

EFFECTS OF TOTAL MIXED RATION WITH DIFFERENT LEVELS OF Lupinus angustifolius L. ON GROWTH PERFORMANCE AND MEAT QUALITY OF MALE BOER GOATS

AINUL YUZAIRI MOHD YUSOF

FPV 2019 23



EFFECTS OF TOTAL MIXED RATION WITH DIFFERENT LEVELS OF Lupinus angustifolius L. ON GROWTH PERFORMANCE AND MEAT QUALITY OF MALE BOER GOATS



AINUL YUZAIRI MOHD YUSOF

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

July 2018

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

 \mathbf{C}



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

EFFECTS OF TOTAL MIXED RATION WITH DIFFERENT LEVELS OF Lupinus angustifolius L. ON GROWTH PERFORMANCE AND MEAT QUALITY OF MALE BOER GOATS

By

AINUL YUZAIRI MOHD YUSOF

July 2018

Chairman: Professor Md Zuki bin Abu Bakar, PhDFaculty: Veterinary Medicine

As one of the major critical success factors in goat production, feed cost and its management have been given attention today to improve efficiencies and productivity of goat production. Total mixed ration (TMR) is an effective way of feeding goats by delivering a complete and nutritionally balanced diet for growing goats in intensive production system. Lupin grain (lupin) has high protein and dietary fiber content with minimal starch and glycemic index. Therefore, it has the potential as an alternative feed ingredient for livestock. However, the comparative data on the performances of Boer goat as affected by giving TMR with different level of lupin are still unavailable. Hence, present study was conducted to evaluate the effects of TMR diets with different lupin inclusion on the growth performance, carcass characteristics and meat quality traits of Boer goat. Twenty-four Boer male goats, 8-9-month-old of 27.83 ± 0.52 kg live body weight were allotted randomly into three treatment groups, namely CON, TMR A and TMR B, of eight animals each. Total mixed ration (TMR) feeds were formulated at 0%, 10% and 30% of lupin inclusion for CON, TMR A and TMR B groups, respectively. The TMR feeds were adjusted to be isocaloric (metabolizable energy: 10.3MJ/kg) and isonitrogenous (crude protein: 16.3%). The animals were fed twice (9.30am and 2.30pm) daily ad libitum during the 103-day feeding trial. All animals were humanely slaughtered according to the Halal slaughtering procedure at the end of the feeding trial. Present study showed that the total feed intake of TMR B (110.76 ± 3.57 kg) animals were significantly lower (P<0.05) than those in CON (135.05 \pm 5.04 kg) and TMR A (125.07 \pm 3.60 kg) groups. However, the final live weight, weight gained, and feed conversion ratio of the animals were unaffected (P>0.05) by the experimental diets. In addition, experimental diets also did not affect (P>0.05) the blood chemical content, carcass characteristics and composition of the Boer goats. In this study, the intramuscular fat in the longissimus dorsi (LD) muscle of TMR B (13.24 \pm 1.28%) was lower (P<0.05) than that of CON $(21.05 \pm 1.49\%)$ and TMR A $(19.63 \pm 1.53\%)$. There was insignificant difference (P>0.05) in the other meat quality traits of the LD muscle among the experimental diets. Meanwhile, the fatty acid composition of LD, biceps branchii (BB) and semitendinosus (ST) muscles were affected by the experimental diets. In LD muscle, the proportions of lauric (C12:0), palmitoleic (C16:1) and linolenic (C18:3n-3) acids were significantly affected (P<0.05) by the experimental diets. Whilst, the proportion of pentadecanoic acid was significantly lower (P<0.05) in CON than that of TMR A in BB muscle. In ST muscle, TMR B had higher (P<0.05) proportion of lauric acid than CON, while the proportion of heptadecanoic acid was higher (P<0.05) in TMR A than that in TMR B. The total saturated fatty acid (SFA), monounsaturated fatty acid (MUFA) and polyunsaturated fatty acid (PUFA) as well as the PUFA: SFA and n-6: n-3 ratios in all the three skeletal muscles were unaffected (P>0.05) by the experimental diets. Present findings demonstrated that feeding TMR diets with the up to 30% inclusion of lupin did not adversely affect the growth performance, carcass characteristics and meat quality of Boer goats. Hence, this finding suggested that TMR feeding with up to 30% of lupin inclusion is suitable for intensive farming of goats and serve as an alternative protein source to the animals.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KESAN RANSUM CAMPURAN LENGKAP (TOTAL MIXED RATION – TMR) DENGAN KANDUNGAN BERBEZA ARAS Lupinus angustifolius L. KE ATAS PRESTASI TUMBESARAN DAN KUALITI DAGING KAMBING BOER JANTAN

Oleh

AINUL YUZAIRI MOHD YUSOF

Julai 2018

Pengerusi : Profesor Md Zuki bin Abu Bakar, PhD Fakulti : Perubatan Veterinar

Sebagai salah satu faktor utama kejayaan kritikal dalam pengeluaran kambing, kos makanan dan pengurusan telah diberi perhatian hari ini untuk meningkatkan kecekapan dan produktiviti pengeluaran kambing. Ransum campuran lengkap (TMR) adalah kaedah yang berkesan untuk pemberian makanan kambing yang mampu memastikan pengambilan nutrien yang lengkap dan seimbang untuk kambing dalam sistem pengeluaran intensif. Bijirin Lupin (lupin) mengandungi kandungan protein dan serabut diet yang tinggi serta kanji dan indeks glisemik yang minimum. Oleh itu, lupin berpotensi digunakan sebagai bahan makanan alternatif untuk ternakan. Namun, kesan TMR dengan pelbagai aras lupin ke atas prestasi kambing Boer masih tidak dikaji. Justeru itu, kajian ini dijalankan untuk menilai kesan TMR dengan pelbagai tahap lupin dalam makanan ke atas prestasi tumbesaran, ciri karkas dan kualiti daging kambing Boer. Dua puluh empat ekor kambing jantan berumur 8-9 bulan dengan purata berat badan 27.83 ± 0.52 kg telah diasingkan secara rawak kepada tiga kumpulan, iaitu CON, TMR A dan TMR B, dengan 8 haiwan dalam setiap kumpulan. Makanan dalam bentuk total mixed ration (TMR) telah diformulasikan dengan mengandungi 0%, 10% dan 30% lupin dalam kumpulan CON, TMR A dan TMR B masing-masing. Makanan TMR juga diselaraskan untuk mengandungi tenaga metabolik (10.3 MJ/kg) dan protein kasar (16.3%) yang sama. Kambing diberi makan dua kali sehari (9.30am dan 2.30pm) secara ad libitum selama 103 hari kajian ini dijalankan. Pada hari ke-103, semua kambing telah disembelih mengikut prosedur penyembelihan Halal. Hasil kajian menunjukkan bahawa jumlah pengambikan makanan kambing dalam kumpulan TMR B (110.76 ± 3.57 kg) adalah lebih rendah (P<0.05) daripada kumpulan CON (135.05 ± 5.04 kg) dan TMR A (125.07 \pm 3.60 kg). Akan tetapi, berat badan akhir, tambahan berat badan dan nisbah perubahan makanan tidak dipengaruhi (P>0.05) oleh makanan yang diberikan. Tambahan pula, tidak terdapat perbezaan tererti (P>0.05) pada kandungan biokimia darah, ciri karkas dan komposisi di antara kumpulan kambing tersebut. Dalam kajian ini, kandungan lemak intraotot dalam otot longissimus dorsi (LD) adalah lebih rendah (P<0.05) dalam TMR B $(13.24 \pm 1.28\%)$ berbanding dengan CON $(21.05 \pm 1.49\%)$ dan TMR A $(19.63 \pm 1.53\%)$.

iii

Namun, tidak terdapat perbezaan tererti (P>0.05) pada ciri-ciri kualiti daging yang lain pada otot LD di antara kumpulan kambing tersebut. Sementara itu, komposisi asid lemak otot LD, biceps branchii (BB) dan semitendinosus (ST) adalah berbeza di antara kumpulan kambing kajian. Kandungan asid laurik (C12:0), palmitoleik (C16:1) dan linolenik (C18:3n-3) dalam otot LD adalah berbeza (P<0.05) di antara kumpulan kambing. Kandungan asid *pentadecanoic* dalam otot BB pula adalah lebih rendah (P<0.05) dalam kumpulan CON berbanding dengan TMR A. Dalam otot ST, kumpulan TMR B mengandungi asid laurik yang lebih tinggi (P<0.05) daripada CON, manakala kandungan asid *heptadecanoic* adalah lebih tinggi (P<0.05) dalam kumpulan TMR A berbanding dengan TMR B. Jumlah asid lemak tepu (SFA), asid lemak monotaktepu (MUFA) dan asid lemak politaktepu (PUFA) serta nisbah n-6: n-3 dan PUFA: SFA tidak mempunyai perbezaan tererti (P>0.05) di antara semua kumpulan kajian. Kajian ini menunjukkan bahawa makanan secara formulasi TMR yang dimasukkan dengan tahap lupin yang berbeza sehingga 30% komposisi diet tidak menjejaskan prestasi tumbesaran. ciri karkas dan kualiti daging kambing Boer. Oleh itu, kajian ini mencadangkan bahawa formulasi secara TMR yang menggunakan lupin sehingga 30% dari komposisi diet sesuai digunakan sebagai sumber protein alternatif bagi penternakan intensif kambing.

ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious and the Most Merciful.

Alhamdulillah, all praises to Allah for the strengths and His blessing in completing this thesis. I would like to thank all those people who Allah choose them in helping me throughout this journey. First of all, I pay my gratitude to the Chairman of the Supervisory Committee, Prof. Dr. Md Zuki bin Abu Bakar, whose guidance, patience and constructive comments was valuable. His timely and efficient contribution helped me conducting the experiment and shape this thesis into its final form and I express my sincerest appreciation for his assistance in any way that I may have asked. I am also deeply indebted to the members of supervisory committee Prof. Dr. Goh Yong Meng, Dr. Yusof Hamali and Dr. Abu Hassan B. Muhammad Ali, for their sincere help in sharing their expertise, providing invaluable advice and guidance in designing the experiment, providing necessary resources to accomplish my research work. I would also like to thank the Government of Malaysia particularly Department of Public Services and Department of Veterinary Services to grant me scholarship of Hadiah Latihan Persekutuan (HLP) and necessary institutional support in continuing my postgraduate study in UPM. My sincere thanks also go to the Faculty of Veterinary Medicine specifically, Prof. Dr. Mohd. Hair Bejo and Prof. Dr. Zunita Zakaria for their kind cooperation and assistance throughout my study process. My sincere appreciation also goes to all the great people involve directly in the experimental and research process at several place including Pusat Pembiakan Kambing Kg. Pah (Dr. Khairil Azman, En Hisham Latif, En. Mohd. Fadil Nazir, En. Suhaimi Ahmad, En. Rosdi Mohd Khamis, En. Jefridin Mansur, En. M. Nizan Aziz, En. Chinnayah Gopal, En. Mat Zaidi Mat Ali, Pn. Dasimah Mahmud, Pn. Roshana Othman, En. Abu Safiaan Shaharuddin, En. Shamsudin Yusof, En Abdul Karim Abu Bakar, En. Tesmin Rashid), Senawang Abattoir, MKAV Salak Tinggi, Feed Analysis Lab IVM, Pre-Clinical and Clinical Pathology Laboratory FPV, UPM. The support and sincere help by them have made the experiment and laboratory work run smoothly as projected. I am really grateful to Allah to send to me great peoples who have provide me their bright thoughts and kind help for shaping up my research, conducting the laboratory, thesis writing and statistical analysis: Dr. Mehdi Ebrahimi, Tn. Hj. Syed Hussein Syed Abdullah, Mr. Leo Teo Kee, Dr. Nur Islam, Dr. Kamal Abdelfatah, Dr. Ahmad Tunio, Prof Madya Dr. Lokman Hakim Idris and En. Sharif Saniman. Specials thanks are due to Dr. Azhari Sheridan Abu Bakar, Dr. Fazilah Aini, Dr. Zarina Mohamed, and Dr. Zeti Yuzreena Abd. Khalid, Dr. Emelia Aini Kamaruzzaman, Dr Muhammad Nazri Khairuddin, Dr Farah Alias and Dr Zurin Azlin Md. Jinin, who had joined the journey together with their encouragement and support from sincere heart as student n friends. Finally, I'm indebted to my family for their understanding and encouragement and love, to my parent: Mohd. Yusof Mohd Sarif, Mother: Pn Timah Bidin, my wife, Roslina Ibrahim and my children: Ikhwan, Fasihah, Dalylla & Ageef. Special thanks, tribute and appreciation to all those their names do not appear here who have contributed to the successful completion of this study.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Md Zuki bin Abu Bakar, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Dr. Yusof Hamali Ahmad

Senior Lecturer Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

Goh Yong Meng, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:	
-		

Name and Matric No.: Ainul Yuzairi Mohd Yusof (GS 29397)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	Professor Dr. Md Zuki bin Abu Bakar
Signature: Name of Member of Supervisory Committee:	Dr. Yusof Hamali Ahmad
Signature: Name of Member of Supervisory Committee:	Associate Professor Dr Goh Yong Meng

TABLE OF CONTENTS

Page
i
iii
v
vi
viii
xii
xiii
xiv

CHAPTER

1.	INTRODUCTION	1
2.	LITERATURE REVIEW	4
	2.1 Introduction	4
	2.2 Goat Meat Production	6
	2.2.1 Boer goat	8
	2.3 Total mixed Ration (TMR)	9
	2.4 Lupin	10
	2.4.1 Lupin as animal feeds	13
	2.5 Meat Quality	16
	2.5.1 Color	17
	2.5.2 Water holding capacity	17
	2.5.3 Texture	17
	2.6 Fatty Acid Composition in Meat	18
3.	MATERIALS AND METHODS	21
	3.1 Experimental Animal Management and Feeding	21
	3.2 Formulation and Preparation of Experimental Diets	21
	3.3 Proximate Analysis of Experimental Diets	24
	3.3.1 Dry matter	24
	3.3.2 Crude protein	24
	3.3.3 Crude fiber	25
	3.3.4 Acid detergent fiber	26
	3.3.5 Neutral detergent fiber	26
	3.3.6 Ether extract	27
	3.3.7 Gross energy	27
	3.3.8 Calcium	28
	3.3.9 Phosphorus	28
	3.4 Growth Performance Evaluation	29
	3.5 Blood Biochemical Content	29
	3.6 Slaughtering	29
	3.7 Carcass Characteristics Evaluation	29
	3.8 Meat Quality Traits Evaluation	30
	3.8.1 pH	30
	3.8.2 Color	30
	3.8.3 Water holding capacity	31

	3.8.4 Texture	31
	3.9 Fatty Acid Composition	31
	3.9.1 Total lipid extraction	31
	3.9.2 Fatty acid methyl ester preparation	32
	3.9.3 Fatty acid methyl ester quantification	32
	3.10 Data Analysis	33
4.	RESULTS AND DISCUSSIONS	34
	4.1 Proximate Analysis of Experimental Diets	34
	4.2 Growth Performance	35
	4.3 Blood Biochemical Content	37
	4.4 Carcass Characteristics and Carcass Composition	38
	4.5 Meat Quality Traits	40
	4.6 Fatty Acid Composition	41
	4.6.1 Fatty acid composition of experimental diets	41
	4.6.2 Fatty acid composition of skeletal muscles	42
5.	CONCLUSIONS	48
REFER	ENCES	49
APPEN	DIX A	57
BIODA	TA OF STUDENT	59

 \bigcirc

LIST OF TABLES

Table		Page
3.1	Ingredients of Experimental Diets and Proximate Values and Composition in Each TMR Treatment	23
4.1	Proximate Analysis of the Experimental Diets.	35
4.2	Effects of Different Levels of Lupin on Growth Performance of Boer Goats.	36
4.3	Effects of Different Levels of Lupin on Blood Biochemical Content of Boer Goats.	38
4.4	Effects of Different Levels of Lupin on Carcass Characteristics of Boer Goats.	39
4.5	Effects of Different Levels of Lupin on Carcass Composition of Boer Goats.	40
4.6	Effects of Different Levels of Lupin on Meat Quality Traits of Boer Goats.	41
4.7	Fatty Acid Composition of Experimental Diets.	42
4.8	Effects of Different Levels of Lupin on Fatty Acid Composition of <i>Longissimus Dorsi</i> Muscle in Boer Goats.	45
4.9	Effects of Different Levels of Lupin on Fatty Acid Composition of <i>Biceps Branchii</i> Muscle in Boer Goats.	46
4.10	Effects of Different Levels of Lupin on Fatty Acid Composition of <i>Semitendinosus</i> Muscle in Boer Goats.	47

C)

LIST OF FIGURES

Figure		Page
2.1	Main Commodity and Externalities Outputs in Small Ruminant Production Systems.	6
2.2	Boer Buck.	8
2.3	The Life Cycle of Lupin.	11
2.4	Narrow-Leaf Lupin Tree.	12
2.5	Structure of Narrow-Leaf Lupin Seed.	13
2.6	Narrow-Leaf Lupin Seed.	13
3.1	Experimental Design	22
A.1	The Goat Shed for the Feeding Trial	57
A.2	The Experimental Goat in The Individual Pen	57
A.3	Total Mixed Ration Used in The Feeding Trial.	57
A.4	Experimental Animals.	58

6

LIST OF ABBREVIATIONS

a*	Redness
ADF	Acid Detergent Fiber
ASL	Australian Sweet Lupin
b*	Yellowness
Ca	Calcium
CON	Control
СР	Crude Protein
CVD	Cardiovascular Diseases
DM	Dry Matter
DVS	Department of Veterinary Services
FAME	Fatty Acid Methyl Esters
FCR	Feed Conversion Ratio
g	Acceleration Of Gravity
h	Hour
HDL	High Density Lipoprotein
IACUC	Institutional Animal Care and Use Committee
IMF	Intramuscular Fat
L*	Lightness
LDL	Low Density Lipoprotein
М	Molar
min	Minute
ME	Metabolizable Energy
MOA	Ministry of Agriculture
MUFA	Monounsaturated Fatty Acids
Mt.	Metric Ton
Ν	Normal
NDF	Neutral Detergent Fiber
Р	Phosphorus
PUFA	Polyunsaturated Fatty Acids
S.E.M.	Standard Error of Mean
SFA	Saturated Fatty Acids
TMR	Total Mixed Ration
UFA	Unsaturated Fatty Acid

 \bigcirc

CHAPTER 1

INTRODUCTION

Livestock production is the integral part for world population to provide protein source for human diet. Hundreds of millions of people in the world involve in livestock production and customarily rely on their animals to provide multiple products and services.

The global livestock industry is focusing to improve production and productivity driven by the expanding human population and better income that create better market for meat and dairy products particularly in developing countries. However, the impact of livestock farming to the environmental issues such as climate change has created concern among professionals to reevaluate the sustainability of livestock products in our diet. In the attempt to increase meat production, more efficient animal production systems, duly managed of natural resources and measures to reduce waste and environmental pollution is necessary approach for us to achieve viable and sustainable animal industry. The current livestock industry has been impelled by least cost of production while trying to meet various product and market segment and requirements to achieve maximal returns (Hayes, et.al. 2013; Wyness et al., 2011; Peacock & Sherman, 2010)

According to Sivasupramaniam (2006), majority of ruminant farmers in Malaysia can be classified under the category of smallholders or traditional farmers. However, since the implementation of ninth and tenth (9th and 10th) Malaysian Plan, there are increased of number of commercial goat farms that using intensive production system. In the intensification of goat farming in Malaysia, farmers faced the challenge of increasing production cost particularly the feed cost to fatten goats for slaughter which usually will require a good quality feed and more concentrates. Forages and fodder feed sources alone in intensive goat production system is inadequate to achieve balanced nutrient for fast growing animals and concentrates need to be supplemented in order to maintain acceptable goat performance. Furthermore, the fact of inconsistent supply in terms of quantity and quality of fodder in goat farms due to climatic changes and land scarcity for fodder production (Röös et al., 2016) can affect goat farmers to find alternative feed sources.

Malaysia still dependent on importation of live goats and frozen meat from sheep and goat to meet the local demand. In 2015, Malaysia imported 32,206.78 Mt. of frozen lamb and mutton from Australia and New Zealand while 145,353 heads of live sheep and goats imported mostly from Australia (Department of Veterinary Services, 2016). With only 12.22% self sufficiency level and 1.43 kg of per kapita consumption in 2015 for sheep and goat meat (Department of Veterinary Services, 2016), the prospect for growth of small ruminant farming seems to be quite engaging.

The increasing feed price particularly grains such as soybean meal and corn as important protein and energy sources for animal feed in the world commodity market recently is a

serious threat to the competitiveness of intensified goat farming in Malaysia. To overcome this problem, farmers can opt to shorten the rearing period to fatten for their goat to achieve market size by applying cost effective feeding management and using faster growth breed with good genetic potential to produce good quality carcass and meat.

To properly fed goat commensurate to their nutritional requirements, there are several opportunities in using good quality and affordable ingredients such as palm kernel expeller, wheat pollard, soya bean meal, soya waste and fishmeal. The nutritional values, anti-nutritional factors, consistencies in nutritional quality and supply and also cost of the ingredient are the important factors in the formulating cost effective feed (Wan Zahari & Wong, 2009)

Lupin seed has been adapted as one of feed ingredients for livestock production in Australia, Japan, and Korea and also in several countries Europe and Asia (Petterson, 2000). Lupin seed has been recognized by its high protein and dietary fiber and minimal starch and glycemic index that have potential as alternative feed ingredients for ruminants, poultry, pig and fish production (van Barneveld, 1999). The limitation of using lupin in TMR feed for goat are the dependency of lupin supply from imported source particularly from Australia since lupin is not suitable to be planted in Malaysian tropical climate. Despite the limitation on the risk of economic viability for using lupin today in Malaysia, the technical feasibility of using lupin in goat feed is still worth to be studied since other common feed source for ruminant concentrate particularly soybean meal are also imported ingredient and subject to price and supply availability fluctuations. The current study has only examined usage of raw lupin grain in TMR to feed male Boer goats.

During the Western Australia-Malaysia Technical Working Group (TWG) meeting that was held on 13th -14th July 2006 in Perth Western Australia, in the areas of technical cooperation between the two countries for feeds and nutrition management, Lupin has been identified as one of potential feed ingredient that can be explored to be used in goat feeding in Malaysia. However, the information on the effects of TMR with different level of lupin inclusion on the performance of goats is still unavailable. Hence, this study aimed to provide information, which can support the strategies to utilize TMR and lupin more efficiently as goat feed in Malaysia. The findings of this study will contribute the knowledge on the usage of TMR and lupin grain in formulating dietary requirement for the Boer goats in order to maximize their genetic potential in the tropical environment.

Hypothesis

By considering the demand of the goat farming sector for good quality feed in developing countries and the necessity of a suitable alternative, it was hypothesized that TMR with *Lupinus angustifolius* (lupin) inclusion can be used as source of protein in animal feed for intensive farming system without causing adverse effects on growth performance, carcass characteristics and meat quality of male Boer goats.

Objective

The current study was carried out to determine the effects of feeding TMR with different levels of *Lupinus angustifolius* (lupin) inclusion on growth performance, carcass characteristics and meat quality of male Boer goats.

Specific Objectives:

- 1. To determine the effects of feeding TMR with different levels of lupin inclusion on the growth performance and carcass characteristics of male Boer goats.
- 2. To investigate the effects of feeding TMR with different levels of lupin inclusion on the meat quality traits of male Boer goats.
- 3. To evaluate the effects of feeding TMR different levels of lupin inclusion on the fatty acid composition in skeletal muscles of male Boer goats.

REFERENCES

- Aberle, E. D., Forrest, J. C., Gerrard, D. E., & Mills, E. W. (2001). Structure and composition of animal tissues. In: *Principles of Meat Science*, 9-116.
- Abuelfatah, K., Zakaria, M. Z. A. B., Meng, G. Y., & Sazili, A. Q. (2014). Changes in Fatty Acid Composition and Distribution of N-3 Fatty Acids in Goat Tissues Fed Different Levels of Whole Linseed. *The Scientific World Journal*, 1–10.
- Agenbag. (2008). Lupin cultivation in the Western and Southern Cape. *Afrikaans*, 79–92.
- Aghwan, Z. a, Alimon, a R., Goh, Y. M., Nakyinsige, K., & Sazili, a Q. (2014). Fatty Acid Profiles of Supraspinatus, Longissimus lumborum and Semitendinosus Muscles and Serum in Kacang Goats Supplemented with Inorganic Selenium and Iodine. Asian-Australasian Journal of Animal Sciences, 27(4), 543–550.
- Ariff, O. M., Hifzan, R. M., Zuki, A. B. M., Jiken, A. J., & Lehan, S. M. (2010). Maturing pattern for body weight, body length and height at withers of Jamnapari and Boer goats. *Pertanika Journal of Tropical Agricultural Science*, 33(2).
- Banskalieva, V., Sahlu, T., & Goetsch, A. L. (2000). Fatty acid composition of goat muscles and fat depots: a review. *Small Ruminant Research*, 37(3), 255–268.
- Ben S. H., & Smith, T. (2008). Feeding strategies to increase small ruminant production in dry environments. *Small Ruminant Research*, 77(2–3), 174–194.
- Beyer, H., Schmalenberg, a. K., Jansen, G., Jorgens, H. U., Uptmoor, R., Broer, I., Ordon, F. (2015). Evaluation of variability, heritability and environmental stability of seed quality and yield parameters of L. angustifolius. *Field Crops Research*, 174, 40–47.
- Bezerra, S. B. L., Véras, A. S. C., de Andrade Silva, D. K., de Andrade Ferreira, M., Pereira, K. P., de Arruda Santos, G. R., Almeida, O. C. (2012). Morphometry and carcass characteristics of goats submitted to grazing in the Caatinga. *Revista Brasileira de Zootecnia*, 41(1), 131–137.
- Boschin, G., D'agostina, A., Annicchiarico, P., Arnoldi, A., 2007. The fatty acid composition on the oil from Lupinus albus cv. Luxe as affected by environmental and agricultural factors. *Eur. Food Res. Technol.* 225, 769–776
- British Department of Health. (1994). Nutritional aspects of cardiovascular diseases.
- Cameron, M. R., Luo, J., Sahlu, T., Hart, S. P., Coleman, S. W., Goetsch, A. L., (2008). Growth and slaughter traits of Boer x Spanish, Boer x Angora, and Spanish goats consuming a concentrate-based diet 1 (2), 1423–1430.
- Casey, N. H., & Niekerk, V., (1988). The Boer Goat . I . Origin , Adaptability , Performance Testing , Reproduction and Milk Production. *Small Ruminant Research*, 1, 291–302.
- Cheetham, N. W. H., Cheung, P. C. K., & Evans, A. J. (1993). Structure of the principal non-starch polysaccharide from the cotyledons of *Lupinus angustifolius* (cultivar Gungurru). *Carbohydrate Polymers*, 22(1), 37–47.
- Clarke, R., Frost, C., Collins, R., Appleby, P. and Peto, R. 1997. Dietary lipids and blood cholesterol: quantitative meta-analysis of metabolic ward studies. *British Medical Journal*, 314: 112 117.
- Cox, R. a, & Garcia-palmieri, M. R. (1990). Cholesterol, Triglycerides, and Associated Lipoproteins. In H. Walker, W. Hall, & J. Hurst (Eds.), *Clinical Methods: The History, Physical, and Laboratory Examinations* (3rd editio, pp. 153–160). Boston: Butterworths.

Department of Veterinary Services. (2016). Perangkaan Ternakan.

- Devendra. C., (2007). Goats: *Biology, Production and Development in Asia*. Academy of Sciences Malaysia
- Devendra, C., & Liang, J. B. (2012). Conference summary of dairy goats in Asia: Current status, multifunctional contribution to food security and potential improvements. *Small Ruminant Research*, 108(1–3), 1–11.
- Dubeuf, J.-P., & Sayadi, S. (2014). Multi-functionality issues for small ruminants: What changes are needed in territorial public policies and training? Report of two round tables on territorial issues and training for the development of goat farming. *Small Ruminant Research*, 121(1), 136–145.
- DVS. (2013). Malaysian Livestock Breeding Policy, 1-42.
- Ebrahimi, M., Rajion, M. A., Goh, Y. M., Sazili, A. Q., & Schonewille, J. T. (2013). Effect of Linseed Oil Dietary Supplementation on Fatty Acid Composition and Gene Expression in Adipose Tissue of Growing Goats, 2013.
- Enser, M., Hallett, K., Hewitt, B., Fursey, G. A. J., &Wood, J. D. (1996). Fatty acid content and composition of English beef, lamb and pork at retail. *Meat Science*, 42, 443–456.
- Ephrem, N., Tegegne, F., Mekuriaw, Y., & Yeheyis, L. (2015). Nutrient intake, digestibility and growth performance of Washera lambs supplemented with graded levels of sweet blue lupin (*Lupinus angustifolius L.*) seed. *Small Ruminant Research*, 130, 101–107.
- Erasmus, J. (2000). Adaptation to various environments and resistance to disease of the Improved Boer goat. *Small Ruminant Research : The Journal of the International Goat Association*, 36(2), 179–187.
- Facciolongo, A. M., Rubino, G., Zarrilli, A., Vicenti, A., Ragni, M., & Toteda, F. (2014). Alternative protein sources in lamb feeding 1. Effects on productive performances, carcass characteristics and energy and protein metabolism. *Progress in Nutrition*, 16(2), 105–115.
- Gellynck, X., Verbeke, W. and Viaene, J. 2004. Quality management in the food supply chain: How does the food industry interact with consumers, retailers and public authorities? In Quality assurance, risk management and environmental control in agriculture and food supply networks, ed. G., Schiefer and U., Rickert, pp. 443 452
- Farrell, D. J., Mannion, P. F., & Perez-Maldonado, R. A. (1999). A comparison of total and digestible amino acids in diets for broilers and layers. *Animal Feed Science* and Technology, 82(1–2), 131–142.
- Fernández, C., & Sánchez, A. (2003). Use of a Total Mixed Ration with Three Sources of Protein as an Alternative Feeding for Dairy Goats on Southeast of Spain, 2(1), 18–24.
- Fletcher, D. L. (2002). Poultry meat quality. *World's Poultry Science Journal*, 58, 131–145.
- Fontanari, G. G., Batistuti, J. P., Cruz, R. J. Da, Saldiva, P. H. N., & Arêas, J. A. G. (2012). Cholesterol-lowering effect of whole lupin (Lupinus albus) seed and its protein isolate. *Food Chemistry*, 132(3), 1521–1526.
- Fychan, R., Marley, C., Lewis, G., Davies, R., Theobald, V., Jones, R., & Abberton, M. (2008a). Effects of feeding concentrate diets containing narrow control diet on the productivity of finishing lambs, , 127–130.
- Fychan, R., Marley, C., Lewis, G., Davies, R., Theobald, V., Jones, R., & Abberton, M. (2008b). Effects of feeding concentrate diets containing narrow-leafed lupin, yellow lupin or soya when compared with a control diet on the productivity of finishing lambs. In "Lupins for Health and Wealth" Proceedings of the 12th

International Lupin Conference (pp. 127–130). 14-18 Sept. 2008, Fremantle, Western Australia: International Lupin Association, Canterbury, New Zealand.

- Givens, D. I., Kliem, K. E., & Gibbs, R. A. (2006). MEAT The role of meat as a source of n À 3 polyunsaturated fatty acids in the human diet, 74, 209–218.
- Glencross, B. D. (2001). Feeding lupins to fish: A review of the nutritional and biological value of lupins in aquaculture feeds.
- Goetsch, A. L., Gipson, T. A., Askar, A. R., Puchala, R., Goetsch, A. L., Gipson, T. A., Puchala, R. (2010). Invited review: Feeding behavior of goats. *Journal of Animal Science*, 88, 361–373.
- Gonzalez, J., & Andres, S. (2003). Rumen degradability of some feed legume seeds. *Animal Research*, 52, 17–25.
- Greter, A. M., Leslie, K. E., Mason, G. J., Mcbride, B. W., & Devries, T. J. (2010). Effect of feed delivery method on the behavior and growth of dairy heifers. *Journal of Dairy Science*, 93(4), 1668–1676.
- Hawthorne, W. (2006). *Pulses nutritional value and their role in the feed industry* (2nd ed.). (W. Hawthorne, Ed.). New South Wales: Pulse Australia Pty Ltd., 10 14.
- Hayes, B. J., Lewin, H. A., & Goddard, and M. E. (2013). The future of livestock breeding: genomic selection for efficiency, reduced emissions intensity, and adaptation. *Trends in Genetics* : TIG, 29(4), 206–214.
- Honikel, K. O. (2004). Water-holding capacity of meat. In M. F. W. Pas, M. E. Everts,
 & H. P. Haagsman (Eds.), *Muscle development of livestock animals: Physiology,* genetics and meat quality (pp. 389–400). Cambridge, USA: CABI Publishing.
- Huidobro, F. R. D., Miguel, E., Blázquez, B., & Onega, E. (2005). A comparison between two methods (Warner-Bratzler and texture profile analysis) for testing either raw meat or cooked meat. *Meat*
- Hunter, R. S., & Harold, R. W. (1987). *The measurement of appearance. A Wiley-Interscience Publication* (2nd edition). John Wiley & Sons.
- Jezierny, D., Mosenthin, R., & Bauer, E. (2010). The use of grain legumes as a protein source in pig nutrition: A review. *Animal Feed Science and Technology*, 157(3–4), 111–128.
- Joo, S. T., Kim, G. D., Hwang, Y. H., & Ryu, Y. C. (2013). Control of fresh meat quality through manipulation of muscle fiber characteristics. *Meat Science*, 95(4), 828– 836.
- Kannan, G., Lee, J. H., & Kouakou, B. (2014). Chevon quality enhancement: Trends in pre- and post-slaughter techniques, *Small Ruminant Research* 121 (2014) 80–88.
- Karami, M., Ponnampalam, E. N., & Hopkins, D. L. (2013). The effect of palm oil or canola oil on feedlot performance, plasma and tissue fatty acid profile and meat quality in goats. *Meat Science*, 94(2), 165–169.
- Karlsson, A., Enfalt, A. C., Essen-Gustavsson, B., Lundstrom, K., Rydhmer, L., & Stern, S. (1993). Muscle histochemical and biochemical properties in relation to meat quality during selection for increased lean tissue growth rate in pigs. *Journal of Animal Science*, 71, 930–938.
- Kaur, B. (2010). Consumer Preference for Goat Meat in Malaysia: Market Oppurtunities and Potential. *Journal of Agribusiness Marketing*, 3, 40–55.
- Kim, J. C., Heo, J. M., Mullan, B. P., & Pluske, J. R. (2012). Performance and intestinal responses to dehulling and inclusion level of Australian sweet lupins (*Lupinus* angustifolius L.) in diets for weaner pigs. Animal Feed Science and Technology, 172(3–4), 201–209.
- Klont, R. E., Brocks, L., & Eikelenboom, G. (1998). Muscle fibre type and meat quality. *Meat Science*, 49, 219–229.

- Koivunen, E., Partanen, K., Perttilä, S., Palander, S., Tuunainen, P., & Valaja, J. (2016). Digestibility and energy value of pea (Pisum sativum L.), faba bean (Vicia faba L.) and blue lupin (narrow-leaf) (Lupinus angustifolius) seeds in broilers. *Animal Feed Science and Technology*, 218, 120–127.
- Kung, L., Maciorowski, K., Powell, K. M., Weidner, S., & Eley, C. L. (1991). Lupin as a protein supplement for growing lambs. *Journal of Animal Science*, 69, 3398– 3405.
- Kwak, B., Kim, H. J., & Park, H. S. (1998). The Effect of Different Lupin Kernel Inclusion Levels on the Growth and Carcass Composition of Growing and Finishing Pigs. Asian-Australasian Journal of Animal Science, 13, 207-212.
- Laaksonen, D. E., Nyyssönen, K., Niskanen, L., Rissanen, T. H., & Salonen, J. T. (2005). Prediction of Cardiovascular Mortality in Middle-aged Men by Dietary and Serum Linoleic and Polyunsaturated Fatty Acids. Archives of Internal Medicine, 165(2), 193–199.
- Lambe, N.R., Ross, D.W., Navajas, E.A., Hyslop, J.J., Prieto, N., Craigie, C., Bunger, L., Simm, G and R. Roehe. 2010. The prediction of Carcass Composition and Tissue Distribution in Beef Cattle Using Ultrasound Scanning at the Start and/or end of the Finishing Period. *Livestock Science*: 131:193-202
- Lawrence, L. (2007). Lupins, Australia's role in world markets. *Australian Commodities* (Vol. 14).
- Lee, M. R. F., Parkinson, S., Fleming, H. R., Theobald, V. J., Leemans, D. K., & Burgess, T. (2016). The potential of blue lupins as a protein source, in the diets of laying hens. *Veterinary and Animal Science*, 1–2(June), 29–35.
- Lestingi, A., Facciolongo, A. M., De Marzo, D., Nicastro, F., & Toteda, F. (2015). The use of faba bean and sweet lupin seeds in fattening lamb feed. 2. Effects on meat quality and fatty acid composition. *Small Ruminant Research*, 131, 2–5.
- Listrat, A., Lebret, B., Louveau, I., Astruc, T., Bonnet, M., Lefaucheur, Bugeon, J. (2016). How muscle structure and composition influence meat and flesh quality. *Scientific World Journal*, 1-14.
- López, E. J., Peraza-Mercado, G., Holguiny, F. M., & Ortiz, M. F. I. (2012). Relationship between live animal weight, warm and cold carcass weight and carcass principal components. *Global Veterinaria*, 9(2), 179–183.
- Malan, S. W. (2000). The improved Boer goat, 36, 165–170.
- Martins, J. M., Riottot, M., de Abreu, M. C., Viegas-Crespo, A. M., Lança, M. J., Almeida, Bento, O. P. (2005). Cholesterol-lowering effects of dietary blue lupin (*Lupinus angustifolius L.*) in intact and ileorectal anastomosed pigs. *Journal of Lipid Research*, 46(7), 1539–1547.
- Mazhangara, I. R., Chivandi, E., & Mupangwa, J. F. (2019). The Potential of Goat Meat in the Red Meat Industry, 1–12.
- Michas, G., Micha, R., & Zampelas, A. (2014). Dietary fats and cardiovascular disease: putting together the pieces of a complicated puzzle. *Atherosclerosis*, 234(2), 320–328.
- Mieczkowska, A., & Smulikowska, S. (2005). The influence of white lupin seeds in diets supplemented with fats of animal or plant origin on the fatty acid composition of broiler tissues. *Journal of Animal and Feed Sciences*, 14(1), 93–107.
- Miller, M. F., Carr, M. F., Ramsey, C. B., Crockett, K. L., & Hoover, L. C. (2001). Consumer thresholds for establishing the value of beef tenderness. *Journal of Animal Science*, 79, 3062-3068.
- MLA (7th Nov. 2018) Malaysian market Retrieved from: https://www.mla.com.au/prices-markets/market-news/in-depth-sheep-and-goatmeat-to-Malaysia/#

- MOA. (2011). *National Agrofood Policy*, NAP 2011-2020. Putrajaya: Unit Penerbitan MOA.
- Mondal, M., & Prakash, B. S. (2004). Effects of long-term GH-releasing factor administration on patterns of GH and LH secretion in growing female buffaloes (Bubalus bubalis). *Reproduction*, 127(1), 45–55.
- Moss, A. R., Givens, D. I., Grundy, H. F., & Wheeler, K. P. a. (1997). The nutritive value for ruminants of lupin seeds from determinate plants and their replacement of soya bean meal in diets for young growing cattle. *Animal Feed Science and Technology*, 68(1–2), 11–23.
- Murphy, S. R., & McNiven, M. A. (1994). Raw or roasted lupin supplementation of grass silage diets for beef steers. *Animal Feed Science and Technology*, 46, 23–35.
- Nissanka N.P.C., Bandara R.M.A.S.and.Disnaka K.G.J.S. (2010) A comparative study on feeding of total mixed ration vs conventional feeding on weight gain in weaned friesian heifers under tropical environment. *The Journal of Agricultural Sciences*, 5(January), 42–51.
- Niekerk, A. V. A. N., & Casey, N. H. (1988). The Boer Goat . II . Growth , Nutrient Requirements , Carcass and Meat Quality, 1, 355–368.
- Nute, G. R., Richardson, R. I., Wood, J. D., Hughes, S. I., Wilkinson, R. G., Cooper, S. L., & Sinclair, L. A. (2007). Effect of dietary oil source on the flavour and the colour and lipid stability of lamb meat. *Meat Science*, 77(4), 547–555.
- Oeckel, M. J. V, Warnants, N., & Boucque, C. V. (1999). Comparison of different methods for measuring water holding capacity and juiciness of pork versus on-line screening methods. *Meat Science*, 51, 313–320.
- Ohlund, I., Hörnell, a, Lind, T., & Hernell, O. (2008). Dietary fat in infancy should be more focused on quality than on quantity. *European Journal of Clinical Nutrition*, 62(9), 1058–1064.
- Offer, G., & Cousins, T. (1992). The mechanism of drip production—Formation of 2 compartments of extracellular-space in muscle post-mortem. *Journal of the Science of Food and Agriculture*, 58, 107–116.
- Owens, F. N., Gill, D. R., Secrist, D. S., Coleman, S. W., Owens, F. N., Gill, D. R., & Secrist, D. S. (1995). Review of some aspects of growth and development of feedlot cattle. *Journal of Animal Science*, 73, 3152–3172.
- Peacock, C., & Sherman, D. M. (2010). Sustainable goat production-Some global perspectives. Small Ruminant Research, 89(2–3), 70–80.
- Petterson, D. (2000). The use of lupins in feeding systems 2000. Asian-Australasian Journal of Animal Sciences, 13(6), 861–882.
- Pi, Z. K., Wu, Y. M., & Liu, J. X. (2005). Effect of pretreatment and pelletization on nutritive value of rice straw-based total mixed ration, and growth performance and meat quality of growing Boer goats fed on TMR. *Small Ruminant Research*, 56(1– 3), 81–88.
- Piedrafita, J., Quintanilla, R., Sanudo, C., Olleta, J. L., Campo, M. M., Panea, B., et al. (2003). Carcass quality of 10 beef cattle breeds of the southwest of Europe in their typical production systems. *Livestock Production Science*, 82, 1–13.
- Pitchford, W. S., Deland, M. P. B., Siebert, B. D., Malau-Aduli, A. E. O., & Bottema, C. D. K. (2002). Genetic variation in fatness and fatty acid composition of crossbred cattle. *Journal of Animal Science*, 80(11), 2825–2832.
- Prandini, A., Morlacchini, M., Moschini, M., Fusconi, G., Masoero, F., Piva, G., Parametri, E. (2005). Raw and extruded pea (Pisum sativum) and lupin seeds as protein sources in weaned piglets' diets: effect on growth rate and blood parameters. *Italian Journal of Animal Science*, 4(4), 385–394.

- Reynolds, K., Chin, A., Lees, K. A., Nguyen, A., Bujnowski, D., & He, J. A. (2006). A meta-analysis of the effect of soy protein supplementation on serum lipids. *American Journal of Cardiology*, 98(5), 633–640.
- Robinson, P. H., & McNiven, M. A. (1993). Nutritive value of raw and roasted sweet white lupins (Lupinus albus) for lactating dairy cows. *Animal Feed Science and Technology*, 43(3–4), 275–290.
- Röös, E., Patel, Spångberg, M., Johanna, Carlsson, Rydhmer, & Lotta, G. (2016). Limiting livestock production to pasture and by-products in a search for sustainable diets. *Food Policy*, 58, 1–13.
- Rosali, M. H., & Azizi, Ahmad Azmin Kamaruddin, M. (2015). Prestasi dan potensi kambing Savanna (Performance and potential of Savanna goat in Malaysia). Buletin Teknologi MARDI, 5(2014), 25–31.
- Salisi, M. S., Saad, M. Z., & Kasim, A. (2012). Implementation of herd health program to improve survival of Boer goats in Malaysia. *Tropical Animal Health and Production*, 44(2), 207–211.
- Salwani, M. S., Sazili, A. Q., Zulkifli, L., Nizam, Z., & Zul Edham, W. (2012). Effects of head-only electrical stunning on the physico-chemical characteristics and desmin degradation of broiler breast muscles at different time postmortem. *Journal* of Animal and Veterinary Advances, 11, 2409-2416.
- Schingoethe, D. J. (2017). A 100-Year Review : Total mixed ration feeding of dairy cows 1. *Journal of Dairy Science*, 100(12), 10143–10150.
- Schmid, A., Collomb, M., Sieber, R. and Bee, G. 2006. Conjugated linoleic acid in meat and meat products: A review. *Meat Science* 73(1): 29 – 41.
- Scott, T. W., Cook, L. J., & Mills, S. C. (1971). Protection of Dietary Polyunsaturated Fatty Acids Against Microbial Hydrogenation in Ruminants. *Journal of the American Oil Chemists' Society*, 48, 358–364.
- Shackelford, S. D., Wheeler, T. L., Meade, M. K., Reagan, J. O., Byrnes, B. L., Shackelford, S. D., ... Koohmaraie, M. (2001). Consumer impressions of Tender Select beef. *Journal of Animal Science*, 79, 2605–2614.
- Shahjalal, M., Galbraith, H., & Topps, J. H. (1992). The effect of changes in dietary protein and energy on growth, body composition and mohair fibre characteristics of British Angora goats. *British Society of Animal Science*, 54(3), 405–412.
- Shahudin, M. S., Ghani, A. A. A., Zamri-Saad, M., Zuki, A. B., Abdullah, F. F. J., Wahid, H., & Hassim, H. A. (2018). The necessity of a herd health management programme for dairy goat farms in Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 41(1), 1–18.
- Shrestha, J. N. B., & Fahmy, M. H. (2007). Breeding goats for meat production. Small Ruminant Research, 67(2–3), 93–112.
- Sirtori, C. R., Lovati, M. R., Manzoni, C., Castiglioni, S., Duranti, M., Magni, C., Arnoldi, A. (2004). Proteins of white lupin seed, a naturally isoflavone-poor legume, reduce cholesterolemia in rats and increase LDL receptor activity in HepG2 cells. *The Journal of Nutrition*, 134(1), 18–23.
- Sivasupramaniam, G. (2006). Goats Undervalued Assets in Asia Proceedings of the Aphca-Ilri Regional Workshop on Goat Production Systems. Presentaion in Asia Proceedings of the Aphca-Ilri Regional Workshop on Goat Production Systems. Luang Prabang, Lao.
- Small, E. (2012). 38. Lupins benefit and harm potentials. *Biodiversity*, 13(1), 54–64.
- Solaiman, S. G. (2010). *Goat science and production* (2010th ed.). Iowa, USA: Wiley-Blackwell Publishing.

- Suchý, P., Straková, E., Kroupa, L., & Vecerek, V. (2008). The fatty acid content of oil from seeds of some lupin varieties. In "Lupins for Health and Wealth" Proceedings of the 12th International Lupin Conference (pp. 188–191).
- Sujak, A., Kotlarz, A., & Strobel, W. (2006). Compositional and nutritional evaluation of several lupin seeds. *Food Chemistry*, 98(4), 711–719.
- Tan. K., L. (2012). Edible Medicinal and Non-Medicinal Plants. Springer.
- Uzun, B., Arslan, C., Karhan, M., & Toker, C. (2007). Fat and fatty acids of white lupin (Lupinus albus L.) in comparison to sesame (Sesamum indicum L.). *Food Chemistry*, 102,45–49.
- Vaclavik, V. A., & Christian, E. W. (2007). Essentials of food science. Springer.
- Van Barneveld, R. J. (1999). Understanding the nutritional chemistry of lupin (Lupinus spp.) seed to improve livestock production efficiency. *Nutrition Research Reviews*, 12(2), 203–230.
- Vicenti, A., Toteda, F., Turi, L. Di, Cocca, C., Perrucci, M., Melodia, L., & Ragni, M. (2009). Use of sweet lupin (Lupinusalbus L. var. Multitalia) in feeding for Podolian young bulls and influence on productive performances and meat quality traits. *Meat Science*, 82(2), 247–251.
- Volek, Z., & Marounek, M. (2011). Effect of feeding growing-fattening rabbits a diet supplemented with whole white lupin (Lupinus albus cv. Amiga) seeds on fatty acid composition and indexes related to human health in hind leg meat and perirenal fat. *Meat Science*, 87(1), 40–45.
- Walker, J., Herthel, K., Parker, P., & Edwards, J. (2011). Lupin growth and development. *Procrop.*
- Wan Zahari, M., & Wong, H. K. (2009). Research and development of animal feed in Malaysia. Wartazoa, 19(4), 171–179.
- Wang, J., Choi, S., & Song, M. (2003). Effects of concentrate to roughage ratio on the formation of cis-9, trans-11 CLA and trans-11 octadecenoic acid in rumen fluid and plasma of sheep when fed high oleic or high linoleic acid oils. *Asian-Australasian Journal of Animal Sciences*, 16(11), 1604–1609.
- Webb, E. C., Casey, N. H., & Simela, L. (2005). Goat meat quality. *Small Ruminant Research*, 60(1–2), 153–166.
- White, C. L., & Staines, V. E. (2007). A review of the nutritional value of lupins for dairy cows. *Australian Journal of Agricultural Research*, 58, 185–202.
- White, P., French, B., & McLarty, A. (2008). *Producing lupins*. (J. Knight, Ed.) (2nd.). Perth, Western Australia: Department of Agriculture and Food, South Perth, Western Australia.
- Wood, J. D., Enser, M., Fisher, A. V., Nute, G. R., Sheard, P. R., Richardson, R. I., Whittington, F. M. (2008). Fat deposition, fatty acid composition and meat quality: A review. *Meat Science*, 78, 343-358.
- Wood, J. D., Richardson, R. I., Nute, G. R., Fisher, a V, Campo, M. M., Kasapidou, E., ... Enser, M. (2004). Effects of fatty acids on meat quality: a review. *Meat Science*, 66(1), 21–32.
- Wyness, L., Weichselbaum, E., O'Connor, a., Williams, E. B., Benelam, B., Riley, H., & Stanner, S. (2011). Red meat in the diet: An update. *Nutrition Bulletin*, 36(1), 34–77.
- Zembayashi, M., Nishimura, K., Lunt, D. K., & Smith, S. B. (1995). Effect of Breed Type and Sex on the Fatty Acid Composition of Subcutaneous and Intramuscular Lipids of Finishing Steers and Heifers. *Journal of Animal Science*, 73, 3325–3332.
- Zembayashi, M., & Nishimura, K. (1996). Genetic and nutritional effects on the fatty acid composition of subcutaneous and intramuscular lipids of steers. *Meat Science* 43, 83-92.

- Zhang, W., Naveena, B. M., Jo, C., Sakata, R., Zhou, G., Banerjee, R., & Nishiumi, T. (2017). Technological demands of meat processing–An Asian perspective. *Meat Science*, 132(April), 35–44.
- Zraly, Z., Pisarikova, B., Trckova, M., Herzig, I., Juzl, M., & Simeonovova, J. (2007). The effect of white lupine on the performance, health,carcass characteristics and meat quality of market pigs. *Veterinarni Medicina*, 52(1), 29–41.

