



UNIVERSITI PUTRA MALAYSIA

**DEVELOPMENT OF DUPLEX POLYMERASE CHAIN REACTION FOR
DETECTION OF *Vibrio alginolyticus* INFECTION IN MARINE FISH**

DIYANA NADHIRAH KHAIRUL PARMAN

FP 2018 13



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DIYANA NADHIRAH KHAIRUL PAR MAN

By

Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Master of Science

October 2017

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

**DEVELOPMENT OF DUPLEX POLYMERASE CHAIN REACTION FOR
DETECTION OF *Vibrio alginolyticus* INFECTION IN MARINE FISH**

By

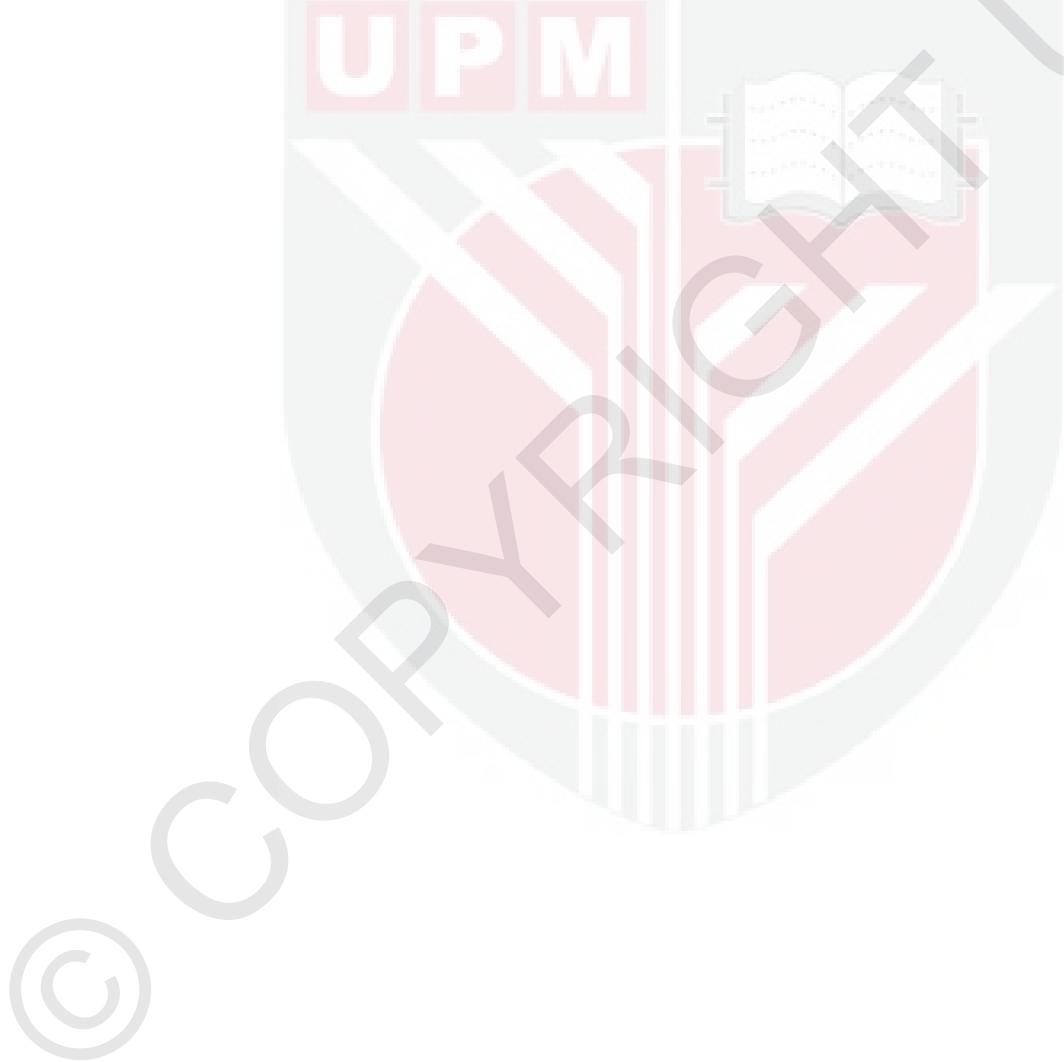
DIYANA NADHIRAH KHAIRUL PARMAN

October 2017

Chairmain : Ina Salwany Md. Yasin, PhD
Faculty : Agriculture

Vibrio alginolyticus species are among key pathogens that causes Vibriosis in a marine culture that can eventually lead to problems in food safety and bacterial disease management. In this study, an effective duplex PCR was developed for simultaneous and rapid detection species specific to *Vibrio alginolyticus* by using *gyrB* and collagenase genes. The *gyrB* and collagenase genes were amplified using purified genomic DNA of *V. alginolyticus* and other *Vibrio* species with an estimated size of 337 bp and 737 bp respectively. The *gyrB* gene was specifically detected for all *Vibrio* sp and collagenase gene was specifically detected only for *V. alginolyticus*. The PCR products were cloned and sequenced. The *Vibrio* species and non-*Vibrio* species were conducted to specificity test, and it revealed that the primer sets were specific to *V. alginolyticus* and *Vibrio* species only. The sensitivity test was accomplished with various DNA concentrations of *V. alginolyticus* (100pg to 1pg), resulting in the lowest value of 1pg. The developed duplex PCR was also tested against artificial infection of groupers that were injected with 10^{10} CFU/ml of *V. alginolyticus*, *V. parahaemolyticus*, and *A. hydrophila*. At 24 hours post-injection, the organs (liver, spleen, and kidney) were homogenized and a colony bacterium on TCBS agar was used in duplex PCR assay. All pairs of primers were shown to be specific and sensitive to *V. alginolyticus* and able to amplify collagenase and *gyrB* genes at 737 bp and 337 bp respectively. The result of artificially infected groupers showed that the duplex PCR was able to detect *V. alginolyticus*. The efficacy of this duplex PCR assay was confirmed from collected diseased groupers from the aquatic environment. It showed that duplex PCR assay was able to amplify *gyrB*

gene for only 12 samples of *Vibrio* species, however no amplification was observed for collagenase gene. Therefore, these 12 samples were sent for sequenced and ran for BLASTn analysis. Based on the analysis, only two isolates were correctly identified as *V. alginolyticus* strains (BK5G3 and BK3G1). The extracted DNA of *V. alginolyticus* strains (BK5G3 and BK3G1) were subjected to duplex PCR assay and the result showed no amplification of collagenase gene. To confirm the presence of collagenase gene, the experiment was further carried out using *Lates calcarifer* as a fish model. However, no amplification was observed for collagenase gene. Therefore, the newly developed duplex PCR assay is able to be used *in-vivo* but unable to be apply in *in-vitro* study. Newly developed duplex PCR assay is, therefore, possible to use in food security analysis and disease detection of aquatic animals.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PEMBANGUNAN TINDAK BALAS BERANTAI POLIMERASE DUPLEK
UNTUK PENGESANAN JANGKITAN *Vibrio alginolyticus* DALAM IKAN
MARIN**

Oleh

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Oktober 2017

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Spesis *Vibrio alginolyticus* adalah antara patogen utama yang menyebabkan Vibriosis dalam kultur marin yang akhirnya boleh membawa masalah kepada keselamatan makanan dan pengurusan penyakit bakteria. Dalam kajian ini, PCR duplek yang berkesan telah dibangunkan untuk pengesanan serentak dan spesifik terhadap *Vibrio alginolyticus* dengan menggunakan *gyrB* dan kolagenase gen. *GyrB* dan kolagenase gen diamplifikasi menggunakan DNA genomik yang ditulenkandari *V. alginolyticus* dan spesies *Vibrio* yang lain dengan anggaran saiz masing-masing 337 bp dan 737 bp. Gen *gyrB* secara khusus dikesan untuk semua *Vibrio* sp dan kolagenase secara khusus dikesan hanya untuk *V. alginolyticus*. Produk PCR telah diklon dan diujukan. Spesies *Vibrio* dan spesies bukan *Vibrio* telah digunakan untuk ujian spesifik, dan ianya mendedahkan bahawa set primer adalah khusus untuk spesies *V. alginolyticus* dan *Vibrio* sahaja. Ujian kepekaan dicapai dengan pelbagai kepekatan DNA *V. alginolyticus* (100pg hingga 1pg), menghasilkan nilai terendah 1pg. PCR duplek yang dibangunkan juga diuji terhadap jangkitan artifisial ikan kerapu yang disuntik dengan 10^{10} CFU / ml *V. alginolyticus*, *V. parahaemolyticus*, dan *A. hydrophila*. Pada 24 jam pasca-suntikan, organ-organ (hati, limpa, dan ginjal) telah dihomogenat dan bakteria koloni di atas TCBS agar digunakan dalam ujian PCR dupleks. Semua pasangan primers ditunjukkan khusus dan sensitif terhadap *V. alginolyticus* dan mampu mengamplifikasi gen kolagenase dan *gyrB* pada 737 bp dan 337 bp masing-masing. Keputusan dari ikan yang dijangkiti secara artifisial menunjukkan bahawa PCR duplek dapat mengesan *V.*

alginolyticus. Keberkesanan ujian PCR duplek ini telah disahkan melalui pengumpulan kerapu berpenyakit dari persekitaran akuatik. Ianya menunjukkan bahawa PCR duplek assai dapat mengamplifikasi gen *gyrB* hanya untuk 12 sampel spesies *Vibrio*, namun tiada amplifikasi diperhatikan untuk gen kolagenase. Oleh itu, 12 sampel telah dihantar untuk penjujukan dan analisis BLASTn dijalankan. Berdasarkan analisis, hanya dua isolat yang berjaya dikenal pasti sebagai strain *V. alginolyticus* (BK5G3 dan BK3G1). DNA yang diekstrak dari strain *V. alginolyticus* (BK5G3 dan BK3G1) digunakan dalam PCR duplek assai dan menunjukkan tiada amplifikasi dari gen kolagenase. Untuk mengesahkan kehadiran gen kolagenase, eksperimen ini terus dijalankan dengan menggunakan *Lates calcarifer* sebagai model ikan. Walau bagaimanapun, tiada amplifikasi dihasilkan bagi gen kolagenase. Oleh itu, ujian PCR dupleks yang baru dibangunkan ini mampu digunakan di dalam *in-vivo* tetapi tidak dapat digunakan dalam kajian *in-vitro*. Oleh itu, pembangunan PCR duplek assai yang baru ini mampu digunakan sebagai analisis keselamatan makanan dan pengesahan penyakit pada haiwan akuatik.

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This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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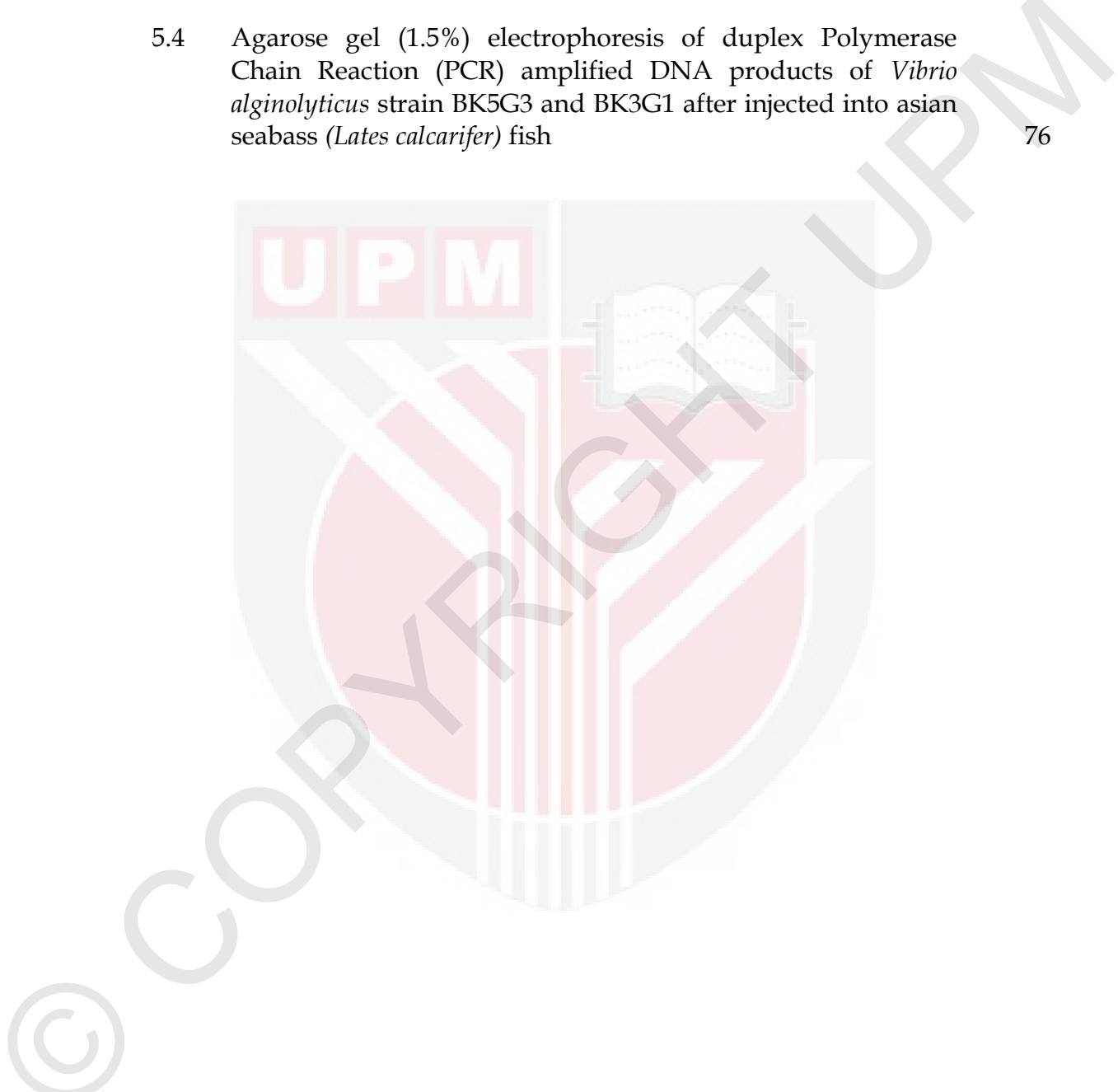
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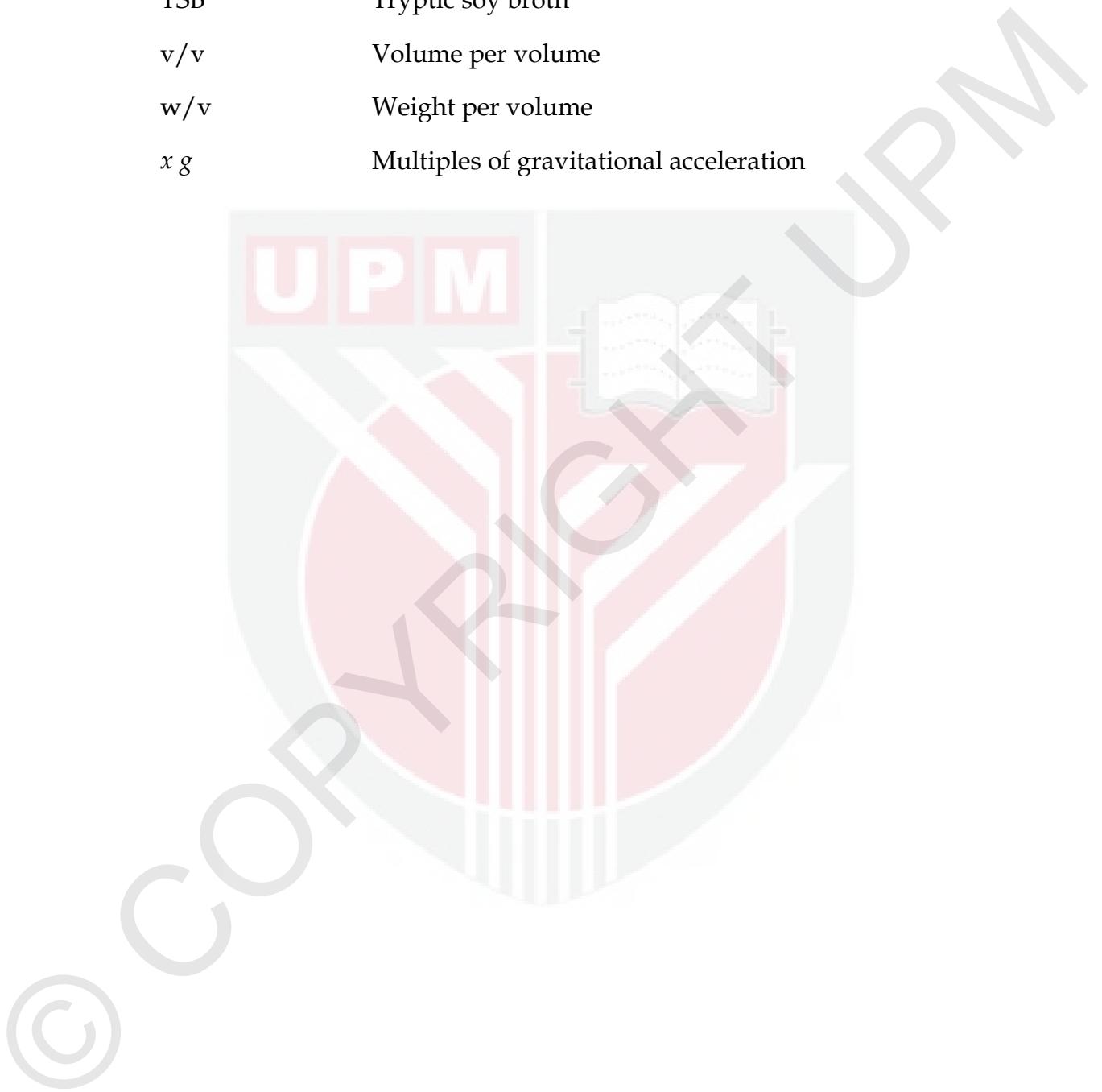
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LIST OF ABBREVIATIONS/NOTATIONS/GLOSSARY OF TERMS

AI	Auto inducer
BLAST	Basic Local Alignment Search Tool
bp	Base pair
CaCl	Calcium chloride
CFU	Colony forming units
dATP	Deoxyadenosine triphosphate
dNTP	Deoxynucleotide triphosphate
DNA	Deoxyribonucleic acid
hrs	Hours
IPTG	Isopropyl β -D-1-thiogalactopyranoside
IUCN	International Union for Conservation of Nature
LB	Luria Bertani
Min	Minute
MgCl ₂	Magnesium chloride
MgSO ₄	Magnesium sulfate
MS-222	Tricaine methanesulfonate solution
NaCl	Sodium chloride
OD	Optical density
OMP	Outer membrane protein
PBS	Phosphate buffer saline
PCR	Polymerase chain reaction
Pmol	Picomole
RNA	Ribonucleic acid
rRNA	Ribosomal ribonucleic acid
s	Second

TBE	Tris-boric EDTA
TCBS	Thiosulfate-citrate-bile salts-sucrose
TSA	Tryptic soy agar
TSB	Tryptic soy broth
v/v	Volume per volume
w/v	Weight per volume
x g	Multiples of gravitational acceleration



CHAPTER 1

INTRODUCTION

Aquaculture sector today remains a growing and important protein based food productions to cater growing human population. The production of world fish culture had expanded at an average annual rate from 6.2% in the year 2000 until 2012 (9.5% in 1990- 2000) from 32.4 million up to 66.6 million tonnes. World aquaculture can be considered into two types; land aquaculture and marine culture. The growth in land aquaculture has overtaken mariculture growth contributing to 50% its fish production in 1980 and increasing to 63% in 2012 (FAO, 2014).

Throughout the global world aquaculture, the sector has provided one-third of the total food fish supply. At the national level, the production of marine fish from Malaysian waters was 1, 482, 899 tonnes immaculate the value of RM 8.336 billion in 2013 whereas the production had falloffs by 1.67 % to 1, 458, 128 tonnes immaculate with a value of RM 8785 billion in 2014 (Dof, 2016). Index of fish consumption had increase from 53.1 kg in year 2011 and is expected to be 61.1 kg in year 2020 which placed Malaysia one of the highest fish consumers in the world (Yusoff, 2015).

Similar to other farming sectors, the occurrence of infectious disease was caused by fungi, bacteria, parasites and other prominent pathogens increased as aquaculture activities intensified and expand. In numerous countries, diseases had become one of the main constrain in cultured aquatic organism, inhibiting both economic and social development (Bondad-Reantaso et al., 2005). There are two ways in which marine fish infectious diseases can impact a species' economic values.

Decreasing potential catch is due to increase in mortality, slower growth, immune defences infested by hosts, and host reactions to infection that can harm both hosts and infectious agents. Secondly, the fish's taste was off-flavour, had bad appearance and risks to human health (Lafferty et al., 2015).

Water quality is the main factor in aquaculture system for disease management. The non-optimum water physic-chemical parameters such as dissolved oxygen, pH, salinity, ammonia, temperature and poor management practice like overfeeding, inadequate nutrition and overcrowding causes stress to fish cultured thus make them susceptible to disease outbreaks (Boyd & Tucker, 1998; Zamri-Saad et al., 2014). Disease

management aspects should be taken seriously, since intensive fish culture areas gave harmful effects towards water quality on those areas (Gorlach-Lira et al., 2013).

The fast growing marine fish such as groupers (*E. fuscoguttatus*) has become farmers' choice for their intensive aquaculture. However, warming of marine and saline inland waters is likely to augment larger amounts of *Vibrio* populations which increased the risk of infection (Roux et al., 2015). Vibriosis has caused the farmers to struggle with losses due to mass mortalities of fry and fingerlings. The common clinical signs related to Vibriosis in groupers including dark skin, pale gills, hemorrhagic around mouth and skin ulceration (Sarjito et al., 2009)

In recent times, bacterial diseases have caused the major mortalities in various fish and shellfish species over all over geographical area (Saeed, 1995; Zhang & Austin, 2000). Vibriosis is a major cause that is related to bacterial disease in mariculture system (Bondad-Reantaso et al. 2005; Zhang et al. 2007; Ponprateep et al. 2009). *Vibrio* infections are acquired through ingestion or through an open wound (Iwamoto et al., 2010). The pathogens frequently associated with mass mortality during outbreaks include *V. alginolyticus*, *V. parahaemolyticus*, *V. harveyi*, and *V. anguillarum* (Alipiah et al., 2016). *Vibrio alginolyticus* exists in marine surroundings, coastal and aquatic atmospheres (Narracci et al., 2013) with wide-reaching distribution. It is also vastly abundant and usually takes over *Vibrio* communities (Lai et al., 2004; Schets et al., 2010; Jones et al., 2013). It has been reported that *V. alginolyticus* is an associated agent with Vibriosis disease that caused mass mortality in the cultivation of yellow croakers (*Pseudosciaena crocea*) and economic losses (Yan et al., 2007). However the usage of PCR method in identifying *V. alginolyticus* is time consuming and can cause misidentification of the emergence of Vibrios species.

Molecular DNA based diagnostic assay, was based on polymerase chain reaction (PCR), have been deliberated and established for fast and accurate identification of *Vibrio* species. This protocol gives comparative benefits to match usual microbiological culture procedures (Kaysner & DePaola, 1998; Jones et al., 2012). The polymerase chain reaction (PCR) was developed and established for speedy detection of *V. alginolyticus* (Robert-Pillot et al. 2002; Liu et al. 2004). Fast and efficient method for detection of this pathogenic *V. alginolyticus* is important in controlling and investigating Vibriosis outbreaks. The aim of this project is to develop duplex PCR assay which can be used in detection of *Vibrio* and *V. alginolyticus* species in single reaction and finally to give a precautionary measures against Vibriosis diseases associated with *V. alginolyticus*. In developing duplex polymerase chain

reaction (PCR) assay for diagnosis of *V. alginolyticus* in marine aquaculture. Three objectives of study were set. The three objectives were:

1. To clone and sequence *gyrB* and collagenase genes of *Vibrio alginolyticus* strain VA2 as gene marker for assay development.
2. To develop duplex PCR assay by using *gyrB* and collagenase genes for *V. alginolyticus* detection.
3. To assess the efficacy of developed duplex PCR assay for identification of *V. alginolyticus* in naturally and artificial infected fish.

Hypotheses of this study were:

H_0 : The efficacy of new developed duplex PCR could be used and identify *V. alginolyticus* in naturally and artificial infected fish.

H_A : The efficacy of new developed duplex PCR could not be used and unable to identify *V. alginolyticus* strain in naturally and artificial infected fish.

REFERENCES

- Adams, A., & Thompson, K. D. (2006). Biotechnology offers revolution to fish health management. *Trends in Biotechnology*.24(5): 201-205.
- Aguirre-Guzman, G., Ruiz, H., & Ascencio, F. (2004).A review of extracellular virulence product of *Vibrio* species important in diseases of cultivated shrimp.*Aquaculture Research*. 35: 1395-1404.
- Aguirre-Guzmán, G., Sánchez-Martínez, J. G., Pérez-Castañeda, R., Palacios-Monzón, A., Trujillo-Rodríguez, T., La Cruz-Hernández, D. & Ivan, N. (2010). Pathogenicity and infection route of *Vibrio parahaemolyticus* in American white shrimp, *Litopenaeus vannamei*. *Journal of the World Aquaculture Society*.41(3): 464 - 470.
- Akiba, M., Kusumoto, M., & Iwata, T. (2011). Rapid identification of *Salmonella enterica* serovars, typhimurium, choleraesuis, infantis, hadar, enteritidis, dublin and gallinarum, by multiplex PCR. *Journal of Microbiological Methods*. 85(1): 9-15.
- Alipiah, N. M., Ramli, N. H. S., Low, C. F., Shamsudin, M. N., & Yusoff, F. M. (2016).Protective effects of sea cucumber surface-associated bacteria against *Vibrio harveyi* in brown-marbled grouper fingerlings.*Aquaculture Environment Interactions*. 8: 147-155.
- Altinok, I. (2011). Multiplex PCR assay for detection of four major bacterial pathogens causing rainbow trout disease. *Diseases of Aquatic Organisms*. 93:199–206
- Amal, M. N. A., Saad, M. Z., Zahrah, A. S., & Zulkafli, A. R. (2015). Water quality influences the presence of *Streptococcus agalactiae* in cage cultured red hybrid tilapia, *Oreochromis niloticus* × *Oreochromis mossambicus*. *Aquaculture Research*. 46(2), 313-323.
- Amalina, N. Z., & Ina-Salwany, M. Y. (2016). Recent advancements in molecular detection of *Vibrio* species in aquatic animals: A review. *Bioscience Biotechnology Research Communications*. 9(3), 349-356.
- Arif, M., Suprapto, H., Sulmartiwi, L., & Sudarno(2016). Bacteria associated with mass mortality of hybrid grouper *Epinephelus* sp. In East Java Province Indonesia.*International Journal of Fisheries and Aquatic Studies*. 4(6): 439-441.

- Austin, B., Austin, D. A., Blanch, A. R., Cerdá, M., Grimont, F., Grimont, P. A. D., Jofre, J., Koblavi, S., Larsen, J.L., Pedersen, K., Tianien, T., Verdonck, L., & Swings, J. (1997). A comparison of methods for the typing of fish pathogenic *Vibrio* spp. *Systematic and Applied Microbiology*. 20(1): 89-101.
- Avendaño-Herrera, R., Magariños, B., Toranzo, A.E., Beaz, R., & Romalde, J.L. (2004). Species-specific polymerase chain reaction primers sets for the diagnosis of *Tenacibaculum maritimum* infection. *Diseases of Aquatic Organisms*. 62: 75-83.
- Baehaki, A., Suhartono, M. T., Syah, D., Sitanggang, A. B., Setyahadi, S., & Meinhardt, F. (2012). Purification and characterization of collagenase from *Bacillus licheniformis* F11. 4. *African Journal of Microbiology Research*. 6(10): 2373-2379.
- Balcázar, J. L., De Blas, I., Ruiz-Zarzuela, I., Cunningham, D., Vendrell, D., & Muzquiz, J. L. (2006). The role of probiotics in aquaculture. *Veterinary Microbiology*. 114(3): 173-186.
- Balcázar, J.L., Gallo-Bueno A., Planas, M., & Pintado, J. (2010). Isolation of *Vibrio alginolyticus* and *Vibrio splendidus* from captive-bred seahorses with disease symptoms. *Antonie Van Leeuwenhoek*. 97(2):207-210.
- Balebona, M. C., Andreu, M. J., M., Bordas, A., Zorrilla, I., Morinigo, M. A., & Borrego, J. J. (1998). Pathogenicity of *Vibrio alginolyticus* for cultured gilt-head sea bream (*Sparus aurata* L.). *Applied and Environmental Microbiology*. 64: 4269-4275.
- Bauer, A. & Rørvik, L.M. (2007). A novel multiplex PCR for the identification of *Vibrio parahaemolyticus*, *Vibrio cholerae* and *Vibrio vulnificus*. *Letters in Applied Microbiology*. 45: 371-375. doi:10.1111/j.1472-765X.2007.02195.x
- Bej, A.K., Patterson, D.P., Brasher, C.W., Vickery, M.C., Jones, D.D. & Kaysner, C.A. (1999) Detection of total and hemolysin-producing *Vibrioparahaemolyticus* in shellfish using multiplex PCR amplification of *tlh*, *tdh* and *trh*. *Journal of Microbiology Methods*. 36: 215-225.
- Bergh, O.F., Nilsen, & Samuelsen, O.B. (2001). Disease, prophylaxis and treatment of *poglossus hippoglossus*: a review. *Diseases of Aquatic organisms*. 48(1):57-74.
- Bhattacharyya, N., & Hou, A. (2013). A pentaplex PCR assay for detection and characterization of *Vibriovulnificus* and *Vibrioparahaemolyticus* isolates. *Letters in Applied Microbiology*. 57(3): 233-240.

- Bondad-Reantaso, M. G., Subasinghe, R. P., Arthur, J. R., Ogawa, K., Chinabut, S., Adlard, R., Tan, Z., & Shariff, M. (2005). Disease and health management in Asian aquaculture. *Veterinary Parasitology*. 132(3): 249-272.
- Boyd, C. E., & Tucker, C. S., (1998). Pond Aquaculture Water Quality Management. Kluwer Academic Publisher, Massachusetts.
- Boyer, S.L., Johansen, J.R., Flechtner, V.R. & Howard, G.L., (2002). Phylogeny and genetic variance in terrestrial *Microcoleus* (*Cyanophyceae*) species based on sequence analysis of the 16S rRNA gene and associated 16S-23S ITS region. *Journal of Phycology*. 38:1222-1235.
- Caburlotto, G., Gennari, M., Ghidini, V., Tafi, M.C. & Lleo, M.M. (2009) Presence of T3SS2 and other virulence-related genes in *tdh*-negative *Vibrio parahaemolyticus* environmental strains isolated from marine samples in the area of the Venetian Lagoon, Italy. *FEMS Microbiology Ecology*. 70: 506-514.
- Caburlotto, G., Lleo, M.M., Gennari, M., Balboa, S., & Romalde, J.L. (2011). The use of multiple typing methods allows a more accurate molecular characterization of *Vibrio parahaemolyticus* strains isolated from the Italian Adriatic Sea. *FEMS Microbiology Ecology*. 77: 611-622. doi:10.1111/j.1574-6941.2011.01142.x
- Cai, S. H., Wu, Z. H., Jian, J. C., & Lu, Y. S. (2007a). Cloning and expression of gene encoding the thermostable direct hemolysin from *Vibrio alginolyticus* strain HY9901, the causative agent of vibriosis of crimson snapper (*Lutjanus erythopterus*). *Journal of Applied Microbiology*. 103: 289-296
- Cai, S. H., Wu, Z. H., Jian, J. C., & Lu, Y. S. (2007b). Cloning and expression of the gene encoding an extracellular alkaline serine protease from *Vibrio alginolyticus* strain HY9901, the causative agent of vibriosis in *Lutjanus erythopterus* (Bloch). *Journal of Fish Diseases*. 30: 493-500.
- Cai, S. H., Lu, Y. S., Wu, Z. H., Jian, J. C. & Huang, Y. C. (2009). A novel multiplex PCR method for detecting virulent strains of *Vibrio alginolyticus*. *Aquaculture Research*. 41: 27-4134.
- Caipang, C. M. A. & Aguana, M. P. N. (2011). Conventional PCR assays for the detection of pathogenic *Vibrio* spp. in shrimp aquaculture in the Philippines. *AACL Bioflux*. 4(3):339-350.

- Castro, N., Toranzo, A. E., & Magariños, B. (2014).A multiplex PCR for the simultaneous detection of *Tenacibaculum maritimum* and *Edwardsiella tarda* in aquaculture.*International Microbiology*. 17: 111-117.
- Castroverde, C. D. M., San Luis, B. B., Monsalud, R. G., & Hedreyda, C. T. (2006). Differential detection of vibrios pathogenic to shrimp by multiplex PCR.*Journal of General and Applied Microbiology*. 52: 273-280
- Cazorla, C., Guigon, A., Noel, M., Quilici, M.L. & Lacassine, F. (2011). Fatal *Vibrio vulnificus* Infections associated with Eating Raw Oysters, New Caledonia. *Emerging Infectious Diseases*. 17: 136-137
- Ch'ng, C.L. & Senoo, S. (2008). Egg and larval development of a new hybrid grouper, tiger grouper *Epinephelus fuscoguttatus* × giant grouper *E. lanceolatus*.*Aquaculture Science*.56(4): 505-515
- Chang, C.-I., Hung, P.-H., Wu, C.-C., Cheng, T.C., & Tsai, J.-M.(2012). Simultaneous Detection of Multiple Fish Pathogens Using a Naked-Eye Readable DNA Microarray.*Sensors*. 12: 2710-2728
- Cheah, Y. K., Noorzaleha, A. S., Lee, L. H., Radu, S., Sukardi, S., & Sim, J. H. (2008). Comparison of PCR fingerprinting techniques for the discrimination of *Salmonella enterica* subsp. *enterica* serovar Weltevreden isolated from indigenous vegetables in Malaysia. *World Journal of Microbiology and Biotechnology*. 24, 327-335. doi:10.1007/s11274-007-9474-8
- Chen, F. R., Liu, P. C., & Lee, K. K. (2000). Lethal attribute of serine protease secreted by *Vibrio alginolyticus* strains in Kurama Prawn *Penaeus japonicus*. *Zeitschrift für Naturforschung*. 55(1-2):94-99
- Chen, X. G., Wu, S. Q., Shi, C. B., & Li, N. Q. (2004). Isolation and identification of pathogenetic *Vibrio Harveyi* from estuary cod *Epinephelus coioides*. *Journal of Fishery Sciences of China/Zhongguo Shuichan Kexue*. 11(4): 313-317
- Chen, H. M. & Lin, C. W. (2007).Hydrogel-coated streptavidin piezoelectric biosensors and applications to selective detection of Strep-Tag displaying cells.*Biotechnology Progress*. 23: 741-748
- Chen, C., Villet, R., Jacoby, G. A., & Hooper, D. C. (2015).Functions of a GyrBA Fusion Protein and Its Interaction with QnrB and Quinolones.*Antimicrobial Agents and Chemotherapy*.59(11): 7124-7127.

- Chun, J., Huq, A.&Colwell, R. R., (1999).Analysis of 16S-23S rRNA intergenic spacer regions of *Vibrio cholera* and *Vibrio mimicus*.*Applied and Environmental Microbiology*.65: 2202-2208.
- Colakoglu, F.A., Sarmasik, A. & Koseoglu, B. (2006). Occurrence of *Vibrio* spp. and *Aeromonas* spp. in shellfish harvested off Dardanelles of Turkey. *Food Control*. 17:648-652
- Croci, L., Suffredini, E., Cozzi, L., Toti, L., Ottaviani, D., Pruzzo, C., Serratore, P., Fischetti, R., Goffredo, E., Loffredo, G. & Mioni, R. (2007). Comparison of different biochemical and molecular methods for the identification of *Vibrio parahaemolyticus*.*Journal of Applied Microbiology*. 102: 229-237.
- Cunningham, C.O. (2004). Use of molecular diagnostic tests in disease control: making the leap from laboratory to field application. In Molecular Aspects of Fish and Marine Biology: Current Trends in the Study of Bacterial and Viral Fish and Shrimp Diseases (Vol. 3) (Leung, K-Y., ed.), pp. 292-312, *World Scientific Publishing Co*.
- De Andrade, T. P. (2011). Novel and emerging trends in the application of diagnostic technologies, epidemiology and disease exclusion in the aquaculture food—Producing industry. *World Aquaculture*.42(2): 58.
- De Silva, S.S., & Soto, D. (2009). Climate change and aquaculture: potential impacts, adaptation and mitigation. *FAO Fisheries Technical Paper*. 530: 137-215.
- De, M., Ghaffar, M.A., Bakar, Y., Zaidi, C., Cob, C., & Das, S.K. (2016). Optimum temperature for the growth form of tiger grouper (*Epinephelusfuscoguttatus*♀)× giant grouper (*E. Lanceolatus*♂) hybrid. *Sains Malaysiana*. 45(4): 541-549.
- DeChaine, E.G.; Bates, A.E.; Shank, T.M. & Cavanaugh, C.M. (2006). Off-axis symbiosis found: Characterization and biogeography of bacterial symbionts of *Bathymodiolus* mussels from Lost City hydrothermal vents. *Environmental Microbiology*. 8: 1902-1912
- Del Cerro, A., Marquez, I., & Guijarro, J. A. (2002).Simultaneous detection of *Aeromonas salmonicida*, *Flavobacterium psychrophilum*, and *Yersinia ruckeri*, three major fish pathogens, by multiplex PCR.*Applied and Environmental Microbiology*. 68(10): 5177-5180.

de-la-Re-Vega, E.; García-Galaz, A.; Díaz-Cinco, M.E. and Sotelo-Mundo, R.R. (2006). White shrimp (*Litopenaeus vannamei*) recombinant lysozyme has antibacterial activity against Gram negative bacteria: *Vibrio alginolyticus*, *Vibrio parahemolyticus* and *Vibrio Cholerae*. *Fish Shellfish Immunology*. 20:405-408

Demarta, A., Respinis, S., Dolina, M., & Peduzzi. R. (2004) Molecular typing of *Yersinia frederiksenii* strains by means of 16S rDNA and *gyrB* genes sequence analyses. *FEMS Microbiol Letter*. 238: 423-428

Department of Fisheries (2016). Senario industri perikanan malaysia. Retrieved December 23, 2016, from <http://www.dof.gov.my/index.php/pages/view/42>

Deshmukh, R. A., Joshi, K., Bhand, S., & Roy, U. (2016). Recent developments in detection and enumeration of waterborne bacteria: a retrospective mini review. *Microbiology Open*. 5(6): 901-922.

Di Pinto, A., Ciccarese, G., Tantillo, G., Catalano, D. & Forte, V.T.(2005). A collagenase-targeted multiplex PCR assay for identification of *Vibrio alginolyticus*, *Vibrio cholerae*, and *Vibrio parahaemolyticus*. *Journal of Food Protection*.68:150-153

Dick, M. H., Guillerm, M., Moussy, F., & Chaignat, C. L. (2012). Review of two decades of cholera diagnostics-how far have we really come?. *PLoS Negl Trop Dis*. 6(10): e1845.

DiMeo, C. A.; Wilbur, A. E.; Holben, W. E.; Feldman, R., Vrijenhoek, R. C. & Cary, S. C. (2000) Genetic variation among endosymbionts of widely distributed *vestimentiferan* tubeworms. *Applied and Environmental Microbiology*. 66: 651-658.

Drancourt, M.; Bollet, C.; Carlioz, A.; Martelin, R.; Gayral, J.P. and Raoult, D. (2000).16S ribosomal DNA sequence analysis of a large collection of environmental and clinical unidentifiable bacterial isolates. *Journal of Clinical Microbiology*. 38(10): 3623-3630.

Ebrahimzadeh Mousavi, H. A.; Akhondzadeh Basti, A.; Mirzargar S. S.; Soltani, M.; Taheri Mirghaed, A.; Esmaeili, H.; Firouzbakhsh, F.(2011). *Vibrio parahaemolyticus* in cultured shrimps and their environment in South Iran. *International Journal of Veterinary Research*. 5(3):149-150.

Egidius, E. (1987). Vibriosis: pathogenicity and pathology. A review. *Aquaculture*. 67(1-2): 15-28.

Engelstad, M. (2004). Vaccination and consumer perception of seafood quality. *Developments in Biological*. 121: 245-254.

Feng, J. M. (2008) Detection of *V. alginolyticus* by PCR and identification of virulent strains by SSCP. Thesis of Master Degree. Guangdong Ocean University, Zhanjiang, China.

Food and Agriculture Organization, FAO.(2014). Opportunities and challenges 2014. *Food and Agriculture Organization of the United Nation. Rome*. 3-30.

Francis-Floyd, R. (2005). Introduction to Fish Health Management, CIR 921: Institute of INT'L. *Journal of Agriculture and Rural Development Food Agriculture Science*. University of Florida, USA.

Frans I., Lievens B., Heusdens C. & Willems K.A. (2008) Detection and identification of fish pathogens: what is the future? *The Israeli Journal of Aquaculture*. 60: 213-229.

Fricker, M., Messelhäuser, U., Busch, U., Scherer, S., & Ehling-Schulz, M. (2007). Diagnostic real-time PCR assays for the detection of emetic *Bacillus cereus* strains in foods and recent food-borne outbreaks. *Applied Environmental Microbiology*. 73: 1892-1898. doi: 10.1128/AEM.02219-06.

Gargouti, A. S., Ab-Rashid, M. N. K., Ghazali, M. F., Mitsuaki, N., Haresh, K. K., & Radu, S. (2015). Detection of *tdh* and *trh* Toxic Genes in *Vibrio alginolyticus* Strain from Mantis Shrimp (*Oratosquilla Oratoria*). *Journal of Nutrition & Food Sciences*. 5(5): 1

Gennari, M., Ghidini, V., Caburlotto, G., & Lleo, M.M. (2012). Virulence genes and pathogenicity islands in environmental *Vibrio* strains nonpathogenic to humans. *FEMS Microbiology Ecology*. 82(3): 563-573.

George, M. R., John, K. R., Iyappan, T., & Jeyaseelan, M. J. P. (2005). Genetic heterogeneity among *Vibrio alginolyticus* isolated from shrimp farms by PCR fingerprinting. *Letters in Applied Microbiology*. 40(5): 369-372.

Germini, A., Masola, A., Carnevali, P., & Marchelli, R. (2009). Simultaneous detection of *Escherichiacoli* O175: H7, *Salmonella* spp., and *Listeriamonocytogenes* by multiplex PCR. *Food Control*. 20(8): 733-738.

- Gomez-Leon, J.; Villamil, L.; Lemos.M.L.; Novoa, B. & Figueras, A. (2005). Isolation of *Vibrioalginolyticus* and *Vibriospandidus* from aquacultured carpet shell clam (*Ruditapesdecussates*) larvae associated with mass mortalities. *Applied Environmental Microbiology*. 71(1):98-104
- Gonzalez, S.F., Osorio, C.R. & Santos, Y. (2003). Development of a PCR-based method for the detection of *Listonella anguillarum* in fish tissue and blood samples. *Diseases of Aquatic Organism*. 55: 109–115.
- Gonzalez, S.F., Krug, M.J., Nielsen, M.E., Santos, Y. & Call, D.R. (2004) Simultaneous detection of marine fish pathogens by using multiplex PCR and a DNA microarray. *Journal of Clinical Microbiology*. 42: 1414-1419.doi: 10.1128/JCM.42.4.1414-1419.2004
- González-Escalona, N., Blackstone, G. M., & DePaola, A. (2006). Characterization of a *Vibrio alginolyticus* strain, isolated from Alaskan oysters, carrying a hemolysin gene similar to the thermostable direct hemolysin-related hemolysin gene (*trh*) of *Vibrio parahaemolyticus*. *Applied and Environmental Microbiology*. 72(12): 7925-7929.
- Gorlach-Lira, K., Pacheco, C., Carvalho, L. C. T., Melo Júnior, H. N., & Crispim, M. C. (2013). The influence of fish culture in floating net cages on microbial indicators of water quality. *Brazilian Journal of Biology*: 73(3): 457-463.
- Gu, D., Liu, H., Yang, Z., Zhang, Y., & Wang, Q. (2016). Chromatin immunoprecipitation sequencing technology reveals global regulatory roles of low-cell-density quorum-sensing regulator *AphA* in the pathogen *Vibrioalginolyticus*. *Journal of Bacteriology*. 7:2985-2999.
- Gubala, A. J. (2006). Multiplex real-time PCR detection of *Vibrio cholerae*. *Journal of Microbiological Methods*. 65: 278–293.
- Haldar, S., Neogi, S. B., Kogure, K., Chatterjee, S., Chowdhury, N., Hinenoya, A., Asakura, M., & Yamasaki, S., (2010). Development of a haemolysin gene-based multiplex PCR for simultaneous detection of *Vibriocampbellii*, *Vibrio Harveyi* and *Vibrio parahaemolyticus*. *Letters of Applied Microbiology*. 50:146-152.
- Hameed, S., Solapure, S., Mukherjee, K., Nandi, V., Waterson, D., Shandil, R., & Ghosh, A. (2014). Optimization of pyrrolamides as mycobacterial GyrB ATPase inhibitors: structure-activity relationship and in vivo efficacy in a mouse model of tuberculosis. *Antimicrobial Agents and Chemotherapy*.58(1), 61-70.

- Harayama, S. & Yamamoto, S., (1996). Phylogenetic Identification of *Pseudomonas* Strains Based on Comparison of *gyrB* and *rpoD* Sequences, In Molecular Biology of Pseudomonads, edited by T. Nakazawa, K. Furukawa, D. Haas, S. Silver, ASM Press, Washington, D.C. 250–258.
- Hasegawa, H., & Häse, C. C. (2009). The extracellular metalloprotease of *Vibrio tubiashii* directly inhibits its extracellular haemolysin. *Microbiology*. 155(7): 2296-2305.
- Hasegawa, H., Lind, E. J., Boin, M. A., & Häse, C. C. (2008). The extracellular metalloprotease of *Vibrio tubiashii* is a major virulence factor for pacific oyster (*Crassostrea gigas*) larvae. *Applied And Environmental Microbiology*. 74(13): 4101-4110.
- Hassan, A. A., Khan, I. U., Abdulmawjood, A., & Lämmle, C. (2003). Inter- and intraspecies variations of the 16S-23S rDNA intergenic spacer region of various streptococcal species. *Systematic and Applied Microbiology*. 26(1): 97-103.
- Heithoff, D.M., Conner, C.P. & Mahan, M.J. (1997).Dissecting the biology of a pathogen during infection.*Trends in Microbiology*.5: 509–513. doi: 10.1016/S0966-842X(97)01153-0.
- Hendiksen, R. S., Price, L. B., Schupp, J. M., Gillece, J. D., Kaas, R. S., Engelthaller, D. M., Bortololia, V., Pearson, T., Wters, A. E., Updhyay, B. P., Shrestha, S. D., Adhikari, S., Shakya, D., Keim, P. S. & Aarestrup, F. M. (2011). Population Genetics of *Vibrio cholera* from Nepal in 2010: Evidence on the Origin of the Haitian Outbreak. *MBio*. 2(4):e00157-11.
- Henke, J. M., & Bassler, B. L. (2004).Bacterial social engagements.*Trends in Cell Biology*.14(11): 648-656.
- Hill, B. J. (2004). The need for effective disease control in international aquaculture.*Developments in Biological*.121: 3-12.
- Hjeltnes, B. & Roberts, R.J. (1993).*Vibriosis :Bacterial diseases of fish*. 109-121. United Kingdom: Blackwell Scientific Publication.
- Hong, G. E., Kim, D. G., Bae, J. Y., Ahn, S. H., Bai, S. C. and Kong, I. S. (2007). PCR detection of the fish pathogen, *Vibrio anguillarum*, using the *amiB* gene, which encodes N-acetylmuramoyl-L-alanine amidase.*FEMS Microbiology Letters*. 269: 201-206. doi: 10.1111/j.1574-6968.2006.00618.x

- Hou, X. L., Cao, Q. Y., Pan, J. C., & Chen, Z. (2006). Classification and identification of *vibrio cholerae* and *vibrio parahaemolyticus* isolates based on *gyrB* gene phylogenetic analysis. *Wei Sheng Wu Xue Bao= Acta Microbiologica Sinica*. 46(6): 884-889.
- Huang, W. M. (1996). Bacterial diversity based on type II DNA topoisomerase genes. *Annual Review of Genetics*. 30: 79–107.
- Huang, X., & Madan, A. (1999). CAP3: A DNA Sequence Assembly Program. *Genome Research*. 9(9): 868–877.
- Hurtado, L. A.; Mateos, M.; Lutz, R. A. and Vrijenhoek, R. C. (2003). Coupling of bacterial endosymbiont and host mitochondrial genomes in the hydrothermal vent clam *Calyptogenamagnifica*. *Applied and Environmental Microbiology*. 69:2058-2064.
- Huss, H.H., Ababouch, L. & Gram, L. (2003). Assessment and management of seafood safety and quality. FAO Fisheries Technical Paper No. 444. Rome, FAO. 230 pp. (available at: <ftp://ftp.fao.org/docrep/fao/006/y4743e/y4743e00.pdf>).
- INFOFISH International.(2012). *Malaysia's Seafood Industry*.1: 57-59.
- IUCN.(2014). IUCN Red List of Threatened Species. Version 2014. 1. IUCN 2014. IUCN Red List of Threatened Species.
- Iwamoto, M., Ayers, T., Mahon, B. E., & Swerdlow, D. L. (2010). Epidemiology of seafood-associated infections in the United States. *Clinical Microbiology Reviews*. 23(2): 399-411.
- Izumi, S., Yamamoto, M., Suzuki, K., Shimizu, A., & Aranishi, F. (2007). Identification and detection of *Pseudomonas plecoglossicida* isolates with PCR primers targeting the *gyrB* region. *Journal of Fish Diseases*.30(7): 391-397.
- Izumi, S., Fujii, H., & Aranishi, F. (2005). Detection and identification of *Flavobacterium psychrophilum* from gill washings and benthic diatoms by PCR-based sequencing analysis. *Journal of Fish Diseases*. 28: 559–564.
- Izumi, S., Yamamoto, M., Suzuki, K., & Shimizu, A. (2007) Identification and detection of *Pseudomonas plecoglossicida* isolates with PCR primers targeting the *gyrB* region. *Journal of Fish Diseases*. 30: 391–397.

- Izumiya, H., Matsumoto, K., Yahiro, S., Lee, J., Morita, M., Yamamoto, S., Arakawa, E., & Ohnishi, M. (2011). Multiplex PCR assay for identification of three major pathogenic *Vibrio* spp., *Vibrio cholerae*, *Vibrio parahaemolyticus*, and *Vibrio vulnificus*. *Molecular and Cellular Probes*. 25(4): 174-176.
- Jackson, C. R., Fedorka-Cray, P. J., & Barrett, J. B. (2004). Use of a genus-and species-specific multiplex PCR for identification of enterococci. *Journal of Clinical Microbiology*. 42(8): 3558-3565.
- Janda, J. M., & Abbott, S. L. (2010). The genus *Aeromonas*: taxonomy, pathogenicity, and infection. *Clinical Microbiology Reviews*. 23: 35-73. 2.
- Jeyasekaran, G., Raj, K. T., Shakila, R. J., Thangarani, A. J., & Sukumar, D. (2011). Multiplex polymerase chain reaction-based assay for the specific detection of toxin-producing *Vibrio cholerae* in fish and fishery products. *Applied Microbiology and Biotechnology*. 90(3): 1111-1118.
- Jofre, A., Martin, B., Garriga, M., Hugas, M., Pla, M., Rodríguez-Lázaro, D., & Aymerich, T. (2005). Simultaneous detection of *Listeria monocytogenes* and *Salmonella* by multiplex PCR in cooked ham. *Food Microbiology*. 22(1): 109-115. doi.org/10.1016/j.fm.2004.04.009.
- Jones, E. H., Feldman, K. A., Palmer, A., Butler, E., Blythe, D., & Mitchell, C. S. (2013). *Vibrio* infections and surveillance in Maryland, 2002-2008. *Public Health Reports*. 128: 537-545.
- Jones, J. L., Hara-Kudo, Y., Krantz, J. A., Benner, R. A., Smith, A. B., & Dambaugh, T. R. (2012). Comparison of molecular detection methods for *Vibrio parahaemolyticus* and *Vibrio vulnificus*. *Food Microbiology*. 30:105-11.
- Kakinuma, K., Fukushima, M., & Kawaguchi, R. (2003). Detection and identification of *Escherichia coli*, *Shigella*, and *Salmonella* by microarrays using the *gyrB* gene. *Biotechnology and Bioengineering*. 83(6): 721-728.
- Kang, Y., Takeda, K., Yazawa, K., & Mikami, Y. (2009). Phylogenetic studies of *Gordonia* species based on *gyrB* and *secA1* gene analyses. *Mycopathologia*. 167(2): 95-105.
- Kasai, H., Watanabe, K., Gasteiger, E., Bairoch, A.M., Isono, K., Yamamoto, S., & Harayama, S. (1998). Construction of the *gyrB* database for the identification and classification of bacteria. *Genomics and Informatics*. 9: 13-21.

- Kawase, T., Miyoshi, S., Sultan, Z. & Shinoda, S. (2004). Regulation system for protease production in *Vibrio vulnificus*. *FEMS Microbiology Letters*. 240: 55–59. doi: 10.1016/j.femsle.2004.09.023
- Keysner, C. A, & DePaola, A. (2001). *Vibrio*. In Compendium of methods for the microbiological examination of foods, 4th ed. New York: American Public Health Association.
- Khamesipour, F. (2014). Detection of *Vibrio* spp. in shrimp from aquaculture sites in Iran using polymerase chain reaction (PCR). *AACL Bioflux*. 7(1).
- Khan, I. U. H., & Edge, T. A. (2007). Development of a novel triplex PCR assay for the detection and differentiation of thermophilic species of *Campylobacter* using 16S-23S rDNA internal transcribed spacer (ITS) region. *Journal of Applied Microbiology*. 103(6): 2561-2569.
- Khoo, C. H., Cheah, Y. K., Lee, L. H., Sim, J. H., Noorzaleha, A. S., & Sidik, M. S. (2009). Virulotyping of *Salmonella enterica* subsp. *enterica* isolated from indigenous vegetables and poultry meat in Malaysia using multiplex-PCR. *Antonie Van Leeuwenhoek*. 96: 441–457. doi: 10.1007/s10482-009-9358-z
- Kim, D. G., Bae, J. Y., Hong, G. E., Min, M. K., Kim, J. K. and Kong, I. S. (2008). Application of the *rpoS* gene for the detection of *Vibrio anguillarum* in flounder and prawn by polymerase chain reaction. *Journal of Fish Diseases*. 31(9): 639-647.
- Kim, H. J., Ryu, J. O., Lee, S. Y., Kim, E. S., & Kim, H. Y. (2015). Multiplex PCR for detection of the *Vibrio* genus and five pathogenic *Vibrio* species with primer sets designed using comparative genomics. *BMC Microbiology*. 15(1): 239.
- Kim, J. S.; Lee, G. G.; Park, J. S.; Jung, Y. H.; Kwak, H. S.; & Kim, S. B. (2007). A novel multiplex PCR assay for rapid and simultaneous detection of five pathogenic bacteria: *Escherichia coli* O157:H7, *Salmonella*, *Staphylococcus aureus*, *Listeriamonocytogenes* and *Vibrio parahaemolyticus*. *Journal of Food Protection*. 70(7):1656–1662.
- Kim, S. K., Yang, J. Y. & Cha, J. (2002). Cloning and sequence analysis of a novel metalloprotease gene from *Vibrio parahaemolyticus* 04. *Gene*. 283: 277–286. doi: 10.1016/S0378-1119(01)00882-4

Kim, S.Y., Lee, S.E., Kim, Y.R., Kim, C.M., Ryu, P.Y., Choy, H.E., Chung, S.S. & Rhee, J.H. (2003). Regulation of *Vibrio vulnificus* virulence by the LuxS quorum-sensing system. *Molecular Microbiology*. 8: 1647-1664. doi: 10.1046/j.1365-2958.2003.03536.x

Kirby, B. M., Everest, G. J., & Meyers, P. R. (2010). Phylogenetic analysis of the genus *Kribbella* based on the *gyrB* gene: proposal of a *gyrB*-sequence threshold for species delineation in the genus *Kribbella*. *Antonie Leeuwenhoek*. 97(2): 131-142.

Kirkup, B.C., Chang, L., Chang, S., Gevers, D., & Polz, M.F. (2010). *Vibrio* chromosomes share common history. *BMC Microbiology*. 10:137. doi: 10.1186/1471-2180-10-137

Kong, R.Y.C., Lee, S.K.Y., Law, T.W.F., Law, S.H.W. & Wu, R.S.S. (2002). Rapid detection of six types of bacterial pathogens in marine waters by multiplex PCR. *Water Research*. 36(11): 2802-2812.

Kumar, H. S., Parvathi, A., Karunasagar, I., & Karunasagar, I. (2006). A *gyrB*-based PCR for the detection of *Vibrio vulnificus* and its application for direct detection of this pathogen in oyster enrichment broths. *International Journal of Food Microbiology*. 111(3): 216-220.

Kumar V., Roy S., Barman, D. & Kumar, A. (2014). Immunoserological and molecular techniques used in fish disease diagnosis- A mini review. *International Journal of Fisheries and Aquatic Studies*. 1: 111-117.

Kustusch, R., Kuehl, C. & Crosa, J. (2011) Power plays: iron transport and energy transduction in pathogenic vibrios. *Biometals*. 24:559-566

La Duc, M.T., Satomi, M., Agata, N., & Venkateswaran, K. (2004). *gyrB* as a phylogenetic discriminator for members of the *Bacillusanthracis-cereus-thuringiensis* group. *Journal of Microbiological Methods*. 56:383-394

Lafferty, K. D., Harvell, C. D., Conrad, J. M., Friedman, C. S., Kent, M. L., Kuris, A. M., Powell E.N., Rondeau, D. & Saksida, S. M. (2015). Infectious diseases affect marine fisheries and aquaculture economics. *Annual Review of Marine Science*. 7: 471-496.

Lafisca, A., Pereira, C. S., Giaccone, V., & Rodrigues, D. D. P. (2008). Enzymatic characterization of *Vibrio alginolyticus* strains isolated from bivalves harvested at Venice Lagoon (Italy) and Guanabara Bay (Brazil). *Revista do Instituto de Medicina Tropical de São Paulo*. 50(4): 199-202.

- Lai, F. C., Wang, Q., Zhou, Y. P., Mu, C. H., Geng, S. N., Zhang, Y. M., et al. (2004). Investigation of the bacteria in the seawater of Xisha in the South China Sea and their antibiotic sensitivity profile. *Di. Yi. Jun. Yi. Da. Xue. Xue.Bao.* 24. 347-348
- Lavilla-Pitogo, C.R., Leano, E.M. & Paner, M.G. (1998). Mortalities of pond-cultured juvenile shrimp, *Penaeusmonodon*, associated with dominance of luminescent vibrios in the rearing environment. *Aquaculture*. 164: 337-349
- Law, J. W. F., Ab Mutalib, N. S., Chan, K. G., & Lee, L. H. (2015). Rapid methods for the detection of foodborne bacterial pathogens: principles, applications, advantages and limitations. *Frontiers in Microbiology*. 5:770. doi: 10.3389/fmicb.2014.00770
- Le Roux, F., Gay, M., Lambert, C., Nicolas, J. L., Gouy, M. & Berthe, F. (2004). Phylogenetic study and identification of *Vibrio splendidus*-related strains based on *gyrB* gene sequences. *Diseases of Aquatic Organisms*. 58(2-3):143-150.
- Lebech, A. M., Hindersson, P., Vuust, J., & Hansen, K. (1991). Comparison of *in vitro* culture and polymerase chain reaction for detection of *Borrelia burgdorferi* in tissue from experimentally infected animals. *Journal of Clinical Microbiology*. 29(4): 731-737.
- Lee, S. H., Hava, D. L., Waldor, M. K., & Camilli, A. (1999). Regulation and temporal expression patterns of *Vibrio cholera* virulence genes during infection. *Cell*. 996: 25-634. doi: 10.1016/S0092-8674(00)81551-2
- Leone, G., van Schijndel, H., van Gemen, B., Kramer, F. R., & Schoen, C. D. (1998). Molecular beacon probes combined with amplification by NASBA enable homogenous, real-time detection of RNA. *Nucleic Acids Research*. 26: 2150-2155. doi: 10.1093/nar/26.9.2150
- Lepage, T., & Gache, C. (1990). Early expression of a collagenase like hatching enzyme gene in the sea urchin embryo. *Embo Journal*. 9: 3003-3012.
- Liu, C.H.; Cheng, W.; Hsu, J.P. & Chen, J.C. (2004). *Vibrio alginolyticus* infection in the white shrimp *Litopenaeus vannamei* confirmed by polymerase chain reaction and 16S rDNA sequencing. *Diseases of Aquatic Organisms*. 61:169-174.

- Liu, L., Ge, M., Zheng, X., Tao, Z., Zhou, S., & Wang, G. (2016). Investigation of *Vibrio alginolyticus*, *V. harveyi*, and *V. parahaemolyticus* in large yellow croaker, *Pseudosciaena crocea* (Richardson) reared in Xiangshan Bay, China. *Aquaculture Reports*. 3: 220-224.
- Lovatelli, A., Phillips, M. J., Arthur, J. R., & Yamamoto, K. (2008). The future of mariculture: a regional approach for responsible development in the Asia-Pacific region. Guangzhou, China, 7–11 March 2006.
- Luo, P., & Hu, C. (2008). *Vibrio alginolyticusgyrB* sequence analysis and *gyrB*-targeted PCR identification in environmental isolates. *Diseases of Aquatic Organisms*. 82(3), 209-216.
- Luo, P., Jiang, H., Wang, Y., Su, T., Hu, C., Ren, C., & Jiang, X. (2013). Prevalence of mobile genetic elements and transposase genes in *Vibrio alginolyticus* from the southern coastal region of China and their role in horizontal gene transfer. *International Microbiology*. 15(4): 199-208
- Maeda, T., Takada, N., Furushita, M., & Shiba, T. (2000). Structural variation in the 16S-23S rRNA intergenic spacers of *Vibrio parahaemolyticus*. *FEMS Microbiology Letters*. 192: 73-77.
- Maeda, T., Matsuo, Y., Furushita, M. & Shiba, T. (2003). Seasonal dynamics in coastal *Vibrio* community examined by rapid and clustering method based on 16S rDNA. *Fisheries Science*. 69: 358-394.
- Mahbub, K. R., Paul, K. P. & Ahmed, M.M. (2011). Prevalence of *Vibrio* spp. and antibiogram of isolates from shrimp rearing ponds in Bangladesh. *Journal of Advanced Scientific Research*. 2:74-80.
- Markoulatos, P., Siafakas, N., & Moncany, M. (2002). Multiplex polymerase chain reaction: a practical approach. *Journal of Clinical Laboratory Analysis*. 16, 47-51. doi: 10.1002/jcla.2058
- Martinez-Murcia, A. J., Monera, A., Saavedra, M. J., Oncina, R., Lopez-Alvarez, M., Lara, E., & Figueras, M. J. (2011). Multilocus phylogenetic analysis of the genus *Aeromonas*. *Systematic and Applied Microbiology*. 34(3), 189-199.
- Mata, A. I., Gibello, A., Casamayor, A., Blanco, M. M., Domínguez, L., & Fernández-Garayzábal, J. F. (2004). Multiplex PCR assay for detection of bacterial pathogens associated with warm-water *Streptococcosis* in fish. *Applied and Environmental Microbiology*. 70(5): 3183-3187.

McCallum, H., Harvell, D., & Dobson, A. (2003). Rates of spread of marine pathogens. *Ecology Letters*. 6: 1062–1067. doi:10.1046/j.1461-0248.2003.00545.x.

McCallum, H.I., Kuris, A., Harvell, C.D., Lafferty, K.D., Smith, G.W., & Porter, J. (2004). Does terrestrial epidemiology apply to marine systems? *Trends in Ecology & Evolution*. 19: 585–591. doi:10.1016/j.tree.2004.08.009.

Michael, I.P., Westenskow, P.D., Hacibekiroglu, S., Cohen, Greenwald, A., Ballios, B.G., Kurihara, T., Li, Z., Warren, C.M., Zhang, P., Aguilar, E., Donaldson, L., Marchetti, V., Baba, T., Hussein, M.S., Sung, K.H., Iruela-Arispe, L.M., Rini, M.J., Van der Kooy, D., Friedlander, M. & Nagy, A. (2014). Local acting Sticky-trap inhibits VEGF dependent pathological angiogenesis in the eye. *EMBO Molecular Medicine*. 6:604–623.

Miyoshi, S. I. (2013). Extracellular proteolytic enzymes produced by human pathogenic *Vibrio* species. *Frontiers in Microbiology*. 4: 339.

Miyoshi, S., Nitanda, Y., Fujii, K., Kawahara, K., Li, T., Maehara, Y., Ramamurthy, T., Takeda, Y. & Shinoda, S. (2008). Differential expression and extracellular secretion of the collagenolytic enzymes by the pathogen *Vibrio parahaemolyticus*. *FEMS Microbiology Letters*. 283: 176–181. doi: 10.1111/j.1574-6968.2008.01159.x

Mukhopadhyay, A., & Mukhopadhyay, U. K. (2007). Novel multiplex PCR approaches for the simultaneous detection of human pathogens: *Escherichia coli* 0157: H7 and *Listeriamonocytogenes*. *Journal of Microbiological Methods*. 68(1): 193-200.

Narracci, M., Acquaviva, M. I., & Cavallo, R. A. (2013). Mar Piccolo of Taranto: *Vibrio* biodiversity in ecotoxicology approach. *Environmental Science and Pollution Research International*. 21: 2378–2385. doi: 10.1007/s11356-013-2049-3.

Nehlah, R. (2016). Development of recombinant outer membrane proteins vaccine and its immunoprotective ability against *Vibrio alginolyticus* in hybrid grouper (*Epinephelus fuscoguttatus* Forsskal X *Epinephelus lanceolatus* Bloch). (Master of Science dissertation), Universiti Putra Malaysia, Malaysia.

- Neogi, S. B., Chowdhury, N., Asakura, M., Hineno, A., Haldar, S., Saidi, S. M. & Yamasaki, S. (2010). A highly sensitive and specific multiplex PCR assay for simultaneous detection of *Vibrio cholerae*, *Vibrioparahaemolyticus* and *Vibrio vulnificus*. *Letters in Applied Microbiology*. 51(3): 293-300.
- Nhung, P. H., Ohkusu, K., Miyasaka, J., Sun, X. S., & Ezaki, T. (2007). Rapid and specific identification of 5 human pathogenic *Vibrio* species by multiplex polymerase chain reaction targeted to *dnaJ* gene. *Diagnostic microbiology and infectious disease*. 59(3): 271-275.
- Nordstrom, J. L., Vickery, M.C., Blackstone, G. M., Murray, S. L., & DePaola, A. (2007). Development of a multiplex real-time PCR assay with an internal amplification control for the detection of total and pathogenic *Vibrio parahaemolyticus* bacteria in oysters. *Applied and Environmental Microbiology*. 73:5840-5847. Doi:10.1128/AEM.00460-07.
- Notomi, T., Okayama, H., Masubuchi, H., Yonekawa, T., Watanabe, K., Amino, N., & Hase, T. (2000). Loop-mediated isothermal amplification of DNA. *Nucleic Acids Research*. 28(12): e63-e63.
- Oates, S. C., Miller, M. A., Byrne, B. A., Chouicha, N., Hardin, D., Jessup, D., Dominik, C., Roug, A., Schriewer, A., Jang, S. S., & Milerr, A. W. (2012). Epidemiology and potential land sea transfer of enteric bacteria from terrestrial to marine species in the Monterey Bay Region of California. *Journal of Wildlife Diseases*. 48: 654-668. doi:10.7589/0090-3558-48.3.654.
- Olivier, G. (2002). Disease interactions between wild and cultured fish—perspectives from the American Northeast (Atlantic provinces). *Bulletin of the European Association of Fish Pathologists*. 22: 103–109.
- Osorio, C., & Toranzo, A. (2002). DNA-based diagnostics in sea farming, p.253-310. In M. Fingerman and R. Nagabhushanam (ed.), *Seafood safety and human health*. Science Publishers, Inc., Enfield, N.H.
- Osorio, C. R., & Klose, K. E. (2000). A region of the trans-membrane regulatory protein *toxR* that tethers the transcriptional activation domain to the cytoplasmic membrane displays wide divergence among *Vibrio* species. *Journal of Bacteriology*. 182: 526-528.
- Osorio, C.R.; Collins, M.D.; Romalde, J.L.; & Toranzo, A.E. (2005). Variation in 16S-23S rRNA intergenic spacer regions in *Photobacterium damselae*: a mosaic-like structure. *Applied and Environmental Microbiology*. 71:636-645.

- Pandove, G., Sahota, P., Vikal, Y., & Kaur, B. (2013). Multiplex PCR water testing kit for rapid, economic and simultaneous detection of *Escherichiacoli*, *Yersinia enterocolitica* and *Aeromonas hydrophila* from drinking water. *Current Science (Bangalore)*. 104(3): 352-358.
- Panicker, G., Call, D. R., Krug, M. J., & Bej, A. K. (2004). Detection of pathogenic *Vibrio* spp. in shellfish by using multiplex PCR and DNA microarrays. *Applied Environment Microbiology* 70, 7436-7444.
- Park, S., Kim, H., Kim, J., Kim, T., & Kim, H. (2007). Simultaneous detection and identification of *Bacillus cereus* group bacteria using multiplex PCR. *Journal of Microbiology and Biotechnology*. 17(7): 1177.
- Park, Y. S., Lee, S. R., & Kim, Y. G. (2006). Detection of *Escherichia coli* O157: H7, *Salmonella* spp., *Staphylococcus aureus* and *Listeria monocytogenes* in kimchi by multiplex polymerase chain reaction (mPCR). *Journal of Microbiology Seoul*. 44(1): 92.
- Park, S. H., Kim, H. J., Cho, W. H., Kim, J. H., Oh, M. H., Kim, S. H., Lee, B. K., Ricke, S. C. & Kim, H. Y. (2009). Identification of *Salmonella enterica* subspecies I, *Salmonella enterica* serovars Typhimurium, Enteritidis and Typhi using multiplex PCR. *FEMS Microbiology Letters*. 301: 137-146. doi:10.1111/j.1574-6968.2009.01809.x
- Parker, J. L., & Shaw, J. G. (2011). *Aeromonas* spp. clinical microbiology and disease. *Journal of Infection*. 62(2): 109-118.
- Payne, S. M., Mey, A. R., & Wyckoff, E. E. (2015) *Vibrio* iron transport: evolutionary adaptation to life in multiple environments. *Microbiology and Molecular Biology Reviews*. 9: 69-90
- Pereira, C. S., Viana, C. M., & Rodrigues, D. D. P. (2007). Pathogenic vibrios in oysters (*Crassostrea rhizophorae*) served at restaurants in Rio de Janeiro: a public health warning. *Revista da Sociedade Brasileira de Medicina Tropical*. 40(3): 300-303. doi: 10.1590/S0037-86822007000300010.
- Ponprateep, S., Somboonwiwat, K. & Tassanakajon, A. (2009). Recombinant anti-lipopolysaccharide factor isoform 3 and the prevention of vibriosis in the black tiger shrimp, *Penaeus monodon*. *Aquaculture*. 289: 219-224

- Pullkinen, K., Suomalainen, L.R., Read, A.F., Ebert, D., Rintamaki P., & Valtonen, E.T.(2010). Intensive fish farming and the evolution of pathogen virulence: the case of columnaris disease in Finland, Proceeding of the Royal, Series B, *Biological Science*. 277: 593-600.
- Puthucheary, S. D., Puah, S. M., & Chua, K. H. (2012). Molecular characterization of clinical isolates of *Aeromonas* species from Malaysia. *PLoS One*. 7(2): e30205.
- Puttinaowarat, S., Thompson, K. D., & Adams, A. (2000). Mycobacteriosis: detection and identification of aquatic *Mycobacterium* species. *Fish Veterinary Journal*. 5: 6-21.
- Qian, R., Chu, W., Mao, Z., Zhang, C., Wei, Y. & Yu, L. (2007). Expression, characterization and immunogenicity of a major outer membrane protein from *Vibrio alginolyticus*. *Acta Biochimica et Biophysica Sinica*. 39(3): 194-200.
- Qian, R., Xiao, Z., Zhang, C., Chu, W., Mao, Z., & Yu, L. (2008). Expression and purification of two major outer membrane proteins from *Vibrio alginolyticus*. *World Journal of Microbiology and Biotechnology*. 24: 245-251.
- Raissy, M., Momtaz, H., Moumeni, M., Ansari, M., & Rahimi, E. (2011). Molecular detection of *Vibrio* spp. in lobster hemolymph. *African Journal of Microbiology Research*. 5(13):1697-1700.
- Raja, N., Shamsudin, M. N., Somarny, W., Rosli, R., Rahim, R. A., & Radu, S. (2001). Detection and molecular characterization of the zot gene in *Vibrio cholerae* and *V. alginolyticus* isolates. *The Southeast Asian Journal of Tropical Medicine and Public Health*. 32:100-104
- Raja, R. A., & Jithendran, K. P. (2015). Aquaculture Disease Diagnosis and Health Management. In *Advances in Marine and Brackishwater Aquaculture* (pp. 247-255). Springer India.
- Rajendhran, J., & Gunasekaran, P. (2011). Microbial phylogeny and diversity: small subunit ribosomal RNA sequence analysis and beyond. *Microbiological Research*. 166(2): 99-110.
- Rasheed, V. (1989). Disease of cultured brown-spotted grouper *Epinephelus tauvina* and silvery black porgy *Acanthopagrus cuvieri* in Kuwait. *Journal of Aquatic Animal Health*. 1(2): 102-107.

Raychaudhuri, S., Jain, V. & Dongre, M. (2006). Identification of a constitutively active variant of *LuxO* that affects production of HA/protease and biofilm development in a non-O1, non-O139 *Vibrio cholera* O110. *Gene*. 369: 126–133. doi: 10.1016/j.gene.2005.10.031.

Robert-Pillot, A., Guenole, A., & Fournier, J.M. (2002). Usefulness of R72H PCR assay for differentiation between *Vibrio parahaemolyticus* and *Vibrio alginolyticus* species: validation by DNA-DNA hybridization. *FEMS Microbiology Letters*. 215: 1-6.

Roberts, D. M., Personne, Y., Ollinger, J., & Parish, T. (2013). Proteases in *Mycobacterium tuberculosis* pathogenesis: potential as drug targets. *Future Microbiology*.8(5): 621-631.

Rodkhum, C., Kayansamruaj, P., Pirarat, N., & Wongtawatchai, J. (2012). Duplex PCR for Simultaneous and Unambiguous Detection of *Streptococcus viriae* and *Streptococcus galactiae* associated with Streptococcosis of Cultured Tilapia in Thailand. *The Thai Journal of Veterinary Medicine*. 42(2): 153-158.

Roszen, L., Nørskov, P., Holmstrøm, K., & Rasmussen, O. F. (1992). Inhibition of PCR by components of food samples, microbial diagnostic assays and DNA-extraction solutions. *International Journal of Food Microbiology*.17(1): 37-45.

Roux, F. L., Wegner, K. M., Baker-Austin, C., Vezzulli, L., Osorio, C. R., Amaro, C., Ritchie, J.M., Defoirdt, T., Destoumieux-Garzón,D., Blokesh, M. & Mazel, D. (2015). The emergence of *Vibrio* pathogens in Europe: ecology, evolution, and pathogenesis (Paris, 11–12th March 2015). *Frontiers in Microbiology*.6: 830.

Rowe-Magnus, D. A., Guerout, A. M., Ploncard, P., Dychinco, B., Davies, J. & Mazel, D. (2001). The evolutionary history of chromosomal super-integrons provides an ancestry for multi resistant integrons. *Proceedings of the National Academy of Sciences*. 98: 652–627. doi: 10.1073/pnas.98.2.652.

Ruimy, R., Breittmayer, V., Elbaxe, P., Lafay, B., Boussemart, O., & Gauthier, M. (1994) Phylogenetic analysis and assessment of the genera *Vibrio*, *Photobacterium*, *Aeromonas*, and *Plesiomonas* deduced from small-subunit rRNA sequences. *International Journal of Systematic and Evolutionary Microbiology*.44:416–426

- Sanjuán, E., & Amaro, C. (2007). Multiplex PCR assay for detection of *Vibiovulnificus* biotype 2 and simultaneous discrimination of serovar E strains. *Applied and Environmental Microbiology*.73(6): 2029-2032.
- Sadok, K., Mejdi, S., Nourhen, S., & Amina, B. (2013). Phenotypic characterization and RAPD fingerprinting of *Vibrio parahaemolyticus* and *Vibrio alginolyticus* isolated during Tunisian fish farm outbreaks. *Folia Microbiologica*.58(1): 17-26.
- Saeed.M.O. (1995).Association of *Vibrio harveyi* with mortalities in cultures marine fish in Kuwait. *Aquaculture*. 136: 21-29.
- Sarjito, S., Radjasa, O. K., Sabdono, A., Prayitno, S. B., & Hutabarat, S. (2009). Phylogenetic diversity of the causative agents of vibriosis associated with groupers fish from Karimunjawa Islands, Indonesia. *Current Research in Bacteriology*. 2(1): 14-21.
- Schets, F. M., Van den Berg, H. H., Demeulmeester, A. A., Van Dijk, E., Rutjes, S. A., van Hooijdonk, H. J., & de Roda Husman, A. M. (2006). *Vibrio alginolyticus* infections in the Netherlands after swimming in the North Sea. *Euro Surveill*. 11(45): 3077.
- Schets, F. M., Van Den Berg, H. H., Marchese, A., Garbom, S., De Roda & Husman, A. M. (2011). Potentially human pathogenic vibrios in marine and fresh bathing waters related to environmental conditions and disease outcome. *International Journal of Hygiene and Environmental Health*.214: 399–406. doi:10.1016/j.ijheh.2011. 05.003.
- Schets, F. M., Van Den Berg, H. H., Rutjes, S. A., & De Roda Husman, A. M. (2010). Pathogenic *Vibrio* species in dutch shellfish destined for direct human consumption. *Journal of Food Protection*. 73: 734-738.
- Sechi, L.A., Dupre, I., Deriu, A., Fadda, G., & Zanetti, S. (2000). Distribution of *Vibrio cholerae* virulence genes among different *Vibrio* species isolated in Sardinia, Italy. *Journal of Applied Microbiology*. 88: 475–481.
- Selvin, J., & Lipton, A.P. (2003) *Vibrioalginolyticus* associated with white spot disease of *Penaeusmonodon*. *Diseases of Aquatic Organisms*. 57(1-2): 147-50.
- Sen, K. (2005) Development of a rapid identification method for *Aeromonas* species by multiplex-PCR. *Canadian Journal of Microbiology*. 51:957-966.

- Senoo, S. (2006). Hybrid production between tiger grouper *Epinephelus fuscoguttatus* × giant grouper *Epinephelus lanceolatus* (fish culture in Southeast Asia 64). *Aquaculture Magazine*. 12: 58-63.
- Senoo, S. (2010). Consideration of artificial egg collection technique on fish IV (fish culture in Southeast Asia 80). *Aquaculture Magazine*. 204: 64-67.
- Shariff, M., Yusoff, F.M., & Gopinath, N. (1997). Aquaculture in Malaysia: Current Trends and Future Outlook. In Proceedings of Second International Seminar on Fisheries Science in Tropical Area. August 19-22. (p. 45-51), Tokyo, Japan.
- Sharma, K., Ramachandra, S., Rathore, G., Verma, D. K., Sadhu, N., & Philipose, K.K (2011). *V. alginolyticus* infection in Asian Seabass (*Lates calcarifer*, Bloch) reared in open sea floating cages in India. *Aquaculture Research*. 44(1): 86-92.
- Shen, G. M., Shi, C. Y., Fan, C., Jia, D., Wang, S. Q., Xie, G. S., Li, G. Y., Mo, Z. L., & Huang, J. (2017). Isolation, identification and pathogenicity of *Vibrio harveyi*, the causal agent of skin ulcer disease in juvenile hybrid groupers *Epinephelus fuscoguttatus* × *Epinephelus lanceolatus*. *Journal of Fish Disease*. doi:10.1111/jfd.12609
- Shi, Y.-H., Chen, J., Li, C.H., Lu, X.-J., & Zhang, D.-M. et al. (2012). Detection of bacterial pathogens in aquaculture samples by DNA microarray analysis. *Aquaculture*. 338-341: 29-35.
- Shinoda S. & Miyoshi S. (2011). Proteases produced by vibrios. *Biocontrol Science*. 16:1-11. <http://dx.doi.org/10.4265/bio.16.1>.
- Silva-Rubio, A., Avendano-Herrera, R., Jaureguiberry, B., Toranzo, A.E., & Magarinos, B. (2008) First description of serotype O3 in *Vibrio anguillarum* strains isolated from salmonids in Chile. *Journal of Fish Diseases*. 31:235-239
- Simpkins, S. A., Chan, A. B., Hays, J., Pöpping, B., & Cook, N. (2000). An RNA transcription-based amplification technique (NASBA) for the detection of viable *Salmonella enterica*. *Letters in Applied Microbiology*. 30:75-79. doi: 10.1046/j.1472-765x.2000.00670.x
- Slack, A. T., Symonds, M. L., Dohnt, M. F., & Smythe, L. D. (2006). Identification of pathogenic *Leptospira* species by conventional or real-time PCR and sequencing of the DNA gyrase subunit B encoding gene. *BMC Microbiology*. 6:95-104.

- Snoussi, M., Noumi, E., Usai, D., Sechi, L. A., Zanetti, S., & Bakhrou, A. (2008). Distribution of some virulence related-properties of *Vibrio alginolyticus* strains isolated from Mediterranean seawater (Bay of Khenis, Tunisia): investigation of eight *Vibrio cholerae* virulence genes. *World Journal of Microbiology and Biotechnology*. 24:2133-2141
- Sobhana, K. S. (2009). Diseases of seabass in cage culture and control measures. pp. 87-93.
- Stager, C. E., & Davis, J. R. (1992). Automated system for identification of microorganisms. *Clinical Microbiology Review*. 5: 302-327.
- Su, T., Luo, P., Ren, C., & Hu, C. (2010). Complete nucleotide sequence of a plasmid pVAE259 from *Vibrio alginolyticus* and analysis of molecular biological characteristic of the plasmid. *Acta Microbiologica Sinica*. 50:162-168
- Sun, K., Zhang, W.W., Hou, J.H., & Sun, L. (2009). Immunoprotective analysis of VhhP2, a *Vibrio harveyi* vaccine candidate. *Vaccine*. 27:2733-2740
- Tabacchioni, S., Ferri, L., Manno, G., Mentasti, M., Cocchi, P., Campana, S., Ravenni, N., Taccetti, G., Dalmastri, C., Chiarini, L., Fani, R., & Bevivino, A. (2008). Use of the gyrB gene to discriminate among species of the *Burkholderia cepacia* complex. *FEMS Microbiology Letters*. 281(2): 175-182.
- Tacão, M., Moura, A., Henriques, I., Savedra, M.J., & Correia, A. (2005). Evaluation of 16S rDNA and gyrB-DGGE for typing members of the genus *Aeromonas*. *FEMS Microbiology Letters*. 246: 11-18
- Takeuchi, H., Shibano, Y., Morihara, K., Fukushima, J., Inami, S., Kell, B., Gilles, A. M., Kawamoto, S., & Okuda, K. (1992). Structural gene and complete amino acid sequence of *Vibrio alginolyticus* collagenase. *Biochemical Journal*. 281: 703-708.
- Tan, C. K., (1998). Overview of Aquaculture in Malaysia. In Nagaraj and Singh (eds.) Aquaculture Practices in Malaysia. Occasional Publication No. 9. Kuala Lumpur: Malaysian Fisheries Society.
- Tani, K., Kobayashi, T., Sakotani, A., Kenzaka, T., & Nasu, M. (2012). Expression of the gyrB gene as an indicator of growth activity of *Escherichia coli*. *Journal of Petroleum and Environmental Biotechnology*. 12: 33-38

Tantillo, G.M.; Fontanarosa, M.; Di Pinto, A. & Musti, M. (2004). Updated perspectives on emerging vibrios associated with human infections. *Letters in Applied Microbiology*. 39: 117-126.

Tapia-Cammas, D., Yañez, A., Arancibia, G., Toranzo, A. E., & Avendaño-Herrera, R. (2011). Multiplex PCR for the detection of *Piscirickettsia salmonis*, *Vibrio anguillarum*, *Aeromonas salmonicida* and *Streptococcus phocae* in Chilean marine farms. *Diseases of Aquatic Organisms*. 97(2): 135-142.

Tarr, C. L., Patel, J. S., Puhr, N. D., Sowers, E. G., Bopp, C. A., & Strockbine, N. A. (2007). Identification of *Vibrio* isolates by a multiplex PCR assay and *rpoB* sequence determination. *Journal of Clinical Microbiology*. 45(1): 134-140.

Teh, C. S. J., Chua, K. H., & Thong, K. L. (2010). Simultaneous differential detection of human pathogenic and nonpathogenic *Vibrio* species using a multiplex PCR based on *gyrB* and *pntA* genes. *Journal of Applied Microbiology*. 108(6): 1940-1945.

Teramura, N., Tanaka, K., Iijima, K., Hayashida, O., Suzuki, K., Hattori, S. & Irie, S. (2011). Cloning of a novel collagenase gene from the gram-negative bacterium *Grimontia (Vibrio) hollisae* 1706B and its efficient expression in *Brevibacillus schoshinensis*. *Journal of Bacteriology*. 193(12): 3049-3056.

Thaithongnum, S., Ratanama, P., Weeradechapol, K., Sukhoom, A., & Vuddhakul, V. (2006) Detection of *V. harveyi* in shrimp postlarvae and hatchery tank water by the most probable number technique with PCR. *Aquaculture*. 261:1-9.

The state of world fisheries and aquaculture (2016). Contributing to food security and nutrition for all. (2016). Rome: Food and Agriculture Organization of the United Nations.

Thompson, F.L., & Swings, J. (2006). Taxonomy of vibrios. In: Thompson FL, Austin B, Swings J, Nishibuchi M (eds) The biology of vibrios. ASM Press, Washington, DC.

Thompson, C. C., Thompson, F. L., Vandemeulebroecke, K., Hoste, B., Dawyndt, P. & Swings, J. (2004). Use of *recA* as an alternative phylogenetic marker in the family Vibrionaceae. *International Journal of Systematic and Evolutionary Microbiology*. 54(3): 919-924.

- Thompson, F. L., Gevers, D., Thompson, C. C., Dawyndt, P., Naser, S., Hoste, B., Munn, C. B. & Swings, J. (2005). Phylogeny and molecular identification of vibrios on the basis of multilocus sequence analysis. *Applied and Environmental Microbiology*. 71: 5107-5115.
- Tolmisky, M. E., & Crosa, J. H. (1991). Regulation of plasmid-mediated iron transport and virulence in *Vibrio anguillarum*. *Biometals*. 4(1): 33-35.
- Toranzo, A. E., Magariños, B., & Romalde, J. L. (2005). A review of the main bacterial fish diseases in mariculture systems. *Aquaculture*. 246(1): 37-61.
- Torresi, M., Acciari, V.A., Piano, A., Serratore, P., Prencipe, V., & Migliorati, G. (2011). Detection of *Vibrio splendidus* and related species in *Chamelea gallina* sampled in the Adriatic along the Abruzzi coastline. *Veterinaria Italiana*. 47: 363-370.
- Travis, J., Potempa, J., & Maeda, H. (1995). Are bacterial proteinases pathogenic factors?. *Trends in Microbiology*. 3:405-7.
- Tsolis, R.M. (2002). Comparative genome analysis of the alpha - proteobacteria: relationships between plant and animal pathogens and host specificity. *Proceedings of the National Academy of Sciences of the United States of America*. 99: 12503-12505.
- Van Ketel, R. J., De Wever, B. & Van Alphen, L. (1990). Detection of *Haemophilus influenzae* in cerebrospinal fluids by polymerase chain reaction DNA amplification. *Journal of Medical Microbiology*. 33:271-276.
- Vantarakis, A., Komninos, G., Venieri, D. & Papapetropoulou, M. (2000). Development of a multiplex PCR detection of *Salmonella* spp. and *Shigella* spp. in mussels. *Letters in Applied Microbiology*. 31(2): 105-109.
- Verschueren, L., Rombout, G., Sorgeloos, P., & Verstraete, W. (2000). Probiotic bacteria as biological control agents in aquaculture. *Microbiology and Molecular Biology Reviews*. 64:655-671.
- Vinothkumar, K., Bhardwaj, A.K., & Ramamurthy, T. (2013). Triplex PCR assay for the rapid identification of 3 major *Vibrio* species, *Vibrio cholerae*, *Vibrioparahaemolyticus*, and *Vibrio fluvialis*. *Diagnostic Microbiology and Infectious Disease*. 76:526-8

- Vitanen, A. M., Arstilla, T.P., Lahesmaa, R. Granfors, K., Skurnik, M., & Toivanen, P. (1991). Application of polymerase chain reaction and immuno-fluorescence techniques to the detection of bacteria in *Yersinia*-triggered reactive arthritis. *Arthritis and Rheumatology*. 34:89-96.
- Vogel, J., Normand, P., Thioulouse, J., Nesme, X., & Grundmann, G.L. (2003). Relationship between spatial and genetic distance in *Agrobacterium* spp in 1 cubic centimeter of soil. *Applied and Environmental Microbiology*. 69: 1482-1487.
- Vongxay, K., He, X., Cheng, S., Zhou, X., Shen, B., Zhang, G., & Fang, W. (2006). Prevalence of *Vibrioparahaemolyticus* in seafoods and their processing environments as detected by duplex PCR. *Journal of the Science of Food and Agriculture*. 86(12): 1871-1877.
- Vuddhakul, V., Nakai, T., Matsumoto, C., Oh, T., Nishino, T., Chen, C.H., Nishibuchi, M., & Okuda, J. (2000). Analysis of *gyrB* and *toxR* Gene Sequences of *Vibrio hollisae* and Development of *gyrB*-and *toxR*-Targeted PCR Methods for Isolation of *V. hollisae* from the Environment and Its Identification. *Applied and Environmental Microbiology*. 66(8): 3506-3514.
- Walker P. J., & Mohan C.V. (2009). Viral disease emergence in shrimp aquaculture: origins, impact and the effectiveness of health management. *Reviews in Aquaculture*. 1: 125-154.
- Wang, Q., Liu, Q., Cao, X., Yang, M., & Zhang, Y. (2008). Characterization of two TonB systems in marine fish pathogen *Vibrio alginolyticus*: their roles in iron utilization and virulence. *Archives of Microbiology*. 190(5): 595-603.
- Ward, L. N., & Bej, A. K. (2006). Detection of *Vibrio parahaemolyticus* in shellfish by use of multiplexed real-time PCR with TaqMan fluorescent probes. *Applied and Environmental Microbiology*. 72(3): 2031-2042.
- Warsen, A.E., Krug, M.J., LaFrentz, S., Stanek, D.R., Loge, F.J., & Call, D.R. (2004). Simultaneous Discrimination between 15 Fish Pathogens by Using 16S Ribosomal DNA PCR and DNA Microarrays. *Applied and Environmental Microbiology*. 70(7): 4216-4221.

- Wei, S., Zhao, H., Xian, Y., Hussain, M. A., & Wu, X. (2014). Multiplex PCR assays for the detection of *Vibrio alginolyticus*, *Vibrio parahaemolyticus*, *Vibrio vulnificus* and *Vibrio cholerae* with an internal amplification control. *Diagnostic Microbiology and Infectious Disease*.79: 115–118. doi: 10.1016/j.diagmicrobio.2014.03.012.
- Whitehead, N. A., Barnard, A. M., Slater, H., Simpson, N. J., & Salmond, G. P. (2001). Quorum-sensing in Gram-negative bacteria. *FEMS Microbiology Reviews* .25: 365–404. doi: 10.1111/j.1574-6976.2001.tb00583.x
- Wilson, T., & Carson, J. (2003) Development of sensitive, high throughput one-tube RT-PCR-enzyme hybridization assay to detect selected bacterial fish pathogens. *Diseases of Aquatic Organisms*.54: 127–134
- Woese, C. R., Kandler, O., & Wheelis, M. L. (1990). Towards a natural system of organisms: proposal for the domains Archaea, Bacteria, and Eucarya. *Proceedings of the National Academy of Sciences*. 87(12): 4576–4579.
- Wu, J. G., Wang, J. F., Zhang, X. H., Zhang, S. S., Hu, X. F., & Chen, J. S. (2010). A *gyrB*-targeted PCR for rapid identification of *Paenibacillus mucilaginosus*. *Applied Microbiology and Biotechnology*. 87(2): 739–747.
- Xie, Z. Y., Hu, C. Q., Chen, C., Zhang L. P. & Ren, C. H. (2005). Investigation Of seven *vibrio* virulence genes among *Vibrio alginolyticus* and *Vibrioparahaemolyticus* strains from the coastal mariculture systems in Guangdong, China. *Letters in Applied Microbiology*. 41: 202–207.
- Yamada, S., Ohashi, E., Agata, N. & Venkateswaran, K. (1999) Cloning and nucleotide sequence analysis of *gyrB* of *Bacillus cereus*, *B. thuringiensis*, *B. mycoides*, and *B. anthracis* and their application to the detection of *B. cereus* in rice. *Applied and Environmental Microbiology*.65:1483–1490
- Yamamoto, S. & Harayama, S. (1995). PCR amplification and direct sequencing of *gyrB* genes with universal primers and their application to the detection and taxonomic analysis of *Pseudomonas putida* strains. *Applied and Environmental Microbiology*. 61:1104–1109.
- Yamamoto, S. & Harayama, S. (1996). Phylogenetic analysis of *Acinetobacter* strains based on the nucleotide sequence of *gyrB* genes and on the amino acid sequences of their products *International Journal of Systematic and Evolutionary Microbiology*.46: 506–511.

- Yamamoto, S., Bouvet, P. J. & Harayama, S. (1999). Phylogenetic structures of the genus *Acinetobacter* based on *gyrB* sequences: comparison with the grouping by DNA-DNA hybridization. *International Journal of Systematic and Evolutionary Microbiology*. 49(1): 87-95.
- Yamazaki, W., Ishibashi, M., Kawahara, R., & Inoue, K. (2008). Development of a loop-mediated isothermal amplification assay for sensitive and rapid detection of *Vibrio parahaemolyticus*. *BMC microbiology*. 8(1): 1.
- Yan, Q., Chen, Q., Ma, S., Zhuang, Z., & Wang, X. (2007). Characteristics of adherence of pathogenic *Vibrio alginolyticus* to the intestinal mucus of large yellow croaker (*Pseudosciaena crocea*). *Aquaculture*. 269: 21-30.
- Yanez, M. A., V. Catalan, D. Apraiz, M. J. Figueras, & A. J. Martinez-Murcia. (2003). Phylogenetic analysis of members of the genus *Aeromonas* based on *gyrB* gene sequences. *International Journal of Systematic and Evolutionary Microbiology*. 53:875-883.
- Yang, H., Chen, J., Yang, G., Zhang, X.H., Liu, R., & Xue, X. (2009) Protection of Japanese flounder (*Paralichthys olivaceus*) against *Vibrio anguillarum* with a DNA vaccine containing the mutated zinc-metalloprotease gene. *Vaccine*. 27:2150-2155
- Ye, Y., Wang, B., Huang, F., Song, Y., Yan, H., Alam, M. J., et al. (2011). Application of in situ loop-mediated isothermal amplification method for detection of *Salmonella* in foods. *Food Control* 22, 438-444. doi: 10.1016/j.foodcont.2010.09.023
- Yeung, P.S., & Boor, K.J. (2004). Epidemiology, pathogenesis, and prevention of foodborne *Vibrio parahaemolyticus* infections. *Foodborne Pathogens and Disease*. 1:74-88.
- Yin, H., Cao, L., Qiu, G., Wang, D., Kellogg, L., Zhou, J., Liu, X., Dai, Z., & Ding, J. (2008). Molecular diversity of 16S rRNA and *gyrB* genes in copper mines. *Archives of Microbiology*. 189: 101-110.
- Yishan, L., Jiaming, F., Zaohe, W., & Jichang, J. (2011). Genotype analysis of collagenase gene by PCR-SSCP in *Vibrio alginolyticus* and its association with virulence to marine fish. *Current Microbiology*. 62(6): 1697-1703.
- Yu, S., Chen, W., Wang, D., He, X., Zhu, X., & Shi, X. (2010). Species-specific PCR detection of the food-borne pathogen *Vibrio parahaemolyticus* using the *irgB* gene identified by comparative genomic analysis. *FEMS Microbiology Letters*. 307(1): 65-71.

- Yusoff, A. (2015). Status of resource management and aquaculture in Malaysia. In Resource Enhancement and Sustainable Aquaculture Practices in Southeast Asia: Challenges in Responsible Production of Aquatic Species: Proceedings of the International Workshop on Resource Enhancement and Sustainable Aquaculture Practices in Southeast Asia 2014 (RESA) (pp. 53-65). Aquaculture Department, Southeast Asian Fisheries Development Center.
- Zamri-Saad, M., Amal, M.N.A., Siti-Zahrah, A., & Zulkafli, A.R., (2014). Control and prevention of *streptococciosis* in cultured tilapia in Malaysia: a review. *Pertanika Journal of Tropical Agricultural Science*.37 (4): 389-410.
- Zanetti, S., Deriu, A., Dupre, I., Sanguinetti, M., Fadda, G.,&Sechi, L.A. (1999). Differentiation of *Vibrio alginolyticus* strains isolated from Sardinian waters by ribotyping and a new rapid PCR fingerprinting method. *Applied Environmental Microbiology*.65:1871-1875.
- Zhang, C., Yu, L. & Qian, R. (2007). Characterization of OmpK, GAPDH and their fusion OmpK-GAPDH derived from *Vibrio harveyi* outer membrane proteins: their immune protective ability against vibriosis in large yellow croaker (*Pseudosciaena crocea*). *Journal of Applied Microbiology*. 103: 1587-1599.
- Zhang, X.H., & Austin, B. (2000) Pathogenicity of *Vibrio harveyi* to salmonids. *Journal of Fish Disease*. 23: 93-102.
- Zhao, X., Lin, C. W., Wang, J., & Oh, D. H. (2014).Advances in rapid detection methods for foodborne pathogens. *Journal of Microbiology and Biotechnology*. 24: 297-312. doi:10.4014/jmb.1310.10013
- Zhao, Z., Chen, C., Chao-Qun, H., Ren, C.H., Zhao, J.J., Zhang, L.P., Jiang, X., Luo, P. & Wang, Q.B. (2010).The type III secretion system of *Vibrioalginolyticus* induces rapid apoptosis, cell rounding and osmotic lysis of fish cells. *Microbiology*.156(9): 2864-2872.
- Zhou , S.,Hou, Z., Li, N. & Qin, Q. (2007). Development of a SYBR Green I real-time PCR for quantitative detection of *Vibrio alginolyticus* in seawater and seafood. *Journal of Applied Microbiology*.103(5): 1897-1906.

BIODATA OF STUDENT

Diyana Nadhirah was born in Shah Alam, Selangor in 20th May 1990 and grew up in Temerloh, Pahang. She began her primary education at Sekolah Kebangsaan Paya Taram from 1997 to 2002 and was selected to continue her secondary education at Sekolah Menengah Kebangsaan Kerdau, Temerloh for three years from 2003 to 2005. She was transferred into Sekolah Menengah Kebangsaan Temerloh Jaya for one year and after that she was transferred into Sekolah Menengah Kebangsaan Temerloh and had graduated on 2007. Then, she attended a two-year matriculation program at Johor Matriculation College (KMJ) before entering Universiti Putra Malaysia (UPM) in Serdang in 2010. There, she studied Agriculture, majored in aquaculture for four years. In 2014, she graduated from UPM with Bachelor's degree with honors.

In 2014, she commenced the Master of Science degree in the field of Aquaculture under the supervision of Dr. Ina Salwany Md Yasin at the Faculty of Agriculture, Universiti Putra Malaysia. She was involved in a research project to study the development of duplex polymerase chain reaction (PCR) for the diagnosis of *Vibrio alginolyticus* infection for marine aquaculture.

LIST OF PUBLICATIONS

Diyana-Nadhirah, K.P., and M.Y. Ina-Salwany. (2016). Molecular Characterization of 16S rRNA and Internal Transcribed Spacer (ITS) Regions of *Aeromonas* spp. Isolated from Cultured Freshwater Fishes in Malaysia. *Int.J.Curr.Microbiol.App.Sci.* 5(9): 431-440. doi: <http://dx.doi.org/10.20546/ijcmas.2016.509.046>

Rabi'atul, A. S, Diyana-Nadhirah, K. P., Nehlah, R., Saleema, M., Zarirah.Z., & Ina-Salwany, M.Y. (2015). Internal Transcribed Spacer (ITS) Gene Sequencing and Pathogenicity Study of *Aeromonas veronii* bv. *sobria* Strain KTG3SB in Juvenile Red Tilapia. *National Seminar on Advances in Fish Health 2015*. 4th – 5th February 2015, Kajang, Selangor, Malaysia.



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