



**UNIVERSITI PUTRA MALAYSIA**

**ENHANCEMENT OF HYDROGEN AND METHANE PRODUCTION FROM  
CO-DIGESTION OF FOOD WASTE AND CHICKEN MANURE**

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**FBSB 2018 66**



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FROM CO-DIGESTION OF FOOD WASTE AND CHICKEN MANURE**

**By**

**MOHD FAIZ BIN MAT SAAD**


**Thesis Submitted to the School of Graduate Studies, Universiti  
Putra Malaysia, in Fulfilment of the Requirements for the Degree  
of Master of Science**

**March 2017**

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*To my parents,  
Mat Saad bin Awang &  
Hasnah binti Mohd Noor*

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

## **ENHANCEMENT OF HYDROGEN AND METHANE PRODUCTION FROM CO-DIGESTION OF FOOD WASTE AND CHICKEN MANURE**

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March 2017

**Chair : Nor'Aini Abd Rahman, PhD**  
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The demand for clean energy from renewable resources stimulates biohydrogen and biomethane production from agro-food waste as an alternative fuel to replace fossil fuel. A combined production of biohydrogen and biomethane has attracted growing attention of researchers and industries worldwide due to their potential as fuel substitute. Biosynthesis of biohydrogen and biomethane from food waste and chicken manure fermentation initiates clean technologies for energy generation thus provide the solution for waste treatment. Despite that, biogas production of hydrogen and methane have limiting factors that relate to soluble metabolites and active microorganisms. This inhibition effects can be overcome by optimizing several factors for biohydrogen and biomethane production. The objectives of this study were to determine the best ratio of food waste and chicken manure for biogas production in batch fermentation and to evaluate the effect of different inoculums and heat treatment upon selected inoculum on biohydrogen and biomethane production besides the microbial diversity in the fermentation using Next Generation Sequencing (NGS) of 16S ribosomal RNA were also carried out. The batch fermentation was conducted using 150 mL serum bottles incubated in anaerobic condition. Food waste with composition ratios of 3:1:1 of carbohydrates, protein and fiber were used as substrate added with chicken manure freshly collected from poultry farm. Biohydrogen and biomethane production were tested for the effects of different substrate ratio, different inoculums and heat treatment on selected inoculums. Temperature and initial pH were kept constant at 35°C and initial pH 7. Biohydrogen and biomethane from food waste and chicken manure was performed at different ratio (40:60, 50:50, 60:40 and 70:30 (v/v)) inoculated with aeration tank sludge (ATS), return activated sludge (RAS) and palm oil mill effluent (POME) sludge. Heat treatment was carried out at 80°C for 20 minutes to eliminate the nonsporing bacteria. Biogas was collected daily throughout 10 days fermentation and the composition of hydrogen and methane in the biogas was

analyzed by gas chromatography. The highest biogas yield obtained was 111.72 NmL/g TSS for the experiment conducted at 50:50 (v/v) substrate ratio added with RAS as inoculum without heat treatment. The highest percentages of hydrogen and methane produced were 53.35% and 52.85%, respectively. Microbial assessment was performed by using Next Generation Sequencing (NGS) of 16S ribosomal RNA technique. *Clostridium* sp. was related to biohydrogen production methanotroph such as *Cyclobacteriaceae*, *Saprosiraceae* and *Chloroflexi* that were inhibited after the heat treatment. Heat treatment of inoculums is not suitable for the production of both biohydrogen and biomethane since it inhibits the methanogens. Thus, controlling operating conditions were important for hydrogen-producing bacteria as well as methanogens for biohydrogen and biomethane production.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

**PENAMBAHBAIKAN PENGHASILAN HIDROGEN DAN METANA  
MELALUI CAMPURAN BERSAMA SISA MAKANAN DAN NAJIS AYAM**

Oleh

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Mac 2017

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**Fakulti : Bioteknologi dan Sains Biomolekul**

Permintaan terhadap tenaga tenaga bersih daripada sumber boleh diperbaharui telah menggalakkan penghasilan gas biohidrogen dan biometana sebagai alternatif kepada bahan bakar daripada bahan api fosil. Kombinasi pemprosesan biohidrogen dan biometana telah semakin menarik minat para penyelidik dan industri seluruh dunia disebabkan oleh potensinya sebagai pengganti bahan bakar. Biohidrogen dan biometana dari penapaian sisa makanan dan najis ayam merupakan teknologi bersih untuk penjaan tenaga seterusnya adalah penyelesaian untuk rawatan sisa. Namun, penghasilan hidrogen dan metana mempunyai faktor kekangan yang berkait dengan metabolit dan mikroorganisma aktif. Kesan kekangan ini boleh ditangani dengan mengoptimumkan beberapa parameter untuk penghasilan biohidrogen dan biometana. Objektif kajian ini dijalankan adalah untuk menentukan nisbah terbaik antara sisa makanan dan najis ayam untuk penghasilan biogas, untuk mengetahui kesan penggunaan inokulum berbeza dan rawatan haba ke atas inokulum terpilih untuk penghasilan biohidrogen dan biometana dan untuk mengetahui kepelbagaian mikrob di dalam fermentasi menggunakan Next Generation Sequencing (NGS) untuk 16S DNA ribosom untuk mengenalpasti dan membandingkan bakteria yang wujud di dalam sampel yang tidak diberi rawatan dengan rawatan haba. Penapaian kelompok telah dijalankan menggunakan botol serum 150 mL yang dieram dalam keadaan anaerobik. Sisa makanan yang diambil dari kafeteria dengan nisbah komposisi 3:1:1 dari karbohidrat, protein dan serat digunakan sebagai substrat dicampur dengan najis ayam yang diambil segar dari ladang ayam. Penghasilan biohidrogen dan biometana diuji untuk kesan menggunakan substrat berbeza, inokulum berbeza dan kesan rawatan haba terhadap inokulum yang dipilih. Suhu dan pH dikekalkan pada 35°C dan pH permulaan 7. Sisa makanan dan najis ayam digabungkan pada nisbah berbeza (40:60, 50:50, 60:40 and 70:30 (v/v)) dan pada tiga inokulum berbeza (enapcemar tangki pengudaraan (ATS), enapcemar diaktifkan kembali (RAS) dan

enapcemar efluen kilang kelapa sawit (POME)) digunakan sebagai inokulum. Rawatan haba dijalankan kepada inokulum yang dipilih pada 80°C selama 20 minit bagi menyingkirkan bakteria yang tidak diinginkan. Biogas dikumpulkan setiap hari selama 10 hari dan komposisi hidrogen dan metana di dalam biogas dianalisa menggunakan kromatografi gas. Penghasilan biogas tertinggi ialah 111.72 NmL/g TSS untuk eksperimen yang dijalankan pada nisbah ratio 50:50 (v/v) yang dicampur bersama RAS sebagai inokulum tanpa rawatan haba. Peratus tertinggi penghasilan biohidrogen dan biometana adalah 53.35% dan 52.85%, setiap satu. Mikroorganisma dinilai menggunakan Next Generation Sequencing (NGS) dari teknik 16S ribosomal RNA. *Clostridium* sp. berkait dengan penghasilan biohidrogen dan metanogen seperti *Cyclobacteriaceae*, *Saprosiraceae* dan *Chloroflexi* bertanggungjawab ke atas penghasilan metana. Rawatan haba didapati tidak sesuai bagi penghasilan serentak biohidrogen dan biometana kerana ianya menghalang metanogen. Justeru, kawalan keadaan semasa penghasilan biohidrogen dan biometana adalah sangat penting untuk membantu pertumbuhan bakteria penghasil hidrogen dan juga metanogen untuk penghasilan biohidrogen dan biometana.



## ACKNOWLEDGEMENTS

*In the Name of Allah, the Most Gracious, the Especially Merciful*

I would like to express my full gratitude and praise to Allah and His Messenger who has guided me along the journey of completing this thesis study. Every person and details that have come along of this work is a result of His blessings. My beloved family, especially my parents, Mat Saad Awang and Hasnah Mohd Noor that have supported me along the way with their forbearance and countless effort to get me through this overwhelming journey such as providing me advices, financial support and much more that cannot be written down in a single page. I am forever indebted to them. They deserve my Gratitude after God and His Messenger.

It is my duty and obligation to thank my supervisor, Associate Professor Dr Nor'Aini Abdul Rahman and my co-supervisor, Dr Mohd Zulkhairi Mohd Yusoff for giving and providing continuous supervision, guidance and sacrificing their time throughout my study. They have been very patient and encouraging in finishing my thesis work. I also wish to acknowledge the laboratory staff especially Mr Rosli Aslim and Madam Aluyah Marzuki for their kindness and assistance with their practical knowledge and advices throughout my study.

I also would like to express my sincere appreciation to my lab mates and friends especially Tengku Roslina Tuan Yusof and Shakir Hatim Saupi for their kind support and companionship. My friends have also helped me along the way in their own unique ways.

Finally, I take this opportunity to thank all the people that I might have missed mentioning in this section for your time and support in producing this work. Thank you to everyone for being there for me, your support will never be forgotten and shall be recompensed by God on the day of debt.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the Master of Science. The members of the Supervisory Committee were as follows:

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Chromatogram of standard mixture gas (b)  
Chromatogram of sample from GC



## LIST OF ABBREVIATIONS

ATP	Adenosine triphosphate
ATS	Aeration tank sludge
Biohydrogen	Biological hydrogen
Biomethane	Biological methane
Biohythane	Biological hydrogen and methane
BLAST	Basic local alignment search tool
BOD	Biological oxygen demand
$C_6H_{12}O_6$	Glucose
$CH_4$	Methane
$CO_2$	Carbon dioxide
COD	Chemical oxygen demand
DNA	Deoxyribonucleic acid
FISH	Fluorescent <i>in situ</i> hybridization
F/M	Food to microorganism
GC	Gas Chromatography
$H_2$	Hydrogen
$H_2SO_4$	Sulphuric acid
HCl	Hydrochloric acid
$HNO_3$	Nitric acid
HPLC	High performance liquid chromatography
MSW	Municipal solid waste
NmL	Normalize volume in mL
NADH	Nicotinamide adenine dinucleotide

NaOH	Sodium hydroxide
NCBI	National Center for Biotechnology Information
O <sub>2</sub>	Oxygen
PCR	Polymerase chain reaction
POME	Palm oil mill effluent
rRNA	Ribosomal ribonucleic acid
SD	Standard deviation
TS	Total solid
TSS	Total suspended solid
TVS	Total volatile solid
VSS	Volatile suspended solid

## LIST OF NOMENCLATURE

$C_{H,i}$	Fraction of hydrogen gas in the headspace of the bottle measured using gas chromatograph in the current (%/100)
$C_{H,i-1}$	Fraction of hydrogen gas in the headspace of the bottle measured using gas chromatograph in the previous time interval (%/100)
$e$	2.718281828
$H$	Cumulative biohydrogen produced (mL)
$P$	Biogas production potential (mL)
$R_m$	Rate of biogas production (mL/h)
$V_{G,i}$	Total biogas volume at current (mL)
$V_{G,i-1}$	Total biogas volume at previous time interval (mL)
$V_H$	The total volume of gas in the headspace of reactor (mL)
$V_{H,i}$	Cumulative biohydrogen production at current (mL)
$V_{H,i-1}$	Cumulative biohydrogen production at previous time intervals (mL)
$V_{M,i}$	Cumulative biomethane production at current (mL)
$V_{M,i-1}$	Cumulative biomethane production at previous time intervals (mL)
$\lambda$	Lag phase (h)
$t$	Fermentation time (h)

## CHAPTER 1

### INTRODUCTION

#### 1.1 Introduction

Majority of the world's energy demand today rely on fossil fuels, which are being depleted. Besides, the energy production from fossil fuels that have impacts on the environment such as greenhouse emissions, global warming, climate change and ozone layer depletion are causing an urgent need for renewable energy (Azbar & Levin, 2012). Hydrogen and methane are two major gaseous energy carriers that can address all the above concern as a viable alternate energy source.

Hydrogen is labelled as the cleanest fuel because when it burnt, it released energy and the only product left is water. Utilization of hydrogen can be described as a growing trend of the future society due to its zero-carbon emission and has two times energy yield than any hydrocarbon fuel (Cai et al., 2004). The development of society based on hydrogen is however has a drawback due to its cost-intensive in both processes and operations. By utilizing the waste materials such as food waste, treatment costs will be reduced due to the abundance and low material cost.

Methane in the other hand is used as a vehicle fuel in natural gas form. As for the chemical industry purpose, methane is usually being used as favorite feedstock for the production of methanol, acetic acid, acetic anhydride and hydrogen. Comparing methane with other fossil fuels, it is claimed as more environmentally friendly than other fossil fuels such as gasoline, petrol and diesel, it is however limited by its narrow range of flammability, high ignition temperature and slow burning speed. These weaknesses in methane are interestingly complements perfectly by the hydrogen because it increases the H/C ratio. By adding hydrogen, the narrow range of flammability of methane can be improved, in that way improving the fuel efficiency. The flame speed of methane will be greatly increased and the high ignition of methane can be reduced by the addition of hydrogen (Falco & Basile, 2015).

Various attempts have been carried out in order to use cheap and renewable sources for a fuel substitute. Substrate such as food waste (Kim et al., 2008; Pan et al., 2008) palm oil mill effluent (Ismail et al., 2010), wastewater (Yang et al., 2007), dairy manure (Amon et al., 2007) and chicken manure (Abouelenien et al., 2010) have been studied to produce biohydrogen and biomethane. The usage of carbon and nitrogen sources readily available in food waste and chicken manure for biohydrogen and biomethane production will generate less economic burden for the future by turning food waste and

chicken manure into energy source while overcome the pollution created by these waste.

One way of producing biohydrogen and biomethane is through anaerobic degradation of complex substrate. Anaerobic digestion is a series of biological process during which microorganisms break down organic matter in the absence of oxygen environment. Anaerobic digestion could be used trimmings and solid waste, with varying degrees of degradation. Through a process called methanogenesis, anaerobic digestion produce methane biogas that can be used as a renewable energy source. Production of biogas from anaerobic digestion of biomass is a technology that can produce renewable energy and also minimize the environmental risks related with manure and food waste. Co-digestion of multiple biosolid wastes, which optimize the nutrients and bacterial diversity can use those wastes of digestive process is an appealing approach for exceeding the order of biotransformation. Many effective co-fermentation processes using different substrates have shown great increase in biogas production, contrast with single digestion of the substrates.

Fermentative biogas production is a very complex process (Wang & Wan, 2009). Thus, serious consideration should be taken to control the surrounding conditions such as temperature, pH, substrate concentration and nutrient supply to achieve high biogas yield (Wang & Wan, 2009). Controlling the conditions of the fermentation is important since it can affect the rates of biogas production, product formation (organic acids, solvent, biohydrogen and biomethane), bacterial activities and growth rates, also may change metabolic pathways of hydrogenase enzyme and methanogenic bacteria (Pan et al., 2008). Fermentative biogas production from mixed culture was simple and less tendency of contamination when compared to single culture. The effectiveness of biogas production from mixed culture as inoculums have been verified by several researchers from anaerobic sludge (Pan et al., 2008; Yusoff et al., 2009) compost (Akutsu et al., 2009; Lee et al., 2008), sewage sludge (Kim et al., 2008) and cattle manure sludge (Cheong & Hansen, 2007). Anaerobic fermentation using mixed culture was suitable for biohydrogen and biomethane production as the substrate was utilized by microorganism with fast rate, simple to operate without requirement of light and oxygen supply (Chong et al., 2009; Valdez-Vazquez & Poggi-Varaldo, 2009).

More than one substrate are simultaneously consumed in the same reactor in co-digestion. If the combination of substrates are selected correctly, co-digestion will give benefits to the process due to more preferable buffering capacity, more suitable carbon – nitrogen ratio, more diverse nutrient content or dilution of inhibiting compounds which will resulting a higher biogas yields and biologically a more balance process (Chen et al., 2008; Karthikeyan & Visvanathan, 2013). In co-digestion, adjustment of pH and moisture in the reactor can also be done (Esposito et al., 2012). Total nutrients and water can be balanced by combining different substrates, e.g. wet nutrient-poor substrates combined with nutrient-rich dry matter. For efficient co-digestion,

substrate characteristics must be known (Karthikeyan & Visvanathan, 2013), e.g. substrates that contain high amount of nitrogen (e.g. chicken manure) and other inorganic nutrients should be digested with low-nitrogen and high-carbon content substrates (e.g. food waste) to minimize the inhibition of ammonium nitrogen. This kind of substrate mixture is proposed to inhibit total pH drops (Esposito et al., 2012). Combination of manures and highly biodegradable food such as food waste will form a good mixture for co-digestion (Karthikeyan & Visvanathan, 2013). Esposito et al. (2012) proved a quicker and more stable digestion process when highly biodegradable substrates and ammonia-rich substrate were co-digested. When pig manure was co-digested with kitchen waste, Kuglarz et al., (2011) observed an increment in methane production rates up to 60%.

## 1.2 Problem Statement

There are many ways that food waste can affect our environment. Food waste is food that initially is for eating being thrown away, generally at consumer or retail level. This is a huge problem in industrialized country, where a cheaper option which is throwing away is often favorable than reusing. In reality, food waste is usually avoidable. Increased generation of food waste is both a national and global problems, with an estimation of 70% from the municipal solid waste is originated from the food waste (Wang et al., 2005). Methane gas, a gas 25 times stronger than carbon dioxide at trapping heat in the atmosphere is produced when wasted food is thrown away and break down together with other organic materials (Stabnikova et al., 2006). This will lead to the releases of greenhouse gases in the environment.

Accumulations of greenhouse gases in the atmosphere are contributing to climate change and global warming worldwide. Food waste can also give a big impact on dumping ground, or landfills, and how they give impact to the surrounding environment. Degradation of food waste releases nutrients, which can drift out of landfill waste and spread out to the surrounding environment. Accumulation of nutrients can be a trouble because they can pollute groundwater and waterways. This will lead to the pollution of the water source (Ding & Wang, 2008).

A Malaysia's food production lifecycle, or supply chain can also contribute on greenhouse and affect the environment. For instance, to produce, harvest, transport, process, package, distribute and market all food products, soils, water, natural resources and energy are used. The energy and resources invested by the supply chain to deliver food to our house is lost with the food leftover (Sakai et al., 2008). One of the factor that contribute total greenhouse gas emission in Malaysia is the food supply chain. Starting from direct emissions of agricultural machinery and those attributed to energy including transportation, production of food, processing and distribution to retailers. By changing the wasteful habits for a greener approach to buy, prepare and



manage the food, significant environmental and greenhouse benefits can be achieved when all people play their parts.

While there are numerous reports and studies that drawn up to solve the food waste generation, avoidance and managements are sparse and difficult to conduct and verify. This is due to a range of factors related to the context and process of food waste generation (Sakai et al., 2008). At present, food waste is considered as common waste which is worthless and is therefore not segregate in a proper way. In addition, where kitchen waste and other organic waste are differentiated from other general wastes, they are often considered together, making it difficult to calculate the proportion of this waste that is food only. On the other hand, separating food waste physically is not practical besides health and safe risk. Due to these factors, it is difficult to manage the food waste generation and to structure the programs for food management practices. One of the wise ideas of managing these wastes is to utilize the organic waste and convert it into value-added and sustainable product such as bioenergy.

Due to the increasing demand and supply of chicken product, Malaysia is facing a serious issue in managing chicken manure. A large amount of chicken manure generated gives environmental problem such as spreading of pathogens, odorous compound and emission of greenhouse gases. In Sinar Harian Kelantan edition dated 26 December 2011, one of the villagers of Kampung Cherang, Chabang Empat, Tumpat Kelantan said that the chicken farm bring the unpleasant odours to the village for the past eight year. It gave negative effect to human health and the quality of life for people living around chicken farm areas. These are some of the examples of the serious environmental problems that need to overcome quickly by the proper treatment of chicken manure. The presence of nitrogen in chicken manure will pollute ground water and increase the level of nitrate in drinking water.

There are many factors limiting biogas production including pH, temperature, headspace volume and nature of substrate (Yusof et al., 2014). Separate biohydrogen and biomethane production has been well researched and documented in the literature. Although some researchers found that carbohydrate rich waste feedstock are appropriate substrate for biogas (hydrogen) production, there are new studies that are being explored with mixed substrate to investigate co-fermentation. Co-digestion also can deal with the limitation of source for substrate and inoculums. Biogas yield and production rate of biogas were vary even for a specific substrate relying on the inoculums.

### **1.3 Objectives of Study**

To date, information for biohydrogen and biomethane production has been studied from different kind of substrate. In this study, food waste and chicken manure were used as substrates for biohydrogen and biomethane production. However, biohydrogen is inhibited by hydrogen consuming bacteria, as well as biomethane production inhibited by several limiting factors. Thus, optimizing parameters one factor at a time are critical to eliminate the inhibition effects to enhance high yield of biogas production. The objectives of this study are :

1. To determine the best ratio of food waste and chicken manure for biogas production in batch fermentation.
2. To determine the effect of different inoculums and heat treatment upon selected inoculum on biohydrogen and biomethane production from co-digestion of food waste and chicken manure in one step fermentation.
3. To determine the microbial diversity in the fermentation using Next Generation Sequencing (NGS) of 16S ribosomal RNA to identify and compare the bacteria present in the samples with untreated and heat treated inoculum.

### **1.4 Scope of Study**

The proposed research of investigating the enhancement of biohydrogen and biomethane production from food waste and chicken manure was carried out in three stages; evaluation of mixing ratio of food waste and chicken manure, determine the effect of different source of inoculums, and assessment on heat treatment for inoculums for co-digestion of food waste and chicken manure. The details scope for each objective was stated below:

#### **1.4.1 Evaluation of Mixing Ratio of Food Waste and Chicken Manure**

The evaluation focused on the comparison of different mixing ratio of food waste and chicken manure besides determining the characteristics of the substrates that were used along the experiments. The mixing ratio of 40:60, 50:50, 60:40 and 70:30 (v/v) of food waste and chicken manure were studied. The statistical analyses were performed using MATLAB software and Microsoft Excel 2010.

#### **1.4.2 Determine the Effect of Different Inoculums**

Three types of different inoculums were tested to get the most suitable inoculums to be co-digested with the substrates. Aeration tank sludge (ATS),

return activated sludge (RAS) and palm oil mill effluent (POME) sludge were used in this study. The best mixing ratio were used to study the effect of different inoculums. The deciding factors that were used in choosing the best inoculums are based on high yield of biohydrogen and biomethane production. The statistical analyses were performed using Statistica 13.0 Software, MATLAB software and Microsoft Excel 2010.

#### **1.4.3 Evaluate the Effect of Heat Treatment on Inoculums**

The production of biohydrogen and biomethane were optimized by studying the effect of heat treatment on the selected inoculums. The inoculums were heat treated at 80°C for 20 minutes. MATLAB software and Microsoft Excel 2010 were used to perform the statistical analyses. The influencing factors of producing the high yield of biohydrogen and biomethane were also discussed.

#### **1.4.4 Assessment of Microbial Diversity**

Microbial characterization study was carried out to determine the phyla of the available microorganisms for the samples of heat treated and non-treated inoculums. The selected samples were sent to First BASE Laboratories Sdn. Bhd. for 16S ribosomal RNA (rRNA) analysis. To perform the data analysis, SILVA119, BlasyN and MEGAN5 software were used. The taxonomy and phylogeny of the samples were also discussed.

## REFERENCES

- Abbasi, T., Tauseef, S. M., & Abbasi, S. . (2012). Biogas Energy.
- Abbassi-Guendouz. (2012). *Link between operating parameters, microorganisms and performances of dry anaerobic digestion - Doctoral thesis*. University of Montpellier II, Sciences et Techniques du Languedoc.
- Abouelenien, F., Fujiwara, W., Namba, Y., Kosseva, M., Nishio, N., & Nakashimada, Y. (2010). Improved methane fermentation of chicken manure via ammonia removal by biogas recycle. *Bioresource Technology*, 101(16), 6368–73.
- Abouelenien, F., Namba, Y., Kosseva, M. R., Nishio, N., & Nakashimada, Y. (2014). Enhancement of methane production from co-digestion of chicken manure with agricultural wastes. *Bioresource Technology*, 159, 80–7.
- Agumuthu, P., Fauziah, S. H., Kindir, K. M., & Aiza, A. N. (2007). Sustainable waste management – Asian perspective. In *Proceeding of the international conference on sustainable solid waste management, Chennai, India*.
- Akassou, M., Kaanane A., Crolla, A., Kinsley C. (2010). Statistical modeling of the impact of some polyphenols on the efficiency of anaerobic digestion and the co-digestion of the wine distillery wastewater with dairy cattle manure and cheese whey *Water Sci Technol*, 62 (2010), 475–483
- Akutsu, Y., Li, Y.-Y., Harada, H., & Yu, H.-Q. (2009). Effects of temperature and substrate concentration on biological hydrogen production from starch. *International Journal of Hydrogen Energy*, 34(6), 2558–2566.
- Alexandratos, N., & Bruinsma, J. (2012). World agriculture towards 2030/2050: the 2012 revision. *ESA Working Paper No. 12-03. Rome, FAO*.
- Amon, T., Amon, B., Kryvoruchko, V., Zollitsch, W., Mayer, K., & Gruber, L. (2007). Biogas production from maize and dairy cattle manure—Influence of biomass composition on the methane yield. *Agriculture, Ecosystems & Environment*, 118(1-4), 173–182.
- Ansorge, W. J. (2009). Next-generation DNA sequencing techniques. *New Biotechnology*, 25(4), 195–203.
- APHA. (2005). *Standard Methods for the Examination of Water & Wastewater*. American Public Health Association.
- Astals, S., Ariso, M., Galí, A., & Mata-Alvarez, J. (2011). Co-digestion of pig manure and glycerine: Experimental and modelling study. *Journal of Environmental Management*, 92(4), 1091–1096.

- AVEC. (2013). Association of Poultry Processors and Poultry Trade in the EU Countries: 2013 Annual Report.
- Azbar, N., & Levin, D. B. (2012). *State of the Art and Progress in Production of Biohydrogen*. Bentham Science Publishers.
- Baghchehsaraee, B., Nakhla, G., Karamanev, D., Margaritis, A., & Reid, G. (2008). The effect of heat pretreatment temperature on fermentative hydrogen production using mixed cultures. *International Journal of Hydrogen Energy*, 33(15), 4064–4073.
- Baharuddin, A. S., Kazunori, N., Abd-Aziz, S., Tabatabaei, M., Abdul Rahman, N. A., Hassan, M. A., ... Shirai, Y. (2009). Characteristics and Microbial Succession in Co-Composting of Oil Palm Empty Fruit Bunch and Partially Treated Palm Oil Mill Effluent. *The Open Biotechnology Journal*, 3(1), 87–95.
- Banks, C. J., Zhang, Y., Jiang, Y., & Heaven, S. (2012). Trace element requirements for stable food waste digestion at elevated ammonia concentrations. *Bioresource Technology*, 104, 127–35.
- Birch, R. R., Biver, C., Campagna, R., Gledhill, W. E., Pagga, U., Steber, J., ... Bontinck, W. J. (1989). Screening of chemicals for anaerobic biodegradability. *Chemosphere*, 19(10-11), 1527–1550.
- Bouallagui, H., Touhami, Y., Ben Cheikh, R., & Hamdi, M. (2005). Bioreactor performance in anaerobic digestion of fruit and vegetable wastes. *Process Biochemistry*, 40(3-4), 989–995.
- Bruce, T., Martinez, I. B., Maia Neto, O., Vicente, A. C. P., Kruger, R. H., & Thompson, F. L. (2010). Bacterial community diversity in the Brazilian Atlantic forest soils. *Microbial Ecology*, 60(4), 840–9.
- Bujoczek, G., Oleszkiewicz, J., Sparling, R., & Cenkowski, S. (2000). High Solid Anaerobic Digestion of Chicken Manure. *Journal of Agricultural Engineering Research*, 76(1), 51–60.
- Cai, M., Liu, J., & Wei, Y. (2004). Enhanced biohydrogen production from sewage sludge with alkaline pretreatment. *Environmental Science & Technology*, 38(11), 3195–202.
- Callaghan, F. J., Wase, D. A. J., Thayanithy, K., & Forster, C. F. (2002). Continuous co-digestion of cattle slurry with fruit and vegetable wastes and chicken manure. *Biomass and Bioenergy*, 22(1), 71–77.
- Calli, B., Mertoglu, B., Inanc, B., & Yenigun, O. (2005). Effects of high free ammonia concentrations on the performances of anaerobic bioreactors. *Process Biochemistry*, 40(3-4), 1285–1292.

- Chang, R. (2005). *The Ideal Gas Equation* (Chemistry ). McGraw-Hill Education.
- Chen, T. H., Chynoweth, P., & Bljetina, R. (1990). Anaerobic digestion of municipal solid waste in a nonmixed solids concentrating digester. *Applied Biochemistry and Biotechnology*, 24-25(1), 533–544.
- Chen, Y., Cheng, J. J., & Creamer, K. S. (2008). Inhibition of anaerobic digestion process: a review. *Bioresource Technology*, 99(10), 4044–64.
- Chen, Chiu-Yue Lin, Min-Cheng Lin, C.-C. (2002). Acid–base enrichment enhances anaerobic hydrogen production process. *Applied Microbiology and Biotechnology*, 58(2), 224–228.
- Cheong, D.-Y., & Hansen, C. L. (2007). Feasibility of hydrogen production in thermophilic mixed fermentation by natural anaerobes. *Bioresource Technology*, 98(11), 2229–39.
- Chong, M.-L., Sabaratnam, V., Shirai, Y., & Hassan, M. A. (2009). Biohydrogen production from biomass and industrial wastes by dark fermentation. *International Journal of Hydrogen Energy*, 34(8), 3277–3287.
- Chua, K., Sahid, E. J. M., & Leong, Y. (2011). Sustainable Municipal Solid Waste Management and GHG Abatement in Malaysia. *Green & Energy Management*.
- Comino, E., Riggio, V.A., Rosso, M., (2012). Biogas production by anaerobic co-digestion of cattle slurry and cheese whey. *Bioresource Technology*, 114, 46–53
- Converse, J. C., Evans, G. W., Robinson, K. L., & Gibbons, W. (1980). Methane production from a large-size on-farm digester for poultry manure Livestock Waste. In *Proceedings of the Fourth International Symposium on livestock wastes*. ASAE Publication, Amarillo, Texas, USA.
- Crabtree, G. W., Dresselhaus, M. S., & Buchanan, M. V. (2004). The Hydrogen Economy. *Physics Today*, 57(December), 39–44.
- Deublein, D., & Steinhauser, A. (2010). *Biogas from Waste and Renewable Resources: An Introduction, 2nd, Revised and Expanded Edition*. Wiley.
- Ding, H., & Wang, J. (2008). Responses of the methanogenic reactor to different effluent fractions of fermentative hydrogen production in a phase-separated anaerobic digestion system. *International Journal of Hydrogen Energy*, 33(23), 6993–7005.
- Duangmanee, T., Padmasiri, S. I., Simmons, J. J., Raskin, L., & Sung, S. (2007). Hydrogen production by anaerobic microbial communities exposed to repeated heat treatments. *Water Environment Research : A*

*Research Publication of the Water Environment Federation*, 79(9), 975–83.

- Dunn, S. (2002). Hydrogen futures: toward a sustainable energy system. *International Journal of Hydrogen Energy*, 27(3), 235–264.
- Esposito, G., Frunzo, L., Panico, A., & Pirozzi, F. (2012). Enhanced biomethane production from co-digestion of different organic wastes. *Environmental Technology*, 33(22-24), 2733–40.
- Evans, G. (2001). *Biowaste and Biological Waste Treatment*. Earthscan.
- Falco, M. De, & Basile, A. (2015). *Enriched Methane: The First Step Towards the Hydrogen Economy* (Vol. 29). Springer.
- Gadhamshetty, V., Johnson, D., Nirmalakhandan, N., Smith, G., & Deng, S. (2009). Dark and acidic conditions for fermentative hydrogen production. *International Journal of Hydrogen Energy*, 34(2), 821–826.
- Ginkel, S. Van, Sung, S., & Lay, J.-J. (2001). Biohydrogen Production as a Function of pH and Substrate Concentration. *Environmental Science & Technology*, 35(24), 4726–4730.
- Han, S.-K., & Shin, H.-S. (2004). Biohydrogen production by anaerobic fermentation of food waste. *International Journal of Hydrogen Energy*, 29(6), 569–577.
- Handelsman, J. (2004). Metagenomics: application of genomics to uncultured microorganisms. *Microbiology and Molecular Biology Reviews: MMBR*, 68(4), 669–85.
- Hassan, M. A., Nawata, O., Shirai, Y., Rahman, N. A. A., Yee, P. L., Ariff, A. Bin, & Karim, M. I. A. (2002). A Proposal for Zero Emission from Palm Oil Industry Incorporating the Production of Polyhydroxyalkanoates from Palm Oil Mill Effluent. *JOURNAL OF CHEMICAL ENGINEERING OF JAPAN*, 35(1), 9–14.
- Henihan, A. M., Leahy, M. J., Leahy, J. J., Cummins, E., & Kelleher, B. P. (2003). Emissions modeling of fluidised bed co-combustion of poultry litter and peat. *Bioresource Technology*, 87(3), 289–294.
- Ho, C. C., & Tan, Y. K. (1985). Anaerobic treatment of palm oil mill effluent by tank digesters. *Journal of Chemical Technology and Biotechnology. Biotechnology*, 35(2), 155–164.
- Hwang, M. H., Jang, N. J., Hyun, S. H., & Kim, I. S. (2004). Anaerobic biohydrogen production from ethanol fermentation: the role of pH. *Journal of Biotechnology*, 111(3), 297–309.

- Ibrahim, J. S., Iortyer, H. A., & Kwaghger, A. (2009). Effect of combining piggery dung and palm oil mill effluent on biogas yield. *Niger. J. Indus. Sys. Stud.*
- Idris, A. (2009, July 1). Waste management: what is the choice: land disposal or biofuel. Universiti Putra Malaysia Press.
- Iortyer, H. A., Ibrahim, J. S., & Kwaghger, A. (2012). Effect of Mixing Ratio of Cattle and Piggery Dung on Biogas Generation, *1*(3), 162–169.
- Ismail, I., Hassan, M. A., Abdul Rahman, N. A., & Soon, C. S. (2010). Thermophilic biohydrogen production from palm oil mill effluent (POME) using suspended mixed culture. *Biomass and Bioenergy*, *34*(1), 42–47.
- Jagadabhi, S. (2011). Dissertation: New effective ways for biogas production.
- Jha, A., Li, J., Nies, L., & Zhang, L. (2013). Research advances in dry anaerobic digestion process of solid organic wastes. *African Journal of Biotechnology*. Academic Journals (Kenya).
- Jo, J. H., Lee, D. S., Park, D., & Park, J. M. (2008). Biological hydrogen production by immobilized cells of *Clostridium tyrobutyricum* JM1 isolated from a food waste treatment process. *Bioresource Technology*, *99*(14), 6666–72.
- Kaparaju, P., Buendia, I., Ellegaard, L., & Angelidakia, I. (2008). Effects of mixing on methane production during thermophilic anaerobic digestion of manure: lab-scale and pilot-scale studies. *Bioresource Technology*, *99*(11), 4919–28.
- Kaparaju, P., & Rintala, J. (2005). Anaerobic co-digestion of potato tuber and its industrial by-products with pig manure. *Resources, Conservation and Recycling*, *43*(2), 175–188.
- Karim, K., Hoffmann, R., Thomas Klasson, K., & Al-Dahhan, M. H. (2005). Anaerobic digestion of animal waste: effect of mode of mixing. *Water Research*, *39*(15), 3597–606.
- Karthikeyan, O. P., & Visvanathan, C. (2013). Bio-energy recovery from high-solid organic substrates by dry anaerobic bio-conversion processes: a review. *Reviews in Environmental Science and Bio/Technology*, *12*(3), 257–284.
- Kawagoshi, Y., Hino, N., Fujimoto, A., Nakao, M., Fujita, Y., Sugimura, S., & Furukawa, K. (2005). Effect of inoculum conditioning on hydrogen fermentation and pH effect on bacterial community relevant to hydrogen production. *Journal of Bioscience and Bioengineering*, *100*(5), 524–30.
- Kebede, A. A. M. C. E. A. (2014). Thermal And Chemical Pre-Treatments Of



Cow Dung And Poultry Litter Enhance Biogas Production In Batch Fermentation.

- Kim, D.-H., Kim, S.-H., & Shin, H.-S. (2009). Hydrogen fermentation of food waste without inoculum addition. *Enzyme and Microbial Technology*, 45(3), 181–187.
- Kim, S., Ko, I., Lee, C., & Shin, H. (2008). Start-up strategy for continuous fermentative hydrogen production: Early switchover from batch to continuous operation. *International Journal of Hydrogen Energy*, 33(5), 1532–1541.
- Kim, S.-H., Han, S.-K., & Shin, H.-S. (2004). Feasibility of biohydrogen production by anaerobic co-digestion of food waste and sewage sludge. *International Journal of Hydrogen Energy*, 29(15), 1607–1616.
- Kuglarz, M., Mrowiec, B., & Bohdziewicz, J. (2011). Influence of kitchen biowaste addition on the effectiveness of animal manure digestion in continuous condition. In *Research and application of new technologies in wastewater treatment and municipal solid waste disposal in Ukraine, Sweden and Poland*.
- Lattin, W. C., & Utgikar, V. P. (2007). Transition to hydrogen economy in the United States: A 2006 status report. *International Journal of Hydrogen Energy*, 32(15), 3230–3237.
- Lee, Y. J., Miyahara, T., & Noike, T. (2001). Effect of iron concentration on hydrogen fermentation. *Bioresource Technology*, 80(3), 227–231.
- Lee, Y.-W., & Chung, J. (2010). Bioproduction of hydrogen from food waste by pilot-scale combined hydrogen/methane fermentation. *International Journal of Hydrogen Energy*, 35(21), 11746–11755.
- Lee, Z.-K., Li, S.-L., Lin, J.-S., Wang, Y.-H., Kuo, P.-C., & Cheng, S.-S. (2008). Effect of pH in fermentation of vegetable kitchen wastes on hydrogen production under a thermophilic condition. *International Journal of Hydrogen Energy*, 33(19), 5234–5241.
- Li, Y., Park, S. Y., & Zhu, J. (2011). Solid-state anaerobic digestion for methane production from organic waste. *Renewable and Sustainable Energy Reviews*, 15(1), 821–826.
- Lin, C., Chang, C., & Hung, C. (2008). Fermentative hydrogen production from starch using natural mixed cultures. *International Journal of Hydrogen Energy*, 33(10), 2445–2453.
- Lin, C., & Lay, C. (2005). A nutrient formulation for fermentative hydrogen production using anaerobic sewage sludge microflora. *International*

*Journal of Hydrogen Energy*, 30(3), 285–292.

- Liu, G., Zhang, R., Li, X., Dong, R., & Sun, Z. (2007). Research progress in anaerobic digestion of high moisture organic solid waste, 9.
- Liu, X., Li, R., Ji, M., & Han, L. (2013). Hydrogen and methane production by co-digestion of waste activated sludge and food waste in the two-stage fermentation process: Substrate conversion and energy yield. *Bioresource Technology*, 146, 317–323.
- Liu, X. Y., Ding, H. B., Sreeramachandran, S., Stabnikova, O., & Wang, J. Y. (2008). Enhancement of food waste digestion in the hybrid anaerobic solid-liquid system. *Water Science and Technology: A Journal of the International Association on Water Pollution Research*, 57(9), 1369–73.
- Liu, Z., Li, Q., Zhang, C., Wang, L., Han, B., Li, B., ... Xing, X. (2014). Effects of operating parameters on hydrogen production from raw wet steam-exploded cornstalk and two-stage fermentation potential for biohythane production. *Biochemical Engineering Journal* 90 , 234-238.
- Logan, B. E., Oh, S.-E., Kim, I. S., & Van Ginkel, S. (2002). Biological Hydrogen Production Measured in Batch Anaerobic Respirometers. *Environmental Science & Technology*, 36(11), 2530–2535.
- Luo, Y., Zhang, H., Salerno, M., Logan, B., & Bruns, M. (2008). Organic loading rates affect composition of soil-derived bacterial communities during continuous, fermentative biohydrogen production. *International Journal of Hydrogen Energy*, 33(22), 6566–6576.
- Ma, F., Naeve, N., Wang, M., Jiang, L., Chen, R., & Zhao, S. (2010). *Hydrogen-Enriched Compressed Natural Gas as a Fuel for Engines*. (P. Potocnik, Ed.). Sciyo.
- Manyi-Loh, C. E., Mamphweli, S. N., Meyer, E. L., Okoh, A. I., Makaka, G., & Simon, M. (2013). Microbial anaerobic digestion (bio-digesters) as an approach to the decontamination of animal wastes in pollution control and the generation of renewable energy. *International Journal of Environmental Research and Public Health*, 10(9), 4390–417.
- Mardis, E. R. (2008). Next-generation DNA sequencing methods. *Annual Review of Genomics and Human Genetics*, 9, 387–402.
- Massé, D. I., Talbot, G., & Gilbert, Y. (2011). On farm biogas production: A method to reduce GHG emissions and develop more sustainable livestock operations. *Animal Feed Science and Technology*, 166-167, 436–445.
- Meijden, C. M. Van Der, Veringa, H., Vreugdenhil, B. J., Drift, a. Van Der, Zwart, R. W. R., & Smit, R. (2009). Production of bio-methane from woody

biomass, (June), 8.

Misi, S. (2001). Batch co-digestion of multi-component agro-wastes. *Bioresource Technology*, 80(1), 19–28.

Montusiewicz, A., Lebiocka, M., & Pawłowska, M. (2008). Characterization of the biomethanization process in selected waste mixtures. *Archives of Environmental Protection*, Vol. 34(no. 3), 49–61.

Mu, Y., Wang, G., & Yu, H.-Q. (2006). Response surface methodological analysis on biohydrogen production by enriched anaerobic cultures. *Enzyme and Microbial Technology*, 38(7), 905–913.

Muyzer, G., & Stams, A. J. M. (2008). The ecology and biotechnology of sulphate-reducing bacteria. *Nature Reviews. Microbiology*, 6(6), 441–54.

Naik, N., & Wung, R. (2013). The Anaerobic Digestion of Organic Municipal Solid Waste in California ., 0–23.

Nasir, I. M., Ghazi, T. I. M., & Omar, R. (2012). Anaerobic digestion technology in livestock manure treatment for biogas production. *Engineering in Life Sciences Special Issue: Biogas, Volume 12(Issue 3)*, 258–269.

Nayono, S. E., Gallert, C., & Winter, J. (2010). Co-digestion of press water and food waste in a biowaste digester for improvement of biogas production. *Bioresource Technology*, 101(18), 6998–7004.

Nielsen, A. M., Christensen, K. V., & Møller, H. B. (2013). Inline NH<sub>3</sub> removal from biogas digesters. *Biomass and Bioenergy*, 50, 10–18.

Nogales, B., Moore, E. R., Llobet-Brossa, E., Rossello-Mora, R., Amann, R., & Timmis, K. N. (2001). Combined use of 16S ribosomal DNA and 16S rRNA to study the bacterial community of polychlorinated biphenyl-polluted soil. *Applied and Environmental Microbiology*, 67(4), 1874–84.

Nwoko, C. . . , & Ogunyemi, S. (2010). Effect of Palm Oil Mill Effluent (POME) on Microbial Characteristics in a Humid Tropical Soil under Laboratory Conditions. *International Journal of Environmental Science and Development*, 1(4), 307–314.

O'Sullivan, C. A., Burrell, P. C., Clarke, W. P., & Blackall, L. L. (2005). Structure of a cellulose degrading bacterial community during anaerobic digestion. *Biotechnology and Bioengineering*, 92(7), 871–8.

Okamoto, M., Miyahara, T., Mizuno, O., & Noike, T. (2000). Biological hydrogen potential of materials characteristic of the organic fraction of municipal solid wastes. *Water Science and Technology: A Journal of the International Association on Water Pollution Research*, 41(3), 25–32.

- Othong, S., Prasertan, P., Karakashev, D., & Angelidaki, I. (2008). 16S rRNA-targeted probes for specific detection of *Thermoanaerobacterium* spp., *Thermoanaerobacterium thermosaccharolyticum*, and *Caldicellulosiruptor* spp. by fluorescent in situ hybridization in biohydrogen producing systems. *International Journal of Hydrogen Energy*, 33(21), 6082–6091.
- Pan, J., Zhang, R., Elmashad, H., Sun, H., & Ying, Y. (2008). Effect of food to microorganism ratio on biohydrogen production from food waste via anaerobic fermentation. *International Journal of Hydrogen Energy*, 33(23), 6968–6975.
- Parawira, W., Murto, M., Read, J. S., & Mattiasson, B. (2007). A study of two-stage anaerobic digestion of solid potato waste using reactors under mesophilic and thermophilic conditions. *Environmental Technology*, 28(11), 1205–16.
- Poretsky, R., Rodriguez-R, L. M., Luo, C., Tsementzi, D., & Konstantinidis, K. T. (2014). Strengths and limitations of 16S rRNA gene amplicon sequencing in revealing temporal microbial community dynamics. *PloS One*, 9(4), e93827.
- Reith, J. H., Wijffels, R. H., & Barten, H. (2003). Bio-methane and biohydrogen: status and perspectives of biological methane and hydrogen production. The Hague, The Netherlands: Dutch Biological Hydrogen Foundation - NOVEM.
- Ren, N., Li, J., Li, B., Wang, Y., & Liu, S. (2006). Biohydrogen production from molasses by anaerobic fermentation with a pilot-scale bioreactor system. *International Journal of Hydrogen Energy*, 31(15), 2147–2157.
- Romm, J. J. (2005). The Hype About Hydrogen: Fact and Fiction in the Race to Save the Climate.
- Sakai, S., Ikematsu, T., Hirai, Y., & Yoshida, H. (2008). Unit-charging programs for municipal solid waste in Japan. *Waste Management (New York, N.Y.)*, 28(12), 2815–25.
- Sakar, S., Yetilmezsoy, K., & Kocak, E. (2009). Anaerobic digestion technology in poultry and livestock waste treatment--a literature review. *Waste Management & Research: The Journal of the International Solid Wastes and Public Cleansing Association, ISWA*, 27(1), 3–18.
- Smil, V. (1999). Crop Residues: Agriculture's Largest Harvest. *BioScience*, 49(4), 299.
- Smith, L. C., Elliot, D. J., & James, A. (1996). Mixing in upflow anaerobic filters and its influence on performance and scale-up. *Water Research*, 30(12), 3061–3073.

- Stabnikova, O., Liu, X.-Y., Wang, J.-Y., & Ivanov, V. (2006). Quantification of methanogens by fluorescence in situ hybridization with oligonucleotide probe. *Applied Microbiology and Biotechnology*, 73(3), 696–702.
- Stroot, P., McMohan, K. D., Mackie, R. I., & Raskin, L. (2001). Anaerobic codigestion of municipal solid waste and biosolids under various mixing conditions—I. digester performance. *Water Research*, 35(7), 1804–1816.
- Sun, D.-L., Jiang, X., Wu, Q. L., & Zhou, N.-Y. (2013). Intragenomic heterogeneity of 16S rRNA genes causes overestimation of prokaryotic diversity. *Applied and Environmental Microbiology*, 79(19), 5962–9.
- Tabatabaei, M., Zakaria, M. R., Rahim, R. A., Wright, A. G., Shirai, Y., Abdullah, N., ... Hassan, M. A. (2009). PCR-based DGGE and FISH analysis of methanogens in an anaerobic closed digester tank for treating palm oil mill effluent. *Electronic Journal of Biotechnology*, 12(3), 0–0.
- Talia, P., Sede, S. M., Campos, E., Rorig, M., Principi, D., Tosto, D., ... Cataldi, A. (2012). Biodiversity characterization of cellulolytic bacteria present on native Chaco soil by comparison of ribosomal RNA genes. *Research in Microbiology*, 163(3), 221–32.
- Tasneem, & Abbasi, S. A. (2011). “Renewable” hydrogen: Prospects and challenges. *Renewable and Sustainable Energy Reviews*, 15(6), 3034–3040.
- Tawfik, A., Salem, A., & El-Qelish, M. (2011). Two stage anaerobic baffled reactors for bio-hydrogen production from municipal food waste. *Bioresource Technology*, 102(18), 8723–6.
- Tchobanoglous, G., Burton, F., & Stensel, H. D. (2002). *Wastewater Engineering: Treatment and Reuse*. McGraw-Hill Education.
- Thompson, R. S. (2008). Hydrogen production by anaerobic fermentation using agricultural and food processing wastes utilizing a two-stage digestion system. Master’s Thesis, Utah State University, 1-89."
- Tringe, S. G., von Mering, C., Kobayashi, A., Salamov, A. A., Chen, K., Chang, H. W., ... Rubin, E. M. (2005). Comparative metagenomics of microbial communities. *Science (New York, N.Y.)*, 308(5721), 554–7.
- Ueno, Y., Haruta, S., Ishii, M., & Igarashi, Y. (2001). Microbial community in anaerobic hydrogen-producing microflora enriched from sludge compost. *Applied Microbiology and Biotechnology*, 57(4), 555–562.
- Valdez-Vazquez, I., & Poggi-Varaldo, H. M. (2009). Hydrogen production by fermentative consortia. *Renewable and Sustainable Energy Reviews*, 13(5), 1000–1013.

- Van Ginkel, S. W., & Logan, B. (2005). Increased biological hydrogen production with reduced organic loading. *Water Research*, 39(16), 3819–26.
- Vavilin, V. A., & Angelidaki, I. (2005). Anaerobic degradation of solid material: importance of initiation centers for methanogenesis, mixing intensity, and 2D distributed model. *Biotechnology and Bioengineering*, 89(1), 113–22.
- Vedrenne, F., Béline, F., Dabert, P., & Bernet, N. (2008). The effect of incubation conditions on the laboratory measurement of the methane producing capacity of livestock wastes. *Bioresource Technology*, 99(1), 146–55.
- Venkata Mohan, S., Vijaya Bhaskar, Y., & Sarma, P. N. (2007). Biohydrogen production from chemical wastewater treatment in biofilm configured reactor operated in periodic discontinuous batch mode by selectively enriched anaerobic mixed consortia. *Water Research*, 41(12), 2652–64.
- Wang, G., Mu, Y., & Yu, H.-Q. (2005). Response surface analysis to evaluate the influence of pH, temperature and substrate concentration on the acidogenesis of sucrose-rich wastewater. *Biochemical Engineering Journal*, 23(2), 175–184.
- Wang, J., & Wan, W. (2009). Factors influencing fermentative hydrogen production: A review. *International Journal of Hydrogen Energy*, 34(2), 799–811.
- Wang, J. Y., Zhang, H., Stabnikova, O., Ang, S. S., & Tay, J. H. (2005). A hybrid anaerobic solid-liquid system for food waste digestion. *Water Science and Technology: A Journal of the International Association on Water Pollution Research*, 52(1-2), 223–8.
- Ward, A. J., Hobbs, P. J., Holliman, P. J., & Jones, D. L. (2008). Optimisation of the anaerobic digestion of agricultural resources. *Bioresource Technology*, 99(17), 7928–40.
- Webb, A. R., & Hawkes, F. R. (1985). The anaerobic digestion of poultry manure: Variation of gas yield with influent concentration and ammonium-nitrogen levels. *Agricultural Wastes*, 14(2), 135–156.
- Winter, C. (2005). Into the hydrogen energy economy? milestones. *International Journal of Hydrogen Energy*, 30(7), 681–685.
- Yaldiz, O., Sozer, S., Caglayan, N., Ertekin, C., & Kaya, D. (2011). Methane Production from Plant Wastes and Chicken Manure at Different Working Conditions of a One-stage Anaerobic Digester. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 33(19), 1802–1813.

- Yang, J. Y., Komaki, M., & Nishimura, C. (2007). Effect of overlayer thickness on hydrogen permeation of Pd60Cu40/V–15NiPd60Cu40/V–15Ni composite membranes. *International Journal of Hydrogen Energy*, 32(12), 1820–1824.
- Yasin M, N. H., Rahman, N. A., Man, H. C., Mohd Yusoff, M. Z., & Hassan, M. A. (2011). Microbial characterization of hydrogen-producing bacteria in fermented food waste at different pH values. *International Journal of Hydrogen Energy*, 36(16), 9571–9580.
- Yasin, N. H. M. (2011). *Optimisation of Physical Parameters and Microbial Community Analysis of Bio-Hydrogen Production from Food Waste*.
- Yasin, N. H. M., Mumtaz, T., Hassan, M. A., & Abd Rahman, N. (2013). Food waste and food processing waste for biohydrogen production: a review. *Journal of Environmental Management*, 130, 375–85.
- Yusof, T. R. T. (2014). *Methane Production from Co-Digestion of Poultry Manure and Food Waste*.
- Yusof, T. R. T., Man, H. C., Rahman, N. A. A., & Hafid, H. S. (2014). Optimization of Methane Gas Production From Co-digestion of Food waste and Poultry Manure Using Artificial Neural Network and Response Surface Methodology. *Journal of Agricultural Science*, 6(7), 27.
- Yusoff, M. Z. M., Hassan, M. A., Abd-Aziz, S., & Rahman, N. A. A. (2009). Start-Up of Biohydrogen Production from Palm Oil Mill Effluent under Non-Sterile Condition in 50 L Continuous Stirred Tank Reactor. *International Journal of Agricultural Research*, 4(4), 163–168.
- Zhang, M.-L., Fan, Y.-T., Xing, Y., Pan, C.-M., Zhang, G.-S., & Lay, J.-J. (2007). Enhanced biohydrogen production from cornstalk wastes with acidification pretreatment by mixed anaerobic cultures. *Biomass and Bioenergy*, 31(4), 250–254.
- Zhu, W., Graney, J., & Salvage, K. (2008). Land-Use Impact on Water Pollution: Elevated Pollutant Input and Reduced Pollutant Retention. *Journal of Contemporary Water Research & Education*, 138(1), 15–21.
- Zurawski, D., Meyer, M., & Stegmann, R. (2005). Fermentative production of biohydrogen from biowaste using digested sewage sludge as inoculums. In *Tenth International Waste Management and Landfill Symposium*.

## BIODATA OF STUDENT



Mohd Faiz bin Mat Saad was born on 29<sup>th</sup> of December 1990 in Military Hospital Terendak Camp, Malacca and is the youngest of a family of 5 siblings. He started his education in Malaysia by attending Sekolah Rendah Kebangsaan Kem Terendak (1) for the primary school requirement from 1997 to 2002. He then attended Sekolah Menengah Kebangsaan Ghafar Baba, Masjid Tanah for the lower secondary school from 2003 to 2005 before continued his upper secondary school at Sekolah Menengah Sains Muar, Johor starting from 2006 to 2007. He then continues his matriculation at Pusat Asasi Sains Universiti Malaya, Kuala Lumpur until 2008. After completing the section of his study life, he then decided to further his studies in Universiti Kuala Lumpur for a Bachelor's Degree in Biosystem Engineering Technology that took four years to complete and was awarded the degree in 2013.

During his Bachelor's Degree period, he was selected as the President of Biosystem Engineering Technology and involved in many student activities. In his final semester, he was awarded with a Dean List Award and decided to continue his study. He then pursued a Master's Degree in Environmental Technology in September 2013. He also has presented posters and attended several workshops and seminars in Malaysia.



## PUBLICATION

**Mat Saad, M. F.**, Abdul Rahman, N. A., Mohd Yusoff, M. Z. (2019). Hydrogen and Methane Production from Co-digestion of Food Waste and Chicken Manure. Polish Journal of Environmental Studies, 28(4), 2805-2814. <https://doi.org/10.15244/pjoes/83670>





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The demand for clean energy from renewable resources stimulates biohydrogen and biomethane production from agro-food waste as an alternative fuel to replace fossil fuel. A combined production of biohydrogen and biomethane has attracted growing attention of researchers and industries worldwide due to their potential as fuel substitute. Biosynthesis of biohydrogen and biomethane from food waste and chicken manure fermentation initiates clean technologies for energy generation thus provide the solution for waste treatment. Despite that, biogas production of hydrogen and methane have limiting factors that relate to soluble metabolites and active microorganisms. This inhibition effects can be overcome by optimizing several factors for biohydrogen and biomethane production. The objectives of this study were to determine the best ratio of food waste and chicken manure for biogas production in batch fermentation and to evaluate the effect of different inoculums and heat treatment upon selected inoculum on biohydrogen and biomethane production. The batch fermentation was conducted using 150 mL serum bottles incubated in anaerobic condition. Food waste with composition ratios of 3:1:1 of carbohydrates, protein and fiber were used as substrate added with chicken manure freshly collected from poultry farm. Biohydrogen and biomethane production were tested for the effects of different substrate ratio, different inoculums and heat treatment on selected inoculums. Biogas was collected daily throughout 10 days fermentation and the composition of hydrogen and methane in the biogas was analyzed by gas chromatography. Microbial assessment was performed by using Next Generation Sequencing (NGS) of 16S ribosomal RNA technique. *Clostridium* sp. was related to biohydrogen production methanotroph such as *Cyclobacteriaceae*, *Saprosiraceae* and *Chloroflexi* that were inhibited after the heat treatment. Heat treatment of inoculums is not suitable for the production of both biohydrogen and biomethane since it inhibits the methanogens. Thus, controlling operating conditions were important for hydrogen-producing bacteria as well as methanogens for biohydrogen and biomethane production.