



UNIVERSITI PUTRA MALAYSIA

**HYDROPHOBIC MODIFICATION OF CHITOSAN NANOPARTICLES FOR
INCREASED THERAPEUTIC DELIVERY AND ANTICANCER
PROPERTIES TOWARDS HUMAN CANCER CELL LINES**

CHA YEE KUEN

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By

CHA YEE KUEN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Philosophy**

July 2020

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Abstract of thesis presented to Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

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Lung cancer is the leading cause of cancer-related deaths for men and second for women worldwide, with approximately 1.6 million of deaths reported annually. Conventional delivery of anticancer drugs for treatment is less effective due to intrinsic obstacles such as poor aqueous solubility and poor cellular accumulation of drugs. Additionally, chemotherapy incurs various concomitant undesirable side effects including cytotoxicity, cardiotoxicity and neurotoxicity. Thus, the utilization of natural anticancer therapeutics as an alternative is required to prevent these undesirable consequences. Silibinin (SLB) is a poorly-soluble plant extract derived from *Silybum marianum* which is commonly used for treatment of liver diseases. SLB exhibits anti-cancer properties by inhibiting cell growth, angiogenesis, proliferation, cell migration and inducing apoptotic death. However, poor solubility and cell accumulation of SLB limits the use of this compound as an alternative anticancer therapeutic. Therefore, the employment of nanotechnological-based applications have aided in the advancement of better diagnosis and more efficient treatment of cancer. Additionally, the utilization of nanoparticulate delivery can reduce the dosage of the natural therapeutic compound, reducing potential side effects of therapeutics. Based on the results, a low encapsulation efficiency was achieved by conventional chitosan nanoparticle (CNP) system. This necessitates modification of the CNP through hydrophobic modification to develop a palmitoyl-chitosan nanoparticle (pCNP) system for enhanced encapsulation efficiency and therapeutic efficacy. Various physico-chemical characterization analyses was conducted to evaluate the pCNP system. Utilization of free amine percentage of pCNP increased by 15% due to the conjugation of –NHS palmitic acid to the polymer, in addition to TPP. The pCNP synthesized revealed an expansion in size

and decrease in polydispersity index (PDI) compared to CNP, indicating greater monodispersity in pCNP. The encapsulation of SLB into pCNP showed an expansion in size and PDI, signifying the heterogeneity of nanoparticles after encapsulation and correlating to morphological analyses. Moreover, the encapsulation efficiency improved 1-fold from 25% of CNP-SLB to 50% of pCNP-SLB. Subsequently, MTT cytotoxicity assay showed enhanced efficacy of pCNP-SLB over CNP-SLB and free SLB. It was found that pCNP-SLB was most competitive in A549 cell line compared to SW620 and 786-O cell lines as shown by the lowest IC_{50} value of 7.77 μM among different cancer cell lines and time points. Further study conducted using A549 cell line has demonstrated that pCNP-SLB has enhanced the anti-invasive effect of SLB through cell migration assay and suggested the synergistic effect between pCNP and SLB. On the other hand, Annexin V apoptosis assay and cell cycle analysis have also shown that pCNP-SLB revealed a correlated results of enhanced apoptotic death and G1 cell cycle arrest in A549 cells with sustained release properties. These results have therefore revealed the enhanced therapeutic efficacy and sustained release properties of pCNP-SLB which required low dosing to achieve high therapeutic efficacy. Additionally, subsequent study on pCNP encapsulation on protocatechuic acid (PCA) has revealed that pCNP can be utilized to other poorly soluble compounds other than SLB. Conclusively, various physico-chemical analyses have indicated the modification on CNP system and signified the successful encapsulation of SLB into pCNP. Besides, the *in vitro* cellular study has suggested an enhanced therapeutic efficacy of pCNP-SLB compared to conventional CNP-SLB and free SLB towards A549 cells with sustained release properties. This system can potentially be developed into novel delivery system for many other hydrophobic therapeutics to aid in pharmaceutical and biomedical fields.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGUBAHSUAIAN HIDROFOBİK NANOPARTIKEL KITOSAN UNTUK
MEMPERTINGKATKAN PENGHANTARAN TERAPEUTİK DAN CIRI-CIRI
ANTI-KANSER TERHADAP TITISAN SEL-SEL KANSER**

Oleh

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Kanser paru-paru adalah penyebab utama kematian yang berkaitan dengan barah untuk lelaki dan kedua untuk wanita di seluruh dunia, dengan kira-kira 1.6 juta kematian tahunan dilaporkan dan 1.8 juta kes baru ditemui setiap tahun. Penyampaian ubat antikanker secara konvensional menjadi kurang berkesan kerana rintangan intrinsik seperti kelarutan berair yang buruk dan pengumpulan ubat sel yang lemah. Selain itu, kemoterapi menimbulkan pelbagai kesan sampingan yang tidak diingini termasuk sitotoksik, kardiotoxik dan neurotoksik. Oleh itu, penggunaan terapi antikanker semula jadi sebagai alternatif diperlukan untuk mengelakkan akibat yang tidak diingini ini. Silibinin (SLB) adalah ekstrak tumbuhan yang kurang larut yang berasal dari *Silybum marianum* yang biasanya digunakan untuk perawatan penyakit hati. Telah ditunjukkan bahawa dalam kajian baru-baru ini, SLB menunjukkan sifat anti-kanser dengan menghalang pertumbuhan sel, angiogenesis, percambahan, migrasi sel dan menyebabkan kematian apoptosis. Walau bagaimanapun, kelarutan dan pengumpulan sel SLB yang lemah telah membatasi penggunaan sebatian semula jadi ini sebagai terapi antikanker semula jadi alternatif. Oleh itu, penggunaan aplikasi berasaskan nanoteknologi telah membantu kemajuan diagnosis yang lebih baik dan rawatan penyakit yang lebih cekap, termasuk barah. Selain itu, penggunaan penghantaran nanopartikel dapat mengurangkan dos sebatian terapi semula jadi, di mana dapat mengurangkan kemungkinan kesan sampingan terapi. Berdasarkan hasilnya, peratusan kecekapan enkapsulasi rendah dicapai oleh sistem nanopartikel kitosan konvensional (CNP). Ini telah menunjukkan keperluan untuk memodifikasi sistem CNP ini melalui modifikasi hidrofobik dengan menggunakan NHS-palmitic acid untuk menghasilkan sistem nanopartikel palmitoyl-kitosan (pCNP) untuk peningkatan kecekapan enkapsulasi

dan keberkesanan terapi. Pelbagai analisis pencirian fiziko-kimia telah dilakukan untuk menilai sistem pCNP. Didapati bahawa penggunaan peratusan amina bebas pCNP meningkat sekitar 15% disebabkan oleh konjugasi -NHS asid palmitik ke polimer sebagai tambahan kepada TPP. PCNP yang disintesis menggunakan kaedah persiapan yang sama telah menunjukkan pengembangan dalam ukuran dan penurunan dalam indeks polidispersi (PDI) dibandingkan dengan CNP, yang menunjukkan monodispersity yang lebih besar dalam pCNP. Enkapsulasi SLB ke dalam pCNP telah menunjukkan pengembangan dalam ukuran dan kenaikan PDI, menandakan heterogenitas nanopartikel setelah enkapsulasi dan berkorelasi sesuai dengan analisis morfologi. Lebih-lebih lagi, kecekapan enkapsulasi telah meningkat sekitar 1 kali ganda dari 25% CNP-SLB menjadi 50% pCNP-SLB. Uji sitotoksitas MTT yang seterusnya menunjukkan keberkesanan pCNP-SLB yang lebih tinggi berbanding CNP-SLB dan SLB percuma. Didapati bahawa pCNP-SLB paling kompetitif dalam sel sel A549 berbanding dengan garis sel SW620 dan 786-O seperti yang ditunjukkan oleh nilai IC_{50} terendah 7.77 μM di antara garis sel kanser dan titik waktu yang berbeza. Kajian lebih lanjut yang dilakukan menggunakan garis sel A549 telah menunjukkan bahawa pCNP-SLB telah meningkatkan kesan anti-invasif SLB melalui ujian migrasi sel dan menyarankan kesan sinergis antara pCNP dan SLB. Sebaliknya, analisis apoptosis Annexin V dan analisis kitaran sel juga menunjukkan bahawa pCNP-SLB mendedahkan hasil yang berkaitan dengan peningkatan kematian apoptosis dan penangkapan kitaran sel G1 pada sel A549 dengan sifat pelepasan berkesinambungan. Oleh itu, hasil ini telah menunjukkan peningkatan keberkesanan terapi dan sifat pelepasan berkesinambungan pCNP-SLB yang memerlukan dos rendah untuk mencapai keberkesanan terapi yang tinggi. Selain itu, kajian seterusnya mengenai enkapsulasi pCNP pada asam protocatechuic (PCA) telah menunjukkan bahawa pCNP dapat digunakan untuk sebatian lain yang kurang larut selain SLB. Kesimpulannya, pelbagai analisis fiziko-kimia telah menunjukkan pengubahsuaian pada sistem CNP dan menandakan kejayaan enkapsulasi SLB ke dalam pCNP. Selain itu, kajian sel *in vitro* telah menunjukkan peningkatan keberkesanan terapi pCNP-SLB berbanding dengan CNP-SLB konvensional dan SLB bebas terhadap sel A549 dengan sifat pelepasan berterusan. Sistem ini berpotensi dikembangkan menjadi sistem penyampaian novel untuk banyak terapi hidrofobik lain untuk membantu bidang farmaseutikal dan bioperubatan.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

786-0	Adenocarcinomic human renal epithelial cell line
A549	Adenocarcinomic human alveolar basal epithelial cell line
A _{288nm}	Absorbance at 288nm
A _{405nm}	Absorbance at 405nm
A _{570nm}	Absorbance at 570nm
ATR-FTIR	Attenuated Total Reflection-Fourier Transform Infrared spectroscopy
CNP	Chitosan nanoparticle
CNP-SLB	Silibinin encapsulated chitosan nanoparticle
CNT	Carbon Nanotube
Conc.	Concentration
CS	Chitosan
CVD	cardiovascular disease
dH ₂ O	Deionized distilled water
DLS	Dynamic light scattering
DMEM	Dulbecco's Modified Eagle Medium
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
E.E	Encapsulation Efficiency
EDTA	Ethylenediaminetetraacetic acid
FBS	Feta bovine serum
FDA	Food and Drug administration
FESEM	Field emission-scanning electron microscopy
FITC	Fluorescein isothiocyanate
HCl	Hydrochloric acid
IC ₅₀	Half maximal inhibitory concentration
MRC-5	Human normal lung fibroblast cell line
MTT	3-(4 5-dimethylthiazol-2-yl)-2 5-diphenyltetrazolium bromide
MW	Molecular weight
NaOH	Sodium hydroxide
NH ₂	Amine
NHS	N-hydroxysuccinimide ester group
NSCLC	Non-small cell lung carcinoma
PBS	Phosphate buffer saline
PCA	Protocatechuic acid
pCNP	Palmitoyl chitosan nanoparticle
pCNP-SLB	Silibinin encapsulated palmitoyl chitosan nanoparticle
pCS	Palmitoyl chitosan
PDI	Polydispersity index
PEG	Polyethylene glycol
PI	Propidium Iodide
PSD	Particle size distribution
RNA	Ribonucleic acid
ROS	Reactive oxygen species
rpm	Revolutions per minute
RPMI	Roswell Park Memorial Institute

SCLC	Small cell lung carcinoma
SDS	Sodium dodecyl sulfate
SEM	Scanning electron microscopy
SLB	Silibinin
SW620	adenocarcinomic human colorectal epithelial cell line
TEM	Transmission electron microscopy
TNBS	2,4,6-Trinitrobenzenesulfonic acid
TPP	Sodium tripolyphosphate
v/v	Volume per volume
w/v	Weight per volume



CHAPTER 1

INTRODUCTION

1.1 Background of study

Over the last decade, the incidence and mortality rate of cancer remained the top leading cause of death worldwide (Kooti *et al.*, 2017). Researchers have formulated several means such as surgery, radiation, and chemotherapy to treat cancer patients, but chemotherapy remains the most significant treatment among these approaches, with synthetic drugs such as anthracyclines being the most potent chemotherapeutic drugs used. The four most prominent anthracyclines are epirubicin, doxorubicin, idarubicin and daunorubicin (McGowan *et al.*, 2017). Yet, the severe side effects following the chemotherapy due to non-specific destruction have become the main constrain for the treatment including cardiotoxicity, nausea, and alopecia (Nurgali *et al.*, 2018; Schirrmacher, 2019). Additionally, some chemotherapeutic compounds such as doxorubicin confer lifetime dose limits of not exceed 550 mg/m², which means that a patient can only receive treatment with cumulative lifetime dose of less than the limit or else will encountered with high risk of cardiotoxicity (Cai *et al.*, 2019). Arising from these issues, research focus has since shifted into the utilization of nanotechnology to increase the efficacy and at the same time upsurge the safety of the treatment. The encapsulation of therapeutics by nanoparticles can potentially increase the bioavailability and cellular uptake, thereby can reduce the total dosage of therapeutics to be consumed by patients (Wolfram *et al.*, 2015).

There are many different kinds of nanoparticle systems being formulated by researchers such as carbon nanotube, solid lipid nanoparticle, chitosan nanoparticle and metal nanoparticle. These nano-scale systems generally are 10-100 nm in diameter and able to encapsulate with various cargos attributed to their own properties. The encapsulation of cargos can protect them from random enzymatic degradation and increase the bioavailability, in turn increase the cellular uptake by increasing cellular accumulation (Behzadi *et al.*, 2017; Foroozandeh & Aziz, 2018; Z. Li *et al.*, 2015). Furthermore, recent studies have reported that nanoparticle systems gain the advantage of enhanced permeability and retention effect (EPR) of tumor cells (Kang *et al.*, 2020; J. Park *et al.*, 2019). This means they can significantly increase the cellular accumulation at targeted tumor by crossing the blood vessels and tumor vasculature system present in the body (Golombek *et al.*, 2018; Kalyane *et al.*, 2019).

Chitosan nanoparticle system has emerged as a promising nanoparticle system for the delivery of various kinds of cargos including therapeutic drugs, natural compounds and genes. Chitosan-based nanoparticles possessed positively surface charge and mucoadhesive characteristics which can promote the penetration across the epithelial barriers to facilitate the delivery of cargos

(Mohammed *et al.*, 2017). Besides, the ease of preparation through ionic gelation method with cross-linker such as sodium tripolyphosphate (TPP) with optimized parameter capable to synthesize monodispersed chitosan nanoparticles (CNP). The CNP system is enriched with free amine groups due to N-acetyl glucosamine of chains of chitosan polymer, which provides a conjugation point for compounds and therapeutic genes that are able to conjugate with the amine groups (Sonia & Sharma, 2011). Additionally, the biodegradable and biocompatible properties of CNP have also eliminated that potential problem of accumulation of CNP after consumption (Pang *et al.*, 2017). Therefore, these advantages of CNP have suggested the choice of CNP as a promising delivery system as compared with other nanoparticle systems. Nevertheless, this system is not an ideal route for encapsulation of poorly water soluble compounds such as silibinin (SLB) due to its hydrophobicity and non-charged surface (Biedermann *et al.*, 2014). Although other nanoparticle system, for example solid lipid nanoparticle system is also one of the well-known nano-system for poorly soluble compounds, chitosan nanoparticle derivative system with positive surface charge is preferred. It is because positively charged nanoparticles appeared to have greater cellular uptake efficacy than negatively charged nanoparticles (Jeon *et al.*, 2018).

Silibinin (SLB) is an active natural compound extracted from *Silybum marianum*, commonly known as milk thistle plant. SLB exists as polyphenolic flavonolignan with prominent hepatoprotective and antioxidant activity (Surai, 2015). This natural compound has gained more attentions from researchers since substantial evidences from past and current study have demonstrated its clinical use worldwide (Ferenci, 2016; Tamayo & Diamond, 2007). Furthermore, the utilization of natural compounds can in turn eliminate or minimize the undesirable side effects of synthetic drugs (Mitra & Dash, 2018). However, the utilization of SLB was hindered by constraints of low water solubility and poor cell accumulation (Di Costanzo & Angelico, 2019). The drawbacks of traditional CNP system of low encapsulation and therapeutic efficacy of low water solubility compounds have led to the formulation of hydrophobically-modified chitosan nanoparticle (pCNP) system through the conjugation of NHS-palmitic acid by using the N-hydroxysuccinimide ester group (-NHS) of palmitic acid. The conjugation of the NHS-palmitic acid introduced the hydrophobic moieties to the chitosan polymer and it was hypothesize will be more competent in encapsulation of SLB through the hydrophobic-hydrophobic interactions between SLB and NHS- groups. Palmitic acid is one of the most common saturated fatty acid that can be found naturally in various diet, and existed as an essential component of cell membranes, secretory and transport lipids (German, 2011). In addition, it was reported that the homeostasis of palmitic acid was regulated by endogenous biosynthesis via de novo lipogenesis to prevent its accumulation after consumption (Carta *et al.*, 2017). Thus, the employment of palmitic acid as hydrophobic moiety in this study can potentially avoid undesirable allergic problem and minimum health related concerns.

1.2 Statement of research problem

The cancer disease remains the major cause of mortality globally since many decades ago. Chemotherapeutic agents have been arisen as most common treatment for cancer disease. However, the employment of natural occurring compounds such as phenolic compounds have gained more attention from researchers due to the undesirable side effects following chemotherapy treatment (Mitra & Dash, 2018; Zhang *et al.*, 2018). Phenolic compounds have low water solubility and low accumulation in targeted cells, leading to inefficient delivery routes in treatments. In order to increase the loading, efficacy and release of such compounds, higher dosing needs to be administered leading to non-specific destruction of normal cells and multidrug resistances upon consumption or injection into the human body.

The delivery of therapeutics need the presence of delivery vehicle to protect themselves from enzymatic degradation and enhance the efficacy. Although various delivery vectors including several nanoparticle systems have been synthesized and utilized for the delivery of different therapeutics, the delivery of poorly water-soluble phenolic compounds however required additional attention due to their poor bioavailability and low cellular accumulation. A holistic delivery system for poorly water-soluble compounds in cancer treatment was able to conjugate with the compounds and possessed controlled release properties after administered by patients. Thus, this study is focused on developing a nano-delivery system where can form hydrophobic linkage with hydrophobic compounds to increase the encapsulation efficiency and possessed controlled release properties after administration.

1.3 Research statement of study

Conventional cancer treatments such as chemotherapy and radiotherapy have occasioning a series of undesirable side effects such as cardiotoxicity, nausea, and hair loss in patients (Nurgali *et al.*, 2018). Therefore, the utilization of natural therapeutic alternatives such as plant extracts, herbs and phenolic compounds as anticancer agents have gained attention from researchers to reduce the dosing of therapeutics through nanoparticle encapsulation (Kooti *et al.*, 2017). Nonetheless, the therapeutic efficacy of some of these natural alternatives was restricted by some limitations such as low water solubility which leads to poor bioavailability and low cellular accumulation. This study is deliberate to synthesize a novel hydrophobically-modified chitosan nanoparticle (pCNP) system to increase the encapsulation efficiency of poorly water soluble compounds and subsequently enhance the therapeutic efficacy and controlled release properties. The pCNP system was synthesized through the conjugation of NHS-ester group of NHS-palmitic acid to the amine group of chitosan. Consequently, the NHS-ester groups in chitosan act as hydrophobic moieties associated with poorly soluble compound through hydrophobic-hydrophobic interactions. This pCNP system could serve as potential delivery vehicle for various poorly soluble compounds to aid in medical and pharmaceutical fields.

1.4 Hypothesis of study

Modification to the structure of CNP, which includes a hydrophobic moiety such as palmitic acid will increase the encapsulation efficiency, compound-nanoparticle interaction, and confer sustained release properties of silibinin from nanoparticles.

1.5 Objectives

The general objective of the study was to develop a hydrophobically-modified chitosan nanoparticle system to increase the loading, controlled release and efficacy of poorly-soluble phenolic compound, silibinin (SLB) for increased anti-cancer activities.

The specific objectives is described as follow:

1. To perform hydrophobic modification of conventionally synthesized chitosan nanoparticles using NHS- conjugation with palmitic acid;
2. To characterize the hydrophobically modified-chitosan nanoparticles using various physico-chemical characterization analysis;
3. To evaluate the cytotoxicity efficacy of SLB, CNP-SLB, and pCNP-SLB in various human cancer cell lines; and
4. To analyze the enhanced therapeutic efficacy and sustained release properties of pCNP-SLB in apoptotic cell death and effect in cell cycle.

REFERENCES

- Abd El-Ghaffar, M. A., Akl, M. A., Kamel, A. M., & Hashem, M. S. (2017). Amino acid combined chitosan nanoparticles for controlled release of doxorubicin hydrochloride. *Egyptian Journal of Chemistry*, 60(4), 507–518.
- Abdel-Hafez, S. M., Hathout, R. M., & Sammour, O. A. (2014). Towards better modeling of chitosan nanoparticles production: Screening different factors and comparing two experimental designs. *International Journal of Biological Macromolecules*, 64, 334–340.
- Abràmoff, M. D., Magalhães, P. J., & Ram, S. J. (2004). Image processing with imageJ. *Biophotonics International*, 11(7), 36–41.
- Adhikari, H. S., & Yadav, P. N. (2018). Anticancer activity of chitosan, chitosan derivatives, and their mechanism of action. *International Journal of Biomaterials*, 2018, 27–38.
- Ai, J. W., Liao, W., & Ren, Z. L. (2017). Enhanced anticancer effect of copper-loaded chitosan nanoparticles against osteosarcoma. *RSC Advances*, 7(26), 15971–15977.
- Akbarzadeh, A., Rezaei-Sadabady, R., Davaran, S., Joo, S. W., Zarghami, N., Hanifehpour, Y., Samiei, M., Kouhi, M & Nejati-Koshki, K. (2013). Liposome: Classification, preparation, and applications. *Nanoscale Research Letters*, 8(1), 1–8.
- Ali, S. W., Joshi, M., & Rajendran, S. (2011). Synthesis and characterization of chitosan nanoparticles with enhanced antimicrobial activity. *International Journal of Nanoscience*, 10(04n05), 979–984.
- Alipour, M., Bigdeli, M., Aligholi, H., Rasouljan, B., & Khaksarian, M. (2019). Sustained release of silibinin loaded in chitosan nanoparticle induced apoptosis in glioma cells. *Journal of Biomedical Materials Research Part A*, jbm.a.36827.
- Amidi, M., Mastrobattista, E., Jiskoot, W., & Hennink, W. E. (2010). Chitosan-based delivery systems for protein therapeutics and antigens. *Advanced Drug Delivery Reviews*, 62(1), 59–82.
- Amin, A., Karpowicz, P. A., Carey, T. E., Arbiser, J., Nahta, R., Chen, Z. G., et al. (2015). Evasion of anti-growth signaling: A key step in tumorigenesis and potential target for treatment and prophylaxis by natural compounds. *Seminars in cancer biology*, 35 Suppl, S55–S77.
- Amirsaadat, S., Pilehvar-Soltanahmadi, Y., Zarghami, F., Alipour, S., Ebrahimnezhad, Z., & Zarghami, N. (2017). Silibinin-loaded magnetic nanoparticles inhibit hTERT gene expression and proliferation of lung cancer cells. *Artificial Cells, Nanomedicine, and Biotechnology*, 45(8), 1649–1656.

- Amreddy, N., Babu, A., Muralidharan, R., Munshi, A., & Ramesh, R. (2017). Polymeric nanoparticle-mediated gene delivery for lung cancer treatment. *Topics in Current Chemistry*, 375(2), 1–23.
- Anand, M., Sathyapriya, P., Maruthupandy, M., & Hameedha Beevi, A. (2018). Synthesis of chitosan nanoparticles by TPP and their potential mosquito larvicidal application. *Frontiers in Laboratory Medicine*, 2(2), 72–78.
- Anitha, A., Rejinold, N. S., Bumgardner, J. D., Nair, S. V., & Jayakumar, R. (2012). Approaches for functional modification or cross-linking of chitosan. *Chitosan-Based Systems for Biopharmaceuticals*, 95(107–124). Chichester, UK: John Wiley & Sons, Ltd.
- Ariff, S. A. Y., Yusoff, K., & Masarudin, M. J. (2017). Encapsulation of miRNA in chitosan nanoparticles as a candidate for an anti-metastatic agent in cancer therapy. *Malaysian Applied Biology*, 46(1), 165–170.
- Arruebo, M., Vilaboa, N., Sáez-gutierrez, B., & Lambea, J. (2011). Assessment of the evolution of cancer treatment therapies. *Cancers*, 3, 3279–3330.
- Azizah, A. M., Hashimah, B., Nirmal, K., Siti Zubaidah, A. R., Puteri, N. A., Nabihah, A., Sukumaran, R., Balqis, B., Nadia, S. M. R., Sharifah, S. S S., Rahayu, O., Nur Alham, O., Azlina, A. A. (2019). Malaysia national cancer registry report (MNCRR) 2012-2016. Ministry of Health. *National Cancer Registry (NCI)*. Publication 5.
- Bácskay, I., Nemes, D., Fenyvesi, F., Váradi, J., Vasvári, G., Fehér, P., *et al.* (2018). Role of cytotoxicity experiments in pharmaceutical development. Cytotoxicity. <https://doi.org/10.5772/intechopen.72539>
- Babaei, M., Eshghi, H., Abnous, K., Rahimizadeh, M., & Ramezani, M. (2017). Promising gene delivery system based on polyethylenimine-modified silica nanoparticles. *Cancer Gene Therapy*, 24(4), 156–164.
- Bai, M. Y., Tang, S. L., Chuang, M. H., Wang, T. Y., & Hong, P. Da. (2018). Evaluation of chitosan derivative microparticles encapsulating superparamagnetic iron oxide and doxorubicin as a pH-Sensitive delivery carrier in hepatic carcinoma treatment: An *in vitro* comparison study. *Frontiers in Pharmacology*, 9(1–14).
- Banerjee, T., Mitra, S., Kumar Singh, A., Kumar Sharma, R., & Maitra, A. (2002). Preparation, characterization and biodistribution of ultrafine chitosan nanoparticles. *International Journal of Pharmaceutics*, 243(1–2), 93–105.
- Bangun, H., Tandiono, S., & Arianto, A. (2018). Preparation and evaluation of chitosan-tripolyphosphate nanoparticles suspension as an antibacterial agent. *Journal of Applied Pharmaceutical Science*, 8(12),
- Banik, B. L., Fattahi, P., & Brown, J. L. (2016). Polymeric nanoparticles: The future of nanomedicine. Wiley Interdisciplinary Reviews. *Nanomedicine and Nanobiotechnology*, 8(2), 271–299.

- Barbosa, A. I., Costa Lima, S. A., & Reis, S. (2019). Application of pH-responsive fucoidan/chitosan nanoparticles to improve oral quercetin delivery. *Molecules*, 24(2).
- Barhoum, A., & Luisa García-Betancourt, M. (2018). Physicochemical characterization of nanomaterials: Size, morphology, optical, magnetic, and electrical properties. Emerging applications of nanoparticles and architectural nanostructures: Current prospects and future trends. Elsevier Inc. <https://doi.org/10.1016/B978-0-323-51254-1.00010-5>
- Bayram, D., Çetin, E. S., Kara, M., Özgöçmen, M., & Candan, I. A. (2017). The apoptotic effects of silibinin on MDA-MB-231 and MCF-7 human breast carcinoma cells. *Human and Experimental Toxicology*, 36(6), 573–586.
- Behzadi, S., Serpooshan, V., Tao, W., Hamaly, M. A., Alkawareek, M. Y., Dreaden, E. C., Brown, D., Alkilany, A. M., Farokhzad, O., & Mahmoudi, M. (2017). Cellular uptake of nanoparticles: journey inside the cell. *Chemical Society Reviews*, 46(14), 4218–4244.
- Ben Yehuda Greenwald, M., Ben Sasson, S., & Bianco-Peled, H. (2013). A new method for encapsulating hydrophobic compounds within cationic polymeric nanoparticles. *Journal of Microencapsulation*, 30(6), 580–588.
- Benelmekki, M. (2014). An introduction to nanoparticles and nanotechnology. Designing Hybrid Nanoparticles. <https://doi.org/10.1088/978-1-6270-5469-0ch1>
- Bera, B. (2015). Nanoporous silicon prepared by vapour phase strain etch and sacrificial technique. *International Journal of Computer Applications, Micro(2015)*, 42–45.
- Berger, J., Reist, M., Mayer, J. M., Felt, O., Peppas, N. A., & Gurny, R. (2004). Structure and interactions in covalently and ionically crosslinked chitosan hydrogels for biomedical applications. *European Journal of Pharmaceutics and Biopharmaceutics*, 57(1), 19–34.
- Berghe, T. Vanden, Vanlangenakker, N., Parthoens, E., Deckers, W., Devos, M., Festjens, N., Guerin, C. J., Brunk, U. T., Declercq, W., & Vandenabeele, P. (2010). Necroptosis, necrosis and secondary necrosis converge on similar cellular disintegration features. *Cell Death and Differentiation*, 17(6), 922–930.
- Bergsbaken, T., Fink, S. L., & Cookson, B. T. (2009). Pyroptosis: Host cell death and inflammation. *Nature Reviews Microbiology*, 7(2), 99–109.
- Bhavana, V., Sudharshan, S. J. S., & Madhu, D. (2017). Natural anticancer compounds and their derivatives in clinical trials. Anticancer plants: clinical trials and nanotechnology (pp. 51–104). Singapore: Springer Singapore.
- Biedermann, D., Vavříková, E., Cvak, L., & Křen, V. (2014). Chemistry of silybin. *Nat. Prod. Rep.*, 31(9), 1138–1157.

- Bijak, M. (2017). Silybin, a major bioactive component of milk thistle (*Silybum marianum* L. Gaernt.) chemistry, bioavailability, and metabolism. *Molecules*, 22(11), 1–11.
- Blanco, E., Shen, H., & Ferrari, M. (2015). Principles of nanoparticle design for overcoming biological barriers to drug delivery. *Nature Biotechnology*, 33(9), 941–951.
- Bohn, K. A., Adkins, C. E., Nounou, M. I., & Lockman, P. R. (2017). Inhibition of VEGF and angiopoietin-2 to reduce brain metastases of breast cancer burden. *Frontiers in Pharmacology*, 8(APR), 1–10.
- Brasselet, C., Pierre, G., Dubessay, P., Dols-Lafargue, M., Coulon, J., Maupeu, J., Vallet-Courbin, A., de Baynast, H., Doco, T., Michaud, P., & Delattre, C. (2019). Modification of chitosan for the generation of functional derivatives. *Applied Sciences (Switzerland)*, 9(7).
- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., & Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: A Cancer Journal for Clinicians*, 68(6), 394–424.
- Brydson, J. A. (1999). Relation of structure to electrical and optical properties. *Plastics Materials* (pp. 110–123). Elsevier.
- Cadena-Herrera, D., Esparza-De Lara, J. E., Ramírez-Ibañez, N. D., López-Morales, C. A., Pérez, N. O., Flores-Ortiz, L. F., & Medina-Rivero, E. (2015). Validation of three viable-cell counting methods: Manual, semi-automated, and automated. *Biotechnology Reports*, 7, 9–16.
- Cai, F., Luis, M., Lin, X., Wang, M., Cai, L., Cen, C., & Biskup, E. (2019). Anthracycline-induced cardiotoxicity in the chemotherapy treatment of breast cancer: Preventive strategies and treatment (Review). *Molecular and Clinical Oncology*, 15–23.
- Cai, J. Y., Li, J., Hou, Y. N., Ma, K., Yao, G. D., Liu, W. W., Hayashi, T., Itoh, K., Tashiro, S., Onodera, S., & Ikejima, T. (2018). Concentration-dependent dual effects of silibinin on kanamycin-induced cells death in *Staphylococcus aureus*. *Biomedicine and Pharmacotherapy*, 102(March), 782–791.
- Cai, Z., Jitkaew, S., Zhao, J., Chiang, H. C., Choksi, S., Liu, J., Ward, Y., Wu, L. G., & Liu, Z. G. (2014). Plasma membrane translocation of trimerized MLKL protein is required for TNF-induced necroptosis. *Nature Cell Biology*, 16(1), 55–65.
- Calzoni, E., Cesaretti, A., Polchi, A., Di Michele, A., Tancini, B., & Emiliani, C. (2019). Biocompatible polymer nanoparticles for drug delivery applications in cancer and neurodegenerative disorder therapies. *Journal of Functional Biomaterials*, 10(1), 1–15.
- Cao, B., Zheng, Y., Xi, T., Zhang, C., Song, W., Burugapalli, K., Yang, H., & Ma, Y. (2012). Concentration-dependent cytotoxicity of copper ions on mouse

fibroblasts *in vitro*: Effects of copper ion release from TCu380A vs TCu220C intra-uterine devices. *Biomedical Microdevices*, 14(4), 709–720.

Carrillo, C., Suñé, J. M., Pérez-Lozano, P., García-Montoya, E., Sarrate, R., Fàbregas, A., Miñarro, M., & Ticó, J. R. (2014). Chitosan nanoparticles as non-viral gene delivery systems: Determination of loading efficiency. *Biomedicine and Pharmacotherapy*, 68(6), 775–783.

Carta, G., Murru, E., Banni, S., & Manca, C. (2017). Palmitic acid: Physiological role, metabolism and nutritional implications. *Frontiers in Physiology*, 8(NOV), 1–14.

Cecchini, M. J., Amiri, M., & Dick, F. A. (2012). Analysis of cell cycle position in mammalian cells. *Journal of Visualized Experiments*, (59), 1–7.

Cerqueira, M. A., Pinheiro, A. C., Pastrana, L. M., & Vicente, A. A. (2019). Amphiphilic modified galactomannan as a novel potential carrier for hydrophobic compounds. *Frontiers in Sustainable Food Systems*, 3(March), 1–8.

Chan Siang, K., & Kok Meng John, C. (2016). A review of lung cancer research in Malaysia. *Medical Journal of Malaysia*, 71(1), 70–78.

Chanphai, P., & Tajmir-Riahi, H. A. (2017). Encapsulation of testosterone by chitosan nanoparticles. *International Journal of Biological Macromolecules*, 98, 535–541.

Chao, C.-Y., & Yin, M.-C. (2009). Antibacterial effects of roselle calyx extracts and protocatechuic acid in ground beef and apple juice. *Foodborne Pathogens and Disease*, 6(2), 201–206.

Chatterjee, D. K., Diagaradjane, P., & Krishnan, S. (2011). Nanoparticle-mediated hyperthermia in cancer therapy. *Therapeutic Delivery*, 2(8), 1001–1014.

Chaudhry, Q., Scotter, M., Blackburn, J., Ross, B., Boxall, A., Castle, L., *et al.* (2008). Applications and implications of nanotechnologies for the food sector. *Food Additives and Contaminants - Part A Chemistry, Analysis, Control, Exposure and Risk Assessment*, 25(3), 241–258.

Chen, C. K., Jones, C. H., Mistriotis, P., Yu, Y., Ma, X., Ravikrishnan, A., *et al.* (2013). Poly(ethylene glycol)-block-cationic polylactide nanocomplexes of differing charge density for gene delivery. *Biomaterials*, 34(37), 9688–9699.

Chen, K.-Y., & Zeng, S.-Y. (2018). Fabrication of quaternized chitosan nanoparticles using tripolyphosphate/genipin dual cross-linkers as a protein delivery system. *Polymers*, 10(11), 1226.

Cheung, R., Ng, T., Wong, J., & Chan, W. (2015). Chitosan: An update on potential biomedical and pharmaceutical applications. *Marine Drugs*, 13(8), 5156–5186.

- Chiarugi, P., & Giannoni, E. (2008). Anoikis: A necessary death program for anchorage-dependent cells. *Biochemical Pharmacology*, 76(11), 1352–1364.
- Chittezhath, M., Deep, G., Singh, R. P., Agarwal, C., & Agarwal, R. (2008). Silibinin inhibits cytokine-induced signaling cascades and down-regulates inducible nitric oxide synthase in human lung carcinoma A549 cells. *Molecular Cancer Therapeutics*, 7(7), 1817–1826.
- Choi, M. E., Price, D. R., Ryter, S. W., & Choi, A. M. K. (2019). Necroptosis: A crucial pathogenic mediator of human disease. *JCI Insight*, 4(15), 1–16.
- Choudhari, Y. M., Detane, S. V., Kulthe, S. S., Godhani, C. C., Inamdar, N. N., Shirolkar, S. M., C Borde, L. C., & Mourya, V. K. (2012). Low molecular weight palmitoyl chitosan: Synthesis, characterization and nanoparticle preparation. *Advanced Materials Letters*, 3(6), 487–492.
- Chu, F. (2014). Pyroptosis. *Encyclopedia of Cancer*. 34, 1–3. Berlin, Heidelberg: Springer Berlin Heidelberg.
- Chuang, C. H., Wang, L. Y., Wong, Y. M., & Lin, E. S. (2018). Anti-metastatic effects of isolinderalactone via the inhibition of MMP-2 and up regulation of NM23-H1 expression in human lung cancer A549 cells. *Oncology Letters*, 15(4), 4690–4696.
- Coates, J. (2000). Interpretation of infrared spectra, a practical approach (R.A. Meyer). John Wiley & Sons Ltd, Chichester, 2000.
- Cohen, G. M. (1997). Caspases: the executioners of apoptosis. *The Biochemical Journal*, 326 (1), 1–16.
- Cuff, S., Bonavia, R., Vazquez-Martin, A., Corominas-Faja, B., Oliveras-Ferraro, C., Cuyàs, E., *et al.* (2013). Silibinin meglumine, a water-soluble form of milk thistle silymarin, is an orally active anti-cancer agent that impedes the epithelial-to-mesenchymal transition (EMT) in EGFR-mutant non-small-cell lung carcinoma cells. *Food and Chemical Toxicology*, 60, 360–368.
- Cunha, R. A., Soares, T. A., Rusu, V. H., Pontes, F. J. S., Franca, E. F., & Lins, R. D. (2012). The molecular structure and conformational dynamics of chitosan polymers: An integrated perspective from experiments and computational simulations. *The Complex World of Polysaccharides, Desiree Nedra Karunaratne, IntechOpen*, DOI: 10.5772/51803.
- Dagne, A., Melkamu, T., Schutten, M. M., Qian, X., Upadhyaya, P., Luo, X., & Kassie, F. (2011). Enhanced inhibition of lung adenocarcinoma by combinatorial treatment with indole-3-carbinol and silibinin in A / J mice. *Carcinogenesis*, 32(4), 561–567.
- Debnath, S. K., Saisivam, S., Debanth, M., & Omri, A. (2018). Development and evaluation of chitosan nanoparticles based dry powder inhalation formulations of prothionamide. *PLoS ONE*, 13(1), 1–12.

- Deep, G., & Agarwal, R. (2010). Antimetastatic efficacy of silibinin: Molecular mechanisms and therapeutic potential against cancer. *Cancer and Metastasis Reviews*, 29(3), 447–463.
- Deep, G., Gangar, S. C., Agarwal, C., & Agarwal, R. (2011). Role of E-cadherin in antimigratory and antiinvasive efficacy of silibinin in prostate cancer cells. *Cancer Prevention Research*, 4(8), 1222–1232.
- Deepa, G., Sivakumar, K. C., & Sajeewan, T. P. (2018). Molecular simulation and *in vitro* evaluation of chitosan nanoparticles as drug delivery systems for the controlled release of anticancer drug cytarabine against solid tumours. 3 *Biotech*, 8(12), 493.
- Degterev, A., Huang, Z., Boyce, M., Li, Y., Jagtap, P., Mizushima, N., Cuny, G. D., Mitchison, T. J., Moskowitz, M. A., & Yuan, J. (2005). Chemical inhibitor of nonapoptotic cell death with therapeutic potential for ischemic brain injury. *Nature Chemical Biology*, 1(2), 112–119.
- Demedts, I. K., Vermaelen, K. Y., & Van Meerbeeck, J. P. (2010). Treatment of extensive-stage small cell lung carcinoma: Current status and future prospects. *European Respiratory Journal*, 35(1), 202–215.
- Di Felice, G., & Colombo, P. (2017). Nanoparticle–allergen complexes for allergen immunotherapy. *International Journal of Nanomedicine*, 12, 4493–4504.
- DiCiccio, J. E., & Steinberg, B. E. (2011). Lysosomal pH and analysis of the counter ion pathways that support acidification. *Journal of General Physiology*, 137(4), 385–390.
- Edwardson, D., Narendrula, R., Chewchuk, S., Mispel-Beyer, K., Mapletoft, J., & Parissenti, A. (2015). Role of drug metabolism in the cytotoxicity and clinical efficacy of anthracyclines. *Current Drug Metabolism*, 16(6), 412–426.
- El-banna, F. S., Mahfouz, M. E., Leporatti, S., El-Kemary, M., & Hanafy, N. A. N. (2019). Chitosan as a natural copolymer with unique properties for the development of hydrogels. *Applied Sciences (Switzerland)*, 9(11), 1–11.
- Elgadir, M. A., Uddin, M. S., Ferdosh, S., Adam, A., Chowdhury, A. J. K., & Sarker, M. Z. I. (2015). Impact of chitosan composites and chitosan nanoparticle composites on various drug delivery systems: A review. *Journal of Food and Drug Analysis*, 23(4), 619–629.
- England, C. G., Miller, M. C., Kuttan, A., Trent, J. O., & Frieboes, H. B. (2015). Release kinetics of paclitaxel and cisplatin from two and three layered gold nanoparticles. *European Journal of Pharmaceutics and Biopharmaceutics*, 92(5), 120–129.
- Escobar, M. L., Echeverría, O. M., & Vázquez-Nin, G. H. (2015). Necrosis as programmed cell death. Cell Death - Autophagy, Apoptosis and Necrosis, Tobias M. Ntuli, IntechOpen, DOI: 10.5772/61483.

- Esquivel, R., Juárez, J., Almada, M., Ibarra, J., & Valdez, M. A. (2015). Synthesis and characterization of new thiolated chitosan nanoparticles obtained by ionic gelation method. *International Journal of Polymer Science*, 2015, 502058.
- Fan, W., Yan, W., Xu, Z., & Ni, H. (2012). Formation mechanism of monodisperse, low molecular weight chitosan nanoparticles by ionic gelation technique. *Colloids and Surfaces B: Biointerfaces*, 90(1), 21–27.
- Fard, J. K., Jafari, S., & Eghbal, M. A. (2015). A review of molecular mechanisms involved in toxicity of nanoparticles. *Advanced Pharmaceutical Bulletin*, 5(4), 447–454.
- Farhangi, M., Kobarfard, F., Mahboubi, A., Vatanara, A., & Mortazavi, S. A. (2018). Preparation of an optimized ciprofloxacin-loaded chitosan nanomicelle with enhanced antibacterial activity. *Drug Development and Industrial Pharmacy*, 44(8), 1273–1284.
- Ferenci, P. (2016). Silymarin in the treatment of liver diseases: What is the clinical evidence? *Clinical Liver Disease*, 7(1), 8–10.
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D. M., Forman, D., & Bray, F. (2015). Cancer incidence and mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. *International Journal of Cancer*, 136(5), E359–E386.
- Fernandes, M., Rosel, D., & Brábek, J. (2014). Translation in solid cancer: are size-based response criteria an anachronism? *Clinical and Translational Oncology*, 17(1).
- Fields, R. (1972). The Rapid Determination of Amino Groups with TNBS. *Methods in Enzymology*, 25(C), 464–468.
- Fink, S. L., & Cookson, B. T. (2005). Apoptosis, pyroptosis, and necrosis: Mechanistic description of dead and dying eukaryotic cells. *Infection and Immunity*, 73(4), 1907–1916.
- Flory, F., Chen, Y.-J., & Lin, H.-J. (2013). Optical thin films containing quantum dots. *Optical Thin Films and Coatings* (p. 493–516e). Elsevier.
- Foroozandeh, P., & Aziz, A. A. (2018). Insight into Cellular Uptake and Intracellular Trafficking of Nanoparticles. *Nanoscale Research Letters*, 13.
- Fouad, Y. A., & Aanei, C. (2017). Revisiting the hallmarks of cancer. *American Journal of Cancer Research*, 7(5), 1016–1036.
- Frantz, S., Ducharme, A., Sawyer, D., Rohde, L. E., Kobzik, L., Fukazawa, R., Tracey, D., Allen, H., Lee, R. T., & Kelly, R. A. (2003). Targeted deletion of caspase-1 reduces early mortality and left ventricular dilatation following myocardial infarction. *Journal of Molecular and Cellular Cardiology*, 35(6), 685–694.

- Frisch, Steven. M.; Francis, H. (1994). Disruption of epithelial cell-matrix interactions induces apoptosis. *The Journal of Cell Biology*, 124(4), 619–626.
- Fu, R., Yan, Y., Roberts, C., Liu, Z., & Chen, Y. (2018). The role of dipole interactions in hyperthermia heating colloidal clusters of densely-packed superparamagnetic nanoparticles. *Scientific Reports*, 8(1), 1–10.
- G.N. Hortobdgyi. (1997). Anthracyclines in the Treatment of Cancer. *Drugs*, 54(4), 1–7.
- Galluzzi, L., Bravo-San Pedro, J. M., Kepp, O., & Kroemer, G. (2016). Regulated cell death and adaptive stress responses. *Cellular and Molecular Life Sciences*, 73(11–12), 2405–2410.
- Galluzzi, L., Vitale, I., Aaronson, S. A., Abrams, J. M., Adam, D., Agostinis, P., et al. (2018). Molecular mechanisms of cell death: recommendations of the Nomenclature Committee on Cell Death 2018. *Cell Death & Differentiation*, 25(3), 486–541.
- Gan, Q., & Wang, T. (2007). Chitosan nanoparticle as protein delivery carrier-Systematic examination of fabrication conditions for efficient loading and release. *Colloids and Surfaces B: Biointerfaces*, 59(1), 24–34.
- Gandalovičová, A., Rosel, D., Fernandes, M., Veselý, P., Heneberg, P., Čermák, V., Petruželka, L., Kumar, S., Sanz-Moreno, V., & Brábek, J. (2017). Migrastatics—anti-metastatic and anti-invasion drugs: Promises and challenges. *Trends in Cancer*, 3(6), 391–406.
- Gao, Y., Theng, S. S., Mah, W.-C., & Lee, C. G. L. (2015). Silibinin down-regulates FAT10 and modulate TNF- /IFN- induced chromosomal instability and apoptosis sensitivity. *Biology Open*, 4(8), 961–969.
- Garg, U., Chauhan, S., Nagaich, U., & Jain, N. (2019). Current advances in chitosan nanoparticles based drug delivery and targeting. *Advanced Pharmaceutical Bulletin*, 9(2), 195–204.
- Ge, Y., Zhang, Y., Chen, Y., Li, Q., Chen, J., Dong, Y., & Shi, W. (2011). Silibinin causes apoptosis and cell cycle arrest in some human pancreatic cancer cells. *International Journal of Molecular Sciences*, 12(8), 4861–4871.
- Gelen, V., Şengül, E., Yıldırım, S., & Atila, G. (2018). The protective effects of naringin against 5-fluorouracil-induced hepatotoxicity and nephrotoxicity in rats. *Iranian Journal of Basic Medical Sciences*, 21(4), 404–410.
- German, J. B. (2011). Dietary lipids from an evolutionary perspective: Sources, structures and functions. *Maternal and Child Nutrition*, 7(SUPPL. 2), 2–16.
- Ghadi, A., Mahjoub, S., Tabandeh, F., & Talebnia, F. (2014). Synthesis and optimization of chitosan nanoparticles: Potential applications in nanomedicine and biomedical engineering. *Caspian Journal of Internal Medicine*, 5(3), 156–161.

- Gilmore, A. P. (2005). Anoikis. *Cell Death and Differentiation*, 12, 1473–1477.
- Giorgi, V. S. I., Peracoli, M. T. S., Peracoli, J. C., Witkin, S. S., & Bannwart-Castro, C. F. (2012). Silibinin modulates the NF-kb pathway and pro-inflammatory cytokine production by mononuclear cells from preeclamptic women. *Journal of Reproductive Immunology*, 95(1–2), 67–72.
- Giustini, A. J., Petryk, A. A., Cassim, S. M., Tate, J. A., Baker, I., & Hoopes, P. J. (2010). Magnetic nanoparticle hyperthermia in cancer treatment. *Nano LIFE*, 1(01n02), 10.1142/S1793984410000067.
- Golombek, S. K., May, J. N., Theek, B., Appold, L., Drude, N., Kiessling, F., & Lammers, T. (2018). Tumor targeting via EPR: Strategies to enhance patient responses. *Advanced Drug Delivery Reviews*, 130, 17–38.
- Gong, Y., Fan, Z., Luo, G., Yang, C., Huang, Q., Fan, K., Cheng, H., Jin, K., Ni, Q., Yu, X., & Liu, C. (2019). The role of necroptosis in cancer biology and therapy. *Molecular Cancer*, 18(1), 1–17.
- Gordon, J., Brown, M., & Reynolds, M. (2018). Cell-based methods for determination of efficacy for candidate therapeutics in the clinical management of cancer. *Diseases*, 6(4), 85.
- Greenwell, M., & Rahman, P. K. S. (2015). Medicinal plants: Their use in anticancer treatment. *International Journal of Pharmaceutical Sciences and Research*, 6(11), 4103–4112.
- Gref, R., Domb, A., Quellec, P., Blunk, T., Müller, R. H., Verbavatz, J. M., & Langer, R. (1995). The controlled intravenous delivery of drugs using PEG-coated sterically stabilized nanospheres. *Advanced Drug Delivery Reviews*, 16(2–3), 215–233.
- Gu, J., Al-Bayati, K., & Ho, E. A. (2017). Development of antibody-modified chitosan nanoparticles for the targeted delivery of siRNA across the blood-brain barrier as a strategy for inhibiting HIV replication in astrocytes. *Drug Delivery and Translational Research*, 7(4), 497–506.
- Guo, K. W., & Tam, H.-Y. (2015). TEM morphology of carbon nanotubes (CNTs) and its effect on the life of micropunch. In *The Transmission Electron Microscope - Theory and Applications*. InTech. <https://doi.org/10.5772/60713>
- Haddad, P. S., Haddad, Y., Vallerand, D., & Brault, A. (2011). Antioxidant and hepatoprotective effects of silibinin in a rat model of nonalcoholic steatohepatitis. Evidence-based complementary and alternative medicine, 2011. <https://doi.org/10.1093/ecam/nep164>
- Hamid Akash, M. S., Rehman, K., & Chen, S. (2015). Natural and synthetic polymers as drug carriers for delivery of therapeutic proteins. *Polymer Reviews*, 55(3), 371–406.
- Hanahan, D., & Weinberg, R. A. (2011). Review hallmarks of cancer : The next generation. *Cell*, 144(5), 646–674.

- Hanahan, D., Weinberg, R. A., & Francisco, S. (2000). The hallmarks of cancer *Cell*, 100(1):57-70.
- Hare, J. I., Lammers, T., Ashford, M. B., Puri, S., Storm, G., & Barry, S. T. (2017). Challenges and strategies in anti-cancer nanomedicine development: An industry perspective. *Advanced Drug Delivery Reviews*, 108, 25–38.
- Hassan, U. A., Hussein, M. Z., Alitheen, N. B., Ariff, S. A. Y., & Masarudin, M. J. (2018). *In vitro* cellular localization and efficient accumulation of fluorescently tagged biomaterials from monodispersed chitosan nanoparticles for elucidation of controlled release pathways for drug delivery systems. *International Journal of Nanomedicine*, 13, 5075–5095.
- Hassan, U. A., Hussein, M. Z., & Masarudin, M. J. (2017). Synthesis of a nanoparticle system for the enhanced accumulation of fluorescently-labelled amino acids encapsulated in monodispersed chitosan nanoparticle system. *Malaysian Applied Biology*, 46(1).
- Hassani, S., Laouini, A., Fessi, H., & Charcosset, C. (2015). Preparation of chitosan-TPP nanoparticles using microengineered membranes - Effect of parameters and encapsulation of tacrine. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 482, 34–43.
- Hassanpour, S. H., & Dehghani, M. (2017). Review of cancer from perspective of molecular. *Journal of Cancer Research and Practice*, 4(4), 127–129.
- Hauck, M. L., La Rue, S. M., Petros, W. P., Poulson, J. M., Yu, D., Spasojevic, I., Pruitt, A. F., Klein, A., Case, B., Thrall, D. E., Needham, D., & Dewhirst, M. W. (2006). Phase I trial of doxorubicin-containing low temperature sensitive liposomes in spontaneous canine tumors. *Clinical Cancer Research*, 12(13), 4004–4010.
- Hayat, M. A. (2016). Overview of autophagy. In *autophagy: cancer, other pathologies, inflammation, immunity, infection, and aging* (Vol. 1, pp. 1–71). Elsevier.
- Higuchi, T. (1961). Rate of release of medicaments from ointment bases containing drugs in suspension. *Journal of Pharmaceutical Sciences*, 50(10), 874–875.
- Hirsch, F. R., Scagliotti, G. V, Mulshine, J. L., Kwon, R., Curran, W. J., Wu, Y., & Paz-ares, L. (2017). Lung cancer : current therapies and new targeted treatments. *The Lancet*, 389(10066), 299–311.
- Hossainzadeh, S., Ranji, N., Naderi Sohi, A., & Najafi, F. (2019). Silibinin encapsulation in polymersome: A promising anticancer nanoparticle for inducing apoptosis and decreasing the expression level of miR - 125b/miR - 182 in human breast cancer cells. *Journal of Cellular Physiology*, (April), 1–14.
- Hou, X., Du, H., Quan, X., Shi, L., Zhang, Q., Wu, Y., Liu, Y., Xiao, J., Li, Y., Lu, L., Ai, X., Zhan, M., Yuan, S., & Sun, L. (2018). Silibinin inhibits NSCLC

- metastasis by targeting the EGFR/LOX pathway. *Frontiers in Pharmacology*, 9(FEB), 1–15.
- Huang, M., Khor, E., & Lim, L. (2004). Uptake and cytotoxicity of chitosan molecules and nanoparticles: Effects of molecular weight and degree of deacetylation, *Pharmaceutical research*, 21(2), 344–353.
- Huang, Y., & Lapitsky, Y. (2011). Monovalent salt enhances colloidal stability during the formation of chitosan/tripolyphosphate microgels. *Langmuir*, 27(17), 10392–10399.
- Hunt, C. J. (2019). Technical Considerations in the Freezing, Low-Temperature Storage and Thawing of Stem Cells for Cellular Therapies. *Transfusion Medicine and Hemotherapy*, 46(3), 134–149.
- Inamura, K. (2017). Lung Cancer: Understanding its molecular pathology and the 2015 WHO classification, 7(August), 1–7.
- Inguscio, V., Panzarini, E., & Dini, L. (2012). Autophagy contributes to the death/survival balance in cancer photodynamic therapy. *Cells*, 1(3), 464–491.
- Irving, G. R. B., Iwuji, C. O. O., Morgan, B., Berry, D. P., Steward, W. P., Thomas, A., Brown, K., & Howells, L. M. (2015). Combining curcumin (C3-complex, Sabinsa) with standard care FOLFOX chemotherapy in patients with inoperable colorectal cancer (CUFOX): Study protocol for a randomised control trial. *Trials*, 16(1), 1–10.
- Islam, S., Bhuiyan, M. A. R., & Islam, M. N. (2017). Chitin and chitosan: structure, properties and applications in biomedical engineering. *Journal of Polymers and the Environment*, 25(3), 854–866.
- Israelachvili, J. N. (2011). Special interactions: Hydrogen-bonding and hydrophobic and hydrophilic interactions. *Intermolecular and Surface Forces*, 4, 151–167.
- Jain, K. K. (2008). Drug Delivery Systems - An Overview (pp. 1–50).
- Jain, K. K. (2017). Nanobiotechnology. Reference Module in Life Sciences (Vol. 58, pp. 7250–7257). Elsevier.
- Jain, R. K., & Stylianopoulos, T. (2010). Delivering nanomedicine to solid tumors. *Nature Reviews Clinical Oncology*, 7(11), 653–664.
- Jain, S. (2012). Toxicity issues related to biomedical applications of carbon nanotubes. *Journal of Nanomedicine & Nanotechnology*, 3(5).
- Janes, K. A., Fresneau, M. P., Marazuela, A., Fabra, A., & Alonso, M. J. (2001). Chitosan nanoparticles as delivery systems for doxorubicin. *Journal of Controlled Release*, 73(2–3), 255–267.

- Jeon, S., Clavadetscher, J., Lee, D.-K., Chankeshwara, S., Bradley, M., & Cho, W.-S. (2018). Surface charge-dependent cellular uptake of polystyrene nanoparticles. *Nanomaterials*, 8(12), 1028.
- Ji, J., Wang, L., Yu, H., Chen, Y., Zhao, Y., Zhang, H., Amer, W. A., Sun, Y., Huang, L., & Saleem, M. (2014). Chemical modifications of chitosan and its applications. *Polymer - plastics technology and engineering*, 53(14), 1494–1505.
- Ji, J., Zuo, P., & Wang, Y. L. (2015). Enhanced antiproliferative effect of carboplatin in cervical cancer cells utilizing folate-grafted polymeric nanoparticles. *Nanoscale Research Letters*, 10(1), 1–8.
- Ji, Z., Lin, G., Lu, Q., Meng, L., Shen, X., Dong, L., Fu, C., & Zhang, X. (2012). Targeted therapy of SMMC-7721 liver cancer *in vitro* and *in vivo* with carbon nanotubes based drug delivery system. *Journal of Colloid and Interface Science*, 365(1), 143–149.
- Jintapattanakit, A., Junyaprasert, V. B., & Kissel, T. (2009). The role of mucoadhesion of trimethyl chitosan and PEGylated trimethyl chitosan nanocomplexes in insulin uptake. *Journal of Pharmaceutical Sciences*, 98(12), 4818–4830.
- Johnsen, K. B., & Moos, T. (2016). Revisiting nanoparticle technology for blood-brain barrier transport: Unfolding at the endothelial gate improves the fate of transferrin receptor-targeted liposomes. *Journal of Controlled Release*, 222, 32–46.
- Jonassen, H., Kjøniksen, A. L., & Hiorth, M. (2012). Effects of ionic strength on the size and compactness of chitosan nanoparticles. *Colloid and Polymer Science*, 290(10), 919–929.
- Jothimani, B., Sureshkumar, S., & Venkatachalapathy, B. (2017). Hydrophobic structural modification of chitosan and its impact on nanoparticle synthesis – A physicochemical study. *Carbohydrate Polymers*, 173, 714–720.
- Kafshgari, M. H., Khorram, M., Khodadoost, M., & Sahar, K. (2011). Reinforcement of chitosan nanoparticles obtained by an ionic cross-linking process. *Iranian Polymer Journal*, 20(5), 445–456.
- Kakkar, S., & Bais, S. (2014). A review on protocatechuic acid and its pharmacological potential. *ISRN Pharmacology*, 2014, 1–9.
- Kalagatur, N. K., Nirmal Ghosh, O. S., Sundararaj, N., & Mudili, V. (2018). Antifungal activity of chitosan nanoparticles encapsulated with Cymbopogon martinii essential oil on plant pathogenic fungi Fusarium graminearum. *Frontiers in Pharmacology*, 9(JUN), 1–13.
- Kalam, M. A., Sultana, Y., Ali, A., Aqil, M., Mishra, A. K., Aljuffali, I. A., & Alshamsan, A. (2013). Part I: Development and optimization of solid-lipid nanoparticles using Box-Behnken statistical design for ocular delivery of

- gatifloxacin. *Journal of Biomedical Materials Research - Part A*, 101 A(6), 1813–1827.
- Kale, R. N., & Bajaj, A. N. (2010). Ultraviolet spectrophotometric method for determination of gelatin crosslinking in the presence of amino groups. *Journal of Young Pharmacists*, 2(1), 90–94.
- Kalyane, D., Raval, N., Maheshwari, R., Tambe, V., Kalia, K., & Tekade, R. K. (2019). Employment of enhanced permeability and retention effect (EPR): Nanoparticle-based precision tools for targeting of therapeutic and diagnostic agent in cancer. *Materials Science and Engineering C*, 98(2018), 1252–1276.
- Kaminsky, V., & Zhivotovsky, B. (2010). To kill or be killed: How viruses interact with the cell death machinery: Symposium. *Journal of Internal Medicine*, 267(5), 473–482.
- Kang, H., Rho, S., Stiles, W. R., Hu, S., Baek, Y., Hwang, D. W., Kashiwagi, S., Kim, M. S., & Choi, H. S. (2020). Size-dependent EPR effect of polymeric nanoparticles on tumor targeting. *Advanced Healthcare Materials*, 9(1), 8–15.
- Karagozlu, M. Z., Karadeniz, F., & Kim, S. K. (2013). Anticancer effects of chitin and chitosan derivatives. *Chitin and chitosan derivatives: advances in drug discovery and developments* (1st ed., Vol. 72). Elsevier Inc.
- Katas, H., & Alpar, H. O. (2006). Development and characterisation of chitosan nanoparticles for siRNA delivery. *Journal of Controlled Release*, 115(2), 216–225.
- Kaur, H., & Kaur, G. (2014). A critical appraisal of solubility enhancement techniques of polyphenols. *Journal of Pharmaceutics*, 2014, 1–14. 45
- Kavi Rajan, R., Hussein, M. Z., Fakurazi, S., Yusoff, K., & Masarudin, M. J. (2019). Increased ROS scavenging and antioxidant efficiency of chlorogenic acid compound delivered via a chitosan nanoparticulate system for efficient *in vitro* visualization and accumulation in human renal adenocarcinoma cells. *International Journal of Molecular Sciences*, 20(19), 4667.
- Kelland, L. (2007). Targeting the limitless replicative potential of cancer: The telomerase/telomere pathway. *Clinical Cancer Research*, 13(17), 4960–4963.
- Kerr, J. F., Wyllie, A. H., & Currie, A. R. (1972). Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *British Journal of Cancer*, 26(4), 239–257.
- Khan, M. A., Zafaryab, M., Mehdi, S. H., Ahmad, I., & Rizvi, M. M. A. (2016). Characterization and anti-proliferative activity of curcumin loaded chitosan nanoparticles in cervical cancer. *International Journal of Biological Macromolecules*, 93, 242–253.

- Khan, M. A., Zafaryab, M., Mehdi, S. H., Quadri, J., & Rizvi, M. M. A. (2017). Characterization and carboplatin loaded chitosan nanoparticles for the chemotherapy against breast cancer *in vitro* studies. *International Journal of Biological Macromolecules*, 97, 115–122.
- Khani, Y., Pourgholam-Amiji, N., Afshar, M., Otroshi, O., Sharifi-Esfahani, M., Sadeghi-Gandomani, H., Vejdani, M., & Salehiniya, H. (2018). Tobacco smoking and cancer types: A review. *Biomedical Research and Therapy*, 5(4), 2142–2159.
- Khdair, A., Hamad, I., Alkhatib, H., Bustanji, Y., Mohammad, M., Tayem, R., & Aiedeh, K. (2016). Modified-chitosan nanoparticles: Novel drug delivery systems improve oral bioavailability of doxorubicin. *European Journal of Pharmaceutical Sciences*, 93, 38–44.
- Kim, B. R., Seo, H. S., Ku, J. M., Kim, G. J., Jeon, C. Y., Park, J. H., Jang, B. H., Park, S. J., Shin, Y. C., & Ko, S. G. (2013). Silibinin inhibits the production of pro-inflammatory cytokines through inhibition of NF- κ B signaling pathway in HMC-1 human mast cells. *Inflammation Research*, 62(11), 941–950.
- Kim, S. N., Rusling, J. F., & Papadimitrakopoulos, F. (2007). Carbon Nanotubes for Electronic and Electrochemical Detection of Biomolecules. *Advanced Materials*, 19(20), 3214–3228.
- Klionsky, D. J. (2008). Autophagy revisited: A conversation with Christian de Duve. *Autophagy*, 4(6), 740–743.
- Kong, B., Seog, J. H., & Lee, S. B. (2015). Experimental considerations on the cytotoxicity of nanoparticles. *Nanomedicine (London, England)*, 6(5), 1–12.
- Kong, F. Y., Zhang, J. W., Li, R. F., Wang, Z. X., Wang, W. J., & Wang, W. (2017). Unique roles of gold nanoparticles in drug delivery, targeting and imaging applications. *Molecules*, 22(9).
- Kooti, W., Servatyari, K., Behzadifar, M., Asadi-Samani, M., Sadeghi, F., Nouri, B., & Zare Marzouni, H. (2017). Effective medicinal plant in cancer treatment, part 2: Review study. *Journal of Evidence-Based Complementary and Alternative Medicine*, 22(4), 982–995.
- Kore, K. J., Bramhakule, P. P., Rachhadiya, R. M., & Shete, R. V. (2011). Evaluation of tissue level antioxidant activity of *Premna serratifolia* leaf in paracetamol intoxicated wistar albino rats. *International Journal of Pharmacy & Life Sciences*, 2, 909.
- Korsmeyer, R. W., Gurny, R., Doelker, E., Buri, P., & Peppas, N. A. (1983). Mechanisms of solute release from porous hydrophilic polymers. *International Journal of Pharmaceutics*, 15(1), 25–35.
- Kroemer, G., El-Deiry, W. S., Golstein, P., Peter, M. E., Vaux, D., Vandenabeele, P., et al. (2005). Classification of cell death: Recommendations of the nomenclature committee on cell death. *Cell Death and Differentiation*, 12, 1463–1467.

- Kroemer, G., & Levine, B. (2008). Autophagic cell death: the story of a misnomer. *Nature Reviews Molecular Cell Biology*, 9(12), 1004–1010.
- Kuen, C., Fakurazi, S., Othman, S., & Masarudin, M. (2017). Increased loading, efficacy and sustained release of silibinin, a poorly soluble drug using hydrophobically-modified chitosan nanoparticles for enhanced delivery of anticancer drug delivery systems. *Nanomaterials*, 7(11), 379.
- Kumar Yadav, S., Kumarivastava, A., Dev, A., Kaundal, B., Roy Choudhury, S., & Karmakar, S. (2017). Nanomelatonin triggers superior anticancer functionality in a human malignant glioblastoma cell line. *Nanotechnology*, 28(36).
- Kushwaha, S. K. S., Ghoshal, S., Rai, A. K., & Singh, S. (2013). Carbon nanotubes as a novel drug delivery system for anticancer therapy : A review, *Brazilian Journal of Pharmaceutical Sciences*, 49(4), 629-643.
- Kwan, Y. P., Saito, T., Ibrahim, D., Al-Hassan, F. M. S., Ein Oon, C., Chen, Y., et al. (2016). Evaluation of the cytotoxicity, cell-cycle arrest, and apoptotic induction by Euphorbia hirta in MCF-7 breast cancer cells. *Pharmaceutical Biology*, 54(7), 1223–1236.
- Lafay, S., & Gil-Izquierdo, A. (2008). Bioavailability of phenolic acids. *Phytochemistry Reviews*, 7(2), 301–311.
- Lai, F., Fadda, A. M., & Sinico, C. (2013). Liposomes for brain delivery. *Expert Opinion on Drug Delivery*, 10(7), 1003–1022.
- Lai, Y., Chiang, P. C., Blom, J. D., Li, N., Shevlin, K., Brayman, T. G., Yiding Hu, Y. Selbo, J. G., & Hu, L. G. (2008). Comparison of *in vitro* nanoparticles uptake in various cell lines and *in vivo* pulmonary cellular transport in intratracheally dosed rat model. *Nanoscale Research Letters*, 3(9), 321–329.
- Lee, D. W., Lim, C., Israelachvili, J. N., & Hwang, D. S. (2013). Strong adhesion and cohesion of chitosan in aqueous solutions. *Langmuir*, 29(46), 14222–14229.
- Li, H., Zhang, Z., Bao, X., Xu, G., & Yao, P. (2018). Fatty acid and quaternary ammonium modified chitosan nanoparticles for insulin delivery. *Colloids and Surfaces B: Biointerfaces*, 170, 136–143.
- Li, X., Wang, X., Chen, D., & Shuzhi, C. (2011). Antioxidant Activity and Mechanism of Protocatechuic Acid *in vitro*. *Functional Foods in Health and Disease* (Vol. 7).
- Li, Z., Jiang, H., Xu, C., & Gu, L. (2015). A review: Using nanoparticles to enhance absorption and bioavailability of phenolic phytochemicals. *Food Hydrocolloids*, 43, 153–164.

- Liang, C. C., Park, A. Y., & Guan, J. L. (2007). *In vitro* scratch assay: A convenient and inexpensive method for analysis of cell migration *in vitro*. *Nature Protocols*, 2(2), 329–333.
- Lin, W., Hyeon, T., Lanza, G. M., Zhang, M., & Meade, T. J. (2009). Magnetic nanoparticles for early detection of cancer by magnetic resonance imaging. *MRS Bulletin*, 34(6), 441–448.
- Liu, R. H. (2004). Potential synergy of phytochemicals in cancer prevention: mechanism of action. *The Journal of Nutrition*, 134(12 Suppl), 3479S–3485S.
- Liu, Z., Chen, K., Davis, C., Sherlock, S., Cao, Q., Chen, X., & Dai, H. (2008). Drug delivery with carbon nanotubes for *in vivo* cancer treatment. *Cancer Research*, 68(16), 6652–6660.
- Liu, Z., Huang, P., Law, S., Tian, H., Leung, W., & Xu, C. (2018). Preventive effect of curcumin against chemotherapy-induced side-effects. *Frontiers in Pharmacology*, 9(NOV), 1–9.
- López-García, J., Lehocký, M., Humpolíček, P., & Sába, P. (2014). HaCaT keratinocytes response on antimicrobial atelocollagen substrates: Extent of cytotoxicity, cell viability and proliferation. *Journal of Functional Biomaterials*, 5(2), 43–57.
- Lu, S., Zhang, Z., Chen, M., Li, C., Liu, L., & Li, Y. (2017). Silibinin inhibits the migration and invasion of human gastric cancer SGC7901 cells by downregulating MMP-2 and MMP-9 expression via the p38MAPK signaling pathway. *Oncology Letters*, 14(6), 7577 – 7582.
- Lustriane, C., Dwivany, F. M., Suendo, V., & Reza, M. (2018). Effect of chitosan and chitosan-nanoparticles on post harvest quality of banana fruits. *Journal of Plant Biotechnology*, 45(1), 36–44.
- Madureira, A. R., Pereira, A., & Pintado, M. (2016). Chitosan nanoparticles loaded with 2,5-dihydroxybenzoic acid and protocatechuic acid: Properties and digestion. *Journal of Food Engineering*, 174, 8–14.
- Malana, M. A., & Zohra, R. (2013). The release behavior and kinetic evaluation of tramadol HCl from chemically cross linked ter polymeric hydrogels. *DARU, Journal of Pharmaceutical Sciences*, 21(1), 1.
- Malhotra, J., Malvezzi, M., Negri, E., Vecchia, C. La, & Boffetta, P. (2016). Risk factors for lung cancer worldwide. *European Respiratory Journal*, 48, 889–902.
- Man, S. M., Karki, R., & Kanneganti, T. D. (2017). Molecular mechanisms and functions of pyroptosis, inflammatory caspases and inflammasomes in infectious diseases. *Immunological Reviews*, 277(1), 61–75.
- Masarudin, M. J., Cutts, S. M., Evison, B. J., & Pigram, P. J. (2015). Factors determining the stability , size distribution , and cellular accumulation of small

- , monodisperse chitosan nanoparticles as candidate vectors for anticancer drug delivery: Application to the passive encapsulation of [(14)C]-doxorubicin. *Nanotechnology, science and applications*, 8, 67–80.
- Mateen, S., Raina, K., & Agarwal, R. (2013). Chemopreventive and anti-cancer efficacy of silibinin against growth and progression of lung cancer. *Nutrition and Cancer*, 65(sup1), 3–11.
- Mateen, S., Tyagi, A., Agarwal, C., Singh, R. P., & Agarwal, R. (2009). Silibinin inhibits human nonsmall cell lung cancer cell growth through cell-cycle arrest by modulating expression and function of key cell-cycle regulators. *Molecular Carcinogenesis*, 49(3),
- Mathematical models of drug release. (2015). Strategies to modify the drug release from pharmaceutical systems (pp. 63–86). Elsevier.
- Mattu, C., Li, R., & Ciardelli, G. (2013). Chitosan nanoparticles as therapeutic protein nanocarriers: The effect of pH on particle formation and encapsulation efficiency. *Polymer Composites*, 34(9), 1538–1545.
- Mayeen, A., Shaji, L. K., Nair, A. K., & Kalarikkal, N. (2018). Morphological characterization of nanomaterials. Characterization of nanomaterials. Elsevier Ltd.
- McGowan, J. V., Chung, R., Maulik, A., Piotrowska, I., Walker, J. M., & Yellon, D. M. (2017). Anthracycline chemotherapy and cardiotoxicity. *Cardiovascular Drugs and Therapy*, 31(1), 63–75.
- Mima, S., Miya, M., Iwamoto, R., & Yoshikawa, S. (1983). Highly deacetylated chitosan and its properties. *Journal of Applied Polymer Science*, 28(6), 1909–1917.
- Mitra, S., & Dash, R. (2018). Natural Products for the Management and Prevention of Breast Cancer. *Evidence-Based Complementary and Alternative Medicine*, 2018, 8324696.
- Mirzaaghaei, S., Foroughmand, A. M., Saki, G., & Shafiei, M. (2019). Combination of epigallocatechin-3-gallate and silibinin: A novel approach for targeting both tumor and endothelial cells. *ACS Omega*, 4(5), 8421–8430.
- Mody, V., Siwale, R., Singh, A., & Mody, H. (2010). Introduction to metallic nanoparticles. *Journal of Pharmacy and Bioallied Sciences*, 2(4), 282.
- Moghaddam, F. A., Atyabi, F., & Dinarvand, R. (2009). Preparation and *in vitro* evaluation of mucoadhesion and permeation enhancement of thiolated chitosan-pHEMA core-shell nanoparticles. *Nanomedicine: Nanotechnology, Biology, and Medicine*, 5(2), 208–215.
- Mohammed, M. A., Syeda, J. T. M., Wasan, K. M., & Wasan, E. K. (2017). An overview of chitosan nanoparticles and its application in non-parenteral drug delivery. *Pharmaceutics*, 9(4).

- Mombeini, M., Saki, G., Khorsandi, L., & Bavarsad, N. (2018). Effects of silymarin-loaded nanoparticles on HT-29 human colon cancer cells. *Medicina*, 54(1), 1.
- Moreno-Vega, A. I., Gómez-Quintero, T., Nuñez-Anita, R. E., Acosta-Torres, L. S., & Castaño, V. (2012). Polymeric and ceramic nanoparticles in biomedical applications. *Journal of Nanotechnology*, 2012, 936041.
- Morris, C. B. (2007). Cryopreservation of animal and human cell lines. *Methods in Molecular Biology (Clifton, N.J.)*, 368, 227–236.
- Mundargi, R. C., Babu, V. R., Rangaswamy, V., Patel, P., & Aminabhavi, T. M. (2008). Nano/micro technologies for delivering macromolecular therapeutics using poly(d,l-lactide-co-glycolide) and its derivatives. *Journal of Controlled Release*, 125(3), 193–209.
- Murawska, M., Skrzypczak, A., & Kozak, M. (2012). Structure and morphology of gold nanoparticles in solution studied by TEM, SAXS and UV-Vis. *Acta Physica Polonica A*, 121(4), 888–892.
- Murphy, J. M., Czabotar, P. E., Hildebrand, J. M., Lucet, I. S., Zhang, J. G., Alvarez-Diaz, et al. (2013). The pseudokinase MLKL mediates necroptosis via a molecular switch mechanism. *Immunity*, 39(3), 443–453.
- Nagarajan, R. (2008). Nanoparticles: Building blocks for nanotechnology. Nanoparticles: Synthesis, stabilization, passivation, and functionalization. *American Chemical Society*, 996, 2–14.
- Nair, R. S., Morris, A., Billa, N., & Leong, C. O. (2019). An evaluation of curcumin-encapsulated chitosan nanoparticles for transdermal delivery. *AAPS PharmSciTech*, 20(2), 1–13.
- Nallamuthu, I., Devi, A., & Khanum, F. (2014). Chlorogenic acid loaded chitosan nanoparticles with sustained release property, retained antioxidant activity and enhanced bioavailability. *Asian Journal of Pharmaceutical Sciences*, 10(3), 203–211.
- Naser, M. D., Eskandari, R., Zolfagharian, H., Mohammad, M., Dounighi, M. N., & Mohammad Sadeghi, M. A. (2012). Preparation and *in vitro* characterization of chitosan nanoparticles containing Mesobuthus eupeus scorpion venom as an antigen delivery system. *The Journal of Venomous Animals and Toxins Including Tropical Diseases*, 18(1), 44–52.
- Nashaat, D., Elsabahy, M., El-Sherif, T., Hamad, M. A., El-Gindy, G. A., & Ibrahim, E. H. (2018). Development and *in vivo* evaluation of chitosan nanoparticles for the oral delivery of albumin. *Pharmaceutical Development and Technology*, 24(3), 329–337.
- Naskar, S., Koutsu, K., & Sharma, S. (2019). Chitosan-based nanoparticles as drug delivery systems: a review on two decades of research. *Journal of Drug Targeting*, 27(4), 379–393.

- Nasrollahzadeh, M., Sajadi, S. M., Sajjadi, M., & Issaabadi, Z. (2019). Applications of nanotechnology in daily life. Interface science and technology (1st ed., Vol. 28). Elsevier Ltd.
- Navya, P. N., Kaphle, A., Srinivas, S. P., Bhargava, S. K., Rotello, V. M., & Daima, H. K. (2019). Current trends and challenges in cancer management and therapy using designer nanomaterials. *Nano Convergence*, 6(1), 23.
- Negrea, P., Caunii, A., Sarac, I., & Butnariu, M. (2015). The study of infrared spectrum of chitin and chitosan extract as potential sources of biomass. *Digest Journal of Nanomaterials and Biostructures*, 10(4), 1129–1138.
- Negrini, S., Gorgoulis, V. G., & Halazonetis, T. D. (2010). Genomic instability an evolving hallmark of cancer. *Nature Reviews Molecular Cell Biology*, 11(3), 220–228.
- Nelson, P. A., & Owen, J. R. (2003). A high-performance supercapacitor/battery hybrid incorporating templated mesoporous electrodes. *Journal of the Electrochemical Society*, 150(10), 1313–1317.
- Nguyen, M. H., Yu, H., Kiew, T. Y., & Hadinoto, K. (2015). Cost-effective alternative to nano-encapsulation: Amorphous curcumin-chitosan nanoparticle complex exhibiting high payload and supersaturation generation. *European Journal of Pharmaceutics and Biopharmaceutics*, 96, 1–10.
- Nurgali, K., Jagoe, R. T., & Abalo, R. (2018). Editorial: Adverse effects of cancer chemotherapy: Anything new to improve tolerance and reduce sequelae? *Frontiers in Pharmacology*, 9(MAR), 1–3.
- Obaidat, R., Al-Jbour, N., Al-Sou'D, K., Sweidan, K., Al-Remawi, M., & Badwan, A. (2010). Some physico-chemical properties of low molecular weight chitosans and their relationship to conformation in aqueous solution. *Journal of Solution Chemistry*, 39(4), 575–588.
- Okamoto, Y., Taguchi, K., Yamasaki, K., Sakuragi, M., Kuroda, S., & Otagiri, M. (2018). Albumin-encapsulated liposomes: A novel drug delivery carrier with hydrophobic drugs encapsulated in the inner aqueous core. *Journal of Pharmaceutical Sciences*, 107(1), 436–445.
- Oliver Metzger, M., Fuchs, D., Tagscherer, K. E., Gröne, H.-J., Schirmacher, P., & Roth, W. (2016). Inhibition of caspases primes colon cancer cells for 5-fluorouracil-induced TNF- α -dependent necroptosis driven by RIP1 kinase and NF- κ B. *Oncogene*, 35(26), 3399–3409.
- Omar Zaki, S. S., Katas, H., & Hamid, Z. A. (2015). Lineage-related and particle size-dependent cytotoxicity of chitosan nanoparticles on mouse bone marrow-derived hematopoietic stem and progenitor cells. *Food and Chemical Toxicology*, 85, 31–44.
- Omidi, S., & Kakanejadifard, A. (2019). Modification of chitosan and chitosan nanoparticle by long chain pyridinium compounds: Synthesis,

- characterization, antibacterial, and antioxidant activities. *Carbohydrate Polymers*, 208(December 2018), 477–485.
- Ong, S., Ming, L., Lee, K., & Yuen, K. (2016). Influence of the encapsulation efficiency and size of liposome on the oral bioavailability of griseofulvin-loaded liposomes. *Pharmaceutics*, 8(3), 25.
- Ortona, O., D'Errico, G., Mangiapia, G., & Ciccarelli, D. (2008). The aggregative behavior of hydrophobically modified chitosans with high substitution degree in aqueous solution. *Carbohydrate Polymers*, 74(1), 16–22.
- Othman, N., Masarudin, M., Kuen, C., Dasuan, N., Abdullah, L., & Md. Jamil, S. (2018). Synthesis and optimization of chitosan nanoparticles loaded with L-ascorbic acid and thymoquinone. *Nanomaterials*, 8(11), 920.
- Pacheco, I., & Buzea, C. (2017). Metal nanoparticles and their toxicity. In metal nanoparticles (237–293). Weinheim, Germany: Wiley-VCH Verlag GmbH & Co. KGaA.
- Pang, Y., Qin, A., Lin, X., Yang, L., Wang, Q., Wang, Z., Shan, Z., Li, S., Wang, J., Fan, S., & Hu, Q. (2017). Biodegradable and biocompatible high elastic chitosan scaffold is cell-friendly both *in vitro* and *in vivo*. *Oncotarget*, 8(22), 35583–35591.
- Paoli, P., Giannoni, E., & Chiarugi, P. (2013). Anoikis molecular pathways and its role in cancer progression. *Biochimica et Biophysica Acta - Molecular Cell Research*, 1833(12), 3481–3498.
- Pardini, B., Kumar, R., Naccarati, A., Novotny, J., Prasad, R. B., Forsti, A., Hemminki, S., Vodicka, P., & Bermejo, J. L. (2011). 5-Fluorouracil-based chemotherapy for colorectal cancer and MTHFR / MTRR genotypes. *Br J Clin Pharmacol.*, 72(1), 162–163.
- Parida, U. K., Rout, N., & Bindhani, B. K. (2013). properties of chitosan nanoparticles induce apoptosis in human lymphoma SUDHL-4 cell line. *Advances in Bioscience and Biotechnology*, 4(12), 1118–1127.
- Park, J., Choi, Y., Chang, H., Um, W., Ryu, J. H., & Kwon, I. C. (2019). Alliance with EPR effect: Combined strategies to improve the EPR effect in the tumor microenvironment. *Theranostics*, 9(26), 8073–8090.
- Park, S. Y., Jun, S. T., & Marsh, K. S. (2001). Physical properties of PVOH/chitosan-blended films cast from different solvents. *Food Hydrocolloids*, 15(4–6), 499–502.
- Patel, M. R., & San Martin-Gonzalez, M. F. (2012). Characterization of ergocalciferol loaded solid lipid nanoparticles. *Journal of Food Science*, 77(1), 8–13.
- Patel, S., & Nanda, R. (2015). Nanotechnology in healthcare: Applications and challenges. *Medicinal Chemistry*, 5(12), 528–533.

- Patra, J. K., Das, G., Fraceto, L. F., Campos, E. V. R., Rodriguez-Torres, M. D. P., Acosta-Torres, L. S., *et al.* (2018). Nano based drug delivery systems: Recent developments and future prospects. *Journal of Nanobiotechnology*, 16(1), 71.
- Peppas, N. A., & Narasimhan, B. (2014). Mathematical models in drug delivery: How modeling has shaped the way we design new drug delivery systems. *Journal of Controlled Release*, 190, 75–81.
- Pesch, B., Kendzia, B., Gustavsson, P., Jöckel, K., Johnen, G., Pohlabein, H., *et al.* (2012). Cigarette smoking and lung cancer-relative risk estimates for the major histological types from a pooled analysis of case-control studies. *International Journal of Cancer*, 131(5), 1210–1219.
- Pham, T. T., Nguyen, T. H., Thi, T. V., Nguyen, T. T., Le, T. D., Vo, D., Nguyen, D. H., Nguyen, C. K., Nguyen, D. C., Nguyen, T. T., & Bach, L. G. (2019). Investigation of chitosan nanoparticles loaded with protocatechuic acid (pca) for the resistance of *pyricularia oryzae* fungus against rice blast. *Polymers*, 11(1), 177.
- Pink, M., Verma, N., Zerries, A., & Schmitz-Spanke, S. (2017). Dose-dependent response to 3-nitrobenzanthrone exposure in human urothelial cancer cells. *Chemical Research in Toxicology*, 30(10), 1855–1864.
- Polachi, N., Bai, G., Li, T., Chu, Y., Wang, X., Li, S., *et al.* (2016). Modulatory effects of silibinin in various cell signaling pathways against liver disorders and cancer – A comprehensive review. *European Journal of Medicinal Chemistry*, 123, 577–595.
- Pooja, D., Babu Bikkina, D. J., Kulhari, H., Nikhila, N., Chinde, S., Raghavendra, Y. M., Sreedhar, B., & Tiwari, A. K. (2014). Fabrication, characterization and bioevaluation of silibinin loaded chitosan nanoparticles. *International Journal of Biological Macromolecules*, 69, 267–273.
- Prick, J., de Haan, G., Green, A. R., & Kent, D. G. (2014). Clonal heterogeneity as a driver of disease variability in the evolution of myeloproliferative neoplasms. *Experimental Hematology*, 42(10), 841–851.
- Raja, M. A., Arif, M., Feng, C., Zeenat, S., & Liu, C. G. (2017). Synthesis and evaluation of pH-sensitive, self-assembled chitosan-based nanoparticles as efficient doxorubicin carriers. *Journal of Biomaterials Applications*, 31(8), 1182–1195.
- Raval, A., Parikh, J., & Engineer, C. (2010). Mechanism of controlled release kinetics from medical devices. *Brazilian Journal of Chemical Engineering*, 27(2), 211–225.
- Rello, S., Stockert, J. C., Moreno, V., Gámez, A., Pacheco, M., Juarranz, A., Cañete, M., & Villanueva, A. (2005). Morphological criteria to distinguish cell death induced by apoptotic and necrotic treatments. *Apoptosis*, 10(1), 201–208.

- Rho, J. K., Choi, Y. J., Jeon, B.-S., Choi, S. J., Cheon, G. J., Woo, S.-K., Kim, H. R., Kim, C. H., Choi, C. M., & Lee, J. C. (2010). Combined treatment with silibinin and epidermal growth factor receptor tyrosine kinase inhibitors overcomes drug resistance caused by T790M mutation. *Molecular Cancer Therapeutics*, 9(12), 3233–3243.
- Risbud, M. V., Hardikar, A. A., Bhat, S. V., & Bhonde, R. R. (2000). Hydrogels as controlled release system for antibiotic delivery. *Journal of Controlled Release*, 68(1), 23–30.
- Rizvi, S. A. A., & Saleh, A. M. (2018). Applications of nanoparticle systems in drug delivery technology. *Saudi Pharmaceutical Journal*, 26(1), 64–70.
- Romanucci, V., Di Fabio, G., & Zarrelli, A. (2019). A new class of synthetic flavonolignan-like dimers: Still few molecules, but with attractive properties. *Molecules*, 24(1).
- Roychowdhury, S., McMullen, M. R., Pisano, S. G., Liu, X., & Nagy, L. E. (2013). Absence of receptor interacting protein kinase 3 prevents ethanol-induced liver injury. *Hepatology*, 57(5), 1773–1783.
- Ryu, S. R., Noda, I., & Jung, Y. M. (2010). What is the origin of positional fluctuation of spectral features: True frequency shift or relative intensity changes of two overlapped bands? *Applied Spectroscopy*, 64(9), 1017–1021.
- Sadiq, A. A., & Abdul Rassol, A. A. (2014). Formulation and evaluation of silibinin loaded solid lipid nanoparticles for peroral use targeting lower part of gastrointestinal tract. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(1), 55–67.
- Saha, K., Kim, S. T., Yan, B., Miranda, O. R., Alfonso, F. S., Shlosman, D., & Rotello, V. M. (2013). Surface functionality of nanoparticles determines cellular uptake mechanisms in mammalian cells. *Small (Weinheim an der Bergstrasse, Germany)*, 9(2), 300–305.
- Sahariah, P., Gaware, V. S., Lieder, R., Jónsdóttir, S., Hjálmarsson, M., Sigurjonsson, O. E., & Másson, M. (2014). The effect of substituent, degree of acetylation and positioning of the cationic charge on the antibacterial activity of quaternary chitosan derivatives. *Marine Drugs*, 12(8), 4635–4658.
- Sahibzada, M. U. K., Sadiq, A., Khan, S., Faidah, H. S., Naseemullah, Khurram, M., et al. (2017). Fabrication, characterization and *in vitro* evaluation of silibinin nanoparticles: An attempt to enhance its oral bioavailability. *Drug Design, Development and Therapy*, 11, 1453–1464.
- Saikia, C., & Gogoi, P. (2015). Chitosan: a promising biopolymer in drug delivery applications. *Journal of Molecular and Genetic Medicine*, S4, 006.
- Salatin, S., & Yari Khosroushahi, A. (2017). Overviews on the cellular uptake mechanism of polysaccharide colloidal nanoparticles. *Journal of Cellular and Molecular Medicine*, 21(9), 1668–1686.

- Saller, R., Meier, R., & Brignoli, R. (2001). The use of silymarin in the treatment of liver diseases. *Drugs*, 61(14), 2035–2063.
- Sara, S., Solmaz, M. D., & Ahmad, Y. K. (2015). Effect of the surface modification, size, and shape on cellular uptake of nanoparticles. *Cell Biology International*, 39(8), 881–890.
- Sarangapani, S., Patil, A., Ngeow, Y. K., Elsa Mohan, R., Asundi, A., & Lang, M. J. (2018). Chitosan nanoparticles' functionality as redox active drugs through cytotoxicity, radical scavenging and cellular behaviour. *Integrative Biology (United Kingdom)*, 10(5), 313–324.
- Sarkar, S. D., Farrugia, B. L., Dargaville, T. R., & Dhara, S. (2013). Physico-chemical/biological properties of tripolyphosphate cross-linked chitosan based nanofibers. *Materials Science and Engineering C*, 33(3), 1446–1454.
- Scazzocchio, B., Vari, R., Filesi, C., D'Archivio, M., Santangelo, C., Giovannini, C., Iacovelli, A., Silecchia, G., Li Volti, G., Galvano, F., & Masella, R. (2011). Cyanidin-3-O- β -glucoside and protocatechuic acid exert insulin-like effects by upregulating PPAR γ activity in human omental adipocytes. *Diabetes*, 60(9), 2234–2244.
- Schirmacher, V. (2019). From chemotherapy to biological therapy: A review of novel concepts to reduce the side effects of systemic cancer treatment (Review). *International Journal of Oncology*, 54(2), 407–419.
- Servat-Medina, L., González-Gómez, A., Reyes-Ortega, F., Sousa, I. M. O., Queiroz, N. de C. A., Zago, P. M. W., *et al.* (2015). Chitosan–tripolyphosphate nanoparticles as *Arrabidaea chica* standardized extract carrier: Synthesis, characterization, biocompatibility, and antiulcerogenic activity. *International Journal of Nanomedicine*, 10, 3897–3909.
- Shaikh, A. Y., & Shih, J. A. (2012). Chemotherapy-induced cardiotoxicity. *Current Heart Failure Reports*, 9(2), 117–127.
- Shameli, K., Ahmad, M. B., Jaffar, E. A., Ibrahim, N. A., Shabanzadeh, P., Rustaiyan, A., *et al.* (2012). Green biosynthesis of silver nanoparticles using *callicarpa maingayi* stem bark extraction. *Molecules*, 17(7), 8506–8517.
- Sharma, A., Boise, L. H., & Shanmugam, M. (2019). Cancer metabolism and the evasion of apoptotic cell death. *Cancers*, 11(8), 1–20.
- Sharma, D., & Singh, J. (2017). Synthesis and characterization of fatty acid grafted chitosan polymer and their nanomicelles for nonviral gene delivery applications. *Bioconjugate Chemistry*, 28(11), 2772–2783.
- Shen, S., & Codogno, P. (2016). The role of autophagy in cell death. Autophagy: cancer, other pathologies, inflammation, immunity, infection, and aging (Vol. 8). Elsevier Inc.

- Shi, J., Gao, W., & Shao, F. (2017). Pyroptosis: Gasdermin-mediated programmed necrotic cell death. *Trends in Biochemical Sciences*, 42(4), 245–254.
- Shukla, S. K., Mishra, A. K., Arotiba, O. A., & Mamba, B. B. (2013). Chitosan-based nanomaterials: A state-of-the-art review. *International Journal of Biological Macromolecules*, 59, 46–58.
- Siegel, R. A., & Rathbone, M. J. (2012). Overview of controlled release mechanisms. In *fundamentals and applications of controlled release drug delivery* (19–43). Boston, MA: Springer US.
- Siepmann, J., & Peppas, N. A. (2012). Modeling of drug release from delivery systems based on hydroxypropyl methylcellulose (HPMC). *Advanced Drug Delivery Reviews*, 64(SUPPL.), 163–174.
- Singh, P., Singh Grewal, A., Pandita, D., & Lather, V. (2018). Synthesis and evaluation of a series of caffeic acid derivatives as anticancer agents. *Future Journal of Pharmaceutical Sciences*, 4(2), 124–130.
- Singh, R. P., Mallikarjuna, G. U., Sharma, G., Dhanalakshmi, S., Tyagi, A. K., Chan, D. C. F., Agarwal, C., & Agarwal, R. (2004). Oral silibinin inhibits lung tumor growth in athymic nude mice and forms a novel chemocombination with doxorubicin targeting nuclear factor κ B-mediated inducible chemoresistance. *Clinical Cancer Research*, 10(24), 8641–8647.
- Singh, R., Singh, S., & Lillard, J. W. (2008). Past, present, and future technologies for oral delivery of therapeutic proteins. *Journal of Pharmaceutical Sciences*, 97(7), 2497–2523.
- Smits, E., Soetekouw, J. A., Pieters, E., Smits, C., de Wijs-Rot, N., & Vromans, H. (2019). The availability of drug by liposomal drug delivery: Individual kinetics and tissue distribution of encapsulated and released drug in mice after administration of PEGylated liposomal prednisolone phosphate. *Investigational new drugs*, 37(5), 890–901.
- Soane, R. J., Frier, M., Perkins, A. C., Jones, N. S., Davis, S. S., & Illum, L. (1999). Evaluation of the clearance characteristics of bioadhesive systems in humans. *International Journal of Pharmaceutics*, 178(1), 55–65.
- Soliman, G. M., Zhang, Y. L., Merle, G., Cerruti, M., & Barralet, J. (2014). Hydrocaffeic acid-chitosan nanoparticles with enhanced stability, mucoadhesion and permeation properties. *European Journal of Pharmaceutics and Biopharmaceutics*, 88(3), 1026–1037.
- Son, J. H., Kim, S.-Y., Jang, H. H., Lee, S. N., & Ahn, K. J. (2018). Protective effect of protocatechuic acid against inflammatory stress induced in human dermal fibroblasts. *Biomedical Dermatology*, 2(1), 1–5.
- Sonia, T. A., & Sharma, C. P. (2011). Chitosan and its derivatives for drug delivery perspective. In *Nat. Prod. Rep.*, 31, 23–53.

- Souto, G. D., Farhane, Z., Casey, A., Efeoglu, E., McIntyre, J., & Byrne, H. J. (2016). Evaluation of cytotoxicity profile and intracellular localisation of doxorubicin-loaded chitosan nanoparticles. *Analytical and Bioanalytical Chemistry*, 408(20), 5443–5455.
- Sreekumar, S., Goycoolea, F. M., Moerschbacher, B. M., & Rivera-Rodriguez, G. R. (2018). Parameters influencing the size of chitosan-TPP nano- and microparticles. *Scientific Reports*, 8(1), 1–11.
- Sridharan, H., & Upton, J. W. (2014). Programmed necrosis in microbial pathogenesis. *Trends in Microbiology*, 22(4), 199–207.
- Sun, L., Chen, Y., Zhou, Y., Guo, D., Fan, Y., Guo, F., Zheng, Y., & Chen, W. (2017). Preparation of 5-fluorouracil-loaded chitosan nanoparticles and study of the sustained release *in vitro* and *in vivo*. *Asian Journal of Pharmaceutical Sciences*, 12(5), 418–423.
- Surai, P. (2015). Silymarin as a natural antioxidant: an overview of the current evidence and perspectives. *Antioxidants*, 4(1), 204–247.
- Sutapa, B., & Samir, M. (2014). Challenges associated with penetration of nanoparticles across cell and tissue barriers: A review of current status and future prospects. *Nano Today*, 9(2), 223–243.
- Taddei, M. L., Giannoni, E., Fiaschi, T., & Chiarugi, P. (2012). Anoikis: An emerging hallmark in health and diseases. *Journal of Pathology*, 226(2), 380–393.
- Tamayo, C., & Diamond, S. (2007). Review of clinical trials evaluating safety and efficacy of milk thistle (*Silybum marianum* [L.] Gaertn.). *Integrative Cancer Therapies*, 6(2), 146–157.
- Tan, J. M., Karthivashan, G., Gani, S. A., Fakurazi, S., & Hussein, M. Z. (2016). *In vitro* drug release characteristic and cytotoxic activity of silibinin-loaded single walled carbon nanotubes functionalized with biocompatible polymers. *Chemistry Central Journal*, 10(1), 1–12.
- Tanaka, T., Tanaka, T., & Tanaka, M. (2011). Potential cancer chemopreventive activity of protocatechuic acid. *Journal of Experimental and Clinical Medicine*, 3(1), 27–33.
- Tang, D., Kang, R., Berghe, T. Vanden, Vandenabeele, P., & Kroemer, G. (2019). The molecular machinery of regulated cell death. *Cell Research*, 29(5), 347–364.
- Teske, E., Rutteman, G. R., Kirpenstein, J., & Hirschberger, J. (2011). A randomized controlled study into the efficacy and toxicity of pegylated liposome encapsulated doxorubicin as an adjuvant therapy in dogs with splenic haemangiosarcoma. *Veterinary and Comparative Oncology*, 9(4), 283–289.

- Tewari-Singh, N., Jain, A. K., Inturi, S., Agarwal, C., White, C. W., & Agarwal, R. (2012). Silibinin attenuates sulfur mustard analog-induced skin injury by targeting multiple pathways connecting oxidative stress and inflammation. *PLoS ONE*, 7(9).
- Thakkar, H., Patel, B., & Thakkar, S. (2010). A review on techniques for oral bioavailability enhancement of drugs. *International Journal of Pharmaceutical Sciences Review and Research*, 4(3), 203–223.
- Thanou, M. M., Kotzé, A. F., Scharringhausen, T., Lueßen, H. L., De Boer, A. G., Verhoef, J. C., & Junginger, H. E. (2000). Effect of degree of quaternization of N-trimethyl chitosan chloride for enhanced transport of hydrophilic compounds across intestinal Caco-2 cell monolayers. *Journal of Controlled Release*, 64(1–3), 15–25.
- Thotakura, N., Dadarwal, M., Kumar, R., Singh, B., Sharma, G., Kumar, P., Katare, O. P., & Raza, K. (2017). Chitosan-palmitic acid based polymeric micelles as promising carrier for circumventing pharmacokinetic and drug delivery concerns of tamoxifen. *International Journal of Biological Macromolecules*, 102, 1220–1225.
- Tiğli Aydın, R. S., & Pulat, M. (2012). 5-fluorouracil encapsulated chitosan nanoparticles for pH-stimulated drug delivery: Evaluation of controlled release kinetics. *Journal of Nanomaterials*, 2012.
- Tiyaboonchai, W. (2003). Chitosan Nanoparticles : a promising system for drug delivery. *Naresuan University Journal*, 11(3), 51–66.
- Trapani, A., Sitterberg, J., Bakowsky, U., & Kissel, T. (2009). The potential of glycol chitosan nanoparticles as carrier for low water soluble drugs. *International Journal of Pharmaceutics*, 375(1–2), 97–106.
- Tsai, M. L., Chen, R. H., Bai, S. W., & Chen, W. Y. (2011). The storage stability of chitosan/tripolyphosphate nanoparticles in a phosphate buffer. *Carbohydrate Polymers*, 84(2), 756–761.
- Tyagi, A., Agarwal, C., Dwyer-Nield, L. D., Singh, R. P., Malkinson, A. M., & Agarwal, R. (2012). Silibinin modulates TNF- α and IFN- γ mediated signaling to regulate COX2 and iNOS expression in tumorigenic mouse lung epithelial LM2 cells. *Molecular Carcinogenesis*, 51(10), 832–842.
- Usman, M. S., Ibrahim, N. A., Shameli, K., Zainuddin, N., & Yunus, W. M. Z. W. (2012). Copper nanoparticles mediated by chitosan: Synthesis and characterization via chemical methods. *Molecules*, 17(12), 14928–14936.
- Vaid, M., Singh, T., Prasad, R., & Katiyar, S. K. (2015). Silymarin inhibits melanoma cell growth both *in vitro* and *in vivo* by targeting cell cycle regulators, angiogenic biomarkers and induction of apoptosis. *Molecular Carcinogenesis*, 54(11), 1328–1339.

- Veisheh, O., Tang, B. C., Whitehead, K. A., Anderson, D. G., & Langer, R. (2015). Managing diabetes with nanomedicine: challenges and opportunities. *Nature Reviews Drug Discovery*, 14(1), 45–57.
- Velussi, M., Cernigoi, A. M., Ariella, D. M., Dapas, F., Caffau, C., & Zilli, M. (1997). Long-term (23 months) treatment with an anti-oxidant drug (silymarin) is effective on hyperinsulinemia, exogenous insulin need and malondialdehyde levels in cirrhotic diabetic patients. *Journal of Hepatology*, 26(4), 871–879.
- Vimal, S., Taju, G., Nambi, K. S. N., Abdul Majeed, S., Sarath Babu, V., Ravi, M., & Sahul Hameed, A. S. (2012). Synthesis and characterization of CS/TPP nanoparticles for oral delivery of gene in fish. *Aquaculture*, 358–359, 14–22.
- Vimala, K., Yallapu, M. M., Varaprasad, K., Reddy, N. N., Ravindra, S., Naidu, N. S., & Raju, K. M. (2011). Fabrication of curcumin encapsulated chitosan-pva silver nanocomposite films for improved antimicrobial activity. *Journal of Biomaterials and Nanobiotechnology*, 2(1), 55–64.
- Wang, A. Z., Langer, R., & Farokhzad, O. C. (2012). Nanoparticle delivery of cancer drugs. *Annual Review of Medicine*, 63(1), 185–198.
- Wang, J. J., Zeng, Z. W., Xiao, R. Z., Xie, T., Zhou, G. L., Zhan, X. R., & Wang, S. L. (2011). Recent advances of chitosan nanoparticles as drug carriers. *International Journal of Nanomedicine*, 6, 765–774.
- Wang, J. yi, Wang, Y., & Meng, X. (2016). Chitosan nanolayered cisplatin-loaded lipid nanoparticles for enhanced anticancer efficacy in cervical cancer. *Nanoscale Research Letters*, 11(1).
- Wang, L., Feng, M., Li, Q., Qiu, C., & Chen, R. (2019). Advances in nanotechnology and asthma. *Annals of Translational Medicine*, 7(8), 180–180.
- Wang, M., Zhao, J., Zhang, L., Wei, F., Lian, Y., Wu, Y., et al. (2017). Role of tumor microenvironment in tumorigenesis. *Journal of Cancer*, 8(5), 761–773.
- Wang, W., Meng, Q., Li, Q., Liu, J., Zhou, M., Jin, Z., & Zhao, K. (2020). Chitosan derivatives and their application in biomedicine. *International Journal of Molecular Sciences*, 21(2).
- Wang, Y. S., Liu, L. R., Jiang, Q., & Zhang, Q. Q. (2007). Self-aggregated nanoparticles of cholesterol-modified chitosan conjugate as a novel carrier of epirubicin. *European Polymer Journal*, 43(1), 43–51.
- Wang F, Wang YC, Dou S, Xiong MH, Sun TM, Wang, & J. (2011). Doxorubicin-tethered responsive gold nanoparticles facilitate intracellular drug delivery for overcoming multidrug resistance in cancer cells. *ACS Nano*, 5(5), 3679–3692.

- Ways, T. M. M., Lau, W. M., & Khutoryanskiy, V. V. (2018). Chitosan and its derivatives for application in mucoadhesive drug delivery systems. *Polymers*, 10(3).
- Wei, L., Li, Q., Tan, W., Dong, F., Luan, F., Guo, Z., Saso, L., Dux, L., Wegrzyn, G., & Csont, T. (2017). Synthesis, characterization, and the antioxidant activity of double quaternized chitosan derivatives. *Molecules*, 22(3).
- Wei, Z., Jiao, D., & Xu, J. (2015). Using Fourier transform infrared spectroscopy to study effects of magnetic field treatment on wheat (*Triticum aestivum* L.) seedlings. *Journal of Spectroscopy*, 2015, 570190.
- Williams, D. B., & Carter, C. B. (2009). Transmission electron microscopy. Introduction to conventional transmission electron microscopy. Boston, MA: Springer US. <https://doi.org/10.1007/978-0-387-76501-3>
- Wing Ying Cheung, C., Gibbons, N., Wayne Johnson, D., & Lawrence Nicol, D. (2012). Silibinin – a promising new treatment for cancer. *Anti-Cancer Agents in Medicinal Chemistry*, 10(3), 186–195.
- Wolfram, J., Zhu, M., Yang, Y., Shen, J., Gentile, E., Paolino, D., *et al.* (2015). Safety of nanoparticles in medicine. *Current Drug Targets*, 16(14), 1671–1681.
- Wong, P. T., & Choi, S. K. (2015). Mechanisms of drug release in nanotherapeutic delivery systems. *Chemical Reviews*, 115(9), 3388–3432.
- Wong, R. S. Y. (2011). Apoptosis in cancer: from pathogenesis to treatment. *Journal of Experimental & Clinical Cancer Research*, 30(1), 87.
- Wu, C.-H., Lan, C. H., Wu, K. L., Wu, Y. M., Jane, W. N., Hsiao, M., & Wu, H. C. (2018). Hepatocellular carcinoma-targeted nanoparticles for cancer therapy. *International Journal of Oncology*, 52(2), 389–401.
- Wu, J.-W., Lin, L.-C., Hung, S.-C., Chi, C.-W., & Tsai, T.-H. (2007). Analysis of silibinin in rat plasma and bile for hepatobiliary excretion and oral bioavailability application. *Journal of Pharmaceutical and Biomedical Analysis*, 45(4), 635–641.
- Wu, W., Zu, Y., Wang, L., Wang, L., Li, Y., Liu, Y., Wu, M., Zhao, X., & Zhang, X. (2017). Preparation, characterization and antitumor activity evaluation of silibinin nanoparticles for oral delivery through liquid antisolvent precipitation. *RSC Advances*, 7(86), 54379–54390.
- Wulandari, I. O., Sulistyarti, H., Safitri, A., Santjojo, D. J. D. H., & Sabarudin, A. (2019). Development of synthesis method of magnetic nanoparticles modified by oleic acid and chitosan as a candidate for drug delivery agent. *Journal of Applied Pharmaceutical Science*, 9(7), 1–11.
- Xu, J., Ma, L., Liu, Y., Xu, F., Nie, J., & Ma, G. (2012). Design and characterization of antitumor drug paclitaxel-loaded chitosan nanoparticles

- by W/O emulsions. *International Journal of Biological Macromolecules*, 50(2), 438–443.
- Xu, J., Xu, B., Shou, D., Xia, X., & Hu, Y. (2015). Preparation and evaluation of vancomycin-loaded N-trimethyl chitosan nanoparticles. *Polymers*, 7(9), 1850–1870.
- Xuan Nui, P., Tan Phuoc, N., Tuyet Nhung, P., Thi Thuy Nga, T., & Thi Van Thi, T. (2016). Synthesis and characterization of chitosan- coated magnetite nanoparticles and their application in curcumin drug delivery. *Advances in Natural Sciences: Nanoscience and Nanotechnology*, 7(4), 1–9.
- Yadav, D., Sandeep, K., Pandey, D., & Dutta, R. K. (2017). Liposomes for drug delivery. *Journal of Biotechnology & Biomaterials*, 7(4), 377–388.
- Yan, G., Elbadawi, M., & Efferth, T. (2020). Multiple cell death modalities and their key features (Review). *World Academy of Sciences Journal*, 39–48.
- Yang, S. T., Guo, W., Lin, Y., Deng, X. Y., Wang, H. F., Sun, H. F., *et al.* (2007). Biodistribution of pristine single-walled carbon nanotubes in vivo. *Journal of Physical Chemistry C*, 111(48), 17761–17764.
- Yao, H., & Kimura, K. (2007). Field emission scanning electron microscopy for structural characterization of 3D gold nanoparticle superlattices. *Modern Research and Educational Topics in Microscopy*, 568–575.
- Yao, Q., Ye, X., Wang, L., Gu, J., Fu, T., Wang, Y., Lai, Y., Wang, Y., Wang, X., Jin, H., & Guo, Y. (2013). Protective effect of curcumin on chemotherapy-induced intestinal dysfunction. *International journal of clinical and experimental pathology*, 6(11), 2342–2349.
- Yao, Y., & Dai, W. (2014). Genomic instability and cancer. *Journal of Carcinogenesis & Mutagenesis*, 5(2), 1–3.
- Yao, Y., Zang, Y., Qu, J., Tang, M., & Zhang, T. (2019). The toxicity of metallic nanoparticles on liver: The subcellular damages, mechanisms, and outcomes. *International Journal of Nanomedicine*, 14, 8787–8804.
- Yoshida, C., Uchida, Y., Ito, T., Takami, T., & Murakami, Y. (2017). Chitosan gel sheet containing polymeric micelles: Synthesis and gelation properties of PEG-grafted chitosan. *Materials*, 10(9).
- Yu, J., Zhu, Y., Wang, L., Peng, M., Tong, S., Cao, X., Qiu, H., & Xu, X. (2010). Enhancement of oral bioavailability of the poorly water-soluble drug silybin by sodium cholate/phospholipid-mixed micelles. *Acta Pharmacologica Sinica*, 31(6), 759–764.
- Zappa, C., & Mousa, S. A. (2016). Non-small cell lung cancer: Current treatment and future advances. *Translational Lung Cancer Research*, 5(3), 288–300.

- Zare, M., Samani, S. M., & Sobhani, Z. (2018). Enhanced intestinal permeation of doxorubicin using chitosan nanoparticles. *Advanced Pharmaceutical Bulletin*, 8(3), 411–417.
- Zargar, V., Asghari, M., & Dashti, A. (2015). A review on chitin and chitosan polymers: structure, chemistry, solubility, derivatives, and applications. *ChemBioEng Reviews*, 2(3), 204–226.
- Zeng, L., An, L., & Wu, X. (2011). Modeling drug-carrier interaction in the drug release from nanocarriers. *Journal of Drug Delivery*, 2011, 1–15.
- Zeng, R., Tu, M., Liu, H.-W., Zhao, J.-H., Zha, Z.-G., & Zhou, C.-R. (2009). Preparation, structure and drug release behaviour of chitosan-based nanofibres. *IET Nanobiotechnology*, 3(1), 8.
- Zhang, H., Mardiyani, S., Chan, W. C. W., & Kumacheva, E. (2006). Design of biocompatible chitosan microgels for targeted pH-mediated intracellular release of cancer therapeutics. *Biomacromolecules*, 7(5), 1568–1572.
- Zhang, H., Oh, M., Allen, C., & Kumacheva, E. (2004). Monodisperse chitosan nanoparticles for mucosal drug delivery. *Biomacromolecules*, 5(6), 2461–2468.
- Zhang, L., Gu, F., Chan, J., Wang, A., Langer, R., & Farokhzad, O. (2008). Nanoparticles in medicine: therapeutic applications and developments. *Clinical Pharmacology & Therapeutics*, 83(5), 761–769.
- Zhang, Q. Y., Wang, F. X., Jia, K. K., & Kong, L. D. (2018). Natural product interventions for chemotherapy and radiotherapy-induced side effects. *Frontiers in pharmacology*, 9, 1253.
- Zhang, W., Zhang, Z., & Zhang, Y. (2011). The application of carbon nanotubes in target drug delivery systems for cancer therapies. *Nanoscale Research Letters*, 6(1), 555.
- Zhang, X. G., Teng, D. Y., Wu, Z. M., Wang, X., Wang, Z., Yu, D. M., & Li, C. X. (2008). PEG-grafted chitosan nanoparticles as an injectable carrier for sustained protein release. *Journal of Materials Science: Materials in Medicine*, 19(12), 3525–3533.
- Zhang, X., Shi, G. F., Liu, X. Z., An, L. J., & Guan, S. (2011). Anti-ageing effects of protocatechuic acid from *Alpinia* on spleen and liver antioxidative system of senescent mice. *Cell Biochemistry and Function*, 29(4), 342–347.
- Zhao, D., Yu, S., Sun, B., Gao, S., Guo, S., & Zhao, K. (2018). Biomedical applications of chitosan and its derivative nanoparticles. *Polymers*, 10(4).
- Zhao, G., & Xie, Z. (2014). Pyroptosis and neurological diseases. *Neuroimmunology and Neuroinflammation*, 1(2), 60.
- Zhaolin, Z., Guohua, L., Shiyuan, W., & Zuo, W. (2018). Role of pyroptosis in cardiovascular disease. *Cell proliferation*, 52(2), e12563.

- Zhu, X., Su, M., Tang, S., Wang, L., Liang, X., Meng, F., Hong, Y., & Xu, Z. (2012). Synthesis of thiolated chitosan and preparation nanoparticles with sodium alginate for ocular drug delivery. *Molecular vision*, 18, 1973–1982.
- Ziegler, U., & Groscurth, P. (2004). Morphological Features of Cell Death. *Physiology*, 19(3), 124–128.
- Zychlinsky, A., Prevost, M. C., & Sansonetti, P. J. (1992). *Shigella flexneri* induces apoptosis in infected macrophages. *Nature*, 358(6382), 167–169.



BIODATA OF STUDENT

The author was born in Hospital Teluk Intan, Perak Darul Ridzuan in 1991. Due to the hardship of life, he has to follow his parent to move to several places before settled down in a small town, Langkap. He went through his primary education in S.J.K (C) Wah Keow, Langkap, and later attended secondary education in S.M.K Dato' Sagor, Langkap. After SPM, he continued his study in S.M.K Horley Methodist Teluk Intan to pursue his pre-university (STPM) course. Fortunately, he was then offered by Universiti Putra Malaysia for his Bachelor degree for 4 years. He received his Bachelor of Science (Honours) in Cell and Molecular Biology course in 2015. Next, after worked as research assistance for about 4 months, he continued his post-graduate study journey in Master of Science. In January 2018, he has successfully converted his student status and started his PhD journey after a series of conversion examination at Faculty of Biotechnology and Biomolecular Sciences.

LIST OF PUBLICATIONS

Paper publications:

- Kuen, C. Y., Fakurazi, S., Othman, S., & Masarudin, M. (2017). Increased Loading, Efficacy and Sustained Release of Silibinin, a Poorly Soluble Drug Using Hydrophobically-Modified Chitosan Nanoparticles for Enhanced Delivery of Anticancer Drug Delivery Systems. *Nanomaterials*, 7(11), 379.
- Othman, N., Masarudin, M., Kuen, C. Y., Dasuan, N., Abdullah, L., & Md. Jamil, S. (2018). Synthesis and Optimization of Chitosan Nanoparticles Loaded with L-Ascorbic Acid and Thymoquinone. *Nanomaterials*, 8(11), 920.
- Kuen, C. Y., Galen, T., Fakurazi, S., Othman, S. S., & Masarudin, M. J. (2020). Increased Cytotoxic Efficacy of Protocatechuic Acid in A549 Human Lung Cancer Delivered via Hydrophobically Modified-Chitosan Nanoparticles As an Anticancer Modality. *Polymers*, 12(9), 1951
- Kuen, C. Y., Fakurazi, S., Othman, S., & Masarudin, M. Enhanced Therapeutic Efficacy of Silibinin via a Nano-Mediated pCNP Delivery System towards A549 Human Lung Cancer Cells. (in the process of submission to International Journal of Nanomedicine)

Conferences and symposium:

1. Participant in International Congress of the Malaysian Society for Microbiology (ICMSM) 2017, Hotel Bangi Putrajaya.
2. Kuen, C., Fakurazi, S., Othman, S., & Masarudin, M. (2019). Hydrophobic modification of chitosan nanoparticles for increased therapeutic delivery, sustained release, and anticancer properties of silibinin against A549 human lung cancer cell line. Oral speaker at the 3rd International Conference on Molecular Biology & Biotech (ICMBB) 2019. 24-25 April 2019. UCSI University, Kuala Lumpur.
3. Kuen, C., Fakurazi, S., Othman, S., & Masarudin, M. (2019). The enhanced therapeutic efficacy of silibinin encapsulated hydrophobically-modified chitosan nanoparticles in A549 human lung cancer cell line. Oral presenter at the Pure and Applied Biology Colloquium (PABC) 2019. 30th October 2019. Faculty of Applied Sciences, Universiti Teknologi MARA, Shah Alam (Best Oral Presenter)



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