



UNIVERSITI PUTRA MALAYSIA

**ENGINEERING RECOMBINANT NEWCASTLE DISEASE VIRUS WITH
PRO-APOPTOTIC GENE TO ENHANCE ONCOLYSIS IN SELECTED
COLORECTAL CANCER CELLS**

JEEVANATHAN KALYANASUNDRAM

FBSB 2020 12



**ENGINEERING RECOMBINANT NEWCASTLE DISEASE VIRUS WITH
PRO-APOPTOTIC GENE TO ENHANCE ONCOLYSIS IN SELECTED
COLORECTAL CANCER CELLS**

By

JEEVANATHAN KALYANASUNDRAM

Thesis submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of
Doctor of Philosophy

February 2020

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

**ENGINEERING RECOMBINANT NEWCASTLE DISEASE VIRUS WITH
PRO-APOPTOTIC GENE TO ENHANCE ONCOLYSIS IN SELECTED
COLORECTAL CANCER CELLS**

By

JEEVANATHAN KALYANASUNDRAM

February 2020

Chairman: Prof. Datin Paduka Khatijah Yusoff, PhD, FASc
Faculty: Biotechnology and Biomolecular Sciences

Anticancer therapy manipulating oncolytic Newcastle disease virus (NDV) has been reported to show exciting prospects. Since NDV is an avian virus, it is generally cleared by the human immune system, thus making it safe for humans. However, this also reduces viral replication in cancer cells, therefore compromising its full oncolytic potential. The cloning of pro-apoptotic gene such as apoptin into the NDV genome may produce a recombinant NDV with enhanced oncolytic ability which could result in higher tumour killing before being cleared by the immune system. In this study, development of local NDV strain AF2240 as a vector for pro-apoptotic, apoptin transgene delivery into tumour was explored in order to improve AF2240 oncolytic ability. The main objective of the study is to develop a recombinant AF2240 virus armed with pro-apoptotic transgene, capable of higher oncolysis and increased potency by using a selection of colorectal cancer cells as model. Colorectal cancer cells such as HT29, SW620 and CaCo-2 was selected due to their heterogeneity in suppressing pro-apoptotic and anti-viral signalling. A recombinant anti-genomic plasmid, pOLTV5(rAF-ApHGy) was constructed by inserting the apoptin gene into the M and F intergenic region within AF2240 anti-genome. Recombinant AF2240 harbouring apoptin transgene, rAF-ApHGy was successfully recovered from the transfection of the anti-genomic plasmid into BSRT7/5 cells and propagated in embryonated chicken eggs. The recovered rAF-ApHGy virus stably harboured the apoptin transgene through 5 egg passages. The replication kinetics was found to be similar to that of the parental AF2240 strain. The apoptin gene was also detected to be expressed in rAF-ApHGy-infected HT29, SW620 and Caco-2 colorectal cells. The cytotoxicity of rAF-ApHGy was detected to be enhanced compared to wildtype strain AF2240 among infected HT29, SW620 and Caco-2 colorectal cells through MTT assay and flow cytometry analysis. Flow cytometric analysis of infected HT29 showed rAF-ApHGy to result in 26.58%, 34.29% and 31.47% more cell death population compared to AF2240 infection at 24, 48 and 72

h.p.i. Similarly, infection of SW620 cells also revealed rAF-ApHGy resulted in 3.05%, 18.74% and 13.37% more cell death population compared to wildtype AF2240 infection at 24, 48 and 72 h.p.i. Infection of Caco-2 cells with rAF-ApHGy was detected to induce 4.94%, 14.08% and 9.02% more cell death compared to AF2240 infection at 24, 48 and 72 h.p.i. These findings were further corroborated by MTT analysis. Virus infection of HT29 particularly, showed increased population entering into necrotic phase which were determined to be secondary necrosis as appose to necroptosis through RIP-PARP-Caspase 8 cell signalling analysis. The latter also revealed increased cleaved Bax in the rAF-ApHGy infected HT29 cells compared to wildtype strain AF2240, indicating the enhanced oncolytic ability of rAF-ApHGy. Based on these findings, it can be concluded that AF2240 can be genetically modified for pro-apoptotic transgene delivery without compromising virus stability. The enhanced oncolytic ability of rAF-ApHGy highlights the potential of this recombinant strain AF2240 to result in higher oncolysis compared to its wildtype counterpart, before being cleared by patient's immune response during oncovirotherapy.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk ijazah Doktor Falsafah

**KEJURUTERAAN REKOMBINAN NEWCASTLE DISEASE VIRUS
DENGAN GEN PRO-APOPTOTIK BAGI MENINGKATKAN ONKOLISIS
SEL KANSER KLOREKTAL TERPILIH**

Oleh

JEEVANATHAN KALYANASUNDRAM

Februari 2020

Pengerusi: Prof. Datin Paduka Khatijah Yusoff, PhD, FASc
Fakulti: Bioteknologi dan Sains Biomolekul

Terapi antikanser menggunakan virus penyakit Newcastle (NDV) dilaporkan mempunyai prospek yang menarik. Oleh kerana NDV adalah virus burung, ia secara umumnya dibasmi oleh sistem keimunan yang menjadikannya selamat bagi manusia. Namun, ini juga mengurangkan replikasi virus dalam sel kanser yang menjelaskan potensi onkolisisnya. Dalam penyelidikan ini, pembangunan strain NDV tempatan AF2240 yang mengandungi transgen apoptin (rAF-ApHGy) dikaji bagi meningkatkan keupayaan onkolisis strain AF2240. Objektif utama kajian ini adalah untuk membangunkan virus AF2240 rekombinan dengan transgen pro-apoptosis, berkeupayaan untuk mencetuskan onkolisis yang lebih tinggi dan lebih mujarab dengan menggunakan pemilihan sel barah kolorektal sebagai model. Sel-sel barah kolorektal seperti HT29, SW620 dan CaCo-2 dipilih kerana heterogenitasnya dalam memberi isyarat pro-apoptosis dan anti-virus. Plasmid anti-genomik rekombinan, pOLTV5(rAF-ApHGy), dikonstruk dengan selitan gen apoptin di antara bahagian gen M-F di dalam anti-genom strain AF2240. Strain AF2240 rekombinan yang mengandungi transgen ApHGy berjaya diperoleh kembali, melalui transfeksi plasmid anti-genomik ke dalam sel BSRT7/5 dan dibiakkan di dalam telur berembryo. Virus rAF-ApHGy yang diperoleh kembali, mengandungi transgen apoptin dengan stabil sepanjang 5 pemindahan dalam telur dan menunjukkan replikasi kinetik selari dengan virus induk strain AF2240. Ekspresi gen apoptin juga dikesan di dalam sel HT29, SW620 dan Caco-2 yang dijangkiti dengan rAF-ApHGy. Kesitotoksan rAF-ApHGy juga didapati lebih baik berbanding virus induk strain AF2240 di kalangan HT29, SW620 dan Caco-2 melalui analisa asai MTT dan aliran sitometri. Kajian aliran sitometri terhadap sel HT29 yang dijangkiti of rAF-ApHGy menunjukkan 26.58%, 34.29% dan 31.47% lebih banyak kematian sel berbanding dengan AF2240 pada 24, 48 dan 72 j.p.i. Begitu juga dengan jangkitan sel SW620 yang menunjukkan 3.05%, 18.74% dan

13.37% lebih tinggi populasi sel mati berbanding dengan AF2240 pada 24, 48 dan 72 j.p.i. Jangkitan sel Caco-2 dengan rAF-ApHGy turut menunjukkan 4.94%, 14.08% dan 9.02% lebih banyak sel mati berbanding dengan AF2240 jangkitan AF2240 pada 24, 48 and 72 j.p.i. Penemuan ini juga disokong oleh asai MTT. Jangkitan sel HT29 dengan virus menunjuk peningkatan populasi sel menjalani fasa nekrosis yang disahkan sebagai nekrosis sekunder dan bukan nekroptosis melalui analisa pengisyarat sel RIP-PARP-Caspase 8. Analisa pengisyarat sel juga mendedahkan peningkatan Bax terpotong dalam jangkitan HT29 dengan rAF-ApHGy berbanding virus induk AF2240, yang menunjukkan peningkatan kebolehan onkolisis rAF-ApHGy. Berdasarkan kepada penemuan ini, strain AF2240 boleh diubahsuai secara genetik tanpa menjelaskan kestabilan virus. Peningkatan keupayaan onkolitik rAF-ApHGy, meningkatkan potensi virus rekombinan strain AF2240 ini dalam penghapusan tumor yang lebih tinggi semasa onkoviroterapi sebelum disingkirkan oleh sistem imun.

ACKNOWLEDGEMENTS

When Lord Vinayagar won the fruit of knowledge by devotedly walking around his parents, one could not help feeling sorry for his brother who literally raced around the world just to lose it. However, my long arduous quest in obtaining this fruit of knowledge thought me why I should indeed circle my parents for the rest of my life. When the steps were steep, the only thing that persisted were their embracing arms, encouraging words and unconditional love. To my beloved little sister, thank you for turning those inconsolable worries into memorable laughters.

I would also like to express my heartfelt appreciation to my supervisor, Prof. Datin Paduka Dr. Khatijah Yusoff, who has worked hard to secure the project funding. It was a privilege to work under her meticulous supervision. I am forever indebted to my supervisory committee member, Prof. Datuk Wira Dr. Raha Abdul Rahim who ushered me into the biotechnology research field. Her support, generosity and modesty despite being a very busy professor, were exemplary for me. I am also in debt to Associate Prof. Dr. Eddie for his meticulous co-supervision and assistance in experimental design, lab work and writing. I also appreciate the support from Dr. Adelene, Dr. Sarah, Dr. Tan and dear old Freddy, throughout this project.

Lab mates come and go, but Lee Chin, Bei Ru and Hapis stayed when I needed them the most. From spinning Tomy to bustling Beckmen, you all were there, for that, I tip my hat with gratitude. To the children of Virology Lab and Microbial Biotechnology Lab, thank you for maintaining a good working environment.

And lastly to Science, thank you for being a sport whenever I cursed you, for I was naïve to hope you'd go easy on me whenever I picked the micropipette.

I certify that a Thesis Examination Committee has met on (date of viva voce) to conduct the final examination of Jeevanathan Kalyanasundram on his thesis entitled "Engineering Recombinant Newcastle Disease Virus, rNDV (rAF2240) for Targeted Pro-Apoptotic Gene Delivery into Colorectal Cancer Cells" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the University Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the degree of Doctor of Philosophy

Members of the Examination Committee were as follows:

Prof. Dr. Rosfarizan Mohamad

Professor

Faculty of Biotechnology and Biomolecular Science

Universiti Putra Malaysia

(Chairman)

Prof. Dr. Rozita Rosli, PhD

Professor

Faculty of Medicine and Health Science

Universiti Putra Malaysia

(Internal Examiner)

Dr. Muhajir Hamid, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Science

Universiti Putra Malaysia

(Internal Examiner)

Prof. Dr. Lorne Babiuk PhD

Professor

Department of Agricultural Food and Nutritional Science

Faculty of Agricultural Life and Environmental Sciences

University of Alberta

Canada

(External Examiner)

ZURIATI AHMAD ZUKARNAIN, PhD

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy
The members of the Supervisory Committee were as follows:

Khatijah binti Mohd Yusoff, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Raha binti Haji Abdul Rahim, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Chia Suet Lin, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 16 July 2020

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____

Date: _____

Name and Matric No.: Jeevanathan Kalyanasundram (GS48372)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Name of Chairman
of Supervisory
Committee:

Khatijah binti Mohd Yusoff

Signature:

Name of Member of
Supervisory
Committee:

Raha binti Haji Abdul Rahim

Signature:

Name of Member of
Supervisory
Committee:

Chia Suet Lin

TABLE OF CONTENTS

	Page	
ABSTRACT	i	
ABSTRAK	iii	
ACKNOWLEDGEMENTS	v	
APPROVAL	vi	
DECLARATION	vii	
LIST OF TABLES	xiv	
LIST OF FIGURES	xv	
LIST OF APPENDICES	xx	
LIST OF ABBREVIATIONS	xxi	
CHAPTER		
1	INTRODUCTION	1
2	LITERATURE REVIEW	4
2.1	Colorectal Cancer	4
2.1.1	Colorectal cancer treatment	5
2.2	Cancer Oncovirotherapy	6
2.3	Newcastle disease virus (NDV)	9
2.4	NDV strain AF2240	11
2.4.1	AF2240 oncolysis	12
2.4.2	AF2240 tumour selectivity	17
2.4.3	AF2240 immunomodulation	20
2.4.4	AF2240 tropism and challenged	22
2.5	Apoptin	25
2.5.1	Structure of apoptin	28
2.5.2	Nuclear translocation and phosphorylation	29
2.5.3	Apoptin-induced apoptosis	31
2.6	Conclusion	34
3	CONSTRUCTION AND RECOVERY OF RECOMBINANT NDV rAF(GFP-ApHGy) HARBOURING APOPTIN GENE AND GFP REPORTER GENE	35
3.1	Introduction	35
3.2	Materials and Method	37
3.2.1	Cloning strategy	37

3.2.1.1	Plasmid, bacterial strain and growth condition	39
3.2.1.2	Plasmid Extraction	40
3.2.1.3	Agarose Gel Electrophoresis	41
3.2.1.4	PCR	41
3.2.1.5	Gel Purification	41
3.2.1.6	RE Digestion and Dephosphorylation	42
3.2.1.7	DNA Ligation	42
3.2.1.8	Transformation of plasmids through heat shock method	43
3.2.1.9	Colony PCR and construct verification	43
3.2.1.10	Large scale plasmid extraction	44
3.2.2	Recombinant NDV rescue	45
3.2.2.1	Cell culture	46
3.2.2.2	Co-transfection and co-culture	47
3.2.2.3	Virus rescue	47
3.2.2.4	Haemagglutination (HA) assay	47
3.2.3	Recombinant virus stability	48
3.2.3.1	Replication kinetics	48
3.2.3.2	Transgene stability	48
3.2.4	Sucrose gradient purification	49
3.2.4.1	Plaque assay	50
3.2.5	Transgene expression analysis	51
3.2.5.1	Colorectal cancer cell maintenance	51
3.2.5.2	Colorectal cancer cell infection	51
3.2.5.3	Apoptin expression analysis	52
3.2.5.3.1	SDS-PAGE analysis	52
3.2.5.3.2	Western Blot analysis	53
3.3	Result	54
3.3.1	Construction of recombinant anti-genomic plasmid	54
3.3.2	Co-transfection and recovery of recombinant virus	58
3.3.3	Stability analysis of recombinant virus	60
3.3.4	Quantification of purified recombinant virus	61
3.3.5	Transgene expression analysis	63

3.4	Discussion	64
3.5	Conclusion	67
4	<i>IN-VITRO ANALYSIS OF ONCOLYTIC EFFICIENCY OF rAF-ApHGy AND rAF-GFP IN COLORECTAL CANCER CELLS</i>	68
4.1	Introduction	68
4.2	Materials and Method	71
4.2.1	Cell maintenance	71
4.2.2	Cell infection	71
4.2.3	Flow cytometry	71
4.2.4	MTT Assay	72
4.3	Results	72
4.3.1	Infection of HT29	72
4.3.1.1	Flow cytometry analysis of HT29 infection	76
4.3.1.2	MTT assay on HT29 infection	79
4.3.2	Infection of SW620	81
4.3.2.1	Flow cytometry analysis of SW620 infection	84
4.3.2.2	MTT assay on SW620 infection	87
4.3.3	Infection of Caco-2	90
4.3.3.1	Flow cytometry analysis of Caco-2 infection	93
4.3.3.2	MTT assay on Caco-2 infection	97
4.3.4	Infection of NHDF	99
4.3.4.1	Flow cytometry analysis of NHDF infection	101
4.3.4.2	MTT assay on NHDF infection	102
4.3.5	Multiple cell lines infection of recombinant and wildtype virus	104
4.4	Discussion	106
4.5	Conclusion	109
5	<i>CELL SIGNALLING COMPARISON BETWEEN rAF-ApHGy AND AF2240 INFECTED HT29 CELLS</i>	110
5.1	Introduction	110
5.2	Materials and Method	113
5.2.1	Cell culture and maintenance	113
5.2.2	Cell infection	113
5.2.3	Western Blot	113

5.3	Results	114
5.4	Discussion	117
5.5	Conclusion	122
6	CONCLUSION, SIGNIFICANCE OF STUDY AND FUTURE RECOMMENDATIONS	123
REFERENCES		125
APPENDICES		156
BIODATA OF STUDENT		172
PUBLICATION		173

LIST OF TABLES

Table		Page
2.1	Cancer cells utilised for AF2240 <i>in-vitro</i> oncolytic analysis.	12
2.2	Preclinical in-vivo studies on apoptin anti-tumour effects	25
3.1	Primer sequences designed by CloneManager 9.0	37
3.2	Bacterial strain and plasmids utilised in this study	38
3.3	Intergenic primers sequence designed by CloneManager9.0	42
4.1	Colorectal cancer cell stage, origin and DNA instability phenotype	69

LIST OF FIGURES

Figures		Page
2.1	Newcastle disease virus structure.	9
2.2	Apoptosis induction by AF2240 in cancer cells	15
2.3	Tumour selective NDV-mediated apoptosis by putative p38 MAPK/NF- κ B/I κ B α pathway	19
2.4	Immunomodulation by AF2240 through tumour cell infection	21
2.5	Apoptin amino acid sequence	28
3.1	Cloning Strategy	38
3.2	Recombinant rAF-ApHGy and rAF-GFP virus recovery methodology outline	46
3.3	Gel electrophoresis profile for (A) ApHGy (B) gfp gene amplification with flanking sequence using the respective primers	54
3.4	Gel electrophoresis profile for NheI digested and dephosphorylated pOLTV5(rAF)	55
3.5	Gel electrophoresis profile of colony PCR for putative transformants potentially harbouring (A) pOLTV5(rAF-ApHGy) and (B) pOLTV5(rAF-GFP)	56
3.6	Gel electrophoresis profile of harbouring pOLTV5(rAF-ApHGy) clone intergene PCR amplification	57
3.7	Gel electrophoresis profile of harbouring pOLTV5(rAF-GFP) clones intergene PCR amplification	57
3.8	Flourescence microscopy observation and analysis of cells transfected with (A) pCITE-GFP plasmid; (B) pOLTV5(rAF-GFP) anti-genomic plasmid and helper plasmids; (C) Distilled water, dH ₂ O	59
3.9	Haemagglutination assay profile	59

3.10	Gel electrophoresis profile of M-F intergene PCR amplification from 5 SPF egg passage cDNA of (A) rAF-ApHGy and (B) rAF-GFP virus	60
3.11	Replication Kinetics profile of rAF-ApHGy and rAF-GFP in comparison to wildtype AF2240	61
3.12	Plaque assay profile of wildtype AF2240, rAF-ApHGy and rAF-GFP.	62
3.13	Profile of plaques formed by (A) AF2240, (B) rAF-ApHGy and (C) rAF-GFP during plaque assay.	63
3.14	Western Blot chemiluminescent profile of infected (A) HT29, (B) SW620 and (C) Caco-2 (MOI=1, otherwise stated)	63
4.1	Microscopic phase contrast picture of (A) mock infected HT29 along with HT29 infected with (B) AF2240, (C) rAF-GFP and (D) rAF-ApHGy at 24, 48 and 72 h.p.i.	73
4.2	Flourescence microscopy picture of (A) mock infected HT29 along with HT29 infected with (B) AF2240, (C) rAF-GFP and (D) rAF-ApHGy at 24, 48 and 72 h.p.i.	74
4.3	Microscopic phase contrast picture of (A) mock infected HT29 along with HT29 infected with (B) strain AF2240, (C) rAF-GFP and (D) rAF-ApHGy observed at 48 h.p.i.	75
4.4	Flow cytometry of PE-Annexin V stained mock, strain AF2440, rAF-GFP and rAF-ApHGy infected HT29 cells harvested at 24, 48 and 72 h.p.i.	77
4.5	Bar chart for comparison of dead cells population detected by flow cytometry of infected HT29 cells harvested at 24, 48 and 72 h.p.i, stained with Annexin-PE staining kit (BD Bioscience, USA).	78
4.6	MTT profile of infected HT29 cells infected with AF2240, rAF-GFP and rAF-ApHGy at (A) 24 h.p.i, (B) 48 h.p.i and (C) 72 h.p.i.	80
4.7	Bar chart for comparison of percentages of viable HT29 cells between AF2240, rAF-GFP and rAG ApHGy infections at 24, 48 and 72 h.p.i based on MTT assay reading at ~ MOI 1 virus dilution.	81

4.8	Microscopy phase contrast picture of (A) mock infected SW620 along with HT29 infected with (B) strain AF2240, (C) rAF-GFP and (D) rAF-ApHGy at 24, 48 and 72 h.p.i.	82
4.9	Flourescence microscopic picture of (A) mock infected SW620 along with SW620 infected with (B) AF2240, (C) rAF-GFP and (D) rAF-ApHGy at 24, 48 and 72 h.p.i.	83
4.10	Microscopic phase contrast picture of (A) mock infected SW620 along with SW620 infected with (B) AF2240, (C) rAF-GFP and (D) rAF-ApHGy observed at 48 h.p.i.	84
4.11	Flow cytometry of PE-Annexin V stained mock, AF2440, rAF-GFP and rAF-ApHGy infected SW620 cells harvested at 24, 48 and 72 h.p.i.	85
4.12	Bar chart for comparison of dead cells population detected by flow cytometry of infected SW620 cells harvested at 24, 48 and 72 h.p.i, stained with Annexin-PE staining kit (BD Bioscience, USA).	87
4.13	MTT profile of infected SW620 cells infected with AF2240, rAF-GFP and rAF-ApHGy at (A) 24 h.p.i, (B) 48 h.p.i and (C) 72 h.p.i.	88
4.14	Bar chart for comparison of percentages of viable SW620 cells between AF2240, rAF-GFP and rAG ApHGy infections based on MTT assay reading at ~ MOI 1 virus dilution at 24, 48 and 72 h.p.i.	89
4.15	Microscopy phase contrast picture of (A) mock infected Caco-2 along with Caco-2 infected with (B) strain AF2240, (C) rAF-GFP and (D) rAF-ApHGy at 24, 48 and 72 h.p.i.	91
4.16	Flourescence microscopic picture of (A) mock infected Caco-2 along with Caco-2 infected with (B) strain AF2240, (C) rAF-GFP and (D) rAF-ApHGy at 24, 48 and 72 h.p.i.	92
4.17	Microscopic phase contrast picture of (A) mock infected Caco-2 along with Caco-2 infected with (B) strain AF2240, (C) rAF-GFP and (D) rAF-ApHGy observed at 48 h.p.i.	93

4.18	Flow cytometry of PE-Annexin V stained mock, strain AF2240, rAF-GFP and rAF-ApHGy infected Caco-2 cells harvested at 24, 48 and 72 h.p.i.	95
4.19	Bar chart for comparison of dead cells population detected by flow cytometry of infected Caco-2 cells harvested at 24, 48 and 72 h.p.i, stained with Annexin-PE staining kit (BD Bioscience, USA).	96
4.20	MTT profile of infected Caco-2 cells infected with strain AF2240, rAF-GFP and rAF-ApHGy at (A) 24 h.p.i, (B) 48 h.p.i and (C) 72 h.p.i.	98
4.21	Bar chart for comparison of percentages of viable Caco-2 cells between AF2240, rAF-GFP and rAG ApHGy infections based on MTT assay reading at ~ MOI 1 virus dilution at 24, 48 and 72 h.p.i.	99
4.22	Microscopic phase contrast picture of (A) mock infected NHDF along with NHDF infected with (B) strain AF2240, (C) rAF-GFP and (D) rAF-ApHGy at 24, 48 and 72 hours post-infection.	100
4.23	Flourescence microscopic picture of (A) mock infected NHDF along with NHDF infected with (B) strain AF2240, (C) rAF-GFP and (D) rAF-ApHGy at 24, 48 and 72 h.p.i.	101
4.24	Flow cytometry of PE-Annexin V stained mock, strain AF2240, rAF-GFP and rAF-ApHGy infected NHDF cells harvested at 24, 48 and 72 h.p.i.	102
4.25	MTT profile of infected NHDF cells infected with strain AF2240, rAF-GFP and rAF-ApHGy at (A) 24 h.p.i, (B) 48 h.p.i and (C) 72 h.p.i.	104
4.26	Bar chart for comparison of dead cells population detected by flow cytometry of infected NHDF, HT29, SW620 and Caco-2 cells infected with strain AF2240, rAF-GFP and rAF-ApHGy respectively at 48 h.p.i. (MOI = 1), stained with Annexin-PE staining kit (BD Bioscience, USA).	106
5.1	Necroptosis induction through RIP-signalling modulation via death receptor pathway	113
5.2	Western blot profile of RIP signalling analysis.	116
5.3	Western blot profile of apoptotic signalling analysis	117

5.4	Pro-apoptotic TNF- α /TNFR1 signalling by HT29 infected with AF2240 and rAF-ApHGY.	120
5.5	Modulation of mitochondrial associated protein in HT29 cells infected with A) AF2240 and B) rAF-ApHGY.	122



LIST OF APPENDICES

Appendix		Page
A	Cloning strategy and <i>gfp</i> and <i>ApHGy</i> gene insert sequence	157
B	List of chemical components and its compositions	160
C	Buffer compositions for SDS-PAGE gel preparation and analysis	162
D	Replicates of plaque assay and flow cytometric profile	163

LIST OF ABBREVIATIONS

~	approximately
°C	degree Celcius
µg	microgram
µL	microlitre
bp	base pairs
cDNA	complementary deoxynucleotide acid
Da	Dalton
dH ₂ O	distilled water
DNA	deoxyribonucleotide acid
dNTP	deoxyribonucleotide triphosphate
EDTA	ethylenediaminetetraacetic acid
g	gravity force
h	hour
h.p.i.	hours post infection/inoculation
HRP	Horse Radish Peroxidase
j.p.i	jam pos infeksi
kb	kilo base pairs
kDa	kilo Dalton
L	litre
LB	Luria-Bertani
M	Molar
µL	microlitre
mA	milliampere
min	minute
mg	milligram
mL	millilitre
mm	millimetre
mM	millimolar
MgCl ₂	magnesium chloride
NaCl	sodium chloride
NaOH	sodium hydroxide
ng	nanogram
OD	Optical Density
PCR	Polymerase Chain Reaction
RE	Restriction enzymes
rpm	revolutions per minute
sec	seconds
Ta	annealing temperature
Tm	melting temperature
V	volt
v/v	volume per volume
W	Watts
w/v	weight per volume

CHAPTER 1

INTRODUCTION

The Malaysian National Cancer Registry Report 2007-2011 listed colorectal cancer as the most common type of cancer among men and second among women (Azizah *et al.*, 2016). Most colorectal cancer pathogenesis involves the unregulated growth of colonic polyps in the inner lining of colon or rectal which interrupts bowel movement, causing abdominal pain, constipation and bloody stool (Shussman and Wexner, 2014). Surgical resection is the principal form of treatment for the removal of primary colorectal tumours, followed by adjunctive radiotherapy and chemotherapy to prevent localised tumour reoccurrence as well as metastases (Cunningham *et al.*, 2010). However, adverse side effects associated with multi-drug regimens and radiation leads to various medical complications and mortality (Gill *et al.*, 2003; Hafner and Debus, 2016).

In order to circumvent such problems, alternative treatments such as targeted therapy, immunotherapy and hormonal therapy have been developed throughout the years (Palumbo *et al.*, 2013). The intracellular mechanisms as well as immunosuppressive tumour microenvironment developed by colorectal cancer cells in order to resist apoptosis for survival (Watson, 2004), provides targets for alternative anticancer therapy (Kultz, 2005; Lockshin and Zakeri, 2007). Strategies to reinstate the otherwise suppressed apoptotic signalling, have been pursued by directing cytotoxic agents towards cancer cells and delivering pro-apoptotic molecules into tumour cells (Gerl and Vaux, 2005).

Whilst numerous alternative anti-tumour therapies showed success in inducing cytotoxicity against tumour cells, naturally occurring oncolytic viruses such the Newcastle disease virus (NDV) exhibit distinct ability to specifically target, replicate and eradicate tumour cells while leaving normal cells unharmed (Mansour *et al.*, 2011; Zamarin and Palese, 2012). Apart from the direct elimination of tumour cells, NDV has an additional immunotherapeutic potential which enables it to indirectly restore tumour immunosurveillance. This highlights the multifaceted feature of NDV oncovirotherapy which includes tumour specific oncolysis and immunogenicity (Zamarin *et al.*, 2014; Schirrmacher, 2016). Besides, oncolytic NDV is also considered to be safe for human application (Zamarin and Palese, 2012). In addition, the advancement in reverse genetics over the past decades, has spurred the explosion of various recombinant viruses armed with specialised characteristics in order to improve oncovirotherapy (Zamarin and Palese, 2012). This includes enhancing the oncolytic ability of viruses such as NDV through genetic modification by incorporating anti-tumour transgenes into its genome (Vigil *et al.*, 2007; Puhler *et al.*, 2008; Zamarin *et al.*, 2009).

The discovery of apoptin which induces tumour specific apoptosis is one of its kind and intensely researched for the past 25 years. The VP3 protein of chicken anemia virus is the protoplast of apoptin which has been shown to selectively induce apoptosis in tumour cells as opposed to normal cells despite being expressed in both type of cells (Danen-Van Oorschot *et al.*, 1997). Since then, apoptin homologs such as human gyrovirus VP3 protein (Bullenkamp *et al.*, 2012), avian reovirus σC protein (Shih *et al.*, 2004), TT virus-derived apoptosis-inducing protein (TAIP) (Kooistra *et al.*, 2004), and porcine circovirus type 2 VP3 protein (Hough *et al.*, 2015) were discovered to display similar specificity in anti-tumour activity. In contrast to native human pro-apoptotic molecules such as granzymes which are directly reactive (Rousalova and Krepela, 2010), apoptin was found to be inducing intrinsic apoptosis by indirectly modulating Bcl-2 associated proteins (Lin *et al.*, 2004; Maddika *et al.*, 2005; Chaabane *et al.*, 2014). This involves increase in cellular pro-apoptotic factors and enhancement of pro-apoptotic signalling through apoptin tumour specific behaviour such as nuclear translocation (Zhuang *et al.*, 1995b), unregulated multimerization (Heckl *et al.*, 2008), co-interacting proteins (Maddika *et al.*, 2007) and DNA binding ability (Leliveld *et al.*, 2003a). Thus, apoptins have been manipulated for anti-cancer applications by delivery through, plasmid DNA (Mitrus *et al.*, 2005), viral (Liu *et al.*, 2012) and bacterial vectors (Cao *et al.*, 2010).

Although NDV is considered safe for human applications, it is worth noting that this oncolytic virus is an avian virus which is subjected to immune-clearance in humans (Zamarain and Palese, 2012).). This could reduce the oncolytic potential of NDV against tumour during clinical oncovirotherapy. Thus, development of a recombinant NDV with enhanced oncolytic ability is essential in order to trigger higher tumour killing ability before being cleared by the human anti-viral response. This study aims to address the need for a more aggressive and potent NDV by cloning in a pro-apoptotic gene such as apoptin into the genome of local Malaysian NDV isolate strain AF2240. The heterogeneity among cultured colorectal cancer cells (Berg *et al.*, 2017), provides a suitable model to test the improved oncolytic recombinant virus tropism in comparison to wild type strain. The hypothesis of this study is the resulting recombinant NDV AF2240 armed with pro-apoptotic gene could result in higher cytotoxicity against cancer cells such as colorectal cancer cells compared to its parental wildtype strain.

The general objectives of this study is to recover two recombinant AFF240 strain harbouring human gyrovirus apoptin, ApHGy transgene (rAF-ApHGy) and green fluorescence protein, gfp reporter gene (rAF-GFP) respectively via reverse genetics. The gene delivery as well as cytotoxicity of both recombinant viruses were investigated by infecting a selection of colorectal cancers, namely, HT29, SW620 and Caco-2. The oncolytic ability rAF-ApHGy against these colorectal cancer cells was also compared to that of rAF-GFP and parental strain AF2240 in order to determine the rAF-ApHGy enhanced oncolytic activity.

The significance of this study is the development of local recombinant NDV AF2240 strain with enhanced oncolysis features which exhibits potential to be utilise as an alternative primary and secondary anti-cancer therapy. Whilst AF2240 has been shown to display oncolytic ability against wide range of tumour cells (Omar *et al.*, 2003), a recombinant virus with potent oncolytic ability would potentially improve AF2240 based treatment by reducing virus dosage, expanding tumour cell tropism as well as treating therapy resistant tumour cells. In addition, the construction of *gfp* reporter gene harbouring AF2240, rAF-GFP, enabled quick assessment of AF2240 gene delivery ability and verify virus infection which together, substantiated rAF-GFP as reliable infection control for *in-vitro* infection analysis.

The general objective of this study is to recover two recombinant AF2240 viruses harbouring *ApHGy* gene (rAF-ApHGy) as well as *gfp* reporter gene (rAF-GFP) through reverse genetics. Both recombinant viruses will be tested for gene delivery and cytotoxicity against a selection of colorectal cancer cell lines in comparison to wild-type AF2240.

The specific objectives of this research are as follows:

1. To construct and recover of recombinant NDV strain AF2240 harbouring:
 - a. apoptin transgene gene (rAF-ApHGy)
 - b. *gfp* reporter gene (rAF-GFP)
2. To compare and analyse cell cytotoxicity induced by recombinant rAF-ApHGy, rAF-GFP and wildtype strain AF2240 through *in-vitro* infection of:
 - a. HT29 (epithelial human colon adenocarcinoma cell)
 - b. SW620 (metastatic epithelial human colon adenocarcinoma cell)
 - c. Caco-2 (epithelial human colon adenocarcinoma cell)
 - d. Normal Human Dermal Fibroblasts (NHDF) cells
3. To conduct and compare cell death signalling analysis between rAF-ApHGy and AF2240 by using HT29 infection as a model.
 - a. Apoptosis/Necroptosis pathway (RIP-signalling)
 - b. Mitochondrial associated-Bcl proteins regulation

REFERENCES

- Abd-Aziz, N., Stanbridge, E.J.and Shafee, N. 2016. Newcastle disease virus degrades HIF-1 α through proteasomal pathways independent of VHL and p53. *Journal of General Virology* 97, 3174-3182.
- Abdul Rahman, M.S., Chee, Y.S.and Lim, S.S. 1976. Observations on the antibody response of breeder flocks to Ranikhet 'STD' vaccination. *Kajian Veterinar* 8, 48-53.
- Abu Hassan, M. R., and Ismail, I. 2016. Incidence and mortality rates of colorectal cancer in Malaysia. *Journal of Global Oncology* Vol.4; 216s-216s
- Ahmad, U., Ahmed, I., Keong, Y.Y., Abd Manan, N.and Othman, F. 2015. Inhibitory and Apoptosis-Inducing Effects of Newcastle Disease Virus Strain AF2240 on Mammary Carcinoma Cell Line. *BioMed Research International* 2015, 1-12.
- Ahmed, I. 2014. Induction of Nitric Oxide and TNF-A in Newcastle Disease Virus (NDV) AF2240 Infected RAW 264.7 Macrophages and their Cytotoxic Activity on MDA-MB-231 Breast Cancer Cell Line. *Journal of Cancer Science & Therapy* 06.
- Alabsi, A.M., Ali, R., Ideris, A., Omar, A.R., Bejo, M.H., Yusoff, K.and Ali, A.M. 2012. Anti-leukemic activity of Newcastle disease virus strains AF2240 and V4-UPM in murine myelomonocytic leukemia in vivo. *Leukemia Research* 36, 634-645.
- Alabsi, A.M., Bakar, S.A.A., Ali, R., Omar, A.R., Bejo, M.H., Ideris, A.and Ali, A.M. 2011. Effects of Newcastle Disease Virus Strains AF2240 and V4-UPM on Cytolysis and Apoptosis of Leukemia Cell Lines. *International Journal of Molecular Sciences* 12, 8645-8660.
- Alexander, J. L., Scott, A. J., Pouncey, A. L., Marchesi, J., Kinross, J., and Teare, J. 2018. Colorectal carcinogenesis: an archetype of gut microbiota-host interaction. *Epidemiol Health*, 12, 865.
- Ali, R., Alabsi, A.M., Ali, A.M., Ideris, A., Omar, A.R., Yusoff, K.and Saif-Ali, R. 2011. Cytolytic Effects and Apoptosis Induction of Newcastle Disease Virus Strain AF2240 on Anaplastic Astrocytoma Brain Tumor Cell Line. *Neurochemical Research* 36, 2051-2062.
- Amaravadi, R.K., Thompson, C.B., 2007. The roles of therapy-induced autophagy and necrosis in cancer treatment. *Clinical cancer research : an official journal of the American Association for Cancer Research* 13, 7271-7279.

- An, S., Nam, K., Choi, S., Bai, C.Z., Lee, Y. and Park, J.S. 2013. Nonviral gene therapy in vivo with PAM-RG4/apoptin as a potential brain tumor therapeutic. *International journal of nanomedicine* 8, 821-834.
- Aref, S., Bailey, K. and Fielding, A. 2016. Measles to the Rescue: A Review of Oncolytic Measles Virus. *Viruses* 8.
- Assayaghi, R.M., Alabsi, A.M. and Ali, A.M. 2016. Apoptosis Induction of Newcastle Disease Virus Strains (AF 2240 & V4-UPM) on HT-29 Human Colorectal Adenocarcinoma Cells. *JCRTO* 4, 1-5.
- Austin, D., Baer, A., Lundberg, L., Shafagati, N., Schoonmaker, A., Narayanan, A., Popova, T., Panthier, J.J., Kashanchi, F., Bailey, C. and Kehn-Hall, K. 2012. p53 Activation following Rift Valley fever virus infection contributes to cell death and viral production. *PloS one* 7, e36327.
- Ayllon, J., Garcia-Sastre, A. and Martinez-Sobrido, L. 2013. Rescue of recombinant Newcastle disease virus from cDNA. *Journal of visualized experiments : JoVE*.
- Azizah, A.M., Nor Saleha, I.T., Noor Hashimah, A., Asmah, Z.A. and Mastulu, W. 2016. Malaysian National Cancer Registry Report 2007-2011 National Cancer Institute, Ministry of Health Malaysia.
- Bakar, S.A.A., T-Johari, S.A.T., Mohamad, N.M., Hamid, M.H.A., Yusoff, M.A.M. and Ali, A.M. 2016. Antiproliferative and Apoptotic Effect of Newcastle Disease Virus (NDV) Strain AF2240 in Human Promyelocytic Leukemia Cells (HL60). *International Journal of Cancer Research* 13, 9-16.
- Bakar, S.A.A., Zawawi, M., Ali, A.M. and Ideris, A. 2012. Induction of apoptosis by Newcastle Disease virus strains AF220 and V4-UPM in human promyelocytic leukemia (HL60) and human t lymphoblastic leukemia (CEM-SS) Cells. *International Journal of Bioengineering and Life Sciences* 6, 135-139.
- Bell, C.W., Jiang, W., Reich, C.F., 3rd, Pisetsky, D.S., 2006. The extracellular release of HMGB1 during apoptotic cell death. American journal of physiology. *Cell physiology* 291, C1318-1325.
- Benedict, C.A., Norris, P.S., Prigozy, T.I., Bodmer, J.L., Mahr, J.A., Garnett, C.T., Martinon, F., Tschoopp, J., Gooding, L.R. and Ware, C.F. 2001. Three adenovirus E3 proteins cooperate to evade apoptosis by tumor necrosis factor-related apoptosis-inducing ligand receptor-1 and -2. *The Journal of biological chemistry* 276, 3270-3278.
- Berg, K.C.G., Eide, P.W., Eilertsen, I.A., Johannessen, B., Bruun, J. Danielsen, S.A., Bjørnslett, M., Meza-Zepeda L.A., Eknæs, M., Lind, G.E., Myklebost,

- O., Skotheim, R.I., Sveen, A. and Lothe, R.A. 2017. Multi-omics of 34 colorectal cancer cell lines - a resource for biomedical studies. *Molecular Cancer* 16, 116.
- Bierman, H.R., Crile, D.M., Dod, K.S., Kelly, K.H., Petrakis, N.L., White, L.P. and Shimkin, M.B. 1953. Remissions in leukemia of childhood following acute infectious disease: staphylococcus and streptococcus, varicella, and feline panleukopenia. *Cancer* 6, 591-605.
- Bluming, A.Z. and Ziegler, J.L. 1971. Regression of Burkitt's lymphoma in association with measles infection. *Lancet (London, England)* 2, 105-106.
- Boame, N., Gresham, G., Jonker, D., Martel, G., Balaa, F., and Asmis, T. 2014. Use of chemotherapy and radiofrequency ablation to treat colorectal cancer metastases: a retrospective review of The Ottawa Hospital Cancer Centre over 7 years. *Current Oncology* 21(4), e557-563.
- Bode, J.G., Brenndörfer, E.D. and Häussinger, D. 2008. Hepatitis C virus (HCV) employs multiple strategies to subvert the host innate antiviral response. *Biological Chemistry* 389.
- Bohle, W., Schlag, P., Liebrich, W., Hohenberger, P., Manasterski, M., Möller, P. and Schirrmacher, V. 1990. Postoperative active specific immunization in colorectal cancer patients with virus-modified autologous tumor-cell vaccine. First clinical results with tumor-cell vaccines modified with live but avirulent newcastle disease virus. *Cancer* 66, 1517-1523.
- Boleij, A., and Tjalsma, H. 2013. The itinerary of *Streptococcus gallolyticus* infection in patients with colonic malignant disease. *Lancet Infectious Disease*, 13(8), 719-724.
- Bordeianou, L., Maguire, L. H., Alavi, K., Sudan, R., Wise, P. E., and Kaiser, A. M. 2014. Sphincter-sparing surgery in patients with low-lying rectal cancer: techniques, oncologic outcomes, and functional results. *Journal of Gastrointestinal Surgery*, 18(7), 1358-1372.
- Bouchet, B.P., Caron de Fromentel, C., Puisieux, A. and Galmarini, C.M. 2006. p53 as a target for anti-cancer drug development. *Critical reviews in oncology/hematology* 58, 190-207.
- Botteri, E., Iodice, S., Bagnardi, V., Raimondi, S., Lowenfels, A. B., and Maisonneuve, P. 2008. Smoking and colorectal cancer: a meta-analysis. *JAMA*, 300(23), 2765-2778.
- Bretagnol, F., Slim, K., and Faucheron, J. L. 2005. Anterior resection with low colorectal anastomosis. To drain or not?. *Ann Chir*, 130(5), 336-339.

- Bridgen, A. 2012. Reverse genetics of RNA viruses : applications and perspectives. Wiley - Blackwell, Chichester, West Sussex, UK; Hoboken, NJ.
- Brunet, A., Bonni, A., Zigmond, M.J., Lin, M.Z., Juo, P., Hu, L.S., Anderson, M.J., Arden, K.C., Blenis, J. and Greenberg, M.E. 1999. Akt promotes cell survival by phosphorylating and inhibiting a Forkhead transcription factor. *Cell* 96, 857-868.
- Buchholz, U.J., Finke, S. and Conzelmann, K.K. 1999. Generation of bovine respiratory syncytial virus (BRSV) from cDNA: BRSV NS2 is not essential for virus replication in tissue culture, and the human RSV leader region acts as a functional BRSV genome promoter. *J Virol* 73, 251-259.
- Bullenkamp, J., Cole, D., Malik, F., Alkhatabi, H., Kulasekararaj, A., Odell, E.W., Farzaneh, F., Gaken, J. and Tavassoli, M. 2012. Human Gyrovirus Apoptin shows a similar subcellular distribution pattern and apoptosis induction as the chicken anaemia virus derived VP3/Apoptin. *Cell death & disease* 3, e296.
- Burek, M., Maddika, S., Burek, C.J., Daniel, P.T., Schulze-Osthoff, K. and Los, M. 2006. Apoptin-induced cell death is modulated by Bcl-2 family members and is Apaf-1 dependent. *Oncogene* 25, 2213-2222.
- Cai, Z., Liu, Z.G., 2018. Detection of MLKL Oligomerization During Programmed Necrosis. *Methods in molecular biology* (Clifton, N.J.) 1857, 85-92.
- Cao, H.D., Yang, Y.X., Lu, L., Liu, S.N., Wang, P.L., Tao, X.H., Wang, L.J. and Xiang, T.X. 2010. Attenuated Salmonella typhimurium carrying TRAIL and VP3 genes inhibits the growth of gastric cancer cells in vitro and in vivo. *Tumori* 96, 296-303.
- Cao, X., Deng, X., May, W.S., 2003. Cleavage of Bax to p18 Bax accelerates stress-induced apoptosis, and a cathepsin-like protease may rapidly degrade p18 Bax. *Blood* 102, 2605-2614.
- Carruthers, N.J., Parker, G.C., Gratsch, T., Caruso, J.A. and Stemmer, P.M. 2015. Protein Mobility Shifts Contribute to Gel Electrophoresis Liquid Chromatography Analysis. *Journal of biomolecular techniques : JBT* 26, 103-112.
- Cassel, W.A., Murray, D.R., Torbin, A.H., Olkowski, Z.L. and Moore, M.E. 1977. Viral oncolysate in the management of malignant melanoma.I. Preparation of the oncolysate and measurement of immunologic responses. *Cancer* 40, 672-679.
- Cassel, W.A., Murray, D.R. and Phillips, H.S. 1983. A phase II study on the postsurgical management of stage II malignant melanoma with a Newcastle disease virus oncolysate. *Cancer* 52, 856-860.

- Cassel, W.A.and Garrett, R.E. 1965. Newcastle disease virus as an antineoplastic agent. *Cancer* 18, 863-868.
- Castedo, M. 2002. Sequential involvement of Cdk1, mTOR and p53 in apoptosis induced by the HIV-1 envelope. *The EMBO Journal* 21, 4070-4080.
- Ch'ng, W.-C., Abd-Aziz, N., Ong, M.-H., Stanbridge, E.J.and Shafee, N. 2015. Human renal carcinoma cells respond to Newcastle disease virus infection through activation of the p38 MAPK/NF-κB/IκB α pathway. *Cellular Oncology* 38, 279-288.
- Chaabane, W., Cieslar-Pobuda, A., El-Gazzah, M., Jain, M.V., Rzeszowska-Wolny, J., Rafat, M., Stetefeld, J., Ghavami, S.and Los, M.J. 2014. Human-gyrovirus-Apopin triggers mitochondrial death pathway--Nur77 is required for apoptosis triggering. *Neoplasia (New York, N.Y.)* 16, 679-693.
- Chaabane, W., Ghavami, S., Małecki, A. and Łos, M.J. 2017. Human Gyrovirus-Apopin Interferes with the Cell Cycle and Induces G2/M Arrest Prior to Apoptosis. *Arch Immunol Ther Exp (Warsz)*;65(6):545-552
- Chaitanya, G.V., Steven, A.J., Babu, P.P., 2010. PARP-1 cleavage fragments: signatures of cell-death proteases in neurodegeneration. *Cell communication and signaling : CCS* 8, 31.
- Chan, F.K., Shisler, J., Bixby, J.G., Felices, M., Zheng, L., Appel, M., Orenstein, J., Moss, B., Lenardo, M.J., 2003. A role for tumor necrosis factor receptor-2 and receptor-interacting protein in programmed necrosis and antiviral responses. *The Journal of biological chemistry* 278, 51613-51621.
- Chand, M., Bhoday, J., Brown, G., Moran, B., and Parvaiz, A. 2012. Laparoscopic surgery for rectal cancer. *Journal of the Royal Society of Medicine*, 105(10), 429-435.
- Chen, L., Eloranta, S., Martling, A., Glimelius, I., Neovius, M., Glimelius, B., and Smedby, K. E. 2018. Short- and long-term risks of cardiovascular disease following radiotherapy in rectal cancer in four randomized controlled trials and a population-based register. *Radiotherapy Oncology*, 126(3), 424-430.
- Chen, G.X., Zhang, S., He, X.H., Liu, S.Y., Ma, C.and Zou, X.P. 2014. Clinical utility of recombinant adenoviral human p53 gene therapy: current perspectives. *Oncotargets and therapy* 7, 1901-1909.
- Chen, K., Luo, Z., Tang, J.and Zheng, S.J. 2011. A critical role of heat shock cognate protein 70 in Apoptin-induced phosphorylation of Akt. *Biochemical and biophysical research communications* 409, 200-204.
- Cherubini, G., Petouchoff, T., Grossi, M., Piersanti, S., Cundari, E., Saggio, I., 2006. E1B55K-deleted adenovirus (ONYX-015) overrides G1/S and G2/M

- checkpoints and causes mitotic catastrophe and endoreduplication in p53-proficient normal cells. *Cell cycle* (Georgetown, Tex.) 5, 2244-2252.
- Chia, S.L., Tan, W.S., Yusoff, K.and Shafee, N. 2012. Plaque formation by a velogenic Newcastle disease virus in human colorectal cancer cell lines. *Acta virologica* 56, 345-347.
- Chia, S.-L., Yusoff, K.and Shafee, N. 2014. Viral persistence in colorectal cancer cells infected by Newcastle disease virus. *Virology Journal* 11, 91.
- Chiche, J., Brahimi-Horn, M.C.and Pouysségur, J. 2010. Tumour hypoxia induces a metabolic shift causing acidosis: a common feature in cancer. *Journal of Cellular and Molecular Medicine* 14, 771-794.
- Ch'ng, W.-C., Stanbridge, E.J., Yusoff, K.and Shafee, N. 2013. The Oncolytic Activity of Newcastle Disease Virus in Clear Cell Renal Carcinoma Cells in Normoxic and Hypoxic Conditions: The Interplay Between von Hippel-Lindau and Interferon- β Signaling. *Journal of Interferon & Cytokine Research* 33, 346-354.
- Christofferson, D.E., Yuan, J., 2010. Necroptosis as an alternative form of programmed cell death. *Current opinion in cell biology* 22, 263-268.
- Chulan, U., Ibrahim, A.L., Mustaffa Babjee, A.and Sheikh-Omar, A.R. 1982. Vaccination against newcastle disease. *Tropical Animal Health and Production* 14, 177-184.
- Corthay, A., Lundin, K.U., Lorvik, K.B., Hofgaard, P.O.and Bogen, B. 2009. Secretion of Tumor-Specific Antigen by Myeloma Cells Is Required for Cancer Immunosurveillance by CD4+ T Cells. *Cancer Research* 69, 5901-5907.
- Corthay, A., Skovseth, D.K., Lundin, K.U., Røsjø, E., Omholt, H., Hofgaard, P.O., Haraldsen, G.and Bogen, B. 2005. Primary Antitumor Immune Response Mediated by CD4+ T Cells. *Immunity* 22, 371-383.
- Cunningham, D., Atkin, W., Lenz, H.J., Lynch, H.T., Minsky, B., Nordlinger, B.and Starling, N. 2010. Colorectal cancer. *Lancet (London, England)* 375, 1030-1047.
- Danenberg, P. V., and Danenberg, K. D. 1978. Effect of 5-, 10-methylenetetrahydrofolate on the dissociation of 5-fluoro-2'-deoxyuridylate from thymidylate synthetase: evidence for an ordered mechanism. *Biochemistry*, 17(19), 4018-4024.
- Danen-Van Oorschot, A., Voskamp, Seelen, M.C., van Miltenburg, M.A.H.M., Bolk, M.W., Tait, S.W., Boesen-de Cock, J.G.R., Rohn, J.L., Borst, J.and Noteborn, M. 2004. Human death effector domain-associated factor

- interacts with the viral apoptosis agonist Apoptin and exerts tumor-preferential cell killing. *Cell Death and Differentiation* 11, 10.
- Danen-Van Oorschot, A.A., Fischer, D.F., Grimbergen, J.M., Klein, B., Zhuang, S., Falkenburg, J.H., Backendorf, C., Quax, P.H., Van der Eb, A.J. and Noteborn, M.H. 1997. Apoptin induces apoptosis in human transformed and malignant cells but not in normal cells. *Proceedings of the National Academy of Sciences of the United States of America* 94, 5843-5847.
- Danen-Van Oorschot, A.A., Zhang, Y., Erkeland, S.J., Fischer, D.F., van der Eb, A.J. and Noteborn, M.H. 1999. The effect of Bcl-2 on Apoptin in 'normal' vs transformed human cells. *Leukemia* 13 Suppl 1, S75-77.
- Danen-Van Oorschot, A.A., Zhang, Y.H., Leliveld, S.R., Rohn, J.L., Seelen, M.C., Bolk, M.W., Van Zon, A., Erkeland, S.J., Abrahams, J.P., Mumberg, D. and Noteborn, M.H. 2003. Importance of nuclear localization of apoptin for tumor-specific induction of apoptosis. *The Journal of biological chemistry* 278, 27729-27736.
- Denisenko, T.V., Sorokina, I.V., Gogvadze, V., Zhivotovsky, B., 2016. Mitotic catastrophe and cancer drug resistance: A link that must to be broken. *Drug resistance updates : reviews and commentaries in antimicrobial and anticancer chemotherapy* 24, 1-12.
- Devireddy, L.R. and Jones, C.J. 1999. Activation of caspases and p53 by bovine herpesvirus 1 infection results in programmed cell death and efficient virus release. *J Virol* 73, 3778-3788.
- Dewson, G. and Kluck, R.M. 2009. Mechanisms by which Bak and Bax permeabilise mitochondria during apoptosis. *Journal of cell science* 122, 2801-2808.
- Diamond, S. J., Enestvedt, B. K., Jiang, Z., Holub, J. L., Gupta, M., Lieberman, D. A., and Eisen, G. M. 2011. Adenoma detection rate increases with each decade of life after 50 years of age. *Gastrointestinal Endoscopy*, 74(1), 135-140.
- Dillon, C.P., Oberst, A., Weinlich, R., Janke, L.J., Kang, T.B., Ben-Moshe, T., Mak, T.W., Wallach, D., Green, D.R., 2012. Survival function of the FADD-CASPASE-8-cFLIP(L) complex. *Cell reports* 1, 401-407.
- Dock, G. 1904. The influence of complicating diseases upon leukemia. *The American Journal of the Medical Sciences* 27, 20.
- Doi, T., Kwon, H.-J., Honda, T., Sato, H., Yoneda, M. and Kai, C. 2016. Measles virus induces persistent infection by autoregulation of viral replication. *Scientific Reports* 6.

- Dortmans, J.C., Rottier, P.J., Koch, G. and Peeters, B.P. 2010. The viral replication complex is associated with the virulence of Newcastle disease virus. *J Virol* 84, 10113-10120.
- Douglas, A.J., Phenix, K., Mawhinney, K.A., Todd, D., Mackie, D.P. and Curran, W.L. 1995. Identification of a 24 kDa protein expressed by chicken anaemia virus. *The Journal of general virology* 76 (Pt 7), 1557-1562.
- Drexler, I., Heller, K., Wahren, B., Erfle, V. and Sutter, G. 1998. Highly attenuated modified vaccinia virus Ankara replicates in baby hamster kidney cells, a potential host for virus propagation, but not in various human transformed and primary cells. *The Journal of general virology* 79 (Pt 2), 347-352.
- Eguchi, Y., Shimizu, S., Tsujimoto, Y., 1997. Intracellular ATP levels determine cell death fate by apoptosis or necrosis. *Cancer Res* 57, 1835-1840.
- Elankumaran, S., Chavan, V., Qiao, D., Shobana, R., Moorkanat, G., Biswas, M. and Samal, S.K. 2010. Type I Interferon-Sensitive Recombinant Newcastle Disease Virus for Oncolytic Virotherapy. *Journal of Virology* 84, 3835-3844.
- Elankumaran, S., Rockemann, D. and Samal, S.K. 2006. Newcastle Disease Virus Exerts Oncolysis by both Intrinsic and Extrinsic Caspase-Dependent Pathways of Cell Death. *Journal of Virology* 80, 7522-7534.
- Elroy-Stein, O., Fuerst, T.R. and Moss, B. 1989. Cap-independent translation of mRNA conferred by encephalomyocarditis virus 5' sequence improves the performance of the vaccinia virus/bacteriophage T7 hybrid expression system. *Proceedings of the National Academy of Sciences of the United States of America* 86, 6126-6130.
- Endo, Y., Sakai, R., Ouchi, M., Onimatsu, H., Hioki, M., Kagawa, S., Uno, F., Watanabe, Y., Urata, Y., Tanaka, N. and Fujiwara, T. 2008. Virus-mediated oncolysis induces danger signal and stimulates cytotoxic T-lymphocyte activity via proteasome activator upregulation. *Oncogene* 27, 2375-2381.
- Erdman, S. E., Rao, V. P., Poutahidis, T., Rogers, A. B., Taylor, C. L., Jackson, E. A., Ge, Z., Lee, C.W., Schauer, D.B., Wogan, G.N., Tannenbaum, S.R. and Fox, J. G. 2009. Nitric oxide and TNF-alpha trigger colonic inflammation and carcinogenesis in *Helicobacter hepaticus*-infected, Rag2-deficient mice. *Proceedings of the National Academy of Sciences of the United States of America*, 106(4), 1027-1032.
- Eshghi, M. J., Fatemi, R., Hashemy, A., Aldulaimi, D., and Khodadoostan, M. 2011. A retrospective study of patients with colorectal polyps. *Gastroenterol Hepatol Bed Bench*, 4(1), 17-22

- Evgin, L., Acuna, S.A., Tanese de Souza, C., Marguerie, M., Lemay, C.G., Ilkow, C.S., Findlay, C.S., Falls, T., Parato, K.A., Hanwell, D., Goldstein, A., Lopez, R., Lafrance, S., Breitbach, C.J., Kirn, D., Atkins, H., Auer, R.C., Thurman, J.M., Stahl, G.L., Lambris, J.D., Bell, J.C. and McCart, J.A. 2015. Complement inhibition prevents oncolytic vaccinia virus neutralization in immune humans and cynomolgus macaques. *Molecular therapy : the journal of the American Society of Gene Therapy* 23, 1066-1076.
- Felt, S.A., Moerdyk-Schauwecker, M.J. and Grdzelishvili, V.Z. 2015. Induction of apoptosis in pancreatic cancer cells by vesicular stomatitis virus. *Virology* 474, 163-173.
- Fountzilas, C., Patel, S. and Mahalingam, D. 2017. Review: Oncolytic virotherapy, updates and future directions. *Oncotarget* 8, 102617-102639.
- Fraser, D., Maher, H.R., Shug, A.L. and Thomas, C.A. 1957. The Infection Of Sub-Cellular *Escherichia coli*, Strain B, With A Dna Preparation From T2 Bacteriophage. *Proceedings of the National Academy of Sciences of the United States of America* 43, 939-947.
- Fu, Z., Shrubsole, M. J., Smalley, W. E., Wu, H., Chen, Z., Shyr, Y., Ness, R.M. and Zheng, W. 2012. Lifestyle factors and their combined impact on the risk of colorectal polyps. *American Journal of Epidemiology*, 176(9), 766-776.
- Fulda, S., 2013. Regulation of cell death in cancer—possible implications for immunotherapy. *Frontiers in oncology* 3, 29.
- Furukawa, Y., Takasu, A., Yura, Y., 2017. Role of autophagy in oncolytic herpes simplex virus type 1-induced cell death in squamous cell carcinoma cells. *Cancer gene therapy* 24, 393-400.
- Gadaleta, P., Perfetti, X., Mersich, S. and Coulombe, F. 2005. Early activation of the mitochondrial apoptotic pathway in Vesicular Stomatitis virus-infected cells. *Virus Res* 109, 65-69.
- Garber, K. 2006. China approves world's first oncolytic virus therapy for cancer treatment. *Journal of the National Cancer Institute* 98, 298-300.
- Garten, W., Berk, W., Nagai, Y., Rott, R. and Klenk, H.D. 1980. Mutational changes of the protease susceptibility of glycoprotein F of Newcastle disease virus: effects on pathogenicity. *The Journal of general virology* 50, 135-147.
- Gerl, R. and Vaux, D.L. 2005. Apoptosis in the development and treatment of cancer. *Carcinogenesis* 26, 263-270.
- Gerlier, D. and Lyles, D.S. 2011. Interplay between innate immunity and negative-strand RNA viruses: towards a rational model. *Microbiology and molecular biology reviews : MMBR* 75, 468-490, second page of table of contents.

- Gey, G.C. 1952. Tissue culture studies of the proliferative capacity of cervical carcinoma and normal epithelium. *Cancer Res* 12, 264-265.
- Ghrici, M., El Zowalaty, M., Omar, A.R.and Ideris, A. 2013a. Induction of apoptosis in MCF-7 cells by the hemagglutinin-neuraminidase glycoprotein of Newcastle disease virus Malaysian strain AF2240. *Oncology Reports* 30, 1035-1044.
- Ghrici, M., El Zowalaty, M., Omar, A.R.and Ideris, A. 2013b. Newcastle disease virus Malaysian strain AF2240 induces apoptosis in MCF-7 human breast carcinoma cells at an early stage of the virus life cycle. *International Journal of Molecular Medicine* 31, 525-532.
- Gill, S., Thomas, R.R.and Goldberg, R.M. 2003. Review article: colorectal cancer chemotherapy. *Alimentary pharmacology & therapeutics* 18, 683-692.
- Goping, I.S., Gross, A., Lavoie, J.N., Nguyen, M., Jemmerson, R., Roth, K., Korsmeyer, S.J., Shore, G.C., 1998. Regulated targeting of BAX to mitochondria. *The Journal of cell biology* 143, 207-215.
- Goto, M., Okazaki, M.and Yazaki, H. 1959. Oncolytic Effect Of Newcastle Disease Virus On Yoshida Sarcoma (I). *Japanese Journal of Microbiology* 3, 171-181.
- Greiner, S., Humrich, J.Y., Thuman, P., Sauter, B., Schuler, G.and Jenne, L. 2006. The highly attenuated vaccinia virus strain modified virus Ankara induces apoptosis in melanoma cells and allows bystander dendritic cells to generate a potent anti-tumoral immunity. *Clinical and experimental immunology* 146, 344-353.
- Grivennikov, S.I., Greten, F.R.and Karin, M. 2010. Immunity, Inflammation, and Cancer. *Cell* 140, 883-899.
- Guan, G.F., Zhao, M., Liu, L.M., Jin, C.S., Sun, K., Zhang, D.J., Yu, D.J., Cao, H.W., Lu, Y.Q.and Wen, L.J. 2013. *Salmonella typhimurium* mediated delivery of Apoptin in human laryngeal cancer. *Int J Med Sci* 10, 1639-1648.
- Haabeth, O.A.W., Bogen, B.and Corthay, A. 2012. A model for cancer-suppressive inflammation. *OncoImmunology* 1, 1146-1155.
- Haabeth, O.A.W., Lorvik, K.B., Hammarström, C., Donaldson, I.M., Haraldsen, G., Bogen, B.and Corthay, A. 2011. Inflammation driven by tumour-specific Th1 cells protects against B-cell cancer. *Nature Communications* 2, 240.
- Habjan, M., Penski, N., Spiegel, M.and Weber, F. 2008. T7 RNA polymerase-dependent and -independent systems for cDNA-based rescue of Rift Valley fever virus. *The Journal of general virology* 89, 2157-2166.

- Hafner, M.F.and Debus, J. 2016. Radiotherapy for Colorectal Cancer: Current Standards and Future Perspectives. *Visceral medicine* 32, 172-177.
- Harty, R.N., Brown, M.E., Hayes, F.P., Wright, N.T.and Schnell, M.J. 2001. Vaccinia virus-free recovery of vesicular stomatitis virus. *Journal of molecular microbiology and biotechnology* 3, 513-517.
- He, S., Wang, L., Miao, L., Wang, T., Du, F., Zhao, L., Wang, X., 2009. Receptor interacting protein kinase-3 determines cellular necrotic response to TNF-alpha. *Cell* 137, 1100-1111.
- Heaton, N.S.and Randall, G. 2010. Dengue Virus-Induced Autophagy Regulates Lipid Metabolism. *Cell Host & Microbe* 8, 422-432.
- Heckl, S., Regenbogen, M., Sturzu, A., Gharabaghi, A., Feil, G., Beck, A., Echner, H.and Nagele, T. 2008. Value of apoptin's 40-amino-acid C-terminal fragment for the differentiation between human tumor and non-tumor cells. *Apoptosis : an international journal on programmed cell death* 13, 495-508.
- Heilman, D.W., Teodoro, J.G.and Green, M.R. 2006. Apoptin nucleocytoplasmic shuttling is required for cell type-specific localization, apoptosis, and recruitment of the anaphase-promoting complex/cycosome to PML bodies. *J Virol* 80, 7535-7545.
- Henson, P.M., Hume, D.A., 2006. Apoptotic cell removal in development and tissue homeostasis. *Trends Immunol* 27, 244-250.
- Holler, N., Zaru, R., Micheau, O., Thome, M., Attinger, A., Valitutti, S., Bodmer, J.L., Schneider, P., Seed, B., Tschoopp, J., 2000. Fas triggers an alternative, caspase-8-independent cell death pathway using the kinase RIP as effector molecule. *Nat Immunol* 1, 489-495.
- Hough, K.P., Rogers, A.M., Zelic, M., Paris, M.and Heilman, D.W. 2015. Transformed cell-specific induction of apoptosis by porcine circovirus type 1 viral protein 3. *The Journal of general virology* 96, 351-359.
- Housman, G., Byler, S., Heerboth, S., Lapinska, K., Longacre, M., Snyder, N., and Sarkar, S. 2014. Drug resistance in cancer: an overview. *Cancers (Basel)*, 6(3), 1769-1792.
- Housseau, F., and Sears, C. L. 2010. Enterotoxigenic Bacteroides fragilis (ETBF)-mediated colitis in Min (Apc+/-) mice: a human commensal-based murine model of colon carcinogenesis. *Cell Cycle*, 9(1), 3-5.
- Howells, A., Marelli, G., Lemoine, N.R., Wang, Y., 2017. Oncolytic Viruses- Interaction of Virus and Tumor Cells in the Battle to Eliminate Cancer. *Frontiers in oncology* 7, 195.

- Hsieh, J. S., Lin, S. R., Chang, M. Y., Chen, F. M., Lu, C. Y., Huang, T. J., Huang, Y.S., Huang, C.J. and Wang, J. Y. 2005. APC, K-ras, and p53 gene mutations in colorectal cancer patients: correlation to clinicopathologic features and postoperative surveillance. *American Surgeon*, 71(4), 336-343.
- Huang, Z., Krishnamurthy, S., Panda, A. and Samal, S.K. 2003. Newcastle disease virus V protein is associated with viral pathogenesis and functions as an alpha interferon antagonist. *J Virol* 77, 8676-8685.
- Huo, D.H., Yi, L.N. and Yang, J. 2008. Interaction with Ppil3 leads to the cytoplasmic localization of Apoptin in tumor cells. *Biochemical and biophysical research communications* 372, 14-18.
- Hwang, I.I.L., Watson, I.R., Der, S.D. and Ohh, M. 2006. Loss of VHL Confers Hypoxia-Inducible Factor (HIF)-Dependent Resistance to Vesicular Stomatitis Virus: Role of HIF in Antiviral Response. *Journal of Virology* 80, 10712-10723.
- Ibrahim, A.L., Chulan, U. and Mustaffa Babjee, A. 1980. Preliminary evaluation on a freeze dried Mukteswar Newcastle disease virus vaccine. *Kajian Veterinar* 57.
- Igney, F.H. and Krammer, P.H. 2002. Death and Anti-Death: Tumour Resistance To Apoptosis. *Nature Reviews Cancer* 2, 277-288.
- Irrinki, K.M., Mallilankaraman, K., Thapa, R.J., Chandramoorthy, H.C., Smith, F.J., Jog, N.R., Gandhirajan, R.K., Kelsen, S.G., Houser, S.R., May, M.J., Balachandran, S., Madesh, M., 2011. Requirement of FADD, NEMO, and BAX/BAK for aberrant mitochondrial function in tumor necrosis factor alpha-induced necrosis. *Molecular and cellular biology* 31, 3745-3758.
- Jamal, M.-H., Ch'ng, W.-C., Yusoff, K. and Shafee, N. 2012. Reduced Newcastle disease virus-induced oncolysis in a subpopulation of cisplatin-resistant MCF7 cells is associated with survivin stabilization. *Cancer Cell International* 12, 35.
- Janssen, K., Hofmann, T.G., Jans, D.A., Hay, R.T., Schulze-Osthoff, K. and Fischer, U. 2007. Apoptin is modified by SUMO conjugation and targeted to promyelocytic leukemia protein nuclear bodies. *Oncogene* 26, 1557-1566.
- Jeuriissen, S.H., Wagenaar, F., Pol, J.M., van der Eb, A.J. and Noteborn, M.H. 1992. Chicken anemia virus causes apoptosis of thymocytes after in vivo infection and of cell lines after in vitro infection. *J Virol* 66, 7383-7388.
- Jiang, J., Cole, D., Westwood, N., Macpherson, L., Farzaneh, F., Mufti, G., Tavassoli, M. and Gaken, J. 2010. Crucial roles for protein kinase C isoforms in tumor-specific killing by apoptin. *Cancer Res* 70, 7242-7252.

- Jin, J.L., Gong, J., Yin, T.J., Lu, Y.J., Xia, J.J., Xie, Y.Y., Di, Y., He, L., Guo, J.L., Sun, J., Noteborn, M.H. and Qu, S. 2011. PTD4-apoptin protein and dacarbazine show a synergistic antitumor effect on B16-F1 melanoma in vitro and in vivo. *European journal of pharmacology* 654, 17-25.
- Johnson, C.F. and Scott, A.D. 1964. Cytological Studies of Newcastle Disease Virus (NDV) in HEp-2 Cells. *Experimental Biology and Medicine* 115, 281-286.
- Jouan-Lanhouet, S., Arshad, M.I., Piquet-Pellorce, C., Martin-Chouly, C., Le Moigne-Muller, G., Van Herreweghe, F., Takahashi, N., Sergent, O., Lagadic-Gossmann, D., Vandenabeele, P., Samson, M., Dimanche-Boitrel, M.T., 2012. TRAIL induces necroptosis involving RIPK1/RIPK3-dependent PARP-1 activation. *Cell Death Differ* 19, 2003-2014.
- Kalid, M., Jahanshiri, F., Omar, A.R. and Yusoff, K. 2010. Gene expression profiling in apoptotic MCF-7 cells infected with Newcastle disease virus. *Global Veterinaria* 5, 334-340.
- Kelly, E. and Russell, S.J. 2007. History of oncolytic viruses: genesis to genetic engineering. *Molecular therapy : the journal of the American Society of Gene Therapy* 15, 651-659.
- Kim, B.M., 2017. The Implications of Several Forms of Programmed Necrosis for Cancer Therapy. *Journal of Cancer Science & Therapy* 9, 6.
- Kim, J.S., He, L., Lemasters, J.J., 2003. Mitochondrial permeability transition: a common pathway to necrosis and apoptosis. *Biochemical and biophysical research communications* 304, 463-470.
- Kim, S.H. and Samal, S.K. 2016. Newcastle Disease Virus as a Vaccine Vector for Development of Human and Veterinary Vaccines. *Viruses* 8.
- Kingsbury, D.W. 1991. The Paramyxoviruses. Springer US, Boston, MA.
- Klanrit, P., Flinterman, M.B., Odell, E.W., Melino, G., Killick, R., Norris, J.S. and Tavassoli, M. 2008. Specific isoforms of p73 control the induction of cell death induced by the viral proteins, E1A or apoptin. *Cell cycle (Georgetown, Tex.)* 7, 205-215.
- Koch, G., van Roozelaar, D.J., Verschueren, C.A., van der Eb, A.J. and Noteborn, M.H. 1995. Immunogenic and protective properties of chicken anaemia virus proteins expressed by baculovirus. *Vaccine* 13, 763-770.
- Kolakofsky, D., Roux, L., Garcin, D. and Ruigrok, R.W. 2005. Paramyxovirus mRNA editing, the "rule of six" and error catastrophe: a hypothesis. *The Journal of general virology* 86, 1869-1877.

- Kono, H., Rock, K.L., 2008. How dying cells alert the immune system to danger. *Nature reviews. Immunology* 8, 279-289.
- Koo, J. H., Leong, R. W., Ching, J., Yeoh, K. G., Wu, D. C., Murdani, A., Chiu, H.M., Chong, V.H., Rerknimitr, R., Goh, K.L., Hilmi, I., Byeon, J.S., Niaz, S.K., Siddique, A., Wu, K.C., Matsuda, T., Makharia, G., Sollano, J., Lee, S.K. and Sung, J. J. 2012. Knowledge of, attitudes toward, and barriers to participation of colorectal cancer screening tests in the Asia-Pacific region: a multicenter study. *Gastrointestinal Endoscopy*, 76(1), 126-135.
- Kooistra, K., Zhang, Y.H., Henriquez, N.V., Weiss, B., Mumberg, D. and Noteborn, M.H. 2004. TT virus-derived apoptosis-inducing protein induces apoptosis preferentially in hepatocellular carcinoma-derived cells. *The Journal of general virology* 85, 1445-1450.
- Kostic, A. D., Chun, E., Robertson, L., Glickman, J. N., Gallini, C. A., Michaud, M., Clancy, T.E., Chung, D.C., Lochhead, P., Hold, G.L., El-Omar, E.M., Brenner, D., Fuchs, C.S., Meyerson, M. and Garrett, W. S. 2013. *Fusobacterium nucleatum* potentiates intestinal tumorigenesis and modulates the tumor-immune microenvironment. *Cell Host Microbe*, 14(2), 207-215.
- Krishnamurthy, S., Takimoto, T., Scroggs, R.A. and Portner, A. 2006. Differentially Regulated Interferon Response Determines the Outcome of Newcastle Disease Virus Infection in Normal and Tumor Cell Lines. *Journal of Virology* 80, 5145-5155.
- Kucharski, T.J., Gamache, I., Gjoerup, O. and Teodoro, J.G. 2011. DNA damage response signaling triggers nuclear localization of the chicken anemia virus protein Apoptin. *J Virol* 85, 12638-12649.
- Kultz, D. 2005. Molecular and evolutionary basis of the cellular stress response. *Annual review of physiology* 67, 225-257.
- Kumar, B., Yadav, A., Lang, J.C., Cipolla, M.J., Schmitt, A.C., Arradaza, N., Teknos, T.N. and Kumar, P. 2012. YM155 Reverses Cisplatin Resistance in Head and Neck Cancer by Decreasing Cytoplasmic Survivin Levels. *Molecular Cancer Therapeutics* 11, 1988-1998.
- Kuusisto, H.V., Wagstaff, K.M., Alvisi, G. and Jans, D.A. 2008. The C-terminus of apoptin represents a unique tumor cell-enhanced nuclear targeting module. *Int J Cancer* 123, 2965-2969.
- Kvansakul, M., Caria, S. and Hinds, M.G. 2017. The Bcl-2 Family in Host-Virus Interactions. *Viruses*, 9(10), 290.
- Lai, C.M. 1985. A study on a velogenic viscerotropic Newcastle disease virus *in vitro* and *in vivo*. PhD Thesis. Universiti Putra Malaysia.

- Lam, H.Y. 2011. Immunomodulatory effects of Newcastle disease virus AF2240 strain on human peripheral blood mononuclear cells. Masters Thesis. Universiti Putra Malaysia.
- Lam, H.Y., Yusoff, K., Yeap, S.K., Subramani, T., Abd-Aziz, S., Omar, A.R.and Alitheen, N.B. 2014. Immunomodulatory Effects of Newcastle Disease Virus AF2240 Strain on Human Peripheral Blood Mononuclear Cells. *International Journal of Medical Sciences* 11, 1240-1247.
- Lamb, R. A., and Kolakofsky, D. 1996. Paramyxoviridae: The viruses and their replication. In B. N. Fields, D. M. Knipe, and P. M. Howley (Eds.), *Fields virology: Third Edition* (3 ed., pp. 1449-1496). Philadelphia: Lippincott-Raven Press.
- Lambright, E.S., Kang, E.H., Force, S., Lanuti, M., Caparrelli, D., Kaiser, L.R., Albelda, S.M.and Molnar-Kimber, K.L. 2000. Effect of preexisting anti-herpes immunity on the efficacy of herpes simplex viral therapy in a murine intraperitoneal tumor model. *Molecular therapy : the journal of the American Society of Gene Therapy* 2, 387-393.
- Lanz, H.L., Suijker, J., Noteborn, M.H.and Backendorf, C. 2012. Proteasomal insensitivity of apoptin in tumor cells. *Biochemical and biophysical research communications* 422, 169-173.
- Laster, S.M., Wood, J.G., Gooding, L.R., 1988. Tumor necrosis factor can induce both apoptic and necrotic forms of cell lysis. *Journal of immunology* (Baltimore, Md. : 1950) 141, 2629-2634.
- Lee, Y.H., Cheng, C.M., Chang, Y.F., Wang, T.Y.and Yuo, C.Y. 2007. Apoptin T108 phosphorylation is not required for its tumor-specific nuclear localization but partially affects its apoptotic activity. *Biochemical and biophysical research communications* 354, 391-395.
- Leliveld, S.R., Dame, R.T., Mommaas, M.A., Koerten, H.K., Wyman, C., Danen-van Oorschot, A.A., Rohn, J.L., Noteborn, M.H.and Abrahams, J.P. 2003. Apoptin protein multimers form distinct higher-order nucleoprotein complexes with DNA. *Nucleic acids research* 31, 4805-4813.
- Leliveld, S.R., Zhang, Y.H., Rohn, J.L., Noteborn, M.H.and Abrahams, J.P. 2003b. Apoptin induces tumor-specific apoptosis as a globular multimer. *The Journal of biological chemistry* 278, 9042-9051.
- Li, J., Liu, Y., Wang, Z., Liu, K., Wang, Y., Liu, J., Ding, H.and Yuan, Z. 2011. Subversion of Cellular Autophagy Machinery by Hepatitis B Virus for Viral Envelopment. *Journal of Virology* 85, 6319-6333.

- Li, X., Jin, N., Mi, Z., Lian, H., Sun, L., Li, X. and Zheng, H. 2006. Antitumor effects of a recombinant fowlpox virus expressing Apoptin in vivo and in vitro. *Int J Cancer* 119, 2948-2957.
- Li, X., Liu, Y., Wen, Z., Li, C., Lu, H., Tian, M., Jin, K., Sun, L., Gao, P., Yang, E., Xu, X., Kan, S., Wang, Z., Wang, Y. and Jin, N. 2010. Potent anti-tumor effects of a dual specific oncolytic adenovirus expressing apoptin in vitro and in vivo. *Molecular cancer* 9, 10.
- Li, Y., Dowbenko, D. and Lasky, L.A. 2002. AKT/PKB phosphorylation of p21Cip/WAF1 enhances protein stability of p21Cip/WAF1 and promotes cell survival. *The Journal of biological chemistry* 277, 11352-11361.
- Lim, K. G. 2014. A review of colorectal cancer research in malaysia. *Medical Journal of Malaysia*, 69 Suppl A, 23-32.
- Lin, B., Kolluri, S.K., Lin, F., Liu, W., Han, Y.H., Cao, X., Dawson, M.I., Reed, J.C. and Zhang, X.K. 2004. Conversion of Bcl-2 from protector to killer by interaction with nuclear orphan receptor Nur77/TR3. *Cell* 116, 527-540.
- Lin, C.Y., Chang, T.W., Hsieh, W.H., Hung, M.C., Lin, I.H., Lai, S.C., Tzeng, Y.J., 2016. Simultaneous induction of apoptosis and necroptosis by Tanshinone IIA in human hepatocellular carcinoma HepG2 cells. *Cell death discovery* 2, 16065.
- Lindsay, J., Esposti, M.D., Gilmore, A.P., 2011. Bcl-2 proteins and mitochondria--specificity in membrane targeting for death. *Biochimica et biophysica acta* 1813, 532-539.
- Liu, L., Wu, W., Zhu, G., Liu, L., Guan, G., Li, X., Jin, N. and Chi, B. 2012. Therapeutic efficacy of an hTERT promoter-driven oncolytic adenovirus that expresses apoptin in gastric carcinoma. *Int J Mol Med* 30, 747-754.
- Liu, X., Yang, W., Guan, Z., Yu, W., Fan, B., Xu, N., Liao, D.J., 2018. There are only four basic modes of cell death, although there are many ad-hoc variants adapted to different situations. *Cell & bioscience* 8, 6.
- Liu, X., Zeidan, Y.H., Elojeimy, S., Holman, D.H., El-Zawahry, A.M., Guo, G.W., Bielawska, A., Bielawski, J., Szulc, Z., Rubinchik, S., Dong, J.Y., Keane, T.E., Tavassoli, M., Hannun, Y.A. and Norris, J.S. 2006. Involvement of sphingolipids in apoptin-induced cell killing. *Molecular therapy : the journal of the American Society of Gene Therapy* 14, 627-636.
- Lockshin, R.A. and Zakeri, Z. 2007. Cell death in health and disease. *J Cell Mol Med* 11, 1214-1224.
- Lomonosova, E., Subramanian, T., Chinnadurai, G., 2002. Requirement of BAX for efficient adenovirus-induced apoptosis. *J Virol* 76, 11283-11290.

- Los, M., Mozoluk, M., Ferrari, D., Stepczynska, A., Stroh, C., Renz, A., Herceg, Z., Wang, Z.Q., Schulze-Osthoff, K., 2002. Activation and caspase-mediated inhibition of PARP: a molecular switch between fibroblast necrosis and apoptosis in death receptor signaling. *Molecular biology of the cell* 13, 978-988.
- Ma, J.L., Han, S.X., Zhao, J., Zhang, D., Wang, L., Li, Y.D. and Zhu, Q. 2012. Systemic delivery of lentivirus-mediated secretable TAT-apoptin eradicates hepatocellular carcinoma xenografts in nude mice. *International journal of oncology* 41, 1013-1020.
- Maddika, S., Bay, G.H., Krocak, T.J., Ande, S.R., Maddika, S., Wiechec, E., Gibson, S.B. and Los, M. 2007. Akt is transferred to the nucleus of cells treated with apoptin, and it participates in apoptin-induced cell death. *Cell proliferation* 40, 835-848.
- Maddika, S., Booy, E.P., Johar, D., Gibson, S.B., Ghavami, S. and Los, M. 2005. Cancer-specific toxicity of apoptin is independent of death receptors but involves the loss of mitochondrial membrane potential and the release of mitochondrial cell-death mediators by a Nur77-dependent pathway. *Journal of cell science* 118, 4485-4493.
- Maddika, S., Panigrahi, S., Wiechec, E., Wesselborg, S., Fischer, U., Schulze-Osthoff, K. and Los, M. 2009. Unscheduled Akt-triggered activation of cyclin-dependent kinase 2 as a key effector mechanism of apoptin's anticancer toxicity. *Molecular and cellular biology* 29, 1235-1248.
- Manjili, M.H., Park, J., Facciponte, J.G. and Subjeck, J.R. 2005. HSP110 induces "danger signals" upon interaction with antigen presenting cells and mouse mammary carcinoma. *Immunobiology* 210, 295-303.
- Mansour, M., Palese, P. and Zamarin, D. 2011. Oncolytic Specificity of Newcastle Disease Virus Is Mediated by Selectivity for Apoptosis-Resistant Cells. *Journal of Virology* 85, 6015-6023.
- Mantovani, A., Sozzani, S., Locati, M., Allavena, P., Sica, A., 2002. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends in Immunology* 23, 549-555.
- Mason, E.J. and Kaufman, N. 1960. The toxic properties of massive inocula of Newcastle disease virus and influenza virus (PR8) for cell strains derived from normal and neoplastic tissue. *American Journal of Pathology* 37.
- Masoud, G.N. and Li, W. 2015. HIF-1 α pathway: role, regulation and intervention for cancer therapy. *Acta Pharmaceutica Sinica B* 5, 378-389.
- McNamara, C.R., Ahuja, R., Osafo-Addo, A.D., Barrows, D., Kettenbach, A., Skidan, I., Teng, X., Cuny, G.D., Gerber, S., Degterev, A., 2013. Akt

- Regulates TNFalpha synthesis downstream of RIP1 kinase activation during necroptosis. *PloS one* 8, e56576.
- Medici, M.A., Sciortino, M.T., Perri, D., Amici, C., Avitabile, E., Ciotti, M., Balestrieri, E., De Smaele, E., Franzoso, G. and Mastino, A. 2003. Protection by herpes simplex virus glycoprotein D against Fas-mediated apoptosis: role of nuclear factor kappaB. *The Journal of biological chemistry* 278, 36059-36067.
- Medrano, R.F.V., Hunger, A., Mendonça, S.A., Barbuto, J.A.M. and Strauss, B.E. 2017. Immunomodulatory and antitumor effects of type I interferons and their application in cancer therapy. *Oncotarget*.
- Megyeri, K., Orosz, L., Kemeny, L., 2007. Vesicular stomatitis virus infection triggers apoptosis associated with decreased DeltaNp63alpha and increased Bax levels in the immortalized HaCaT keratinocyte cell line. *Biomedicine & pharmacotherapy*; 61, 254-260.
- Meier, O. and Greber, U.F. 2003. Adenovirus endocytosis. *The journal of gene medicine* 5, 451-462.
- Meyyapan, N. 2003. Oncolytic effect of Newcastle disease virus on the MCF-7 and MDA-MB-231 breast cancer cell lines. Masters Thesis, Universiti Putra Malaysia.
- Mills, C.D., Kincaid, K., Alt, J.M., Heilman, M.J. and Hill, A.M. 2000. M-1/M-2 Macrophages and the Th1/Th2 Paradigm. *The Journal of Immunology* 164, 6166-6173.
- Mitrus, I., Missol-Kolka, E., Plucienniczak, A. and Szala, S. 2005. Tumour therapy with genes encoding apoptin and E4orf4. *Anticancer research* 25, 1087-1090.
- Miwa, M., Ura, M., Nishida, M., Sawada, N., Ishikawa, T., Mori, K., Shimma, N., Umeda, I., Ishitsuka, H. 1998. Design of a novel oral fluoropyrimidine carbamate, capecitabine, which generates 5-fluorouracil selectively in tumours by enzymes concentrated in human liver and cancer tissue. *European Journal of Cancer*, 34(8), 1274-1281.
- Moehler, M.H., Zeidler, M., Wilsberg, V., Cornelis, J.J., Woelfel, T., Rommelaere, J., Galle, P.R. and Heike, M. 2005. Parvovirus H-1-induced tumor cell death enhances human immune response in vitro via increased phagocytosis, maturation, and cross-presentation by dendritic cells. *Human gene therapy* 16, 996-1005.
- Mohamed, Z., Ahmad, R., Yoke, N. S., Zakaria, Z., Ahmad, H., and Yew, T. H. 2003. A nonsense mutation in exon 8 of the APC gene (Arg283Ter) causes

- clinically variable FAP in a Malaysian Chinese family. *Cancer Sciences*, 94(8), 725-728.
- Mok, H.-P.and Lever, A.M.L. 2007. Chromatin, gene silencing and HIV latency. *Genome Biology* 8, 228.
- Molouki, A., Hsu, Y.-T., Jahanshiri, F., Abdullah, S., Rosli, R.and Yusoff, K. 2011. The matrix (M) protein of newcastle disease virus binds to human bax through its BH3 domain. *Virology Journal* 8, 385.
- Molouki, A., Hsu, Y.-T., Jahanshiri, F., Rosli, R.and Yusoff, K. 2010. Newcastle Disease Virus Infection Promotes Bax Redistribution to Mitochondria and Cell Death in HeLa Cells. *Intervirology* 53, 87-94.
- Molouki, A.and Yusoff, K. 2012. NDV-induced apoptosis in absence of Bax; evidence of involvement of apoptotic proteins upstream of mitochondria. *Virology Journal* 9, 179.
- Morales, J., Li, L., Fattah, F.J., Dong, Y., Bey, E.A., Patel, M., Gao, J., Boothman, D.A., 2014. Review of poly (ADP-ribose) polymerase (PARP) mechanisms of action and rationale for targeting in cancer and other diseases. *Critical reviews in eukaryotic gene expression* 24, 15-28.
- Motalleb, G., Othman, F., Aini, I., Asmah, R., Anushia, S., Zolkapli, E.and Saidi, M. 2009a. Detection of Newcastle disease virus (NDV-AF2240) in lung during intratumoral injection in 4t1 breast cancer in Balb/c mice. *Malaysian Journal of Microscopy* 5, 41-52.
- Motalleb, G., Othman, F., Ideris, A.and Rahmat, A. 2009b. Dissemination of Newcastle disease virus (NDV-AF2240) in liver during intratumoral injection of xenotransplant breast cancer in BALB/c Mice. *Yaktek Medical Journal* 11, 303-310.
- Murphy, P.M. 2001. Viral exploitation and subversion of the immune system through chemokine mimicry. *Nature Immunology* 2, 116-122.
- Murray, D.R., Cassel, W.A., Torbin, A.H., Olkowski, Z.L.and Moore, M.E. 1977. Viral oncolysate in the management of malignant melanoma.II. Clinical studies. *Cancer* 40, 680-686.
- Murulitharan, K., Yusoff, K., Omar, A.R.and Molouki, A. 2013. Characterization of Malaysian velogenic NDV strain AF2240-I genomic sequence: a comparative study. *Virus Genes* 46, 431-440.
- Murulitharan, K. 2015. Characterisation and Rescue of a Recombinant Newcastle Disease Virus Strain AF2240-I. PhD Thesis, Universiti Putra Malaysia.

- Mustapha, M. A., Shahpuddin, M., Nurfatimah, S., Aziz, A., Aizat, A., Bhavaraju, M.K., Naik, V., Zakaria, Z., Shanwani, A., Hassan, M. and Ankathil, R. 2011. Polymorphism in the Tumor Necrosis Factor Alpha Promoter Region and Its Influence on Colorectal Cancer Predisposition Risk in Malaysian Population. *International Medical Journal*, 18(4).
- Mustapha, M. A., Shahpuddin, S. N., Aziz, A. A., and Ankathil, R. 2012. Risk modification of colorectal cancer susceptibility by interleukin-8 -251T>A polymorphism in Malaysians. *World Journal of Gastroenterology*, 18(21), 2668-2673.
- Nagai, Y., Hamaguchi, M. and Toyoda, T. 1989. Molecular biology of Newcastle disease virus. *Progress in veterinary microbiology and immunology* 5, 16-64.
- Nakaya, T., Cros, J., Park, M.S., Nakaya, Y., Zheng, H., Sagrera, A., Villar, E., Garcia-Sastre, A. and Palese, P. 2001. Recombinant Newcastle disease virus as a vaccine vector. *J Virol* 75, 11868-11873.
- Nelson, N.J. 1999. Scientific Interest in Newcastle Disease Virus Is Reviving. *JNCI Journal of the National Cancer Institute* 91, 1708-1710.
- Neumann, G. and Kawaoka, Y. 2004. Reverse genetics systems for the generation of segmented negative-sense RNA viruses entirely from cloned cDNA. *Current topics in microbiology and immunology* 283, 43-60.
- Newhauser, W. D., Berrington de Gonzalez, A., Schulte, R., and Lee, C. 2016. A Review of Radiotherapy-Induced Late Effects Research after Advanced Technology Treatments. *Frontiers in Oncology*, 6, 13.
- Nikoletopoulou, V., Markaki, M., Palikaras, K., Tavernarakis, N., 2013. Crosstalk between apoptosis, necrosis and autophagy. *Biochimica et biophysica acta* 1833, 3448-3459.
- Nizam, Z. M., Abdul Aziz, A. A., Kaur, G., Abu Hassan, M. R., Mohd Sidek, A. S., Yeh, L. Y., Mazuwin, M. and Ankathil, R. 2013. Contribution of the MLH1 -93G>a promoter polymorphism in modulating susceptibility risk in Malaysian colorectal cancer patients. *Asian Pacific Journal of Cancer Prevention*, 14(2), 619-624.
- Noteborn, M.H., Todd, D., Verschueren, C.A., de Gauw, H.W., Curran, W.L., Veldkamp, S., Douglas, A.J., McNulty, M.S., van der, E.A. and Koch, G. 1994. A single chicken anemia virus protein induces apoptosis. *J Virol* 68, 346-351.
- Noteborn, M.H., Verschueren, C.A., Koch, G. and Van der Eb, A.J. 1998. Simultaneous expression of recombinant baculovirus-encoded chicken anaemia virus (CAV) proteins VP1 and VP2 is required for formation of

- the CAV-specific neutralizing epitope. *The Journal of general virology* 79 (Pt 12), 3073-3077.
- Nudson, W.A., Rovnak, J., Buechner, M.and Quackenbush, S.L. 2003. Walleye dermal sarcoma virus Orf C is targeted to the mitochondria. *The Journal of general virology* 84, 375-381.
- Ochiai, H., Campbell, S.A., Archer, G.E., Chewning, T.A., Dragunsky, E., Ivanov, A., Gromeier, M.and Sampson, J.H. 2006. Targeted therapy for glioblastoma multiforme neoplastic meningitis with intrathecal delivery of an oncolytic recombinant poliovirus. *Clinical cancer research : an official journal of the American Association for Cancer Research* 12, 1349-1354.
- Ogasawara, T., Gotoh, B., Suzuki, H., Asaka, J., Shimokata, K., Rott, R.and Nagai, Y. 1992. Expression of factor X and its significance for the determination of paramyxovirus tropism in the chick embryo. *Embo j* 11, 467-472.
- Omar, A.R., Ideris, A., Ali, A.M., Othman, F., Yusoff, K., Abdullah, J.M., Wali, H.S.M.W., Zawawi, M.and Meyappan, N. 2003. An Overview on the Development of Newcastle Disease Virus as an Anti-Cancer Therapy. *Malaysian Journal of Medical Sciences* 10, 4-12.
- Osterlund, P., Orpana, A., Elomaa, I., Repo, H., and Joensuu, H. 2002. Raltitrexed treatment promotes systemic inflammatory reaction in patients with colorectal carcinoma. *Br J Cancer*, 87(6), 591-599.
- Othman, F., Ideris, A., Motalleb, G., Eshak, Z.and Rahmat, A. 2009. Oncolytic effect of Newcastle disease virus AF2240 strain on the MCF-7 breast cancer cell line. *Yaktek Medical Journal* 12, 17-24.
- Othman, F., Omar, A.R., Patimah, I.and Aini, I. 2002. Microscopic evaluation of Newcastle disease virus (NDV) a killer in chicken but a possible live saver in human. *Journal of Electron Microscopy Society Thailand* 16.
- Palumbo, M.O., Kavan, P., Miller, W.H., Jr., Panasci, L., Assouline, S., Johnson, N., Cohen, V., Patenaude, F., Pollak, M., Jagoe, R.T.and Batist, G. 2013. Systemic cancer therapy: achievements and challenges that lie ahead. *Frontiers in pharmacology* 4, 57.
- Pan, Y., Fang, L., Fan, H., Luo, R., Zhao, Q., Chen, H.and Xiao, S. 2010. Antitumor effects of a recombinant pseudotype baculovirus expressing Apoptin in vitro and in vivo. *Int J Cancer* 126, 2741-2751.
- Pandurangan, A. K., Divya, T., Kumar, K., Dineshbabu, V., Velavan, B., and Sudhandiran, G. 2018. Colorectal carcinogenesis: Insights into the cell death and signal transduction pathways: A review. *World Journal of Gastrointestinal Oncology*, 10(9), 244-259.

- Pasquinucci, G. 1971. Possible effect of measles on leukaemia. *Lancet (London, England)* 1, 136.
- Pekarsky, Y., Hallas, C., Palamarchuk, A., Koval, A., Bullrich, F., Hirata, Y., Bichi, R., Letofsky, J. and Croce, C.M. 2001. Akt phosphorylates and regulates the orphan nuclear receptor Nur77. *Proceedings of the National Academy of Sciences of the United States of America* 98, 3690-3694.
- Peng, D.J., Sun, J., Wang, Y.Z., Tian, J., Zhang, Y.H., Noteborn, M.H. and Qu, S. 2007. Inhibition of hepatocarcinoma by systemic delivery of Apoptin gene via the hepatic asialoglycoprotein receptor. *Cancer gene therapy* 14, 66-73.
- Phoolcharoen, W. and Smith, D.R. 2004. Internalization of the dengue virus is cell cycle modulated in HepG2, but not vero cells. *Journal of Medical Virology* 74, 434-441.
- Pietersen, A.M., van der Eb, M.M., Rademaker, H.J., van den Wollenberg, D.J., Rabelink, M.J., Kuppen, P.J., van Dierendonck, J.H., van Ormondt, H., Masman, D., van de Velde, C.J., van der Eb, A.J., Hoeben, R.C. and Noteborn, M.H. 1999. Specific tumor-cell killing with adenovirus vectors containing the apoptin gene. *Gene therapy* 6, 882-892.
- Ploegh, H.L. 1998. Viral Strategies of Immune Evasion. *Science* 280, 248-253.
- Pol, J., Kroemer, G. and Galluzzi, L. 2016. First oncolytic virus approved for melanoma immunotherapy. *Oncimmunology* 5, e1115641.
- Poon, I.K., Oro, C., Dias, M.M., Zhang, J. and Jans, D.A. 2005. Apoptin nuclear accumulation is modulated by a CRM1-recognized nuclear export signal that is active in normal but not in tumor cells. *Cancer Res* 65, 7059-7064.
- Porotto, M., Salah, Z., DeVito, I., Talekar, A., Palmer, S.G., Xu, R., Wilson, I.A. and Moscona, A. 2012. The second receptor binding site of the globular head of the Newcastle disease virus hemagglutinin-neuraminidase activates the stalk of multiple paramyxovirus receptor binding proteins to trigger fusion. *J Virol* 86, 5730-5741.
- Prasetyo, A.A., Kamahora, T., Kuroishi, A., Murakami, K. and Hino, S. 2009. Replication of chicken anemia virus (CAV) requires apoptin and is complemented by VP3 of human torque teno virus (TTV). *Virology* 385, 85-92.
- Prehaud, C., Lay, S., Dietzschold, B. and Lafon, M. 2003. Glycoprotein of Nonpathogenic Rabies Viruses Is a Key Determinant of Human Cell Apoptosis. *Journal of Virology* 77, 10537-10547.

- Prestwich, R.J., Harrington, K.J., Pandha, H.S., Vile, R.G., Melcher, A.A. and Errington, F. 2008. Oncolytic viruses: a novel form of immunotherapy. *Expert review of anticancer therapy* 8, 1581-1588.
- Prince, A.M. and Ginsberg, H.S. 1957. Studies on the cytotoxic effect of Newcastle disease virus (NDV) on Ehrlich ascites tumor cells. I. Characteristics of the virus-cell interaction. *Journal of Immunology* 79, 94-106.
- Puhler, F., Willuda, J., Puhlmann, J., Mumberg, D., Romer-Oberdorfer, A. and Beier, R. 2008. Generation of a recombinant oncolytic Newcastle disease virus and expression of a full IgG antibody from two transgenes. *Gene therapy* 15, 371-383.
- Racaniello, V.R. and Baltimore, D. 1981. Cloned poliovirus complementary DNA is infectious in mammalian cells. *Science* 214, 916-919.
- Radecke, F., Spielhofer, P., Schneider, H., Kaelin, K., Huber, M., Dotsch, C., Christiansen, G. and Billeter, M.A. 1995. Rescue of measles viruses from cloned DNA. *Embo j* 14, 5773-5784.
- Rangaswamy, U.S., Wang, W., Cheng, X., McTamney, P., Carroll, D. and Jin, H. 2017. Newcastle Disease Virus Establishes Persistent Infection in Tumor Cells In Vitro : Contribution of the Cleavage Site of Fusion Protein and Second Sialic Acid Binding Site of Hemagglutinin-Neuraminidase. *Journal of Virology* 91, e00770-00717.
- Rath, A., Glibowicka, M., Nadeau, V.G., Chen, G. and Deber, C.M. 2009. Detergent binding explains anomalous SDS-PAGE migration of membrane proteins. *Proceedings of the National Academy of Sciences of the United States of America* 106, 1760-1765.
- Rathmell, J.C. and Thompson, C.B. 1999. The Central Effectors Of Cell Death In The Immune System. *Annual Review of Immunology* 17, 781-828.
- Reading, P.C., Khanna, A. and Smith, G.L. 2002. Vaccinia virus CrmE encodes a soluble and cell surface tumor necrosis factor receptor that contributes to virus virulence. *Virology* 292, 285-298.
- Reeve, P. and Poste, G. 1971. Studies on the cytopathogenicity of Newcastle disease virus: relation between virulence, polykaryocytosis and plaque size. *The Journal of general virology* 11, 17-24.
- Reichard, K.W., Lorence, R.M., Cascino, C.J., Peeples, M.E., Walter, R.J., Fernando, M.B., Reyes, H.M. and Greager, J.A. 1992. Newcastle disease virus selectively kills human tumor cells. *Journal of Surgical Research* 52, 448-453.
- Ren, G., Tian, G., Liu, Y., He, J., Gao, X., Yu, Y., Liu, X., Zhang, X., Sun, T., Liu, S., Yin, J. and Li, D. 2016. Recombinant Newcastle Disease Virus Encoding

IL-12 and/or IL-2 as Potential Candidate for Hepatoma Carcinoma Therapy.
Technology in cancer research & treatment 15, Np83-94.

Ricca, J.M., Oseledchyk, A., Walther, T., Liu, C., Mangarin, L., Merghoub, T., Wolchok, J.D. and Zamarin, D. 2018. Pre-existing Immunity to Oncolytic Virus Potentiates Its Immunotherapeutic Efficacy. *Molecular therapy : the journal of the American Society of Gene Therapy* 26, 1008-1019.

Rodriguez-Grille, J., Busch, L.K., Martinez-Costas, J. and Benavente, J. 2014. Avian reovirus-triggered apoptosis enhances both virus spread and the processing of the viral nonstructural muNS protein. *Virology* 462-463, 49-59.

Rogers, C., Fernandes-Alnemri, T., Mayes, L., Alnemri, D., Cingolani, G., Alnemri, E.S., 2017. Cleavage of DFNA5 by caspase-3 during apoptosis mediates progression to secondary necrotic/pyroptotic cell death. *Nat Commun* 8, 14128.

Rohn, J.L., Zhang, Y.H., Aalbers, R.I., Otto, N., Den Hertog, J., Henriquez, N.V., Van De Velde, C.J., Kuppen, P.J., Mumberg, D., Donner, P. and Noteborn, M.H. 2002. A tumor-specific kinase activity regulates the viral death protein Apoptin. *The Journal of biological chemistry* 277, 50820-50827.

Rollano Penalosa, O.M., Lewandowska, M., Stetefeld, J., Ossysek, K., Madej, M., Bereta, J., Sobczak, M., Shojaei, S., Ghavami, S. and Los, M.J. 2014. Apoptins: selective anticancer agents. *Trends in molecular medicine* 20, 519-528.

Roohani, K., Tan, S.W., Yeap, S.K., Ideris, A., Bejo, M.H. and Omar, A.R. 2015. Characterisation of genotype VII Newcastle disease virus (NDV) isolated from NDV vaccinated chickens, and the efficacy of LaSota and recombinant genotype VII vaccines against challenge with velogenic NDV. *Journal of Veterinary Science* 16, 447.

Rothenberg, M. L., Meropol, N. J., Poplin, E. A., Van Cutsem, E., and Wadler, S. 2001. Mortality associated with irinotecan plus bolus fluorouracil/leucovorin: summary findings of an independent panel. *Journal of Clinical Oncology*, 19(18), 3801-3807.

Rothenberg, M. L., Oza, A. M., Bigelow, R. H., Berlin, J. D., Marshall, J. L., Ramanathan, R. K., Hart, L.L., Gupta, S., Garay, C.A., Burger, B.G., Le Bail, N. and Haller, D. G. 2003. Superiority of oxaliplatin and fluorouracil-leucovorin compared with either therapy alone in patients with progressive colorectal cancer after irinotecan and fluorouracil-leucovorin: interim results of a phase III trial. *Journal of Clinical Oncology*, 21(11), 2059-2069.

Rousalova, I. and Krepela, E. 2010. Granzyme B-induced apoptosis in cancer cells and its regulation (review). *International journal of oncology* 37, 1361-1378.

- Samson, A.C., Levesley, I.and Russell, P.H. 1991. The 36K polypeptide synthesized in Newcastle disease virus-infected cells possesses properties predicted for the hypothesized 'V' protein. *The Journal of general virology* 72 (Pt 7), 1709-1713.
- San Román, K., Villar, E.and Muñoz-Barroso, I. 1999. Acidic pH Enhancement of the Fusion of Newcastle Disease Virus with Cultured Cells. *Virology* 260, 329-341.
- Sanchez-Felipe, L., Villar, E.and Munoz-Barroso, I. 2012. alpha2-3- and alpha2-6-N-linked sialic acids allow efficient interaction of Newcastle Disease Virus with target cells. *Glycoconjugate journal* 29, 539-549.
- Sanchez-Felipe, L., Villar, E.and Munoz-Barroso, I. 2014. Entry of Newcastle Disease Virus into the host cell: role of acidic pH and endocytosis. *Biochimica et biophysica acta* 1838, 300-309.
- Schirrmacher, V. 2016. Fifty Years of Clinical Application of Newcastle Disease Virus: Time to Celebrate! *Biomedicines* 4.
- Schirrmacher, V.and Fournier, P. 2009. Newcastle Disease Virus: A Promising Vector for Viral Therapy, Immune Therapy, and Gene Therapy of Cancer, Gene Therapy of Cancer. *Humana Press*, pp. 565-605.
- Schloer, G.M.and Hanson, R.P. 1968. Relationship of plaque size and virulence for chickens of 14 representative Newcastle disease virus strains. *J Virol* 2, 40-47.
- Schnell, M.J., Mebatson, T.and Conzelmann, K.K. 1994. Infectious rabies viruses from cloned cDNA. *Embo j* 13, 4195-4203.
- Schoop, R.A., Kooistra, K., Baatenburg De Jong, R.J.and Noteborn, M.H. 2004. Bcl-xL inhibits p53- but not apoptin-induced apoptosis in head and neck squamous cell carcinoma cell line. *Int J Cancer* 109, 38-42.
- Shen, D.W., Pouliot, L.M., Hall, M.D.and Gottesman, M.M. 2012. Cisplatin Resistance: A Cellular Self-Defense Mechanism Resulting from Multiple Epigenetic and Genetic Changes. *Pharmacological Reviews* 64, 706-721.
- Shih, W.L., Hsu, H.W., Liao, M.H., Lee, L.H.and Liu, H.J. 2004. Avian reovirus sigmaC protein induces apoptosis in cultured cells. *Virology* 321, 65-74.
- Shoae-Hassani, A., Keyhanvar, P., Seifalian, A.M., Mortazavi-Tabatabaei, S.A., Ghaderi, N., Issazadeh, K., Amirmozafari, N.and Verdi, J. 2013. lambda Phage nanobioparticle expressing apoptin efficiently suppress human breast carcinoma tumor growth in vivo. *PloS one* 8, e79907.

- Shobana, R., Samal, S.K.and Elankumaran, S. 2013. Prostate-specific antigen-retargeted recombinant newcastle disease virus for prostate cancer virotherapy. *J Virol* 87, 3792-3800.
- Shussman, N.and Wexner, S.D. 2014. Colorectal polyps and polyposis syndromes. *Gastroenterology report* 2, 1-15.
- Siegel, R. L., Miller, K. D., and Jemal, A. 2015. Cancer statistics, 2015. *CA: A Cancer Journal for Clinicians*, 65(1), 5-29.
- Silva, M.T., 2010. Secondary necrosis: the natural outcome of the complete apoptotic program. *FEBS letters* 584, 4491-4499.
- Singh, P.K., Doley, J., Kumar, G.R., Sahoo, A.P.and Tiwari, A.K. 2012. Oncolytic viruses & their specific targeting to tumour cells. *The Indian journal of medical research* 136, 571-584.
- Sinkovics, J.G.and Horvath, J.C. 2000. Newcastle disease virus (NDV): brief history of its oncolytic strains. *Journal of Clinical Virology* 16, 1-15.
- Skaletskaya, A., Bartle, L.M., Chittenden, T., McCormick, A.L., Mocarski, E.S.and Goldmacher, V.S. 2001. A cytomegalovirus-encoded inhibitor of apoptosis that suppresses caspase-8 activation. *Proceedings of the National Academy of Sciences* 98, 7829-7834.
- Soldani, C., Lazze, M.C., Bottone, M.G., Tognon, G., Biggiogera, M., Pellicciari, C.E., Scovassi, A.I., 2001. Poly(ADP-ribose) polymerase cleavage during apoptosis: when and where? *Experimental cell research* 269, 193-201.
- Son, K.N., Becker, R.P., Kallio, P., Lipton, H.L., 2008. Theiler's virus-induced intrinsic apoptosis in M1-D macrophages is Bax mediated and restricts virus infectivity: a mechanism for persistence of a cytopolytic virus. *J Virol* 82, 4502-4510.
- Sosna, J., Voigt, S., Mathieu, S., Lange, A., Thon, L., Davarnia, P., Herdegen, T., Linkermann, A., Rittger, A., Chan, F.K., Kabelitz, D., Schutze, S., Adam, D., 2014. TNF-induced necroptosis and PARP-1-mediated necrosis represent distinct routes to programmed necrotic cell death. *Cellular and molecular life sciences : CMSL* 71, 331-348.
- Spanos, C. P., Mamopoulos, A., Tsapas, A., Syrakos, T., and Kiskinis, D. 2008. Female fertility and colorectal cancer. *International Journal of Colorectal Disease*, 23(8), 735-743.
- Steward, M., Vipond, I.B., Millar, N.S.and Emmerson, P.T. 1993. RNA editing in Newcastle disease virus. *The Journal of general virology* 74 (Pt 12), 2539-2547.

- Stojdl, D.F., Lichty, B.D., tenOever, B.R., Paterson, J.M., Power, A.T., Knowles, S., Marius, R., Reynard, J., Poliquin, L., Atkins, H., Brown, E.G., Durbin, R.K., Durbin, J.E., Hiscott, J. and Bell, J.C. 2003. VSV strains with defects in their ability to shutdown innate immunity are potent systemic anti-cancer agents. *Cancer cell* 4, 263-275.
- Sturmer, T., Buring, J. E., Lee, I. M., Gaziano, J. M., and Glynn, R. J. 2006. Metabolic abnormalities and risk for colorectal cancer in the physicians' health study. *Cancer Epidemiology Biomarkers Prevention*, 15(12), 2391-2397.
- Sun, J., Yan, Y., Wang, X.T., Liu, X.W., Peng, D.J., Wang, M., Tian, J., Zong, Y.Q., Zhang, Y.H., Noteborn, M.H. and Qu, S. 2009. PTD4-apoptin protein therapy inhibits tumor growth in vivo. *Int J Cancer* 124, 2973-2981.
- Sun, L., Wang, H., Wang, Z., He, S., Chen, S., Liao, D., Wang, L., Yan, J., Liu, W., Lei, X., Wang, X., 2012. Mixed lineage kinase domain-like protein mediates necrosis signaling downstream of RIP3 kinase. *Cell* 148, 213-227.
- Takahashi, J.S., Pinto, L.H. and Vitaterna, M.H. 1994. Forward and reverse genetic approaches to behavior in the mouse. *Science* 264, 1724-1733.
- Taniguchi, T., Palmieri, M. and Weissmann, C. 1978. A Qbeta DNA-containing hybrid plasmid giving rise to Qbeta phage formation in the bacterial host [proceedings]. *Annales de microbiologie* 129 b, 535-536.
- Todd, D., Creelan, J.L., Mackie, D.P., Rixon, F. and McNulty, M.S. 1990. Purification and biochemical characterization of chicken anaemia agent. *The Journal of general virology* 71 (Pt 4), 819-823.
- Toth, C.A. and Thomas, P. 1992. Type I interferon resistance in a colorectal cancer cell line is associated with a more aggressive phenotype *in vivo*. *British Journal of Cancer* 65, 365-368.
- Toyoda, T., Sakaguchi, T., Imai, K., Inocencio, N.M., Gotoh, B., Hamaguchi, M. and Nagai, Y. 1987. Structural comparison of the cleavage-activation site of the fusion glycoprotein between virulent and avirulent strains of Newcastle disease virus. *Virology* 158, 242-247.
- Trombetta, E.S., Ebersold, M., Garrett, W., Pypaert, M. and Mellman, I. 2003. Activation of lysosomal function during dendritic cell maturation. *Science* 299, 1400-1403.
- Trump, B.F., Berezesky, I.K., 1995. Calcium-mediated cell injury and cell death. FASEB journal : official publication of the Federation of American Societies for Experimental Biology 9, 219-228.

- Trump, B.F., Berezesky, I.K., Chang, S.H., Phelps, P.C., 1997. The pathways of cell death: oncosis, apoptosis, and necrosis. *Toxicologic pathology* 25, 82-88.
- Turley, S.J., Inaba, K., Garrett, W.S., Ebersold, M., Unternaehrer, J., Steinman, R.M. and Mellman, I. 2000. Transport of peptide-MHC class II complexes in developing dendritic cells. *Science* 288, 522-527.
- van der Eb, M.M., Pietersen, A.M., Speetjens, F.M., Kuppen, P.J., van de Velde, C.J., Noteborn, M.H. and Hoeben, R.C. 2002. Gene therapy with apotin induces regression of xenografted human hepatomas. *Cancer gene therapy* 9, 53-61.
- van Gennip, H.G., van Rijn, P.A., Widjojoatmodjo, M.N. and Moormann, R.J. 1999. Recovery of infectious classical swine fever virus (CSFV) from full-length genomic cDNA clones by a swine kidney cell line expressing bacteriophage T7 RNA polymerase. *Journal of virological methods* 78, 117-128.
- Vanden Berghe, T., Vanlangenakker, N., Parthoens, E., Deckers, W., Devos, M., Festjens, N., Guerin, C.J., Brunk, U.T., Declercq, W., Vandenabeele, P., 2010. Necroptosis, necrosis and secondary necrosis converge on similar cellular disintegration features. *Cell Death Differ* 17, 922-930.
- Vandenabeele, P., Galluzzi, L., Vanden Berghe, T., Kroemer, G., 2010. Molecular mechanisms of necroptosis: an ordered cellular explosion. *Nature reviews. Molecular cell biology* 11, 700-714.
- Vercammen, D., Brouckaert, G., Denecker, G., Van de Craen, M., Declercq, W., Fiers, W., Vandenabeele, P., 1998. Dual signaling of the Fas receptor: initiation of both apoptotic and necrotic cell death pathways. *The Journal of experimental medicine* 188, 919-930.
- Vigil, A., Park, M.S., Martinez, O., Chua, M.A., Xiao, S., Cros, J.F., Martinez-Sobrido, L., Woo, S.L. and Garcia-Sastre, A. 2007. Use of reverse genetics to enhance the oncolytic properties of Newcastle disease virus. *Cancer Res* 67, 8285-8292.
- Vikhanskaya, F., Falugi, C., Valente, P. and Russo, P. 2002. Human papillomavirus type 16 E6-enhanced susceptibility to apoptosis induced by TNF in A2780 human ovarian cancer cell line. *Int J Cancer* 97, 732-739.
- Wadia, J.S., Wagner, M.V., Ezhevsky, S.A. and Dowdy, S.F. 2004. Apoptin/VP3 contains a concentration-dependent nuclear localization signal (NLS), not a tumorigenic selective NLS. *J Virol* 78, 6077-6078.
- Wang, C., Wang, W., Wang, J., Zhan, H., Jiang, L., Yan, R., Hou, Z., Zhu, H., Yu, L., Shi, Y., Ding, M. and Ke, C. 2013. Apoptin induces apoptosis in nude mice allograft model of human bladder cancer by altering multiple bladder

- tumor-associated gene expression profiles. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine* 34, 1667-1678.
- Weerasinghe, P., Buja, L.M., 2012. Oncosis: an important non-apoptotic mode of cell death. *Experimental and molecular pathology* 93, 302-308.
- Weller, T.H., Robbins, F.C. and Enders, J.F. 1949. Cultivation of poliomyelitis virus in cultures of human foreskin and embryonic tissues. *Proceedings of the Society for Experimental Biology and Medicine. Society for Experimental Biology and Medicine (New York, N.Y.)* 72, 153-155.
- Wheelock, E.F. and Dingle, J.H. 1964. Observations on the Repeated Administration of Viruses to a Patient with Acute Leukemia. *New England Journal of Medicine* 271, 645-651.
- Wheelock, E.F. and Tamm, I. 1959. Mitosis and division in HeLa cells infected with influenza or Newcastle disease virus. *Virology* 8, 532-536.
- Williams, C.B., Yeh, E.S. and Soloff, A.C. 2016. Tumor-associated macrophages: unwitting accomplices in breast cancer malignancy. *npj Breast Cancer* 2.
- Wood, D.E. and Newcomb, E.W. 2000. Cleavage of Bax enhances its cell death function. *Experimental cell research* 256, 375-382.
- Wu, A. H., Paganini-Hill, A., Ross, R. K., and Henderson, B. E. 1987. Alcohol, physical activity and other risk factors for colorectal cancer: a prospective study. *British Journal of Cancer*, 55(6), 687-694
- Wu, X. and Rupprecht, C.E. 2008. Glycoprotein gene relocation in rabies virus. *Virus Res* 131, 95-99.
- Wu, Y., Zhang, X., Wang, X., Wang, L., Hu, S., Liu, X. and Meng, S. 2012. Apoptin enhances the oncolytic properties of Newcastle disease virus. *Intervirology*; 55(4): 276-86.
- Xiong, Y., Shan, J., Liu, J., Zhao, K., Chen, S., Xu, W., Zhou, Q., Yang, M. and Lei, X. 2017. Californium-252 neutron intracavity brachytherapy alone for T1N0 low-lying rectal adenocarcinoma: A definitive anal sphincter-preserving radiotherapy. *Scientific Reports*, 7, 40619.
- Yan, Y. and Samal, S.K. 2008. Role of intergenic sequences in newcastle disease virus RNA transcription and pathogenesis. *J Virol* 82, 1323-1331.
- Yang, J., Zhao, Y., Zhang, L., Fan, H., Qi, C., Zhang, K., Liu, X., Fei, L., Chen, S., Wang, M., Kuang, F., Wang, Y., Wu, S., 2018. RIPK3/MLKL-Mediated Neuronal Necrosis Modulates the M1/M2 Polarization of

- Microglia/Macrophages in the Ischemic Cortex. Cerebral cortex (New York, N.Y. : 1991) 28, 2622-2635.
- Ying, L., Huaxia, W., Xiang, M., Hongjie, F., Huang, W., Yanrong, L., Yinjie, S., Chun, M., Lei, T., Cuiping, S., Xusheng, Q. and Chan, D. 2017. RIP1 is a central signaling protein in regulation of TNF-α/TRAIL mediated apoptosis and necroptosis during Newcastle disease virus infection. *Oncotarget*.
- Yu, X. and He, S. 2016. The interplay between human herpes simplex virus infection and the apoptosis and necroptosis cell death pathways. *Virol J* 13, 77.
- Yuan, L., Zhao, H., Zhang, L. and Liu, X. 2013. The efficacy of combination therapy using adeno-associated virus-mediated co-expression of apoptin and interleukin-24 on hepatocellular carcinoma. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine* 34, 3027-3034.
- Zahary, M. N., Kaur, G., Abu Hassan, M. R., Singh, H., Naik, V. R., and Ankathil, R. 2012. Germline mutation analysis of MLH1 and MSH2 in Malaysian Lynch syndrome patients. *World Journal of Gastroenterology*, 18(8), 814-820.
- Zamarin, D., Holmgard, R.B., Subudhi, S.K., Park, J.S., Mansour, M., Palese, P., Merghoub, T., Wolchok, J.D. and Allison, J.P. 2014. Localized Oncolytic Virotherapy Overcomes Systemic Tumor Resistance to Immune Checkpoint Blockade Immunotherapy. *Science Translational Medicine* 6, 226ra232-226ra232.
- Zamarin, D., Martinez-Sobrido, L., Kelly, K., Mansour, M., Sheng, G., Vigil, A., Garcia-Sastre, A., Palese, P. and Fong, Y. 2009. Enhancement of oncolytic properties of recombinant newcastle disease virus through antagonism of cellular innate immune responses. *Molecular therapy : the journal of the American Society of Gene Therapy* 17, 697-706.
- Zamarin, D. and Palese, P. 2012. Oncolytic Newcastle disease virus for cancer therapy: old challenges and new directions. *Future microbiology* 7, 347-367.
- Zaslavsky, E., Hershberg, U., Seto, J., Pham, A.M., Marquez, S., Duke, J.L., Wetmur, J.G., tenOever, B.R., Sealfon, S.C. and Kleinstein, S.H. 2010. Antiviral Response Dictated by Choreographed Cascade of Transcription Factors. *The Journal of Immunology* 184, 2908-2917.
- Zeng, J., Fournier, P. and Schirrmacher, V. 2002. Induction of Interferon- α and Tumor Necrosis Factor-Related Apoptosis-Inducing Ligand in Human Blood Mononuclear Cells by Hemagglutinin-Neuraminidase but Not F Protein of Newcastle Disease Virus. *Virology* 297, 19-30.

- Zhang, K.J., Qian, J., Wang, S.B. and Yang, Y. 2012. Targeting Gene-Viro-Therapy with AFP driving Apoptin gene shows potent antitumor effect in hepatocarcinoma. *Journal of biomedical science* 19, 20.
- Zhang, Y., Chen, X., Gueydan, C., Han, J., 2018. Plasma membrane changes during programmed cell deaths. *Cell research* 28, 9-21.
- Zhao, H., Jaffer, T., Eguchi, S., Wang, Z. and Linkermann, A. 2015. Role of necroptosis in the pathogenesis of solid organ injury. *Cell death & disease*; 6, e1975
- Zhao, J., Han, S.X., Ma, J.L., Ying, X., Liu, P., Li, J., Wang, L., Zhang, Y., Ma, J., Zhang, L. and Zhu, Q. 2013. The role of CDK1 in apoptin-induced apoptosis in hepatocellular carcinoma cells. *Oncol Rep* 30, 253-259.
- Zhao, J., Jitkaew, S., Cai, Z., Choksi, S., Li, Q., Luo, J., Liu, Z.G., 2012. Mixed lineage kinase domain-like is a key receptor interacting protein 3 downstream component of TNF-induced necrosis. *Proceedings of the National Academy of Sciences of the United States of America* 109, 5322-5327.
- Zhu, Y., Roshal, M., Li, F., Blackett, J. and Planelles, V. 2003. Upregulation of survivin by HIV-1 Vpr. *Apoptosis : an international journal on programmed cell death* 8, 71-79.
- Zhuang, S.M., Shvarts, A., Jochemsen, A.G., van Oorschot, A.A., van der Eb, A.J. and Noteborn, M.H. 1995a. Differential sensitivity to Ad5 E1B-21kD and Bcl-2 proteins of apoptin-induced versus p53-induced apoptosis. *Carcinogenesis* 16, 2939-2944.
- Zhuang, S.M., Shvarts, A., van Ormondt, H., Jochemsen, A.G., van der Eb, A.J. and Noteborn, M.H. 1995. Apoptin, a protein derived from chicken anemia virus, induces p53-independent apoptosis in human osteosarcoma cells. *Cancer Res* 55, 486-489.
- Zimmerman, R., Peng, D.J., Lanz, H., Zhang, Y.H., Danen-Van Oorschot, A., Qu, S., Backendorf, C. and Noteborn, M. 2012. PP2A inactivation is a crucial step in triggering apoptin-induced tumor-selective cell killing. *Cell death & disease* 3, e291.
- Zitvogel, L., Kepp, O., Kroemer, G., 2010. Decoding cell death signals in inflammation and immunity. *Cell* 140, 798-804.
- Zwahlen, D. R., Bischoff, L. I., Gruber, G., Sumila, M., and Schneider, U. 2016. Estimation of second cancer risk after radiotherapy for rectal cancer: comparison of 3D conformal radiotherapy and volumetric modulated arc therapy using different high dose fractionation schemes. *Radiation Oncology*, 11(1), 149.

BIODATA OF STUDENT

Jeevanathan Kalyanasundram was born on the 19th September 1986 in Perak, Malaysia. He completed his primary education in Sekolah Kebangsaan Kerteh, Kemaman Terengganu. He pursued his secondary education in Sekolah Menengah Sains Sultan Mahmud, Kuala Terengganu with following completion of Higher Certificate of Education, HCE in Sekolah Menengah Sultan Ismail (I), Kemaman Terengganu. He was awarded Bachelor's Degree in Biotechnology by Universiti Malaysia Sabah (UMS) in 2009. He then worked as a Research Assistant in Institute for Medical Research (IMR) from the year 2009 to 2011. Inspired by biotechnology researches in IMR, he then decided to embark on research field from 2011 to 2015 and obtained Master of Science studies in the field of Genetic Engineering and Biomolecular Sciences from UPM. He then further extended his studies by pursuing Doctor of Philosophy in the field of Genetic Engineering and Biomolecular Sciences at UPM.

PUBLICATION

Academic:

Kalyanasundram, J., Hamid, A., Yusoff, K. and Chia, S-L. 2018. Newcastle disease virus strain AF2240 as an oncolytic virus: A review. *Acta Tropica* 183:126-133.

Poster:

Kalyanasundram, J., Chan, L-C., Chia, S-L., Raha, A.R. and Yusoff, K. 2017. Engineering recombinant Newcastle Disease Virus for pro-apoptotic gene delivery into tumor cells. *PUTRA Cancer Research Symposium 2017*.

Kalyanasundram, J., Chia, S-L., Raha, A.R. and Yusoff, K. 2016. Engineering Recombinant Newcastle Disease Virus rNDV (rAF2240) for Green Flourescence Protein (GFP) delivery into tumor Cells. *33rd Symposium of Malaysian Society for Microbiology*.