

UNIVERSITI PUTRA MALAYSIA

TOXICITY OF SYNTHETIC INSECTICIDES ON THIRD INSTAR LARVAE OF Chrysoperla nipponensis (NEUROPTERA : CHRYSOPIDAE)

NADIAH BINTI JALILUDIN

FP 2017 43

TOXICITY OF SYNTHETIC INSECTICIDES ON THIRD INSTAR LARVAE OF

Chrysoperla nipponensis

(NEUROPTERA: CHRYSOPIDAE)



NADIAH BINTI JALILUDIN

FACULTY OF AGRICULTURE UNIVERSITI PUTRA MALAYSIA SERDANG, SELANGOR DARUL EHSAN 2016/2017

TOXICITY OF SYNTHETIC INSECTICIDES ON THIRD INSTAR LARVAE OF

Chrysoperla nipponensis

(NEUROPTERA: CHRYSOPIDAE)



A project report submitted to Faculty of Agriculture, Universiti Putra Malaysia, in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Agricultural Science.

Faculty of Agriculture Universiti Putra Malaysia 2016/2017

ENDORSEMENT

This project report entitled "Toxicity of synthetic insecticides on third instar larvae of *Chrysoperla nipponensis*" is prepared by Nadiah binti Jaliludin and submitted to Faculty of Agriculture in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Agricultural Science.

| Student's name | Student's signature |
|-----------------------------------|---------------------|
| | |
| Certified by: | |
| | |
| | |
| (Supervisor's signature) | |
| (Prof. Dr. Dzolkhifli bin Omar) | |
| (Department of Plant Protection) | |
| Date: | |
| | |

ACKNOWLEDGEMENT

Alhamdulillah, the most precious word to Allah for His blessing upon me which allowed me accomplished this final year project and thesis writing completely within 2 semesters. On this opportunity, I would like to express my profound gratitude to my project supervisor, Prof. Dr. Dzolkhifli bin Omar for his fully commitment, guidance and encouragement in conducting this project throughout this year. He is qualified to be a worth lecturer and guider in my study. A special thanks to laboratory assistance of Toxicology Laboratory, Mr Jakasi Sarbini and Mr Zaki Yusof for their kindness in guiding me about the usage of tools and chemical materials within the laboratory clearly and safely. Besides, special gratitude are dedicated to Mr Syafique Ahmad Memon for their assistant and guidance during the period of culturing the green lacewing. Not forgotten to Dr Norhayu binti Asib in which assist me to complete this final year project. Moreover, thanks to the post graduates students of crop protection department, who have directly and indirectly helped me during conducting the laboratory works. Last but not least, deeply appreciated is dedicated to my beloved family members, Faridah binti Baba, Nasrah binti Jaliludin, Syufri bin Selamat and dear friends who had given me a full support moral in completing this project. May the almighty cherish all of you with His divine blessings.

TABLE OF CONTENTS

| CONTENT | PAGE |
|--|-----------------------|
| ENDORSEMENT | i |
| ACKNOWLEDGEMENT | ii |
| TABLE OF CONTENT | iii |
| LIST OF TABLES | v |
| LIST OF FIGURES | vi |
| ABSTRACT ABSTRAK | vii viii |
| 1.0 INTRODUCTION | 1 |
| 1.1 Background of study | 1 |
| 1.2 Problem statement | 1 |
| 1.3 Objective | 2 |
| 1.4 Significant of study | 2 |
| 2.0 LITERATURE REVIEW | 3 |
| 2.1 Morphology of Chrysoperla | nipponensis 3 |
| 2.2 Food sources | 3 |
| 2.3 The important of <i>Chrysoperi</i> | la nipponensis 4 |
| 2.4 Habitat of green lacewing | 4 |
| 2.5 Biological control in Integra | ted Pest Management 5 |
| 2.6 Serious pest problem in agric | culture activity 6 |

| 2.7 Insecticides | 8 |
|---|----|
| 2.7.1 Cypermethrin | 8 |
| 2.7.2 Spinosad | 9 |
| 3.0 MATERIALS AND METHODS | 10 |
| 3.1 Plant and planting material | 10 |
| 3.2 Culture of insect pests | 10 |
| 3.3 Rearing of Chrysoperla nipponensis | 10 |
| 3.4 Preparation of insecticides | 11 |
| 3.5 Bioassay | 11 |
| 3.6 Statistical analysis | 12 |
| 4.0 RESULTS AND DISCUSSION | 13 |
| 4.1 Mortality of larvae Chrysoperla nipponensis after exposed to | |
| cypermethrin and spinosad | 13 |
| 4.2 Toxicity of synthetic insecticides against third instar larvae of | |
| Chrysoperla nipponensis | 15 |
| 4.3 Duration of pupation for Chrysoperla nipponensis | 20 |
| 4.4 Adult emergence of Chrysoperla nipponensis | 22 |
| 5.0 CONCLUSION | 24 |
| REFERENCES | 25 |
| APPENDICES | 29 |

LIST OF TABLES

| Table | | Page |
|-------|--|------|
| 1 | Table4.1.1LC50ofcypermethrinagainstChrysoperlanipponensis. | 14 |
| 2 | Table 4.1.2 LC ₅₀ of spinosad against <i>Chrysoperla nipponensis</i> . | 14 |
| 3 | Table 4.2.1 Effect of cypermethrin on the third instar larvae mortality of <i>Chrysoperla nipponensis</i> at 24 hours. | 15 |
| 4 | Table 4.2.2 Effect of cypermethrin and spinosad on the third instar larvae mortality of <i>Chrysoperla nipponensis</i> at 48 hours. | 16 |
| 5 | Table 4.2.3 Effect of cypermethrin and spinosad on the third instar larvae mortality of <i>Chrysoperla nipponensis</i> at 72 hours. | 17 |
| 6 | Table 4.2.4 Effect of cypermethrin and spinosad on the third instar larvae mortality of <i>Chrysoperla nipponensis</i> at 96 hours. | 18 |
| 7 | Table 4.2.5 Effect of cypermethrin and spinosad on the third instar larvae mortality of <i>Chrysoperla nipponensis</i> at 120 hours. | 19 |
| 8 | Table 4.3.1 Effect of cypermethrin and spinosad on duration of pupation for <i>Chrysoperla nipponensis</i> | 20 |
| 9 | Table 4.4.1 Effect of cypermethrin and spinosad on adult emergence of Chrysoperla nipponensis | 22 |
| | | |

LIST OF FIGURES

| Figure | | Page |
|--------|--|------|
| 1 | Figure2.1 Structural compound of cypermethrin | 8 |
| 2 | Figure 2.2 Structural compound of spinosad | 9 |
| 3 | Figure 3.1 The leave of Capsicum annum was cut into | 11 |
| | small size based on nucleon delta surface. | |
| 4 | Figure 3.2 The treated leaves were placed inside the holes | 12 |
| | and one of third instar was put in each hole. | 12 |

C

ABSTRACT

Green lacewing *Chrysoperla niponensis* is well known as insect predator of pests in the field crop and widely used as biological control agent. Usually an adult feeds on nectar, pollen and sugar excretions. The larvae are considered as active worker and having huge appetite to feed on aphids, red spider mites, thrips, mealybugs and whiteflies. The intensive use of insecticides to control the pest could reduce the number of biological control agents through residual contact with insecticides. Therefore, the purpose of this study was to evaluate toxicity of cypermethrin and spinosad and to identify the most toxic insecticide against the larvae of *C. nipponensis*. The insecticides that had been used are cypermethrin and spinosad. The experiment was conducted by using leaf-dip bioassay method. The mortality was recorded after 24, 48, 72, 96 and 120 hours and the results were subjected to probit analysis and ANOVA with means separation by Tukey. The most toxic insecticide against third instar larvae was obtained from cypermethrin with LC_{50} 30.28 ppm while the less toxic was examined from spinosad with LC_{50} 182.23 ppm.

ABSTRAK

'Green lacewing' atau nama sainstifik ialah Chrysoperla niponensis terkenal sebagai serangga pemangsa bagi perosak tanaman makanan dan digunakan secara meluas sebagai agen kawalan biologi. Biasanya serangga dewasa memakan madu, debunga dan kumuhan gula dari bunga tumbuhan. Manakala larva tersebut dikenali sebagai serangga yang aktif dan mempunyai selera makan yang banyak untuk memakan aphids, hama labah-labah merah, thrips, koya dan lalat putih. Serangga ini boleh didapati di kawasan pertanian seperti di ladang sayur-sayuran. Walau bagaimanapun, penggunaan racun serangga secara intensif boleh mengurangkan bilangan agen kawalan biologi melalui sisa racun serangga yang terdapat di tanaman. Oleh itu, tujuan eksperimen ini adalah untuk menilai ketoksikan beberapa racun serangga sintetik serta mengenal pasti racun serangga yang paling toksik terhadap C. nipponensis. Racun serangga yang telah digunakan adalah cypermethrin dan spinosad. Racun ini sering digunakan dalam penanaman sayur-sayuran. Eksperimen ini dijalankan dengan menggunakan kaedah bioassay. Kematian telah direkodkan untuk setiap 24, 48, 72, 96 dan 120 jam dan keputusan yang di perolehi di analisis dengan menggunakan Probit dan ANOVA serta cara perbandingan dengan Tukey. Racun serangga yang paling toksik terhadap larva instar ketiga adalah daripada cypermethrin dengan LC₅₀ 30.280 ppm manakala racun yang kurang toksik adalah daripada spinosad dengan LC₅₀ 182.23 ppm.

1.0 INTRODUCTION

1.1 Background of study

Green lacewing *Chrysoperla nipponensis* is well known as predatory of pest in the field crop and widely used as biological control of pests. This predator has complete metamorphosis from the formation of eggs, larvae, pupae and adults. Usually adults feed on nectar, pollen and sugar excretion of insects. The larvae are considered as active worker and have huge appetite to feed on aphids, red spider mites, thrips, mealybug and whitefly (Garrett, 2005).

According to Salim (2016), biological control is related to the use of natural enemies in the field to reduce the number of insect pest and at the same time, it helps to increase crop yield through lowering the damaged crop. There are several examples of biological control agents such as parasitic wasps, lady beetle and green lacewing. They are useful in killing and consuming insect pests during their development.

1.2 Problem statement

Instead of biological control method, most farmers use chemical control because of its effectiveness to kill the insect pest in the field. The overuse of insecticides can cause elimination of beneficial insects in the natural environment. After that, biological control agents are difficult to survive when conventional insecticides are applied to control the pest. Thus, the toxicity of conventional insecticides should be evaluated.

1.3 Objective

The objectives of this study were consequently as below :

- 1. To evaluate toxicity effects of several synthetics insecticides on third instar larvae *Chrysoperla nipponensis*.
- 2. To identify the most toxic insecticides on C. nipponensis

1.4 Significant of study

This study was to focus on the use on synthetic insecticides such as cypermethrin and spinosad that currently being commonly used to control insect pest population in vegetable production in Malaysia. Apart from that, the experiment would will assist the farmer to choose the insecticides with no detrimental effect toward the green lacewing.



REFERENCES

Alford, D.V. (2011) Plant pests. HarperCollins, UK.

- Alghamdi, A. (2013) Genetic Diversity of Chrysoperla sp. at East of Red Sea Using Cytochrome Oxidase Subunit I (COI) Gene. International Journal of Science and Research (IJSR). 4(3).1639-1642.
- Boucher, T.J. (2012) University Connecticut. http://ipm.uconn.edu/documents/raw2/Spinos ad/Spinosad.php?aid=9. Retrieved 18 september 2016.
- Brooks S. J. and Barnard P.C. (1990). The green lacewings of the world: a generic review (Neuroptera: Chrysopidae). Bulletin of the British Museum Natural History (Entomology). 59(2): 117-286
- Brooks S. J. (1997). An Overview of the Current Status of Chrysopidae (Neuroptera) systematics. Journal of Applied Entomology. 44 (2): 267-275.
- Bunch, T. R.; Bond, C.; Buhl, K.; Stone, D. 2014. Spinosad General Fact Sheet; National
Pesticide Information Center, Oregon State University Extension Services.
Retrieved 18 September 2016 from
http://npic.orst.edu/factsheets/spinosadgen.html.
- Cypermethrin. 1998. National Pesticides Information Center. Oregon State University Extension Services. Retrieved 18 September 2016 from http://npic.orst.edu/factsheets/cypermethrin.pdf.

Caroline, C. 2005. Insecticides Factsheet. Journal of Pesticides Reform . 2. : 10-15.

- Choi, Y. C., Ham, E.H., Lee, J. S., Lee, B.W., Ahn, T.H., Jin, H.Y. and Song, J. H. 2013.
 A study on application of Pseudococcus comstocki (Kuwana) (Hemiptera: Pseudococcidae) by new native natural enemy (Chrysoperla nipponensis (Okamoto)) (Neuroptera: Chrysopidae) on tropical plant (Trachycarpus fortunei (Hook)). Korean Journal of Nature Conservation Vol.7 No.2 pp.147-150
- Choi, M. Y., Mochizuki, A., & Henry, C. S. (2015). The green lacewing, Chrysoperla nipponensis in nature and in an insectary population in Korea: Song types and mitochondrial COI haplotypes. Journal of Asia-Pacific Entomology, 18(2), 151-155.

- Diehl, E., Sereda, E., Wolters, V., & Birkhofer, K. (2013). Effects of predator specialization, host plant and climate on biological control of aphids by natural enemies: a meta-analysis. Journal of Applied Ecology, 50(1), 262-270.
- DiTomaso, J. M., Van Steenwyk, R. A., Nowierski, R. M., Vollmer, J. L., Lane, E., Chilton, E., ... & Dionigi, C. P. (2016). Enhancing the effectiveness of biological control programs of invasive species through a more comprehensive pest management approach. Pest Management Science.
- Drees, B.M and Jackman, J. 1998. Field Guide to Texas Insects Houstan, Texas :Gulf Publishing Company. Retrieved 18 September 2016 from http://texasinsects.tamu.edu/bimg125.html.
- Ferron, P., & Deguine, J. P. (2005). Crop protection, biological control, habitat management and integrated farming. A review. Agronomy for Sustainable 25(1), 17-24.
- Fossen, M. 2006. Environmental Fate of Imidacloprid. Retrieved 18 September 2016 from http://www.cdpr.ca.gov/docs/emon/pubs/fatememo/Imidclprdfate2.pdf.
- Garrett, H. and Beck, C.M. 2005 .Texas Bug Book : The Good, The Bad and Ugly.Austin :Texas University Press.
- Garzón, A., Medina, P., Amor, F., Viñuela, E., & Budia, F. (2015). Toxicity and sublethal effects of six insecticides to last instar larvae and adults of the biocontrol agents Chrysoperla carnea (Stephens)(Neuroptera: Chrysopidae) and Adalia bipunctata (L.)(Coleoptera: Coccinellidae). Chemosphere, 132, 87-93.
- Gervais, J. A.; Luukinen, B.; Buhl, K.; Stone, D. 2010. Imidacloprid General Fact Sheet; National Pesticide Information Center, Oregon State University Extension Services. http://npic.orst.edu/factsheets/imidagen.html. Retrieved 18 September 2016.

Gilbert, S. 2014. Carbaryl . Toxipidea Connecting Science and People . Retrieved 18 September 2016 from http://www.toxipedia.org/display/toxipedia/Pesticides.

Henry, C. S., Brooks, S.J., Duelli, P., Johnson, J. B., Wells, M. M., & Mochizuki, A.(2013).
Obligatory duetting behaviour in the Chrysoperla carnea-group of cryptic species (Neuroptera: Chrysopidae): its role in shaping evolutionary history. Biological Reviews, 88(4), 787-808.

- Heu, R.A., M.T. Fukada and P. Conant, 2007. Papaya mealybug, *Paracoccus marginatus* Williams and Granara de Willink 1992 (Hemiptera: Pseudococcidae). New Pest Advisory. 4(3). State of Hawaii Department of Agriculture, Honolulu, HI. Available from http://hawaii.gov/hdoa/pi/ppc/npa-1/npa04-03-PMB.pdf[Accessed 14 August 2009].
- Holman, J. (2009) Host plant catalog of aphids. Palaearctic Region. Springer Science.
- Ma, W., & Abdulai, A. (2016). Adoption of Integrated Pest Management Technology and Farm Economic Performance. The Impact of Agricultural Cooperatives on the Adoption of Technologies and Farm Performance of Apple Farmers in China, 65.
- Mastoi, M. I., Azura, A. N., Muhammad, R., Idris, A. B., & Ibrahim, Y. (2011). First report of papaya mealybug Paracoccus marginatus (Hemiptera: Pseudococcidae) from Malaysia. Aust J Basic Appl Sci, 5(7), 1247-1250.
- Menzler-Hokkanen, I. (2006). Socioeconomic significance of biological control. In An ecological and societal approach to biological control (pp. 13-25). Springer Netherlands.
- Miller, D.R. and G.L. Miller, 2002. Redescription of Paracoccus marginatus Williams and Granara de Willink (Hemiptera: Coccoidea: Pseudococcidae) including descriptions of the immature stages and adult male. Proceedings of Entomological Society of Washington, 104(1): 1-23.
- Oswald J.D. (2013). LDL Neuropterida Species of the World (version Sep 2007). In: Species 2000 & IT IS Catalogue of Life, 11th March 2013 (Roskov Y., Kunze T., Paglinawan L., Orrell T., Nicolson D., Culham A., Bailly N., Kirk P., Bourgoin T., Baillargeon G., Hernandez F., De Wever A., eds). Digital resource at www.catalogueoflife.org/col/. Species 2000: Reading, UK.
- Principi, M.M. and M. Canard, 1984. Feeding Habits. In: Biology of Chrysopidae, Canard, M., Y. Semeria and T.R. New (Ed.). Dr. W. Junk Publishers, The Hague, The Netherlands, pp: 76-92.
- Rabasse, J. M., & van Steenis, M. J. (1999). Biological control of aphids. In Integrated Pest and Disease Management in Greenhouse Crops (pp. 235-243). Springer Netherlands.

- Rimoldi, F., Schneider, M. I., & Ronco, A. E. (2008). Susceptibility of *Chrysoperla externa* eggs (Neuroptera: Chrisopidae) to conventional and biorational insecticides. Environmental entomology, 37(5), 1252-1257.
- Salim, M., Gökçe, A., Naqqash, M. N., & Bakhsh, A. (2016). An overview of biological control of economically important lepidopteron pests with parasitoids.
- Wells, M. M., & Henry, C. S. (1998). Songs, reproductive isolation, and speciation in cryptic species of insects. Endless forms: species and speciation, 217-233.
- Zehnder, G., Gurr, G. M., Kühne, S., Wade, M. R., Wratten, S. D., & Wyss, E. (2007). Arthropod pest management in organic crops. Annu. Rev. Entomol., 52, 57-80.

