



***ASSESSMENT OF DRINKING WATER AND WASTEWATER QUALITY IN  
DAIRY CATTLE FARMS AND ITS ADVERSE EFFECTS ON HOST CELL  
RESPONSE IN MICE***

**NAGACHANDRA RAO A/L GOPI NAIDU**

**FPV 2020 3**



**ASSESSMENT OF DRINKING WATER AND WASTEWATER QUALITY IN  
DAIRY CATTLE FARMS AND ITS ADVERSE EFFECTS ON HOST CELL  
RESPONSE IN MICE**

By

**NAGACHANDRA RAO A/L GOPI NAIDU**

**Thesis Submitted to the School of Graduate Studies, University Putra Malaysia  
In Fulfillment of the Requirement of the Degree of Master of Veterinary  
Science**

**November 2019**

## **COPYRIGHT**

All material contained in this thesis, including without limitation text, logos, icons photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial use of material may only be made with the expression, prior written permission of University Putra Malaysia.

Copyright © University Putra Malaysia



## **DEDICATION**

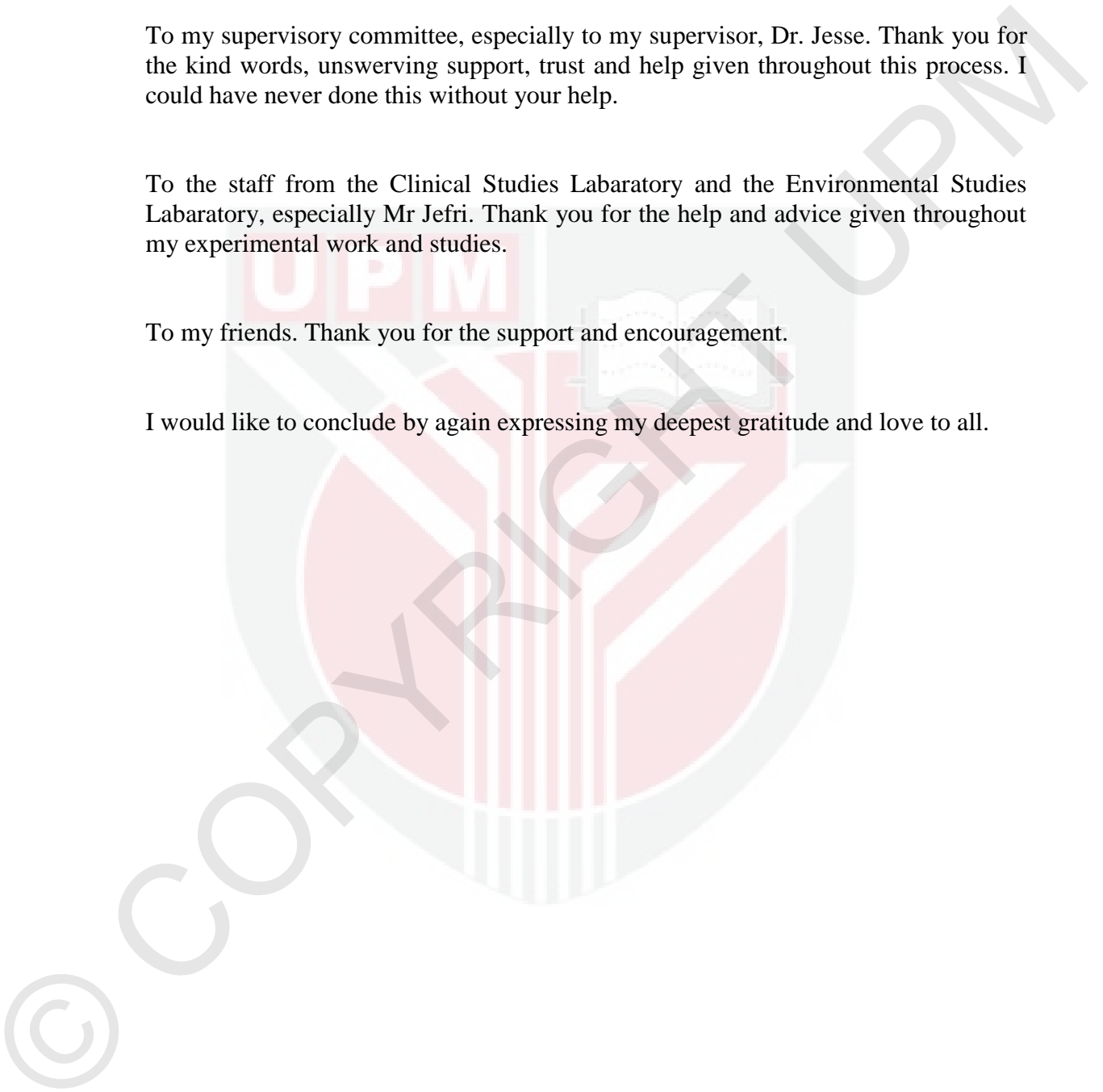
To my beloved family. Thank you for all the support, trust and encouragement throughout my study.

To my supervisory committee, especially to my supervisor, Dr. Jesse. Thank you for the kind words, unswerving support, trust and help given throughout this process. I could have never done this without your help.

To the staff from the Clinical Studies Laboratory and the Environmental Studies Laboratory, especially Mr Jefri. Thank you for the help and advice given throughout my experimental work and studies.

To my friends. Thank you for the support and encouragement.

I would like to conclude by again expressing my deepest gratitude and love to all.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in Fulfillment of the requirement for the degree of Master of Veterinary Science

**ASSESSMENT OF DRINKING WATER AND WASTEWATER QUALITY IN DAIRY CATTLE FARMS AND ITS ADVERSE EFFECTS ON HOST CELL RESPONSE IN MICE**

By

**NAGACHANDRA RAO A/L GOPI NAIDU**

**November 2019**

**Chairman : Professor Faez Firdaus Jesse Abdullah, PhD**  
**Faculty : Veterinary Medicine**

Water is an essential medium for animal metabolism as well as an important cleaning medium and it must be maintained at certain quality to avoid adverse effects on the animals. According to the National Water Quality Standards, water source with a classification of Class III and less is deemed suitable for animal consumption. The chances for water quality to be substandard is as the possibility of farmers using water from a questionable water source exists. Therefore, there is an urgent need to explore the possible adverse effects in the livestock from drinking the substandard drinking water. In our best knowledge, most cattle farms in Malaysia do not practice wastewater treatment prior releasing it into water bodies such as drains, rivers and lakes and this can lead to environmental pollution and adverse health effects to the fauna in the immediate surrounding. In this study 7 cattle farms were selected and both drinking water and wastewater samples were sampled for laboratory analysis. The water samples were analysed in-situ and in the lab for 15 water quality parameters which are dissolved oxygen, pH, salinity, electrical conductivity, turbidity, biological oxygen demand, chemical oxygen demand, total suspended solid, total dissolved solid, ammoniacal nitrogen, nitrate, phosphates, total coliform, iron and magnesium content. The results were then compared with the National Water Quality Standards and the overall classification for each water sample was decided. Then, the water classification was further narrowed to three categories for the purpose of this study, namely, good category (Class I and Class II), moderate category (Class III) and unsatisfactory category (Class IV and Class V). The results for drinking water were as follows; 1(14.29%) Class II, 2 (28.57%) Class III, 2 (28.57%) Class IV and 2 (28.57%) Class V. Meanwhile, all the wastewater samples in this study were categorized as Class V. A total of 35 female mice were divided into 5 groups; Group 1, 2, 3, 4 and 5 with 7 mice in each group. All the mice were gavage fed with 0.25 ml of water samples three times daily for a period of 30 days. Group 1 was fed with sterile deionized distilled water as the negative control, Group 2 was fed with the water sample from the good

category, Group 3 was fed with the water sample from moderate category and Group 4 was fed with the water sample from unsatisfactory category. Meanwhile, Group 5 was fed with one of the wastewater samples. The mice were observed at regular intervals and any mice that require humane end-points were euthanized and blood was collected through cardiac puncture before post-mortem examination and preservation of visceral organs and the brain. At the end of the 30 days, all surviving mice were sacrificed and blood, visceral organs and the brain was preserved for serological and histological examinations.

All treatment groups showed weight loss in comparison to the control group with the group treated with the wastewater showed most significant weight loss ( $p < 0.05$ ). Mice in groups 1, 2 and 3 did not show any significant clinical signs changes throughout the experimental period. In this experiment, 57% ( $n=4$ ) of the mice from group 4 exhibited moderate emaciation and mild ruffled fur at the end of the experiment while all the mice ( $n=7$ ) from group 5 exhibited clinical signs of severe ruffled fur, dehydration and severe emaciation and were euthanized to minimize pain. No significant gross lesions were observed in mice from groups 1, 2 and 3. Only 57% ( $n=4$ ) of the mice from group 4 showed signs of moderate dehydration while all the mice from group 5 showed signs of severe dehydration. 28.5% ( $n=2$ ) of the mice from group 2 and 14.2% ( $n=1$ ) of the mice from group 4 showed subcutaneous abscesses. Several coliform bacteria were isolated and identified from the abscesses.

Only mice from Group 5 showed significant decrease ( $p < 0.05$ ) in the serum estrogen levels in comparison to the control while no significant changes were observed in treatment groups in comparison to the control group for the serum progesterone levels.

In terms serum Ig G concentrations, only the wastewater treatment group showed significant elevation ( $p < 0.05$ ) in comparison to the control group. The treatment groups 3, 4 and 5 showed significant elevation ( $p < 0.05$ ) in comparison to the control group in serum Ig M concentrations. The treatment groups 4 and 5 showed significant elevation ( $p < 0.05$ ) in serum IL-12 concentrations in comparison to the control group.

Groups 3 and 4 showed significant elevation ( $p < 0.05$ ) in serum Hp concentrations in comparison to the control group while groups 3, 4 and 5 showed significant elevation ( $p < 0.05$ ) in SAA concentrations in comparison to the control group.

Histopathological analysis revealed that both the kidneys and the livers from the mice in group 3 showed moderate inflammation and degeneration cellular changes. Severe inflammation and degeneration were observed in both the kidneys and livers in mice from groups 4 and 5 while moderate inflammation were observed in uterus and ovaries of mice from groups 4 and 5. At the same time, mild presence of inflammatory cells and oedema was observed in groups 3, 4 and 5.

In conclusion, this study had proven that the drinking water provided for cattle consumption in most of the cattle farms are subpar as per the requirement set by the National Water Quality Standards. At the same time, due to a lack of wastewater treatment prior to its release to the environment, the quality of the wastewater is very poor and can become a source of pollution especially in cases where the farm is located nearby raw drinking water source areas. Based on the clinicopathological and serological symptoms exhibited by the mice in the treatment groups fed different qualities of drink water obtained from the dairy cattle farms, there is a high risk that there would be a negative impact in terms of production, reproduction and health from long term consumption of drinking water with subpar quality. Therefore, constant evaluation is required to ensure that the drinking water provided to the cattle in the dairy cattle farms are up to the standard for their consumption and the wastewater in the farms should be treated and evaluated before being released to the environment to avoid pollution.

**Keywords:** Drinking water; wastewater; National Water Quality Standards; dairy cattle farm, veterinary aspect, host cell response, mice model

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia bagi memenuhi keperluan untuk ijazah Master Sains Veterinar

**PENILAIAN KUALITI AIR MINUMAN DAN AIR KUMBAHAN DI  
LADANG-LADANG LEMBU TENUSU DAN KESAN BURUKNYA DENGAN  
RESPON SEL HOS DI TIKUS**

Oleh

**NAGACHANDRA RAO A/L GOPI NAIDU**

**November 2019**

**Pengerusi : Profesor Faez Firdaus Jesse Abdullah, PhD**  
**Fakulti : Perubatan Veterinar**

Air adalah medium yang penting di dalam metabolisme haiwan selain daripada peranannya sebagai medium pembersihan di dalam lading. Berdasarkan Piawaian Kualiti Air Kebangsaan Malaysia, apa-apa sumber air dengan kualiti keseluruhan Kelas III dan ke bawah adalah sesuai untuk digunakan sebagai air minuman haiwan ternakan. Kemungkinan untuk kualiti air yang kurang berkualiti adalah tinggi kerana kebanyakan peladang menggunakan sumber air yang boleh dipersoalkan. Oleh itu, ianya adalah penting untuk mengkaji kemungkinan kesan buruk di dalam haiwan ternakan yang minum air daripada sumber yang boleh dipersoalkan. Kebanyakan ladang lembu di Malaysia tidak merawat air kumbahan sebelum melepaskan air tersebut ke dalam badan-badan air seperti longkang, sungai dan tasik. Sebanyak 7 buah ladang lembu tenusus telah dipilih untuk kajian ini dan sampel air minuman dan air kumbahan telah dikumpul untuk analisis. Pemeriksaan sampel air tersebut telah dijalankan di ladang dan juga di makmal untuk 15 parameter berikut; oksigen terlarut, pH, kemasinan, kekonduksian elektrik, kekeruhan, keperluan oksigen biologi, keperluan oksigen kimia, jumlah pepejal tidak terlarut, jumlah pepejal terlarut, nitrogen ammonia, nitrat, fosfat, jumlah bakteria koliform, besi dan magnesium. Penemuan daripada penilaian di ladang dan makmal telah dibandingkan dengan Piawaian Kualiti Air Kebangsaan Malaysia dan pengkelasan keseluruhan untuk sampel-sampel air tersebut telah dibuat. Pada waktu yang sama, pengkelasan kualiti air tersebut telah diubahsuai untuk kajian ini kepada kategori bagus (Kelas I dan Kelas II), kategori sederhana (Kelas III) dan kategori tidak memuaskan (Kelas IV dan Kelas V). Keputusan penilaian kualiti air minuman adalah seperti berikut; 1(14.29%) Kelas II, 2 (28.57%) Kelas III, 2 (28.57%) Kelas IV dan 2 (28.57%) Kelas V. Kesemua sample air kumbahan pula telah dikelaskan sebagai Kelas V.



Sebanyak 35 ekor tikus betina telah dibahagikan kepada 5 kumpulan; Kumpulan 1, 2, 3, 4 dan 5 dengan 7 tikus di dalam setiap kumpulan. Kesemua tikus tersebut telah digavaj dengan 0.25 ml sampel air tiga kali sehari selama 30 hari. Kumpulan 1 telah diberi air tulen steril sebagai kumpulan kawalan negatif, Kumpulan 2 telah diberi sampel air daripada kategori memuaskan, Kumpulan 3 telah diberi sampel air daripada kategori sederhana, Kumpulan 4 telah diberi sampel air daripada kategori tidak memuaskan dan Kumpulan 5 telah diberi sampel air kumbahan. Kesemua tikus tersebut telah diperhatikan dari masa ke masa dan mana-mana tikus yang mempamerkan gejala klinikal yang memerlukan euthanasia telah dimatikan dengan cara dislokasi tulang vertebra serviks dan sampel darah telah dikumpulkan melalui penusukan jantung sebelum pemeriksaan post-mortem dan organ yang penting telah dipelihara di dalam cecair formalin 10%. Pada akhir tempoh eksperimen, kesemua tikus yang masih hidup telah dimatikan dan sampel darah, dan organ telah dikumpul untuk pemeriksaan serologi dan histologi.

Kesemua kumpulan rawatan menunjukkan kekurangan berat berbanding dengan kumpulan kawalan negatif dengan kumpulan yang dirawat dengan air kumbahan menunjukkan kekurangan berat yang ketara ( $p < 0.05$ ). Tikus di dalam kumpulan 1, 2 dan 3 tidak menunjukkan tanda-tanda klinikal yang ketara sepanjang tempoh eksperimen. 57% ( $n=4$ ) tikus daripada kumpulan 4 menunjukkan tanda-tanda kekurusan yang sederhana dan bulu tidak rata yang ringan. Pada masa yang sama, kesemua tikus daripada kumpulan 5 ( $n=7$ ) menunjukkan tanda-tanda dehidrasi, kekurusan dan bulu tidak rata yang teruk sebelum mereka dimatikan. Tiada perubahan ketara yang telah dapat diperhatikan secara kasar pada kesemua tikus-tikus daripada kumpulan 1, 2 dan 3. Hanya 57% ( $n=4$ ) daripada tikus-tikus daripada kumpulan 4 menunjukkan tanda-tanda dehidrasi yang sederhana manakala, kesemua tikus-tikus daripada kumpulan 5 menunjukkan tanda-tanda dehidrasi yang teruk. 28.5% ( $n=2$ ) tikus-tikus daripada kumpulan 2 dan 14.2% ( $n=1$ ) tikus-tikus daripada kumpulan 4 telah menunjukkan abses pada lapisan subkutaneus. Beberapa jenis bakteria jenis koliform telah berjaya diasingkan dan dikenalpasti daripada abses-abses tersebut.

Hanya tikus-tikus daripada kumpulan 5 menunjukkan penurunan yang ketara ( $p < 0.05$ ) dalam konsentrasi serum hormon estrogen berbanding dengan kumpulan kawalan. Dari segi konsentrasi serum hormon progesteron, kesemua kumpulan rawatan menunjukkan penurunan berbanding dengan kumpulan kawalan tanpa apa-apa penurunan ketara.

Hanya kumpulan yang dirawat dengan air kumbahan menunjukkan kenaikan yang ketara ( $p < 0.05$ ) di dalam konsentrasi serum Ig G berbanding dengan kumpulan kawalan. Kenaikan di dalam konsentrasi serum Ig M di dalam tikus-tikus daripada kumpulan 3, kumpulan 4 dan kumpulan 5 adalah ketara ( $p < 0.05$ ) berbanding dengan kumpulan kawalan. Kenaikan konsentrasi serum IL-12 di dalam tikus-tikus daripada kumpulan 4 and 5 adalah ketara ( $p < 0.05$ ) berbanding dengan kumpulan kawalan.

Kumpulan 3 dan 4 menunjukkan kenaikan yang ketara ( $p < 0.05$ ) di dalam konsentrasi serum Hp berbanding dengan kumpulan kawalan. Kumpulan rawatan 3, 4 dan 5 menunjukkan kenaikan yang ketara ( $p < 0.05$ ) di dalam konsentrasi serum SAA berbanding dengan kumpulan kawalan.

Analisa histopatologi telah mendedahkan yang hati dan buah pinggang daripada tikus-tikus daripada kumpulan 3 telah menunjukkan tanda-tanda keradangan dan degenerasi dari segi perubahan selular. Keradangan dan degenerasi yang teruk telah diperhatikan di dalam hati dan buah pinggang daripada tikus-tikus daripada kumpulan 4 dan 5 manakala keradangan yang sederhana telah diperhatikan di dalam uterus dan ovari daripada tikus-tikus daripada kumpulan 4 dan 5. Pada waktu yang sama, keradangan dan edema yang ringan telah diperhatikan di dalam kumpulan rawatan 3, 4 dan 5.

Kesimpulannya, kajian ini telah membuktikan bahawa air minuman yang diberikan kepada lembu-lembu di kebanyakan ladang-ladang lembu tenusu tidak mengikut standard yang telah ditentukan oleh Piawaian Kualiti Air Kebangsaan Malaysia. Pada waktu yang sama, ketiadaan sistem rawatan air kumbahan sebelum pelepasan air kumbahan tersebut menyebabkan kualiti air kumbahan tersebut adalah sangat teruk dan ianya boleh menjadi sumber untuk pencemaran, terutamanya apabila ladang tersebut berada berdekatan dengan sumber bekalan air minuman mentah. Berdasarkan tanda-tanda klinikal, patologi dan serologi yang telah dipamerkan oleh tikus-tikus daripada kumpulan-kumpulan yang telah diberikan dengan air minuman yang berlainan kualiti daripada ladang-ladang lembu tenusu, mempunyai risiko yang tinggi untuk impak negatif dari segi produksi, reproduksi dan kesihatan di dalam lembu-lembu tenusu sekiranya penggunaan air minuman yang berkualiti rendah. Oleh itu, penilaian yang berterusan adalah perlu untuk memastikan bahawa kualiti air minuman yang dibekalkan kepada lembu-lembu di ladang lembu tenusu adalah sesuai dan air kumbahan daripada ladang-ladang ini dirawat dan dianalisa sebelum ianya dilepaskan ke persekitaran bagi mengelakkan pencemaran.

**Kata kunci:** Air minuman; air kumbahan; Piawaian Kualiti Air Kebangsaan Malaysia; ladang lembu tenusu, aspek veterinar, respon sel hos, model tikus

## ACKNOWLEDGEMENTS

I would like thank the god without whom there would be nothing.

I would like to express my sincerest gratitude to my supervisor, Professor Dr. Faez Firdaus Jesse Abdullah and my supervisory committee members, Professor Dr. Abd Wahid Haron, Dr. Annas Salleh and Dr Wan Lutfi Wan Johari.

My gratitude also goes to Dr. Lawan Adamo, Dr. Eric Lim, Dr Yusuf Abba, Dr Idris Hambali, Dr. Muhammad Umar, Dr. Ali Dhia Mazra, Mr Mohd Jefri Norsidin and Mr Yap Keng Chee.

I would like to express my utmost appreciation to the Faculty of Veterinary Medicine, UPM and their Department of Veterinary Clinical studies as well as the Faculty of Environmental Studies and Department of Environmental Studies for affording with the opportunity to conduct my studies in their facilities.

I would also like to express my appreciation to the Universiti Putra Malaysia, School of Graduate Studies and the Ministry of Higher Education for providing with the opportunity to pursue my further studies.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Veterinary Science. The members of the Supervisory committee were as follows:

**Faez Firdaus Jesse Abdullah, PhD**

Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**Abd Wahid Haron, PhD**

Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

**Annas Bin Salleh, PhD**

Senior Lecturer  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

**Wan Lutfi Wan Johari, PhD**

Senior Lecturer  
Faculty of Environmental Studies  
Universiti Putra Malaysia  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 09 April 2020

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No: Nagachandra Rao a/l Gopi Naidu, GS 44417

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of  
Chairman of  
Supervisory Committee: Professor  
Faez Firdaus Jesse Abdullah

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory Committee: Professor  
Dr. Abd Wahid Haron,

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory Committee: Dr. Annas Bin Salleh

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory Committee: Dr. Wan Lutfi Wan Johari

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iv
<b>ACKNOWLEDGEMENTS</b>	vii
<b>APPROVAL</b>	viii
<b>DECLARATION</b>	x
<b>LIST OF TABLES</b>	xiv
<b>LIST OF FIGURES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xvi
 <b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Background	1
1.2 Problem Statement	2
1.3 Objectives	3
<b>2 LITERATURE REVIEW</b>	<b>4</b>
2.1 Background information of Malaysian cattle industry	4
2.1.1 Scope of Malaysian Cattle Industry	4
2.1.2 Hygiene standards of Malaysian cattle industry	5
2.2 Water	6
2.2.1 Role of water in cattle production	6
2.2.2 Drinking water sources in cattle farms	7
2.2.3 Factors affecting water consumption	8
2.2.4 Water contaminants	8
2.3 Drinking water and wastewater management of Malaysian cattle industry	9
2.3.1 Drinking water quality standards for cattle industry	9
2.3.2 Drinking water management in cattle farms.	9
2.3.3 Wastewater management in cattle farms	10
2.4 Clinical Examination of Mice	10
2.4.1 Normal and abnormal behaviour	10
2.4.2 Humane end-points	10
2.5 General Histopathology	11
2.6 Reproductive Efficiency	11
2.6.1 Female reproductive organs	11
2.6.2 Female reproductive hormones	11
2.7 Acute Phase Proteins	12
2.7.1 The acute phase protein response	12
2.7.2 Positive acute phase proteins	12
2.7.3 Negative acute phase	12
2.7.4 Function of acute phase proteins	12
2.8 Cytokine Response (IL-12)	12
2.9 Antibody Response	13



<b>3</b>	<b>MATERIALS AND METHODS</b>	14
3.1	Experimental Design	14
3.2	Study Approval	16
3.3	Cattle Farms Selection and Location	16
3.4	Water Sample Collection & In-situ Examination	16
3.5	Water Sample Analysis	17
3.5.1	Biological Oxygen Demand (BOD)	17
3.5.2	Chemical Oxygen Demand (COD)	18
3.5.3	Total Coliform	19
3.5.4	Total Soluble Solids (TSS)	19
3.5.5	Total Dissolved Solids (TDS)	20
3.5.6	Magnesium and Iron	20
3.5.7	Nitrate	21
3.5.8	Phosphate	21
3.5.9	Ammoniacal Nitrogen	21
3.6	Management and feeding of experimental animals	21
3.7	Gavage feeding of mice	22
3.8	Clinical examination of mice	23
3.8.1	Clinical examination and observation of mice	23
3.8.2	Humane end-point of mice	23
3.9	Reproductive hormone analysis	23
3.9.1	Estradiol RIA kit assay	23
3.9.2	Progesterone RIA kit assay	24
3.10	Immune response analyses	24
3.10.1	Mice Interleukin 12 (IL-12) ELISA kit assay	24
3.10.2	Mice Immunoglobulin G (Ig G) ELISA kit assay	24
3.10.3	Mice Immunoglobulin M (Ig M) ELISA kit assay	25
3.11	Acute phase protein response analyses	26
3.11.1	Mice haptoglobin (Hp) ELISA kit assay	26
3.11.2	Mice Serum Amyloid A (SAA) ELISA kit assay	26
3.12	Histopathological examination of mice	27
3.12.1	Post-mortem examination	27
3.12.2	Histopathology	27
3.12.3	Lesion Scoring	27
<b>4</b>	<b>RESULTS</b>	29
4.1	Water Quality Analysis	29
4.1.1	Drinking Water Quality Analysis	29
4.1.2	Wastewater Quality Analysis	31
4.2	Mice Clinicopathological Results	33
4.2.1	Changes in Body Weight	33
4.2.2	Clinical Signs and Responses	34
4.2.3	Gross Pathology	34
4.3	Responses of Female Reproductive Hormones (Estrogen and Progesterone)	35
4.3.1	Estrogen	35
4.3.2	Progesterone	35
4.4	Responses of Immunoglobulin G, Immunoglobulin M and Interleukin-12	36



4.4.1	Immunoglobulin G	36
4.4.2	Immunoglobulin M	37
4.4.3	Interleukin 12	38
4.5	Responses of Haptoglobin (Hp) and Serum Amyloid A (SAA)	39
4.5.1	Haptoglobin (Hp)	39
4.5.2	Serum Amyloid A (SAA)	40
4.6	Histopathological Changes of Mice	41
<b>5</b>	<b>DISCUSSION</b>	<b>51</b>
5.1	Water Quality Analysis	51
5.1.1	Drinking water quality analysis	51
5.1.2	Wastewater quality analysis	53
5.2	Mice Result	55
5.2.1	Changes in body weight	55
5.2.2	Clinical signs and responses	55
5.2.3	Gross Pathology	55
5.3	Responses of female reproductive hormones (estrogen and progesterone)	55
5.3.1	Estrogen	55
5.3.2	Progesterone	56
5.4	Responses of Immunoglobulin G, Immunoglobulin M and Interleukin 12	56
5.4.1	Immunoglobulin G	56
5.4.2	Immunoglobulin M	57
5.4.3	Interleukin 12	57
5.5	Response of haptoglobin (Hp) and serum amyloid A (SAA)	57
5.5.1	Haptoglobin	57
5.5.2	Serum amyloid A	57
5.6	Histopathological responses of mice	58
<b>6</b>	<b>SUMMARY, GENERAL CONCLUSION AND RECOMMENDATION</b>	<b>60</b>
	<b>REFERENCES</b>	<b>62</b>
	<b>BIODATA OF STUDENT</b>	<b>70</b>
	<b>PUBLICATION</b>	<b>71</b>

## LIST OF TABLES

<b>Table</b>		<b>Page</b>
3.4.1	In-situ parameters and the machine/ probes used to measure them	17
3.12.3.1	Histology lesion scoring	28
4.1.1.1	The table describes the results of parameters tested for samples of drinking water collected from selected cattle farms	30
4.1.2.1	The table describes the results of parameters tested for samples of wastewater collected from the selected cattle	32
4.6.1	The table shows the histopathological analysis results of the brain from mice from different treatment groups	41
4.6.2	The table shows the histopathological analysis results of the lungs from mice from different treatment groups	42
4.6.3	The table shows the histopathological analysis results of the spleen from mice from different treatment groups	42
4.6.4	The table shows the histopathological analysis results of the intestines from mice from different treatment groups	43
4.6.5	The table shows the histopathological analysis results of the stomach from mice from different treatment groups	43
4.6.6	The table shows the histopathological analysis results of the heart from mice from different treatment groups	44
4.6.7	The table shows the histopathological analysis results of the liver from mice from different treatment groups	44
4.6.8	The table shows the histopathological analysis results of the kidney from mice from different treatment groups	45
4.6.9	The table shows the histopathological analysis results of the uterus and ovaries from mice from different treatment groups	47

## LIST OF FIGURES

Figure		Page
3.1.1	The flow chart describing the experimental design of this study	16
3.6.1	The flow chart describing the management and feeding of the experimental animals	22
4.2.1.1	Mean changes of body weight in mice from different treatment groups.	33
4.2.3.1	A post-mortem image of a mice from group 4	34
4.3.1.1	Serum estrogen concentrations in mice from different treatment groups.	35
4.3.2.1	Serum progesterone concentrations in mice from different treatment groups	36
4.4.1.1	Serum Immunoglobulin G concentrations in mice from different treatment groups	37
4.4.2.1	Serum Immunoglobulin M concentrations in mice from different treatment groups	38
4.4.3.1	Serum Interleukin 12 concentrations in mice from different treatment groups	39
4.5.1.1	Serum Haptoglobin concentrations in mice from different treatment groups	40
4.5.2.1	Serum Amyloid A concentrations in mice from different treatment groups	41
4.6.1	Histomicrograph of the brain showing normal morphology in (A) control, (B) Group 2 and (C) Group 5. H & E x 200.	42
4.6.2	Histomicrograph of the lung of mice from group 5 showing mild interstitial inflammatory cell infiltration and oedema H & E x 200.	43
4.6.3	Histomicrograph of the spleen of mice from group 5 showing congestion in the red pulp with lymphocytes in the white pulp (arrows). H & E x 200.	44

4.6.4	Histomicrograph of intestine of mice from group 5 showing mild vacuolation (degeneration) (arrows). H & E x 200.	45
4.6.5	Histomicrograph of the heart of mice from group 5 showing normal morphology of cardiomyocytes. H & E x 200.	46
4.6.6	Histomicrograph of the liver of mice from group 5 showing necrotic cells (purple arrow) with kuppfer cell proliferation (red arrow). H & E x 200.	47
4.6.7	Histomicrograph of the kidney of mice from group 5 showing interstitial inflammatory cell infiltration tubular degeneration and necrosis (arrows). H & E x 200.	49
4.6.8	Histomicrograph of the uterus showing epithelia vacuolation with transformation of the epithelia (Metaplasia, yellow arrow) with few aggregates of degenerating neutrophils in the mucosal layer (red arrow). H & E x 200.	50

## LIST OF ABBREVIATIONS

ml	Mililiter
L	Liter
µl	Microliter
µg	Microgram
GAHP	Good Animal Husbandry Practices
SALT	Livestock Farm Accreditation Scheme
MyGAP	Malaysia Good Agricultural Practices
SALM	Good Agricultural Practices
SPLAM	Malaysia Aquaculture Farm Certification Scheme
DVS	Department of Veterinary Services
FAO	Food and Agriculture Organization of the United Nations
GAP	Good Animal Practices
CAP	Common Agricultural Policy
Il-12	Interleukin 12
Ig M	Immunoglobulin M
Ig G	Immunoglobulin G
Hp	Haptoglobin
SAA	Serum Amyloid A
DO	Dissolved Oxygen
BOD	Biological Oxygen Demand (BOD)
COD	Chemical Oxygen Demand
g	gram
TSS	Total Soluble Solid
TDS	Total Dissolved Solid

°C	Degree Celsius
ELISA	Enzyme-linked immunosorbent assay
RIA	Radioimmunoassay
ppt	Parts per trillion
μS/m	Micro Siemens per meter
CFU	Colony forming unit
NTU	Nephelometric turbidity unit
ICP	Inductively coupled plasma



# CHAPTER 1

## INTRODUCTION

### 1.1 Background

Water makes up to 90% of all molecules in the body and about 70% of the live body weight. It is also considered to be the most important of all the nutrients as it is needed in the most regular basis and in the highest volume (Sejian, V *et al.*, 2012). Domesticated animals can live about 60 days without food but they can only survive about 7 days without water (Church & Pond, 1974), therefore, it is advised for livestock to be given water in *ad libitum* as they can easily suffer from stress or dehydration due to lack of sufficient, clean drinking water (Sejian, V *et al.*, 2012).

Physiologically, water plays an important role in the transport of nutrients between cells as well as a vital medium for intracellular metabolism (Gropper *et al.*, 2009). There were some studies carried out and had shown that there are positive correlation between regular access to clean drinking water and performance such as growth, reproduction and milk production (Ensley *et al.*, 2000; Schutz *et al.*, 2012). Animals that has access to sufficient clean water are also usually less prone to illness and diseases due to reduced contact with pathogens and toxins in the water (Schutz *et al.*, 2012). Till date there is still no study was carried out in Malaysia to determine the quality of drinking water in ruminant livestock.

The common water sources in livestock farms are tap water, underground water, surface water such as lakes, ponds, rivers, swamps and drains as well as feed that is high in moisture such as green chop, pasture and silage (usgs.gov, 2005). Underground water typically contain less particulate matter such as leaves, soil and bugs as the grounds acts as a fine filtration mechanism as compared to the surface while on the other hand, it will also have a higher concentration of dissolved substances as the chemicals and minerals are leached off the ground as the water moves through it (usgs.gov, 2016). The natural composition of both ground water and surface water can be altered due to pollution through human activities, causing the infiltrations of contaminants such as pesticide, fertilizers, animal wastes and chemicals (Sasakova *et al.*, 2018). The quality of the tap water follows the standards of drinking water quality that is safe for human consumption as set by the various governments (moh.gov.my, 2004). Currently, there is a minimum drinking water quality that had been set by the National Water Quality Standards that is considered safe for livestock consumption in Malaysia (WEPA, 2006). This, in turn, raises a need to assess the quality of the ground water and surface water sources before they are deemed suitable for livestock consumption.

In the United Kingdom, United States and Canada, there are strict guidelines available for drinking water quality in livestock industry as well for sampling and testing

guidelines (Higgins *et al*, 2008; Wright, 2007). This is because most cattle farmers used ground water and surface water sources as drinking water to their livestock without any treatment, raising a need to check for the suitability of the water sources for livestock consumption. . Treatment such as filtration, chlorination or reverse osmosis must be performed in cases of substandard drinking water sources before it is deemed safe for consumption (Qadir *et al*, 2007)

Wastewater is water that has been adversely affected in quality through anthropogenic influence that originates from domestic, industrial, commercial or agricultural activities, surface runoff or stormwater and from sewer inflow or infiltration (WHO, 2014). Water is also used extensively in most livestock farming for the purpose of cleaning, animal cooling and sometimes for manure transfer leading to the production of copious amount of wastewater (FAO, 2019). Water is also used extensively in most livestock farming for the purpose of cleaning and this wastewater has been released into the rivers, drains and lakes without any proper treatment. These practices will lead to environmental pollution and as a medium of disease transfer. The proximity of the wastewater to the drinking water sources may increase the chances of polluting the drinking water. Currently, there is a severe lack of information on the drinking water and wastewater quality in the ruminant livestock industry of Malaysia.

Oral inoculation of contaminated water into an animal model such as mice and rats had been done in many instances in order to determine the possible effects from the consumption of the water (O'Donnell *et al.*, 2015). The clinicopathological effects from drinking the different water qualities can be an important indicator for the possible effects that the animals might suffer from the consumption of these water samples. For host cell responses such as acute phase proteins, antibodies, proinflammatory cytokines and reproductive hormones due to the effect of water contaminants in animals has not been studied till date. Therefore this study was designed to determine the quality of drinking water and wastewater in selected dairy cattle farms and to elucidate the clinicopathological and host cell responses due to oral inoculations of different qualities drinking water and wastewater using animal models.

## **1.2 Problem Statement**

Till date, there are still lack of study related to determining quality of drinking water and wastewater in livestock farming in Malaysia. Therefore, this study was designed to fill up the gap knowledge of water quality research in the ruminant livestock industry in Malaysia.



### 1.3 Objectives

The objectives of this study are:

1. To determine the water quality of the drinking water and the wastewater from selected dairy cattle farms.
2. To determine the clinical responses in mice inoculated with three different quality of water samples; namely good, moderate and unsatisfactory quality, from the assessment of objective (1) and a wastewater sample.
3. To determine the host cell responses in mice model from orally inoculated with three different quality of water samples; namely good, moderate and unsatisfactory quality, from the assessment of objective (1) and a wastewater sample focusing towards assessment of female reproductive hormones, acute phase protein and immunoglobulin concentration responses.

## REFERENCES

- Ahuja, S. (2019). Advances in water purification techniques: meeting the needs of developed and developing countries. Ahuja Consulting, Calabash, NC, United States.
- Aksoy, A. N., Toker, A., Celik, M., Aksoy, M., Halıcı, Z., & Aksoy, H. (2014). The effect of progesterone on systemic inflammation and oxidative stress in the rat model of sepsis. *Indian journal of pharmacology*, 46(6), 622–626. <https://doi.org/10.4103/0253-7613.144922>
- Anderson D. M., Gilbert, P. M. & Burkholder, J. M. (2002). Harmful algal blooms and eutrophication: Nutrient sources, composition and consequences. *Estuaries*, Volume 25, Issue 4, pg. 704-726.
- Anderson, W. B., Huck, P. M., Dixon, D. G., Mayfield, C. I. (2003). Endotoxin inactivation in water by using medium pressure UV lamps. *Appl. Environ. Microbiol.* 6:3002 – 3004.
- Archive.stats.govt.nz (2019)
- Bilotta, G. S. & Brazier, R. E. (2008). Understanding the influence of suspended solids on water quality and aquatic biota. *A Journal of International Water Association: Water Research*, 42, pg. 2849-2861.
- Binta, M. G. & Mushi, E. Z. (2012). Environmental factors associated with nitrate poisoning in livestock in Botswana. *Journal of Petroleum & Environmental Biotechnology*, Vol 3, Issue 6.
- Blumenthal, U. J., Mara, D. D., peasey, A., Ruiz-Palacios, G. & Stott, R. (2000). Guidelines for the microbiological quality of treated wastewater used in agriculture: Recommendations for revising WHO guidelines. *Bulletin of the World health Organization*, Geneva, 78 (9), pg. 1104-1116.
- Bouaziz-Ketata, H., Salah, G. B., Salah, H. B., Marrekchi, R., Jamoussi, K., Boudawara, T., Fakhfekh, F. & Zeghal, N. (2014). Nitrate-induced biochemical and histopathological changes in the liver of rats: Ameliorative effect of *Hyparrhenia hirta*. *Biomedical and Environmental Sciences*, Vol. 27, Issue 9, pg. 695-706.
- Bove, F., Shim, Y., Zeitz, P. 2002. Drinking water contaminants and adverse pregnancy outcomes: a review. *Environ. Health Perspect.* 110: 61–74.
- Brake, J. P., Hess, J. B. 2001. Evaluating water quality for poultry. Alabama Cooperative Extension System. Auburn University. Publ. ARN-1201. 4 pp.
- Byers, S. L., Wiles, M. V., Dunn, S. L & Taft, R. A (2012). Mouse estrous cycle identification: Tool and images. *PLOS One* 7 (4). e35538.

- Carter, T. A., Sneed, R. E. 1996. Drinking water quality for poultry. Published by: North Carolina Cooperative Extension Service. <http://www.bae.ncsu.edu/programs/extension/evans/ps&t-42.html>.
- Chew, K. W., Chia, S. R., Yen, H., Naomanbhay, S. Ho, Y. & Show, P. L. (2019). Transformation of biomass waste into sustainable organic fertilizers. *Sustainability*, 11(8), 2266
- Centers for Disease Control and Prevention, the Public Health Service, the U.S. Department of Health and Human Services, or the American Water Works Association. 2013. <http://www.cdc.gov/healthywater/emergency/dwa-comm-toolbox/index.html>.
- Chooi, K. E. (1993). Postmortem examination of transgenic mice. *Methods of Molecular Biology*, Volume 18, pg. 407-411.
- Church, D.C., Pond, W.G. 1974. Introduction to nutrition. In: Basic animal nutrition and feeding, John Wiley & Sons, Inc., New York. 3rd Ed.
- DAHD, In (2016). Livestock Population 2017. Department of Animal Husbandry and Dairying, India.
- Dan, G., Lall, S. B., Rao, D. N. 2000. Humoral and Cell Mediated Immune Response to Cadmium in Mice. *Drug Chem. Toxicol.* 23:349-360.
- Defra, UK (2018). Farming Statistics: Livestock Populations at 1 December 2017, United Kingdom. Department for Environmental Food & Rural Affairs, United Kingdom.
- Dey, P. (2018). Basic and advanced laboratory Techniques in histopathology and cytology. Springer Nature Singapore Pvt. Ltd., Singapore.
- Diersing, N. 2009. "Water Quality: Frequently Asked Questions." Florida Brooks National Marine Sanctuary, Key West, FL.
- DLD Th (2018). Number of livestock inventory in Thailand on 2017. Department of Livestock Development, Ministry of Agriculture and Cooperatives, Thailand.
- Dosm, My (2017). Supply and Utilization Accounts Selected Agricultural Commodities, Malaysia 2013-2017. Department of Statistics Malaysia.
- Dosm, My (2013). The Pig Industry in Malaysia. EPA and the Centers for Disease Control and Prevention (CDC), 2017.
- Eller, P., Eller, K., Kirsch, A. H., Patsch, J. J., Wolf, A. M., Tagwerker, A., Stanzl, U., Kaindl, R., Kahlenberg, V., Mayer, G., Patsch, J. R., & Rosenkranz, A. R. (2011). A murine model of phosphate nephropathy. *The American journal of pathology*, 178(5), 1999–2006. <https://doi.org/10.1016/j.ajpath.2011.01.024>
- El-Refaiy, A. I., & Eissa, F. I. (2013). Histopathology and cytotoxicity as biomarkers in treated rats with cadmium and some therapeutic agents. *Saudi journal of biological sciences*, 20(3), 265–280. doi:10.1016/j.sjbs.2013.02.004.

- Enderlein, U. S., Enderlein, R. E. & Williams, W. P. (1996). Water quality requirements, 2<sup>nd</sup> ed. Vol. 2. World Health Organization, Geneva.
- Ensley, S. M. (2000). Relationships of drinking water quality to production and reproduction in dairy herds. Retrospective Theses and Dissertations, Iowa State University.
- Fadiran, A. O., Dlamini, S. C. & Mavuso, A. (2008). A comparative study of the phosphate levels in some surface and ground water bodies of Switzerland. *Chemical Society of Ethiopia*, 22 (2), pg. 197-206.
- FAO (2019). Water use in livestock production system and supply chain. *Food and Agriculture Organization of the United Nations*, Rome, Italy.
- Goodrich, J. A., Lykins, B. W. & Clark, R. M. (1991). Drinking water from agriculturally contaminated groundwater. *Journal of Environmental Quality*, Vol. 20, No. 4, pg. 707-717.
- Grandjean, A. C., Campbell, S. M. 2004. Hydration: Fluids for Life. A monograph by the North American Branch of the International Life Science Institute. ILSI North America: Washington DC.
- Gropper, S. S., Smith, J. L. & Groff, J. L. (2009). Advanced nutrition and human metabolism. Wasworth, Belmont, California.
- Häussinger, D. 1996. The role of cellular hydration in the regulation of cell function. *Biochem. J.* 313: 697–710.
- Higgins, S. F. (2008). Drinking water quality guidelines for cattle. University of Kentucky, College of agriculture, United Kingdom.
- Hotchkiss, A. K., Ankley, G. T., Wilson, V. S., Hartig, P. C., Durhan, E. J., Jensen, K. M., Martinovi, D. & Gray, L. E. (2008). Of mice and men (and mosquitofish): Antiandrogens and androgens in the environment. *Bioscience*, Volume 58, Issue 11, pg. 1037-1050.
- Huang, Y. F., Ang, S. Y., Lee, K. M. & Lee, T. S. (2015). Research and practices in water quality: Quality of water resources in Malaysia. IntechOpen.
- Ibrahim, N. (2014). Trends on natural organic matter in drinking water sources and its impact. *International Journal of Scientific Research in Environmental Sciences*, 2 (3), pg. 94-106.
- Jadhav, S. H., Sarkar, S.N., Patil, R. D., Tripathi, H. C. 2007. Effects of subchronic exposure via drinking water to a mixture of eight water-contaminating metals: a biochemical and histopathological study in male rats. *Arch. Environ. Contam. Toxicol.* 53: 667–677.
- Jafarinejad, S. (2016). Cost estimation and economical evaluation of three configurations of activated sludge process for a wastewater treatment plant (WWTP) using simulation. *Applied Water Science*, 016, pg. 446-448.

- Je'quier, E., Constant, F. 2010. Water as an essential nutrient: the physiological basis of hydration. *European J. Clin. Nut.* 64: 115–123.
- Joan, F. K., Nancy, L. B., Susan, S. H., Kristin, S. L., John, K. L., Molly, A. M. 2009. Estimated Use of Water in the United States in 2005. U.S. Department of the Interior, U.S. Geological Survey. 1-52.
- Johnson, P. D. & Besselsen, D. G. (2002). Practical aspects of experimental design in animal research. *Institute of Laboratory Animal Research Journal*, Volume 43, Issue 4, pg. 202-206.
- Kenny, D. A., Humpherson, P. G., Leese, H. J., Tomos, A. D., Diskin, M. G. & Sreenan, J. M. (2002). Effect of elevated systemic concentrations of ammonia and urea on the metabolite and ionic composition of oviductal fluid in cattle. *Biology of Reproduction*, 66, pg. 1797-1804.
- Klein-Schneegans, A., Gaveriaux, C., Fonteneau, P. & Looor, F. (1988). Indirect double sandwich ELISA for the specific and quantitative measurement of mouse Ig M, Ig A and Ig G subclasses. *Journal of Immunological Methods*, Volume 119, Issue 1, pg. 117-125.
- Knight, R. L., Payne, V. W. E., Borer, R. E. , Clarke, R. A. & Pries, J. H. (2000). Constructed wetlands for livestock wastewater management. *Ecological Engineering*, 15 (1-2), pg. 41-55.
- Kramer, E. 2009. Water and feed intake in dairy cows-model evaluation and potential for health monitoring. M.S. Universitat zu Kiel, Kiel, Germany. 89pp.
- Landefeld, M., Bettinger, J. 2000. Water effects on livestock performance. *J. Agri. Nat. Res.* Columbus, oh. 43210-1084.
- Lardner, H. A., Kirychuk, B. D., Braul, L., Willms, W. D., Yarotski, J. 2005. The effect of water quality on cattle performance on pasture. *Australian J. Agricultural Res.* 56: 97–104.
- Leonard, F. C., Quinn, P. J., Ellis, W. A. 1992a. Possible effect of pH on the survival of leptospire in cattle urine. *Vet. Record* 131: 53-54.
- Leonard, F. C., Quinn, P. J., Ellis, W. A., O'Farrell, K. 1992b. Duration of urinary excretion of leptospire by cattle naturally or experimentally infected with *Leptospira interrogans* serovar hardjo. *Vet. Record* 131: 435-439.
- Luo, X., Yan, Q., Wang, C., Luo, C., Zhou, N. & Jian, C. (2015). Treatment of ammonia nitrogen wastewater in low concentration by two-stage ionization. *International Journal of Environmental Research: Public Health*, 12, pg. 11975-11987.
- Mako, A, Kocsis, M., Barna, G. & Toth, G. (2017). Mapping the storing and filtering capacity of European soils. Joint Research Centre, European Union.
- Mara, D., Horan, N. J. 2003. *The Handbook of Water and Wastewater, Microbiology*, Academic. Press, San Diego, California, USA.

- Mc Cue, M. D., Terblanche, J. S. & Benoit, J. B. (2017). Learning to starve: Impacts of food limitation beyond the stress period. *Journal of Experimental Biology*, 220, pg. 4330-4338.
- Michael, J. D., Ronald, D. S. 2014. *Veterinary Immunology: Principles and Practice*, Second Edition. CRC Press Textbook Color Illustrations. 336-328 Pp ISBN 9781482224627 - CAT# K22381.
- Ming, T. T., Hyun, K. T. & Joo, L. M. (2007). Characterization of livestock wastewater at various stages of wastewater treatment plant. *The Malaysian Journal of Analytical Sciences*, Vol 11, No. 1, pg. 23-28.
- MLA, Au (2018). Livestock distribution maps. Meat & Livestock Department Australia.
- MOH My. 2004. National Standard for Drinking Water Quality. Engineering Services Division. Ministry of health. Malaysia. Pp1-11.
- Montain, S. J., Maughan, R. J., Sawka, M. N. 1996. Fluid replacement strategies for exercise in hot weather. *Athletic Therapy Today*. 1:24–27.
- Nakade, U. P., Garg, S. K., Sharma, A., Choudhury, S., Yadav, R. S., Gupta, K., & Sood, N. (2015). Lead-induced adverse effects on the reproductive system of rats with particular reference to histopathological changes in uterus. *Indian journal of pharmacology*, 47(1), 22–26. <https://doi.org/10.4103/0253-7613.150317>
- Nasiadek, M., Danilewicz, M., Sitarek, K., Świątkowska, E., Daragó, A., Stragierowicz, J., & Kilanowicz, A. (2018). The effect of repeated cadmium oral exposure on the level of sex hormones, estrous cyclicity, and endometrium morphometry in female rats. *Environmental science and pollution research international*, 25(28), 28025–28038. <https://doi.org/10.1007/s11356-018-2821-5>
- NASS, US (2018). Overview of US livestock, poultry and aquaculture production in 2018 and statistics on major commodities. National Agricultural Statistics Service of the US Department of Agriculture.
- National Research Council (1992). Recognition and assessment of pain, stress and distress. National Academy Press, Washington, DC, pg. 32-53.
- Ngatia, L, Grace, J. M., Moriasi, D. & Taylor, R. (2018). Nitrogen and phosphorus eutrophication in marine ecosystem. IntechOpen.
- Nooraisyah, S., Nurulhuda, M. O., Mohd-Shaharizan, M. S., Azlina-Azma, I. A. 2014. Biosecurity Status of Small Ruminant Farms in Peninsular Malaysia. 26th Veterinary association Malaysia Congress. 5: 127-129.
- Norsyahariati, N. D. N. & Anijiofor, S. C. (2017). Livestock wastewater generation and farm management: The gap ananlysis. *Acta horticulturae*, 1152(1):265-272.



- Olkowski, A. A. (2009). *Livestock water quality: A field guide for cattle , horse, poultry and swine*. University of Saskatchewan, Canada.
- O'Donnell, H., Pham, O. H., Benoun, J. M., Raveslout-Chávez, M. M., & McSorley, S. J. (2015). Contaminated water delivery as a simple and effective method of experimental Salmonella infection. *Future microbiology*, *10*(10), 1615–1627. <https://doi.org/10.2217/fmb.15.93>
- Patel, B. G., Rudnicki, M., Yu, J., Shu, J. & Taylor, R. N. (2017). Progesterone resistance in endometriosis: origins, consequence and interventions. *Nordic Federation of Societies of Obstetrics and Gynecology, Acta Obstetrica et Gynecologica Scandinavica*, *96*, pg. 623-632.
- Patterson, H. H., Johnson, P. S, Epperson, W. B., Haigh, R. D. 2004. Effect of Total Dissolved Solids and Sulfates in Drinking Water for Growing Steers. South Dakota Beef Report. Paper 6.
- Peters, G. A., Yi, L., Skomorovska-Prokvolit, Y., Patel, B., Amini, P., Tan, H., & Mesiano, S. (2017). Inflammatory Stimuli Increase Progesterone Receptor-A Stability and Transrepressive Activity in Myometrial Cells. *Endocrinology*, *158*(1), 158–169. <https://doi.org/10.1210/en.2016-1537>
- Qadir, M., Sharma, B. R., Bruggeman, A., Choukr-Allah, R. & Karajeh, F. (2007). Non-conventional water resources and opportunities for water augmentation to achieve food security in water scarce in water scarce countries. *Agricultural Water Management*, Volume 87, Issue 1, pg..2-22.
- Rusydi, A. F. (2018). Correlation between conductivity and total dissolved solid in various types of water: A review. *IOP conf. Series: Earth and Environmental Science* *118*, pg. 102-019
- Rzymiski, P., Niedzielski, P. Rzymiski, P., Tomczyk, K., Kozak, L. & Poniedzialek, B. (2016). Metal accumulation in the human uterus varies by pathology and smoking status. *Fertility and Sterility*, Vol 105, No.6, pg. 1511-1518.
- Sanders, E. C., Yuan, Y. & Pitchford. A. (2013). Fecal coliform and *E. coli* concentrations in effluent-dominated streams of the Upper Santa Cruz Watershed. *MDPI: Water Journal*, *5*, pg. 243-261.
- Sasakova, N., Gregova, G., Takacova, D., Mojziso, J., Papajova, I., Venglovsky, J, Szaboova, T & Kovacova, S. (2018). Pollution of surface and ground water sources related to agricultural activity. *Frontiers in Sustainable Food System*, Vol 2, pg 42.
- Schutz, K. (2012). Effects of providing clean water on the health and productivity of cattle. Report for the Northland Regional Council, New Zealand.
- Sejian, V., Naqvi, S. M. K., Ezeji, T., Lakritz, J. & Lal, R. (2012). *Environmental stress and amelioration in livestock production*. Springer Heidelberg, New York, Dordrecht (London).

- Shih, A. W. Y., Mc Farlane, A. & Verhovsek, M (2013). Haptoglobin testing in hemolysis: Measurement and interpretation. *American Journal of Hematology*, Vol. 89, Issue 4, pg. 443-447.
- Shils, M. E., Shike, M., Ross, A.C., Caballero, B., Cousins, R. J. 2006. Modern Nutrition: In Health and Disease, Lippincott Williams & Wilkins, Philadelphia. 2006. 10th Ed.
- Stats NZ (2017). Agricultural production statistics: June 2017 (final).
- Statista Jakarta (2017). Livestock Statistics, Jakarta, Indonesia 2017. Kementerian Pertanian Republik Indonesia.
- Su, H., Li, Z., Kenston, S. S. F., Shi, H., Wang, Y., Song, X., Gu, Y., Barber, T., Aldinger, J., Zou, B., Ding, M., Zhao, J. & Lin, X. (2017). Joint toxicity of different heavy metal mixtures after a short-term oral repeated-administration in rats. *Int J Environ Res Public Health*, Vol 14 (10), 1164.
- Takata, S., Wada, H. Tamura, M., Koide, T., Higaki, M. Mikura, S. I. & Yasutake, T. (2011). Kinetics of c-reactive protein (CRP) and serum amyloid A (SAA) in patients with community-acquired pneumonia (CAP), as presented with biologic half-life times. *Biomarkers*, Vol 16 (6), pg 530-535.
- Tui, S. H., Valbuena, D., Masikati, P., Descheemaeker, K., Nyamangara, J., Erenstein, O., Rooyen, A. & Nkomboni, D. (2015). Economic trade-offs of biomass use in crop-livestock systems: Exploring more sustainable options in semi-arid Zimbabwe. *Agricultural Systems*, Vol. 134, pg. 48-60.
- Tyrrel, S. F. & Quinton, J. N. (2003). Overland flow transport of pathogens from agricultural land receiving fecal wastes. *Journal of Applied Microbiology*, 94, pg. 87S-93S.
- Uhlar, C. M. & Whitehead, A. S. (2001). Serum amyloid A, the major vertebrate acute-phase reactant. *European Journal of Biochemistry*, Volume 265, Issue 2, pg. 501-523.
- Unc, A. & Goss, M. J. (2003). Transport of bacteria from manure and protection of water sources. *Applied Soil Ecology*, Volume 25, Issue 1, pg. 1-18.
- Underwood, E. J. & Suttle, N. F. (1999). The mineral nutrition of livestock, 3rd edition. CABI Publishing, New York, USA.
- Usgs.gov.edu. 2005. Livestock water use. U.S. Department of the Interior | U.S. Geological Survey.
- Usgs.gov.edu. 2016. Contamination of groundwater. U.S. Department of the Interior | U.S. Geological Survey.
- Vodela, J. K., Rende, J. A., Lenz, S. D., Mchel Henney, W. H., Kemppainen, B. W. 1997. Drinking water contaminants. *Poult. Sci.* 76: 1474-1492.



- Vanotti, M. B., Garcia, M. C., Szogi, A. A., Hunt, P. G. & Millner, P. D. (2012). Method for recovery of phosphorus from animal waste. Annual Water Environment Federation Technical Exhibition and Conference, New Orleans, Oct 3, 2012.
- Water, Sanitation and Health Protection and the Human Environment World Health Organization Geneva. 2005
- Watanabe, T. (2016). The liver in systemic diseases: Physiological and pathological interactions between liver and kidney. Springer, Tokyo, Japan.
- WEPA (2006). National Water Quality Standards. Water Environment Protection Agency.
- Wright, C. L. (2007). Management of water quality for beef cattle. *Veterinary clinics of North America Food Animal Practice*, 23 (1), pg. 91-103.
- WHO. 2004. Guidelines for drinking water: Water Quality, 3rd ed. Vol. 1. World Health Organization, Geneva.
- WHO. 2015. Guidelines for drinking water: Sodium in Drinking Water. World Health Organization, Geneva. 2nd ed. Vol 2.