

UNIVERSITI PUTRA MALAYSIA

EFFECTS OF PRE- GELATINIZED STARCH INCLUSION IN PELLET ON GROWTH AND PHYSIOLOGICAL PARAMETERS OF TWO COMMERCIAL AQUACULTURE SPECIES

NAGAKANMANI A/P MUNIANDY

FP 2018 114



EFFECTS OF PRE- GELATINIZED STARCH INCLUSION IN PELLET ON GROWTH AND PHYSIOLOGICAL PARAMETERS OF TWO COMMERCIAL AQUACULTURE SPECIES

By

NAGAKANMANI A/P MUNIANDY

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

COPYRIGHT

All material contain within the thesis, including without limitation text, logos, icon, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copy right holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EFFECTS OF PRE- GELATINIZED STARCH INCLUSION IN PELLET ON GROWTH AND PHYSIOLOGICAL PARAMETERS OF TWO COMMERCIAL AQUACULTURE SPECIES

By

NAGAKANMANI D/O MUNIANDY

February 2018

Chairman: S. M. Nurul Amin, PhD

Faculty : Agriculture

A 9-week study was conducted to compare the use of dietary corn starch (CS) or tapioca starch (TS), with or without being pre-gelatinized (PG), on the growth, feeding efficiencies, plasma biochemistry, whole-body proximate composition, muscle cholesterol, muscle fatty acid composition, intestinal short chain fatty acids (SCFA), and liver glycogen and histopathology of red hybrid tilapia (*Orecohromis* sp.). Triplicate groups of 20 fish (initial mean weight = 0.739 ± 0.01 g) were fed their respective diets to satiation. Various pellet characteristics were also measured that included bulk density (BD), expansion ratio (ER), pellet durability index (PDI), water solubility index (WSI), water absorption index (WAI), and water stability (WS) while the surface microstructure was examined.

Results showed that tilapia fed the TS diet had significantly lower growth (p < 0.05) than all other treatments, but was significantly improved when pre-gelatinized. In the PG dietary treatments, intestinal SCFA significantly decreased while plasma glucose, cholesterol and triglycerides as well as liver glycogen were significantly higher compared to the native starch diets. Whole-body proximate composition and muscle cholesterol were unaffected by dietary treatments, although fish in the CS treatment had significantly higher amounts of long chain polyunsaturated fatty acids compared to the other treatments. The PG diets had significantly higher PDI, WS, WSI and BD compared to native starch diets. The surface morphology generally showed a smoother surface for the PG diets.

While dietary TS was inferior compared to CS for tilapia, the PG-TS diet significantly improved their growth and feeding efficiencies and moreover the PG diets led better pellet characteristics that are likely to have important implications to the production of aquafeeds. Therefore, for the second experiment the tapioca starch was chosen and

tested with African catfish. Tilapia and African catfish are the common commercial species that been farmed world wide.

A 7-week study was conducted to compare the use of dietary tapioca starch and pre gelatinized tapioca starch with or without IMO on the growth, feeding efficiencies and muscle proximate composition of African catfish, *Clarias Gariepinus*. Triplicate groups of 15 fish (initial mean weight = 6.2 ± 0.3 g) were fed their respective diets to satiation. Various pellet characteristics were also measured that included bulk density (BD), pellet durability index, water solubility index (WSI), water absorption index (WAI), and water stability (WS).

Results showed that there are no significance differences between treatments in growth of catfish (p > 0.05). The native starches with and without iso- maltose shows significantly lower water stability while the pre- gelatinized starch with and without iso- maltose showing high water stability. The water absorption index also shows significantly different between the native starches and pre- gelatinized starch. In overall, pre- gelatinized starches reduced the feed intake without compromising growth. As well, there were some negative effects to the nutritive value of the fish but it was mitigated by the addition of the prebiotic. Therefore, the pre- gelatinized tapioca starch can reduce the cost of production and as well help to maintain the water quality.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Sarjana Sains

KESAN PENGGUNAAN KANJI PRA- GEL DALAM PELLET TERHADAP TUMBESARAN DAN CIRI- CIRI FIZIKAL DUA SPESIES KOMERSIAL AKUAKULTUR

Oleh

NAGAKANMANI A/P MUNIANDY

Februari 2018

Pengerusi: S. M. Nurul Amin, PhD

Fakulti : Pertanian

Satu kajian selama 9 minggu dijalankan untuk membandingkan penggunaan kanji jagung (CS) dan kanji ubi kayu yang sudah digel atau yang belum digel (PG), terhadap pertumbuhan, kecekapan pemakanan, biokimia plasma, komposisi proximat seluruh badan, kolesterol otot, komposisi asid lemak, asid lemak rantaian pendek usus (SCFA) dan glikogen hati serta histopatologi tilapia hybrid merah (*Oreochromis sp.*).

Sebuah akuarium diisikan dengan 20 ekor ikan (berat minimum awal = 0.739 ± 0.01 g) dan diberikan makanan rawatan yang disediakan sehingga kenyang serta setiap rawatan digandakan kepada tiga kali ganda. Pelbagai ciri-ciri pellet diperiksa iaitu ketumpatan pukal (BD), kadar pengembangan (ER), daya ketahanan pellet (PDI), indeks kelarutan air (WSI), indeks penyerapan air (WAI), dan kestabilan air (WS) serta mikrostruktur permukaan.

Hasil kajian menunjukkan tilapia yang diberi makan makanan TS mempunyai pertumbuhan yang kurang dengan ketara (p<0.05) daripada makanan rawatan yang lain, tetapi pertumbuhannya bertambah baik apabila digel. Dalam rawatan pemakanan PG, SCFA usus menurun secara ketara sementara glukosa, kolesterol dan trigliserida plasma serta glikogen hati lebih tinggi berbanding dengan diet kanji asli atau yang tidak digel. Komposisi proximat dan kolesterol otot tidak ada perbezaan antara rawatan pemakanan, walaupun ikan dalam rawatan CS mempunyai rantaian panjang asid lemak tak tepu yang jauh lebih tinggi dibandingkan dengan rawatan lain. Diet PG mempunyai PDI, WS, WSI dan BD yang tinggi berbanding dengan diet kanji yang asli. Walaupun diet TS menunjukan pertumbuhan yang kurang baik berbanding dengan diet CS, tetapi diet PGTS menunjukkan pertumbuhan yang baik dan kecekapan pemakanan serta ciriciri pellet yang lebih baik dimana ia boleh mempunyai implikasi yang penting terhadap pengeluaran makanan ikan. Oleh sebab itu, untuk eksperimen yang kedua, kanji ubi

kayu dipilih untuk diuji dengan ikan keli. Ikan tilapia dan keli dipilih kerana kedua spesies ini merupakan spesies komersial yang diternak secara luas di seluruh dunia.

Satu lagi kajian dijalankan selama 7 minggu untuk membandingkan penggunaan kanji ubi kayu asli dan yang digel dengan atau tanpa IMO pada pertumbuhan, kecekapan pemakanan dan komposisi proximat otot ikan keli Afrika, *Clarias gariepinus*. Sebanyak 15 ekor ikan dimasukkan ke dalam satu akuarium (berat minimum awal = 6.2 ± 0.3 g) dan diberi makanan rawatan sehingga kenyang serta setiap rawatan digandakan kepada tiga kali ganda. Pelbagai ciri- ciri pellet juga diuji termasuk ketumpatan pukal (BD), kadar pengembangan (ER), daya ketahanan pellet (PDI), indeks kelarutan air (WSI), indeks penyerapan air (WAI), dan kestabilan air (WS).

Hasil kajian menunjukkan bahawa tiada perbezaan yang dikenal pasti dari segi pertumbuhan antara rawatan. Kanji asli dengan dan tanpa iso- maltose menunjukkan kestabilan air yang rendah manakala kanji yang digel dengan dan tanpa iso maltose menunjukana kestabilan air yang tinggi. Indeks penyerapan air (WAI) juga menunjukkan perbezaan yang ketara antara rawatan. Secara keseluruhan, kanji yang digel mengurangkan pengambilan makanan tetapi tidak menjejaskan pertumbuhan ikan. Selain itu, terdapat beberapa kesan negatif terhadap nilai pemakanan ikan tetapi ia dapat diatasi dengan penambahan prebiotic. Oleh itu, kanji ubi kayu yang digel boleh mengurangkan kos pengeluaran dan membantu untuk mengekalkan kualiti air.

ACKNOWLEDGEMENTS

First and foremost, I would like to express my heartfelt gratitude to His Almighty for I have finally completed my thesis project for master degree successfully.

This thesis would not have been possible without the help, support and patience of my supervisor, Dr. S. M. Nurul Amin. His good advice, support and guidance have been invaluable on both an academic and a personal level, for which I am extremely grateful I am very much indebted to him for his unwavering support, guidance and valuable advice in completing this thesis. Without his counsel, this thesis would not be able to achieve its objective

I would like to thank my thesis committee member Dr, Nicholas Romano for his advices without his quality and friendly supervisions, this work would not have come to completion. A special thanks to Prof. Dr. Mohd Salleh Kamarudin, En. Salleh, En. Zawawi, En. Azman, Pn. Syafika, Cik Hafizah and all staff of Aquaculture Department for their assistance and cooperation which led to the smooth running of this experiment. A sincere thank to all my friends for their kindness and moral support during my study.

Last but not least, my beloved families who have given me their unequivocal support throughout and indirect support, as always, for which my mere expression of thanks likewise does not suffice. Thank you very much.

This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the degree of Master of Science. The members of Supervisory Committee were as follows:

S. M. Nurul Amin, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Nicholas Romano, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 16 August 2018

Declaration by Graduate Student

I hereby declare that:

- the thesis is my original work;
- quotations, illustrations and citation have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writing, seminar papers, manuscripts, poster, reports, lecture notes, learning modules or any other material as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/ fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. This thesis has undergone plagiarism detection software.

Signature:	Date:

Name and Matric No.: Nagakanmani A/P Muniandy, GS44426

TABLE OF CONTENTS

					Page
ABSTRACT					i
ABSTRAK					iii
ACKNOWL	EDGEMEN	NTS			v
APPROVAL	ı				vi
DECLARAT	TON				viii
LIST OF TA	BLES				XV
LIST OF FIG	GURES				xvi
LIST OF AB	BREVIAT	IONS			xvii
CHAPTER					
1	INTD	ODUCT	ION		1
1	1.1		ound of stu	ndu -	1
	1.1	_	ant of stud		1
	1.3	_	ant of stuc		2
	1.3	hypothe		talid	2
	1.4		h objectiv	es	2
2	L <mark>ITE</mark>	RATURI	E REVIE	W	3
	2.1	Aquacu	lture		3
	2.2	Carbohy	ydrates		3
		2.2.1		drate usage in	5
		222	aquacult		6
		2.2.2	15	f carbohydrates Saccharides	6
			2.2.2.1		6
		2.2.2	2.2.2.2	Polysaccharides	7
	2.2	2.2.3 Starch	Carbony	drates potential	9
	2.3		C4 1	14	10
		2.3.1		usage in aquaculture	11
		2.3.2	• 1	f starches	13
				Native starches	13
			2.3.2.2	Pre- gelatinized starch	14
	2.4	Fish		Starter	15
		2.4.1	Red hyb	rid tilapia,	16
		2.4.5	Oreochr	omis spp	
		2.4.2	African gariepin	catfish, <i>Clarias</i> us	16

3	GEN	ERAL M	ETHODO	DLOGY	18
	3.1	Introdu	ction		18
	3.2	Method	ls		18
		3.2.1	Location	of study	18
		3.2.2	Water qu	ality parameters	18
		3.2.3	Data coll	lection	19
		3.2.4	Chemica	l analysis	19
			3.2.4.1	Determination of moisture	19
			3.2.4.2	Determination of crude protein	19
			3.2.4.3	Determination of	19
			3.2.4.4	lipid Determination of crude fibre	20
			3.2.4.5	Determination of	20
	2.2	a		ash	20
	3.3	Statistic	cal analysis		20
4	IMDI	OVEM	CNIT	TO PELLET	21
4		ROVEMI RACTEI		TO PELLET AND SUBSEQUENT	21
	EFFE	ECTS	ON	GROWTH AND	
	PHY	SIOLOG	Y OF TI	LAPIA (Oreochromis	
		BOHYDI		PRE-GELATINIZED	
	4.1	Introdu			21
	4.2	Materia	l methods		22
		4.2.1	Formulat	tion of diets	22
			4.2.1.1	Proximate analysis	23
			4.2.1.2	Carbohydrate	23
			1212	analysis	2.4
		4 2 2	4.2.1.3	Fatty acid analysis aracteristics	24 25
		4.2.2			
			4.2.2.1	Bulk density (BD)	25
			4.2.2.2	Expansion ratio (ER)	25
			4.2.2.3	Pellet durability index (PDI	25
			4.2.2.4	Water absorption index (WAI) and water solubility index (WSI)	25
			4.2.2.5	Water stability	26
			4.2.2.6	Microstructure	26
			4227	Protein solubility	26

	4.2.3	Tilapia, Oreochromis spp		27
		4.2.3.1	Animal source and experimental design	27
		4.2.3.2	Plasma biochemistry	27
		4.2.3.3	Muscle proximate	28
		4.2.3.4	Intestinal short chain fatty acids (SCFA)	28
		4.2.3.5	Liver histopathology	28
		4.2.3.6	Cholesterol	28
		4.2.3.7	Fatty acid	29
4.3	Results	3		30
	4.3.1	Experime	ental diets	30
		4.3.1.1	Formulation of diets, proximate and carbohydrate	30
			analysis	
		4.3.1.2	Fatty acid analysis	31
	4.3.2	Pellet cha	racteristics	32
	4.3.3	Tilapia, C	Oreochromis sp	36
		4.3.3.1	Growth, survival, and feeding efficiencies	36
		4.3.3.2	Whole body proximate, muscle	37
			cholesterol and body indices	
		4.3.3.3	Plasma biochemistry	37
		4.3.3.4	Muscle fatty acid composition	38
		4.3.3.5	Liver histopathology	40
		4.3.3.6	Intestinal short chain fatty acid (SCFA)	42
4.4	Discus	sion		44
15	Const			16

TAPIO ISOM THE SUBS FEED NUTE	OCA [ALTOO] PELLET EQUENT PING RITIVE	STA LIGOSAC T CHARA T EFFE	ENCIES AND OF AFRICAN	47
5.1	Introduc	tion		47
5.2	Material	and metho	ds	49
	5.2.1	Formulation	on of diets	49
		5.2.1.1	Proximate analysis	50
	5.2.2	Pellet char	racteristics	50
	5.2.3	gariepinu		51
		5.2.3.1	Animal source and experimental design	51
		5.2.3.2	Plasma biochemistry	51
		5.2.3.3	Muscle proximate	52
		5.2.3.4	Intestinal short chain fatty acids (SCFA)	52
		5.2.3.5	Liver histopathology	52
		5.2.3.6	Cholesterol	52
		5.2.3.7	Liver digestive enzyme	53
5.3	Results			53
	5.3.1	Experimen		53
		5.3.1.1	Formulation of diets and proximate analysis	53
	5.3.2	Pellet char	3	54
	5.3.3	African ca	atfish, <i>Clarias</i>	58
		5.3.3.1	Growth, survival, feeding efficiencies and hepatosomatic index	58
		5.3.3.2	Muscle body proximate	58
		5.3.3.3	Plasma biochemistry	59
		5.3.3.4	Intestinal short chain fatty acid	60
		5.3.3.5	Liver histopathology	61
		5.3.3.6	Liver digestive enzyme	62

	5.4	Discussion	64
	5.5	Conclusion	69
6		ERAL DISCUSSION, CONCLUSIONS RECOMMENDATIONS	70
REFERENCES	3		72
BIODATA OF STUDENT			
LIST OF PUBI	LICATION	ON	82

LIST OF TABLES

Table		Page
4.1	Feed formulations in percentage.	23
4.2	Ingredient formulation (g) and proximate composition (% dry weight) of the experimental diets.	30
4.3	Fatty acid composition (%) of the experimental diets	31
4.4	Physical properties of the extruded pellets with different pregelatinized or native carbohydrates sources.	32
4.5	Growth performance and feeding efficiencies of tilapia fed diets with different pre-gelatinized or native carbohydrate sources.	36
4.6	Whole body proximate composition (% wet weight), muscle cholesterol (µg ml-1) and body indices of tilapia fed diets with different pre-gelatinized or native carbohydrate sources.	37
4.7	Plasma biochemistry (mmol 1-1) of the muscle from tilapia fed diets with different pre-gelatinized or native carbohydrate sources.	38
4.8	Fatty acid composition (%) of the muscle from tilapia fed diets with different pre-gelatinized or native carbohydrate sources.	39
5.1	Feed formulations in percentage	50
5.2	Ingredient formulation (g) and proximate composition (% dry weight) of the experimental diets.	53
5.3	Physical properties of the pelletized pellets whether native or pregelatinized tapioca starch with or without Iso-maltose.	55
5.4	Mean (±SE) growth performance, feeding efficiencies and hepatosomatic index (HSI) of African catfish (<i>Clarias gariepinus</i>) juveniles fed diets with native and pre- gelatinized tapioca starch with or without Iso- maltose.	58
5.5	Mean (±SE) proximate composition (% wet weight) of African catfish (<i>Clarias gariepinus</i>) juveniles fed diets with native and pre- gelatinized tapioca starch with or without Iso- maltose.	59
5.6	Mean (±SE) plasma biochemistry parameters in African catfish (<i>Clarias gariepinus</i>) juveniles fed diets with native and pregelatinized tapioca starch with or without Iso- maltose.	59

LIST OF FIGURES

Figur	re	Pag
4.1	Surface structures of a) native corn starch, b) pre- gelatinized corn starch, c) native tapioca starch and d) pre- gelatinized tapioca starch at magnification $\times 30$.	33
4.2	Cross section structures of a) native corn starch, b) pre-gelatinized corn starch, c) native tapioca starch and d) pre- gelatinized tapioca starch at magnification ×40.	34
4.3	Surface structures of a) native corn starch, b) pre- gelatinized corn starch, c) native tapioca starch and d) pre- gelatinized tapioca starch at magnification ×250.	35
4.4	Tilapia fed with the a) native corn starch, b) pre- gelatinized corn starch, c) native tapioca starch and d) pre- gelatinized tapioca starch after the Periodic- acid Schiff staining at magnification × 20.	41
4.5	PAS staining intensity.	42
4.6	Intestinal short chain fatty acids (SCFA)	43
5.1	Surface structures of a) native tapioca starch, b) native tapioca starch with Iso- maltose, c) pre- gelatinized tapioca starch and d) pre- gelatinized tapioca starch with Iso- maltose at magnification ×30.	56
5.2	Surface structures of a) native tapioca starch, b) native tapioca starch with Iso- maltose, c) pre- gelatinized tapioca starch and d) pre- gelatinized tapioca starch with Iso- maltose at magnification ×250.	57
5.3	Intestinal short chain fatty acids (SCFA).	60
5.4	African catfish fed with the a) native tapioca starch, b) Native tapioca starch with Iso- maltose, c) pre- gelatinized tapioca starch and d) pre-gelatinized tapioca starch with Iso- maltose after the Periodic- acid Schiff staining at magnification × 20.	61
5.5	Periodic- acid Shciff staining intensity.	62
5.6	Liver enzyme activity.	63

LIST OF ABBREVIATIONS

ANOVA Analysis of variance FCR Food conversion ratio PG Pre- gelatinized

 $\begin{array}{ccc} g & & Gram \\ h & & Hour \\ H_2SO_4 & & Sulfuric acid \end{array}$

ind

kg L

Min

SGR

IUCN International Union for Conservation of

Nature Individual Kilogram Liter Minute

Mg/L Milligram per liter

ml Milliliter

NaOH Sodium hydroxide

pcs Pieces

PER Protein efficiency ratio

Part per million ppm PDI Pellet durability index WS Water stability BD Bulk density PS Protein solubility Water absorption index WAI WSI Water solubility index SE Standard error

Sp. Species

UPM Universiti Putra Malaysia
USA United States of America

Specific growth rate

⁰C Degree Celsius

% Percentage
< Less than
> More than

CHAPTER 1

INTRODUCTION

1.1 Background of study

The main sources of energy in most animal diets are carbohydrates. The carbohydrates can be classified based on the structure, composition, constituent sugars and degree of polymerisation. It also can be classified based on the glycosidic linkage such as non-monomer carbohydrates like oligosaccharides which is lactose and maltose, polysaccharides which is starch, chitin and cellulose, and the other one is monomer sugars like glucose and fructose (Englyst, 1996). Carbohydrates properties are the vital importance for their nutritional effects. For example, the absorption rate and digestion, the viscosity, structural features fermentation ability in the gastrointestinal tract and water binding capacity (Asp. 1996).

The energy storage nutrient is starch. For example in wheat, approximately 60% of the total grain is a starch which is used as energy storage nutrient and constitutes (Novus, 1992), where the glucose molecules linked together by α -glycosidic bonds is composed and the enzymatic activities in fish are influenced. Several fish species appeared to produce positive effects on digestibility and growth by the inclusion of dietary carbohydrate (Li *et al.*, 2013). There might have negative effects on growth, health, metabolism and nutrient utilization with the inappropriate amount of carbohydrates in aquafeed (Li *et al.*, 2012).

There are several factors like types, environmental conditions and source of carbohydrate that varies with the digestion and metabolism of carbohydrates digestion and in addition of fish species as well (Hutchins, 1998). Generally fish species that utilized carbohydrates more efficiently is warm water fish. They can utilize at higher levels than cold water fish and marine fish species (Wilson, 1994). As well high dietary levels of carbohydrates are efficiently utilized by the omnivorous fish species compared to carnivorous fish species (Enes *et al.*, 2011). So far, the dietary carbohydrate level has been not particularly defined.

1.2 Significant of study

The most abundant and inexpensive sources in formulated feed are carbohydrates. It also can be easily available. Several fish species can utilize it efficiently as well (Zhao, 2011). Other than that, Azaza, 2013 found that there is protein sparing effects in tilapia and salmonid fish species when fed with carbohydrates. The most expensive ingredients are protein in the formulated aquafeed, and thus increasing the cost of the feed production. Starch is the well utilized carbohydrates source in aquafeed which can reduced cost of production and as well been used as a binder in feed. Reduced in cost of production is not only because the starch is inexpensive and it spare protein but it also influenced the pellet characteristic. In where, gelatinized starch improved the pellet characteristics.

1.3 Problem statement and hypothesis

Starch is a carbohydrate made up of glucose units linked together by glicosidic bond and the most used carbohydrates sources in aquafeed. Utilization of carbohydrates is much more variable. It is probably related to natural feeding habits of the fish. Pelleting quality of the diet and the fish growth might have the benefit by the incorporation of the carbohydrate (Wilson, 1994). Usually native starch will be used in producing the aquafeed. Pre- gelatinized starches are rarely used and the characteristic of pre- gelatinized starches are unknown. Pre- gelatinized starches are more potential in growth and characteristic of the pellets compared to the native starches.

1.4 Research objective

The general purpose of this study was to examine the potential differences between native starches and pre-gelatinizes starches. The specific objectives were:.

- 1. To assess the potential differences of native and pre- gelatinized starches (corn starch and tapioca starch) on the pellet characteristic when extruded.
- 2. To assess the potential differences of native and pre-gelatinized starches (corn starch and tapioca starch) when extruded on the growth and various physiological parameters of tilapia.
- 3. To assess the potential differences of native and pre-gelatinized tapioca starch with and without prebiotic on the pellet characteristic when cold pelleted.
- 4. To assess the potential differences of native and pre- gelatinized tapioca starch with or without prebiotic on the growth and various physiological parameters of African catfish when cold pelleted.

REFERENCES

- Aas, T. S., Terjesen, B. F., Sigholt, T., Hillestad, M., Holm, J., Refstie, S., Baeverfjord, G. (2011). Nutritional responses in rainbow trout (*Oncorhynchus mykiss*) fed diets with different physical qualities at stable or variable environmental condition. *Aquaculture nutrition*, 17, 657-670.
- Abd El-Khalek, E., Kalmar, I., Van Weyenberg, S., Werquin, G., & Janssens, G. (2009). Effect of starch gelatinisation on nutrient digestibility and plasma metabolites in pigeons. *Journal of animal physiology and animal nutrition*, 93(3)., 359-365.
- Alvarez, M. J., Lopez-Bote, C. J., Diez, A., Corraze, G., Arze, J., Dias, J., Kaushik, S. & Bautista, S. J. (1999). The partial substitution of digestible protein with gelatinized starch as an energy source reduces susceptibility to lipid oxidation in rainbow trout (*Oncorhynchus mykiss*) and sea bass (*Dicentrarchus labrax*) muscle. *J. Ani. Sci.* 77, 3322-3329.
- Amirkolaie, A. K., Verreth, J. A. J., Schrama, J. W. (2006). Effect of gelatinization degree and inclusion level of dietary starch on the characteristics of digesta and faeces in Nile tilapia (*Oreochromis niloticus* (*L*.)). *Aquaculture* 260, 194-205.
- Anderson, J., Jackson, A. J., Matty, A. J., Capper, B. S. (1984). Effects of dietary carbohydrate and fibre on the tilapia *Oreochromis niloticus (Linn.)*. *Aquaculture* 37, 303-314.
- Anderson, R.A. (1982). Water absorption and solubility and amylograph characteristics of roll-cooked small grain products. *Cereal Chem.* 59, 265-269.
- AOAC. (1997). Official Methods of Analysis of AOAC International, 16th ed. AOAC International. Arlington, Virginia, USA.: In: Cunniff, P.A. (Ed.).
- Araba, M. & Dale, N. (1990). Evaluation of protein solubility as an indicator of over processing soybean meal. *Poultry science*, 69, 76-83.
- Arnesen, P., Krogdahl, A. & Sundby, A. (1995). Nutrient digestibilities, weight gain and plasma and liver levels of carbohydrate in Atlantic salmon (*Salmo salar*, *L*) fed diets containing oats and maize. *Aquaculture Nutrition.*, 1, 151-158.
- Asp, N. G. (1996). Dietary carbohydrates: Classification by chemistry and physiology. *Food Chemistry 57(1)*, 9-14.
- Azaza, M. S., Khiari, N., Dhraief, M. N., Aloui, N., Kraïem, M. M. & Elfeki, A. (2013). Growth performance, oxidative stress indices and hepatic carbohydrate metabolic enzymes activities of juvenile Nile tilapia (*Oreochromis niloticus*), in response to dietary starch to protein ratios. *Aquaculture Research*, 1-14.
- Bergot, F. & Breque, J. (1983). Digestibility of starch by rainbow trout: effects of the physical state of starch and the intake level. *Aquaculture*, *34*. 203–212., 203–212.
- Bergot, F.& Breque, J. (1979). Carbohydrate in rainbow trout diets: effects of the level and source of carbohydrate and the number of meals on the growth and body composition. *Aquaculture 18*, 1557-167.
- Bertolini Andrea C. (2009). *Starches characterization, properties and application*. London: CRC Press.
- Booth, M.A., Allan, G.L., Frances, J. & Parkinson, S. (2001). Replacement of fish meal in diets for Australian silver perch, *Bidyanus bidyanus IV*. Effects of dehulling and protein concentration on digestibility of grain legumes. *Aquaculture*, 196, 67-85.

- Boyd, C. E. & Tucker, C. S. (1998). Chapter 14: Waste Management. In *Pond aquaculture water quality management*. (pp. 541- 573). Massachusetts, USA.: Kluwer Academic Publishers.
- Buddington, R. K. & Hilton, J. W. (1987). Intestinal adaptations of rainbow trout to changes in dietary carbohydrate. *Am. J. Physiol.*, 253, G489-G496.
- Buddington, R.K., Krogdahl, A & Bakke-McKellep, A.M. (1997). The intestine of carnivorous fish: structure and functions and the relations with diet. *Acta Physiol. Scand.*, 161, 67-80.
- Burel, C., Boujard, T., Tulli, F. & Kaushik, S. J. (2000). Digestibility of extruded peas, extruded lupin, and rapeseed meal in rainbow trout (*Oncorhynchus mykiss*) and turbot (*Psetta maxima*). *Aquaculture* 188, 285-298.
- Case, S. E., Hannann, D. D., Schwartz, S. J. (1992). Effect of starch gelatinzation on physical properties of wheat and corn-based products. *Cereal chemistry*, 69, 401-404.
- Chang, J.C.-H., Wu, S.-M., Tseng, Y.-C., Lee, Y.-C., Baba, O., Hwang, P.-P. (2007). Regulation of glycogen metabolism in gills and liver of the euryhaline tilapia (*Oreochromis mossambicus*) during acclimation to seawater. *J. Environ Biol.* 210, 3494-3504.
- Chen, W. W., Romano, N., Ebrahimi, M. & Natrah, I. (2017). The effects of dietary fructooligosaccharide on growth, intestinal short chain fatty acids level and hepatopancreatic condition of the giant freshwater prawn (*Macrobrachium rosenbergii*) post-larvae. *Aquaculture*, 456, 95-101.
- Cheng, Z.J. & Hardy, R.W. (2003). Effects of extrusion processing of feed ingredients on apparent digestibility coefficients of nutrients for rainbow trout (*Oncorhynchus mykiss*). Aquacult. Nutr., 9, 77-87.
- Colonna, P., Doublier, J.L., Melcion, J.P., de Monredon, F., Mercier, F. (1984). Extrusion cooking and drum drying of wheat starch. I. Physical and macromolecular modifications. *Cereal Chem.* 61, 538-543.
- Conway, H. F. & Anderson, P.A. (1973). Protein-fortified extruded food products. Cereal Sci. Today 18, 94.
- Da Silva, R. F., & Vázquez, F. J. S. (2014). Biology of European Sea Bass. In *Nutrition and Dietary Selection*. (p. 252).
- Dabrowski, K. & Guderley, H. (2002). Intermediary metabolism.In Halver, J.E., (Eds.), London: Academic Press.
- de Cruz, C.R., Kamarudin, M.S., Saad, C.R., Ramezani-Fard, E. (2015). Effects of extruder die temperature on the physical properties of extruded fish pellets containing taro and broken rice starch. *Animal Feed Sci. Tech.* 199, 137-145.
- Degani G, Ben Zvi Y & Levanon D. (1989). The effect of different protein levels and temperature on feed utilization, growth and bosy composition of *Clarias gariepinus* (Burchell 1822). *Aquaculture* 76, 293-301.
- Deng, D.-F., Refstie, S. & Hung, S.S.O. (2001). Glycemia and Glycosuric reponses in White sturgeon (*Acipenser transmontanus*) after oral administration of simple and complex carbohydrates. *Aquaculture 199*, 107-117.
- Douzals, J.P., Marechal, P. A., Coquille, J.C., and Gervais, P. (1996). Microscopic study of starch gelatinization under high hydrostatic pressure. *Journal of Agriculture and food chemistry*, 44, 1403-1408.
- Dukic´, A., Mens, R., Adriaensens, P., Foreman, P., Gelah, J., Remon, J. P., & Vervaet,
 C. (2007). Development of starch-based pellets via extrustion/spheronisation.
 European Journal of Pharmaceutics and Biopharmaceutics, 66, 83-94.

- Ebrahimi, M., Raijon, M. A., Meng, G. Y., Farjam, A. S. (2014). Omega-3 fatty acid enriched chevon (goat meat) lowers plasma cholesterol levels and alters gene expressions in rats. *BioMed Res. Intern.* 2014, 1-8.
- Edwin H. Robinson, Menghe H. Li, Bruce B. Manning. (2001). A practical guide to nutrition, feeds and feeding of catfish. *Bulletin 1113*, 10.
- Enes, P., Panserat, S., Kaushik, S. & Oliva-Teles, A. (2008). Growth performance and metabolic utilization of diets with native and waxy maize starch by gilthead seabream (*Sparus auratus*) juveniles. *Aquaculture*, 274., 101-108.
- Enes, P., Panserat, S., Kaushik, S. & Oliva-Teles, A. (2011). Dietary carbohydrate utilization by European sea bass (*Dicentrarchus labrax*) and gilthead sea bream (*Sparus aurata*) Juveniles. *Reviews in Fisheries Science* 19(3), 201-215.
- Englyst, H.N. & Hudson, G.J. (1996). The classification and measurement of dietary carbohydrates. *Food Chemistry* 57(1), 15-21.
- Erfanullah & Jafri, A.K. (1998). Growth, feed conversion, body composition and nutrient retention efficiencies in fingerling catfish, *Heteropneustes fossilis* (Bloch), fed different sources of dietary carbohydrate. *Aquaculture Res.*, 30, 43-49.
- Fagbenro, O., & Jauncey, K. (1995). Water stability, nutrient leaching and nutritional properties of moist fermented fish silage diets. *Aquacultural Engineering*, 14, 143-153.
- FAO. (2014). World aquaculture production of fish, crustacean, molluscs, etc by principal species in 2013.
- FAO. (2016). Aquaculture summary, the state of world fisheries and aquaculture report. Food and agriculture organization of the United Nations.
- Fernandez, F., Miquel, A.G., Guinea, J. & Martinez, R. (1998). Digestion and digestibility in gilthead sea bream (*Sparus aurata*): the effect of diet composition and ration size. *Aquaculture* 166, 67-84.
- Furuichi, M. and Yone, Y. (1980). Effect of dietary dextrin levels on the growth and feed efficiency, the chemical composition of liver and dorsal muscle and the absorption of dietary protein and dextrin in fishes. *Bull. Jpn. Sot. Sci. Fish*, 46., 225-229.
- Furuichi, M. and Yone, Y. (1982b). Effect of insulin on blood sugar levels of fishes. *Bull. Jpn. Sot. Sci. Fish.*, 48., 1289-1291.
- Furuichi, M. and Yone, Y.,. (1982a). Availability of carbohydrate in nutrition of carp and red sea bream. *Bull. Jpn. Sot. Sci. Fish.*, 48, 945-948.
- Gao, W., Liu, Y.J., Tian, L.X., Mai, K.S., Liang, G.Y., Yang, H.J., Huai, M.Y. & Luo, W.J. (2010). Effect of dietary carbohydrate-to-lipid ratios on growth performance, body composition, nutrient utilization and hepatic enzymes activities of herbivorous grass carp (*Ctenopharyngodon idella*). *Aquaculture Nutrition* 16(3), 327-333.
- Garci'a-Gallego, M.; Bazoco, J.; Akharbach, H.; Sua' rez, M.D.; Sanz, A. (1994). Utilization of different carbohydrates by the European eel (*Anguilla anguilla*). *Aquaculture 124*, 99-108.
- Geraylou, Z., Souffreau, C., Rurangwa, E., Maes, G.E., Spanier, K.I., Courtin, C.M., Delcour, J.A., Buyse, J., Ollevier, F. (2013). Prebiotic effects of arabinoxylan oligosaccharides on juvenile Siberian sturgeon (*Acipenser baerii*) with emphasis on the modulation of the gut microbiota using 454 pyrosequencing. *FEMS Microbiol. Ecol.* 86, 357-371.
- Gerhardt, P., Murray, R.G.E., Wood, W.A., Krieg, N.R. (1994). Methods for General and Molecular Bacteriology. *American Society for Microbiology, Washington, DC.*, 518.

- Glencross, B., Blyth, D., Tabrett, S., Bourne, N., Irvin, S., Anderson, M., Fox-Smith, T., Smullen, R. (2012). An assessment of cereal grains and other starch sources in diets for barramundi (*Lates calcarifer*) implications for nutritional and functional qualities of extruded feeds. *Aquacult. Nutri.* 18, 388-399.
- Glencross, B.D. (2006). Nutritional management of barramundi *Lates calcarifer* a review. *Aquacult. Nutr.*, 12, 291-309.
- Glencross, B.D., Booth, M. & Allan, G.L. (2007). A feed is only as good as its ingredients a review of ingredient evaluation for aquaculture feeds. *Aquacult. Nutr.*, 13, 17-34.
- Gonzalez, C., Allan, G. (2007). Preparing farm-made fish feed. *New South Wales Department of Primary Industries.*, 1-21.
- Grisdale-Helland, B. & Helland, S.J. (1997). Replacement of protein by fat and carbohydrate in diets for Atlantic salmon (*Salmo salar*) at the end of the freshwater stage. *Aquaculture*, 152, 167-180.
- Grisdale-Helland, B. & Helland, S.J. (1998). Macronutrient utilization by Atlantic halibut (*Hippoglossus hippoglossus*): diet digestibility and growth of 1 kg fish. *Aquaculture*, 166, 57-65.
- Guha, M., Zakiuddin, A.S., Bhattacharya, S. (1997). Twin-screw extrusion of rice flour without a die: Effect of barrel temperature and screw speed on extrusion and extrudate characteristics. *J. Food Eng.* 32, 251-267.
- Hemre, G.-I., Lie, O. & Lambertsen, G. (1990). Digestibility of different carbohydrates sources in cod (*Gadus morhua*) and its relation to glucose content in blood and urine. *Fisk Dir. Skr. Ser. Ern.*, 3-9.
- Hemre, G.-I., Lie, Ø., Lied, E. & Lambertsen, G. (1989). Starch as an energy source in feed for cod (*Gadus morhua*): digestibility and retention. *Aquaculture*, 80, 261-270.
- Hemre, G.-I., Lie, Ø., Lied, E., Lambertsen, G. (1989). Starch as an energy source in feed for cod (*Gadus morhua*): digestibility and retention. *Aquaculture* 80, 261-270.
- Hemre, G.-I., Mommsen, T.P. & Krogdahl, A. (2002). Carbohydrates in fish nutrition: effects on growth, glucose metabolism and hepatic enzymes. *Aquaculture Nutrition* 8, 175-194.
- Hemre, G.I., Sandnes, K., Torrissen, O. & Waagbo, R. (1995). Carbohydrate nutrition in Atlontic salmon (Salmo solar). Aquaculture and Fish Management(26), 154-156.
- Hilton, J.W., Cho, C.Y., Slinger, S.J. (1981). Effect of extrusion processing and steam pelleting diets on pellet durability, pellet water absorption and the physiological response of rainbow trout (*Salmo gairdneri* R). Aquaculture 25, 185-194.
- Hixson, SM. (2014). Fish nutrition and current issues in aquaculture: the balance in provividing safe and nutritious seafood, in an environmentally sustainable manner. *Journal Aquaculture Research Development* 5, 234.
- Hoseinifar, S.H., Eshaghzadeh, H., Vahabzadeh, H., Peykaran Mana, N. (2015a). Modulation of growth performances, survival, digestive enzyme activities and intestinal microbiota in common carp (*Cyprinus carpio*) larvae using short chain fructooligosaccharide. *Aquac. Res.* 47, 3246-3253.
- Hoseinifar, S.H., Esteban, M.Á., Cuesta, A., Sun, Y.Z. (2015). Prebiotics and fish immune response: A review of current knowledge and future perspectives. *Reviews in Fisheries Science and Aquaculture 23*, 315-328.
- Hua, K. & Bureau, D.P. (2009). A mathematical model to explain variations in estimates of starch digestibility and predict digestible starch content of salmonid fish feeds. *Aquaculture*, 294., 282–287.

- Hutchins, C.G., Rawles, S.D. & Gatlin, D.M. (1998). Effects of dietary carbohydrate kind and level on growth, body composition and glycemic response of juvenile sunshine bass (*Morone chrysops* female x *Morone saxatilis* male). *Aquaculture* 161(1-4), 187-199.
- Isaac Wayne Witus, Leong Wan Vun. (2016). Aquaculture in Malaysia. *A short Review on Current Policy ang Legislation*, 150 -154.
- Jeong, K.S., Takeuchi, T., Okamoto, N. & Watanabe, T. (1992). The effect of dietary gelatinized rations at different dietary energy levels on growth and characteristics of blood in carp fingerlings. *Bull. Jpn Soc. Sci. Fish*, 58., 945-951.
- Kapoor. B. G. (1976). The alimentary canal and digestion in teleosts. *Adv. Mar. Bio.* 13, 109-239.
- Kaushik, S.J. and Médale. (1994). Energy requirements, utilization and dietary supply to salmonids. *Aquaculture* 124, 81-97.
- Kihara, M., Sakata, T. (1997). Fermentation of dietary carbohydrates to short-chain fatty acids by gut microbes and its influence on intestinal morphology of a detritivorous teleost tilapia (*Oreochromis niloticus*). *Comp. Biochem. Physiol.* 118A, 1201-1207.
- Kihara, M., Sakata, T. (2002). Production of short-chain fatty acids and gas from various oligosaccharides by gut microbes of carp (*Cyprinus carpio L.*) in microscale batch culture. *Comp. Biochem. Physiol.* 132A, 333-340.
- Krogdahl, A., Hemre, G.I. & Mommsen, T.P. (2005). Carbohydrate in fish nutrition: digestion and absorption in post-larval stages. *Aquaculture nutrition.*, 11, 103-122.
- Krogdahl, A., Sundby, A. & Olli, J.J. (2004). Atlantic salmon (*Salmo salar*, L) and rainbow trout (*Oncorhynchus mykiss*) digest and metabolize nutrients differently depending on water salinity and dietary starch level. *Aquaculture*, 229, 335–360.
- Kumar, S., Sahu, N.P., Pal, A.K., Choudhury, D., Mukherjee, S.C. (2006). Studies on digestibility and digestive enzyme activity in *Labeo rohita* (Hamilton) juveniles: effect of microbial α-amylase supplementation in non-gelatinized or gelatinized corn-based diet at two protein levels. *Fish Physiol. Biochem.* 32, 209-220.
- Kumar, S.; Sahu, N. P.; Pal, A. K.; Choudhury, D.; Yengkokpam, S.; Mukherjee, S. C. (2005). Effect of dietary carbohydrate on haematology, respiratory burst activity and histological changes in L. rohita juveniles. *Fish and Shellfish Immunology* 19., 331-344.
- Kumar, V., Sahu, N., Pal, A., Kumar, S., & Gupta, S. (2008). Gelatinized to non-gelatinized starch ratio in the diet of *Labeo rohita*: effect on digestive and metabolic response and on growth. *Journal of animal physiology and animal nutrition*, 92(4), 492-501.
- Kumar, V., Sahu, N.P., Pal, A.K., Kumar, S., Sharma, P., Chettri, J.K., Sinha, A.K. (2009). Non-gelatinized starch influences the deposition of n-3 fatty acids in the muscle of a tropical freshwater fish, *Labeo rohita*. *J. Anim. Physiol. Animal Nutr.* 93, 659-668.
- Kumar, V.; Sahu, N. P.; Pal, A. K.; Kumar, S. (2007). Immunomodulation of *Labeo rohita* juveniles due to dietary gelatinized and non-gelatinized starch. *Fish and Shellfish Immunology* 23., 341-353.
- Leenhouwers, J.I., Ter Veld, M., Verreth, J.A.J., Schrama, J.W. (2007). Digesta characteristics and performance of African catfish (*Clarias gariepinus*) fed cereal grains that differ in viscosity. *Aquaculture 264*, 330-341.
- Li, J., Tan, B., Mai, K. (2009). Dietary probiotic *Bacillus OJ* and isomaltooligosaccharies influence the intestine microbial populations, immune

- responses and resistance to white spot syndrome virus in shrimp (*Litopenaeus vannamei*). *Aquaculture 291*, 35-40.
- Li, X.F., Liu, W.B., Lu, K.L., Xu, W.N. & Wang, Y. (2012). Dietary carbohydrate/lipid ratios affect stress, oxidative status and non-specific immune responses of fingerling blunt snout bream (*Megalobrama amblycephala*). Fish & Shellfish Immunology 33(2), 316-323.
- Li, X.F., Wang, Y., Liu, W.-B., Jiang, G.-Z. & Zhu, J. (2013). Effects of dietary carbohydrate/lipid ratios on growth performance, body composition and. *Aquaculture Nutrition* 2, 1-12.
- Liti, D.M., Mugo, R.M., Munguti, J.M., Waidbacher, H. (2006). Growth and economic performance of Nile tilapia (*Oreochromis niloticus L.*) fed on three brans (maize, wheat and rice) in fertilized ponds. *Aquacult. Nutr. 12*, 239-245.
- Long, L., Yang, J., Li, Y., Guan, Wu, F. (2015). Effect of biofloc technology on growth, digestive enzyme activity, hematology and immune response of genetically improved farmed tilapia (*Oreochromis niloticus*). Aquaculture 448, 135-141.
- Mingchun ren, Habte- Michael Habte- Tsion, Jun Xie, Bo Liu, Qunlan Zhou, Xianping Ge, Lliangkun Pa, Ruli Chenn. (2015). Effects of dietary carbohydrate source on growth performance, diet digestibility and liver glucose enzyme activity in blunt snout bream, *Megalobrama amblycephala*. Aquaculture 438, 75-81.
- Misra, S.; Sahu, N. P.; Pal, A. K.; Xavier, B.; Kumar, S. (2006). Pre- and post-challenge immuno-haematological changes in *Labeo rohita* juveniles fed gelatinised or non-gelatinised carbohydrate with n-3 PUFA. *Fish and Shellfish Immunology* 21, 346-356.
- Mjoun, K., Rosentrater, K.A. (2011). Extruded aquafeeds containing distillers dried grains with solubles: effects on extrudate properties and processing behavior. *J. Sci. Food Agricult.* 91, 2865-2874.
- Mohapatra, M.; Sahu, N. P.; Chaudhari, A. (2003). Utilization of gelatinized carbohydrate in diets in *Labeo rohita* fry. *Aquaculture Nutrition* 8., 1-8.
- Ng, W. K., Romano, N. (2013). A review of the nutrition and feeding management of farmed tilapia throughout the culture cycle. *Rev. Aquaculture*, 5, 220-254.
- Nordrum, S., Bakke-McKellep, A.M., Krogdahl, A. & Buddington, R.K. (2000). Effects of soybean meal and salinity on intestinal transport of nutrients in Atlantic salmon (*Salmo salar L.*) and rainbow trout (*Oncorhynchus mykiss*). *Comp. Biochem. Physiol. B, 125*, 317-335.
- Nordrum, S., Krogdahl, A., Røsjø, C., Olli, J.J. & Holm, H. (2000). Effects of methionine, cysteine and medium chain triglycerides on nutrient digestibility, absorption of amino acids along the intestinal tract and nutrient retention in Atlantic salmon (*Salmo salar L.*) under pair-feeding regime. *Aquaculture*, 186, 341-360.
- Nordrum, S., Olli, J.J., Røsjø, C., Holm, H. & Krogdahl, A. (2003). Effects of graded levels of medium chain triglycerides and cysteine on growth, digestive processes and nutrient utilization in seawater reared Atlantic salmon (*Salmo salar*, *L*) under ad libitum feeding regime. *Aquaculture nutrition.*, 9, 263-274.
- Novus. (1992). Raw material compendium. Ist edn. Novus International, St Louis.
- NRC, (national research council). (1983). United States Canadian Tables of Feed Composition. Committee on Animal Nutrition, Washington, D.C., 1983 Nutrient requirements of warmwater fishes and shellfishes. Washington, D.C.: National Academy Press.
- NRC, (national research council). (2011). *Nutrient requirements of fish and shrimp*. Washington D.C.: National Academy Press.

- Obaldo, L. G., & Masuda, R. (2006). Effect of diet size on feeding behavior and growth of Pacific white shrimp, *Litopenaeus vannamei*. *Journal of Applied Aquaculture*, 18, 101-110.
- P.J. White, A. Tziotis. (2004). New corn starches. In A. C. Eliasson, *Starch in Food, Structure, Function and Applications* (pp. 295-320). Cambridge: CRC press.
- Palmer, T.N. and Ryman, B.E. (1972). Studies of oral glucose intolerance in fish. *J. Fish Biol.*, 4, 311-319.
- Paul M. Insel, R. Elaine Turner, Don Rodd. (2006). Discovering Nutrition. Jones & Bartlett Learning.
- Paula Enes, Helena Peres. (2011). Growth, feed utilization and glycemic response in European Sea bass, *Dicentrarchus labrax*, Juveniles fed carbohydrate of different complexities. *Journal of the world aquaculture society*, 873-879.
- Peres, H., & Oliva-Teles, A. (2002). Utilization of raw and gelatinized starch by European sea bass (*Dicentrarchus labrax*) juveniles. *Aquaculture*, 96(3), 287-299.
- Pérez-Jiménez, A., Abellán, E., Arizcun, M., Cardenete, G., Morales, A.E., Hidalgo, M.C. (2015). Nutritional and metabolic responses in common dentex (*Dentex dentex*) fed on different types and levels of carbohydrates. *Comp. Biochem. Physiol.* 184A, 56-64.
- Pfeffer, E. (1995). Carbohydrate utilization and its determination. *J. Appl. Ichthyol.*, 11, 175–182.
- Pieper. A., Pfeffer.E. (1980). Studies on the comparative efficiency of utilization of gross energy from some carbohydrates, proteins and fats by rainbow trout (*Salmo gairdneri R.*). Aquaculture 20, 323-332.
- Pillay, T.V.R. (1990). *Aquaculture: Principle and practices*. London: Fishing Book News.
- Reinitz, G. (1983). Evaluation of sodium bentonite in practical diets for rainbow trout. *Prog.Fish-Cult.*, 45, 100–102.
- Ren, M., Habte-Tsion, H.-M., Xie, J., Liu, B., Zhou, Q., Ge, X., Pan, L., Chen, R. (2015). Effects of dietary carbohydrate source on growth performance, diet digestibility and liver glucose enzyme activity in blunt snout bream *Megalobrama amblycephala. Aquaculture 438*, 75-81.
- Robb, D.H.F., Crampton, V.O. (2013). On-farm feeding and feed management: perspectives from the fish feed industry. On-farm feeding and feed management in aquaculture. In M.R. Hasan and M.B. New, eds. *FAO Fisheries and Aquaculture Technical Paper No. 583. Rome, FAO.*, 489-518.
- Romano, N., Simon, W., Ebrahimi, M., Fadel, A.H.I., Chong, C.M., Kamarudin, M.S. (2016). Dietary sodium citrate improved oxidative stability in red hybrid tilapia (*Oreochromis sp.*) but reduced growth, health status, intestinal short chain fatty acids and induced liver damage. *Aquaculture 458*, 170-176.
- Rosa, D.S., Guedes, C.G.F., and Pedroso, A.G. (2004). Gelatinized and nongelatinized corn starch/ poly (caprolactone) blends: Characterization by rheological, mechanical and morphological properties. *Polimer: science and technology, 14*, 217-224.
- Rudel, L.L, & Morris, M.D. (1973). Determination of cholesterol using opthalaldehyde. *Journal of Lipid Research*, 14, 364-366.
- Safriel, O. & Bruton, M.N. (1984). A cooperative aquaculture research programme for South Africa. *South African National Scientific Programmes Report* 89., 79.
- Saha, S., Roy, R.N., Sen, S.K., Ray, A.K. (2006). Characterization of cellulase-producing bacterial from the digestive tract of tilapia, *Oreochromis mossambica*

- (Peters) and grass carp, Ctenopharyngodon idella (Valenciennes). Aquacult. Res. 37, 380-388.
- Sanchez Muros MJ, Garcia-rejon L, Lupia-ez J.A, De la Higuera M. (1996). Long term nutritional effects on the primary liver and kidney metabolism in rainbow trout (*Oncorhynchus mykiss*).ll. Adaptive response of glucose 6- phosphate dehydrogenase activity to high carbohydrate/ low protein and high fat/ non carbohydrate diets. *Aquaculture Nutrition*, 2., 193-200.
- Sayed, Abdel Fattah M. El-. (2006). *Tilapia culture*. Egypt: CABI publishing.
- Shiau, S.Y and Y.H. Lin. (2001). Carbohydrate utilization and its protein sparing effect in diets for grouper (*Epinephelus malabaricus*). *Animal science* 73, 299-304.
- Shiau, Shi Yen. (1997). Utilization of carbohydrates in warmwater fish with particular reference to tilapia *Oreochromis niloticus* x *O. aureus. Aquaculture* 151, 79-96.
- Shimeno, S. (1974). Studies on carbohydrate metabolism in fishes. *REPORTS OF THE FISHERIES LABORATORY, KOCHI UNIVERSITY*, 2-107.
- Shimeno, S., Mima, T., Kawabata, T., Takaku, H. (1993). Inclusion of oligosaccharide in soybean meal diet for fingerling yellowtail. *Suisanzoshoku 41*, 423-427.
- Sinha, A.K., Kumar, V., Makkar, H.P.S., Boeck, G.D., Becker, K. (2011). Non-starch polysaccharides and their role in fish nutrition a review. *Food Chem. 127*, 1409-1426.
- Skelton, P. (2001). A complete guide to the freshwater fishes of Southern Africa. Cape Town: Struik Publishers.
- Smith, L.S. (1989). Digestive functions in teleost fish. In *Fish Nutrition*, 2nd edn (ed by J. E. Halver) (pp. 332-423). Academic Press London.
- Sriburi, P., Hill, S. E., & Barclay, F. (1999). Depolymerization of cassava starch. *Carbohydrate Polymers*, 38, 211-218.
- Stone, D.A.J. (2003). Dietary carbohydrate utilization by fish. fish Science, 337-369.
- Storebakken, T., Shearer, K.D., Refstie, S., Lagocki, S. & McCool, J. (1998). Interaction between salinity, dietary carbohydrate source and carbohydrate concentration on the digestibility of macronutrients and energy in rainbow trout (*Oncorhynhus mykiss*). Aquaculture 163, 347-359.
- Storebakken, Trond. (1985). Binders in fish feeds: Effect of alginate and guar gum on growth, digestibility, feed intake and passage through the gastrointestinal tract of rainbow trout. *Aquaculture*, 47, 11-26.
- Svihus, B. (2014). Nutritive and Digestive Effects of Starch and Fiber in Whole Wheat. 81.
- Takeuchi, T., Jeong, K.-S., & Watanbe, T. (1990). Availability of extruded carbohydrate ingredients to rainbow trout *Oncorhynchus mykiss* and carp *Cyprinus carpio. Nippon Suisan Gakkaishi*, 56, 1839-1845.
- Tako, M., Tamaki, Y., Teruya, T., & Takeda, Y. (2014). The principles of starch gelatinization and retrogradation. *Food and Nutritional Sciences*, *5*, 280-291.
- Thodesen, J. & Storebakken, T. (1998). Digestibility of diets with precooked rye or wheat by Atlantic salmon, *Salmo salar L. Aquacult. Nutr.*, 4., 123-126.
- Thompson. (2000). On the non-random nature of amylopectin branching. *Carbohydrate Polymer*, 43, 223-239.
- Thongprajukaew, K., Kovitvadhi, U., Kovitvadhi, S., Somsueb, P., Rungruangsak-Torrissen, K. (2011). Effects of different modified diets on growth, digestive enzyme activities and muscle compositions in juvenile Siamese fighting fish (*Betta splendens Regan*, 1910). Auaculture 322-323,1-9.
- Thorpe, A. and Ince, B.W. (1974). The effects of pancreatic hormones, catecholamines, and glucose loading on blood metabolites in the northern pike (*Esox Lucius L.*). *Gen. Comp. Endocrinol.*, 23, 29-44.

- Thorpe, A. and Ince, B.W. (1976). Plasma insulin levels in teleosts determined by a charcoal separation radioimmunoassay technique. *Gen. Comp. Endocrinol.*, 30, 332-339.
- Tian, L.-X., Y.-J. Liu, and S.S.O. Hung. (2004). Utilization of glucose and corn starch by juvenile grass carp. *North American Journal of Aquaculture* 66, 141-145.
- Turan, F and I. Akyurt. (2005). Effects of red clover extract on growth performance and body composition of African catfish *Clarias gariepinus*. Fish science, 71., 618-620.
- Van der Waal, B.C.W. (1998). Survival strategies of sharptooth catfish *Clarias* gariepinus in desiccating pans in the northern Kruger National Park. Koedoe. *African Protected Area Conservation and Science* 41, 131-138.
- Venou, B., Alexis, M. N., Fountoulaki, E., Nengas, I., Apostolopoulou, M., & Castritsi Cathariou, I. (2003). Effect of extrusion of wheat and corn on gilthead sea bream (*Sparus aurata*) growth, nutrient utilization efficiency, rates of gastric evacuation and digestive enzyme activities. *Aquaculture* 225, 207-223.
- Venou, B., Alexis, M. N., Fountoulaki, E., Haralabous, J.(2009). Performance factors, body composition and digestion characteristic of gilthead sea bream (*Sparus aurata*) fed pelleted or extruded diets. *Aquaculture nutrition* 15,390-401.
- Viola, S., N. Gur and G. Zohar. (1986). Effects of pelleting temperature, binders and basic grains on water-stability of pellets and on growth of tilapia. *Bamidgeh*, 39, 19-26
- Volpe, M.G., Varricchio, E., Coccia, E., Santagata, G., Di Stasio, M., Mallinonico, M., Paolucci, M. (2012). Manufacturing pellets with different binders: Effect on water stability and feeding response in juvenile Cherax albidus. *Aquaculture 324-325*, 104-110.
- W.M., Bahnassey Y.A and Breene. (1994). 'Rapid viscoanalyzer (RVA) pasting profiles of wheat, corn, waxy corn, tapioca, and amaranth starches (A. hypochondriacus and A. cruentus) in the presence of konjac flour, gellan, guar, xanthan, and locust bean gums. *Starch*, 19, 134-141.
- Williams, K.C., Barlow, C.G., Rodgers, L., Hockings, I., Agcopra, C. & Ruscoe, I. (2003). Asian seabass *Lates calcarifer* perform well when fed pellet diets high in protein and lipid. *Aquaculture* 225, 191-206.
- Wilson, R.P. (1994). Utilization of dietary carbohydrate by fish. *Aquaculture 124(1-4)*, 67-80.
- Wilson, R.P. and Poe, W.E. (1987). Apparent inability of channel catfish to utilize dietary mono- and di-saccharides as energy sources. *J. Nutr.*, 117, 280-285.
- Wood, J.F. (1987). The functional properties of raw materials and their effects on the production and quality of feed pellets. *Animal Feed Science and Technology, 18*, 1-17.
- Yamamoto, T., Shima, T., Furuita, H., Suzuki, N. & Shiraishi, M. (2001). Nutrient digestibility values of a test diet determined by manual feeding and self-feeding in rainbow trout and common carp. *Fish Sci.*, 67, 355-357.
- Yengkokpam, S.; Sahu, N. P.; Pal, A. K.; Mukherjee, S.C.; Debnath, D. (2007). Gelatinized carbohydrates in the diet of *Catla catla* fingerlings: effect of levels and sources on nutrient utilization, body composition and tissue enzyme activities. *Asian Australasian Journal of Animal Science* 20, 89-99.
- Zhao, H.X., Cao, J.M., Liu, X.H., Zhu, X., Chen, S.C., Lan, H.B. & Wang, A.L. (2011). Effect of supplemental dietary zinc sources on the growth and carbohydrate utilization of tilapia *Oreochromis niloticus* x *Oreochromis aureus*. *Aquaculture Nutrition* 17(1), 64-72.

BIODATA OF STUDENT

Nagakanmani D/O Muniandy was born at Temerloh District, Pahang. The district is well known as 'Temerloh Bandar Ikan Patin'. She completed her primary school at Sekolah Kebangsaan Kuala Kaung (1996- 2001), Lanchang. Then she went to Sekolah Menengah Kebangsaan Datuk Bahaman (2002-2006), Lanchang for her secondary school. At the secondary school she finds that Science and Mathematics as her loved subjects and she grow the most interest in it. She successfully completed her Sijil Pelajaran Malaysia (SPM) on 2006 and continued with the Sijil Tinggi Pelajaran Malaysia at Sekolah Menengah Kebangsaan Mentakab (2007-2008), Mentakab.

In 2009 until 2013 she have been continued her study in Bio-industrial technology at Universiti Malaysia Kelantan. She had been graduated in Bachelor of Science (Hons) in Bio- industrial Technology successfully on 2013. While she pursuing her degree, her interest was to help her father in farming. Her family background was from the agriculture sector, so that she eagerly wants to help the farmers to produce the productive product and supply a good product to the community.

LIST OF PUBLICATION

Kanmani. N, Romano. N, Ebrahimi. M, Amin. S. M. N, Kamarudin. M. S, Karami. A and Kumar. V. 2017. Improvement of pellet characteristic by dietary pregelatinized starch and their subsequent effects on growth and physiology in tilapia. *Food Chemistry*, 239. 1037-1046.

Romano. N, Kanmani. N, Ebrahimi. M, Chong. C. M, Teh. J. C, Hoseinifar. S. H, Amin. N. S. M, Kamarudin. M. S, Kumar. V. 2018. Combination of dietary pre-gelatinized starch and isomaltooligosaccharides improved pellet characterisitics, subsequent feeding efficiencies and physiological status in African catfish, *Clarias gariepinus*, juveniles. *Aquaculture*, 484. 293-302.

