century by the Portuguese and to the East (Indonesia) by the Dutch in the 19th century (Sundram *et al.*, 2003).

Commercial cultivation began in the early twentieth century and despite the long breeding cycle (10 to 12years) and large land requirement for field trials (Mayes *et al.*, 1997), high yield breeding materials (up to 12 tonnes per hectare per year (t/ha-1yr-1) (Corley and Tinker., 2003) have been developed in less than 100 years. As such, the largely undomesticated oil palm is an ideal candidate for genomic-based tools including expressed sequence tags (ESTs) (Jouannic., 2005) (Ho., 2007) (Low., 2008) and transcriptome sequencing of the oil palm fruit during development, maturation and ripening (Tranbarger., 2011) to harness the potential of this remarkably productive crop. Furthermore, oil palm is rich with vitamin E that consists of two major molecules which are tocopherols and tocotrienols. In the biosynthesis of vitamin E, both molecules are synthesized through two important converging pathways that fused the side chain building block and head group together (Schnieder., 2005).

Homogentisate geranylgeranyl transferase (HGGT) is the enzyme that initiates tocotrienol synthesis in monocot seeds such as oil palm. This enzyme is related to homogentisate phytyltransferase (HPT), which catalyzes the prenylation step in tocopherol synthesis. The application of DNA markers could greatly improve precision and efficiency of selection, leading to accelerated development of new yielding planting materials. While in breeding, the use of DNA markers are based on the knowledge of the relation between genotypic and phenotypic variation. Compare to other vegetable, oil palm is the richest source of tocotrienols in Vitamin E and it can enhance the palm oil commercial value in the world market. Therefore, the improvement of Vitamin E of Malaysian commercial oil palm needs to be done through scientific research. The objective of this study is to determine the gene structure of oil palm HGGT gene. The gene structures used from previous study were used as guideline to obtain the best gene structure. Hence, the information generated can assist in the development of molecular markers for Vitamin E improvement.

REFERENCES

- Alberts B, Johnson A, Lewis J, Raff M, Roberts K, Walter P (2002). *Molecular Biology of the Cell* (Fourth ed.). New York: Garland Science. ISBN 978-0-8153-3218-3.
- Anonymous, (2008, May 5). *Sequencing error*. Retrieved from <https://www.broadinstitute.org>.
- Cahoon, E.B., Hall, S.E., Ripp, K.G., Ganzke, T.S., Hitz, W.D. and Coughlan, S.J. (2003). Metabolic redesign of vitamin E biosynthesis in plants for tocotrienol production and increased antioxidant content, *Nat. Biotechnol.* 21, 1082-1087.
- Cochard, B., Amblard, P. and Durand-Gasselien, T. Oil palm genetic improvement and sustainable development. *Oleagineux Corps Gras Lipides* 12, 141–147 (2005).
- Corley, R. H. V. and Tinker, P. B. In *The Oil Palm* 4th edn, 1–26 (Blackwell Science, 2003).
- Dransfield, J. *Genera Palmarum: The Evolution and Classification of Palms* (Royal Botanic Gardens Kew, 2008).
- Gupta, P. K., Roy, J. K. and Prasad M. 2001. Single nucleotide polymorphisms: A new paradigm for molecular marker technology and DNA polymorphism detection with emphasis on their use in plants. *Current Science* 80:523:535.
- Hardon, J. J. 1974. The oil palm. In: *Handbook of plant introduction in tropical crops*. F.A.O Agricultural Studies 93, Rome.
- Hartley, C. *The Oil Palm* 47–94 (Longman, 1988).
- Ho, C. –L. Analysis and functional annotation of expressed sequence tags (ESTs) from multiple tissues of oil palm (*Elaeis guineensis* Jacq.). *BMC Genomics* 8, 381 (2007).
- Horvath, G., Wessjohann, L., Bigirimana, J., Jansen, M., Guisez, Y., Caubergs, R., Horemans, N. 2006 June. Differential distribution of tocopherols and tocotrienols in photosynthetic and non-photosynthetic tissues. *Phytochemistry*. 67(12):1185-95.

- Hunter, S. C. and Cahoon, E.B. (2007) Enhancing vitamin E in oilseeds: unraveling tocopherol and tocotrienol biosynthesis. 42,97-108.
- Jouannic, S. Analysis of expressed sequence tags from oil palm (*Elaeis guineensis*).FEBS Lett. 579, 2709–2714 (2005).
- Khanna, S. (2005). Stroke. 36(10): 2258-6.
- Kiple, KF., and Ornelas, KC. (2000) The Cambridge World History of Food. New York: Cambridge University Press.
- Low, E. T. L. Oil palm (*Elaeis guineensis* Jacq.) tissue culture ESTs: identifying genes associated with callogenesis and embryogenesis. BMC Plant Biol. 8, 62 (2008).
- Mayes, S., Jack, P. L., Corley, R. H. & Marshall, D. F. Construction of a RFLP genetic linkage map for oil palm (*Elaeis guineensis* Jacq.). Genome 40, 116–122 (1997).
- Purseglove, J. W. In Tropical Crops (Monocotyledons) 416–510 (Longman, 1972).
- Rajanaidu, N. (1994). PORIM Oil Palm Genebank. PORIM, Bangi.
- Schneider, C. 2005. Chemistry and biology of vitamin E. Molecular Nutrition of Food Research 49:7-30.
- Semagn, K., Bjornstad, A., & Ndjiondjop, M. N. (2006). An overview of molecular marker methods for plants. 5(25), 2540-2568.
- Sen, C. K (2000). J.Biol.Chem. 275(17): 13049-55.
- Soll, J. and Schultz, G. (1980) 2-Methyl-6-phytylquinol and 2,3-dimethyl-5-phytylquinol as precursors of tocopherol synthesis in spinach chloroplasts. Phytochemistry. 19, 215-218.
- Soll, J., Kemmerling, M. and Schultz, G. (1980) Tocopherol and plastoquinone synthesis in spinach chloroplasts subfractions. Arch. Biochem. Biophys. 204, 544-550.
- Soll, J. (1987) α -Tocopherol and plastoquinone synthesis in chloroplast membranes. Methods Enzymol. 148, 383-392.
- Srivastava, J. K. (2006). Biochem Biophys Res Commun. 346(2): 447-53

Sundram, K., Sambanthamurthi, R., and Tan, YA. (2003). Palm fruit chemistry and nutrition. *Asia Pac J Clin Nutr.* 12(3):355-62.

Tranbarger, T. J. Regulatory mechanisms underlying oil palm fruit mesocarp maturation, ripening, and functional specialization in lipid and carotenoid metabolism. *Plant Physiol.* 156,564–584 (2011).

Wendel, J. F., Cronn, R. C., Alvarez, I., Liu, B., Small, R. L., & Senchina, D. S. (2002). Letter to the Editor Intron Size and Genome Size in Plants. 2346-2352.

Yang, W., Cahoon, R. E., Hunter, S. C., Zhang, C., Han, J., Borgschulte, T., & Cahoon, E. B. (2011). Vitamin E biosynthesis: functional characterization of the monocot homogentisate geranylgeranyl transferase. *The Plant Journal: for Cell and Molecular Biology.* 65(2), 206-17.

Zeven, A. C. The origin of the oil palm. *J. Niger. Inst. Oil Palm Res.* 4, 218–225 (1965).