

SAFETY AND EFFICACY OF RECOMBINANT NEWCASTLE DISEASE VIRUS EXPRESSING HUMAN INTERLEUKIN-12 AS A POTENTIAL VACCINE IN BREAST CANCER

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By

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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

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In this developing era, breast cancer still remains a life-threatening disease globally. However, oncolytic virotherapy has taken over interest as a promising non-conventional alternative to treating breast cancers. Newcastle disease virus (NDV), an avian paramyxovirus has been demonstrated with significant oncolytic activity against cancer based on numerous preclinical studies. Today, genetically modified viruses coding for immunomodulatory agents, such as cytokines or chemokines, have come into focus. Such engineered viruses are able to promote efficient immune responses against tumour cells. The overall of this project aims to study the effects of a recombinant NDV expressing human interleukin 12 (rAF-IL12) in the apoptotic and metastatic process in MCF7 and MDA-MB231 human breast cancer cell lines. The parental NDV AF2240 was used as a positive control in this study. Notably, rAF-IL12 was able to maintain its stability when passaged in specific pathogen free (SPF) eggs up to ten passages. Furthermore it is considered safe as it selectively induced cytotoxic effects in chicken and breast cancer cell lines while sparing non-cancerous breast cell line as demonstrated through the MTT assay. The stability of each passaged rAF-IL12 was verified via haemagglutination assay (HA), mean death time (MDT) and intracerebral pathogenicity index (ICPI), while the IL12 was quantified through Enzyme-Linked Immunosorbent Assay (ELISA). Comparable to AF2240, rAF-IL12 was also able to induce apoptosis significantly based on several apoptotic assays. Both rAF-IL12 and AF2240 managed to increase the percentage of G2/M and S phase in the cell cycle analysis while inducing the percentage of apoptosis. Although both AF2240 and rAF-IL12 demonstrated comparable in vitro apoptosis results, rAF-IL12 possessed significant (p<0.05) antimetastatic activity in comparison to AF2240 based on metastasis related assays including the in vitro scratch assay, migration/invasion assay, human umbilical vein endothelial cell (HUVEC) tube formation and rat aortic ring assay. Additionally, to further evaluate the anti-tumour and anti-metastatic mechanism of rAF-IL12, in vivo studies were conducted using 4T1-challenged BALB/c mice as a model of this study. The rAF-IL12 was proven to function as an improved tumour vaccine as it significantly (p<0.05) reduced the size of tumour in comparison to the parental AF2240 virus. Apoptotic results showed that the number of cancer cells in the tumour significantly (p<0.05) reduced after 28 days of intra-tumoural treatment with rAF-IL12 (2^7 HAU). In addition, rAF-IL12 was able to inhibit the migration of cancer cells to other vital organs as opposed to the untreated group. To further elucidate the apoptotic and anti-metastatic mechanism of rAF-IL12 at molecular level, NanoString nCounter was conducted. Even though both AF2240 and rAF-IL12 exhibited similar mechanism of action, rAF-IL12 was more potent than AF2240 in terms of apoptosis and anti- metastasis activity. In conclusion, both rAF-IL12 and AF2240 were able to inhibit the proliferation of breast cancer cells and induce apoptosis in the *in vitro* studies; however, rAF-IL12 was able to demonstrate significant (p<0.05) improved functionality in comparison to AF2240 alone based on the *in vivo* studies. This proved that interleukin 12 was able to increase the immune response against tumour cells by inducing cell death, anti-angiogenesis and anti-metastasis effects, further improving the function of AF2240 as a potential vaccine in breast cancer.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

KESELAMATAN DAN KECEKAPAN REKOMBINAN VIRUS PENYAKIT SAMPAR MENGEKSPRESI INTERLEUKIN-12 MANUSIA SEBAGAI VAKSIN BERPOTENSI DI DALAM KANSER PAYU DARA

Oleh

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Dalam era membangun ini, kanser payudara masih menjadi penyakit yang mengancam nyawa di seluruh dunia. Walaubagaimanapun, terapi viro onkolitik telah mengambil alih kepentingan sebagai alternatif yang tidak konvensional bagi merawat kanser payudara. Virus penyakit sampar (NDV) merupakan paramyxovirus burung telah menunjukkan aktiviti onkolitik yang ketara terhadap kanser berdasarkan pelbagai kajian pra-klinikal. Pada masa kini, virus pengubahsuaian genetik untuk ejen imunomodulator, seperti sitokin atau chemokin telah menjadi tumpuan. Ini adalah kerana virus kejuruteraan itu dapat menggalakkan tindak balas imun yang cekap terhadap sel-sel tumor. Projek ini bertujuan untuk mengkaji kesan NDV rekombinan yang mengekspresi interleukin 12 manusia (rAF-IL12) melalui proses apoptotik dan metastatik dalam sel-sel kanser payudara MCF7 dan MDA-MB231. NDV AF2240 digunakan sebagai pemalar positif dalam kajian ini. Dilihat rAF-IL12 mampu mengekalkan kestabilannya apabila dilancarkan dalam telur bebas patogen spesifik (SPF) hingga sepuluh petikan dan keselamatan secara selektif melalui kesan sitotoksik di dalam sel-sel kanser ayam dan payudara tanpa memberi kesan sitotoksik pada sel payudara bukan kanser seperti yang ditunjukkan melalui asai MTT. Kestabilan setiap rAF-IL12 disahkan melalui asai hemaglutinan (HA), purata waktu kematian (MDT) and indeks patogenisiti antara serebral (ICPI), manakala interleukin 12 dikuantiti melalui asai imunoserapan berkaitan enzim (ELISA). Sebanding dengan AF2240, raf-IL12 juga dapat menyebabkan apoptosis dengan ketara berdasarkan beberapa ujian apoptosis. Kedua-dua raf-IL12 dan AF2240 berjaya meningkatkan peratusan fasa G2/ M dan S dalam analisis kitaran sel sambil meningkatkan peratusan apoptosis. Walaupun kedua-dua AF2240 dan rAF-IL12 menunjukkan hasil apoptosis in vitro yang sebanding, rAF-IL12 mempunyai aktiviti anti-metastatik yang ketara (p<0.05) berbanding AF2240 berdasarkan ujian metastasis berkaitan termasuk asai gores in vitro, asai migrasi/ invasi, pembentukan tiub human umbilical vein endothelial cell (HUVEC) dan asai cincin aorta tikus. Selain itu, untuk menilai lagi mekanisma anti-tumor dan anti-metastatik rAF-IL12, kajian in vivo dijalankan menggunakan tikus BALB/ c yang dicabar 4T1 sebagai model kajian ini. Terbukti rAF-IL12 berfungsi sebagai vaksin tumor yang lebih baik kerana ia secara signifikan (p<0.05) dapat mengurangkan saiz tumor secara ketara berbanding dengan virus AF2240. Keputusan apoptotik menunjukkan bahawa bilangan sel kanser dalam tumor berkurangan secara signifikan (p<0.05) 28 hari selepas rawatan intra-tumoral dengan raf-IL12 (2⁷ HAU). Di samping itu, rAF-IL12 mampu menghalang penghijrahan sel-sel kanser ke organ penting berbanding kumpulan yang tidak dirawat. Bagi menerangkan mekanisme apoptotik dan anti-metastatik raf-IL12 pada tahap molekul, NanoString nCounter telah dijalankan. Walaupun kedua-dua AF2240 dan rAF-IL12 mempamerkan mekanisma tindakan yang sama, raf-IL12 lebih berkesan secara signifikan (p<0.05) berbanding AF2240 dari segi apoptosis dan aktiviti anti-metastasis. Kesimpulannya, kedua-dua rAF-IL12 dan AF2240 dapat menghalang percambahan selsel kanser payudara dan mendorong apoptosis dalam kajian in vitro, bagaimanapun, rAF-IL12 dapat menunjukkan fungsi peningkatan yang lebih ketara (p<0.05) berbanding dengan AF2240 sahaja berdasarkan kajian in vivo. Ini membuktikan bahawa interleukin 12 dapat meningkatkan tindak balas imun terhadap sel tumor dengan menggalakkan kematian sel, anti-angiogenesis dan kesan anti-metastasis, sambil meningkatkan lagi fungsi AF2240 sebagai vaksin berpotensi terhadap kanser payudara.

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LIST OF ABBREVIATIONS

AO Acridine Orange

ANG-2 Angiotensin 2

ATCC Animal Tissue Culture Collection

ATM Ataxia telangiectasia mutated

BAX Bcl-2 Associated X-Protein

BCL2 B-cell lymphoma 2

BRCA1 Breast Cancer 1

BrdU Bromodeoxyuridine

CO₂ Carbon dioxide

CXCL-1 Chemokine Ligand 1

EGFP Enhanced green fluorescent protein

F Fusion glycoprotein

DNA Deoxyribonucleic acid

ELISA Enzyme-Linked Immunosorbent Assay

DMEM Dulbecco's Modified Eagle Medium

FITC Fluorescent isothiocyanate

HA Haemagglutination

HEGF Human Endothelial Growth Factor

HN Haemagglutinin-neuraminidase glycoprotein

HUVEC Human Umbilical Vein Endothelial Cells

IC₅₀ Half maximal inhibitory concentration

IACUC Institutional Animal Care and Use Committee

ICPI Intracerebral Pathogenecity Index

IFN-γ Interferon-gamma

IL-12 Interleukin-12

IL-2 Interleukin-2

JC-1 5,5',6,6'-tetrachloro-1,1',3,3'-

tetraethylbenzimidazolylcarbocyanine iodide

L Large polymerase protein

M Matrix protein

MDT Mean Death Time

MTT 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide

N Nucleocapsid protein

NDV Newcastle Disease Virus

OIE World Organisation for Animal Health

Phosphoprotein

PBS Phosphate buffer saline

PCR Polymerase chain reaction

PI Propidium Iodide

RNA Ribonucleic acid

RNAse Ribonuclease

RT-QPCR Real Time quantitative PCR

SSC Saline-sodium citrate

SPSS Statistical Package for the Social Sciences

TMB 3,3',5,5'-Tetramethylbenzidine

TRAIL TNF-related apoptosis inducing ligand

TUNEL Terminal deoxynucleotidyl transferase dUTP nick end

labeling

VEGF A Vascular endothelial growth factor A

CHAPTER 1

INTRODUCTION

Cancer is defined as uncontrollable cell division and the invasion ability of these cells to other tissues causing tumour mass formation, metastasis and vascularization (Vogelstein and Kinzler, 2004). In addition, angiogenesis (formation of new blood vessels) is also an important process for growth, development and to cause tumours to become malignant (Jyothi, 2012). After lung cancer, breast cancer ranks the second most common cancer estimating 1.7 million cases since 2012, worldwide and is the fifth cause of death (522,000 deaths) (Ferlay *et al.*, 2015). One of the main approaches to treat breast cancer is chemotherapy. Chemotherapy utilizes anti-breast cancer drugs either given intravenously or orally (Maughan *et al.*, 2010). The drugs travel through the bloodstream reaching breast cancer cells (Maughan *et al.*, 2010). However, chemotherapy induces cytotoxic effects to the cancer cells as well as normal cells (Jyothi, 2012). In addition, resistance towards the drug(s), disturbed biodistribution and clearance of the drug may arise (Alfarouk *et al.*, 2015; Jyothi, 2012). Therefore, the development of new therapeutic strategies is required to overcome the latter complications.

Oncolytic virotherapy is an adjunctive strategy to minimize tumour burden through selective virus replication in rapidly proliferating cells (Aurelian, 2013). It is currenty studied to be used as a future novel strategy in cancer treatments that can be later combined with conventional therapies for improved efficacy. Various viruses from different families are currently being studied as oncolytic agents at pre-clinical and clinical levels (Haddad, 2017; Singh *et al.*, 2012; Sugiyama *et al.*, 2012). Newcastle disease virus (NDV), also known as avian paramyxovirus serotype 1 (APMV-1), is a single negative-stranded RNA virus of nearly 15 kb, which belongs to the *Avulavirus* genus member in the family of *Paramyxoviridae*. NDV is a promising oncotherapeutic agent for human cancers as it is normally non-pathogenic to humans (Ghrici *et al.*, 2013). AF2240 is a viscerotropic velogenic strain of NDV which was isolated during an outbreak in the Malaysia in the 1960s resulting in high mortality rate and is now being used as the challenge virus in vaccine trials in Malaysia (Molouki *et al.*, 2011).

NDV is known to efficiently infect and kill cancer cells and is consequently being investigated as a novel cancer therapy (oncolytic virotherapy) (Schirrmacher *et al.*, 2014). The first oncolytic study on NDV was reported back in 1965 by Cassel and Garret (1965) whereby the virus was studied as an antineoplastic agent on Ehrlich ascites carcinoma of the mouse, the human adenocarcinoma and HeLa cells. The virus kills the human cancer cells by apoptosis *via* either the intrinsic or extrinsic pathway resulting in the cell death (Ekert and Vaux, 1997). NDV possesses several advantages over other oncolytic viruses such as poxvirus, measles, HSV-1 or reovirus (Zamarin and Palese, 2012). It is considered to be safe for use in cancer treatments since it is an avian and non-pathogenic to humans and does not affect the immune system. The problem of pre-existing immunity can be overcome as 96% of the human population is seronegative towards NDV (Charan *et al.*, 1981; Miller *et al.*, 1971).

Vaccines are known to be highly effective. The production of viral vaccines involves either modifying living attenuated virus or using viruses which are chemically inactivated. The drawback of utilising such conventional methods in vaccine development relates to safety, efficiency and cost of the vaccine (Huang *et al.*, 2003). Therefore, the development of genetically engineered viruses has come into focus as such viruses improve in safety, efficiency and cost.

The first study of genetically engineered virus was conducted to induce immune response against hepatitis B in chimpanzees using recombinant engineered to exhibit hepatitis B surface antigen in animal cells (Moss *et al.* 1984). Ever since then, several studies have been made in the fields of various viral vectors which were evaluated for immunotherapeutic and vaccine applications for a wide range of diseases (Choi and Chang, 2013). Methods of producing infectious paramyxoviruses from c-DNA clones (reverse genetics) have been developed during the last 20 years (Schirrmacher *et al.*, 2014). Some of the genetically engineered NDV vaccines reported include Hitchner B1strain armed with IL2 in treating malignant melanoma (Zamarin et al., 2009), LaSota strain armed with IL-2, TNF-related apoptosis inducing ligand (TRAIL), IL-2-TRAIL, or enhanced green fluorescent protein (EGFP) as anti-neoplastic (Bai *et al.*, 2014) and LaSota strain armed with IFN-λ1 against gastric adenocarcinoma cells (Bu *et al.*, 2016). However, no studies have been reported on NDV AF2240 armed with IL12 against breast cancer yet.

In animal models and in clinical testing, cytokines have been found to be effective immunomodulators, suggesting that anti-tumour effect is closely related to its expression levels (Pan et al., 2016). Interleukin-12 (IL12) is a heterodimeric cytokine produced specifically by phagocytic cells and antigen-presenting cells. It exhibits a crucial role in activating anti-tumour immunity as it increases differentiation of T-cells, initiates natural killer cells as well as functions to inhibit angiogenesis and metastatic process enhancing anti-tumour activity in pre-clinical models (Derin et al., 2018). Additionally, IL12 was able to efficiently eradicate experimental liver cancer through intra-tumoural injections while reducing systemic toxicity (Sangro et al., 2004). Promising anti-tumour properties of IL12 have been proven on cancers such as liver, colorectal and pancreatic (Sangro et al., 2004), renal cancer (Gollob et al., 2000) and gastrointestinal primary malignancy (Lenzi et al., 2002). However, further in-depth mechanisms including anti-metastatic effects of IL12 are yet to be revealed, especially in breast cancer. The safety profile of NDV AF2240 expressing IL12 should also be tested although the anti-cancer activities of both AF2240 and IL12 are promising. The problem statement of this study is that IL12 has potent anti-tumour activities with strong clinical relevance to cancer therapy. However, the operating mechanism of NDV armed with IL12 is not fully understood. Other than that, the anti-tumour effect of NDV armed with IL12 in breast cancer has not been reported yet. Therefore, the hypothesis of this study is; the newly developed recombinant NDV expressing human IL12 genes is safe and stable with the ability to selectively replicate in cancer cells and induce anti-tumour effect. . Hence, the novel recombinant NDV armed with human IL12 as a potential cancer vaccine in breast cancer studies will be the first reported.

The objectives of this study are:

- 1. To assess the pathogenicity and stability of rAF-IL12 in chickens through multiple passaging *in vivo*.
- 2. To investigate the *in vitro* cytotoxic effects and the mechanism of rAF-IL12 in the induction of cell death and anti-metastatic abilities in MCF-7 and MDA-MB231 breast cancer cell lines.
- 3. To determine the *in vivo* anti-tumour activity of rAF-IL12 in 4T1-breast cancer challenged mice.



REFERENCES

- Abu, N., Akhtar, M. N., Yeap, S. K., Lim, K. L., Ho, W. Y., Zulfadli, A. J., et al. & Alitheen, N. B. (2014). Flavokawain A induces apoptosis in MCF-7 and MDA-MB231 and inhibits the metastatic process in vitro. PLoS One, 9(10), e105244.
- Ahmad, A. S., Ormiston-Smith, N., & Sasieni, P. D. (2015). Trends in the lifetime risk of developing cancer in Great Britain: comparison of risk for those born from 1930 to 1960. *British journal of cancer*, 112(5), 943-947.
- Ahmad, U., Ahmed, I., Keong, Y. Y., Abd Manan, N., & Othman, F. (2015). Inhibitory and apoptosis-inducing effects of Newcastle disease virus strain AF2240 on mammary carcinoma cell line. *BioMed research international*, 2015.
- Alabsi, A. M., Bakar, S. A. A., Ali, R., Omar, A. R., Bejo, M. H., Ideris, A., & Ali, A. M. (2011). Effects of Newcastle disease virus strains AF2240 and V4-UPM on cytolysis and apoptosis of leukemia cell lines. *International journal of molecular sciences*, 12(12), 8645-8660.
- Alabsi, A. M., Ali, R., Ideris, A., Omar, A. R., Bejo, M. H., Yusoff, K., & Ali, A. M. (2012). Anti-leukemic activity of Newcastle disease virus strains AF2240 and V4-UPM in murine myelomonocytic leukemia in vivo. *Leukemia research*, 36(5), 634-645.
- Alatrash, G., Hutson, T. E., Molto, L., Richmond, A., Nemec, C., Mekhail, T., Bukowski, R. M., et al. (2004). Clinical and immunologic effects of subcutaneously administered interleukin-12 and interferon alfa-2b: phase I trial of patients with metastatic renal cell carcinoma or malignant melanoma. *Journal of clinical oncology*, 22(14), 2891-2900.
- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., & Walter, P. (2002). The cytoskeleton and cell behavior.
- Albini, A., & Benelli, R. (2007). The chemoinvasion assay: a method to assess tumor and endothelial cell invasion and its modulation. *Nature Protocols*, 2, 504–511.
- Alexander, D. J. (2001). Newcastle disease. British poultry science, 42(1), 5-22.
- Ali, R., Alabsi, A. M., Ali, A. M., Ideris, A., Omar, A. R., Yusoff, K., & Saif-Ali, R. (2011). Cytolytic effects and apoptosis induction of Newcastle disease virus strain AF2240 on anaplastic astrocytoma brain tumor cell line. *Neurochemical research*, *36*(11), 2051.
- Alfarouk, K. O., Stock, C. M., Taylor, S., Walsh, M., Muddathir, A. K., Verduzco, D, et al. & Reshkin, S. J. (2015). Resistance to cancer chemotherapy: failure in drug response from ADME to P-gp. Cancer cell international, 15(1), 71.

- Alkayyal, A. A., Mahmoud, A. B., & Auer, R. C. (2016). Interleukin-12-expressing oncolytic virus: a promising strategy for cancer immunotherapy. *Journal of Taibah University Medical Sciences*, 11(3), 187-193.
- Anand, P., Kunnumakara, A. B., Sundaram, C., Harikumar, K. B., Tharakan, S. T., Lai, O. S., Aggarwal, B. B., et al. (2008). Cancer is a preventable disease that requires major lifestyle changes. *Pharmaceutical research*, 25(9), 2097-2116.
- Andtbacka, R. H., Kaufman, H. L., Collichio, F., Amatruda, T., Senzer, N., Chesney, J., Milhem, M., et al. (2015). Talimogene laherparepvec improves durable response rate in patients with advanced melanoma. *Journal of clinical oncology*, *33*(25), 2780-2788.
- Arnaoutova, I., George, J., Kleinman, H. K., & Benton, G. (2009). The endothelial cell tube formation assay on basement membrane turns 20: state of the science and the art. *Angiogenesis*, 12(3), 267-274.
- Arnaoutova, I., & Kleinman, H. K. (2010). In vitro angiogenesis: endothelial cell tube formation on gelled basement membrane extract. *Nature protocols*, 5(4), 628.
- Arnold, A., & Papanikolaou, A. (2005). Cyclin D1 in breast cancer pathogenesis. *Journal of clinical oncology*, 23(18), 4215-4224.
- Arur, S., Uche, U. E., Rezaul, K., Fong, M., Scranton, V., Cowan, A. E., Han, D. K., et al. (2003). Annexin I is an endogenous ligand that mediates apoptotic cell engulfment. *Developmental cell*, 4(4), 587-598.
- Asad, S. (2017). Exploring the molecular mechanism (s) involved in Wolbachia-Aedes aegypti-dengue virus interactions.
- Atkins, M. B., Robertson, M. J., Gordon, M., Lotze, M. T., DeCoste, M., DuBois, J. S., Mier, J. W. et al. (1997). Phase I evaluation of intravenous recombinant human interleukin 12 in patients with advanced malignancies. *Clinical Cancer Research*, 3(3), 409-417.
- Aubry, J., Blaecke, A., Lecoanet-Henchoz, S., Jeannin, P., Herbault, N., Caron, G., Moine, V., & Bonnefoy, J. (1999) Annexin V used for measuring apoptosis in the early events of cellular cytotoxicity. *Cytometry*, *37*, 197–204.
- Aurelian, L. (2013). Oncolytic virotherapy: the questions and the promise. Oncolytic virotherapy, 2, 19.
- Ayllon, J., García-Sastre, A., & Martínez-Sobrido, L. (2013). Rescue of recombinant Newcastle disease virus from cDNA. *Journal of visualized experiments: JoVE*, (80).
- Bai, F. L., Yu, Y. H., Tian, H., Ren, G. P., Wang, H., Zhou, B., et.al & Li, D. S. (2014). Genetically engineered Newcastle disease virus expressing interleukin-2 and

- TNF-related apoptosis-inducing ligand for cancer therapy. Cancer biology & therapy, 15(9), 1226-1238.
- Barajas, M., Mazzolini, G., Genové, G., Bilbao, R., Narvaiza, I., Schmitz, V., Prieto, J., et al. (2001). Gene therapy of orthotopic hepatocellular carcinoma in rats using adenovirus coding for interleukin 12. *Hepatology*, *33*(1), 52-61.
- Bartlett, D. L., Liu, Z., Sathaiah, M., Ravindranathan, R., Guo, Z., He, Y., & Guo, Z. S. (2013). Oncolytic viruses as therapeutic cancer vaccines. *Molecular cancer*, *12*(1), 103.
- Bauman, G., Charette, M., Reid, R., Sathya, J., & of Cancer, T. R. G. G. (2005). Radiopharmaceuticals for the palliation of painful bone metastases—a systematic review. *Radiotherapy and oncology*, 75(3), 258-E1.
- Beral, V., Bull, D., Doll, R., & Key, T. (1997). Breast cancer and hormone replacement therapy: collaborative reanalysis of data from 51 epidemiological studies of 52 705 women with breast cancer and 108 411 women without breast cancer. *The Lancet*, 350(9084), 1047.
- Bian, J., Wang, K., Kong, X., Liu, H., Chen, F., Hu, M., Meng, S., et al. (2011). Caspase-and p38-MAPK-dependent induction of apoptosis in A549 lung cancer cells by Newcastle disease virus. *Archives of virology*, 156(8), 1335-1344.
- Binder, B. J., & Landman, K. A. (2009). Exclusion processes on a growing domain. *Journal of theoretical biology*, 259(3), 541-551.
- Biroccio, A., Candiloro, A., Mottolese, M., Sapora, O., Albini, A., Zupi, G., & Del Bufalo, D. (2000). Bcl-2 overexpression and hypoxia synergistically act to modulate vascular endothelial growth factor expression and in vivo angiogenesis in a breast carcinoma line. *The FASEB Journal*, 14(5), 652-660.
- Bossy-Wetzel, E., & Green, D. R. (2000). Detection of apoptosis by annexin V labeling. *Methods in enzymology*, 322, 15-18.
- Boyden, S. (1962). The chemotactic effect of mixtures of antibody and antigen on polymorphonuclear leucocytes. *Journal of Experimental Medicine*, 115, 453–466.
- Bray, F., & Møller, B. (2006). Predicting the future burden of cancer. *Nature Reviews Cancer*, 6(1), 63-74.
- Bridle, B.W., Boudreau, J.E., Lichty, B.D., et al. (2009). Vesicular stomatitis virus as a novel cancer vaccine vector to prime antitumor immunity amenable to rapid boosting with adenovirus. *Molecular Therapy*, *17*, 1814–1821.
- Brown, I., Shalli, K., McDonald, S. L., Moir, S. E., Hutcheon, A. W., Heys, S. D., & Schofield, A. C. (2004). Reduced expression of p27 is a novel mechanism of docetaxel resistance in breast cancer cells. *Breast Cancer Research*, 6(5), R601.

- Bu, X., Li, M., Zhao, Y., Liu, S., Wang, M., Ge, J., et. al & Yan, Y. (2016). Genetically engineered Newcastle disease virus expressing human interferon-λ1 induces apoptosis in gastric adenocarcinoma cells and modulates the Th1/Th2 immune response. Oncology reports, 36(3), 1393-1402.
- Buijs, P., van Nieuwkoop, S., Vaes, V., Fouchier, R., van Eijck, C., & Hoogen, B. V. D. (2015). Recombinant immunomodulating lentogenic or mesogenic oncolytic Newcastle disease virus for treatment of pancreatic adenocarcinoma. *Viruses*, 7(6), 2980-2998.
- Bukreyev, A., Huang, Z., Yang, L., Elankumaran, S., Claire, M., Murphy, B. R., Samal, & S. K., Collins, P. L. (2005). Recombinant Newcastle disease virus expressing a foreign viral antigen is attenuated and highly immunogenic in primates. *Journal of Virology*, 79, 13275–13284.
- Caruso, M., Pham-Nguyen, K., Kwong, Y. L., Xu, B., Kosai, K. I., Finegold, M., Chen, S. H., et al. (1996). Adenovirus-mediated interleukin-12 gene therapy for metastatic colon carcinoma. *Proceedings of the National Academy of Sciences*, 93(21), 11302-11306.
- Cassel, W. A., & Garrett, R. E. (1965). Newcastle disease virus as an antineoplastic agent. *Cancer*, 18(7), 863-868.
- Chan, K.M., Rajab, N.F., Ishak, M.H.A., Ali, A.M., Yusoff, K., Din, L.B., Inayat-Hussain, S.H. (2006) Goniothalamin induces apoptosis in vascular smooth muscle cells. *Chemico-Biological Interactions*, 159, 129–140.
- Chan, L. L., Wilkinson, A. R., Paradis, B. D., & Lai, N. (2012). Rapid image-based cytometry for comparison of fluorescent viability staining methods. *Journal of fluorescence*, 22(5), 1301-1311.
- Chan, L. L. Y., Kuksin, D., Laverty, D. J., Saldi, S., & Qiu, J. (2015). Morphological observation and analysis using automated image cytometry for the comparison of trypan blue and fluorescence-based viability detection method. *Cytotechnology*, 67(3), 461-473.
- Chen, H. (2005). Boyden chamber assay. Methods in Molecular Biology, 294, 15-22.
- Chen, W. L., Kuo, K. T., Chou, T. Y., Chen, C. L., Wang, C. H., Wei, Y.H., & Wang, L. S. (2012). The role of cytochrome c oxidase subunit Va in nonsmall cell lung carcinoma cells: association with migration, invasion and prediction of distant metastasis. *BMC Cancer*, 12, 273.
- Choi, Y., & Chang, J. (2013). Viral vectors for vaccine applications. *Clinical and experimental vaccine research*, 2(2), 97-105.
- Claesson- Welsh, L., & Welsh, M. (2013). VEGFA and tumour angiogenesis. *Journal of internal medicine*, 273(2), 114-127.

- Clarke, R.G., Lund, E.K., Johnson, I.T., & Pinder, A.C. (2000). Apoptosis can be detected in attached colonic adenocarcinoma HT29 cells using Annexin V binding, but not by TUNEL assay or sub-G0 DNA content. *Cytometry*, *39*, 141–150.
- Coates, A. S., Winer, E. P., Goldhirsch, A., Gelber, R. D., Gnant, M., Piccart-Gebhart, M., Baselga, J., et al. (2015). Tailoring therapies—improving the management of early breast cancer: St Gallen International Expert Consensus on the Primary Therapy of Early Breast Cancer 2015. *Annals of oncology*, 26(8), 1533-1546.
- Cohen, G. M., Sun, X. M., Snowden, R. T., Dinsdale, D., & Skilleter, D. N. (1992). Key morphological features of apoptosis may occur in the absence of internucleosomal DNA fragmentation. *Biochemical Journal*, 286(2), 331-334.
- Colombo, M. P., & Trinchieri, G. (2002). Interleukin-12 in anti-tumor immunity and immunotherapy. *Cytokine & growth factor reviews*, 13(2), 155-168.
- Gottrup, F., Ågren, M. S., & Karlsmark, T. (2000). Models for use in wound healing research: a survey focusing on in vitro and in vivo adult soft tissue. *Wound Repair and Regeneration*, 8(2), 83-96.
- Coolen, N. A., Schouten, K. C., Boekema, B. K., Middelkoop, E., & Ulrich, M. M. (2010). Wound healing in a fetal, adult, and scar tissue model: a comparative study. *Wound Repair and Regeneration*, 18(3), 291-301.
- Cooper, S. (2000). The continuum model and G1-control of the mammalian cell cycle. In *Progress in cell cycle research*(pp. 27-39). Springer, Boston, MA.
- Cuadrado-Castano, S., Sanchez-Aparicio, M. T., García-Sastre, A., & Villar, E. (2015). The therapeutic effect of death: Newcastle disease virus and its antitumor potential. *Virus research*, 209, 56-66.
- Darzynkiewicz, Z., Bruno, S., Del Bino, G., Gorczyca, W., Hotz, M. A., Lassota, P., & Traganos, F. (1992). Features of apoptotic cells measured by flow cytometry. *Cytometry*, *13*(8), 795-808.
- Dasgupta, A., Nomura, M., Shuck, R., & Yustein, J. (2016). Cancer's Achilles' Heel: Apoptosis and Necroptosis to the Rescue. *International journal of molecular sciences*, 18(1), 23.
- Daud, A. I., DeConti, R. C., Andrews, S., Urbas, P., Riker, A. I., Sondak, V. K., Heller, R., et al. (2008). Phase I trial of interleukin-12 plasmid electroporation in patients with metastatic melanoma. *Journal of clinical oncology*, 26(36), 5896.
- De Laurentiis, M., Cancello, G., D'Agostino, D., et al. (2008). Taxane-based combinations as adjuvant chemotherapy of early breast cancer: a metaanalysis of randomized trials. *Journal of Clinical Oncology*, 26(1), 44-53.

- De Leeuw, O. S., Koch, G., Hartog, L., Ravenshorst, N., & Peeters, B. P. (2005). Virulence of Newcastle disease virus is determined by the cleavage site of the fusion protein and by both the stem region and globular head of the haemagglutinin–neuraminidase protein. *Journal of General Virology*, 86(6), 1759-1769.
- Del Vecchio, M., Bajetta, E., Canova, S., Lotze, M. T., Wesa, A., Parmiani, G., & Anichini, A. (2007). Interleukin-12: biological properties and clinical application. *Clinical Cancer Research*, *13*(16), 4677-4685.
- Dias, S., Boyd, R., & Balkwill, F. (1998). IL- 12 regulates VEGF and MMPs in a murine breast cancer model. *International Journal of Cancer*, 78(3), 361-365.
- Doudna, J. A., & Charpentier, E. (2014). The new frontier of genome engineering with CRISPR-Cas9. Science, 346(6213), 1258096.
- Du, T., Shi, G., Li, Y. M., Zhang, J. F., Tian, H. W., Wei, Y. Q., Yu, D. C., et al. (2014). Tumor-specific oncolytic adenoviruses expressing granulocyte macrophage colony-stimulating factor or anti-CTLA4 antibody for the treatment of cancers. *Cancer gene therapy*, 21(8), 340.
- Duda, D. G., Sunamura, M., Lozonschi, L., Kodama, T., Egawa, S. I., Matsumoto, G., S. et al. (2000). Direct in vitro evidence and in vivo analysis of the antiangiogenesis effects of interleukin 12. *Cancer research*, 60(4), 1111-1116.
- Early Breast Cancer Trialists' Collaborative Group. (2006). Effects of radiotherapy and of differences in the extent of surgery for early breast cancer on local recurrence and 15-year survival: an overview of the randomised trials. *The Lancet*, 366(9503), 2087-2106.
- Ekert, P. G., & Vaux, D. L. (1997). Apoptosis and the immune system. British medical bulletin, 53(3), 591-603.
- Elankumaran, S., Rockemann, D., & Samal, S. K. (2006). Newcastle disease virus exerts oncolysis by both intrinsic and extrinsic caspase-dependent pathways of cell death. *Journal of virology*, 80(15), 7522-7534.
- Elmore, S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic pathology*, 35(4), 495-516.
- Engeland, M.V., Nieland, L.J.W., Ramaekers, F.C.S, Schutte, B., & Reutelingsperger, C.P.M. (1998) Annexin V-Affinity assay: a review on an apoptosis detection system based on phosphatidylserine exposure. *Cytometry*, *31*, 1–9.
- Eskes, R., Antonsson, B., Osen-Sand, A., Montessuit, S., Richter, C., Sadoul, R., Martinou, J. C., et al. (1998). Bax-induced cytochrome C release from mitochondria is independent of the permeability transition pore but highly dependent on Mg2+ ions. *The Journal of cell biology*, *143*(1), 217-224.

- Fantozzi, A., & Christofori, G. (2006). Mouse models of breast cancer metastasis. *Breast Cancer Research*, 8(4), 212.
- Fakhari, S., Mahmoodi, M., Hosseini, J., Hosseini Zijoud, S. M., Khoshdel, A., Tahamtan, M., Hakhamaneshi, M. S., et al. (2014). Aloeemodin induces apoptosis through the up-regulation of fas in the human breast cancer cell line MCF-7. *Life Science Journal*, 11(2).
- Felcht, M., Fischer, J. C., Michels, M., Weinhold, M., & Zouboulis, C. C. (2005). Malignant melanoma--a medical students' viewpoint. *Journal der Deutschen Dermatologischen Gesellschaft= Journal of the German Society of Dermatology: JDDG*, 3(6), 421-430.
- Fellmann, C., & Lowe, S. W. (2014). Stable RNA interference rules for silencing. Nature cell biology, 16(1), 10.
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Bray, F., et al. (2015). Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *International journal of cancer*, *136*(5), E359-E386.
- Fidler, I. J. (2000). Angiogenesis and cancer metastasis. *Cancer journal (Sudbury, Mass.)*, 6, S134-41.
- Foglieni, C., Meoni, C., & Davalli, A. M. (2001). Fluorescent dyes for cell viability: an application on prefixed conditions. *Histochemistry and cell biology*, 115(3), 223-229.
- Ford, D., Easton, D. F., Stratton, M., Narod, S., Goldgar, D., Devilee, P., Sobol, H., et al. (1998). Genetic heterogeneity and penetrance analysis of the BRCA1 and BRCA2 genes in breast cancer families. *The American Journal of Human Genetics*, 62(3), 676-689.
- Freeman, A. I., Zakay-Rones, Z., Gomori, J. M., Linetsky, E., Rasooly, L., Greenbaum, E., Galun, E., et al. (2006). Phase I/II trial of intravenous NDV-HUJ oncolytic virus in recurrent glioblastoma multiforme. *Molecular therapy*, *13*(1), 221-228.
- Friedl, P. & Weigelin, B. (2008). Interstitial leukocyte migration and immune function. *Nature Immunology*, 9, 960–969.
- Jemal, A., Siegel, R., Xu, J., & Ward, E. (2010). Cancer statistics, 2010. *CA: a cancer journal for clinicians*, 60(5), 277-300.
- Galanis, E. (2010). Therapeutic potential of oncolytic measles virus: promises and challenges. *Clinical Pharmacology & Therapeutics*, 88, 620–625.
- Ganz, Patricia A., Lorna Kwan, Annette L. Stanton, Julienne E. Bower, and Thomas R. Belin. "Physical and psychosocial recovery in the year after primary treatment of breast cancer." *Journal of Clinical Oncology* 29, no. 9 (2011): 1101.

- Gaston, D. C., Odom, C. I., Li, L., Markert, J. M., Roth, J. C., Cassady, K. A., Parker, J. N., et al. (2013). Production of bioactive soluble interleukin-15 in complex with interleukin-15 receptor alpha from a conditionally-replicating oncolytic HSV-1. *PLoS One*, 8(11), 81768.
- Gerber, S. A., Moran, J. P., Frelinger, J. G., Frelinger, J. A., Fenton, B. M., & Lord, E. M. (2003). Mechanism of IL-12 mediated alterations in tumour blood vessel morphology: analysis using whole-tissue mounts. *British journal of cancer*, 88(9), 1453.
- Ghasemi, M., Emadian, O., Naghshvar, F., Bekhradnia, A., Abediankenari, S., Larijani, L. V., & Moghimpour, R. (2011). Immunohistochemical expression of vascular endothelial growth factor and its correlation with tumor grade in breast ductal carcinoma. *Acta Medica Iranica*, 49(12), 776-779.
- Ghrici, M., El Zowalaty, M., Omar, A. R., & Ideris, A. (2013). Induction of apoptosis in MCF-7 cells by the hemagglutinin-neuraminidase glycoprotein of Newcastle disease virus Malaysian strain AF2240. *Oncology reports*, 30(3), 1035-1044.
- Gollob, J. A., Mier, J. W., Veenstra, K., McDermott, D. F., Clancy, D., Clancy, M., & Atkins, M. B. (2000). Phase I trial of twice-weekly intravenous interleukin 12 in patients with metastatic renal cell cancer or malignant melanoma: ability to maintain IFN-γ induction is associated with clinical response. *Clinical Cancer Research*, 6(5), 1678-1692.
- Gottrup, F., Ågren, M. S., & Karlsmark, T. (2000). Models for use in wound healing research: a survey focusing on in vitro and in vivo adult soft tissue. *Wound Repair and Regeneration*, 8(2), 83-96.
- Gould, S. E., Junttila, M. R., & de Sauvage, F. J. (2015). Translational value of mouse models in oncology drug development. Nature medicine, 21(5), 431.
- Graaf, J. F., de Vor, L., Fouchier, R. A. M., & van den Hoogen, B. G. (2018). Armed oncolytic viruses: A kick-start for anti-tumor immunity. *Cytokine & growth factor reviews*.
- Gubler, U., Chua, A. O., Schoenhaut, D. S., Dwyer, C. M., McComas, W., Motyka, R., Familletti, P. C., et al. (1991). Coexpression of two distinct genes is required to generate secreted bioactive cytotoxic lymphocyte maturation factor. *Proceedings of the National Academy of Sciences*, 88(10), 4143-4147.
- Gyorffy, S., Palmer, K., Podor, T. J., Hitt, M., & Gauldie, J. (2001). Combined treatment of a murine breast cancer model with type 5 adenovirus vectors expressing murine angiostatin and IL-12: a role for combined anti-angiogenesis and immunotherapy. *The Journal of Immunology*, *166*(10), 6212-6217.
- Haas, C., Ertel, C., Gerhards, R., & Schirrmacher, V. (1998). Introduction of adhesive and costimulatory immune functions into tumor cells by infection with Newcastle Disease Virus. *International journal of oncology*, *13*(6), 1105-1120.

- Haddad, D. (2017). Genetically engineered vaccinia viruses as agents for cancer treatment, imaging, and transgene delivery. Frontiers in oncology, 7, 96.
- Haughland, R., (2002). Handbook of Fluorescent Probes and Research Products *Molecular Probes*, 9, 1-9.
- Hellums, E. K., Markert, J. M., Parker, J. N., He, B., Perbal, B., Roizman, B., ... & Gillespie, G. Y. (2005). Increased efficacy of an interleukin-12-secreting herpes simplex virus in a syngeneic intracranial murine glioma model. *Neuro-oncology*, 7(3), 213-224.
- Hernández-Verdugo, S., Guevara-González, R. G., Rivera-Bustamante, R. F., & Oyama, K. (2001). Screening wild plants of Capsicum annuum for resistance to pepper huasteco virus (PHV): presence of viral DNA and differentiation among populations. *Euphytica*, 122(1), 31-36.
- Holen, I., Speirs, V., Morrissey, B., & Blyth, K. (2017). In vivo models in breast cancer research: progress, challenges and future directions. *Disease models & mechanisms*, 10(4), 359-371.
- Hortobagyi, G. N. (1998). Treatment of breast cancer. *New England Journal of Medicine*, 339(14), 974-984.
- Huang, C. Y. H., Butrapet, S., Tsuchiya, K. R., Bhamarapravati, N., Gubler, D. J., & Kinney, R. M. (2003). Dengue 2 PDK-53 virus as a chimeric carrier for tetravalent dengue vaccine development. *Journal of virology*, 77(21), 11436-11447.
- Hutchinson, L. (2010). Breast cancer: challenges, controversies, breakthroughs.
- Imanishi, Y., Hu, B., Xiao, G., Yao, X., & Cheng, S. Y. (2011). Angiopoietin-2 promotes initial growth and survival of breast cancer metastases to the lung through the integrin-linked kinase (ilk)-akt-b cell lymphoma 2 (bcl-2) pathway. *Journal of Biological Chemistry*, jbc-M111.
- Janke, M., Peeters, B., De Leeuw, O., Moorman, R., Arnold, A., Fournier, P., & Schirrmacher, V. (2007). Recombinant Newcastle disease virus (NDV) with inserted gene coding for GM-CSF as a new vector for cancer immunogene therapy. *Gene therapy*, 14(23), 1639.
- Jamal, M. H., Ch'ng, W. C., Yusoff, K., & Shafee, N. (2012). Reduced Newcastle disease virus-induced oncolysis in a subpopulation of cisplatin-resistant MCF7 cells is associated with survivin stabilization. *Cancer cell international*, 12(1), 35.
- Kaufman, H. L., Kohlhapp, F. J., & Zloza, A. (2015). Oncolytic viruses: a new class of immunotherapy drugs. *Nature reviews Drug discovery*, *14*(9), 642.

- Keller, R. (2005). Cell migration during gastrulation, *Current Opinion in Cell Biology*, 17, 533–541.
- Kenis, H., van Genderen, H., Bennaghmouch, A., Rinia, H. A., Frederik, P., Narula, J., Reutelingsperger, C. P., et al. (2004). Cell surface expressed phosphatidylserine and annexin A5 open a novel portal of cell entry. *Journal of Biological Chemistry*.
- Keller, R. (2005). Cell migration during gastrulation. *Current opinion in cell biology*, *17*(5), 533-541.
- Khanna, C., & Hunter, K. (2005). Modeling metastasis in vivo. *Carcinogenesis*, 26(3), 513-523.
- Kim, B., Tang, Q., Biswas, P. S., Xu, J., Schiffelers, R. M., Xie, F. Y., Rouse, B. T., et al. (2004). Inhibition of ocular angiogenesis by siRNA targeting vascular endothelial growth factor pathway genes: therapeutic strategy for herpetic stromal keratitis. *The American journal of pathology*, *165*(6), 2177-2185.
- Kerr, J. F., Wyllie, A. H., & Currie, A. R. (1972). Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *British journal of cancer*, 26(4), 239.
- Kilinc, M. O., Aulakh, K. S., Nair, R. E., Jones, S. A., Alard, P., Kosiewicz, M. M., & Egilmez, N. K. (2006). Reversing tumor immune suppression with intratumoral IL-12: activation of tumor-associated T effector/memory cells, induction of T suppressor apoptosis, and infiltration of CD8+ T effectors. *The Journal of Immunology*, 177(10), 6962-6973.
- Kirn, D., Niculescu-Duvaz, I., Hallden, G., & Springer, C. J. (2002). The emerging fields of suicide gene therapy and virotherapy. *Trends in molecular medicine*, 8(4), 68-73.
- Kobayashi, M., Fitz, L., Ryan, M., Hewick, R. M., Clark, S. C., Chan, S., Loudon, R., Sherman, F., Perussia, B., & Trinchieri, (1989). Identification and purification of natural killer cell stimulatory factor (NKSF), a cytokine with multiple biologic effects on human lymphocytes. *Journal of Experimental Medicine*, *170*, 827-845.
- Kouwenhoven, B. (1993). Newcastle disease. *Virus infections of birds. St. Louis: Elsevier Science*, 341-61.
- Kramer, N., Walzl, A., Unger, C., Rosner, M., Krupitza, G., Hengstschläger, M., & Dolznig, H. (2013). In vitro cell migration and invasion assays. *Mutation Research/Reviews in Mutation Research*, 752(1), 10-24.
- Kubota, Y., Kleinman, H. K., Martin, G. R., & Lawley, T. J. (1988). Role of laminin and basement membrane in the morphological differentiation of human endothelial

- cells into capillary-like structures. The Journal of cell biology, 107(4), 1589-1598.
- Kumar, R., Tiwari, A. K., Chaturvedi, U., Kumar, G. R., Sahoo, A. P., Rajmani, R. S., Kumar, S., et al. (2012). Velogenic newcastle disease virus as an oncolytic virotherapeutics: in vitro characterization. *Applied biochemistry and biotechnology*, 167(7), 2005-2022.
- Lacroix, M., & Leclercq, G. (2004). Relevance of breast cancer cell lines as models for breast tumours: an update. *Breast cancer research and treatment*, 83(3), 249-289.
- Lamparska-Przybysz, M., Gajkowska, B., & Motyl, T. (2006). BID-deficient breast cancer MCF-7 cells as a model for the study of autophagy in cancer therapy. *Autophagy*, 2(1), 47-48.
- Larocca, C., & Schlom, J. (2011). Viral Vector-based Therapeutic Cancer Vaccines. *Cancer journal (Sudbury, Mass.)*, 17(5), 359.
- Lasfargues EY, Ozzello L. (1958). Cultivation of human breast carcinomas. *Journal of the National Cancer Institute*, 21, 1131–1147.
- Lasek, W., Zagożdżon, R., & Jakobisiak, M. (2014). Interleukin 12: still a promising candidate for tumor immunotherapy?. *Cancer Immunology*, *Immunotherapy*, 63(5), 419-435.
- Lech, P.J., Russell, S.J. (2010). Use of attenuated paramyxoviruses for cancer therapy. *Expert Review of Vaccines*, 9, 1275–1302.
- LeDoux, J. (Ed.). (2008). Gene Therapy Protocols: Volume 1: Production and In Vivo Applications of Gene Transfer Vectors (Vol. 433). Humana Press.
- Lee, K. M., & Kim, Y. K. (2006). The role of IL-12 and TGF-β1 in the pathophysiology of major depressive disorder. *International Immunopharmacology*, *6*(8), 1298-1304.
- Lenzi, R., Rosenblum, M., Verschraegen, C., Kudelka, A. P., Kavanagh, J. J., Hicks, M. E., Platsoucas, C. D., et al. (2002). Phase I study of intraperitoneal recombinant human interleukin 12 in patients with Müllerian carcinoma, gastrointestinal primary malignancies, and mesothelioma. *Clinical cancer research*, 8(12), 3686-3695.
- Levenson, A. S., & Jordan, V. C. (1997). MCF-7: the first hormone-responsive breast cancer cell line. *Cancer research*, *57*(15), 3071-3078. Lewis, A. M., Varghese, S., Xu, H., & Alexander, H. R. (2006). Interleukin-1 and cancer progression: the emerging role of interleukin-1 receptor antagonist as a novel therapeutic agent in cancer treatment. *Journal of translational medicine*, *4*(1), 48.

- Lim, S., & Kaldis, P. (2013). Cdks, cyclins and CKIs: roles beyond cell cycle regulation. *Development*, 140(15), 3079-3093.
- Ling, E., Shirai, K., Kanekatsu, R., & Kiguchi, K. (2003). Classification of larval circulating hemocytes of the silkworm, Bombyx mori, by acridine orange and propidium iodide staining. *Histochemistry and cell biology*, 120(6), 505-511.
- Liu, X. H., & Rose, D. P. (1996). Differential expression and regulation of cyclooxygenase-1 and-2 in two human breast cancer cell lines. *Cancer research*, 56(22), 5125-5127.
- Lou, E. (2003). Oncolytic herpes viruses as a potential mechanism for cancer therapy. *Acta Oncologica*, 42, 660–671.
- Lucas, M.L., Heller, L., Coppola, D., et al. (2002). IL-12 plasmid delivery by in vivo electroporation for the successful treatment of established subcutaneous B16.F10 melanoma. *Molecular Therapy*, 5, 668-675.
- Lv, Y. G., Yu, F., Yao, Q., Chen, J. H., & Wang, L. (2010). The role of survivin in diagnosis, prognosis and treatment of breast cancer. *Journal of thoracic disease*, 2(2), 100.
- Madigan MP, Ziegler RG, Benichou J, Byrne C, Hoover RN. (1995). Proportion of breast cancer cases in the United States explained by well-established risk factors. *Journal of National Cancer Institute*, 87,1681-5.
- Maae, E., Nielsen, M., Steffensen, K. D., Jakobsen, E. H., Jakobsen, A., & Sørensen, F. B. (2011). Estimation of immunohistochemical expression of VEGF in ductal carcinomas of the breast. *Journal of Histochemistry & Cytochemistry*, 59(8), 750-760.
- Mamon, H. J., Dahlberg, W., Azzam, E. I., Nagasawa, H., Muto, M. G., & Little, J. B. (2003). Differing effects of breast cancer 1, early onset (BRCA1) and ataxiatelangiectasia mutated (ATM) mutations on cellular responses to ionizing radiation. *International journal of radiation biology*, 79(10), 817-829.
- Manning, H. C., Buck, J. R., & Cook, R. S. (2016). Mouse models of breast cancer: platforms for discovering precision imaging diagnostics and future cancer medicine. Journal of nuclear medicine: official publication, Society of Nuclear Medicine, 57(Suppl 1), 60S.
- Mansour, M., Palese, P., & Zamarin, D. (2011). Oncolytic specificity of Newcastle disease virus is mediated by selectivity for apoptosis-resistant cells. *Journal of virology*, 85(12), 6015-6023.
- Markert, J. M., Cody, J. J., Parker, J. N., Coleman, J. M., Price, K. H., Kern, E. R., Cartner, S. C., et al. (2012). Preclinical evaluation of a genetically engineered herpes simplex virus expressing interleukin-12. *Journal of virology*, 86(9), 5304-5313.

- Marshall, J. (2011). Transwell((R)) invasion assays, *Methods in Molecular Biology*, 769, 97–110.
- Mascotti, K., McCullough, J., & Burger, S. R. (2000). HPC viability measurement: trypan blue versus acridine orange and propidium iodide. *Transfusion*, 40(6), 693-696.
- Maughan, K. L., Lutterbie, M. A., & Ham, P. S. (2010). Treatment of breast cancer. *Chemotherapy*, *51*, 53.
- McPherson, K., Steel, C. M., & Dixon, J. M. (2009). 5 Breast cancer—epidemiology, risk factors, and genetics. *ABC of Breast Diseases*, 69, 24.
- Mincey, B. A., Palmieri, F. M., & Perez, E. A. (2002). Adjuvant therapy for breast cancer: recommendations for management based on consensus review and recent clinical trials. *The Oncologist*, 7(3), 246-250.
- Mohamad, N. E., Abu, N., Rahman, H. S., Ky, H., Ho, W. Y., Lim, K. L., et al. & Yeap, S. K. (2015). Nanostructured lipid carrier improved in vivo anti-tumor and immunomodulatory effect of Zerumbone in 4T1 challenged mice. RSC Advances, 5(28), 22066-22074.
- Molouki, A., Hsu, Y. T., Jahanshiri, F., Rosli, R., & Yusoff, K. (2010). Newcastle disease virus infection promotes Bax redistribution to mitochondria and cell death in HeLa cells. *Intervirology*, 53(2), 87-94.
- Molouki, A., Hsu, Y. T., Jahanshiri, F., Abdullah, S., Rosli, R., & Yusoff, K. (2011). The matrix (M) protein of newcastle disease virus binds to human bax through its BH3 domain. *Virology journal*, 8(1), 385.
- Morimoto, L. M., White, E., Chen, Z., Chlebowski, R. T., Hays, J., Kuller, L., Stefanick, M. L., et al. (2002). Obesity, body size, and risk of postmenopausal breast cancer: the Women's Health Initiative (United States). *Cancer Causes & Control*, 13(8), 741-751.
- Mosmann, T. 1983. Rapid Colorimetric Assay for Cellular Growth and Survival: Application to Proliferation and Cytotoxicity Assays. *Journal of ImmunologicalMethods* 65: 55-63.
- Moss, B., Smith, G. L., Gerin, J. L., & Purcell, R. H. (1984). Live recombinant vaccinia virus protects chimpanzees against hepatitis B. *Nature*, *311*(5981), 67.
- Moutasim, K. A., Nystrom, M. L., & Thomas, G. J. (2011). Cell migration and invasion assays. *Cancer Cell Culture: Methods and Protocols*, 333-343.
- Muneesh, T., Vishva, M.D. (1995). Fas- and tumor necrosis factor-induced apoptosis is inhibited by the poxvirus crmA gene product. *American Society for Biochemistry and Molecular Biology*, 270, 3255–3260.

- Mur, C., Martínez- Carpio, P. A., Fernández- Montolí, M. E., Ramon, J. M., Rosel, P., & Navarro, M. A. (1998). Growth of MDA- MB- 231 cell line: different effects of TGF- β1, EGF and estradiol depending on the length of exposure. *Cell biology international*, 22(9- 10), 679-684.
- Murulitharan, K., Yusoff, K., Omar, A. R., & Molouki, A. (2013). Characterization of Malaysian velogenic NDV strain AF2240-I genomic sequence: a comparative study. *Virus genes*, 46(3), 431-440.
- Myers, R., Greiner, S., Harvey, M., et al. (2005). Oncolytic activities of approved mumps and measles vaccines for therapy of ovarian cancer. *Cancer Gene Therapy*, 12, 593–599.
- Nakaya, T., Cros, J., Park, M. S., Nakaya, Y., Zheng, H., Sagrera, A., Palese, P., et al. (2001). Recombinant Newcastle disease virus as a vaccine vector. *Journal of virology*, 75(23), 11868-11873.
- Nastala, C. L., Edington, H. D., McKinney, T. G., Tahara, H., Nalesnik, M. A., Brunda, M. J., Storkus, W. J., et al. (1994). Recombinant IL-12 administration induces tumor regression in association with IFN-gamma production. *The Journal of Immunology*, 153(4), 1697-1706.
- Nounou, M. I., ElAmrawy, F., Ahmed, N., Abdelraouf, K., Goda, S., & Syed-Sha-Qhattal, H. (2015). Breast cancer: conventional diagnosis and treatment modalities and recent patents and technologies. Breast cancer: basic and clinical research, 9, BCBCR-S29420.
- Oez, S., Platzer, E., & Welte, K. (1990). A quantitative colorimetric method to evaluate the functional state of human polymorphonuclear leukocytes. *Blut: Zeitschrift für die Gesamte Blutforschung*, 60(2), 97-102.
- Oki, T., Nishimura, K., Kitaura, J., Togami, K., Maehara, A., Izawa, K., Kiyonari, H., et al.(2014). A novel cell-cycle-indicator, mVenus-p27K-, identifies quiescent cells and visualizes G0–G1 transition. *Scientific reports*, 4, 4012.
- Omar, S., Khaled, H., Gaafar, R., Zekry, A. R., Eissa, S., & El Khatib, O. (2003). Breast cancer in Egypt: a review of disease presentation and detection strategies.
- Othman, F., Ideris, A., Motalleb, G., Eshak, Z. B., & Rahmat, A. (2010). Oncolytic effect of Newcastle disease virus AF2240 strain on the MCF-7 breast cancer cell line. *Yakhteh Medical Journal*, 12(1), 17-24.
- Page, D. L., & Dupont, W. D. (1990). Premalignant conditions and markers of elevated risk in the breast and their management. *Surgical Clinics of North America*, 70(4), 831-851.
- Paldurai, A., Kumar, S., Nayak, B., & Samal, S. K. (2010). Complete genome sequence of highly virulent neurotropic Newcastle disease virus strain Texas GB. *Virus genes*, 41(1), 67-72.

- Park, M. S., Steel, J., García-Sastre, A., Swayne, D., & Palese, P. (2006). Engineered viral vaccine constructs with dual specificity: avian influenza and Newcastle disease. *Proceedings of the National Academy of Sciences*, 103(21), 8203-8208.
- Park, S. R. S., Widness, M., Levine, A. D., & Patterson, C. E. (2011). T cell, IL-12, and IFN-γ Driven Viral Clearance in Measles Virus-Infected Brain Tissue. *Journal of virology*.
- Parker, J. N., Gillespie, G. Y., Love, C. E., Randall, S., Whitley, R. J., & Markert, J. M. (2000). Engineered herpes simplex virus expressing IL-12 in the treatment of experimental murine brain tumors. *Proceedings of the National Academy of Sciences*, 97(5), 2208-2213.
- Ponce, M. L. (2009). Tube Formation: an In Vitro Matrigel Angiogenesis Assay. *Methods in Molecular Biology*, 467, 183-188.
- Pulaski, B. A., & Ostrand-Rosenberg, S. (2001). Mouse 4T1 Breast Tumor Model. *Current Protocols in Immunology*, 1-7.
- Pulido, J., Kottke, T., Thompson, J., Galivo, F., Wongthida, P., Diaz, R. M., Harrington, K., et al. (2012). Using virally expressed melanoma cDNA libraries to identify tumor-associated antigens that cure melanoma. *Nature biotechnology*, *30*(4), 337.
- Rangaswamy, U. S., Wang, W., Cheng, X., McTamney, P., Carroll, D., & Jin, H.(2017). Newcastle disease virus establishes persistent infection in tumor cells in vitro: contribution of the cleavage site of fusion protein and second sialic acid binding site of hemagglutinin-neuraminidase. *Journal of virology*, JVI-00770.
- Rajan, J. V., Wang, M., Marquis, S. T., & Chodosh, L. A. (1996). Brca2 is coordinately regulated with Brca1 during proliferation and differentiation in mammary epithelial cells. *Proceedings of the National Academy of Sciences*, 93(23), 13078-13083.
- Rasoli, M., Yeap, S. K., Tan, S. W., Moeini, H., Ideris, A., Bejo, M. H., Omar, A. R., et al. (2014). Alteration in lymphocyte responses, cytokine and chemokine profiles in chickens infected with genotype VII and VIII velogenic Newcastle disease virus. *Comparative immunology, microbiology and infectious diseases*, *37*(1), 11-21.
- Ratei, R., Karawajew, L., Lacombe, F., Jagoda, K., Poeta, G. D., Kraan, J., Santiago, M. D., Kappelmayer, J., Björklund, E., Ludwig, W., Gratama, J., & Orfao, A. (2006). Normal Lymphocytes from Leukemic Samples as an Internal Quality Control for Fluorescence Intensity in Immunophenotyping of Acute Leukemias. Cytometry 70, 1-9.
- Ravindra, P. V., Tiwari, A. K., Ratta, B., Bais, M. V., Chaturvedi, U., Palia, S. K., Chauhan, R. S., et al. (2009). Time course of Newcastle disease virus-induced apoptotic pathways. *Virus research*, *144*(1), 350-354.

- Reed, A. E. M., Kutasovic, J. R., Lakhani, S. R., & Simpson, P. T. (2015). Invasive lobular carcinoma of the breast: morphology, biomarkers and omics. Breast cancer research, 17(1), 12.
- Ren, G., Tian, G., Liu, Y., He, J., Gao, X., Yu, Y., Yin, J., et al. (2016). Recombinant Newcastle disease virus encoding IL-12 and/or IL-2 as potential candidate for hepatoma carcinoma therapy. *Technology in cancer research & treatment*, 15(5), NP83-NP94.
- Ring, C. J. A., & Blair, E. D. (2000). Viruses as vehicles and expressors of genetic material. *Genetically Engineered Viruses: Development and Applications*, 1-4.
- Ring, C. J. (2002). Cytolytic viruses as potential anti-cancer agents. *Journal of general virology*, 83(3), 491-502.
- Robertson, M. J., & Ritz, J. (1996). Interleukin 12: basic biology and potential applications in cancer treatment. *The Oncologist* (1 & 2), 88-97.
- Rodriguez-Madoz, J. R., Liu, K. H., Quetglas, J. I., Ruiz-Guillen, M., Otano, I., Crettaz, J., et al. Prieto, J. (2009). Semliki forest virus expressing interleukin-12 induces antiviral and antitumoral responses in woodchucks with chronic viral hepatitis and hepatocellular carcinoma. *Journal of virology*, 83(23), 12266-12278.
- Rossant, J., & Howard, L. (2002). Signaling pathways in vascular development. *Annual Review of Cell and Developmental Biology*, 18, 541–573.
- Roy, P. (2012). Diagnosis and control of Newcastle disease in developing countries. *World's Poultry Science Journal*, 68(04), 693-706.
- Ruben, R.L., Neubuer, R.H. (1987). Semiautomated colorimeteric assay for *in vitro* screening of anticancer compounds. *Cancer Treatment Reports*, 71, 1141–1149.
- Russell, S.J., & Peng, K.W. (2009). Measles virus for cancer therapy. *Current Topics in Microbiology and Immunology*, 330, 213–241.
- Samal, S. K. (2011). Newcastle disease and related avian paramyxoviruses. *The biology of paramyxoviruses*, 69-114.
- Sánchez-Felipe, L., Villar, E., & Muñoz-Barroso, I. (2014). Entry of Newcastle Disease Virus into the host cell: role of acidic pH and endocytosis. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1838(1), 300-309.
- Sangro, B., Mazzolini, G., Ruiz, J., Herraiz, M., Quiroga, J., Herrero, I., Olagüe, C., et al. (2004). Phase I trial of intratumoral injection of an adenovirus encoding interleukin-12 for advanced digestive tumors. *Journal of clinical oncology*, 22(8), 1389-1397.
- Sankaranarayanan, R., & Ferlay, J. (2013). Burden of breast and gynecological cancers in low-resource countries. *In Breast and Gynecological Cancers* (1-17).

- Schirrmacher, V., Bihari, A. S., Stücker, W., & Sprenger, T. (2014). Long-term remission of prostate cancer with extensive bone metastases upon immuno-and virotherapy: A case report. *Oncology letters*, 8(6), 2403-2406.
- Shafi, G., Hasan, T. N., Syed, N. A., Al-Hazzani, A. A., Alshatwi, A. A., Jyothi, A., & Munshi, A. (2012). Artemisia absinthium (AA): a novel potential complementary and alternative medicine for breast cancer. *Molecular biology reports*, 39(7), 7373-7379.
- Sinkovics, J. G., & Horvath, J. C. (2000). Newcastle disease virus (NDV): brief history of its oncolytic strains. *Journal of clinical virology*, *16*(1), 1-15.
- Smyth, M. J., Cretney, E., Kershaw, M. H., & Hayakawa, Y. (2004). Cytokines in cancer immunity and immunotherapy. *Immunological reviews*, 202(1), 275-293.
- Solomon, M., Wofford, J., Johnson, C., Regan, D., & Creer, M. H. (2010). Factors influencing cord blood viability assessment before cryopreservation. *Transfusion*, *50*(4), 820-830.
- Song, K. Y., Wong, J., Gonzalez, L., Sheng, G., Zamarin, D., & Fong, Y. (2010). Antitumor efficacy of viral therapy using genetically engineered Newcastle disease virus [NDV (F3aa)-GFP] for peritoneally disseminated gastric cancer. *Journal of Molecular Medicine*, 88(6), 589-596.
- Soliman, N. A., & Yussif, S. M. (2016). Ki-67 as a prognostic marker according to breast cancer molecular subtype. *Cancer biology & medicine*, *13*(4), 496.
- Sonnenblick, A., Fumagalli, D., Sotiriou, C., & Piccart, M. (2014). Is the differentiation into molecular subtypes of breast cancer important for staging, local and systemic therapy, and follow up?. Cancer treatment reviews, 40(9), 1089-1095.
- Sorensen, E. W., Gerber, S. A., Frelinger, J. G., & Lord, E. M. (2010). IL-12 suppresses vascular endothelial growth factor receptor 3 expression on tumor vessels by two distinct IFN-γ-dependent mechanisms. *The journal of immunology*, ji_0903210.
- Soule, H. D., Vazquez, J., Long, A., Albert, S., & Brennan, M. (1973). A human cell line from a pleural effusion derived from a breast carcinoma 2. *Journal of the National Cancer Institute*, *51*(5), 1409-1416.
- Stem, A. S., Podlaski, F. J., Hulmes, J. D., Pan, Y.C. E., Quinn, P. M., Wolitzky, A. G., Familletti, P. C., Stremlo, D. L., Truitt, I., Chizzonite, R., & Gately, M. K. (1990). Purification to homogeneity and partial characterization of cytotoxic lymphocyte maturation factor from human B-lymphoblastoid cells. *Proceedings of the National Academy of Sciences of the United States of America*, 87, 6808-6812.
- Stern, A. S., Podlaski, F. J., Hulmes, J. D., Pan, Y. C., Quinn, P. M., Wolitzky, A. G., Chizzonite, R., et al. (1990). Purification to homogeneity and partial characterization of cytotoxic lymphocyte maturation factor from human B-

- lymphoblastoid cells. *Proceedings of the National Academy of Sciences*, 87(17), 6808-6812.
- Sugiyama, M., Morita, M., Yoshida, R., Ando, K., Egashira, A., Takefumi, O., ... & Maehara, Y. (2012). Patterns and time of recurrence after complete resection of esophageal cancer. *Surgery today*, 42(8), 752-758.
- Tao, K., Fang, M., Alroy, J. & Sahagian, G. G. (2008). Imagable 4T1 Model for the Study of Late Stage Breast Cancer. *BMC Cancer*, 8, 1-19.
- Tayeb, S., Zakay-Rones, Z., & Panet, A. (2015). Therapeutic potential of oncolytic Newcastle disease virus: a critical review. *Oncolytic virotherapy*, *4*, 49.
- Telford, W.G., King, L.E., Fraker, P.J. (1994). Rapid quantitation of apoptosis in pure and heterogeneous cell populations using flow cytometry. *Journal of Immunological Methods*, 172, 1–16.
- Thompson, D. W. (1994). Genetic epidemiology of breast cancer. *Cancer*, 74(S1), 279-287.
- Trinchieri G. (1994). Interleukin-12: a cytokine produced by antigenpresenting cells with immunoregulatory functions in the generation of T-helper cells type 1 and cytotoxic lymphocytes. *Blood*, 84(12), 4008-4027.
- Tugues, S., Burkhard, S. H., Ohs, I., Vrohlings, M., Nussbaum, K., Vom Berg, J., Becher, B., et al. (2015). New insights into IL-12-mediated tumor suppression. *Cell Death & Differentiation*, 22(2), 237-246.
- Twentyman, P. R., & Luscombe, M. (1987). A study of some variables in a tetrazolium dye (MTT) based assay for cell growth and chemosensitivity. *British journal of cancer*, 56(3), 279.
- Tyrer, J., Duffy, S. W., & Cuzick, J. (2004). A breast cancer prediction model incorporating familial and personal risk factors. *Statistics in medicine*, 23(7), 1111-1130.
- Uemura, A., Takehara, T., Miyagi, T., Suzuki, T., Tatsumi, T., Ohkawa, K., Hayashi, N., et al. (2010). Natural killer cell is a major producer of interferon γ that is critical for the IL-12-induced anti-tumor effect in mice. *Cancer immunology, immunotherapy*, 59(3), 453.
- Van der Burg, S. H., Arens, R., Ossendorp, F., van Hall, T., & Melief, C. J. (2016). Vaccines for established cancer: overcoming the challenges posed by immune evasion. *Nature reviews cancer*, 16(4), 219.
- Van Engeland, M., Nieland, L. J., Ramaekers, F. C., Schutte, B., & Reutelingsperger, C. P. (1998). Annexin V- affinity assay: a review on an apoptosis detection system based on phosphatidylserine exposure. *Cytometry: The Journal of the International Society for Analytical Cytology*, 31(1), 1-9.

- Van Rikxoort, M., Michaelis, M., Wolschek, M., Muster, T., Egorov, A., Seipelt, J., Cinatl Jr, J., et al. (2012). Oncolytic effects of a novel influenza A virus expressing interleukin-15 from the NS reading frame. *PLoS One*, 7(5), 36506.
- Varghese, S., Rabkin, S. D., Liu, R., Nielsen, P. G., Ipe, T., & Martuza, R. L. (2006). Enhanced therapeutic efficacy of IL-12, but not GM-CSF, expressing oncolytic herpes simplex virus for transgenic mouse derived prostate cancers. *Cancer gene therapy*, 13(3), 253.
- Vermes, I., Haanen, C., Steffens-Nakken, H., & Reutelingsperger, C. (1995) A novel assay for apoptosis flow cytometric detection of phosphatidylserine early apoptotic cells using fluorescein labeled expression on Annexin V. *Journal of Immunological Methods*, 184, 39–51.
- Vogelstein, B., & Kinzler, K. W. (2004). Cancer genes and the pathways they control. *Nature medicine*, 10(8), 789.
- Wallen, C. A., Higashikubo, R., & Dethlefsen, L. A. (1982). Comparison of two flow cytometric assays for cellular RNA—acridine orange and propidium iodide. *Cytometry: The Journal of the International Society for Analytical Cytology*, 3(3), 155-160.
- Walrath, J. C., Hawes, J. J., Van Dyke, T., & Reilly, K. M. (2010). Genetically engineered mouse models in cancer research. In Advances in cancer research (Vol. 106, pp. 113-164). Academic Press.
- Washburn, B., & Schirrmacher, V. (2002). Human tumor cell infection by Newcastle Disease Virus leads to upregulation of HLA and cell adhesion molecules and to induction of interferons, chemokines and finally apoptosis. *International journal of oncology*, 21(1), 85-93.
- Weiss, J. M., Subleski, J. J., Wigginton, J. M., & Wiltrout, R. H. (2007). Immunotherapy of cancer by IL-12-based cytokine combinations. *Expert opinion on biological therapy*, 7(11), 1705-1721.
- Wolf, S.F., Temple, P.A., & Kobayashi, M. et al. (1991). Cloning of cDNA for natural killer cell stimulatory factor, a heterodimeric cytokine with multiple effects on T and natural killer cells. *Journal of Immunology*, *146*, 3074-3081.
- Wong HH, Lemoine NR, & Wang Y. (2010). Oncolytic viruses for cancer therapy: overcoming the obstacles. *Viruses*, 2(1), 78-106.
- "World Population Prospects Population Division United Nations". esa.un.org.

 December 12, 2018 from https://www.un.org/development/desa/publications/world-population-prospects-the-2017-revision.html

- Worldwide cancer statistics retrieved December 12, 2018 from https://www.cancerresearchuk.org/health-professional/cancer-statistics/worldwide-cancer
- Wu, Y., He, J., An, Y., Wang, X., Liu, Y., Yan, S., Yin, J., et al. (2016). Recombinant Newcastle disease virus (NDV/Anh-IL-2) expressing human IL-2 as a potential candidate for suppresses growth of hepatoma therapy. *Journal of pharmacological sciences*, 132(1), 24-30.
- Yaacov, B., Elihaoo, E., Ben-Shlomo, M., Greenbaum, I., Panet, A., & Zakay-Rones, Z. (2008). Selective oncolytic effect of an attenuated Newcastle disease virus (NDV-HUJ) in lung tumors. *Cancer gene therapy*, 15(12), 795.
- Zaharoff, D. A., Hoffman, B. S., Hooper, H. B., Benjamin, C. J., Khurana, K. K., Hance, K. W., Greiner, J. W., et al. (2009). Intravesical immunotherapy of superficial bladder cancer with chitosan/interleukin-12. *Cancer research*, 69(15), 6192-6199.
- Zamarin, D., Vigil, A., Kelly, K., García-Sastre, A., & Fong, Y. (2009). Genetically engineered Newcastle disease virus for malignant melanoma therapy. *Gene therapy*, 16(6), 796.
- Zeng, J., Fournier, P., & Schirrmacher, V. (2002). Induction of interferon-α and tumor necrosis factor-related apoptosis-inducing ligand in human blood mononuclear cells by hemagglutinin-neuraminidase but not F protein of Newcastle disease virus. *Virology*, 297(1), 19-30.
- Zhao, H., & Peeters, B. P. (2003). Recombinant Newcastle disease virus as a viral vector: effect of genomic location of foreign gene on gene expression and virus replication. *Journal of General Virology*, 84(4), 781-788.
- Zitvogel, L., Tahara, H., Robbins, P. D., Storkus, W. J., Clarke, M. R., Nalesnik, M. A.,
 & Lotze, M. T. (1995). Cancer immunotherapy of established tumors with IL12. Effective delivery by genetically engineered fibroblasts. *The Journal of Immunology*, 155(3), 1393-1403.

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LIST OF PUBLICATIONS / AWARDS

- Zahiah, M. A., Tan, S. W., Noorjahan, B. M. A. & Yeap, S. K. Safety, stability and efficacy of recombinant Newcastle disease virus expressing human interleukin 12 (rAF-il12) as an anti-breast cancer vaccine. The 3rd AMDI IBSC 2018 on Emerging Infectious Diseases, January 19-20, 2018.
- Awarded "Best Oral Presenter" at The 3rd AMDI IBSC 2018 on Emerging Infectious Diseases, January 19-20, 2018.
- Zahiah, M.A, Muhammad, A.C.A, Sheau, W.T, Swee, K.Y, Noorjahan, B.A. et al. & Khatijah, Y. (2018). Safety and cytotoxicity effects of a recombinant NDV strain AF2240 expressing human interleukin 12 (Submitted to Scientific Reports, Nature). Submission Date: 4th July 2018; Submitted with Revision: 1st October 2018.
- Zahiah, M.A, Muhammad, A.C.A, Sheau, W.T, Swee, K.Y, Noorjahan, B.A. et al. & Khatijah, Y. Cytotoxic and anti-metastatic effects of a recombinant NDV strain AF2240 expressing human interleukin 12 in two breast cancer cell lines, MCF-7 and MDA-MB231 in vitro. (Manuscript in Preparation).
- Zahiah, M.A, Muhammad, A.C.A, Sheau, W.T, Swee, K.Y, Noorjahan, B.A. et al. & Khatijah, Y. In vivo anti-tumour effects of a recombinant NDV strain AF2240 expressing human interleukin 12 in 4T1 breast cancer cell-challenged mice. (Manuscript in Preparation).



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