

# CHARACTERIZATION OF NOVEL OIL PALM DEFENSE-RELATED TRANSCRIPTION FACTORS DURING BIOTROPHIC AND NECROTROPHIC INFECTION PHASES OF Ganoderma boninense

# **NURSHAFIKA BINTI MOHD SAKEH**

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By

NURSHAFIKA BINTI MOHD SAKEH

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirement for the Degree of Doctor of Philosophy

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# TO MY LOVELY PARENTS.

You both have sacrificed through blood, sweat and tears to nurture me into whom I have become today.

May Allah rewards both of you Jannatul Firdaus.

# CHARACTERIZATION OF NOVEL OIL PALM DEFENSE-RELATED TRANSCRIPTION FACTORS DURING BIOTROPHIC AND NECROTROPHIC INFECTION PHASES OF Ganoderma boninense

By

### NURSHAFIKA BINTI MOHD SAKEH

January 2020

Chair : Siti Nor Akmar Abdullah, PhD

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Basal stem rot (BSR) disease, caused by Ganoderma boninense has been pinpointed to be one of the major factors that contribute to the decline in yield of oil palm. Hemibiotroph pathogen such as G. boninense manipulates the host defense mechanisms to strategically infect host plant by switching from biotrophic to necrotrophic phase. Recognizing the early infection phase of G. boninense by identifying phase-specific oil palm transcription factors (TFs) may offer opportunities to tackle or attenuate the progress of infection. The fragmentary information on molecular interactions between plant and hemibiotroph should be explicated to identify the key molecular mechanisms of pathogenesis and plant immunity. Thus, this study is an attempt to recognize specific TFs as 'key' biomarkers in identifying oil palm defense mechanisms during biotrophic and necrotrophic infection phases of G. boninense. Artificial infection of G. boninense on oil palm was performed by using Ganoderma-inoculated rubber wood blocks (RWBs) and bare RWBs serves as mock-treatment presenting abiotic stress. In order to identify oil palm defense response during early interaction with G. boninense, transcriptomic analysis of root tissues at different time points of 3, 7 and 11 days post inoculation (d.p.i) was carried out. High-throughput RNA-seq data analysis has revealed two distinguishable expression profiles of oil palm genes that formed the basis for deducing biotrophic phase at early interaction (3 d.p.i) which switched to initiation of necrotrophic phase at later stage (11 d.p.i) of infection. Based on the findings, the present study focused on identifying differentially expressed genes (DEGs) encoding TFs from the generated RNA-seq data. A total of 106 upregulated and 108 downregulated DEGs of TFs were identified. There are four established defense-related pathways that have been presented whereby reported genes involved in cell wall modification, reactive oxygen species (ROS)-mediated signaling, programmed cell death (PCD) and plant innate immunity were differentially expressed. The genes were found to be either upregulated or downregulated during the two distinct infection phases. Multiplex semi-quantitative RT-PCR was conducted to screen for defense-related TFs independent of abiotic stress. Normalized band intensity of Ganoderma-treated (GT) samples were compared to the mock-treated (MT) samples to estimate the mRNA expression level between groups. The expression patterns of eight candidate TFs genes including EgJUB1, EgERF113,

EgTCP15, EgNAC29, EgEIN3, EgMYC2, EgNAC83 and EgMYB122 were further quantified via quantitative Real-Time PCR (qPCR). Both EgJUB1 and EgERF113 were found to be specifically upregulated under biotic stress at all time points. The findings discovered upregulation of EgJUB1 during biotrophic phase while EgERF113 demonstrated prominent upregulation as oil palm switches to defense against necrotrophic phase. Characterization of EgJUB1 and EgERF113 was performed via in vivo Yeast One-Hybrid (Y1H) assay and in vitro electrophoretic mobility shift assay (EMSA). JUB1 has been reported to bind to NAC binding site (NACBS) motif during abiotic stress. The present study is the first report on the binding activity of EgJUB1 to secondary wall NAC binding element 1 (SNBE1) which was present in the promoter region of EgHSFC-2b having similar expression profile as EgJUB1. SNBE1 motifs with single nucleotide change at either 5<sup>th</sup> or 18<sup>th</sup> position have been found in the promoter regions of a few TFs that co-expressed with EgJUB1, including EgHSFB-4b and EgGAMYB X2. Meanwhile EgERF113 binds to GCC-box and DRE/CRT motifs promoting plasticity in upregulating downstream defense-related genes against G. boninense attack. Sequence analysis revealed the presence of NAC DNA binding domain (DBD) in EgJUB1. Amino acid change from phenylalanine (F) to tryptophan (W) at 14<sup>th</sup> position of EgERF113 DBD proved the binding specificity to both GCC-box and DRE/CRT motifs. Prolonged treatment revealed oil palm seedlings succumbing to the G. boninense infection. Mature basidiomata of G. boninense was observed at 24 weeks post inoculation (w.p.i) and the infection culminated in plant death. Overall, our findings propose EgJUB1 and EgERF113 as key TFs in orchestrating the oil palm defense mechanisms during biotrophic and necrotrophic infection phases of hemibiotrophic G. boninense, respectively.

# Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

# PENCIRIAN FAKTOR TRANSKRIPSI BERKAITAN PERTAHANAN POKOK KELAPA SAWIT YANG BAHARU SEMASA JANGKITAN FASA BIOTROFIK DAN NEKROTROFIK OLEH Ganoderma boninense

# Oleh

### NURSHAFIKA BINTI MOHD SAKEH

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Penyakit reput pangkal batang (BSR), disebabkan oleh Ganoderma boninense telah ditunjukkan sebagai salah satu faktor utama yang menyumbang kepada penurunan hasil kelapa sawit. Patogen hemibiotrof seperti G. boninense memanipulasi mekanisme pertahanan perumah untuk menjangkiti perumah tumbuhan secara strategik dengan menukar fasa dari biotrofik kepada nekrotrofik. Mengenali fasa awal jangkitan G. boninense dengan mengenal pasti faktor transkripsi (TFs) kelapa sawit yang berfasa spesifik boleh menawarkan peluang untuk menangani atau melemahkan perkembangan jangkitan. Maklumat serpihan terhadap interaksi molekular antara tumbuhan dan hemibiotrof harus diperincikan untuk mengenalpasti mekanisme molekular yang utama daripada patogenesis dan imuniti tumbuhan. Oleh itu, kajian ini adalah percubaan untuk mengenal TFs spesifik sebagai 'kunci' penanda bio dalam mengenal pasti mekanisme pertahanan kelapa sawit semasa jangkitan fasa biotrofik dan nekrotrofik G. boninense. Jangkitan buatan G. boninense ke atas kelapa sawit telah dilakukan menggunakan blok kayu getah (RWB) terjangkit inokulum G. boninense dan RWB kosong sebagai rawatan olokan mewakili tekanan abiotik. Untuk mengenal pasti respon pertahanan kelapa sawit semasa interaksi awal dengan G. boninense, analisis transkriptomik pada tisu akar pada titik masa yang berlainan iaitu 3, 7, dan 11 hari-selepas-inokulasi (d.p.i) telah dijalankan. Analisis data RNA-seq keupayaan celusan tinggi telah mendedahkan dua profil ekspresi yang jelas daripada gen kelapa sawit yang membentuk asas kepada mendeduksi fasa biotrofik pada jangkitan awal (3 d.p.i) yang mana ditukar kepada permulaan fasa nektorofik pada peringkat kemudian (11 d.p.i). Berdasarkan kepada penemuanpenemuan ini, kajian kini memfokuskan pada mengenal pasti gen diekspres secara ketara (DEGs) yang mengekod TFs dari data RNA-seq yang terhasil. Sejumlah 106 DEGs dikawal naik dan 108 DEGs dikawal turun daripada TFs telah dikenalpasti. Terdapat empat laluan berkaitan pertahanan yang kukuh telah dibentangkan yang mana gen yang dilaporkan terlibat dalam pengubahsuaian dinding sel, isyarat spesis oksigen reaktif (ROS)-perantara, kematian sel terprogram, dan keimunan semulajadi tumbuhan telah diekspres secara ketara. Gen ini didapati samada dikawal naik atau dikawal turun dalam dua fasa jangkitan yang berbeza tersebut. RT-PCR semi-kuantitatif multipleks telah dijalankan untuk menyaring TFs berkaitan pertahanan yang bebas tekanan abiotik.

Keamatan jalur ternormalkan daripada sampel terawat Ganoderma (GT) dibanding dengan sampel olokan (MT) untuk menganggarkan tahap ekspresi mRNA antara kumpulan. Pola ekspresi lapan calon gen TFs termasuk EgJUB1, EgERF113, EgTCP15, EgNAC29, EgEIN3, EgMYC2, EgNAC83 dan EgMYB122 selanjutnya dikuantitikan melalui PCR-masa nyata kuantitatif (qPCR). Kedua-dua EgJUB1 dan EgERF113 telah dikesan dikawal naik secara spesifik di bawah tekanan biotik pada semua titik masa. Dapatan-dapatan tersebut menunjukkan kawalan naik EgJUB1 semasa fasa biotrofik sementara EgERF113 menunjukkan kawalan naik yang ketara apabila kelapa sawit. menukar pertahanan menentang fasa nekrotrofik. Pencirian EgJUB1 dan EgERF113 telah dijalankan melaui asai Yis Satu-Hibrid (Y1H) in vivo dan asai syif kegerakan elektroforesis (EMSA) in vitro. JUB1 telah dilaporkan mengikat kepada motif tapak ikatan NAC (NACBS) semasa tekanan abiotik. Kajian ini adalah laporan pertama ke atas aktiviti ikatan EgJUB1 kepada dinding sekunder element ikatan 1 NAC (SNBE1) yang mana berada dalam kawasan promoter EgHSFC-2b dengan profil ekspresi yang sama seperti EgJUB1. Motif SNBE1 dengan pertukaran nukleotida tunggal samada pada kedudukan kelima atau kelapan telah ditemukan dalam kawasan promoter beberapa TFs yang terekspres bersama EgJUBI, termasuk EgHSFB-4b dan EgGAMYB X2. Sementara itu EgERF113 mengikat pada kotak GCC dan motif DRE/CRT yang menggalakkan keplastikan dalam mengawal naik gen berkaitan pertahanan hilir melawan serangan G. boninense. Analysis jujukan menunjukkan kehadiran domain ikatan DNA (DBD) NAC dalam EgJUB1. Perubahan asid amino daripada fenilalanina kepada triptofan pada kedudukan ke-14 DBD EgERF113 membuktikan kespesifikan ikatan pada kedua-dua kotak GCC dan motif DRE/CRT. Rawatan berpanjangan menyebabkan anak benih kelapa sawit alah kepada jangkitan G. boninense. Basidiomata matang G. boninense telah diperhati pada 24 minggu selepas inokulasi (w.p.i) dan jangkitan tersebut berakhir dengan kematian tumbuhan. Kesimpulannya, penemuan kami mencadangkan EgJUB1 dan EgERF113 sebagai TFs utama dalam mengatur mekanisme pertahanan kelapa sawit masing-masing semasa fasa jangkitan biotrofik dan nekrotrofik oleh hemibiotrofik G. boninense.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follow:

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protein kinase; CESA: Cellulose synthase A; CML: Calmodulin-like; CSLD: Cellulose synthase-like protein D; ETI: Effector-triggered immunity; GST: Glutathione S-transferase; NPR1: NONEXPRESSOR OF PATHOGENESIS-RELATED GENE 1; SOD: Superoxide dismutase; RBOH: Respiratory burst oxidase homolog protein; ROS: Reactive oxygen species; MC9: Metacaspase 9; PTI: PAMP-triggered immunity.

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4.1 Deduced nucleotide and amino acid sequence of EgJUB1.

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- 4.6 Electrophoretic mobility shift assay (EMSA) of EgJUB1 with SNBE1 probe. Lane 1 to 3: EBNA control system. Lane 4 to 8: EgJUB1 test system. Lane 2 and 3 were positive and negative control for EMSA, respectively. Shifted band in lane 6 indicated direct binding of EgJUB1 to biotinylated SNBE1 probe, but none when tested with untransformed nuclear extract

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- 47 Deduced nucleotide and amino acid sequence of EgERF113. Translation of nucleotide to amino acid sequence was carried using Out translate tool in **ExPASv** https://web.expasy.org/translate/. The pink shading indicates specific amino acid predicted for binding to both GCC-box and DRE/CRT motifs. The basic amino acids of putative nuclear localization signal **PWGKWAAEIRDPRKAARV** underlined with red.
- Multiple sequence alignment of the deduced amino acid 4.8 sequence of EgERF113 protein with other ERF113-related proteins. Structure based alignment was constructed using Expresso of T-Coffee http://tcoffee.crg.cat/apps/tcoffee/do:expresso. The asterisks indicate fully conserved residues. The colons indicate conservation of strong group. The full stops indicate the conservation of weak group. The dashes indicate no consensus. The red shadings indicate reliable and consistent alignment. Yellow and green shadings indicate average reliability while blue shadings indicate very poor alignment. Pink shadings indicate specific amino acid for binding to both GCC-box and DRE/CRT motifs at specific location. The putative nuclear localization signal is shown by double-headed arrow below the sequences. The deduced amino acids sequences of ERF113 are derived from different species; Elaeis guineensis (Eg), Musa acuminata (Ma), Phoenix dactylifera (Pd), Ananas comosus (Ac), Asparagus officinalis (Ao), Populus trichocarpa (Pt), Hevea brasiliensis (Hb), Theobroma cacao (Tc) and Arabidopsis thaliana (At).
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DRE/CRT: Dehydration response element/C-repeat; EIN3: Ethylene insensitive 3; ET: Ethylene; ETI: Effector-triggered immunity; HR: Hypersensitive response; HSFs: Heat stress transcription factors; HSP: Heat shock protein; JA: Jasmonic acid; NPR1: NONEXPRESSOR OF PATHOGENESIS-RELATED GENE 1; PR: Pathogenesis-related; PRRs: Pattern recognition receptors; PTI: PAMP-triggered immunity; SNBE: Secondary wall NAC binding element; SA: Salicylic acid; TSS: Transcription start site.



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#### LIST OF ABBREVIATIONS

Alanine Α

AbA Aureobasidin A

AbAr Aureobasidin A resistance gene

ABA Abscisic acid ANOVA Analysis of variance APS Ammonium persulfate

AP2/ERF APETALA2/ethylene responsive factor

ATAAurin tricarboxylic acid

Avr Avirulence

**BCA** Biological control agent bHLH basic Helix-loop-helix

BLAST Basic Local Alignment Search Tool

**hZIP** basic leucine zipper  $Ca^{2+}$ Calcium ion

**CAMTA** Calmodulin-binding transcription activator

Centimeter cm <sup>60</sup>Co Cobalt-60

CTR1 CONSTITUTIVE TRIPLE RESPONSE 1

Base pair bp BSR Basal stem rot Calcium chloride CaCl<sub>2</sub>

complementary Deoxyribonucleic acid cDNA **CDPK** Calcium dependent protein kinase

Coding sequence CDS Cellulose synthase **CESA** Calmodulin-like CML COR Coronatine

CSL Cellulose synthase-like  $C_{T}$ Threshold cycle

Cu

Copper

**CWDE** Cell wall degrading enzyme

d Day Da Dalton

**DAMP** Damage-associated molecular pattern

DNA-binding domain DBD

Differentially expressed gene DEG

DMSO Dimethyl sulfoxide DNA Deoxyribonucleic acid Days post inoculation d.p.i

DREB Dehydration responsive element-binding DRE/CRT Dehydration-Responsive Element/C-Repeat

Glutamic acid

**EBNA** Epstein-Barr Nuclear Antigen EDS1 Enhanced Disease Susceptibility 1 **EDTA** Ethylenediaminetetraacetic acid

Ethylene-insensitive 3 EIN3

EIL EIN3-like

EMSA Electrophoretic Mobility Shift Assay
ENA European Nucleotide Archive
ERE Ethylene-Response Element
ERF Ethylene Responsive Factor
EST Expressed Sequence Tag

ET Ethylene

ETI Effector-triggered immunity

EU European Union F Phenylalanine

Fe Iron fmole femtomole

Felda Federal Land Development Authority

FFB Fresh fruit bunch
G Glutamic acid

GanoDROP Ganoderma and Disease Research of Oil Palm GAPDH Glyceraldehyde 3-phosphate dehydrogenase

gDNA Genomic deoxyribonucleic acid GDP Gross Domestic Product

GO Gene Ontology

GST Glutathione S-transferase
GT Ganoderma-treated

Gy Gray h hour

ha hectare

HLH Helix-loop-helix
HR Hypersensitive response

HSF Heat stress transcription factor

HSP Heat shock protein

Ile Isoleucine

IPTG Isopropyl β-d-1-thiogalactopyranoside

JA Jasmonic acid

JAMA Japan Automobile Manufacturers Association

JAZ Jasmonic acid-ZIM domain

JUB1 JUNGBRUNNEN 1

kb kilobase LB Luria-Bertani

LD-PCR Long-distance Polymerase Chain Reaction

Leu Leucine
LiAc Lithium acetate
LiCl Lithium chloride

M Molar

MAMP Microbe-associated molecular pattern MAPK Mitogen-activated protein kinase

MC Metacaspase
MEA Malt extract agar
ME Mercaptoethanol

min Minute

MgCl<sub>2</sub> Magnesium chloride

MIC Minimal Inhibitory Concentration

MITC Methyl isothiocyanate

Mn Manganese

MPOB Malaysian Palm Oil Board MSD Manganese superoxide dismutase

MT Mock-treated

MTI MAMP-triggered immunity

MW Molecular weight
MYB Myeloblastosis
MYC Myelocytomatosis
N Total sample size
n Sample size
NaCl Sodium chloride

NADH Nicotinamide adenine dinucleotide dehydrogenase

NaOH Sodium hydroxide

NAC No apical meristem (NAM), Arabidopsis transcription

activation factor (ATAF1/2), and cup-shaped cotyledon

(CUC2)

NACBS NAC binding site

NCBI National Centre of Biotechnology Information

NGS Next Generation Sequencing
NIH National Institute of Health
NLS Nuclear localization signals

nm Nanometer nM Nanomolar

NPR1 NONEXPRESSOR OF PATHOGENESIS-RELATED GENE

1

ns Not significant

NST NAC SECONDARY WALL THICKENING PROMOTING

FACTOR1

NTC Non-template control
OD Optical density
ORF Open reading frame

pmole picomole

PAMP Pathogen-associated molecular pattern

PDA Potato dextrose agar
PEG Polyethylene glycol
pI Isoelectric point

PlantTFDB Plant Transcription Factor Database

POFA Palm oil fuel ash
PCD Programmed cell death
PR Pathogenesis-related
PRR Pattern recognition receptor
PTI PAMP-triggered immunity

qPCR Quantitative Real-Time Polymerase Chain Reaction

R Resistance

R<sup>2</sup> Regression coefficient

RAP2.6 RELATED TO APETALA 2.6 RAP2.6L RELATED TO APETALA 2.6-LIKE

RAV Related to Abscisic Acid Insensitive 3/Vivaparous 1

RBOH Respiratory burst oxidase homolog protein

ROS Reactive Oxygen Species RT Room temperature

RT-PCR Reverse Transcriptase Polymerase Chain Reaction

RWB Rubber wood block

s second SA Salicylic acid

SAGE Serial Analysis of Gene Expression
SAR Systemic acquired resistance
SBS Sequencing by synthesis
SBL Sequencing by ligation
SD Synthetic dropout
SDS Sodium dodecyl sulfate

Si Silicon

SEM Standard error of mean SNAC Stress-responsive NAC

SNBE Secondary wall NAC binding element

S.O.C Super Optimal broth with Catabolite repression

SOD Superoxide dismutase

t tonne

TAE Tris-Acetate-EDTA

Tb Terabase

TBE Tris-borate-EDTA

TCP Teosinate branched 1, Cycloidea, Proliferating Cell Factor

TdT Terminal deoxynucleotidyl transferase

TE Tris-EDTA

TEMED N,N,N',N'-tetramethylethane-1,2-diamine
TESH Thermally enhanced sustainable hybrid

TF Transcription factor

TIR-NBS-LRR Toll/interleukin-1 receptor-nucleotide-binding site-leucine-

rich repeat

TPM Transcript per kilobase million

TPL TOPLESS

TPLR TOPLESS-related
TSS Transcription start site

U Unit

UBQ Ubiquitein-β-chain

URA Uracil V Voltan

VND VASCULAR-RELATED NAC-DOMAIN

W Tryptophan

w.p.i Weeks post inoculation

YPDA Yeast Extract Peptone Dextrose Adenine

Y1H Yeast One-Hybrid

μ Micro

x g Gravities (Unit for relative centrifugal force)

°C Degree Celsius

#### **CHAPTER 1**

#### INTRODUCTION

Oil palm (*Elaeis guineensis*) is economically important for countries especially in Southeast Asian region. Malaysia has become the world second largest palm oil exporter accounting 44% of world's export and contributing 39% of world palm oil production (Malaysian Palm Oil Council, 2017). The country achieved the highest export revenue in 2017 from palm oil and palm oil products of approximately USD18.5 billion (Malaysian Palm Oil Board, 2018). Oil palm has been recognized as one of the world's largest edible oil source as well as precursor of biodiesel fuel (Kurnia et al., 2016; Stichnothe et al., 2014). To date, oil palm is the most efficient oil-bearing crop with 5 to 9 times higher yield per ha yearly than other vegetable oil crops such as soybean, sunflower and rapeseed (Lam et al., 2019).

However, the sustainable production of palm oil is hampered severely due to basal stem rot (BSR) disease caused by pathogenic and destructive fungus, *Ganoderma boninense*. Malaysia and Indonesia have suffered highest economic loss from the disease ranging from RM225 million to RM1.5 billion (up to USD 361 million) per year in both countries (Morel et al., 2016, Arif et al., 2011). The vast spread of BSR from one oil palm plantation to another in both countries is so serious and almost uncontrollable. *G. boninense* is a basidiomycete soil-borne pathogen which is the prevalent species causing BSR in oil palm (Ishaq et al., 2014; Hushiarian et al., 2013). The spreading of *G. boninense* in oil palm plantation involves dikaryotic mycelium spreading through root-to-root contact, dispersal of basidiospore and the presence of secondary inoculum in the soil (Isaac et al., 2018; Chong et al., 2017a).

The main constraint in managing the BSR disease is the failure in identifying *Ganoderma*-infected palm trees at the earliest stage of infection since there is no visible symptoms. The pathogenic fungus is categorized as hemibiotroph wherein initial biotrophic lifestyle is needed for a period of time before switching to subsequent necrotrophic phase (Chong et al., 2017a). The dynamic intermediate lifestyle of hemibiotroph made it possible for *G. boninense* to adapt and manipulate the host plant defense mechanisms which almost ultimately result in the plant succumbing to the infection. During the biotrophic phase (early infection), *G. boninense* survives by parasitically extracting nutrients from the living host plant while keeping it viable. The earliest symptoms are only visible on the foliage as the infection progresses up to 60-70%, reaching the later stage of necrotrophic phase with half of the stem base already ravaged (Chong et al., 2017a). BSR is the most common manifestation of *G. boninense* infection whereby it affects and decays the bole and produces multiple spears as well as fruiting bodies (Hushiarian et al., 2013).

Cultural practices including soil mounding, sanitation and trenching have been commonly applied as one of BSR disease management strategies although Chong et al. (2017b) has highlighted the risks of disease recurrence. Chemical control using

fungicides is the next choice of controlling BSR in oil palm. Studies on systemic fungicide such as hexaconazole have shown strong inhibition towards *Ganoderma* growth (Maluin et al., 2019; Mustafa et al., 2018). The application of fungicides on fields using trunk injection technique was also shown to have positive impacts in reducing the BSR incidence (Jelani et al., 2018). However, growing concerns on the negative impacts of fungicides on beneficial soil microbes, health and environmental hazards as well as development of fungicides resistance results in the intense need for green sustainable alternatives. As a result, anti-fungal compounds synthesized by bacteria (termed as biofungicides) are actively being studied as continuous efforts in reducing the use of synthetic fungicides (Lee et al., 2018; Pramudito et al., 2018). Alternatively, biocontrol agents (BCA) of soil-borne *Trichoderma spp.* have been numerously proven to exhibit antagonistic effect on *G. boninense* growth while promoting plant resistance against the fungal infection (Musa et al., 2018; Alizadeh et al., 2014; Priwiratama and Susanto, 2014). However, little is known about the oil palm defense mechanisms during different infection phases staged by this hemibiotroph pathogen, *G. boninense*.

The most promising molecular approach in managing *Ganoderma*-infected palm trees as well as minimizing the disease progress caused by *G. boninense* is most likely through identifying the infection at earlier stage (Kushairi et al., 2019). Phytohormones crosstalk of salicylic acid (SA) and jasmonic acid- ethylene (JA-ET) signalling has been widely studied to determine the fine-tuning defense response against biotrophic and necrotrophic infection phases, respectively (Häffner et. al., 2015). However, there is poor molecular information available explaining the defense regulation by transcription factors (TFs) during transition from biotrophic to necrotrophic state. The fragmentary information on molecular interactions between plant and hemibiotroph should be explicated to identify the key molecular mechanisms of both pathogenesis and plant immunity. A recent high-throughput RNA-seq analysis has pointed out the counter-act defense mechanisms executed by plant during transition from biotrophic to necrotrophic phase (Bahari et al., 2018). In the attempt of finding novel phase-specific biomarkers, this study focus on the regulation of defense-related TFs.

TFs are the 'master switches', which regulate the downstream genes transcription processes. Alteration of the genes expression by the TFs may either over-express or suppress different stress-related genes (Ng et al., 2018; Tsuda and Somssich, 2015). These pathogen-defensive genes are responsible in the production of antimicrobial secondary metabolites and antifungal proteins (Pandey et al., 2016; Alves et al., 2014). There are six major families of TFs recognized to regulate defense response against pathogens including NAC, bHLH, MYB, WRKY, AP2/ERF and bZIP (Ng et al., 2018). To date, no study has reported comprehensive transcriptomic profiling of oil palm defense-related TFs against *G. boninense*. Recognizing the infection phase-specific TFs may enables researchers to underpin the regulation of downstream target genes and offer opportunities to tackle or attenuate the progress of infection. The TFs can be used as biomarkers in plant breeding and engineering to develop *Ganoderma*-resistant palms. Thus, this study aims to recognize specific oil palm TFs as 'key' biomarkers associated with biotrophic and necrotrophic infection phases of hemibiotroph *G. boninense*.

The objectives of this study are;

- 1. To perform high-throughput transcript analysis via Next Generation Sequencing (NGS) and to determine differentially expressed genes (DEGs) of TFs during early infection of *G. boninense*
- 2. To identify oil palm TFs regulated specifically under biotic stress via quantitative Real-Time PCR (qPCR)
- 3. To carry out molecular characterization of an oil palm TF with potential in regulating defense response during the biotrophic and necrotrophic infection phase of *G. boninense*, respectively.



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The author, Nurshafika binti Mohd Sakeh was born on 24<sup>th</sup> of December, 1989 in, Kuala Lumpur, Malaysia. She received her early education at Sekolah Kebangsaan Dato' Demang Hussin, Melaka and attended Sekolah Menengah Kebangsaan Bukit Katil, Melaka before being accepted to Sekolah Menengah Sains Muar, Johor for her secondary education. She completed her matriculation level at Kolej Matrikulasi Perak (KMPK) before pursuing her study at tertiary level in Universiti Putra Malaysia (UPM). Shafika graduated with Bachelor of Science in Biochemistry (Hons.) in year 2010. During her degree study, she was honored with the Best Thesis and Best Final Year Project in Biochemistry for her final year project entitled the "Development of An Inhibitive Enzyme Assay For Heavy Metals Using Plant Protease". The awards were presented by the Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences (FBSB), UPM and by the Malaysian Society for Biochemistry and Molecular Biology (MSBMB). In 2011, she was awarded with two scholarships to pursue her Master's degree which were Graduate Research Fellowship (GRF) by UPM as well as MyBrain15 by the Ministry of Education, Malaysia. She completed her Master's study in the field of Animal Cell Biotechnology at FBSB, UPM under supervision of Assoc. Prof. Dr. Syahida Ahmad. Shafika started her Doctoral study on February 2015 and successfully graduated on January 2020 in the field of Molecular Biotechnology. She enrolled her full-time PhD study under the Institute of Tropical Agriculture and Food Security, UPM and was supervised by Prof. Datin Dr. Siti Nor Akmar Abdullah. Her Doctoral study was fully funded by MyPhD scholarship from the Ministry of Education, Malaysia.

## LIST OF PUBLICATIONS

## Journal

- Mohamad Nazri Abdul Bahari, **Nurshafika Mohd Sakeh**, Siti Nor Akmar Abdullah, Redzyque Ramza Ramli and Saied Kadkhodaei. (2018). Transciptome profiling at early infection of *Elaeis guineensis* by *Ganoderma boninense* provides novel insights on fungal transition from biotrophic to necrotrophic phase. *BMC Plant Biology*, 18(1): 377 (**Joined first author**)
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# **Oral Paper**

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Redzyque Ramza Ramli, Siti Nor Akmar Abdullah, Mohamad Nazri Abdul Bahari, **Nurshafika Mohd Sakeh**. (2017). Expression of Selected Groups of Pathogenesis-related (PR) Proteins at Early Stage of *Ganoderma boninense* Infection of Oil Palm (*Elaeis Guineensis* Jacq.) Seedlings. Universiti Putra Malaysia (pp 175-180). Perpustakaan Negara Malaysia **ISBN 978-967-960-427-6.** 

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# Poster

Nurshafika Mohd Sakeh, Siti Nor Akmar Abdullah, Mohamad Nazri Abdul Bahari, Azzreena Mohamad Azzeme, Noor Azmi Shaharuddin, Idris Abu Seman. (2018). Novel Transcription Factors Regulating Oil Palm (*Elaeis guineensis Jacq.*) Defense Response Against Hemibiotroph *Ganoderma boninense*. International Conference and Workshop on Comparative Genomics and Interactomics for Agriculture (ICCGIA) held in Universiti Putra Malaysia. -Best Poster Award



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