



UNIVERSITI PUTRA MALAYSIA

**ANTIOXIDANT AND ANTICANCER PROPERTIES OF CRUDE
EXTRACTS FROM *DEDALU* (*Helixanthera parasitica* Lour.)**

THIRUBUVANESVARI DURAIVELU

FS 2020 9



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By

THIRUBUVANESVARI DURAIVELU

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

March 2019

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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March 2019

Chair : Professor Rusea Go, PhD
Faculty : Science

Helixanthera parasitica Lour, recognized as a hemiparasitic mistletoe from the family Loranthaceae is a mystique plant that has been utilized ethnobotanically as medicinal plant in Southeast Asia countries. Up to now and to the best of our knowledge very scarce research has been conducted on this species to exploit its further medicinal potential particularly on exploring its antioxidant capacity and towards one of the leading non-communicable disease (NCD), that has remained difficult to treat but preventable, the breast cancer. The present study was designed to prepare different extracts from the leaves and stem of *H. parasitica* and to determine their total phenol, flavonoids content, antioxidant activity as well as selecting the best extracts towards anticancer assay by applying established methods. Different extracts were prepared by maceration technique with powdered leaves and stem sample. Extraction of both plant part were performed separately using solvents of different polarity, namely, distilled water (dH₂O), methanol (70% and 100), acetone (70% and 100%) and petroleum ether. The parameters targeted on the plant extracts were determined using Total Phenolic Content (TPC)/ Folin-Ciocalteu reagent (FCR), Total Flavonoid Content (TFC)/ aluminium chloride (AlCl₃), 2, 2-diphenyl-1-picrylhydrazyl (DPPH), ferric reducing antioxidant power (FRAP) and MTT (for anticancer assay) method. Highest extract yield was obtained from leaf extracts (679.13 ± 9.60 mg/g DW) and the yields of extraction by various solvents decreased in the following order: 70% acetone > 70% methanol > 100% methanol > 100% acetone > dH₂O > petroleum ether. Leaf extract from pure acetone solvent was found to exhibit highest TPC (99.45 ± 12.87 mg GAE/g DW), TFC (17.57 ± 0.63 mg QE/g DW) and DPPH (81.61 ± 1.22 %) with no significant difference found between leaf and stem evaluated from TPC and DPPH. This extract also showed slightly lower FRAP value (6.73 ± 0.36 mg TE/g DW), but without significant difference compared to stem. In terms of 50% inhibition concentration of DPPH (IC₅₀), the lowest value was retrieved from 100% acetone stem extract (42.66 ± 4.82 µg/mL), however this value is insignificant (p > 0.05) with the top scoring extract, 100% acetone leaf (45.21 ± 4.91 µg/mL) which are both almost similar to the potent standard antioxidant Trolox (48.22 ± 0.79 µg/mL) with

insignificant difference. A correlation study of the extracts demonstrated very strong significant positive correlation between phenolic content (TPC) and antioxidant potential especially with DPPH ($r = 0.928$) and followed by FRAP ($r = 0.901$). Crude extracts of *H. parasitica* was found to be highly cytotoxic or antiproliferative to Michigan Cancer Foundation-7 breast cancer cells (MCF-7) when tested for MTT IC_{50} particularly with selected 100% acetone stem extract at $97.47 \pm 2.87 \mu\text{g/mL}$, while 5-fluorouracil used as positive standard revealed an IC_{50} of $3.31 \pm 0.15 \mu\text{g/mL}$. Interestingly, all the 3 tested extracts (100% acetone, leaf; 100% acetone, stem and 100% methanol, leaf) had no cytotoxic effect up to the maximum extract concentration tested on normal dermal fibroblast cells ($IC_{50} > 500 \mu\text{g/mL}$). Current study suggested that extraction solvents and plant parts (for Objective 1 and 3) played significant effect on extractability which depends to the preference of the research, either in terms of targeting maximal extract yield or active phytochemicals. This study proposed the presence of water with intermediate polar solvent (70% acetone) and leaf part could provide increased extraction yield while 100% acetone is suitable for bioactive compound recovery. Not forgotten, *H. parasitica* also revealed to be a potent source exhibiting anticancer activities but further research need to be done to purify and characterize the extract to identify the potent polar therapeutic compound or candidate against breast cancer. Meanwhile, this report is the first in describing total phenols, flavonoids content, extended assessment of antioxidant activity and anticancer property against breast cancer cell line from extracts of *H. parasitica* especially on species from Malaysia which may account for some of the medical claims attributed to this plant. Hence, this research and outcome may become a follow up from previous discontinued studies on the exploration of this potent source and lead for new nutraceutical and pharmaceutical approach.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**CIRI ANTIOKSIDAN DAN ANTIKANSER PELBAGAI EKSTRAK MENTAH
*DEDALU (Helixanthera parasitica Lour.)***

Oleh

THIRUBUVANESVARI DURAIVELU

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Helixanthera parasitica Lour sejenis pokok dedalu hemiparasit berasal daripada famili Loranthaceae dikenali sebagai tumbuhan unik yang digunakan secara meluas bagi tujuan ethnobotani ataupun sebagai tumbuhan ubatan di negara Asia Tenggara. Berdasarkan pengetahuan kami, terlalu sedikit penyelidikan saintifik yang dijalankan terhadap spesies ini terutamanya untuk mengeksplorasi potensinya sebagai sumber antioksidan dan keupayaannya untuk menentang salah satu punca utama kematian populasi dunia iaitu penyakit kanser payu dara yang kekal sebagai penyakit yang sukar diubati tetapi mampu dicegah. Dalam kajian ini, pelbagai ekstrak berasaskan daun dan batang *H. parasitica* telah diuji bagi mengetahui jumlah sebatian fenolik, flavonoid, aktiviti antioksidan dan juga pemilihan ekstrak terbaik bagi tujuan ujian antikanser dengan mengaplikasikan teknik yang sah. Pelbagai ekstrak mentah telah disediakan menerusi teknik rendaman dengan menggunakan serbuk daun dan batang. Proses pengekstrakan dijalankan dengan mengguna pakai pelarut yang berbagai kepolaran, iaitu air suling (dH_2O), metanol (70% dan 100%), aseton (70% dan 100%) dan petroleum eter, sekiranya disusun pada kadar polariti yang menurun. Teknik Jumlah Kandungan Fenolik (TPC), Jumlah Kandungan Flavonoid (TFC), 2, 2-difenil-1-pikrilhidrazil (DPPH), Kuasa Antioksidan Penurunan Ferik (FRAP) dan MTT (ujian antikanser) telah digunakan bagi memenuhi objektif kajian yang disasarkan. Hasil ekstrak tertinggi diperolehi daripada ekstrak daun $679.13 \pm 9.60 \text{ mg/g DW}$ dan pelarut pengekstrak terbaik dapat disusun pada kadar menurun iaitu 70% aseton > 70% metanol > 100% metanol > 100% aseton > dH_2O > petroleum eter. Aseton tulen (100%) dapat digelar sebagai pelarut terbaik dari segi Jumlah Kandungan Fenolik ($99.45 \pm 12.87 \text{ mg GAE/g DW}$), Jumlah Kandungan Flavonoid ($17.57 \pm 0.63 \text{ mg QE/g DW}$) dan peratusan 2, 2-difenil-2-pikrilhidrazil ($81.61 \pm 1.22 \%$) dengan tiada perbezaan signifikan yang dilihat dari segi daun dan batang. Ekstrak ini juga menunjukkan nilai Kuasa Antioksidan Penurunan Ferik ($6.73 \pm 0.36 \text{ mg TE/g DW}$) yang kurang sedikit berbanding batang, tetapi nilai tersebut juga tidak mempunyai perbezaan yang signifikan. Daripada sudut kepekatan ekstrak yang membawa kepada pengurangan 50% DPPH (IC_{50}), nilai atau kepekatan terendah diperolehi dengan

menggunakan ekstrak batang di dalam pelarut 100% aseton ($42.66 \pm 4.82 \mu\text{g/mL}$), tetapi nilai ini tidak mempunyai perbezaan signifikan jika dibandingkan dengan ekstrak 100% aseton daun ($45.21 \pm 4.91 \mu\text{g/mL}$) yang mana kedua-dua nilai ekstrak ini hampir sama dengan antioksidan piawai Trolox ($48.22 \pm 0.79 \mu\text{g/mL}$) yang turut tidak menunjukkan perbezaan signifikan. Kajian korelasi bagi ekstrak *H. parasitica* memaparkan korelasi signifikan positif yang sangat tinggi di antara kuantiti sebatian fenolik (TPC) dan aktiviti antioksidan terutamanya dengan DPPH ($r = 0.928$) dan diikuti dengan FRAP ($r = 0.901$). Ekstrak mentah *H. parasitica* menunjukkan aktiviti sitotoksik/antiproliferatif yang tinggi terhadap sel kanser payu dara Michigan Cancer Foundation-7 (MCF-7) semasa diuji dengan asai MTT di mana IC_{50} terendah diperolehi apabila ekstrak 100% aseton batang ($97.47 \pm 2.87 \mu\text{g/mL}$) digunakan, sementara 5-fluorouracil yang dijadikan piawai positif merekodkan nilai IC_{50} , $3.31 \pm 0.15 \mu\text{g/mL}$. Kesan ekstrak ke atas sel normal fibroblas pula tidak menunjukkan kematian sel sehingga pada kepekatan ekstrak maksimum yang diuji ($\text{IC}_{50} > 500 \mu\text{g/mL}$). Kajian ini mencadangkan pelarut pengekstrakan dan bahagian pokok (untuk Objektif 1 dan 3) memainkan peranan daripada segi kuantiti ekstrak dan jenis hasil yang diperolehi yang bergantung kepada objektif penyelidikan, sama ada untuk mendapatkan hasil ekstrak mentah maksimum atau menyasarkan sebatian bioaktifnya. Sehubungan dengan itu, kajian ini menunjukkan campuran komponen air bersama dengan pelarut sederhana polar berupaya menghasilkan jumlah ekstrak mentah yang maksima manakala pelarut tulen aseton adalah sesuai untuk mengekstrak sebatian bioaktif pokok *H. parasitica*. Tambahan lagi, ekstrak pokok ini juga menunjukkan potensi yang tinggi sebagai sumber antiproliferatif sel kanser payudara, dan penyelidikan tambahan perlu dijalankan untuk mengenalpasti agen bioaktif spesifik yang membawa kepada kesan sitotoksik ini dan seterusnya menjadi calon antikanser yang berpotensi. Selain itu, laporan ini merupakan manuskrip saintifik pertama yang menerajui jumlah sebatian fenolik, flavonoid, penambahan penilaian antioksidan dan aktiviti antikanser terhadap kanser payu dara bagi ekstrak mentah *H. parasitica* khususnya bagi spesies yang berasal dari Malaysia yang mana selari dengan pengetahuan perubatan tradisional yang dikaitkan dengan pokok ini. Di samping itu, kajian dan keputusan yang ditunjukkan mampu mengusul semula usaha untuk menerajui potensi perubatan pokok ini kepada penemuan nutraseutikal dan farmaseutikal.

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LIST OF ABBREVIATIONS

ANNOVA	Analysis of variance
BC	Breast cancer
CE	Catechin equivalent
CGM	Complete growth medium
°C	Degree celsius
DIM	Diindolylmethane
DMSO	Dimethyl sulfoxide
MTT	3-(4,5-dimethylthiazole-2-yl)-2,5-diphenyl tetrazolium bromide
DPPH	2,2-diphenyl-1-picrylhydrazyl
dH ₂ O	Distilled water
DW	Dry weight
DMEM	Dulbecco's modified eagle medium
EGCG	Epigallocatechin gallate
EMT	Epithelial-mesenchymal transition
E2F	Eukaryote transcriptional factor
FRAP	Ferric reducing antioxidant power
FBS	Fetal bovine serum
5-FU	5-fluorouracil
GC	Guanine-Cytosine
GAE	Gallic acid equivalent
G2/M	Gap 2/mitotic phase
g	Gram
100% Ace	Hundred percent acetone
100%	Hundred percent methanol
MeOH	
IC ₅₀	Inhibition concentration 50%
MCF-7	Michigan Cancer Foundation-7
μL	Microliter
mg	Milligram
mL	Milliliter
%	Percentage
Obj	Objective
PtEth	Petroleum ether
PBS	Phosphate buffer solution
QE	Quercetin equivalent
Rb	Retinoblastoma
rpm	Revolution per minute
70% Ace	Seventy percent acetone
70% MeOH	Seventy percent methanol
S	Synthesis phase
TA	Thymine-Adenine
TPTZ	2,4,6-tripyridyl-s-triazine
TFC	Total flavonoid content
TPC	Total phenolic content
TE	Trolox equivalent
w/v	Weight over volume

CHAPTER 1

INTRODUCTION

1.1 Overview of study

In the course of recent decades (over 30 years) there has been a developing substantial shift in disease burden from communicable to multiple non-communicable chronic diseases (NCDs) among world population (Murray et al., 2012). Nearly two-thirds of deaths recorded worldwide accounts due to NCDs (World Health Organization, 2013). Cancer or malignant growth is one of the leading major cause of death under NCDs which is generally described as an abnormal growth of tissue and cell arising from uncontrolled proliferation autonomously, ensuing in a progressive rise in the magnitude of cell divisions. Among them, breast cancer is the most frequent and number one leading cause of cancer among women that hits high death rate globally, particularly in non-developing nations because of deffered diagnosis and populace upsurge (Nordin, Kadir, Zakaria, Abdullah, & Abdullah, 2018). Oxidative stress is one of the phenomenon often associated with pathways related to carcinogenesis. Reactive free radical species like oxygen and nitrogen is crucial to mediate certain essential roles under a controlled regulation by the human body, but the risk arises when overproduction or an unregulated release is manifested in the form of various life-threatening disorders predominantly cancer.

There is no denial on the availability of existing natural and synthetic compounds which have the power to quench or scavenge these free radicals providing a great scope for correcting such oxidative imbalances. However, usage of synthetic antioxidants like Butylated Hydroxyanisole (BHA), butylated hydroxy toluene (BHT), propyl gallate (PG) and tert-butylhydroquinone causes an alarming concern on its toxicity and carcinogenicity emerged due to their extensive use which necessitates the venture to search for better alternatives.

Additionally, problem with combating against cancer also turns very challenging and requires strenuous effort, particularly in the development of treatments for aggressive and rapidly multiplying tumours. The ability of breast cancer cells inducing metastasis and being invasive as well as relapses as a result of having resistant disease (Early Breast Cancer Trialists' Collaborative Group, 2005) are often blamed as the factors liable for the mortality allied with this disease. Eventhough, existing chemotherapy has implicated relevant outcome on cancer treatment, it exhibits low specificity and is restricted by dose-limiting toxicity (Khandanlou, Murthy, Saranath, & Damani, 2018) and the worst complication arises when drug resistance phenomenon takes over. Even, some analogs that have given a major renewable natural source and mostly utilized in breast cancer treatment was then known to be unproductive in some cases after a period of time which further urge an extensive exploration from natural source.

Alternative traditional and complementary treatment using medicines derived from herbal and natural products such as Ayurveda and Chinese medicine have been used widely in every culture across different times and civilization since ancient immemorial times globally. Asian countries specifically Thailand, India, Indonesia, China, and Malaysia are exceptionally well-known for precious ethnobotanical knowledges. Moreover, Malaysia sets its name to be one of the 17 mega biodiversity countries in the world, enriched in biodiversity as a potential source of both primary and secondary therapeutic metabolites like phenols and flavonoids which exhibit myriad biological properties. Emergence of increasing evidences which indicates that medicinal plant-derived products inhibit tumorigenesis and related to inflammatory processes at various stages emphasizes the significance of these products in chemotherapy and also as therapeutic agents targeted for free radical induced pathologies and disease prevention. Approximately 65% of all the approved drugs used in the treatment namely chemotherapy or prevention of cancers originates from natural products or natural product derivatives (Nurhanan et al., 2008), of which plants contribute around 25% (Balunas & Kinghorn, 2005).

Natural antioxidants or drugs apart from being safe and causing minimal adverse effect, these compounds receive great attention because of its economic affordability as well as being more efficient than their synthetic counterparts. In addition, not only a plethora of drugs are of plant origin but also the entire plant extracts in some cases could be used as promising therapeutic modalities taking into consideration their bioactivity as well as target specificity (Bhat, Gul, Lohamror, Qureshi, & Ghazi, 2018). Other than that, the use of plant extracts may impart an advantage over using one isolated compound as drugs, due to the availability of various components possessing numerous possible intracellular targets which may work individually or synergistically to combat diseases induced by oxidative stress and improving health (Nordin et al., 2018). Furthermore, complementary practises such as consuming vitamins or herbs for cancer treatment or to relieve its side effects is not something neoteric.

Therefore, this has invoked the quest for continuous search targeting wild species of plant exhibiting unique behavior such as parasitism, for naturally occurring novel drugs providing high efficacy to drug resistant tumors with minimal side effects.

1.2 Problem statements

Mistletoe, a group of parasitic plant species used as medicinal herbs are not sufficiently explored and studied in Malaysia for their therapeutics and pharmacological properties. Mistletoes are often and typically utilized by traditional healers as traditional medicines without any written proof who need to hide the formulas, plant types and treatments from the public (Kwanda, Noikotr, Sudmoon, Tanee, & Chaveerach, 2013). Additionally, parasitic plants are potential traditional medicine through fatal to their hosts. Keeping this view in mind, the current attempt was directed on therapeutic exploration on Malaysian mistletoe, *Helixanthera parasitica* Lour. which is known to be a hemiparasitic shrub in the family of Loranthaceae.

This shrub is chosen due its ethnopharmacological significance, identified therapeutic property and phytochemicals discovered, apart from other potent antioxidative compounds. However, the information gathered on this potential mistletoe shrub remains scarce due to very few scientific researches conducted. It is important to notify that, the last study performed on *H. parasitica* dates 15 years ago in Thailand and another significant study on its phytochemistry dates back in 1994 originating from China and no publication is revealed from Malaysia. However, those limited studies focused on scientific experimentation of its therapeutic and phytochemistry without considering extract preparation and plant parts. Selection of a suitable extraction solvent plays a role for the standardization of herbal products as it is utilized in the removal of desirable soluble constituents, leaving out those not required. Further, different plant part of *H. parasitica* were evaluated for phytochemical values in order to valorize all the plant as functional source of extraction. Optimization of parameters are critical for upscaling purposes to bring *H. parasitica* from bench scale to pilot plant level and subsequently to be adapted readily in clinical research program. Hence, post-harvest treatment variability namely extraction solvents and profiling the composition of different plant part extracts are applied in this research taking into consideration of its great significance towards obtaining a maximum yield of phytocompounds especially antioxidants that correspond to the therapeutic potential of local *H. parasitica*.

Additionally, thus far, no empirical submission has been reported on the antioxidative levels thoroughly and cytotoxic properties of any *Helixanthera* species against breast cancer cell. Other than being used for folkhore benefits, scientific reports exposing the therapeutic potential of *H. parasitica* on anticancer responses and newly discovered compound prompted us to hypothesize that *H. parasitica* plant may exhibit vast range of antioxidant and antiproliferative activities. With this background and in the view of exploration for an alternative medicine, *H. parasitica* was tested to fulfil the imperative need to discover the real potential of the local herbs through its naturally occurring phytocompounds for antioxidant and anti-breast cancer activity. It is important when looking at the side effects of current treatment, damages upon healthy cells and rising therapeutic resistances. Current research also taken to support ideas of using plants synergistically with the available modern drugs to subsequently improve the effect on pharmacological, toxicological and prognosis of treatment if this research and extended studies proves it to be safe and effective. This effort will demonstrate a scientific documentary on post-harvest prep and therapeutic properties of one of the Malaysian mistletoe *H. parasitica*, having a potential medicinal value, that could lead towards drug discovery and being handfull in human healthcare.

1.3 Objectives of study

Thus, the objectives of this study are:

1. To determine the best extraction solvent and plant part based on optimum extract yield obtained from *H. parasitica*.
2. To determine the best extract based on extraction solvent and plant part by quantifying phenolic and flavonoid compounds as well as antioxidant potential.

3. To determine the anticancer potential and cytotoxicity effect on breast cancer (MCF-7) and healthy fibroblast cell lines using selected extracts of *H. parasitica* based on phenolic quantification, flavonoid contents and antioxidant activity.



REFERENCES

- Abaza, L., Youssef, N. B., Manai, H., Haddada, F. M., Methenni, K., & Zarrouk, M. (2011). Chétoui olive leaf extracts: influence of the solvent type on phenolics and antioxidant activities. *Grasas y Aceites*, 62(1), 96-104.
- Abdulkadira, A. R., Mata, N., Hasanb, M. M., & Jahana, M. S. (2016). In vitro antioxidant activity of the ethanolic extract from fruit, stem, and leaf of *Solanum torvum*. *Science Asia*, 42(3), 184-189.
- Abugri, D. A., Tiimob, B. J., Apalangya, V. A., Pritchett, G., & McElhenney, W. H. (2013). Bioactive and nutritive compounds in *Sorghum bicolor* (Guinea corn) red leaves and their health implication. *Food Chemistry*, 138(1), 718-723.
- Acharya, A., Das, I., Chandhok, D., & Saha, T. (2010). Redox regulation in cancer: a double-edged sword with therapeutic potential. *Oxidative Medicine & Cellular Longevity*, 3(1), 23-34.
- Agati, G., Matteini, P., Goti, A., & Tattini, M. (2007). Chloroplast-located flavonoids can scavenge singlet oxygen. *New Phytologist*, 174(1), 77-89.
- Ahmad, N., & Mukhtar, H. (1999). Green tea polyphenols and cancer: biologic mechanisms and practical implications. *Nutrition Reviews*, 57(3), 78-83.
- Ahmadian-Kouchaksaraie, Z., Niazmand, R., & Najafi, M. N. (2016). Optimization of the subcritical water extraction of phenolic antioxidants from *Crocus sativus* petals of saffron industry residues: Box-Behnken design and principal component analysis. *Innovative Food Science & Emerging Technologies*, 36, 234-244.
- Ajabnoor, G., Macanas-Pirard, P., Shotton, C., & Coley, H. (2008). Paclitaxel-resistant MCF-7 cells show a caspase-less phenotype but retain sensitivity to many anticancer agents. *American Association for Cancer Research*, 68(9), 2657
- Aksoy, L., Kolay, E., Ağılönü, Y., Aslan, Z., & Kargioğlu, M. (2013). Free radical scavenging activity, total phenolic content, total antioxidant status, and total oxidant status of endemic *Thermopsis turcica*. *Saudi Journal of Biological Sciences*, 20(3), 235-239.
- Al-Juhaimi, F., & Ghafoor, K. A. S. H. I. F. (2011). Total phenols and antioxidant activities of leaf and stem extracts from coriander, mint and parsley grown in Saudi Arabia. *Pakistan Journal of Botany*, 43(4), 2235-2237.
- Albrecht, H., Yoder, J. I., & Phillips, D. A. (1999). Flavonoids Promote Haustoria Formation in the Root Parasite *Triphysaria versicolor*. *Plant Physiology*, 119(2), 585-592.

- Ali, A.A. & Alqurainy F. (2006). Activities of antioxidants in plants under environmental stress. In: Motohashi N, editor. *The lutein-prevention and treatment for diseases* (p. 187–256). India: Transworld Research Network.
- Allothman, M., Bhat, R., & Karim, A. A. (2009). Antioxidant capacity and phenolic content of selected tropical fruits from Malaysia, extracted with different solvents. *Food Chemistry*, 115(3), 785-788.
- Alsarhan, A., Sultana, N., Al-Khatib, A., & Kadir, M. R. A. (2014). Review on some Malaysian traditional medicinal plants with therapeutic properties. *Journal of Basic & Applied Sciences*, 10, 149.
- Amabeoku, G. J., Leng, M. J., & Syce, J. A. (1998). Antimicrobial and anticonvulsant activities of *Viscum capense*. *Journal of Ethnopharmacology*, 61(3), 237-241.
- Ameer Omar, Z., Salman Ibrahim, M., & Quek Ko Jin, A. M. (2015). *Loranthus ferrugineus*: a mistletoe from traditional uses to laboratory bench. *Journal of Pharmacopuncture*, 18(1), 7-18.
- Anokwuru, C. P., Anyasor, G. N., Ajibaye, O., Fakoya, O., & Okebugwu, P. (2011). Effect of extraction solvents on phenolic, flavonoid and antioxidant activities of three nigerian medicinal plants. *Nature & Science*, 9(7), 53-61.
- Anokwuru, C. P., Esiaba, I., Ajibaye, O., & Adesuyi, A. O. (2011). Polyphenolic content and antioxidant activity of *Hibiscus sabdariffa* calyx. *Research Journal of Medicinal Plant*, 5(5), 557-566.
- Antoniou, A. C., & Easton, D. F. (2006). Models of genetic susceptibility to breast cancer. *Oncogene*, 25(43), 5898-5905.
- Anwar, F., Jamil, A., Iqbal, S., & Sheikh, M. A. (2006). Antioxidant activity of various plant extracts under ambient and accelerated storage of sunflower oil. *Grasas y Aceites*, 57(2), 189-197.
- Anwar, F., Kalsoom, U., Sultana, B., Mushtaq, M., Mehmood, T., & Arshad, H. A. (2013). Effect of drying method and extraction solvent on the total phenolics and antioxidant activity of cauliflower (*Brassica oleracea* L.) extracts. *International Food Research Journal*, 20(2), 653-659.
- Arnous, A., Makris, D. P., & Kefalas, P. (2002). Correlation of pigment and flavanol content with antioxidant properties in selected aged regional wines from Greece. *Journal of Food Composition & Analysis*, 15(6), 655-665.
- Aron, P. M., & Kennedy, J. A. (2008). Flavan-3-ols: Nature, occurrence and biological activity. *Molecular Nutrition & Food Research*, 52(1), 79-104.
- Atienza, J. M., Yu, N., Wang, X., Xu, X., & Abassi, Y. (2006). Label-free and real-time cell-based kinase assay for screening selective and potent receptor tyrosine kinase inhibitors using microelectronic sensor array. *Journal of Biomolecular Screening*, 11(6), 634-643.

- Aziz, A., Zulkiflee, A.B. & Yusof, F.Z.M. (2017, December) Concentration and time dependent cytotoxic effect of methanolic crude extracts of pseuduvaria macrophylla on the human cancer cell line. Paper presented at the *International Congress on Sciences & Technology 2017* (ICST 2017), Bali, Indonesia. Retrieved from http://eprints.um.edu.my/18510/1/Atiqah_Bali_paper_oct_2017_revised.pdf.
- Baba, S. A., & Malik, S. A. (2015). Determination of total phenolic and flavonoid content, antimicrobial and antioxidant activity of a root extract of *Arisaema jacquemontii* Blume. *Journal of Taibah University for Science*, 9(4), 449-454.
- Badar, F., Faruqui, Z. S., Uddin, N., & Trevan, E. A. (2011). Management of breast lesions by breast physicians in a heavily populated South Asian developing country. *Asian Pacific Journal of Cancer Prevention*, 12(3), 827-32.
- Badr, J. M. (2015). Chemical constituents of *Phragmanthera austroarabica* AG Mill and JA Nyberg with potent antioxidant activity. *Pharmacognosy Research*, 7(4), 335.
- Bagchi, D., Bagchi, M., Stohs, S. J., Das, D. K., Ray, S. D., Kuszynski, C. A., ... & Pruess, H. G. (2000). Free radicals and grape seed proanthocyanidin extract: importance in human health and disease prevention. *Toxicology*, 148(2-3), 187-197.
- Bai, M., Liu, S. F., Wang, W., Wang, X. B., Huang, X. X., & Song, S. J. (2017). Two new lactones from whole herbs of *Patrinia villosa* Juss. *Phytochemistry Letters*, 22, 145-148.
- Baldwin, A. S. (2001). Control of oncogenesis and cancer therapy resistance by the transcription factor NF- κ B. *The Journal of Clinical Investigation*, 107(3), 241-246.
- Balunas, M. J., & Kinghorn, A. D. (2005). Drug discovery from medicinal plants. *Life Sciences*, 78(5), 431-441.
- Banerjee, S. K., & Bonde, C. G. (2011). Total phenolic content and antioxidant activity of extracts of *Bridelia retusa* Spreng Bark: Impact of dielectric constant and geographical location. *Journal of Medicinal Plants Research*, 5(5), 817-822.
- Bar-Sela, G. (2011). White-Berry Mistletoe (*Viscum album* L.) as complementary treatment in cancer: Does it help?. *European Journal of Integrative Medicine*, 3(2), e55-e62.
- Barboza, G. E., Cantero, J. J., Núñez, C., Pacciaroni, A., & Ariza Espinar, L. (2009). Medicinal plants: A general review and a phytochemical and ethnopharmacological screening of the native Argentine Flora. *Kurtziana*, 34(1-2), 7-365.
- Barhé, T. A., & Tchouya, G. F. (2016). Comparative study of the anti-oxidant activity of the total polyphenols extracted from *Hibiscus sabdariffa* L., *Glycine max* L.

- Merr., yellow tea and red wine through reaction with DPPH free radicals. *Arabian Journal of Chemistry*, 9(1), 1-8.
- Barlow, B. (2012). *What is a mistletoe ?* Retrieved from <http://www.cpbr.gov.au/mistletoe/what-is-mistletoe.html>.
- Barlow, B. A. (1983). Biogeography of Loranthaceae and Viscaceae. In: M. Calder & P. Bernhardt (Eds.), *The Biology of Mistletoe* (pp. 19-46). Sydney: Academic Press.
- Beckers, G. J. M., & Spoel, S. H. (2006). Fine-tuning plant defence signalling: salicylate versus jasmonate. *Plant biology*, 8(1), 1-10.
- Beevi, S. S., Narasu, M. L., & Gowda, B. B. (2010). Polyphenolics profile, antioxidant and radical scavenging activity of leaves and stem of *Raphanus sativus* L. *Plant Foods for Human Nutrition*, 65(1), 8-17.
- Benzie, I. F., & Strain, J. J. (1996). The ferric reducing ability of plasma (FRAP) as a measure of “antioxidant power”: the FRAP assay. *Analytical biochemistry*, 239(1), 70-76.
- Bezerra, A. N. S., Massing, L. T., de Oliveira, R. B., & Mourão, R. H. V. (2017). Standardization and anti-inflammatory activity of aqueous extract of *Psittacanthus plagiophyllus* Eichl. (Loranthaceae). *Journal of Ethnopharmacology*, 202, 234-240.
- Bhat, M. Y., Gul, M. Z., Lohamror, L. R., Qureshi, I. A., & Ghazi, I. A. (2018). An in vitro Study of the Antioxidant and Antiproliferative Properties of *Artemisia absinthium*-A Potent Medicinal Plant. *Free Radicals & Antioxidants*, 8(1), 18-25.
- Bhatt, A. P., Redinbo, M. R., & Bultman, S. J. (2017). The role of the microbiome in cancer development and therapy. *CA: A Cancer Journal for Clinicians*. 67(4), 326-344.
- Bhattacharyya, A., Chattopadhyay, R., Mitra, S., & Crowe, S. E. (2014). Oxidative stress: an essential factor in the pathogenesis of gastrointestinal mucosal diseases. *Physiological Reviews*, 94(2), 329-354.
- Bhebbhe, M., Füller, T. N., Chipurura, B., & Muchuweti, M. (2016). Effect of solvent type on total phenolic content and free radical scavenging activity of black tea and herbal infusions. *Food Analytical Methods*, 9(4), 1060-1067.
- Biesaga, M. (2011). Influence of extraction methods on stability of flavonoids. *Journal of Chromatography A*, 1218(18), 2505-2512.
- Blois, M. S. (1958). Antioxidant determinations by the use of a stable free radical. *Nature*, 181(4617), 1199.
- Boeing, J. S., Barizão, É. O., e Silva, B. C., Montanher, P. F., de Cinque Almeida, V., & Visentainer, J. V. (2014). Evaluation of solvent effect on the extraction of

- phenolic compounds and antioxidant capacities from the berries: application of principal component analysis. *Chemistry Central Journal*, 8(1), 48.
- Bolton, J. L., Trush, M. A., Penning, T. M., Dryhurst, G., & Monks, T. J. (2000). Role of quinones in toxicology. *Chemical Research in Toxicology*, 13(3), 135-160.
- Borchardt, J. K. (2002). The beginnings of drug therapy: Ancient mesopotamian medicine. *Drug News Perspect*, 15(3), 187-192.
- Borneo, R., León, A. E., Aguirre, A., Ribotta, P., & Cantero, J. J. (2009). Antioxidant capacity of medicinal plants from the Province of Córdoba (Argentina) and their in vitro testing in a model food system. *Food Chemistry*, 112(3), 664-670.
- Brewer, M. S. (2011). Natural antioxidants: sources, compounds, mechanisms of action, and potential applications. *Comprehensive Reviews in Food Science & Food Safety*, 10(4), 221-247.
- Brown, J. E., & Kelly, M. F. (2007). Inhibition of lipid peroxidation by anthocyanins, anthocyanidins and their phenolic degradation products. *European Journal of Lipid Science & Technology*, 109(1), 66-71.
- Brown, N. S., Jones, A., Fujiyama, C., Harris, A. L., & Bicknell, R. (2000). Thymidine phosphorylase induces carcinoma cell oxidative stress and promotes secretion of angiogenic factors. *Cancer Research*, 60(22), 6298-6302.
- Bucić-Kojić, A., Planinić, M., Tomas, S., Bilić, M., & Velić, D. (2007). Study of solid–liquid extraction kinetics of total polyphenols from grape seeds. *Journal of Food Engineering*, 81(1), 236-242.
- Buergers, R., Rosentritt, M., & Handel, G. (2007). Bacterial adhesion of *Streptococcus mutans* to provisional fixed prosthodontic material. *Journal of Prosthetic Dentistry*, 98(6), 461-469.
- Buetti-Dinh, A., Pivkin, I. V., & Friedman, R. (2015a). S100A4 and its role in metastasis—computational integration of data on biological networks. *Molecular BioSystems*, 11(8), 2238-2246.
- Buetti-Dinh, A., Pivkin, I. V., & Friedman, R. (2015b). S100A4 and its role in metastasis—simulations of knockout and amplification of epithelial growth factor receptor and matrix metalloproteinases. *Molecular BioSystems*, 11(8), 2247-2254.
- Bujor, O. C., Giniès, C., Popa, V. I., & Dufour, C. (2018). Phenolic compounds and antioxidant activity of lingonberry (*Vaccinium vitis-idaea* L.) leaf, stem and fruit at different harvest periods. *Food Chemistry*. 252, 356-365.
- Burkill, I. H. (1966). *A dictionary of the economic products of the Malay Peninsula*. (Vol. 2). Kuala Lumpur, Malaysia: Ministry of Agriculture and Cooperatives.

- Bussing, A. (2000) *Mistletoe: the genus Viscum, Medicinal and Aromatic Plants*. Amsterdam, Netherland: Harwood Academic.
- Büssing, A., Schaller, G., & Pfüller, U. (1998). Generation of reactive oxygen intermediates (ROI) by the thionins from *Viscum album* L. *Anticancer Research*, 18(6A), 4291-4296.
- Büssing, A., Vervecken, W., Wagner, M., Wagner, B., Pfüller, U., & Schietzel, M. (1999). Expression of mitochondrial Apo2. 7 molecules and caspase-3 activation in human lymphocytes treated with the ribosome-inhibiting mistletoe lectins and the cell membrane permeabilizing viscotoxins. *Cytometry: The Journal of the International Society for Analytical Cytology*, 37(2), 133-139.
- Cacace, J. E., & Mazza, G. (2003). Optimization of extraction of anthocyanins from black currants with aqueous ethanol. *Journal of Food Science*, 68(1), 240-248.
- Cai, X., Xiao, C., Xue, H., Xiong, H., Hang, Y., Xu, J., & Lu, Y. (2018). A comparative study of the antioxidant and intestinal protective effects of extracts from different parts of Java tea (*Orthosiphon stamineus*). *Food Science & Nutrition*, 6(3), 579-584.
- Cairns, R.A., Harris, I., McCracken, S., & Mak, T.W. (2011). Cancer cell metabolism. *Cold Spring Harbor Symposia Quantitative Biology*, 76, 299–311.
- Calder, D. M. (1983). Mistletoes in focus: an introduction. *The Biology of Mistletoes*, 4, 1-18.
- Cao, G., Sofic, E., & Prior, R. L. (1997). Antioxidant and prooxidant behavior of flavonoids: structure-activity relationships. *Free Radical Biology & Medicine*, 22(5), 749-760.
- Caragay, A. B. (1992). Cancer-preventive foods and ingredients. *Food Technology*, 46(4), 65-68.
- Castro-Vargas, H. I., Rodríguez-Varela, L. I., Ferreira, S. R., & Parada-Alfonso, F. (2010). Extraction of phenolic fraction from guava seeds (*Psidium guajava* L.) using supercritical carbon dioxide and co-solvents. *The Journal of Supercritical Fluids*, 51(3), 319-324.
- Cemaluk, E. A., & Nwankwo, N. E. (2012). Phytochemical properties of some solvent fractions of petroleum ether extract of the African mistletoe (*Loranthus micranthus* Linn) leaves and their antimicrobial activity. *African Journal of Biotechnology*, 11(62), 12595-12599.
- Chen, Z. (2008). Research of antioxidative capacity in essential oils of plants. *China Condiment*, 11, 40–43
- Choi, S. H., Lee, S. H., Kim, H. J., Lee, I. S., Kozukue, N., Levin, C. E., & Friedman, M. (2010). Changes in free amino acid, phenolic, chlorophyll, carotenoid, and glycoalkaloid contents in tomatoes during 11 stages of growth and inhibition

- of cervical and lung human cancer cells by green tomato extracts. *Journal of Agricultural & Food Chemistry*, 58(13), 7547-7556.
- Coleman, M. P., Quaresma, M., Berrino, F., Lutz, J. M., De Angelis, R., Capocaccia, R., ... & the CONCORD Working Group (2008). Cancer survival in five continents: a worldwide population-based study (CONCORD). *The Lancet Oncology*, 9(8), 730-756.
- Coley, H. M. (2009). Mechanisms and consequences of chemotherapy resistance in breast cancer. *European Journal of Cancer Supplements*, 7(1), 3-7.
- Corlett, R. T. (2005). Interactions between birds, fruit bats and exotic plants in urban Hong Kong, South China. *Urban Ecosystems*, 8(3-4), 275-283.
- Cragg, G. M., & Newman, D. J. (2002). Drugs from nature: past achievements, future prospects. In *Advances in Phytomedicine* (Vol. 1, pp. 23-37). New York, USA: Elsevier.
- Cragg, G. M., & Newman, D. J. (2013). Natural products: a continuing source of novel drug leads. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1830(6), 3670-3695.
- Cumming, R. C., Lightfoot, J., Beard, K., Youssoufian, H., O'Brien, P. J., & Buchwald, M. (2001). Fanconi anemia group C protein prevents apoptosis in hematopoietic cells through redox regulation of GSTP1. *Nature Medicine*, 7(7), 814.
- Cuvelier, M. E., Maillard, M. N., & Berset, C. (1996, September). *Synergistic and antagonistic effect in pure phenolic antioxidant mixtures and in plant extracts*. Paper presented at the 1st Congress of the European Section of AOCS, Dijon, France.
- Dalar, A., Türker, M., & Konczak, I. (2012). Antioxidant capacity and phenolic constituents of *Malva neglecta* Wallr. and *Plantago lanceolata* L. from Eastern Anatolia Region of Turkey. *Journal of Herbal Medicine*, 2(2), 42-51.
- Dai, J., & Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15(10), 7313-7352.
- Darwin, C. (1859). On the origins of species by means of natural selection. *London: Murray*, 247, 1859.
- Das, N., Islam, M. E., Jahan, N., Islam, M. S., Khan, A., Islam, M. R., & Parvin, M. S. (2014). Antioxidant activities of ethanol extracts and fractions of *Crescentia cujete* leaves and stem bark and the involvement of phenolic compounds. *BMC Complementary & Alternative Medicine*, 14(1), 45.
- Daud, A., Gallo, A., & Riera, A. S. (2005). Antimicrobial properties of *Phrygilanthus acutifolius*. *Journal of Ethnopharmacology*, 99(2), 193-197.

- Deeni, Y. Y., & Sadiq, N. M. (2002). Antimicrobial properties and phytochemical constituents of the leaves of African mistletoe (*Tapinanthus dodoneifolius* (DC) Danser) (Loranthaceae): an ethnomedicinal plant of Hausaland, Northern Nigeria. *Journal of Ethnopharmacology*, 83(3), 235-240.
- del Baño, M. J., Lorente, J., Castillo, J., Benavente-García, O., del Río, J. A., Ortuño, A., ... & Gerard, D. (2003). Phenolic diterpenes, flavones, and rosmarinic acid distribution during the development of leaves, flowers, stems, and roots of *Rosmarinus officinalis*. Antioxidant activity. *Journal of Agricultural & Food Chemistry*, 51(15), 4247-4253.
- Delaviz, H., Mohammadi, J., Ghalamfarsa, G., Mohammadi, B., & Farhadi, N. (2017). A review study on phytochemistry and pharmacology applications of *Juglans regia* plant. *Pharmacognosy Reviews*, 11(22), 145.
- Deng, X. H., Song, H. Y., Zhou, Y. F., Yuan, G. Y., & Zheng, F. J. (2013). Effects of quercetin on the proliferation of breast cancer cells and expression of survivin in vitro. *Experimental & Therapeutic Medicine*, 6(5), 1155-1158.
- Deng, Y., Zhao, Y., Padilla-Zakour, O., & Yang, G. (2015). Polyphenols, antioxidant and antimicrobial activities of leaf and bark extracts of *Solidago canadensis* L. *Industrial Crops & Products*, 74, 803-809.
- Devkota, M. P., & Kunwar, R. M. (2006). Diversity, distribution and host range of mistletoes in Godawari-Phulchoki Area, Kathmandu, Nepal. *Journal of Japanese Botany*, 81(5), 255.
- Dizdaroglu, M., Jaruga, P., Birincioglu, M., & Rodriguez, H. (2002). Free radical-induced damage to DNA: mechanisms and measurement1, 2. *Free Radical Biology & Medicine*, 32(11), 1102-1115.
- Djeridane, A., Yousfi, M., Nadjemi, B., Boutassouna, D., Stocker, P., & Vidal, N. (2006). Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds. *Food Chemistry*, 97(4), 654-660.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., & Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of Food & Drug Analysis*, 22(3), 296-302.
- Dobiáš, P., Pavlíková, P., Adam, M., Eisner, A., Beňová, B., & Ventura, K. (2010). Comparison of pressurised fluid and ultrasonic extraction methods for analysis of plant antioxidants and their antioxidant capacity. *Open Chemistry*, 8(1), 87-95.
- Doughari, J. H. (2012). Phytochemicals: Extraction methods, basic structures and mode of action as potential chemotherapeutic agents. In V. Rao (Ed.). *Phytochemicals-A global perspective of their role in nutrition and health* (pp. 1-32). Retrieved from <http://www.intechopen.com/books/phytochemicals-a-global-perspective-of-their-role-in-nutrition-and-health/phytochemicalsextract>

ion-methods-basic-structures-and-mode-of-action-as-potentialchemotherapeuti
c-

- Droge, W. (2002). Free radicals in the physiological control of cell function. *Physiological Reviews*, 82(1), 47-95.
- Du, M. Q., Carmichael, P. L., & Phillips, D. H. (1994). Induction of activating mutations in the human c-Ha-ras-1 proto-oncogene by oxygen free radicals. *Molecular Carcinogenesis*, 11(3), 170-175.
- Duarte, T. L., Cooke, M. S., & Jones, G. D. (2009). Gene expression profiling reveals new protective roles for vitamin C in human skin cells. *Free Radical Biology & Medicine*, 46(1), 78-87.
- Duo, J., Ying, G. G., Wang, G. W., & Zhang, L. (2012). Quercetin inhibits human breast cancer cell proliferation and induces apoptosis via Bcl-2 and Bax regulation. *Molecular Medicine Reports*, 5(6), 1453-1456.
- Early Breast Cancer Trialists' Collaborative Group. (2005). Effects of chemotherapy and hormonal therapy for early breast cancer on recurrence and 15-year survival: an overview of the randomised trials. *The Lancet*, 365(9472), 1687-1717.
- Fan, W., Chang, J., & Fu, P. (2015). Endocrine therapy resistance in breast cancer: current status, possible mechanisms and overcoming strategies. *Future Medicinal Chemistry*, 7(12), 1511-1519.
- Ferguson, L. R. (2010). Chronic inflammation and mutagenesis. *Mutation Research/Fundamental & Molecular Mechanisms of Mutagenesis*, 690(1), 3-11.
- Ferhat, R., Lekbir, A., Ouadah, H., Kahoul, M. A., Khlalfa, L., Laroui, S., & Alloui-Lombarkia, O. (2017). Effect of extraction solvent on total phenolic content, total flavonoid content, and antioxidant activities of Algerian pomace olive oil. *International Food Research Journal*, 24(6), 2295-2303.
- Ferlay, J., Shin, H. R., Bray, F., Forman, D., Mathers, C., & Parkin, D. M. (2010). Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *International Journal of Cancer*, 127(12), 2893-2917.
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., ... & Bray, F. (2015). Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *International Journal of Cancer*, 136(5), E359-E386.
- Fernández-Agulló, A., Pereira, E., Freire, M. S., Valentao, P., Andrade, P. B., González-Álvarez, J., & Pereira, J. A. (2013). Influence of solvent on the antioxidant and antimicrobial properties of walnut (*Juglans regia* L.) green husk extracts. *Industrial Crops & Products*, 42, 126-132.
- Fernández, T., Wagner, M. L., Varela, B. G., Ricco, R. A., Hajos, S. E., Gurni, A. A., & Alvarez, E. (1998). Study of an Argentine Mistletoe, the hemiparasite

- Ligaria cuneifolia* (R. et P.) Tiegh. (Loranthaceae). *Journal of Ethnopharmacology*, 62(1), 25-34.
- Ferruzzi, M. G., Böhm, V., Courtney, P. D., & Schwartz, S. J. (2002). Antioxidant and antimutagenic activity of dietary chlorophyll derivatives determined by radical scavenging and bacterial reverse mutagenesis assays. *Journal of Food Science*, 67(7), 2589-2595.
- Flockhart, D. A., & Meyer, U. A. (2005). Clinical Perspectives. In *Pharmacogenomics*, (2nd ed.) (pp. 247-263). Boca Raton, FL: Taylor & Francis
- Florence, T.M. (1995). The role of free radicals in disease. *Australian & New Zealand Journal Ophthalmology*, 23(1), 3-7.
- Frankel, E. N. (2005). *Lipid oxidation* (2nd ed.). Cambridge, UK: Oily Press.
- Fратиanni, F., Tucci, M., De Palma, M., Pepe, R., & Nazzaro, F. (2007). Polyphenolic composition in different parts of some cultivars of globe artichoke (*Cynara cardunculus* L. var. scolymus (L.) Fiori). *Food Chemistry*, 104(3), 1282-1286.
- Frei B. (1997). Reactive oxygen species and antioxidant vitamins. Linus Pauling Institute. Oregon State University. Retrieved from <http://lpi.oregonstate.edu/fw97/reactive.html>.
- Friedman, R. (2016). Drug resistance in cancer: molecular evolution and compensatory proliferation. *Oncotarget*, 7(11), 11746.
- Fruehauf, J. P., & Meyskens, F. L. (2007). Reactive oxygen species: a breath of life or death? *Clinical Cancer Research*, 13(3), 789-794.
- Fujii, T., Wakaizumi, M., Ikami, T., & Saito, M. (2008). Amla (*Embllica officinalis* Gaertn.) extract promotes procollagen production and inhibits matrix metalloproteinase-1 in human skin fibroblasts. *Journal of Ethnopharmacology*, 119(1), 53-57.
- Fukunaga, T., Nishiya, K., Kajikawa, I., Takeya, K. & Itokawa H. (1989). Studies on the constituents of Japanese mistletoes from different host trees, and their antimicrobial and hypotensive properties. *Chemical & Pharmaceutical Bulletin*, 37(6), 1543-1546.
- Galanakis, C. M., Goulas, V., Tsakona, S., Manganaris, G. A., & Gekas, V. (2013). A knowledge base for the recovery of natural phenols with different solvents. *International Journal of Food Properties*, 16(2), 382-396.
- Galati, G., & O'brien, P. J. (2004). Potential toxicity of flavonoids and other dietary phenolics: significance for their chemopreventive and anticancer properties. *Free Radical Biology & Medicine*, 37(3), 287-303.
- García-Pérez, J., López-Abente, G., Gómez-Barroso, D., Morales-Piga, A., Romaguera, E. P., Tamayo, I., Fernández-Navarroab, P. & Ramis, R. (2015). Childhood

- leukemia and residential proximity to industrial and urban sites. *Environmental Research*, 140, 542-553.
- Garcia-Salas, P., Morales-Soto, A., Segura-Carretero, A., & Fernández-Gutiérrez, A. (2010). Phenolic-compound-extraction systems for fruit and vegetable samples. *Molecules*, 15(12), 8813-8826.
- Gayathri, P., Dhana Sekar, V., Sudhakar, K., Sangeetha, M., & Chamundeeswari, D. (2018). Effect of ethanolic extract of *Reissantia indica* on the human breast cancer cell line (MCF-7). *The Journal of Phytopharmacology*, 7(1), 10-12.
- Genestra, M. (2007). Oxyl radicals, redox-sensitive signalling cascades and antioxidants. *Cellular Signalling*, 19(9), 1807-1819.
- Ghagane, S. C., Puranik, S. I., Kumbar, V. M., Nerli, R. B., Jalalpure, S. S., Hiremath, M. B., ... & Aladakatti, R. (2017). In vitro antioxidant and anticancer activity of *Leea indica* leaf extracts on human prostate cancer cell lines. *Integrative Medicine Research*, 6(1), 79-87.
- Goldenberg, M. M. (1999). Trastuzumab, a recombinant DNA-derived humanized monoclonal antibody, a novel agent for the treatment of metastatic breast cancer. *Clinical Therapeutics*, 21(2), 309-318.
- González, R., Ballester, I., López-Posadas, R., Suárez, M. D., Zarzuelo, A., Martinez-Augustin, O., & Medina, F. S. D. (2011). Effects of flavonoids and other polyphenols on inflammation. *Critical reviews in food science and nutrition*, 51(4), 331-362.
- Gopalakrishnan, A., Ram, M., Kumawat, S., Tandan, S. K., & Kumar, D. (2016). Quercetin accelerated cutaneous wound healing in rats by increasing levels of VEGF and TGF- β 1. *Indian Journal of Experimental Biology*, 54(3), 187-195.
- Gray, A. M., & Flatt, P. R. (1999). Insulin-secreting activity of the traditional antidiabetic plant *Viscum album* (mistletoe). *Journal of Endocrinology*, 160(3), 409-414.
- Gupta, S. C., Hevia, D., Patchva, S., Park, B., Koh, W., & Aggarwal, B. B. (2012). Upsides and downsides of reactive oxygen species for cancer: the roles of reactive oxygen species in tumorigenesis, prevention, and therapy. *Antioxidants & Redox Signaling*, 16(11), 1295-1322.
- Halliwell, B. (2006). Oxidative stress and neurodegeneration: where are we now? *Journal of Neurochemistry*, 97(6), 1634-1658.
- Halliwell, B. (2007). Biochemistry of oxidative stress. *Biochemical Society Transactions*, 35, 1147-1150.
- Halliwell, B., & Gutteridge, J. M. (1999). *Free radicals in biology and medicine* (3rd ed.). New York, NY: Oxford University Press.

- Halliwell, B., & Gutteridge, J. M. (2007). *Free radicals in biology and medicine* (4th ed.). Oxford, England: Oxford University Press.
- Hamade, K., Karam, L., Habib, J., Merhi, R. A., & Elkak, A. (2017). Cytotoxic effect of *Berberis libanotica* roots extracts on human cancer cells and antioxidant activities. *The Journal of Phytopharmacology*, 6(6), 311-317.
- Hanasaki, Y., Ogawa, S., & Fukui, S. (1994). The correlation between active oxygens scavenging and antioxidative effects of flavonoids. *Free Radical Biology & Medicine*, 16(6), 845-850.
- Harborne, J. B., & Williams, C. A. (2000). Advances in flavonoid research since 1992. *Phytochemistry*, 55(6), 481-504.
- Hata, R. I., & Senoo, H. (1989). L-ascorbic acid 2-phosphate stimulates collagen accumulation, cell proliferation, and formation of a three-dimensional tissuelike substance by skin fibroblasts. *Journal of Cellular Physiology*, 138(1), 8-16.
- Hata, R. I., Sunada, H., Arai, K., Sato, T., Ninomiya, Y., Nagai, Y., & Senoo, H. (1988). Regulation of collagen metabolism and cell growth by epidermal growth factor and ascorbate in cultured human skin fibroblasts. *European Journal of Biochemistry*, 173(2), 261-267.
- Hawksworth, F. G. (1983). Mistletoes as forest parasites. In M. Calder & P. Bernhardt (Eds.), *The biology of mistletoes*, (pp. 317-333). Sydney, Australia: Academic Press.
- Heim, K. E., Tagliaferro, A. R., & Bobilya, D. J. (2002). Flavonoid antioxidants: chemistry, metabolism and structure-activity relationships. *The Journal of Nutritional Biochemistry*, 13(10), 572-584.
- Heinrich, M., Barnes, Gibbons, S., & Williamson, E. M. (1993). *Fundamentals of Pharmacognosy and Phytotherapy* (2nd ed.). Edinburgh, UK: Elsevier Health Sciences.
- Hertog, M. G., Hollman, P. C., & Van de Putte, B. (1993). Content of potentially anticarcinogenic flavonoids of tea infusions, wines, and fruit juices. *Journal of Agricultural & Food Chemistry*, 41(8), 1242-1246.
- Hirshfield, K. M., Rebbeck, T. R., & Levine, A. J. (2010). Germline mutations and polymorphisms in the origins of cancers in women. *Journal of Oncology*, 2010, 1-11.
- Horax, R., Hettiarachchy, N., & Chen, P. (2010). Extraction, quantification, and antioxidant activities of phenolics from pericarp and seeds of bitter melons (*Momordica charantia*) harvested at three maturity stages (immature, mature, and ripe). *Journal of Agricultural & Food Chemistry*, 58(7), 4428-4433.
- Hostanska, K., Hajto, T., Spagnoli, G. C., Fischer, J., Lentzen, H., & Herrmann, R. (1995). A plant lectin derived from *Viscum album* induces cytokine gene

- expression and protein production in cultures of human peripheral blood mononuclear cells. *Natural Immunity*, 14(5-6), 295-304.
- Housman, G., Byler, S., Heerboth, S., Lapinska, K., Longacre, M., Snyder, N. & Sarkar, S. (2014). Drug resistance in cancer: an overview. *Cancers*, 6(3), 1769-1792.
- Howes, R. M. (2009). Dangers of Antioxidants in Cancer Patients: A Review. Retrieved from http://philica.com/display_article.php?article_id=153.
- Hsu, J. D., Kao, S. H., Ou, T. T., Chen, Y. J., Li, Y. J., & Wang, C. J. (2011). Gallic acid induces G2/M phase arrest of breast cancer cell MCF-7 through stabilization of p27Kip1 attributed to disruption of p27Kip1/Skp2 complex. *Journal of Agricultural & Food Chemistry*, 59(5), 1996-2003.
- Huang, D., Ou, B., & Prior, R. L. (2005). The chemistry behind antioxidant capacity assays. *Journal of Agricultural & Food Chemistry*, 53(6), 1841-1856.
- Huaxing, Q., Hua-hsing, C., Hua-xing, K., & Gilbert, M. G. (2003). Loranthaceae. *Flora of China*, 5, 220-239.
- Hussain, J., Ali, L., Khan, A. L., Rehman, N. U., Jabeen, F., Kim, J. S., & Al-Harrasi, A. (2014). Isolation and bioactivities of the flavonoids morin and morin-3-O- β -d-glucopyranoside from *Acridocarpus orientalis*—A wild Arabian medicinal plant. *Molecules*, 19(11), 17763-17772.
- Iacopini, P., Baldi, M., Storch, P., & Sebastiani, L. (2008). Catechin, epicatechin, quercetin, rutin and resveratrol in red grape: Content, in vitro antioxidant activity and interactions. *Journal of Food Composition & Analysis*, 21(8), 589-598.
- Ichihara, M. K., Murakami, S., Nakanishi K., Takahashi, Y., Yamaguchi, M., Shioya, T., ... & Hino, A. (2013). Antioxidant Capacity and Polyphenol Content of Extracts from Crops Cultivated in Japan, and the Effect of Cultivation Environment. *Food Science & Technology Research*, 19(1), 69-79.
- Iloki-Assanga, S. B., Lewis-Luján, L. M., Lara-Espinoza, C. L., Gil-Salido, A. A., Fernandez-Angulo, D., Rubio-Pino, J. L., & Haines, D. D. (2015). Solvent effects on phytochemical constituent profiles and antioxidant activities, using four different extraction formulations for analysis of *Bucida buceras* L. and *Phoradendron californicum*. *BMC Research Notes*, 8(1), 396.
- Iqbal, J., Abbasi, B. A., Batool, R., Mahmood, T., Ali, B., Khalil, A. T., ... & Ahmad, R. (2018). Potential phytochemicals for developing breast cancer therapeutics: Nature's healing touch. *European Journal of Pharmacology*, 15, 125-148.
- Iqbal, J., Abbasi, B. A., Mahmood, T., Kanwal, S., Ali, B., & Khalil, A. T. (2017). Plant-derived anticancer agents: A green anticancer approach. *Asian Pacific Journal of Tropical Biomedicine*. 7(12), 1129-1150.

- Ismail, H. I., Chan, K. W., Mariod, A. A., & Ismail, M. (2010). Phenolic content and antioxidant activity of cantaloupe (*Cucumis melo*) methanolic extracts. *Food Chemistry*, 119(2), 643-647.
- Ismail, N.N., Utra, U., Hoong, C. L., & Esa, A. M. (2018). Effect of Tea Polyphenols on α -Amylase Activity in Starch Hydrolysis. *Sains Malaysiana*, 47(4), 731-739.
- Iwakuma, T. & Lozano, G. (2003). MDM2, An Introduction. *Molecular Cancer Research*, 1, 993-1000.
- Iwalokun, B. A., Hodonu, S. A., Nwoke, S., Ojo, O., & Agomo, P. U. (2011). Evaluation of the possible mechanisms of antihypertensive activity of *Loranthus micranthus*: an African mistletoe. *Biochemistry Research International*, 2011, 1-9
- Jahan, M. S., Zawawi, D. D., & Abdulkadir, A. R. (2015). Effect of chlorophyll content and maturity on total phenolic, total flavonoid contents and antioxidant activity of *Moringa oleifera* leaf (Miracle tree). *Journal of Chemical & Pharmaceutical Research*, 7(5), 1147-1152.
- Jahan, M. S., Zawawi, D. D., & Abdulkadir, A. R. (2015a). DPPH antioxidant activity, total phenolic and total flavonoid content of different part of Drumstic tree (*Moringa oleifera* Lam.). *Journal of Chemical & Pharmaceutical Research*, 7(4), 1423-1428.
- Jahan, M. S., Zawawi, D. D., & Abdulkadir, A. R. (2015b). Effect of chlorophyll content and maturity on total phenolic, total flavonoid contents and antioxidant activity of *Moringa oleifera* leaf (Miracle tree). *Journal of Chemical & Pharmaceutical Research*, 7(5), 1147-1152.
- Jan, S., Khan, M. R., Rashid, U., & Bokhari, J. (2013). Assessment of antioxidant potential, total phenolics and flavonoids of different solvent fractions of *Monothea buxifolia* fruit. *Osong Public Health & Research Perspectives*, 4(5), 246-254.
- Janssen, O., Scheffler, A., & Kabelitz, D. (1993). In vitro effects of mistletoe extracts and mistletoe lectins. Cytotoxicity towards tumor cells due to the induction of programmed cell death (apoptosis). *Arzneimittel-Forschung*, 43(11), 1221-1227.
- Jaradat, N. A., Shawahna, R., Hussein, F., & Al-Lahham, S. (2016). Analysis of the antioxidant potential in aerial parts of *Trigonella arabica* and *Trigonella berythea* grown widely in Palestine: A comparative study. *European Journal of Integrative Medicine*, 8(5), 623-630.
- Jeffery, E. H., Brown, A. F., Kurilich, A. C., Keck, A. S., Matusheski, N., Klein, B. P., & Juvik, J. A. (2003). Variation in content of bioactive components in broccoli. *Journal of Food Composition & Analysis*, 16(3), 323-330.
- Johansson, S., Gullbo, J., Lindholm, P., Ek, B., Thunberg, E., Samuelsson, G., ... & Claeson, P. (2003). Small, novel proteins from the mistletoe *Phoradendron*

- tomentosum* exhibit highly selective cytotoxicity to human breast cancer cells. *Cellular & Molecular Life Sciences CMLS*, 60(1), 165-175.
- Johri, B. M., & Bhatnagar, S. P. (1972). *Botanical Monograph No. 8.: Loranthaceae*. New Delhi, India: Council of Scientific and Industrial Research.
- Justine, V. T. (2016). *Evaluation of Different Post-Harvest Treatments on Malaysian Mistletoe (Scurrula ferruginea (Jack) Dans.) Leaves Extracts Yield, Antioxidant and Antimicrobial Performance* (Master's thesis). Universiti Putra Malaysia, Serdang, Malaysia.
- Justine, V.T., Mustafa, M., & Go, R. (2018). Effect of Antimicrobial Activities on the various solvents extracts of leaves of *Scurrula ferruginea* (Jack) Danser (Loranthaceae). *Pertanika Journal of Tropical Agricultural Science*, 41(2), 677-686.
- Kafaru, E. (1993, June 3). 'Mistletoe-an example of an all-purpose herb' Herbal Remedies. *Guardian Newspaper*, pp. 11.
- Kalia, K., Sharma, K., Singh, H. P., & Singh, B. (2008). Effects of extraction methods on phenolic contents and antioxidant activity in aerial parts of *Potentilla atosanguinea* Lodd. and quantification of its phenolic constituents by RP-HPLC. *Journal of Agricultural & Food Chemistry*, 56(21), 10129-10134.
- Kampa, M., Hatzoglou, A., Notas, G., Damianaki, A., Bakogeorgou, E., Gemetzi, C., ... & Castanas, E. (2000). Wine antioxidant polyphenols inhibit the proliferation of human prostate cancer cell lines. *Nutrition & Cancer*, 37(2), 223-233.
- Kaneria, M. J., Bapodara, M. B., & Chanda, S. V. (2012). Effect of extraction techniques and solvents on antioxidant activity of pomegranate (*Punica granatum* L.) leaf and stem. *Food Analytical Methods*, 5(3), 396-404.
- Karimi, E., Jaafar, H. Z., & Ahmad, S. (2011). Phytochemical analysis and antimicrobial activities of methanolic extracts of leaf, stem and root from different varieties of *Labisa pumila* Benth. *Molecules*, 16(6), 4438-4450.
- Kasai, H. (1997). Analysis of a form of oxidative DNA damage, 8-hydroxy-2'-deoxyguanosine, as a marker of cellular oxidative stress during carcinogenesis. *Mutation Research/Reviews in Mutation Research*, 387(3), 147-163.
- Katsarou, A., Rhizopoulou, S., & Kefalas, P. (2012). Antioxidant potential of the aerial tissues of the mistletoe *Loranthus europaeus* Jacq. *Records of Natural Products*, 6(4), 394-397.
- Kaur, S., & Mondal, P. (2014). Study of total phenolic and flavonoid content, antioxidant activity and antimicrobial properties of medicinal plants. *Journal of Microbiology & Experimentation*, 1(1), 00005-00010.
- Kchaou, W., Abbès, F., Blecker, C., Attia, H., & Besbes, S. (2013). Effects of extraction solvents on phenolic contents and antioxidant activities of Tunisian

- date varieties (*Phoenix dactylifera* L.). *Industrial Crops & Products*, 45, 262-269.
- Kew Science (2017). *Helixanthera parasitica* Lour. Retrieved from <http://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:548786-1>.
- Khaled, A. S., Nahla, S. A. A., Ibrahim, A. S., Mohamed-Elamir, F. H., Mostafa, M. E. M., & Faiza, M. H. (2015). Green technology: economically and environmentally innovative methods for extraction of medicinal & aromatic plants (MAP) in Egypt. *Journal of Chemical & Pharmaceutical Research*, 7(5), 1050-1074.
- Khan, M., Ganie, S. A., Wani, I. H., Ganai, B. A., Masood, A., Zargar, M. A., ... & Hamid, R. (2012). Free radical scavenging activity of *Elsholtzia densa*. *Journal of Acupuncture & Meridian Studies*, 5(3), 104-111.
- Khan, M., Maryam, A., Zhang, H., Mehmood, T., & Ma, T. (2016). Killing cancer with platycodin D through multiple mechanisms. *Journal of Cellular & Molecular Medicine*, 20(3), 389-402.
- Khandanlou, R., Murthy, V., Saranath, D., & Damani, H. (2018). Synthesis and characterization of gold-conjugated *Backhousia citriodora* nanoparticles and their anticancer activity against MCF-7 breast and HepG2 liver cancer cell lines. *Journal of Materials Science*, 53(5), 3106-3118.
- Khoddami, A., Wilkes, M. A., & Roberts, T. H. (2013). Techniques for analysis of plant phenolic compounds. *Molecules*, 18(2), 2328-2375.
- Khokhar, S., & Magnusdottir, S. G. M. (2002). Total phenol, catechin, and caffeine contents of teas commonly consumed in the United Kingdom. *Journal of Agricultural & Food Chemistry*, 50(3), 565-570.
- Kienle, G. S., & Kiene, H. (2010). Influence of *Viscum album* L (European mistletoe) extracts on quality of life in cancer patients: a systematic review of controlled clinical studies. *Integrative Cancer Therapies*, 9(2), 142-157.
- Kim, Y. K., Kim, Y. S., Choi, S. U., & Ryu, S. Y. (2004). Isolation of flavonol rhamnosides from *Loranthus tanakae* and cytotoxic effect of them on human tumor cell lines. *Archives of Pharmacal Research*, 27(1), 44-47.
- Klayman, D. L., Lin, A. J., Acton, N., Scovill, J. P., Hoch, J. M., Milhous, W. K., ... & Dobek, A. S. (1984). Isolation of artemisinin (qinghaosu) from *Artemisia annua* growing in the United States. *Journal of Natural Products*, 47(4), 715-717.
- Kneifel, W., Czech, E., & Kopp, B. (2002). Microbial contamination of medicinal plants-a review. *Planta Medica*, 68(01), 5-15.
- Köksal, E., Bursal, E., Dikici, E., Tozoglu, F., & Gulcin, I. (2011). Antioxidant activity of *Melissa officinalis* leaves. *Journal of Medicinal Plants Research*, 5(2), 217-222.

- Kolář, P., Shen, J. W., Tsuboi, A., & Ishikawa, T. (2002). Solvent selection for pharmaceuticals. *Fluid Phase Equilibria*, 194, 771-782.
- Kovalchuk, O., Filkowski, J., Meservy, J., Ilnytsky, Y., Tryndyak, V. P., Vasylyshyn, C., & Pogribny, I. P. (2008). Involvement of microRNA-451 in resistance of the MCF-7 breast cancer cells to chemotherapeutic drug doxorubicin. *Molecular Cancer Therapeutics*, 7(7), 2152-2159.
- Krishnamoorthi, R., & Elumalai, K. (2018). In-vitro Anticancer Activity of Ethyl Acetate Extract of *Aerva lanata* against MCF-7 Cell Line. *International Journal of Pharma Research & Health Sciences*, 6(1), 2286-89.
- Ksouri, R., Falleh, H., Megdiche, W., Trabelsi, N., Mhamdi, B., Chaieb, K., ... & Abdelly, C. (2009). Antioxidant and antimicrobial activities of the edible medicinal halophyte *Tamarix gallica* L. and related polyphenolic constituents. *Food & Chemical Toxicology*, 47(8), 2083-2091.
- Kuijt, J. (1969). The biology of parasitic flowering plants. University of California Press, Berkeley. *The biology of parasitic flowering plants. University of California Press, Berkeley*.
- Kulkarni, A. C., Kuppusamy, P., & Parinandi, N. (2007). Oxygen, the lead actor in the pathophysiologic drama: enactment of the trinity of normoxia, hypoxia, and hyperoxia in disease and therapy. *Antioxidants & Redox Signaling*, 9(10), 1717-1730.
- Kumar, S., Saini, M., Kumar, V., Prakash, O., Arya, R., Rana, M., & Kumar, D. (2012). Traditional medicinal plants curing diabetes: A promise for today and tomorrow. *Asian Journal of Traditional Medicines*, 7, 178-88.
- Kumoro, A. C., Hasan, M., & Singh, H. (2009). Effects of solvent properties on the Soxhlet extraction of diterpenoid lactones from *Andrographis paniculata* leaves. *Science Asia*, 35, 306-309.
- Kunwar, R. M., Adhikari, N., & Devkota, M. P. (2005). Indigenous use of mistletoes in tropical and temperate region of Nepal. *Banko Janakari*, 15(2), 38-42.
- Kwanda, N., Noikotr, K., Sudmoon, R., Tanee, T., & Chaveerach, A. (2013). Medicinal parasitic plants on diverse hosts with their usages and barcodes. *Journal of Natural Medicines*, 67(3), 438-445.
- Lakhanpal, P., & Rai, D. K. (2007). Quercetin: a versatile flavonoid. *Internet Journal of Medical Update*, 2(2), 22-37.
- Lapornik, B., Prošek, M., & Wondra, A. G. (2005). Comparison of extracts prepared from plant by-products using different solvents and extraction time. *Journal of Food Engineering*, 71(2), 214-222.
- Lee, S. E., Hwang, H. J., Ha, J. S., Jeong, H. S., & Kim, J. H. (2003). Screening of medicinal plant extracts for antioxidant activity. *Life Sciences*, 73(2), 167-179.

- Lemmens, R.H.M.J., & Bunyapraphatsara, N. (2003). *Plant Resources of South-East Asia: Medicinal and Poisonous Plants 3*. Leiden, The Netherlands: Backhuys.
- Li, Y., & Schellhorn, H. E. (2007). New developments and novel therapeutic perspectives for vitamin C. *The Journal of Nutrition*, 137(10), 2171-2184.
- Liangqiong, L., Meirong, L., & Wentao, F. (1994). Studies on the Chemical Constituents of Libanjisheng (*Helixanthera parasitica*) [J]. *Chinese Traditional & Herbal Drugs*, 6.
- Lim, Y. C., Rajabalaya, R., Lee, S. H. F., Tennakoon, K. U., Le, Q. V., Idris, A., ... & David, S. R. (2016). Parasitic mistletoes of the genera *Scurrula* and *Viscum*: from bench to bedside. *Molecules*, 21(8), 1048.
- Lippert, T. H., Ruoff, H. J., & Volm, M. (2011). Current status of methods to assess cancer drug resistance. *International Journal of Medical Sciences*, 8(3), 245.
- Lirdprapamongkol, K., Mahidol, C., Thongnest, S., Prawat, H., Ruchirawat, S., Srisomsap, C., ... & Svasti, J. (2003). Anti-metastatic effects of aqueous extract of *Helixanthera parasitica*. *Journal of Ethnopharmacology*, 86(2-3), 253-256.
- Lisiewska, Z., Kmiecik, W., & Korus, A. (2006). Content of vitamin C, carotenoids, chlorophylls and polyphenols in green parts of dill (*Anethum graveolens* L.) depending on plant height. *Journal of Food Composition & Analysis*, 19(2-3), 134-140.
- Liu, Z. Q., Ma, L. P., Zhou, B., Yang, L., & Liu, Z. L. (2000). Antioxidative effects of green tea polyphenols on free radical initiated and photosensitized peroxidation of human low density lipoprotein. *Chemistry & Physics of Lipids*, 106(1), 53-63.
- Liyana-Pathirana, C. M., & Shahidi, F. (2006). Importance of insoluble-bound phenolics to antioxidant properties of wheat. *Journal of Agricultural & Food Chemistry*, 54(4), 1256-1264.
- Lohézic-Le Dévéhat, F., Bakhtiar, A., Bezivin, C., Amoros, M., & Boustie, J. (2002). Antiviral and cytotoxic activities of some Indonesian plants. *Fitoterapia*, 73(5), 400-405.
- Lü, J. M., Lin, P. H., Yao, Q., & Chen, C. (2010). Chemical and molecular mechanisms of antioxidants: experimental approaches and model systems. *Journal of Cellular & Molecular Medicine*, 14(4), 840-860.
- Lu, Y., & Foo, L. Y. (1999). Rosmarinic acid derivatives from *Salvia officinalis*. *Phytochemistry*, 51(1), 91-94.
- Lu, Y., Jiang, F., Jiang, H., Wu, K., Zheng, X., Cai, Y., ... & To, S. S. T. (2010). Gallic acid suppresses cell viability, proliferation, invasion and angiogenesis in human glioma cells. *European Journal of Pharmacology*, 641(2-3), 102-107.

- Lupea, A. X., Pop, M., & Cacig, S. (2008). Structure-radical scavenging activity relationships of flavonoids from *Ziziphus* and *Hydrangea* extracts. *Revista de Chimie*, 59(3), 309-313.
- Luximon-Ramma, A., Bahorun, T., Crozier, A., Zbarsky, V., Datla, K. P., Dexter, D. T., & Aruoma, O. I. (2005). Characterization of the antioxidant functions of flavonoids and proanthocyanidins in Mauritian black teas. *Food Research International*, 38(4), 357-367.
- Ma, H., Das, T., Pereira, S., Yang, Z., Zhao, M., Mukerji, P., & Hoffman, R. M. (2009). Efficacy of dietary antioxidants combined with a chemotherapeutic agent on human colon cancer progression in a fluorescent orthotopic mouse model. *Anticancer Research*, 29(7), 2421-2426.
- MacConaill, L. E., & Garraway, L. A. (2010). Clinical implications of the cancer genome. *Journal of Clinical Oncology*, 28(35), 5219.
- Macheix, J., Fleuriet, A., & Billot, J. (1990). Fruit phenolics. Boca Raton: CRC Press.
- Madeja, J., Harmata, K., Kołaczek, P., Karpinska-Kolaczek, M., Piatek, K., & Naks, P. (2009). Bracken (*Pteridium aquilinum* (L.) Kuhn), mistletoe (*Viscum album* (L.)) and bladder-nut (*Staphylea pinnata* (L.))-mysterious plants with unusual applications. Cultural and ethnobotanical studies. In J-P. Morel & A. M. Mercuri (Eds.), *Plants and Culture: seeds of the cultural heritage of Europe*. (pp. 207-215), Bari, Italy: Edipuglia.
- Mahon, F. X., Hayette, S., Lagarde, V., Belloc, F., Turcq, B., Nicolini, F., ... & Pasquet, J.M. (2008). Evidence that resistance to nilotinib may be due to BCR-ABL, Pgp, or Src kinase overexpression. *Cancer Research*, 68(23), 9809-9816.
- Makwana, B. A., Darjee, S., Jain, V. K., Kongor, A., Sindhav, G., & Rao, M. V. (2017). A comparative study: Metal nanoparticles as fluorescent sensors for biomolecules and their biomedical application. *Sensors & Actuators B: Chemical*, 246, 686-695.
- Mandryk, D. (2005). *Canadian Christmas Tradition: Festives recipes and stories from coast to coast*. Canmore, Alberta, Canada: Altitude Publishing Canada Ltd.
- Manolitsas, T. P., Englefield, P., Eccles, D. M., & Campbell, I. G. (1997). No association of a 306-bp insertion polymorphism in the progesterone receptor gene with ovarian and breast cancer. *British Journal of Cancer*, 75(9), 1398.
- Mansoori, B., Mohammadi, A., Davudian, S., Shirjang, S., & Baradaran, B. (2017). The Different Mechanisms of Cancer Drug Resistance: A Brief Review. *Advanced Pharmaceutical Bulletin*, 7(3), 339.
- Marvibaigi, M., Amini, N., Supriyanto, E., Jamil, S., Abdul Majid, F. A., & Khangholi, S. (2014). Total phenolic content, antioxidant and antibacterial properties of *Scurrula ferruginea* extracts. *Jurnal Teknologi*, 5, 65-72.

- Matkowski, A. (2008). Plant in vitro culture for the production of antioxidants—a review. *Biotechnology Advances*, 26(6), 548-560.
- Meads, M. B., Gatenby, R. A., & Dalton, W. S. (2009). Environment-mediated drug resistance: a major contributor to minimal residual disease. *Nature Reviews Cancer*, 9(9), 665.
- Medini, F., Fellah, H., Ksouri, R., & Abdelly, C. (2014). Total phenolic, flavonoid and tannin contents and antioxidant and antimicrobial activities of organic extracts of shoots of the plant *Limonium delicatulum*. *Journal of Taibah University for Science*, 8(3), 216-224.
- Merculieff, Z., Ramnath, S., Sankoli, S. M., Venkataramgowda, S., Murthy, G. S., & Ceballos, R. M. (2014). Phytochemical, antioxidant and antibacterial potential of *Elaeagnus kologa* (Schlecht.) leaf. *Asian Pacific Journal of Tropical Medicine*, 7, S599-S602.
- Michor, F., Nowak, M. A., & Iwasa, Y. (2006). Evolution of resistance to cancer therapy. *Current Pharmaceutical Design*, 12(3), 261-271.
- Miller, E. R., Pastor-Barriuso, R., Dalal, D., Riemersma, R. A., Appel, L. J., & Guallar, E. (2005). Meta-analysis: high-dosage vitamin E supplementation may increase all-cause mortality. *Annals of Internal Medicine*, 142(1), 37-46.
- Mohammedelnour, A. A., Mirghani, M.E.S., Kabbashi, N. A., Alam, M. Z., Musa, K. H., & Abdullah, A. (2017). Effect of solvent types on phenolics content and antioxidant activities of *Acacia polyacantha* gum. *International Food Research Journal*, 24, S369-S377.
- Molyneux, P. (2004). The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. *Songklanakarin Journal of Science & Technology*, 26(2), 211-219.
- Mothana, R. A., Al-Said, M. S., Al-Rehaily, A. J., Thabet, T. M., Awad, N. A., Lalk, M., & Lindequist, U. (2012). Anti-inflammatory, antinociceptive, antipyretic and antioxidant activities and phenolic constituents from *Loranthus regularis* Steud. ex Sprague. *Food Chemistry*, 130(2), 344-349.
- Moustapha, B., Marina, G. A. D., Raúl, F. O., Raquel, C. M., & Mahinda, M. (2011). Chemical constituents of the Mexican mistletoe (*Psittacanthus calyculatus*). *Molecules*, 16(11), 9397-9403.
- Mukherjee, P. K., Satheesh Kumar, N., & Heinrich, M. (2008). Plant made pharmaceuticals (PMPs)-Development of natural health products from biodiversity. *Indian Journal of Pharmaceutical Education & Research*, 42(2), 113-121.
- Mukherji, S.M. & Singh, S.P. (1986). *Reaction mechanism in organic chemistry*. Madras, India: Macmillan India Press.
- Munhoz, V. M., Longhini, R., Souza, J. R., Zequi, J. A., Mello, E. V., Lopes, G. C., & Mello, J. C. (2014). Extraction of flavonoids from *Tagetes patula*: process

- optimization and screening for biological activity. *Revista Brasileira de Farmacognosia*, 24(5), 576-583.
- Muniyandi, K., George, E., Mudili, V., Kalagatur, N. K., Anthuvan, A. J., Krishna, K., ... & Natarajan, G. (2017). Antioxidant and anticancer activities of *Plectranthus stocksii* Hook. f. leaf and stem extracts. *Agriculture & Natural Resources*, 51(2), 63-73.
- Murray, C. J., Vos, T., Lozano, R., Naghavi, M., Flaxman, A. D., Michaud, C., ... & Aboyans, V. (2012). Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. *The Lancet*, 380(9859), 2197-2223.
- Musa, A. F., Dillon, J., Taib, M. E. M., Yunus, A. M., Nordin, R. B., & Hay, Y. K. (2018). Hypotheses, rationale, design, and methods for evaluation of a randomized controlled trial using Tocotrienol, an isomer of Vitamin E derived from palm oil, on the prevention of atrial fibrillation after coronary artery bypass grafting surgery. *F1000Research*, 7, 215-237.
- Musa, K. H., Abdullah, A., Jusoh, K., & Subramaniam, V. (2011). Antioxidant activity of pink-flesh guava (*Psidium guajava* L.): effect of extraction techniques and solvents. *Food Analytical Methods*, 4(1), 100-107.
- Naczki, M., & Shahidi, F. (2006). Phenolics in cereals, fruits and vegetables: Occurrence, extraction and analysis. *Journal of Pharmaceutical & Biomedical Analysis*, 41(5), 1523-1542.
- Nakatani, N. (2003). Biologically functional constituents of spices and herbs (2002's JSNFS award for excellence in research). *Journal of Japanese Society of Nutrition & Food Science*, 56(6), 389-395.
- National Department of Statistics Malaysia. (2017, October 31). *Statistics on Causes of Death, Malaysia, 2017*. Retrieved from https://www.dosm.gov.my/v1/index.php?r=column/cthemByCat&cat=401&bu_id=Y3psYUI2VjU0ZzRhZU1kcVFMMThGUT09&menu_id=L0pheU43NWJwRWVSZklWdzQ4TlhUUT09.
- Newman, D. J., & Cragg, G. M. (2012). Natural products as sources of new drugs over the 30 years from 1981 to 2010. *Journal of Natural Products*, 75(3), 311-335.
- Niseteo, T., Komes, D., Belščak-Cvitanović, A., Horžić, D., & Budeč, M. (2012). Bioactive composition and antioxidant potential of different commonly consumed coffee brews affected by their preparation technique and milk addition. *Food Chemistry*, 134(4), 1870-1877.
- Nordin, M. L., Kadir, A. A., Zakaria, Z. A., Abdullah, R., & Abdullah, M. N. H. (2018). In vitro investigation of cytotoxic and antioxidative activities of *Ardisia crispa* against breast cancer cell lines, MCF-7 and MDA-MB-231. *BMC Complementary & Alternative Medicine*, 18(1), 87-97.

- Nurhanan, M. Y., Asiah, O., Ilham, M. M., Syarifah, M. S., Norhayati, I., & Sahira, H. L. (2008). Anti-proliferative activities of 32 Malaysian plant species in breast cancer cell lines. *Journal of Tropical Forest Science*, 20, 77-81.
- O'Neill, A. R., & Rana, S. K. (2016). An ethnobotanical analysis of parasitic plants (Parijibi) in the Nepal Himalaya. *Journal of Ethnobiology & Ethnomedicine*, 12(1), 14-29.
- Obeidat, M., Shatnawi, M., Al-alawi, M., Al-Zu'bi, E., Al-Dmoor, H., Al-Qudah, M., ... & Otri, I. (2012). Antimicrobial activity of crude extracts of some plant leaves. *Research Journal of Microbiology*, 7(1), 59-67.
- Ogunmefun, O. T., Fasola, T. R., Saba, A. B., & Oridupa, O. A. (2013). The ethnobotanical, phytochemical and mineral analyses of *Phragmanthera incana* (Klotzsch), a species of mistletoe growing on three plant hosts in South-Western Nigeria. *International Journal of Biomedical Science: IJBS*, 9(1), 33-40.
- Ohashi, K., Winarno, H., Mukai, M., & Shibuya, H. (2003). Preparation and cancer cell invasion inhibitory effects of C16-alkynic fatty acids. *Chemical & Pharmaceutical Bulletin*, 51(4), 463-466.
- Okpanachi, G. O., Oyi, A. R., Musa, H., Abdulsamad, A., Sani, M. B. & Ya'u, J. (2018). Phytochemical Screening and Hypoglycemic Property of *Globimetula braunii* (Loranthaceae) Leaf Extracts. *Journal of Pharmaceutical Research International*, 22(1), 1-11.
- Omeje, E. O., Osadebe, P. O., Esimone, C. O., Nworu, C. S., Kawamura, A., & Proksch, P. (2012). Three hydroxylated lupeol-based triterpenoid esters isolated from the Eastern Nigeria mistletoe parasitic on *Kola acuminata*. *Natural Product Research*, 26(19), 1775-1781.
- Onunogbo, C., Ohaeri, O. C., Eleazu, C. O., & Eleazu, K. C. (2012). Chemical composition of mistletoe extract (*Loranthus micranthus*) and its effect on the protein, lipid metabolism and the antioxidant status of alloxan induced diabetic rats. *The Journal of Medical Research*, 1(4), 057-063.
- Osadebe, P. O., Abba, C. C., & Agbo, M. O. (2012). Antimotility effects of extracts and fractions of Eastern Nigeria mistletoe (*Loranthus micranthus* Linn). *Asian Pacific Journal of Tropical Medicine*, 5(7), 556-560.
- Oxyradicals, L. M. (2000). DNA damage. *Carcinogenesis*, 21(3), 361-370.
- Pacher, P., Beckman, J. S., & Liaudet, L. (2007). Nitric oxide and peroxynitrite in health and disease. *Physiological Reviews*, 87(1), 315-424.
- Pagliosa, C. M., Vieira, M. A., Podestá, R., Maraschin, M., Zeni, A. L. B., Amante, E. R., & Amboni, R. D. D. M. C. (2010). Methylxanthines, phenolic composition, and antioxidant activity of bark from residues from mate tree harvesting (*Ilex paraguariensis* A. St. Hil.). *Food Chemistry*, 122(1), 173-178.

- Parimala, S., & Selvan, A. T. (2017). Anticancer and Antioxidant activity of *Cissus pallida* and *Cissus vitegenia*. *Journal of Pharmacognosy & Phytochemistry*, 6(4), 1521-1526.
- Park, J. H., Hyun, C. K., & Shin, H. K. (1998). Cytotoxicity of heat-treated Korean mistletoe. *Cancer Letters*, 126(1), 43-48.
- Park, J. H., Hyun, C. K., & Shin, H. K. (1999). Cytotoxic effects of the components in heat-treated mistletoe (*Viscum album*). *Cancer Letters*, 139(2), 207-213.
- Parthasarathy, S., Santanam, N., Ramachandran, S., & Meilhac, O. (1999). Oxidants and antioxidants in atherogenesis: an appraisal. *Journal of Lipid Research*, 40(12), 2143-2157.
- Parwati, N. W. M., Lindayani, I. K., Ratnawati, R., Winarsih, S., & Nurseta, T. (2015). Possible effect of tea plant parasite, *Scurrula atropurpurea* (Blume) Danser, on growth inhibition of culture HeLa cells in vitro through DNA repair and apoptosis intrinsic pathways mechanism. *Asian Pacific Journal of Tropical Disease*, 5(9), 743-746.
- Patwardhan, B. (2005). Ethnopharmacology and drug discovery. *Journal of Ethnopharmacology*, 100(1-2), 50-52.
- Peng Bin (Executive Ed.). (2013). *Case studies on good practices of biodiversity conservation in ASEAN and China*. Beijing, China: China-ASEAN Cooperation Centre & ASEAN Centre for Biodiversity.
- Pham-Huy, L. A., He, H., & Pham-Huy, C. (2008). Free radicals, antioxidants in disease and health. *International Journal of Biomedical Science: IJBS*, 4(2), 89.
- Phillips, C. L., Combs, S. B., & Pinnell, S. R. (1994). Effects of ascorbic acid on proliferation and collagen synthesis in relation to the donor age of human dermal fibroblasts. *Journal of Investigative Dermatology*, 103(2), 228-232.
- Phongpaichit, S., Nikom, J., Rungjindamai, N., Sakayaroj, J., Hutadilok-Towatana, N., Rukachaisirikul, V., & Kirtikara, K. (2007). Biological activities of extracts from endophytic fungi isolated from *Garcinia* plants. *FEMS Immunology & Medical Microbiology*, 51(3), 517-525.
- Phongtongpasuk, S., & Poadang, S. (2014). Extraction of antioxidants from *Peperomia pellucida* L. Kunth. *Science & Technology Asia*, 19(3), 38-43.
- Pim, L.H. (2017, March 19). One in 19 Malaysian women at risk of breast cancer. *New Straits Times*. Retrieved from <http://www.theborneopost.com/2017/03/19/one-in-19-malaysian-women-at-risk-of-breast-cancer/>.
- Pisoschi, A. M., & Pop, A. (2015). The role of antioxidants in the chemistry of oxidative stress: A review. *European Journal of Medicinal Chemistry*, 97, 55-74.

- Poljsak, B., Šuput, D., & Milisav, I. (2013). Achieving the balance between ROS and antioxidants: when to use the synthetic antioxidants. *Oxidative medicine and cellular longevity*, 2013, 1-11.
- Pulido, R., Bravo, L., & Saura-Calixto, F. (2000). Antioxidant activity of dietary polyphenols as determined by a modified ferric reducing/antioxidant power assay. *Journal of Agricultural & Food Chemistry*, 48(8), 3396-3402.
- Putiyanan, S., & Maxwell, J. F. (2007). Survey and Herbarium Specimens of Medicinal Vascular Flora of Doi Chiang Dao. *Chiang Mai University Journal of Natural Sciences (Thailand)*, 6(1), 159-169.
- Qin, P. Z., & Warncke, K. (2015). *Electron paramagnetic resonance investigations of biological systems by using spin labels, spin probes, and intrinsic metal ions* (Vol. 564). MA, USA: Academic Press.
- Rafat, A., Philip, K., & Muni, S. (2010). Antioxidant potential and content of phenolic compounds in ethanolic extracts of selected parts of *Andrographis paniculata*. *Journal of Medicinal Plants Research*, 4(3), 197-202.
- Rosely, N. F. N. (2014). Mistletoe abundance, distribution and their associations with trees along roadside in Penang, Malaysia. *Tropical Ecology*, 55(2), 255-262.
- Randhir, R., Lin, Y. T., & Shetty, K. (2004). Phenolics, their antioxidant and antimicrobial activity in dark germinated fenugreek sprouts in response to peptide and phytochemical elicitors. *Asia Pacific Journal of Clinical Nutrition*, 13(3), 295-307.
- Rao, C. V. (1963). On the morphology of the calyculus. *Journal of the Indian Botanical Society*, 42, 618-28.
- Rates, S. M. K. (2001). Plants as source of drugs. *Toxicon*, 39(5), 603-613.
- Re, R., Pellegrini, N., Proteggente, A., Pannala, A., Yang, M., & Rice-Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biology & Medicine*, 26(9-10), 1231-1237.
- Reid, L. M., Mather, D. E., Arnason, J., Hamilton, T., & Bolton, J. R. (1992). Changes in phenolic constituent in maize silk infected with *Fusarium graminearum*. *Canadian Journal of Botany*, 70, 1697-1700.
- Renaud, S.C., Guéguen, R., Schenker, J., & d'Houtaud, A. (1998). Alcohol and mortality in middle-aged men from eastern France. *Epidemiology*, 9(2), 184-188.
- Rice-Evans, C. A., Miller, N. J., & Paganga, G. (1996). Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology & Medicine*, 20(7), 933-956.

- Rice-evans, C. A., Miller, N. J., Bolwell, P. G., Bramley, P. M., & Pridham, J. B. (1995). The relative antioxidant activities of plant-derived polyphenolic flavonoids. *Free Radical Research*, 22(4), 375-383.
- Riley, L., & Cowan, M. (2014). *Noncommunicable diseases country profiles 2014*. Retrieved from https://apps.who.int/bitstream/handle/10665/128038/9789241507509_eng.pdf.
- Rivlin, N., Brosh, R., Oren, M., & Rotter, V. (2011). Mutations in the p53 tumor suppressor gene: important milestones at the various steps of tumorigenesis. *Genes & Cancer*, 2(4), 466-474.
- Rodakiewicz-Nowak, J. (2000). Phenols oxidizing enzymes in waterrestricted media. *Topics in Catalysis*, 11/12, 419–434. doi:10.1023/ A:1027291629302.
- Rodrigues, C. E., Aracava, K. K., & Abreu, F. N. (2010). Thermodynamic and statistical analysis of soybean oil extraction process using renewable solvent. *International Journal of Food Science & Technology*, 45(11), 2407-2414.
- Rustaie, A., Hadjiakhoondi, A., Akbarzadeh, T., Safavi, M., Samadi, N., Sabourian, R., & Khanavi, M. (2018). Phytochemical constituents and biological activities of *Salvia suffruticosa*. *Research Journal of Pharmacognosy*, 5(2), 25-32.
- Ryan, L., & Sutherland, S. (2011). Comparison of the effects of different types of soya milk on the total antioxidant capacity of black tea infusions. *Food Research International*, 44(9), 3115-3117.
- Sabharwal, S. S., & Schumacker, P. T. (2014). Mitochondrial ROS in cancer: initiators, amplifiers or an Achilles' heel?. *Nature Reviews Cancer*, 14(11), 709.
- Şahin, B., Aygün, A., Gündüz, H., Şahin, K., Demir, E., Akocak, S., & Şen, F. (2017). Cytotoxic Effects of Platinum Nanoparticles Obtained from Pomegranate Extract by the Green Synthesis Method on the MCF-7 Cell Line. *Colloids & Surfaces B: Biointerfaces*, 163, 119-124.
- Sampath, S., Veeramani, V., Krishnakumar, G. S., Sivalingam, U., Madurai, S. L., & Chellan, R. (2017). Evaluation of in vitro anticancer activity of 1, 8-Cineole–containing n-hexane extract of *Callistemon citrinus* (Curtis) Skeels plant and its apoptotic potential. *Biomedicine & Pharmacotherapy*, 93, 296-307.
- Sánchez-González, I., Jiménez-Escrig, A., & Saura-Calixto, F. (2005). In vitro antioxidant activity of coffees brewed using different procedures (Italian, espresso and filter). *Food Chemistry*, 90(1-2), 133-139.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M., & Latha, L. Y. (2011). Extraction, isolation and characterization of bioactive compounds from plants' extracts. *African Journal of Traditional, Complementary & Alternative Medicines*, 8(1), 1-10.

- Saxena, D. K., Sharma, S. K., & Sambi, S. S. (2011). Kinetics and thermodynamics of cottonseed oil extraction. *Grasas y Aceites*, 62(2), 198-205.
- Scalzo, J., Politi, A., Pellegrini, N., Mezzetti, B., & Battino, M. (2005). Plant genotype affects total antioxidant capacity and phenolic contents in fruit. *Nutrition*, 21(2), 207-213.
- Schink, M., & Dehus, O. (2017). Effects of mistletoe products on pharmacokinetic drug turnover by inhibition and induction of cytochrome P450 activities. *BMC Complementary & Alternative Medicine*, 17(1), 521.
- Schmitt, M. W., Loeb, L. A., & Salk, J. J. (2016). The influence of subclonal resistance mutations on targeted cancer therapy. *Nature Reviews Clinical Oncology*, 13(6), 335.
- Schober, P., Boer, C., & Schwarte, L. A. (2018). Correlation coefficients: appropriate use and interpretation. *Anesthesia & Analgesia*, 126(5), 1763-1768.
- Schumacker, P. T. (2006). Reactive oxygen species in cancer cells: live by the sword, die by the sword. *Cancer Cell*, 10(3), 175-176.
- Shahidi, F. & Naczki, M. (1995). Food phenolics: Sources, Chemistry, Effects and Applications. Lancaster, PA: Technomic.
- Shamsi, T. N., Parveen, R., & Fatima, S. (2017). Panchakola Reduces Oxidative Stress in MCF-7 Breast Cancer and HEK293 Cells. *Journal of Dietary Supplements*, 15(5), 704-714.
- Shan, B., Cai, Y. Z., Sun, M., & Corke, H. (2005). Antioxidant capacity of 26 spice extracts and characterization of their phenolic constituents. *Journal of Agricultural & Food Chemistry*, 53(20), 7749-7759.
- Sharma, P., Hu-Lieskovan, S., Wargo, J. A., & Ribas, A. (2017). Primary, adaptive, and acquired resistance to cancer immunotherapy. *Cell*, 168(4), 707-723.
- Siddhuraju, P., & Becker, K. (2003). Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (*Moringa oleifera* Lam.) leaves. *Journal of Agricultural & Food Chemistry*, 51(8), 2144-2155.
- Siddique, N. A., Mujeeb, M., Najmi, A. K., & Akram, M. (2010). Evaluation of antioxidant activity, quantitative estimation of phenols and flavonoids in different parts of *Aegle marmelos*. *African Journal of Plant Science*, 4(1), 001-005.
- Siddiqui, Z.S. & Arif-uz-Zama. (2004). Effects of benlate systemic fungicide on seed germination, seedling growth, biomass and phenolic contents in two cultivars of *Zea mays* L.. *Pakistan Journal of Botany*, 36(3), 577-582.

- Siegel, R.L, Miller, K.D., Fedewa, S.A., Ahnen, D.J., Meester, R.G., Barzi, A., & Jemal, A. (2017). Colorectal cancer statistics, *CA: A Cancer Journal for Clinicians*, 67(3), 177-193.
- Siegfried, S. A., & Schroeder, J. R. (2018). Toxicity of thieves oils to mcf-7 and mda-mb-231 breast cancer cells. *American Journal of Essential Oils & Natural Products*, 6(1), 01-08.
- Silva-Beltrán, N. P., Ruiz-Cruz, S., Cira-Chávez, L. A., Estrada-Alvarado, M. I., Ornelas-Paz, J. D. J., López-Mata, M. A., ... & Márquez-Ríos, E. (2015). Total phenolic, flavonoid, tomatine, and tomatidine contents and antioxidant and antimicrobial activities of extracts of tomato plant. *International Journal of Analytical Chemistry*, 2015, 1-10.
- Singleton, V. L., & Rossi, J. A. (1965). Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American journal of Enology and Viticulture*, 16(3), 144-158.
- Sipe Jr, H. J., Jordan, S. J., Hanna, P. M., & Mason, R. P. (1994). The metabolism of 17 β -estradiol by lactoperoxidase: a possible source of oxidative stress in breast cancer. *Carcinogenesis*, 15(11), 2637-2643.
- Siti, H. N., Kamisah, Y., & Kamsiah, J. (2015). The role of oxidative stress, antioxidants and vascular inflammation in cardiovascular disease (a review). *Vascular Pharmacology*, 71, 40-56.
- Sneader, W. (2005). *Drug discovery: a history*. West Sussex, England: John Wiley & Sons.
- Soare, J. R., Dinis, T. C., Cunha, A. P., & Almeida, L. (1997). Antioxidant activities of some extracts of *Thymus zygis*. *Free Radical Research*, 26(5), 469-478.
- Sodde, V. K., Lobo, R., Kumar, N., Maheshwari, R., & Shreedhara, C. S. (2015). Cytotoxic activity of *Macrosolen parasiticus* (L.) Danser on the growth of breast cancer cell line (MCF-7). *Pharmacognosy Magazine*, 11(Suppl. 1), S156-S160.
- Soto-Vaca, A., Gutierrez, A., Losso, J. N., Xu, Z., & Finley, J. W. (2012). Evolution of phenolic compounds from color and flavor problems to health benefits. *Journal of Agricultural & Food Chemistry*, 60(27), 6658-6677.
- Sourani, Z., Pourgheysari, B., Beshkar, P., Shirzad, H., & Shirzad, M. (2016). Gallic acid inhibits proliferation and induces apoptosis in lymphoblastic leukemia cell line (C121). *Iranian Journal of Medical Sciences*, 41(6), 525-530.
- Sreeramulu, D., & Raghunath, M. (2010). Antioxidant activity and phenolic content of roots, tubers and vegetables commonly consumed in India. *Food Research International*, 43(4), 1017-1020.
- Sri, M. S. H. W. M., & Hartati, W. (1999). Toxicity screening using Brine Shrimp Lethality Test (BST) of some benalu species traditionally utilized to treat

- tumour in Indonesia. *Journal of the Medical Sciences (Berkala ilmu Kedokteran)*, 31(01), 17-22.
- Start, A. N. (2011). Some observations on an urban mistletoe *Dendrophthoe pentandra* (L.) Miq.(Loranthaceae) in Thailand. *Natural History Bulletin of the Siam Society*, 57, 81-86.
- Steinmetz, K. A., & Potter, J. D. (1991). Vegetables, fruit, and cancer. I. Epidemiology. *Cancer Causes & Control*, 2(5), 325-357.
- Stohs, S.J., & Bagchi, D. (1995). Oxidative mechanisms in the toxicity of metal ions. *Free Radical Biology Medicine*, 18(2), 321-336.
- Su, H. J., Hu, J. M., Anderson, F. E., Der, J. P., & Nickrent, D. L. (2015). Phylogenetic relationships of Santalales with insights into the origins of holoparasitic Balanophoraceae. *Taxon*, 64(3), 491-506.
- Su, Z. Y., Shu, L., Khor, T. O., Lee, J. H., Fuentes, F., & Kong, A. N. T. (2012). A perspective on dietary phytochemicals and cancer chemoprevention: oxidative stress, nrf2, and epigenomics. In *Natural Products in Cancer Prevention and Therapy* (pp. 133-162). Berlin, Heidelberg: Springer.
- Sultana, B., Anwar, F., & Przybylski, R. (2007). Antioxidant activity of phenolic components present in barks of *Azadirachta indica*, *Terminalia arjuna*, *Acacia nilotica*, and *Eugenia jambolana* Lam. trees. *Food Chemistry*, 104(3), 1106-1114.
- Suwarso, E. (2014). The Apoptosis Effects of Ethylacetate Extract of *Eleutherine bulbosa* (Mill.) Urb. Against T47D Cells. *International Journal of PharmTech Research*. 7(3), 535-539.
- Tagne, R. S., Telefo, B. P., Nyemb, J. N., Yemele, D. M., Njina, S. N., Goka, S. M. C., ... & Farooq, A. D. (2014). Anticancer and antioxidant activities of methanol extracts and fractions of some Cameroonian medicinal plants. *Asian Pacific Journal of Tropical Medicine*, 7, S442-S447.
- Tagoe, E.A., Arko-Boham, B., Adjorogbe, B., Aryee, N.A., Quaye, O., Adadey, S.M., & Lampitey, J.N.C. (2017). Glutathione S-Transferase T1 and M1 Gene Polymorphisms among Breast Cancer Susceptible Ghanaians. *Clinical Investigation (London)*, 7(3), 119-125.
- Tao, S. F., He, H. F., & Chen, Q. (2015). Quercetin inhibits proliferation and invasion acts by up-regulating miR-146a in human breast cancer cells. *Molecular & Cellular Biochemistry*, 402(1-2), 93-100.
- Tara, C. A., Wagner, M. L., Horacio, M. A., Roberto, P. I. N. O., & Gurni, A. A. (1994). Estudio farmacológico de un agente vasoactivo presente en *Ligaria cuneifolia* var, *cuneifolia*. *Acta Farmaceutica Bonaerense*, 13(2), 91-5.
- Tatiya, A. U., Tapadiya, G. G., Kotecha, S., & Surana, S. J. (2011). Effect of solvents on total phenolics, antioxidant and antimicrobial properties of *Bridelia retusa*

- Spreng. stem bark. *Indian Journal of Natural Products & Resources*, 2(4), 442-447.
- Tattini, M., Galardi, C., Pinelli, P., Massai, R., Remorini, D., & Agati, G. (2004). Differential accumulation of flavonoids and hydroxycinnamates in leaves of *Ligustrum vulgare* under excess light and drought stress. *New Phytologist*, 163(3), 547-561.
- Teh, S. S., Bekhit, A. E. D., & Birch, J. (2014). Antioxidative polyphenols from defatted oilseed cakes: effect of solvents. *Antioxidants*, 3(1), 67-80.
- Tekin, G., & Ozturk, B. R. (2017). Effects of Quercetin and Hesperetin on MCF-7 Cell Proliferation by Using Real-Time Cell Analyzer. *Journal of Basic & Clinical Pharmacy*, 8(3), 121-126.
- Temple, N.J. (2000). Antioxidants and disease: more questions than answers. *Nutrition Research*, 20, 449- 459.
- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L., & Byrne, D. H. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition & Analysis*, 19(6-7), 669-675.
- The Plant List (2018). *Helixanthera*. Retrieved from <http://www.theplantlist.org/1.1/browse/A/Loranthaceae/Helixanthera/>.
- Thoo, Y. Y., Ho, S. K., Liang, J. Y., Ho, C. W., & Tan, C. P. (2010). Effects of binary solvent extraction system, extraction time and extraction temperature on phenolic antioxidants and antioxidant capacity from mengkudu (*Morinda citrifolia*). *Food Chemistry*, 120(1), 290-295.
- Tir, R., Dutta, P. C., & Badjah-Hadj-Ahmed, A. Y. (2012). Effect of the extraction solvent polarity on the sesame seeds oil composition. *European Journal of Lipid Science & Technology*, 114(12), 1427-1438.
- Tiwari, P., Kumar, B., Kaur, M., Kaur, G., & Kaur, H. (2011). Phytochemical screening and extraction: a review. *Internationale pharmaceutica Scientia*, 1(1), 98-106.
- Tofigh, R., Akhavan, S., Tarban, N., Sadrabadi, A. E., Jalili, A., Moridi, K., & Tutunchi, S. (2017). Doxorubicin Induces Apoptosis through down Regulation of miR-21 Expression and Increases miR-21 Target Gene Expression in MCF-7 Breast Cancer Cells. *Journal of Clinical Medicine*, 8, 386-394.
- Torre, L. A., Bray, F., Siegel, R. L., Ferlay, J., Lortet-Tieulent, J., & Jemal, A. (2015). Global cancer statistics, 2012. *CA: A Cancer Journal for Clinicians*, 65(2), 87-108.
- Townsend, D. M., & Tew, K. D. (2003). The role of glutathione-S-transferase in anti-cancer drug resistance. *Oncogene*, 22(47), 7369-7375.

- Trachootham, D., Lu, W., Ogasawara, M. A., Valle, N. R. D., & Huang, P. (2008). Redox regulation of cell survival. *Antioxidants & redox signaling*, 10(8), 1343-1374.
- Trueba, G. P., Sánchez, G. M., & Giuliani, A. (2004). Oxygen free radical and antioxidant defense mechanism in cancer. *Frontiers in Bioscience*, 9(10), 2029-2044.
- Turkmen, N., Velioglu, Y. S., Sari, F., & Polat, G. (2007). Effect of extraction conditions on measured total polyphenol contents and antioxidant and antibacterial activities of black tea. *Molecules*, 12(3), 484-496.
- Uma, D. B., Ho, C. W., & Wan Aida, W. M. (2010). Optimization of extraction parameters of total phenolic compounds from henna (*Lawsonia inermis*) leaves. *Sains Malaysiana*, 39(1), 119-128.
- Unschuld, P. U. (1986). *Medicine in China: a history of pharmaceutics. Comparative studies of health systems and medical care* (1st ed.). CA, USA: University of California Press.
- Valko, M. M. H. C. M., Morris, H., & Cronin, M. T. D. (2005). Metals, toxicity and oxidative stress. *Current Medicinal Chemistry*, 12(10), 1161-1208.
- Valko, M., Leibfritz, D., Moncol, J., Cronin, M. T., Mazur, M., & Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *The International Journal of Biochemistry & Cell Biology*, 39(1), 44-84.
- Valko, M., Rhodes, C., Moncol, J., Izakovic, M. M., & Mazur, M. (2006). Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chemico-Biological Interactions*, 160(1), 1-40.
- Van Acker, S. A., Koymans, L. M., & Bast, A. (1993). Molecular pharmacology of vitamin E: structural aspects of antioxidant activity. *Free Radical Biology & Medicine*, 15(3), 311-328.
- Velioglu, Y. S., Mazza, G., Gao, L., & Oomah, B. D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *Journal of Agricultural & Food Chemistry*, 46(10), 4113-4117.
- Von Staszewski, M., Pilosof, A. M., & Jagus, R. J. (2011). Antioxidant and antimicrobial performance of different Argentinean green tea varieties as affected by whey proteins. *Food Chemistry*, 125(1), 186-192.
- Wan, P. J., & Wakelyn, P. J. (Eds.). (1997). *Technology and solvents for extracting oilseeds and nonpetroleum oils*. Champaign, IL: The American Oil Chemists Society.
- Wang, H., & Helliwell, K. (2001). Determination of flavonols in green and black tea leaves and green tea infusions by high-performance liquid chromatography. *Food Research International*, 34(2-3), 223-227.

- Wastuti, S., Suwarso, E. & Nasution, P. (2014). The cytotoxic test and apoptotic of *Solanum sanitwongsei* Craib against HeLa cells. *International Journal of PharmTech Research*, 7(1), 16-21.
- Wätjen, W., Michels, G., Steffan, B., Niering, P., Chovolou, Y., Kampkötter, A., ... & Kahl, R. (2005). Low concentrations of flavonoids are protective in rat H4IIE cells whereas high concentrations cause DNA damage and apoptosis. *The Journal of Nutrition*, 135(3), 525-531.
- Weidinger, A., & Kozlov, A. V. (2015). Biological activities of reactive oxygen and nitrogen species: oxidative stress versus signal transduction. *Biomolecules*, 5(2), 472-484.
- Werner, R. (2002). *Medicines in Malay Villages*. Kuala Lumpur, Malaysia: Universiti of Malaya Press.
- Wiert, C. (2006). *Medicinal plants of Asia and the Pacific*. Boca Raton, FL: CRC Press.
- Willcox, J. K., Ash, S. L., & Catignani, G. L. (2004). Antioxidants and prevention of chronic disease. *Critical Reviews in Food Science & Nutrition*, 44(4), 275-295.
- Williams, S. S. (1990). Mistletoe. Garden line porpouri miscellaneous/Mistletoe. Retrieved from <http://gardenline.usask.ca/misc/mistletoe.html>.
- Wogan, G. N., Hecht, S. S., Felton, J. S., Conney, A. H., & Loeb, L. A. (2004). Environmental and chemical carcinogenesis. *Seminars in Cancer Biology*, 14(6), 473-486.
- Wojdyło, A., Oszmiański, J., & Czemerys, R. (2007). Antioxidant activity and phenolic compounds in 32 selected herbs. *Food Chemistry*, 105(3), 940-949.
- Wong, C. C., Li, H. B., Cheng, K. W., & Chen, F. (2006). A systematic survey of antioxidant activity of 30 Chinese medicinal plants using the ferric reducing antioxidant power assay. *Food Chemistry*, 97(4), 705-711.
- Wong, D. Z. H., Kadir, H. A., Lee, C. L., & Goh, B. H. (2012). Neuroprotective properties of *Loranthus parasiticus* aqueous fraction against oxidative stress-induced damage in NG108-15 cells. *Journal of Natural Medicines*, 66(3), 544-551.
- Wong, S. P., Leong, L. P., & Koh, J. H. W. (2006). Antioxidant activities of aqueous extracts of selected plants. *Food Chemistry*, 99(4), 775-783.
- World Health Organization. (2018). *Global Health Observatory (GHO) data*. Retrieved from http://www.who.int/gho/ncd/mortality_morbidity/ncd_total/en.
- World Health Organization. (2013). *Global Action Plan for the Prevention and Control of Noncommunicable Diseases 2013–2020*. Retrieved from https://apps.who.int/iris/bitstream/handle/10665/94384/9789244506233_rus.pdf.

- Xiao, Y., Fan, Y., Chen, B., Zhang, Q., & Zeng, H. (2010). Polysaccharides from *Scurrula parasitica* L. inhibit sarcoma S180 growth in mice. *Zhongguo Zhong yao za zhi= Zhongguo zhongyao zazhi= China Journal of Chinese Materia Medica*, 35(3), 381-384.
- Xie, P. J., Huang, L. X., Zhang, C. H., Ding, S. S., Deng, Y. J., & Wang, X. J. (2018). Skin-care effects of dandelion leaf extract and stem extract: Antioxidant properties, tyrosinase inhibitory and molecular docking simulations. *Industrial Crops & Products*, 111, 238-246
- Yang, Y., Chen, M., & Sha, C. (2011). Triterpenoids and triterpenoid saponins of *Viscum liquidambaricolum*. *Zhongguo Zhong yao za zhi= Zhongguo zhongyao zazhi= China Journal of Chinese Materia Medica*, 36(2), 162-165.
- Yilmaz, Y., & Toledo, R. T. (2006). Oxygen radical absorbance capacities of grape/wine industry byproducts and effect of solvent type on extraction of grape seed polyphenols. *Journal of Food Composition & Analysis*, 19(1), 41-48.
- Young, I. S., & Woodside, J. V. (2001). Antioxidants in health and disease. *Journal of Clinical Pathology*, 54(3), 176-186.
- Zafra-Stone, S., Yasmin, T., Bagchi, M., Chatterjee, A., Vinson, J. A., & Bagchi, D. (2007). Berry anthocyanins as novel antioxidants in human health and disease prevention. *Molecular Nutrition & Food Research*, 51(6), 675-683.
- Zahreddine, H., & Borden, K. (2013). Mechanisms and insights into drug resistance in cancer. *Frontiers in pharmacology*, 4, 28.
- Zainal Ariffin, O., & Nor Saleha, I. T. (2011). *National cancer registry report 2007*. Putrajaya, Malaysia: National Cancer Registry, Ministry of Health.
- Zhang, S. S., Gong, Z. J., Li, W. H., Wang, X., & Ling, T. Y. (2012). Antifibrotic effect of curcumin in TGF- β 1-induced myofibroblasts from human oral mucosa. *Asian Pacific Journal of Cancer Prevention*, 13(1), 289-294.
- Zhao, F., Zhao, J., Song, L., Zhang, Y. Q., Guo, Z., & Yang, K. H. (2017). The induction of apoptosis and autophagy in human hepatoma SMMC-7721 cells by combined treatment with vitamin C and polysaccharides extracted from *Grifola frondosa*. *Apoptosis*, 22(11), 1461-1472.
- Zhao, Y. L., Wang, X. Y., Sun, L. X., Fan, R. H., Bi, K. S., & Zhi, G. Y. (2012). Cytotoxic constituents of *Viscum coloratum*. *Zeitschrift für Naturforschung C*, 67(3-4), 129-134.
- Zieliński, H., & Kozłowska, H. (2000). Antioxidant activity and total phenolics in selected cereal grains and their different morphological fractions. *Journal of Agricultural & Food Chemistry*, 48(6), 2008-2016.
- Zorofchian Moghadamtousi, S., Hajrezaei, M., Abdul Kadir, H., & Zandi, K. (2013). *Loranthus micranthus* Linn.: biological activities and

phytochemistry. *Evidence-Based Complementary & Alternative Medicine*, 2013, 1-9.

- Zoubi, M. S. A., Zavaglia, K., Mazanti, C., Hamad, M. A., Batayneh, K. A., Aljabali, A. A., & Bevilacqua, G. (2017). Polymorphisms and mutations in GSTP1, RAD51, XRCC1 and XRCC3 genes in breast cancer patients. *The International Journal of Biological Markers*, 32(3), 337-343.
- Zuo, Y., Chen, H., & Deng, Y. (2002). Simultaneous determination of catechins, caffeine and gallic acids in green, Oolong, black and pu-erh teas using HPLC with a photodiode array detector. *Talanta*, 57(2), 307-316.

BIODATA OF STUDENT

The student was born in 1992, January 27th at Hospital Lumut, Perak, Malaysia. She attended her primary education at S.K. Toh Indera Wangsa Ahmad, Batu Gajah, and followed by secondary education at S.M.K. Toh Indera Wangsa Ahmad, Batu Gajah. Her interest in pursuing higher studies enabled herself to enroll into Foundation in Agricultural Program at Universiti Putra Malaysia and right away continuing her degree program in Bachelor Science (Honours) Major in Biology from 2011 to 2015.

The student has ample of experiences conducting research related to algae namely Spirulina, dinoflagellates, genetic related studies, plant sciences and several other fields. One of her biggest involvement in research is through her participation in researches conducted at University of Chungnam, Daejeon, South Korea. Candidate has huge interest and passion in performing biology related studies and now her interest is focused to healthcare and plant ethnobotany. She has equipped herself with knowledge on biochemistry and cell handling through participation in classes and coach from postgraduate colleagues. Looking at her interest and capability, the author pursued her studies in degree of Master in Science under the field of Cancer Biology and together related to Biodiversity and Conservation of Natural Resources in 2016. With her studies focused on postharvest treatments, antioxidative properties and anticancer responses of wild medicinal mistletoe plant, the author has completed her Master of Science program with a Thesis entitled “Antioxidant and Anticancer Properties of Crude Extracts from *Helizanthera parasitica* Lour. (Dedalu)”.



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