



UNIVERSITI PUTRA MALAYSIA

***CHARACTERIZATION OF SECRETORY STRUCTURES AND
ESSENTIAL OILS IN AERIAL PARTS OF TORCH GINGER [*Etilingera
elatior* (Jack) R.M. Sm.] AT DIFFERENT DEVELOPMENTAL STAGES***

LEE YEE LING

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By

LEE YEE LING

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

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Sm.] AT DIFFERENT DEVELOPMENTAL STAGES**

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June 2019

Chair : Associate Professor Phebe Ding, PhD
Faculty : Agriculture

Torch ginger (*Etilingera elatior*) is an aromatic plant popularly used as flavouring in food preparation. It has been a subject for many scientific studies aimed at investigating their chemical composition of the essential oils (EO). However, the lack of systematic analysis involving different plant parts and developmental stages may have contributed to variation on the major compounds reported by different groups of researchers. Furthermore, identification on the type of secretory structures responsible for accumulating and storing the EO have not been carried out hitherto. Therefore, this study was carried out to address the current research gaps. First, the presence of secretory cells, glandular and non-glandular trichomes were microscopically identified as the main secretory system for EO of torch ginger. The discovery of glandular trichomes is the first report on the EO secretory structure in Zingiberaceae. The histochemical tests indicated the secretory contents of the secretory structures comprised of terpenes and lipophilic that are typical characteristic of EO.

Second, the variation in chemical composition of aerial parts (leaves, peduncle and inflorescence head) of torch ginger at different developmental stages were analysed using GC-MS. The EO leaves from different position were also sampled repeatedly at 8 and 18 WAE to study the changes in EO content. The EOs in leaves, peduncle and inflorescence head were predominated by 1-dodecanol and *n*-dodecanal as the two major compounds irrespective of developmental stages. However, the α -pinene content influenced markedly on the EO in inflorescence head and peduncle during the initial stage of flowering (10.9 and 21.1%, respectively), which then decreased substantially in the following stages (content ranged between 0.3 to 0.7%; and 0.4 to 4.1%, respectively). Subsequent multivariate analysis using principal component analysis (PCA) was able to distinguish the differences among the EO samples. Most notably, α -pinene was identified as the key compound that characterized

the EO in inflorescence head, while *n*-dodecanal, 2-undecanone and tetradecanoic acid were the key characterizing compounds in the peduncle.

Finally, the antioxidant activities of the EO were evaluated using 1,1-diphenyl-2-picrylhydrazil (DPPH) radical scavenging assay and β -carotene bleaching method. Partial Least Squares (PLS) were employed to correlate the antioxidant activities and the chemical compositions of EO. The leaves at 18 WAE and peduncle at tight bud stage exhibited potent radical scavenging and inhibitory activities. Sesquiterpenes (*(E)*- β -farnesene, α -humulene and β -elemene) and monoterpenes (α -pinene and α -terpineol) were correlated as the active compounds that contributed towards the antioxidant activities. On the other hand, the inflorescence head at full bloom stage was shown to be potent inhibitory activities compared to radical scavenging with (*E*)-caryophyllene, (*E*, *E*)- α -farnesene, *n*-dodecanal and 1-dodecanol correlated as the active compounds. In summary, the findings were able to establish the variation in chemical composition of torch ginger as a function of plant developmental stages and their implication on the antioxidant activities.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**CIRI STRUKTUR REMBESAN DAN MINYAK PATI DALAM BAHAGIAN ATAS
POKOK KANTAN [*Etilingera elatior* (Jack) R.M. Sm.] PADA PERINGKAT
PERTUMBUHAN YANG BERBEZA**

Oleh

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Kantan (*Etilingera elatior*) merupakan pokok beraroma tempatan yang terkenal sebagai penggunaan perasa dalam penyediaan makanan. Ia merupakan subjek penyelidikan saintifik dalam penyiasatan komposisi kimia dalam minyak pati tersebut. Namun demikian, analisis sistematik jarang melibatkan bahagian pokok dan peringkat tumbesaran yang mungkin menyebabkan perbezaan dalam laporan sebatian utama oleh pelbagai pihak saintis. Tambahan pula, identiti jenis struktur perembesan yang bertanggungjawab sebagai tempat pengumpulan dan penyimpanan minyak pati masih belum dikenalpasti sehingga kini. Oleh itu, tujuan kajian ini dijalankan adalah untuk mengatasi jurang penyelidikan semasa. Pertama, kehadiran sel rembesan, trikome berketulan dan tidak berketulan telah dikenalpasti melalui mikroskop sebagai struktur rembesan utama minyak pati dalam pokok kantan. Penemuan trikome berketulan adalah laporan yang pertama sebagai struktur rembesan minyak pati dalam Zingiberaceae. Kajian histokimia menyatakan isi kandungan dalam struktur rembesan tersebut terdiri daripada lipofilik dan terpen yang merupakan ciri tipikal minyak pati.

Kedua, variasi dalam komposisi kimia dari bahagian atas pokok (daun, tangkai pokok dan bunga) kantan pada tahap tumbesaran yang berbeza dikaji dengan menggunakan GC-MS. Minyak pati daun daripada kedudukan yang berbeza juga disampel secara berulang pada minggu 8 dan 18 selepas kemunculan untuk mengkaji perubahan dalam minyak pati. Minyak pati dari daun, tangkai pokok dan bunga terdiri daripada 1-dodekanol dan *n*-dodekanol sebagai sebatian utama dalam komposisi kimia pada semua tahap tumbesaran pokok. Namun demikian, kandungan α -pinen mempengaruhi kandungan minyak pati di dalam bunga dan tangkai pokok semasa tahap pembungaan awal (masing-masing 10.9 dan 21.1%), dan kemudian menurun dengan ketara pada tahap pembungaan seterusnya (kandungan masing-masing antara 0.3 hingga 0.7% dan 0.4 hingga 4.1%). Analisis data multivariat menggunakan prinsipal komponen analisa (PCA) berjaya membezakan kandungan antara sampel minyak pati kantan. α -Pinen dikenalpasti sebagai sebatian utama yang

mencirikan minyak pati dalam bunga, manakala *n*-dodekanal, 2-undekanone and tetradekanoic asid merupakan sebatian pencirian dalam tangkai pokok.

Akhirnya, aktiviti antioksidan daripada minyak pati ditentukan dengan menggunakan kaedah pemerangkapan radikal 1,1-difenil-2-picrylhydrazil (DPPH) dan pelunturan β -karotena. Separa analisis kuasa dua (PLS) digunakan untuk kajian korelasi antara aktiviti antioksidan dengan kandungan komposisi kimia dalam minyak pati. Daun pada 18 minggu selepas kemunculan dan tangkai pokok pada peringkat kudup menunjukkan potensi aktiviti pemusnahan radikal dan perencatan. Sesquiterpen (*(E)*- β -farnesen, α -humulen and β -elemen) dan monoterpen (α -pinen and α -terpineol) menunjukkan korelasi sebagai sebatian aktif yang menyumbang terhadap aktiviti antioksidan. Selain itu, bunga pada tahap pembungaan penuh menunjukkan potensi perencatan berbanding pemerangkapan radikal dimana '*(E)*-caryophyllene', (*E, E*)- α -farnesen, *n*-dodekanal and 1-dodekanol merupakan sebatian aktif. Kesimpulannya, penemuan dalam kajian ini dapat menunjukkan variasi dalam komposisi kimia minyak pati daripada kantan sebagai fungsi peringkat tumbesaran pokok dan implikasi terhadap aktiviti antioksidan.

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I certify that a Thesis Examination Committee has met on 20 June 2019 to conduct the final examination of Lee Yee Ling on her thesis entitled "Characterization of Secretary Structures and Essential Oils in Aerial Parts of Torch Ginger [*Etilingera elatior* (Jack) R.M. Sm.] at Different Developmental Stages" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy).

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LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
BHT	butylated hydroxytoluene
DPPH	1,1-diphenyl-2-picrylhydrazyl
EO	Essential oil
FAA	Formaldehyde-acetic acid-alcohol
FFNSC	Flavour and Fragrance Natural Synthetics and Compounds
GC-MS	Gas chromatography-mass spectrometry
GT	Glandular trichomes
IC ₅₀	Inhibition concentration at 50%
nGT	Non-glandular trichomes
NIST	National Institute of Standards and Technology
PC	Principal component
PCA	Principal component analysis
PLS	Partial least squares
Q ²	Predictive ability
R ²	Goodness of fit
SC	Secretory cells
SD	Standard deviation
SEM	Scanning electron microscope
VIP	Variable Importance in Projection
UV-Vis	Ultraviolet-visible

CHAPTER 1

INTRODUCTION

Essential oils (EO) are natural products made up of complex mixtures of volatile to semi-volatile compounds formed by aromatic plants as secondary metabolites (Zuzarte and Salgueiro 2015). The composition of EO often varies substantially, considering the presence of constituents can be about dozens to potentially hundreds occurring at different concentrations that define the quality of EO. That being said, characterizing the chemical composition of EO pose several challenges due to the inherent factors involved in regulating the production of secondary metabolites in plant (Maietti *et al.* 2013).

First, synthesis and storage of EO normally occurs in specialized secretory tissues in vascular plants (Maffei 2010). The presence of such structures is a typical characteristic of aromatic plant and has been for some time, played a key role in plant taxonomy, particularly trichomes (Behnke 1984; Wagner *et al.* 2004). For example, presence of peltate and capitate glandular trichomes are generally associated with mint family Lamiaceae (Werker *et al.* 1985; Fahn 1988; Ascensão *et al.* 1995) and secretory cavities in family Myrtaceae and Rutaceae. In addition, structural diversity of secretory cells also account for certain chemical class of the secretory products. Studies have demonstrated that peltate glandular trichomes is indeed the major site of monoterpene biosynthesis, particularly in *Mentha* species (Turner *et al.* 2000). Occasionally, the distribution of secretory type within a plant varies substantially and the process of oil accumulation could affect the oil yields and quality (Li *et al.* 2013).

Second, a marked dissimilarity in the composition of EO is strongly associated with plant developmental stages (Figueiredo *et al.* 2008). As plant progresses from juvenile to mature stage, modification in the metabolic system takes place resulting in higher rates of cyclization and dehydration of the compounds (Máñez *et al.* 1991), hence the changes in chemical composition of EO. A classic instance has been demonstrated in developing leaves of peppermint (*Mentha × piperita* L.) where composition of monoterpenes, specifically limonene and menthone, significantly altered with plant age. The two major monoterpene compounds are present in the young leaves of peppermint, but soon decline as plant age advances and menthol becomes the dominant monoterpene constituent (Gershenzon *et al.* 2000).

The metabolic modifications stemmed from the presence of certain biosynthetic precursors that regulate different compounds dominating at certain plant age. Investigation on monoterpene variation in white micromeria (*Micromeria fruticosa*) demonstrates that formation of monoterpene alcohols (i.e. geraniol, citronellol) are probably blocked in young leaves, and the enzymes involved in the formation of the monoterpene alcohols only appear late in leaf development (Dudai *et al.* 2001). Consequently, characterizing the EO chemical composition as a function of developmental stages is imperative given the timeline of plant growth and development also crucially determines the proper time for harvesting raw material, which in this case, the aromatic plant intended for EO production (Sangwan *et al.* 2001; Chalchat and Ozcan 2008; Verma *et al.* 2012).

Third, the plant organ(s) from which EO are derived: green parts (leaves and stems), flowers, bark, wood, fruits, pericarp or seed only, or roots also largely affect the chemical composition of a given EO (Novak *et al.* 2005; Ogunwande *et al.* 2005). The variability is particularly evident and comprehensible given that production of those secondary metabolites are naturally functional oriented clues as part of the plant's mechanism maneuvering towards survival of the fittest. For example, metabolites produced in vegetative parts (leaves and stems) are exclusively associated with plant defence. On the contrary, floral metabolites which are associated with reproductive system are involved in both attractions of pollinators, as well as defence against florivores and pathogens (Muhlemann *et al.* 2014). Hence, variation in chemical composition is expected from which the EO are derived from.

Integration of these three facets in characterizing the chemical composition are lacking especially in torch ginger (*Etilingera elatior*). It is an aromatic herb whose numerous appeals include an ostentatious bloom with attractive ornamental display, along with reputed medicinal properties and nutritions that have been proven scientifically. An extensive review on the literature regarding the EO analysis of torch ginger resulted in an equivocal outcome where variation exists over the reports of compounds predominating in the torch ginger's EO. Clarification accounting for the differences have to be resolved to justify the plant's merit as emerging natural source of chemical and pharmaceutical feedstocks (Abdelwahab *et al.* 2010). Furthermore, the type of secretory structures responsible for accumulation and storage of EO for *Etilingera* have yet to be identified hitherto.

EO possesses wide range of biological activities and antioxidant activities is not only used as tool to understand and estimate their validity as remedies (Ilić *et al.* 2015), but most of the variety groups of volatile compounds have been physicochemically characterized as antioxidant (Guimarães *et al.* 2010). Studies have demonstrated that the attributes of the antioxidant activities are directed toward the chemical profile of EO where the presence of active compounds and their interaction within the EO might contribute towards varying degree of activities.

While the laid out facets on characterization of torch ginger EO profile are built on fundamental basis of plant science, it is hope that this research could provide clarification to account for shortcomings from previous analyses while simultaneously produce resourceful information that can be exploited in the EO production and quality management. Furthermore, new discovery can also be benefited for scientific community.

Objectives

1. To identify the type of secretory structures and examine their distribution at different developmental stages,
2. To elucidate the histochemical content of the secretory structures,
3. To evaluate the chemical composition of essential oils from different plant parts at different developmental stages,
4. To evaluate the antioxidant activities of the extracted essential oils.

REFERENCES

- Abdelwahab SI, Zaman FQ, Mariod AA, Yaacob M, Ahmed H, Khamis S, Ahmed AH, Khamis S. 2010. Chemical composition, antioxidant and antibacterial properties of the essential oils of *Etilingera elatior* and *Cinnamomum pubescens* Kochummen. *Journal of the Science of Food and Agriculture* 90: 2682–2688.
- Abu-Asab MS, Cantino PD. 1987. Phylogenetic implications of leaf anatomy in subtribe *Melittidinae* (Labiatae) and related taxa. *Journal of the Arnold Arboretum*: 1–34.
- Alonso A, Marsal S, Julià A. 2015. Analytical methods in untargeted metabolomics: state of the art in 2015. *Frontiers in Bioengineering and Biotechnology* 3: 23.
- Amiri H. 2010. Antioxidant activity of the essential oil and methanolic extract of *Teucrium orientale* (L.) subsp. *taylori* (Boiss.) Rech. f. *Iranian Journal of Pharmaceutical Research* 9: 417–423.
- Amorati R, Foti MC, Valgimigli L. 2013. Antioxidant activity of essential oils. *Journal of Agricultural and Food Chemistry* 61: 10835–10847.
- Amorati R, Pedulli GF, Cabrini L, Zambonin L, Landi L. 2006. Solvent and pH effects on the antioxidant activity of caffeic and other phenolic acids. *Journal of Agricultural and Food Chemistry* 54: 2932–2937.
- de Andrade Wagner M, Loeuille BFP, Siniscalchi CM, Melo-de-Pinna GF, Pirani JR. 2014. Diversity of non-glandular trichomes in subtribe *Lychnophorinae* (Asteraceae: Vernonieae) and taxonomic implications. *Plant Systematics and Evolution* 300: 1219–1233.
- Andre CM, Hausman J-F, Guerriero G. 2016. *Cannabis sativa*: The plant of the thousand and one molecules. *Frontiers in Plant Science* 7: 19.
- Andreucci AC, Ciccarelli D, Desideri I, Pagni AM. 2008. Glandular hairs and secretory ducts of *Matricaria chamomilla* (Asteraceae): morphology and histochemistry. *Annales Botanici Fennici* 45: 11–18.
- Ascensão L, Marques N, Pais M. 1995. Glandular trichomes on vegetative and reproductive organs of *Leonotis leonurus* (Lamiaceae). *Annals of Botany* 75: 619–626.
- Ascensão L, Mota L, Castro M. 1999. Glandular Trichomes on the leaves and flowers of *Plectranthus ornatus*: Morphology, distribution and histochemistry. *Annals of Botany* 84: 437–447.

- Atkinson R, Arey J. 2003. Gas-phase tropospheric chemistry of biogenic volatile organic compounds: A review. *Atmospheric Environment* 37: 197–219.
- Avanci NC, Luche DD, Goldman GH, Goldman MH. 2010. Jasmonates are phytohormones with multiple functions, including plant defense and reproduction. *Genetics and Molecular Research* 9: 484–505.
- Baldwin IT, Halitschke R, Paschold A, Von Dahl CC, Preston CA. 2006. Volatile signaling in plant-plant interactions: "talking trees" in the genomics era. *Science* 311: 812–815.
- Baschieri A, Daci M, Laure J, Tonfack F, Valgimigli L, Amorati R. 2017. Explaining the antioxidant activity of some common non-phenolic components of essential oils. *Food Chemistry* 232: 656–663.
- Baser KHC, Buchbauer G. 2010. *Handbook of Essential Oils Science, Technology, and Applications*. CRC Press.
- Behnke HD. 1984. Plant trichomes-structure and ultrastructure; general terminology, taxonomic applications, and aspects of trichome-bacteria interaction in leaf tips of *Dioscorea*. Rodriguez, E., Healey, P, L., Mehta, I ed (s). *Biology and chemistry of plant trichomes*. Plenum Press: New York.
- Beretta G, Artali R, Facino RM, Gelmini F. 2011. An analytical and theoretical approach for the profiling of the antioxidant activity of essential oils : The case of *Rosmarinus officinalis* L. *Journal of Pharmaceutical and Biomedical Analysis* 55: 1255–1264.
- Berger RG. 2007. *Flavours and Fragrances: Chemistry, Bioprocessing and Sustainability*. Springer Science & Business Media.
- Bezerra-Silva PC, Dutra KA, Santos GKN, et al. 2016. Evaluation of the activity of the essential oil from an ornamental flower against *Aedes aegypti*: Electrophysiology, molecular dynamics and behavioral assays. *PLoS ONE* 11: 1–15.
- Bicas JL, Neri-Numa IA, Ruiz ALTG, De Carvalho JE, Pastore GM. 2011. Evaluation of the antioxidant and antiproliferative potential of bioflavors. *Food and Chemical Toxicology* 49: 1610–1615.
- Bicchi C, Liberto E, Matteodo M, et al. 2008. Quantitative analysis of essential oils: a complex task. *Flavour and Fragrance Journal* 23: 382–391.
- Boege K. 2005. Influence of plant ontogeny on compensation to leaf damage. *American Journal of Botany* 92: 1632–1640.

- Bombo AB, Appezzato-da-Gloria B, Aschenbrenner AK, Spring O. 2016. Capitulate glandular trichomes in *Aldama discolor* (Heliantheae - Asteraceae): Morphology, metabolite profile and sesquiterpene biosynthesis. *Plant Biology* 18: 455–462.
- Bosabalidis A, Tsekos I. 1982. Ultrastructural studies on the secretory cavities of *Citrus deliciosa* ten. II. Development of the essential oil-accumulating central space of the gland and process of active secretion. *Protoplasma* 112: 63–70.
- Boukhris Maher, Ben Nasri-Ayachi M, Mezghani I, Bouaziz M, Boukhris Makki, Sayadi S. 2013. Trichomes morphology, structure and essential oils of *Pelargonium graveolens* L. (Geraniaceae). *Industrial Crops and Products* 50: 604–610.
- Bouwmeester HJ, Gershenzon J, Konings MCJM, Croteau R. 1998. Biosynthesis of the monoterpenes limonene and carvone in the fruit of caraway - I. Demonstration of enzyme activities and their changes with development. *Plant Physiology* 117: 901–912.
- Brouat C, Mckey D, Bessiere J-M, Pascal L, Hossaert-McKey M. 2000. Leaf volatile compounds and the distribution of ant patrolling in an ant-plant protection mutualism: Preliminary results. *Acta Oecologica* 21: 349–357.
- Burton GW, Ingold KU. 1981. Autoxidation of biological molecules. 1. Antioxidant activity of vitamin E and related chain-breaking phenolic antioxidants in vitro. *Journal of the American Chemical Society* 103: 6472–6477.
- Burton GW, Ingold KU. 1986. Vitamin E: application of the principles of physical organic chemistry to the exploration of its structure and function. *Accounts of Chemical Research* 19: 194–201.
- Caissard JC, Meekijironenroj A, Baudino S, Anstett MC. 2004. Localization of production and emission of pollinator attractant on whole leaves of *Chamaerops humilis* (Arecaceae). *American Journal of Botany* 91: 1190–1199.
- Chalchat JC, Ozcan MM. 2008. Comparative essential oil composition of flowers, leaves and stems of basil (*Ocimum basilicum* L.) used as herb. *Food Chemistry* 110: 501–503.
- Chan E, Chiang WEI. 2009. Bioactivities and chemical constituents of leaves of some *Etilingera* Species (Zingiberaceae) in Peninsular Malaysia.: 1–305.
- Chan E, Lim Y, Omar M. 2007. Antioxidant and antibacterial activity of leaves

- of *Etilingera* species (Zingiberaceae) in Peninsular Malaysia. *Food Chemistry* 104: 1586–1593.
- Chan EWC, Lim YY, Wong LF, *et al.* 2008. Antioxidant and tyrosinase inhibition properties of leaves and rhizomes of ginger species. *Food Chemistry* 109: 477–483.
- Chan EW, Ng VP, Tan VV, Low YY. 2011. Antioxidant and antibacterial properties of *Alpinia galanga*, *Curcuma longa*, and *Etilingera elatior* (Zingiberaceae). *Pharmacognosy Journal* 3: 54–61.
- Chau FT, Chan HY, Cheung CY, Xu CJ, Liang Y, Kvalheim OM. 2009. Recipe for uncovering the bioactive components in herbal medicine. *Analytical Chemistry* 81: 7217–7225.
- Chen F, Tholl D, D'Auria JC, Farooq A, Pichersky E, Gershenzon J. 2003. Biosynthesis and emission of terpenoid volatiles from *Arabidopsis* flowers. *The Plant Cell* 15: 481–494.
- Chen Y, Wu YG, Xu Y, *et al.* 2014. Dynamic accumulation of sesquiterpenes in essential oil of *Pogostemon cablin*. *Brazilian Journal of Pharmacognosy* 24: 626–634.
- Choi H-K, Yoon J-H, Kim Y-S, Kwon DY. 2006. Metabolomic profiling of Cheonggukjang during fermentation by ¹H NMR spectrometry and principal components analysis. *Process Biochemistry* 42: 263–266.
- Chong HW, Rezaei K, Chew BL, Lim V. 2018. Chemometric profiling of *Clinacanthus nutans* leaves possessing antioxidant activities using Ultraviolet visible Spectrophotometry. *Chiang Mai Journal of Science* 45: 1377–1388.
- Choon SY, Ding P. 2016. Growth stages of torch ginger (*Etilingera elatior*) Plant. *Sains Malaysiana* 45: 507–515.
- Choon SY, Ding P, Mahmud TMM, Shaari K. 2016. Phenological growth stages of torch ginger (*Etilingera elatior*) inflorescence. *Pertanika Journal Tropical Agricultural Science* 39: 73–78.
- Ciccarelli D, Garbari F, Pagni AM. 2008. The flower of *Myrtus communis* (Myrtaceae): Secretory structures, unicellular papillae, and their ecological role. *Flora* 203: 85–93.
- Colby SM, Alonso WR, Katahira EJ, McGarvey DJ, Croteau R. 1993. 4S-limonene synthase from the oil glands of spearmint (*Mentha spicata*). cDNA isolation, characterization, and bacterial expression of the catalytically active monoterpene cyclase. *Journal of Biological Chemistry* 268: 23016–23024.

- Cordella CBY. 2012. PCA: The basic building block of chemometrics. *Analytical Chemistry*.1–46.
- Corsi G, Bottega S. 1999. Glandular hairs of *Salvia officinalis*: new data on morphology, localization and histochemistry in relation to function. *Annals of Botany* 84: 657–664.
- Croteau R, Karp F, Wagschal KC, Satterwhite DM, Hyatt DC, Skotland CB. 1991. Biochemical characterization of a spearmint mutant that resembles peppermint in monoterpene content. *Plant physiology* 96: 744–52.
- Croteau R, Kutchan TM, Lewis NG. 2000. Chapter 24: Natural Products (Secondary Metabolites) In: *Biochemistry & Molecular Biology of Plants*.1250–1318.
- Croteau R, Venkatachalam K V. 1986. Metabolism of monoterpenes: Demonstration that (+)-cis-isopulegone, not piperitenone, is the key intermediate in the conversion of (-)-isopiperitenone to (+)-pulegone in peppermint (*Mentha piperita*). *Archives of Biochemistry and Biophysics* 249: 306–315.
- Crozier A, Clifford MN. 2006. *Terpenes, Plant Secondary Metabolites*. John Wiley & Sons.
- Dai X, Wang G, Yang DS, *et al.* 2010. TrichOME: A comparative omics database for plant trichomes. *Plant Physiology* 152: 44–54.
- Dalin P, Ågren J, Bjorkman C, Huttunen P, Karkkainen K. 2008. Leaf trichome formation and plant resistance to herbivory In: *Induced Plant Resistance to Herbivory*.89–105.
- Davies PJ, Gan S. 2012. Towards an integrated view of monocarpic plant senescence. *Russian Journal of Plant Physiology* 59: 467–478.
- Dawidowicz AL, Olszowy M. 2014. Does antioxidant properties of the main component of essential oil reflect its antioxidant properties? The comparison of antioxidant properties of essential oils and their main components. *Natural Product Research* 28: 1952–1963.
- de Sousa Galvão M, Narain N, Do Socorro Porto dos Santos M, Nunes ML. 2011. Volatile compounds and descriptive odor attributes in umbu (*Spondias tuberosa*) fruits during maturation. *Food Research International* 44: 1919–1926.
- Demarco D. 2017. Floral glands in *Asclepiads*: structure, diversity and evolution. *Acta Botanica Brasílica* 31: 477–502.

- Deschamps C, Gang D, Dudareva N, Simon JE. 2006. Developmental regulation of phenylpropanoid biosynthesis in leaves and glandular trichomes of basil (*Ocimum basilicum* L.). *International Journal of Plant Sciences* 167: 447–454.
- Desurmont GA, Laplanche D, Schiestl FP, Turlings TCJ. 2015. Floral volatiles interfere with plant attraction of parasitoids: ontogeny-dependent infochemical dynamics in *Brassica rapa*. *BMC Ecology*: 1–11.
- Diezel C, Allmann S, Baldwin IT. 2011. Mechanisms of optimal defense patterns in *Nicotiana attenuata*: Flowering attenuates herbivory-elicited ethylene and jasmonate signaling. *Journal of Integrative Plant Biology* 53: 971–983.
- Dorman HJ, Deans SG. 2000. Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *Journal of Applied Microbiology* 88: 308–16.
- Dubey VS, Luthra R. 2001. Biotransformation of geranyl acetate to geraniol during palmarosa (*Cymbopogon martinii*, Roxb. wats. var. motia) inflorescence development. *Phytochemistry* 57: 675–680.
- Dudai N, Larkov O, Ravid U, Putievsky E, Lewinsohn E. 2001. Developmental control of monoterpene content and composition in *Micromeria fruticosa* (L.) Druce. *Annals of Botany* 88: 349–354.
- Dudareva N, Klempien A, Muhlemann JK, Kaplan I. 2013. Biosynthesis, function and metabolic engineering of plant volatile organic compounds. *New Phytologist* 198: 16–32.
- Dudareva N, Negre F, Nagegowda DA, Orlova I. 2006. Plant volatiles: recent advances and future perspectives. *Critical Reviews in Plant Sciences* 25: 417–440.
- Dudareva N, Pichersky E. 2000. Biochemical and molecular genetic aspects of floral scents. *Plant Physiology* 122: 627–633.
- Dudareva N, Pichersky E, Gershenzon J. 2004. Biochemistry of plant volatiles. *Plant Physiology* 135: 1893–1902.
- Eliasson M, Rannar S, Trygg J. 2011. From data processing to multivariate validation - essential steps in extracting interpretable information from metabolomics data. *Current Pharmaceutical Biotechnology* 12: 996–1004.
- Emami SA, Javadi B, Hassanzadeh MK. 2007. Antioxidant activity of the essential oils of different parts of *Juniperus communis* subsp. *hemisphaerica* and *Juniperus oblonga*. *Pharmaceutical Biology* 45:

769–776.

- Estiarte M, Penuelas J. 1999. Excess carbon: the relationship with phenotypical plasticity in storage and defense functions of plants. *Orsis* 14: 159–203.
- Evert RF. 2006. *Esau's Plant Anatomy: meristem, cells, and tissues of the plant body: their structure, function, and development*. Hoboken, New Jersey: John Wiley & Sons.
- Fahn A. 1988. Secretory tissues in vascular plants. *New Phytologist* 108: 229–257.
- Ferguson DK. 2009. *Anatomy of Flowering Plants: An Introduction to Structure and Development (3rd edition)*.
- Figueiredo AC, Barroso JG, Pedro LG, et al. 2008. Factors affecting secondary metabolite production in plants: volatile components and essential oils. *Flavour and Fragrance Journal* 23: 213–226.
- Fischer R, Nitzan N, Chaimovitch D, Rubin B, Dudai N. 2011. Variation in essential oil composition within individual leaves of sweet basil (*Ocimum basilicum* L.) is more affected by leaf position than by leaf age. *Journal Agricultural and Food Chemistry* 59: 4913–22.
- Foti MC, Ingold KU. 2003. Mechanism of inhibition of lipid peroxidation by gamma-terpinene, an unusual and potentially useful hydrocarbon antioxidant. *Journal of Agricultural and Food Chemistry* 51: 2758–65.
- Fujimura Y, Kurihara K, Ida M, et al. 2011. Metabolomics-driven nutraceutical evaluation of diverse green tea cultivars. *PLoS ONE* 6.
- Fujita Y, Koeduka T, Aida M, Suzuki H, Iijima Y, Matsui K. 2017. Biosynthesis of volatile terpenes that accumulate in the secretory cavities of young leaves of Japanese pepper (*Zanthoxylum piperitum*): Isolation and functional characterization of monoterpene and sesquiterpene synthase genes. *Plant Biotechnology* 34: 17–28.
- Gahan PB. 1984. *Plant histochemistry and cytochemistry*. Academic Press.
- Gang DR, Wang J, Dudareva N, et al. 2001. An investigation of the storage and biosynthesis of phenylpropenes in sweet basil. *Plant Physiology* 125: 539–555.
- Ganjewala D, Luthra R. 2009. Geranyl acetate esterase controls and regulates the level of geraniol in lemongrass (*Cymbopogon flexuosus* nees ex steud.) mutant cv. GRL-1 leaves. *Zeitschrift fur Naturforschung - Section C Journal of Biosciences* 64: 251–259.

- Gatsuk LE, Smirnova O V, Vorontzova LI, Zaugolnova LB, Zhukova LA. 1980. Age states of plants of various growth forms: a review. *The Journal of Ecology*: 675–696.
- Geng SL, Cui ZX, Shu B, Zhao S, Yu XH. 2012. Histochemistry and cell wall specialization of oil cells related to the essential oil accumulation in the bark of *Cinnamomum cassia* Presl. (Lauraceae). *Plant Production Science* 15: 1–9.
- Gersbach PV, Wyllie SG, Sarafis V. 2001. A new histochemical method for localization of the site of monoterpene phenol accumulation in plant secretory structures. *Annals of Botany* 88: 521–525.
- Gershenzon J, Maffei M, Croteau R. 1989. Biochemical and histochemical localization of monoterpene biosynthesis in the glandular trichomes of spearmint (*Mentha spicata*). *Plant Physiology*, 89: 1351–1357
- Gershenzon J. 1994. Metabolic costs of terpenoid accumulation in higher plants. *Journal of Chemical Ecology* 20: 1281–1328.
- Gershenzon J, Dudareva N. 2007. The function of terpene natural products in the natural world. *Nature Chemical Biology* 3: 408–414.
- Gershenzon J, McConkey ME, Croteau RB. 2000. Regulation of monoterpene accumulation in leaves of peppermint. *Plant Physiology* 122: 205–214.
- Glas JJ, Schimmel BCJ, Alba JM, Escobar-Bravo RR, Schuurink RC, Kant MR. 2012. Plant glandular trichomes as targets for breeding or engineering of resistance to herbivores. *International Journal of Molecular Sciences* 13: 17077–17103.
- Gonthier M-P, Verny M-A, Besson C, Rémésy C, Scalbert A. 2003. Chlorogenic acid bioavailability largely depends on its metabolism by the gut microflora in rats. *The Journal of nutrition* 133: 1853–1859.
- González WL, Negritto MA, Suárez LH, Gianoli E. 2008. Induction of glandular and non-glandular trichomes by damage in leaves of *Madia sativa* under contrasting water regimes. *Acta Oecologica* 33: 128–132.
- Goodger JQD, Senaratne SL, Nicolle D, Woodrow IE. 2018. Differential metabolic specialization of foliar oil glands in *Eucalyptus brevistylis* Brooker (Myrtaceae). *Tree physiology* 38: 1451–1460.
- Griffin SG, Wyllie SG, Markham JL, Leach DN. 1999. The role of structure and molecular properties of terpenoids in determining their antimicrobial activity. *Flavour and Fragrance Journal* 14: 322–332.

- Guimarães R, Sousa MJ, Ferreira ICFR. 2010. Contribution of essential oils and phenolics to the antioxidant properties of aromatic plants. *Industrial Crops and Products* 32: 152–156.
- Gupta AK, Ganjewala D. 2015. A study on developmental changes in essential oil content and composition in *Cymbopogon flexuosus* cultivar Suvarna. *Acta Biologica Szegediensis* 59: 119–125.
- Haleagrahara N, Jackie T, Chakravarthi S, Rao M, Kulur A. 2010. Protective effect of *Etlingera elatior* (torch ginger) extract on lead acetate--induced hepatotoxicity in rats. *The Journal of toxicological sciences* 35: 663–671.
- Hanhineva K, Rogachev I, Kokko H, *et al.* 2008. Non-targeted analysis of spatial metabolite composition in strawberry (*Fragaria x ananassa*) flowers. *Phytochemistry* 69: 2463–2481.
- Holopainen JK. 2004. Multiple functions of inducible plant volatiles. *Trends in Plant Science* 9: 529–533.
- Hu ML, Wang YQ, Bai M, Wang YL, Wu H. 2015. Variations in volatile oil yields and compositions of *Magnolia zenii* Cheng flower buds at different growth stages. *Trees* 29: 1649–1660.
- Huang M, Sanchez-Moreiras AM, Abel C, *et al.* 2012. The major volatile organic compound emitted from *Arabidopsis thaliana* flowers, the sesquiterpene (E)- β -caryophyllene, is a defense against a bacterial pathogen. *New Phytologist* 193: 997–1008.
- Huchelmann A, Boutry M, Hachez C. 2017. Plant glandular trichoms: natural cell factories of high biotechnological interest. *Plant Physiology* 175: 6–22.
- Hüsni K, Bacser C, Demirci F, *et al.* 2007. Chemistry of Essential Oils In: Berger RG, ed. *Flavours and Fragrances: Chemistry, Bioprocessing and Sustainability*. Berlin, Heidelberg: Springer Berlin Heidelberg, 43–86.
- Hussain a, Anwar F, Hussain Sherazi ST, Przybylski R. 2008. Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*) essential oils depends on seasonal variations. *Food Chemistry* 108: 986–995.
- Ibanez S, Dötterl S, Anstett MC, Baudino S, Caissard JC, Gallet C, Després L. 2010. The role of volatile organic compounds, morphology and pigments of globe-flowers in the attraction of their specific pollinating flies. *New Phytologist* 188: 451–463.

- Ibrahim H, Setyowati FM. 1999. *Etilingera*. *Plant resources of Southeast Asia* 13: 123–126.
- Ichie T, Inoue Y, Takahashi N, Kamiya K, Kenzo T. 2016. Ecological distribution of leaf stomata and trichomes among tree species in a Malaysian lowland tropical rain forest. *Journal of Plant Research* 129: 625–635.
- Ilić MD, Jovanović VPS, Mitić VD, Jovanović OP, Mihajilov-Krstev TM, Marković MS, Stojanović GS. 2015. Comparison of chemical composition and biological activities of *Seseli rigidum* fruit essential oils from Serbia. *Open Chemistry* 13: 42–51.
- Isman MB. 2000. Plant essential oils for pest and disease management. *Crop Protection* 19: 603–608.
- Isman MB, Miresmailli S, MacHial C. 2011. Commercial opportunities for pesticides based on plant essential oils in agriculture, industry and consumer products. *Phytochemistry Reviews* 10: 197–204.
- Jaafar FM, Osman CP, Ismail NH, Awang K. 2007. Analysis of essential oils of leaves, stems, flowers and rhizomes of *Etilingera elatior* (Jack) R. M. Smith. *The Malaysian Journal of Analytical Sciences* 11: 269–273.
- Janošević D, Budimir S, Alimpić A, Marin P, Al Sheef N, Giweli A, Duletić-Laušević S. 2016. Micromorphology and histochemistry of leaf trichomes of *Salvia aegyptiaca* (Lamiaceae). *Archives of Biological Science* 68: 291–301.
- Janta P, Kulsing C, Nhujak T. 2018. Characterization of volatile compounds in Tom Yum soup by headspace-solid phase microextraction-gas chromatography-mass spectrometry combined with sensory evaluation techniques. *Preprints*.
- Jensen WAWA. 1962. *Botanical histochemistry: principles and practice*. WH Freeman.
- Jezler CN, Batista RS, Alves PB, Silva DC, Costa LCB. 2013. Histochemistry, content and chemical composition of essential oil in different organs of *Alpinia zerumbet*. *Ciência Rural* 43: 1811–1816.
- Jia S, Wang Y, Hu J, *et al.* 2016. Mineral and metabolic profiles in tea leaves and flowers during flower development. *Plant Physiology and Biochemistry* 106: 316–326.
- Johansen DA. 1940. *Plant Microtechnique*. New York: McGraw-Hill.
- Judzentiene A, Budiene J, Butkiene R, Kupcinskiene E, Laffont-Schwob I,

- Masotti V. 2018. Caryophyllene oxide-rich essential oils of Lithuanian *Artemisia campestris* ssp. *campestris* and their toxicity. *Natural Product Communications* 5: 1981–1984.
- Kalogeropoulou A. 2011. *Pre-Processing And Analysis Of High-Dimensional Plant Metabolomics Data*. Thesis.
- Kamali H, Golmakani E, Golshan A, Mohammadi A, Sani TA. 2014. Optimization of ethanol modified supercritical carbon dioxide on the extract yield and antioxidant activity from *Biebersteinia multifida* DC. *Journal of Supercritical Fluids* 91: 46–52.
- Kang JH, Shi F, Jones AD, Marks MD, Howe GA. 2010. Distortion of trichome morphology by the hairless mutation of tomato affects leaf surface chemistry. *Journal of Experimental Botany* 61: 1053–1064.
- Karp F, Mihaliak CA, Harris JL, Croteau R. 1990. Monoterpene biosynthesis: Specificity of the hydroxylations of (-)-limonene by enzyme preparations from peppermint (*Mentha piperita*), spearmint (*Mentha spicata*), and perilla (*Perilla frutescens*) leaves. *Archives of Biochemistry and Biophysics* 276: 219–226.
- Kedare SB, Singh RP. 2011. Genesis and development of DPPH method of antioxidant assay. *Journal of Food Science and Technology* 48: 412–422.
- Keeling CI, Bohlmann J. 2006. Diterpene resin acids in conifers. *Phytochemistry* 67: 2415–2423.
- Kessler A, Baldwin IT. 2000. Defensive function of herbivore-induced plant volatile emissions in nature. *Proceedings of the National Academy of Sciences of the United States of America*. *Plant Cell* 41: 415–50.
- Khaleel C, Tabanca N, Buchbauer G. 2018. α -Terpineol, a natural monoterpene: A review of its biological properties. *Open Chemistry* 16: 349–361.
- Khanom MM, Ueda Y. 2008. Bioconversion of aliphatic and aromatic alcohols to their corresponding esters in melons (*Cucumis melo* L. cv. Prince melon and cv. Earl's favorite melon). *Postharvest Biology and Technology* 50: 18–24.
- Khaw SH. 2001. The genus *Etilingera* (Zingiberaceae) in Peninsular Malaysia including a new species. *Gardens' Bulletin Singapore* 53: 191–239.
- Kikuzawa K, Ackerly D. 1999. Significance of leaf longevity in plants. *Plant Species Biology* 14: 39–45.

- Kjonaas R, Martinkus-Taylor C, Croteau R. 1982. Metabolism of monoterpenes: conversion of l-menthone to l-menthol and d-menthol by stereospecific dehydrogenases from peppermint (*Mentha piperita*) Leaves. *Plant Physiology* 69: 1013–7.
- Kjonaas RB, Venkatachalam K V., Croteau R. 1985. Metabolism of monoterpenes: oxidation of Isopiperitenol to and subsequent Isomerization to piperitenone by soluble enzyme preparations from peppermint (*Mentha piperita*) Leaves. *Archives of Biochemistry and Biophysics* 238: 49–60.
- Kleingesinds CK, Gobara BNK, Mancilha D, Rodrigues MA, Demarco D, Mercier H. 2018. Impact of tank formation on distribution and cellular organization of trichomes within *Guzmania monostachia* rosette. *Flora*.
- Knudsen JT, Eriksson R, Gershenzon J, Ståhl B. 2006. Diversity and distribution of floral scent. *The Botanical Review* 72: 1–120.
- Koeduka T. 2014. The phenylpropene synthase pathway and its applications in the engineering of volatile phenylpropanoids in plants. *Plant Biotechnology* 31: 401–407.
- Kolosova N, Sherman D, Karlson D, Dudareva N. 2001. Cellular and subcellular localization of S-adenosyl-L-methionine:benzoic acid carboxyl methyltransferase, the enzyme responsible for biosynthesis of the volatile ester methylbenzoate in snapdragon flowers. *Plant Physiology* 126: 956–964.
- Kooke R, Keurentjes JJB. 2012. Multi-dimensional regulation of metabolic networks shaping plant development and performance. *Journal of Experimental Botany* 63: 3353–3365.
- Koroch AR, Rodolfo Juliani H, Zygadlo JA. 2007. Bioactivity of Essential Oils and Their Components In: Berger RG, ed. *Flavours and Fragrances: Chemistry, Bioprocessing and Sustainability*. Berlin, Heidelberg: Springer Berlin Heidelberg, 87–115.
- Kramer PJ, Kozlowski TT. 1979. Physiology of woody plants. New York: Academic Press.
- Krauß S, Vetter W. 2018. Phytol and phytol fatty acid esters: occurrence, concentrations, and relevance. *European Journal of Lipid Science and Technology* 120.
- Kreutzmann S, Svensson VT, Thybo AK, Bro R, Petersen MA. 2008. Prediction of sensory quality in raw carrots (*Daucus carota* L.) using multi-block LS-ParPLS. *Food Quality and Preference* 19: 609–617.

- Lachumy SJT, Sasidharan S, Sumathy V, Zuraini Z. 2010. Pharmacological activity, phytochemical analysis and toxicity of methanol extract of *Etilingera elatior* (torch ginger) flowers. *Asian Pacific Journal of Tropical Medicine* 3: 769–774.
- Lange BM. 2015. The evolution of plant secretory structures and the emergence of terpenoid chemical diversity. *Annual Reviews of Plant Biology* 66: 1–21.
- Lange BM, Turner GW. 2013. Terpenoid biosynthesis in trichomes-current status and future opportunities. *Plant Biotechnology Journal* 11: 2–22.
- Larayetan RA, Okoh OO, Sadimenko A, Okoh AI. 2017. Terpene constituents of the aerial parts, phenolic content, antibacterial potential, free radical scavenging and antioxidant activity of *Callistemon citrinus* (Curtis) Skeels (Myrtaceae) from Eastern Cape Province of South Africa. *BMC Complementary and Alternative Medicine* 17: 1–9.
- Larsen K, Ibrahim H, Khaw SH, Saw LG. 1999. *Gingers of peninsular Malaysia and Singapore*. Natural History Publications (Borneo).
- Laurus L. 1979. Ultrastructure and development of oil cells in. *Botanical Journal of the Linnean Society* 78: 81–40.
- Law SR, Chrobok D, Juvany M, Delhomme N, Linden P, Brouwer B, Ahad A, Moritz T, Jansson S, Garderstrom P, Keech O. 2018. Darkened leaves use different metabolic strategies for senescence and survival. *Plant Physiology* 177: 132-150.
- Lerdau M, Litvak M, Palmer P, Monson R. 1997. Controls over monoterpene emissions from boreal forest conifers. *Tree Physiology* 17: 563--569.
- Lewinsohn E, Ziv-raz I, Dudai N, Tadmor Y, Lastochkin E, Larkov O, Chaimovitch, Ravid U, Putievsky E, Pichersky E, Shoham Y. 2000. Biosynthesis of estragole and methyl-eugenol in sweet basil (*Ocimum basilicum* L). Developmental and chemotypic association of allylphenol O-methyltransferase activities. *Plant Science* 160: 27–35.
- Li Y, Kong D, Huang R, Liang H, Xu C, Wu H. 2013. Variations in essential oil yields and compositions of *Cinnamomum cassia* leaves at different developmental stages. *Industrial Crops and Products* 47: 92–101.
- Li W, Zhang H, Li X, *et al.* 2017. Intergrative metabolomic and transcriptomic analyses unveil nutrient remobilization events in leaf senescence of tobacco. *Scientific Reports* 7: 1–17.
- Liber Z, Carović-Stanko K, Politeo O, *et al.* 2011. Chemical characterization and genetic relationships among *Ocimum basilicum* L. cultivars.

Chemistry & Biodiversity 8: 1978–1989.

- Lim TK. 2014. *Etilingera elatior* In: *Edible Medicinal and Non Medicinal Plants*.834–843.
- Łotocka B, Geszprych A. 2004. Anatomy of the vegetative organs and secretory structures of *Rhaponticum carthamoides* (Asteraceae). *Botanical Journal of the Linnean Society* 144: 207–233.
- Ludewig F, Flügge U-I. 2013. Role of metabolite transporters in source-sink carbon allocation. *Frontiers in Plant Science* 4: 1–16.
- Lv SD, Wu YS, Song YZ, et al. 2014. Multivariate analysis based on GC-MS fingerprint and volatile composition for the quality evaluation of Pu-Erh green Tea. *Food Analytical Methods* 8: 321–333.
- Machado SR, Gregório EA, Guimarães E. 2006. Ovary peltate trichomes of *Zeyheria montana* (Bignoniaceae): Developmental ultrastructure and secretion in relation to function. *Annals of Botany* 97: 357–369.
- Machado SR, Souza CV de, Guimarães and E. 2016. A reduced, yet functional, nectary disk integrates a complex system of floral nectar secretion in the genus *Zeyheria* (Bignoniaceae). *Acta Botanica Brasílica* 31: 344-357.
- Madala NE, Piater LA, Steenkamp PA, Dubery IA. 2014. Multivariate statistical models of metabolomic data reveals different metabolite distribution patterns in isonitrosoacetophenone-elicited *Nicotiana tabacum* and *Sorghum bicolor* cells. *SpringerPlus* 3: 1–10.
- Maffei ME. 2010. Sites of synthesis, biochemistry and functional role of plant volatiles. *South African Journal of Botany* 76: 612–631.
- Maggi F, Papa F, Cristalli G, Sagratini G, Vittori S, Giuliani C. 2010. Histochemical localization of secretion and composition of the essential oil in *Melittis melissophyllum* L. subsp. *melissophyllum* from Central Italy. *Flavour and Fragrance Journal* 25: 63–70.
- Mahmoud SS, Croteau RB. 2001. Metabolic engineering of essential oil yield and composition in mint by altering expression of deoxyxylulose phosphate reductoisomerase and menthofuran synthase. *Proceedings of the National Academy of Sciences* 98: 8915–8920.
- Mahmoud SS, Croteau RB. 2002. Strategies for transgenic manipulation of monoterpene biosynthesis in plants. *Trends in Plant Science* 7: 366–373.
- Maietti S, Rossi D, Guerrini A, Useli C, Romagnoli C, Poli F, Bruni R,

- Sacchetti G. 2013. A multivariate analysis approach to the study of chemical and functional properties of chemo-diverse plant derivatives: lavender essential oils. *Flavour and Fragrance Journal* 28: 144–154.
- Máñez S, Jiménez A, Villar A. 1991. Volatiles of *Sideritis mugronensis* flower and leaf. *Journal of Essential Oil Research* 3: 395–397.
- Marcati CR, Angyalossy-alfonso V, Benetati L. 2001. Anatomia comparada do lenho de *Copaifera langsdorffii* Desf. (Leguminosae-Caesalpinoideae) de floresta e cerrado. *Revista Brasileira de Botanica* 24: 311–320.
- Maree J, Kamatou G, Gibbons S, Viljoen A, Van Vuuren S. 2014. The application of GC-MS combined with chemometrics for the identification of antimicrobial compounds from selected commercial essential oils. *Chemometrics and Intelligent Laboratory Systems* 130: 172–181.
- Marinho CR, Zacaro AA, Ventrella MC. 2011. Secretory cells in *Piper umbellatum* (Piperaceae) leaves: A new example for the development of idioblasts. *Flora* 206: 1052–1062.
- Maróstica MR, Rocha TAA, Franchi GC, Nowill A, Pastore GM, Hyslop S. 2009. Antioxidant potential of aroma compounds obtained by limonene biotransformation of orange essential oil. *Food Chemistry* 116: 8–12.
- Mayekiso B, Mhinana Z, Magwa ML. 2009. The structure and function of trichomes in the leaf of *Salvia repens* Burch. Ex Benth. *African Journal of Plant Science* 3: 190–199.
- McCall AC, Fordyce JA. 2010. Can optimal defence theory be used to predict the distribution of plant chemical defences? *Journal of Ecology* 98: 985–992.
- McCaskill D, Gershenzon J, Croteau R. 1992. Morphology and monoterpene biosynthetic capabilities of secretory cell clusters isolated from glandular trichomes of peppermint (*Mentha piperita* L.). *Planta* 187: 445–454.
- McConkey ME, Gershenzon J, Croteau RB. 2000. Developmental regulation of monoterpene biosynthesis in the glandular trichomes of peppermint. *Plant Physiology* 122: 215–24.
- Milani JF, Rocha JF, de Pádua Teixeira S. 2012. Oleoresin glands in copaíba (*Copaifera trapezifolia* Hayne: Leguminosae), a Brazilian rainforest tree. *Trees - Structure and Function* 26: 769–775.
- Mithöfer A, Boland W. 2012. Plant defense against herbivores: chemical

aspects. *Annual Reviews of Plant Biology* 63: 431–450.

- Mohamad H, Ali AM, Lajis NH, Sukari MA, Yap YH, Kikuzaki H, Nakatani N. 2005. Antitumour-promoting and cytotoxic constituents of *Etligeria elatior*. *Malaysian Journal of Medical Sciences* 12: 6–12.
- Monzote L, Stamberg W, Staniek K, Gille L. 2009. Toxic effects of carvacrol, caryophyllene oxide, and ascaridole from essential oil of *Chenopodium ambrosioides* on mitochondria. *Toxicology and Applied Pharmacology* 240: 337–347.
- Moraes CM De, Mescher MC, Tumlinson JH. 2001. Caterpillar-induced nocturnal plant volatiles repel conspecific females. *Nature* 410: 577–580.
- Morrow GW. 2016. *Bioorganic Synthesis: An Introduction*. Oxford University Press.
- Morsy NFS. 2017. Chemical structure, quality indices and bioactivity of essential oil constituents In: *Active Ingredients from Aromatic and Medicinal Plant*. IntechOpen, 175–206.
- Mota L, Figueiredo AC, Pedro LG, Barroso JG, Ascensão L. 2013. Glandular trichomes, histochemical localization of secretion, and essential oil composition in *Plectranthus grandidentatus* growing in Portugal. *Flavour and Fragrance Journal* 28: 393–401.
- Muhlemann JK, Klempien A, Dudareva N. 2014. Floral volatiles: From biosynthesis to function. *Plant, Cell and Environment* 37: 1936–1949.
- Naidoo Y, Karim T, Heneidak S, Sadashiva CT, Naidoo G. 2012. Glandular trichomes of *Ceratotheca triloba* (Pedaliaceae): Morphology, histochemistry and ultrastructure. *Planta* 236: 1215–1226.
- Neinhuis C, Edelman HG. 1996. Methanol as a rapid fixative for the investigation of plant surfaces by SEM. *Journal of Microscopy* 184: 14–16.
- Nishizawa A, Honda G, Kobayashi Y, Tabata M. 1992. Genetic control of peltate glandular trichome formation in *Perilla frutescens*. *Planta Medica* 58: 188–191.
- Novak J, Draxler L, Göhler I, Franz CM. 2005. Essential oil composition of *Vitex agnus-castus* - Comparison of accessions and different plant organs. *Flavour and Fragrance Journal* 20: 186–192.
- Ogunwande IA, Olawore NO, Adeleke KA, Ekundayo O. 2005. Volatile constituents from the leaves of *Eucalyptus cloeziana* F. Muell and *E.*

propinqua Deane & Maiden from Nigeria. *Flavour and Fragrance Journal* 20: 637–639.

- Ojeda-Sana AM, van Baren CM, Elechosa MA, Juárez MA, Moreno S. 2013. New insights into antibacterial and antioxidant activities of rosemary essential oils and their main components. *Food Control* 31: 189–195.
- Okada T, Afendi FM, Altaf-UI-Amin M, Takahashi H, Nakamura K, Kanaya S. 2010. Metabolomics of medicinal plants: the importance of multivariate analysis of analytical chemistry data. *Current Computer-Aided Drug Design* 6: 179–96.
- Okoh SO, Iweriebor BC, Okoh OO, Nwodo UU, Okoh AI. 2016. Antibacterial and antioxidant properties of the leaves and stem essential oils of *Jatropha gossypifolia* L. *BioMed Research International* 9.
- Padalia RC, Verma RS, Chauhan A, Chanotiya CS. 2013. Changes in aroma profiles of 11 Indian *Ocimum* taxa during plant ontogeny. *Acta Physiologiae Plantarum* 35: 2567–2587.
- Paiva EAS. 2016. How do secretory products cross the plant cell wall to be released? A new hypothesis involving cyclic mechanical actions of the protoplast. *Annals of Botany* 117: 533–540.
- Palmer-Young EC, Veit D, Gershenzon J, Schuman MC. 2015. The sesquiterpenes (*E*)- β -farnesene and (*E*)- α -bergamotene quench ozone but fail to protect the wild tobacco *Nicotiana attenuata* from ozone, UVB, and drought stresses. *PLoS ONE* 10.
- Park S, Park AR, Im S, Han YJ, Lee S, Back K, Kim J, Kim YS. 2014. Developmentally regulated sesquiterpene production confers resistance to *Colletotrichum gloeosporioides* in ripe pepper fruits. *PLoS ONE* 9: 1–10.
- Paul MJ, Foyer CH. 2001. Sink regulation of photosynthesis. *Journal of Experimental Botany* 52: 1383–1400.
- Perez-Cacho PR, Rouseff RL. 2008. Fresh squeezed orange juice odor: A review. *Critical Reviews in Food Science and Nutrition* 48: 681–695.
- Pichersky E, Gershenzon J. 2002. The formation and function of plant volatiles: Perfumes for pollinator attraction and defense. *Current Opinion in Plant Biology* 5: 237–243.
- Pljevljakušić D, Rančić D, Ristić M, Vujisić L, Radanović D, Dajić-Stevanović Z. 2012. Rhizome and root yield of the cultivated *Arnica montana* L., chemical composition and histochemical localization of essential oil. *Industrial Crops and Products* 39: 177–189.

- Plowden C. 2003. Production ecology of Copaiba (*Copaifera* Spp.) oleoresin in the Eastern Brazilian Amazon. *Economic Botany* 57: 491–501.
- Postek MT, Tucker SC. 1983. Ontogeny and ultrastructure of secretory oil cells in *Magnolia grandiflora* L. *Botanical Gazette* 144: 501–512.
- Poulsen AD. 2012. *Etilingera of Sulawesi*. Natural History Publisher.
- Prieto MA, Rodríguez-Amado I, Vázquez JA, Murado MA. 2012. β -Carotene assay revisited. Application to characterize and quantify antioxidant and prooxidant activities in a microplate. *Journal of Agricultural and Food Chemistry* 60: 8983–8993.
- Qiu Y, Reed D. 2014. Gas Chromatography in Metabolomics Study In: Guo XBT-A in GC, ed. Rijeka: InTech, Ch. 04.
- Raal A, Orav A, Arak E. 2007. Composition of the essential oil of *Salvia officinalis* L. from various European countries. *Natural Product Research* 21: 406–411.
- Rajaonarivony JIM, Croteau R. 1992. Characterization and mechanism of (4S)-limonene synthase, a monoterpene cyclase from the glandular trichomes of peppermint (*Ventha*). *Archives of Biochemistry and Biophysics* 296: 49–57.
- Rawsthorne S. 2002. Carbon flux and fatty acid synthesis in plants. *Progress in Lipid Research* 41: 182–196.
- Razali MTA, Zainal ZA, Maulidiani M, *et al.* 2018. Classification of raw stingless bee honeys by bee species origins using the NMR- and LC-MS-based metabolomics approach. *Molecules* 23: 1–18.
- Reinhard J, Srinivasan M V, Zhang S. 2004. Olfaction: scent-triggered navigation in honeybees. *Nature* 427: 411.
- Remashree AB, Unnkrishnan K, Ravindran PN. 1999. Development of oil cell and ducts in ginger. *Journal of Spices and Aromatic Crops* 8: 163–170.
- Rizzo WB. 2015. Importance for epidermal structure and function. *Biochimica et Biophysica Acta* 1841: 377–389.
- Rodrigues-Corrêa KC da S, de Lima JC, Fett-Neto AG. 2012. Pine oleoresin: Tapping green chemicals, biofuels, food protection, and carbon sequestration from multipurpose trees. *Food and Energy Security* 1: 81–93.
- Rodrigues G, Fernandes F, Costa JGM, Rodrigues FFG, Campos AR. 2013.

Study of the interference between *Plectranthus* species essential oils from Brazil and aminoglycosides. *Evidence-Based Complementary and Alternative Medicine* 2013: 7.

- Rodrigues L, Monteiro P, Póvoa O, Teixeira G, Moldão M, Figueiredo AC, Monteiro A. 2008. Morphology of secretory structures and essential oil composition in *Mentha cervina* L. from Portugal. *Flavour and Fragrance Journal* 23: 340–347.
- Rodrigues L, Póvoa O, Teixeira G, Figueiredo AC, Moldão M, Monteiro A. 2013. Trichomes micromorphology and essential oil variation at different developmental stages of cultivated and wild growing *Mentha pulegium* L. populations from Portugal. *Industrial Crops and Products* 43: 692–700.
- Ruberto G, Baratta MT. 2000. Antioxidant activity of selected essential oil components in two lipid model systems. *Food Chemistry* 69: 167–174.
- Rubiolo P, Sgorbini B, Liberto E, Cordero C, Bicchi C. 2010. Essential oils and volatiles: Sample preparation and analysis. A review. *Flavour and Fragrance Journal* 25: 282–290.
- Russell M, Southwell IA, Waterman PG. 2011. α -Phellandren-8-ol, a Rare monoterpene alcohol from the oil of *Prostanthera staurophylla* F. Muell. *Journal of Essential Oil Research* 13: 446–447.
- Rusydi A, Talip N, Latip J, Rahman RA, Sharif I. 2013. Morphology of trichomes in *Pogostemon cablin* Benth. (Lamiaceae). *Australian Journal of Crop Science* 7: 744–749.
- Saccenti E, Hoefsloot HCJ, Smilde AK, Westerhuis JA, Hendriks MMWB. 2014. Reflections on univariate and multivariate analysis of metabolomics data. *Metabolomics* 10: 361–374.
- Sangwan NS, Farooqi AHA, Shabih F, Sangwan RS. 2001. Regulation of essential oil production in plants. *Plant Growth Regulation* 34: 3–21.
- Santos Tozin LR dos, de Melo Silva SC, Rodrigues TM. 2016. Non-glandular trichomes in Lamiaceae and Verbenaceae species: morphological and histochemical features indicate more than physical protection. *New Zealand Journal of Botany* 54: 446–457.
- Sardans J, Llusà J, Niinemets Ü, Owen S, Peñuelas J. 2010. Foliar mono- and sesquiterpene contents in relation to leaf economic spectrum in native and alien species in Oahu (Hawai'i). *Journal of Chemical Ecology* 36: 210–226.

- Savage TJ, Hamilton BS, Croteau R. 1996. Biochemistry of short-chain alkanes. *Plant Physiology* 110: 179–186.
- Schiestl FP. 2015. Ecology and evolution of floral volatile-mediated information transfer in plants. *New Phytologist* 206: 571–577.
- Schiestl FP, Ayasse M. 2001. Post-pollination emission of a repellent compound in a sexually deceptive orchid: A new mechanism for maximising reproductive success? *Oecologia* 126: 531–534.
- Sen S, Dehingia M, Talukdar NC, Khan M. 2017. Chemometric analysis reveals links in the formation of fragrant bio-molecules during agarwood (*Aquilaria malaccensis*) and fungal interactions. *Nature Publishing Group*: 1–14.
- Senatore F. 2000. Oli essenziali. *EMSI, Roma*.
- Serrato-Valenti G, Bisio A, Cornara L, Ciarallo G. 1997. Structural and histochemical investigation of the glandular trichomes of *Salvia aurea* L. leaves, and chemical analysis of the essential oil. *Annals of Botany* 79: 329–336.
- Shaaban H a. EE, El-Ghorab AH, Shibamoto T. 2012. Bioactivity of essential oils and their volatile aroma components: Review. *Journal of Essential Oil Research* 24: 203–212.
- Shahidi F, Chandrasekara A. 2010. Hydroxycinnamates and their in vitro and in vivo antioxidant activities. *Phytochemistry Reviews* 9: 147–170.
- Shahidi F, Zhong Y. 2015. Measurement of antioxidant activity. *Journal of Functional Foods* 18: 757–781.
- da Silva AC, Ristin. R, Lopes PM, Monteiro, de Azevedo MM, Ari. B, Costa DC, Ristin. M, Alviano CS, Ale., Alviano DS, Ale. 2012. Biological activities of α -pinene and β -pinene enantiomers. *Molecules* 17: 6305–6316.
- Simmons AT, Gurr GM. 2005. Trichomes of *Lycopersicon* species and their hybrids: Effects on pests and natural enemies. *Agricultural and Forest Entomology* 7: 265–276.
- Singh N, Luthra R, Sangwan RS. 1991. Mobilization of starch and essential oil biogenesis during leaf ontogeny of lemongrass (*Cymbopogon flexuosus* stapf.). *Plant and Cell Physiology* 32: 803–811.
- Sivakumar R, Jebanesan A, Govindarajan M, Rajasekar P. 2011. Larvicidal and repellent activity of tetradecanoic acid against *Aedes aegypti* (Linn.) and *Culex quinquefasciatus* (Say.) (Diptera: Culicidae). *Asian Pacific Journal of Tropical Medicine* 4: 706–710.

- Smirnova O V, Chistyakova AA, Zaugolnova LB, Evstigneev OI, Popadiouk R V, Romanovsky AM. 1999. Ontogeny of a tree. *Botan. Zh* 84: 8–20.
- Smith CA, Want EJ, O'Maille G, Abagyan R, Siuzdak G. 2006. XCMS: Processing mass spectrometry data for metabolite profiling using nonlinear peak alignment, matching, and identification. *Analytical Chemistry* 78: 779–787.
- Smolinska A, Hauschild a C, Fijten RR, Dallinga JW, Baumbach J, van Schooten FJ. 2014. Current breathomics-a review on data pre-processing techniques and machine learning in metabolomics breath analysis. *Journal of Breath Research* 8: 27105.
- Southwell IA. 2005. 25 Years of natural product R&D with New South Wales agriculture. *Molecules* 10: 1232–1241.
- Spicher L, Almeida J, Gutbrod K, *et al.* 2017. Essential role for phytol kinase and tocopherol in tolerance to combined light and temperature stress in tomato. *Journal of Experimental Botany* 68: 5845–5856.
- Stratmann JW, Bequette CJ. 2016. Hairless but no longer clueless: understanding glandular trichome development. *Journal of Experimental Botany* 67: 5285–5287.
- Sun P, Schuurink RC, Caissard JC, Huguency P, Baudino S. 2016. My Way: Noncanonical biosynthesis pathways for plant volatiles. *Trends in Plant Science* 21: 884–894.
- Svoboda K, Svoboda T, Syred A. 2001. A closer look - Secretory Structures of aromatic and medicinal plants. *HerbalGram* 53: 34–43.
- Szymanski DB, Klis DA, Larkin JC, Marks MD. 1998. *cot1*: A regulator of *Arabidopsis* trichome initiation. *Genetics* 149: 565–577.
- Takai H, Iwama R, Kobayashi S, Horiuchi H, Fukuda R, Ohta A. 2012. Construction and characterization of a *Yarrowia lipolytica* mutant lacking genes encoding cytochromes P450 subfamily 52. *Fungal Genetics and Biology* 49: 58–64.
- Talbot MJ, White RG. 2013. Methanol fixation of plant tissue for Scanning Electron Microscopy improves preservation of tissue morphology and dimensions. *Plant Methods* 9: 1–7.
- Tattini M, Matteini P, Saracini E, Traversi ML, Giordano C, Agati G. 2007. Morphology and biochemistry of non-glandular trichomes in *Cistus salvifolius* L. leaves growing in extreme habitats of the Mediterranean basin. *Plant Biology* 9: 411–419.

- Teixeira G, Correia AI, Vasconcelos T, Feijão D, Madureira AM. 2013. *Lavandula stoechas* subsp. *luisieri* and *L. pedunculata* - phytochemical study, micromorphology and histochemistry. *Revista de Ciências Agrárias* 36: 220–228.
- Tholl D, Boland W, Hansel A, *et al.* 2006. Practical approaches to plant volatile analysis. *The Plant Journal* 45: 540–560.
- Tissier A. 2018. Plant secretory structures: more than just reaction bags. *Current Opinion in Biotechnology* 49: 73–79.
- Tozin LR dos S, Rodrigues TM. 2017. Morphology and histochemistry of glandular trichomes in *Hyptis villosa* Pohl ex Benth. (Lamiaceae) and differential labeling of cytoskeletal elements. *Acta Botanica Brasílica* 31: 330–343.
- Turner Glenn W, Gershenzon J, Croteau RB. 2000. Development of peltate glandular trichomes of peppermint. *Plant Physiology* 124: 665–679.
- Turner G W, Gershenzon J, Croteau RB. 2000. Distribution of peltate glandular trichomes on developing leaves of peppermint. *Plant Physiology* 124: 655–64.
- Turner G, Gershenzon J, Nielson EE, Froehlich JE, Croteau R. 1999. Limonene Synthase, the enzyme responsible for monoterpene biosynthesis in peppermint, is localized to leucoplasts of oil gland secretory cells. *Plant Physiology* 120: 879–886.
- Valgimigli L, Valgimigli M, Gaiani S, Pedulli GF, Bolondi L. 2012. Essential oils: an overview on origins, chemistry, properties and uses. *Essential Oils as Natural Food Additives*: 1–24.
- Vancanneyt G, Sanz C, Farmaki T, *et al.* 2001. Hydroperoxide lyase depletion in transgenic potato plants leads to an increase in aphid performance. *Proceedings of the National Academy of Sciences of the United States of America* 98: 8139–8144.
- Veiga VF, Pinto AC. 2002. O Genero *Copaifera* L. *Química Nova* 25: 273–286.
- Vekiari SA, Protopapadakis EE, Papadopoulou P, Papanicolaou D, Panou C, Vamvakias M. 2002. Composition and seasonal variation of the essential oil from leaves and peel of a cretan lemon variety. *Journal of Agricultural and Food Chemistry* 50: 147–153.
- Ventrella MC, Marinho CR. 2008. Morphology and histochemistry of glandular trichomes of *Cordia verbenacea* DC. (Boraginaceae) leaves. *Revista Brasil Botany* 1: 457–467.

- Verma RS, Padalia RC, Chauhan A. 2012. Variation in the volatile terpenoids of two industrially important basil (*Ocimum basilicum* L.) cultivars during plant ontogeny in two different cropping seasons from India. *Journal of the Science of Food and Agriculture* 92: 626–631.
- Villas-Bôas SG, Mas S, Åkesson M, Smedsgaard J, Nielsen J. 2005. Mass spectrometry in metabolome analysis. *Mass Spectrometry Reviews* 24: 613–646.
- Voelckel C, Jander G. 2014. Chapter 8 - Costs of resistance in plants: from theory to evidence In: *Annual Plant Reviews*.263–307.
- Vogt T. 2010. Phenylpropanoid biosynthesis. *Molecular Plant* 3: 2–20.
- Wagner GJ, Wang E, Shepherd RW. 2004. New approaches for studying and exploiting an old protuberance, the plant trichome. *Annals of Botany* 93: 3–11.
- Wang W, Li N, Luo M, Zu Y, Efferth T. 2012. Antibacterial activity and anticancer activity of *Rosmarinus officinalis* L. essential oil compared to that of its main components. *Molecules* 17: 2704–2713.
- Wang Z-Y, Soanes DM, Kershaw MJ, Talbot NJ. 2007. Functional analysis of lipid metabolism in *Magnaporthe grisea* reveals a requirement for peroxisomal fatty acid β -oxidation during appressorium-mediated plant infection. *Molecular Plant-Microbe Interactions* 20: 475–491.
- Wang Y, Xu L, Shen H, Wang J, Liu W, Zhu X. 2015. Metabolomic analysis with GC-MS to reveal potential metabolites and biological pathways involved in Pb & Cd stress response of radish roots. *Nature Publishing Group*: 1–13.
- Wei A, Shibamoto T. 2007. Antioxidant activities and volatile constituents of various essential oils. *Journal of Agricultural and Food Chemistry* 55: 1737–1742.
- Werker E. 2000. Trichome diversity and development. *Advances in botanical research* 31: 1–35.
- Werker E, Ravid U, Putievsky E. 1985. Structure of glandular hairs and identification of the main components of their secreted material in some species of the Labiatae. *Israel Journal of Botany* 34: 31–45.
- Western TL, Skinner DJ, Haughn GW. 2000. Differentiation of mucilage secretory cells of the *Arabidopsis* seed coat. *Plant Physiology* 122: 345–356.
- Wheelock AM, Wheelock CE. 2013. Trials and tribulations of 'omics data

analysis: assessing quality of SIMCA-based multivariate models using examples from pulmonary medicine. *Molecular bioSystems* 9: 2589–96.

- Widhalm JR, Jaini R, Morgan JA, Dudareva N. 2015. Rethinking how volatiles are released from plant cells. *Trends in Plant Science* 20: 545–550.
- Wiedemuth K, Müller J, Kahlau A, *et al.* 2005. Successive maturation and senescence of individual leaves during barley whole plant ontogeny reveals temporal and spatial regulation of photosynthetic function in conjunction with C and N metabolism. *Journal of Plant Physiology* 162: 1226–1236.
- Wijekoon MMJO, Bhat R, Karim AA., Fazilah A. 2013. Chemical composition and antimicrobial activity of essential oil and solvent extracts of torch ginger inflorescence (*Etilingera elatior* Jack.). *International Journal of Food Properties* 16: 1200–1210.
- Wong KC, Sivasothy Y, Boey PL, Osman H, Sulaiman B. 2010. Essential Oils of *Etilingera elatior* (Jack) R. M. Smith and *Etilingera littoralis* (Koenig) Giseke. *Journal of Essential Oil Research* 22: 461–466.
- Wong KC, Yap YF, Ham LK. 1993. The essential oil of young flower shoots of *Phaeomeria speciosa* Koord. *Journal of Essential Oil Research* 5: 135–138.
- Wu Z, Li H, Yang Y, Zhan Y, Tu D. 2013. Variation in the components and antioxidant activity of *Citrus medica* L. var. *sarcodactylis* essential oils at different stages of maturity. *Industrial Crops and Products* 46: 311–316.
- Xia J, Sinelnikov I V, Han B, Wishart DS. 2015. MetaboAnalyst 3.0-making metabolomics more meaningful. *Nucleic Acids Research* 43: W251–W257.
- Yamashita I, Nemoto Y, Yoshikawa S. 1976. Formation of volatile alcohols and esters from aldehydes in strawberries. *Phytochemistry* 15: 1633–1637.
- Yamawo A, Suzuki N, Tagawa J, Hada Y. 2012. Leaf ageing promotes the shift in defence tactics in *Mallotus japonicus* from direct to indirect defence. *Journal of Ecology* 100: 802–809.
- Yang J, Nie Q, Ren M, *et al.* 2013. Metabolic engineering of *Escherichia coli* for the biosynthesis of alpha-pinene. *Biotechnology for Biofuels* 6:1.
- Yanishlieva NV., Marinova EM, Gordon MH, Raneva VG. 1999. Antioxidant activity and mechanism of action of thymol and carvacrol in two lipid

systems. *Food Chemistry* 64: 59–66.

- Yeats, H. 2012. The history and cultivation of *Etilingera* - the torch gingers - at the Royal Botanic Garden Edinburgh. *The Journal of Botanic Garden Horticulture* 11:71-86.
- Yosr Z, Hnia C, Rim T, Mohamed B. 2013. Changes in essential oil composition and phenolic fraction in *Rosmarinus officinalis* L. var. *typicus* Batt. organs during growth and incidence on the antioxidant activity. *Industrial Crops and Products* 43: 412–419.
- Zhang X, Grey PH, Krishnakumar S, Oppenheimer DG. 2005. The *IRREGULAR TRICHOME BRANCH* loci regulate trichome elongation in Arabidopsis. *Plant and Cell Physiology* 46: 1549–1560.
- Zhang X, Sawhney VK, Davis AR. 2014. Annular floral nectary with oil producing trichomes in *Salvia Farinaceae* (Lamiaceae): anatomy, histochemistry, ultrastructure and significance. *American Journal of Botany* 101: 1849–1867.
- Zhao Q, Chen X. 2016. A new function of plant trichomes. *Nature Publishing Group*: 1–2.
- Zoghbi MDGB, Andrade EHA. 2005. Volatiles of the *Etilingera elatior* (Jack) R. M. SM. and *Zingiber spectabile* Griff.: Two Zingiberaceae cultivated in the Amazon. *Journal of Essential Oil Research* 17: 209–211.
- Zuzarte M, Salgueiro L. 2015. Essential Oils Chemistry In: de Sousa DP. *Bioactive Essential Oils and Cancer*. Springer International Publishing Switzerland, 19–59.

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LIST OF PUBLICATIONS

Lee YL, Ding, P, Abas F, Ismail IS. 2020. Chemical composition of essential oils in torch ginger's [*Etilingera elatior* (Jack) R. M. Smith] leaves and inflorescence at different developmental stages. (To be submitted).

Lee YL, Ding, P, Abas F, Ismail IS. 2019. Floral morphology and secretory structures of *Etilingera elatior* (Jack) R. M. Smith (Zingiberaceae). *Protoplasma* (under review).

Lee YL, Ding P. 2016. Production of essential oil in plants: ontogeny, secretory structures and seasonal variations. *Pertanika Journal of Scholarly Research Reviews* 2(1): 1-10.

Conference Presentation

Lee YL, Ding P. Screening analysis of essential oil of *Etilingera elatior* – variations among plant parts and ontogeny. 3rd ISHS Southeast Asia Symposium on Quality Management in Postharvest Systems, Siem Reap, Cambodia, 13-15 August 2015.



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