



**UNIVERSITI PUTRA MALAYSIA**

**EVALUATION OF TWO COMMERCIAL IMMUNOCHROMATOGRAPHIC  
RAPID TESTS FOR DETECTION OF IgM ANTIBODIES AGAINST  
LEPTOSPIRES IN HUMAN SERUM**

**SITI NUR ALIA BINTI RAMLI**

**FPSK(m) 2019 69**



**EVALUATION OF TWO COMMERCIAL IMMUNOCHROMATOGRAPHIC  
RAPID TESTS FOR DETECTION OF IgM ANTIBODIES AGAINST  
LEPTOSPIRES IN HUMAN SERUM**

By

SITI NUR ALIA BINTI RAMLI

Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of  
Master of Science

November 2018

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfilment of the requirement for the degree of Master of Science

**EVALUATION OF TWO COMMERCIAL IMMUNOCHROMATOGRAPHIC  
RAPID TESTS FOR DETECTION OF IgM ANTIBODIES AGAINST  
LEPTOSPIRES IN HUMAN SERUM**

By

**SITI NUR ALIA BINTI RAMLI**

**November 2018**

**Chairman : Associate Professor Vasantha Kumari Neela, PhD**  
**Faculty : Medicine and Health Sciences**

Leptospirosis is a zoonotic disease which shows the increasing cases over the recent years. In Malaysia, leptospirosis became a notifiable disease in 2011. The treatment of leptospirosis is not difficult, however, the diagnosis is a big challenge. Leptospirosis is a biphasic infection, where the first phase (acute or septic phase) is from 3 to 7 days and followed by the second phase that happens from 7 to 14 days. The patients are mostly asymptomatic in the first few days. During infection, the bacteria can be found in the blood. Then, it moves to kidney and starts shedding in the urine. Antibodies start to develop into detectable amount during the second phase. Diagnosis of leptospirosis involves testing for the presence of bacteria or their DNA in the first phase and antibodies in the second phase. Since the actual onset of illness is not clear, laboratory diagnosis should include both microbiological and serological tests. Clinical diagnosis is another big challenge as the symptoms are similar to several febrile illnesses in the tropics. However, clinical suspicion and positive laboratory investigation are very important to start the therapy. The microbiological examination involves culturing the bacteria and PCR based detection which needs DNA from the clinical samples. However, microbiology is not useful for rapid and early diagnosis as *Leptospira* needs minimum 2 to 4 weeks to grow, while PCR needs DNA extraction, technicalities, and expensive PCR machines and reagents. In the context of serology, microscopic agglutination test, which is the gold standard, is technically tedious as well as the interpretations are very subjective. Most importantly, it also requires a large of live cultures. From the hospital point of view, a rapid test, which at least be able to give a laboratory indication, will be sufficient to direct the treatment options. Many hospitals do not have the facilities to perform MAT or PCR and the assays ; moreover, will take few days. To date, there are several commercial rapid tests which detect IgM specific to *Leptospira*

antigens. The sensitivity and specificity of those tests vary geographical regions. The present study was aimed at evaluating the diagnostic accuracy of two rapid tests namely Leptocheck-WB (Zephyr Biomedicals, India) and ImmuneMed IgM Duo Rapid (IMMUNEMED, Inc Korea) for detection of acute leptospirosis in human. The study used two approaches namely prospective and retrospective evaluation to determine the diagnostic accuracy in the two rapid tests. In the prospective approach, the evaluation was performed on clinically suspected leptospirosis patients from a single hospital and used MAT and qPCR as the reference test. The accuracy of Serion ELISA *classic* also had been evaluated and results of diagnostic accuracy were compared to Leptocheck-WB and ImmuneMed IgM Duo Rapid. In the retrospective approach, the confirmed leptospirosis positive samples were obtained from the Public Health Laboratory Kota Bharu. The samples were evaluated and used only MAT results as the reference test. In the prospective evaluation, among the 50 blood samples tested from clinically suspected leptospirosis cases, six were positive MAT and 13 were positive qPCR. The Leptocheck-WB showed sensitivity and specificity of 47.3% and 80.65% respectively. ImmuneMed IgM Duo Rapid showed sensitivity and specificity of 15.79% and 90.32% respectively. The results of sensitivity and specificity of Leptocheck-WB and ImmuneMed IgM Duo in prospective evaluation samples were compared to the result from Serion ELISA *classic*. Serion ELISA *classic* showed 26.32% sensitivity and 83.87% specificity. For retrospective samples, Leptocheck-WB showed sensitivity and specificity of 90.72% and 76.32% respectively. ImmuneMed IgM Duo Rapid showed sensitivity and specificity of 40.21% and 89.47 respectively. In conclusion, from the present study for both prospective and retrospective samples, Leptocheck-WB was found to be more sensitive in detecting acute leptospirosis in human while ImmuneMed IgM Duo Rapid was found to be more specific in detecting acute leptospirosis.

**Keywords:** Leptospirosis, Rapid Diagnostic test, Immunochromatographic Test; Serological Tests; Serum IgM Antibodies, Prospective samples, Retrospective sample.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

**PENILAIAN TERHADAP DUA UJIAN IMMUNOKROMATOGRAFIK SEGERA  
UNTUK MENGENALPASTI ANTIBODI IgM TERHADAP *LEPTOSPIRA*  
DALAM SERUM MANUSIA**

Oleh

**SITI NUR ALIA BINTI RAMLI**

**November 2018**

**Pengerusi : Profesor Madya Vasantha Kumari Neela, PhD**  
**Fakulti : Perubatan dan Sains Kesihatan**

Leptospirosis adalah wabak zoonosis yang menunjukkan peningkatan kes dalam beberapa tahun lepas. Di Malaysia, Leptospirosis telah menjadi wabak yang perlu diambil perhatian pada tahun 2011. Perawatan leptospirosis tidak susah, walaubagaimanapun, diagnosisnya adalah cabaran yang besar. Leptospirosis adalah penyakit dwifasa, yang mana pada fasa pertama (fasa akut atau septik) adalah dari 3 hingga 7 hari dan diikuti dengan fasa kedua yang berlaku dari 7 hingga 14 hari. Pesakit kebanyakannya bersifat asimptomatik pada beberapa hari pertama. Ketika jangkitan, bakteria boleh dijumpai dalam darah. Kemudian, ia bergerak ke buah pinggang dan mula menggelupas di dalam darah. Antibodi mula terbina dalam bentuk yang mudah dikesan ketika fasa kedua. Diagnosis Leptospirosis melibatkan ujian untuk mengesan bakteria atau DNA pesakit dalam fasa pertama dan antibodi pada fasa kedua. Oleh kerana mula penyakit ini tidak begitu jelas, diagnosis makmal sepatutnya melibatkan kedua dua ujian mikrobiologi dan serologi. Diagnosis klinikal ialah merupakan satu lagi ujian besar kerana simptomnya sama dengan beberapa penyakit febril di Kawasan tropika. ‘Clinical Suspision’ dan siasatan ‘Positive laboratory’ adalah sangat penting untuk memulakan terapi. Pemeriksaan mikrobiologi adalah melibatkan pengkulturan bakteria dan pengesan berdasarkan PCR yang memerlukan DNA daripada sampel klinikal. Walaubagaimanapun, mikrobiologi tidak berkesan untuk diagnosis awal dan pantas kerana *Leptospira* memerlukan 2 hingga 4 minggu minimum untuk pertumbuhan; pada masa yang sama PCR memerlukan ekstrak DNA, i.e teknikal serta mesin PCR yang mahal dan bahan uji tak organik. Dalam konteks serologi, ujian aglutinasi mikroskop, yang merupakan standard emas, adalah rumit secara teknikalnya dan interpretasinya sangat subjektif. Yang paling penting, ia juga memerlukan kultur hidup yang banyak. Dalam konteks pandangan hospital, uji cepat, yang sekurang-kurangnya memberikan penanda makmal, sudah cukup untuk menunjukkan pilihan rawatan. Kebanyakan hospital tidak mempunyai kemudahan untuk menjalankan

MAT dan PCR serta menilai keberkesanannya; selain itu, ia memakan masa beberapa hari. Kini, terdapat beberapa uji cepat komersial yang mengesan spesifik IgM kepada antigen Leptospira. Sensitiviti dan spesifikasi ujian tersebut berlainan mengikut Kawasan geografi. Kajian ini bertujuan untuk menilai ketepatan diagnosis dua uji cepat iaitu Leptocheck-WB (Zephyr Biomedicals, India) and ImmuneMed IgM Duo Rapid (IMMUNEMED, Inc Korea) untuk pengesan Leptospirosis akut pada manusia. Kajian ini menggunakan dua pendekatan iaitu penilaian prospektif dan retrospektif untuk menegalkan pasti ketepatan diagnosis dalam dua uji cepat. Dari segi pendekatan prospektif, penilaian telah dilakukan ke atas pesakit leptospirosis yang disyaki secara klinikal dari sebuah hospital dan telah menggunakan MAT dan qPCR sebagai rujukan ujian. Ketepatan Serion ELISA classic juga telah dinilai dan keputusan ketepatan diagnosis dibandingkan dengan Leptocheck-WB dan ImmuneMed IgM Duo Rapid. Dari segi pendekatan retrospektif, sampel positif leptospirosis yang telah dipastikan diambil daripada Makmal Kesihatan Awam Kota Bharu. Sampel tersebut dinilai dan menggunakan hanya hasil keputusan MAT sebagai rujukan ujian. Dalam penilaian prospektif, daripada 50 sampel darah dari kes yang disuspek leptospirosis secara klinikal, enam antaranya positif kepada MAT dan 13 positif kepada qPCR. Leptocheck-WB telah menunjukkan sensitiviti dan spesifikasi masing-masing adalah 47.3% dan 80.65%. ImmuneMed IgM Duo Rapid telah menunjukkan sensitiviti dan spesifikasi masing-masing adalah 15.79% dan 90.32%. Keputusan sensitiviti dan spesifikasi Leptocheck-WB dan ImmuneMed IgM Duo dalam sampel penilaian prospectif telah dibandingkan dengan keputusan daripada Serion ELISA *classic*. Serion ELISA classic menunjukkan keputusan sensitiviti pada 26.32% dan spesifikasi pada 83.87%. Dari sudut lain bagi sampel retrospectif, Leptocheck-WB telah menunjukkan sensitiviti dan spesifikasi masing-masing 90.72% dan 76.32%. ImmuneMed IgM Duo Rapid telah menunjukkan sensitiviti dan spesifikasi masing-masing 40.21% and 89.47. Kesimpulannya, berdasarkan kedua-dua kajian prospektif dan retrospektif ini, Leptocheck-WB dilihat lebih sensitif dalam pengesan leptospirosis akut pada manusia, manakala ImmuneMed IgM Duo Rapid lebih spesifik dalam mengesan leptospirosis.

**Kata kunci:** Leptospirosis, Ujian Diagnostik Pantas, Ujian Immunokromatografik, Ujian Serologi; Serum IgM Antibodi, sampel Prospektif, sampel Retrospektif.

## **ACKNOWLEDGEMENTS**

Firstly, and most important, I would like to show my gratitude to the Almighty **ALLAH S.W.T.**, the creator that has always guided me to work on the right path, giving me strength and his blessing throughout this challenging research work. Secondly, I would like to show my greatest appreciation to my supervisory committee, Prof. Madya Dr. Vasantha Kumari Neela (main supervisor), and Dr. Siti Norbaya Masri for their guidance and supervision from the beginning till the end of the study.

Many thanks to my family especially my husband Muhammad Khairul Anuar Hj Yusop for his support, understanding and sacrifice he made to take care our loving baby Batrisya Qowiy. I also would like to thank my parents, Ramli Othman and Siti Ajar Ismail, my parent in-laws Hj Yusop and Hjh Nongnisa, as well my family members for their prayers and support.

This appreciation is also dedicated to all the lectures, my laboratory mates (Dr. Narcisse, Tasya, Huda, Amira and Norliza) and staff of Department of Medical Microbiology and Parasitology (Kak Iman and Kak farah) for their help and support. Special thanks to Public Health Laboratory from Kota Bharu Kelantan and Ministry of Education.

I aslo take this opportunity to thank Long Term Reseachr Grant Scheme (LRGS) (NO. Grant 5526403) for providing research grant and University Putra Malaysia for the facilities and opportunity to pursue this project.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Vasantha Kumari Neela, PhD**

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Chairman)

**Siti Norbaya Masri, PhD**

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 11 April 2019

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_

Date: \_\_\_\_\_

Name and Matric No.: Siti Nur Alia binti Ramli, GS45010

### **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Name of Chairman  
of Supervisory  
Committee:

---

Associate Professor

Dr. Vasantha Kumari Neela

Signature:

Name of Member  
of Supervisory  
Committee:

---

Associate Professor

Dr. Siti Norbaya Masri

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xi
<b>LIST OF FIGURES</b>	xii
<b>LIST OF ABBREVIATIONS</b>	xiv
 <b>CHAPTER</b>	
<b>1      INTRODUCTION</b>	<b>1</b>
1.1     Study Background	1
1.2     Problem Statement	2
1.3     Research Justification	2
1.4     Objectives	3
1.4.1    General Objective	3
1.4.2    Specific objectives	3
<b>2      LITERATURE REVIEW</b>	<b>4</b>
2.1     History and background of Leptospirosis and <i>Leptospira</i>	4
2.2     Leptospirosis Incidence	5
2.3 <i>Leptospira</i>	7
2.3.1    Structure	7
2.3.2    Growth	8
2.4     Leptospirosis	9
2.4.1    Diagnosis	9
2.4.2    Conventional Method	10
2.4.2.1    Dark-field microscopy	10
2.4.2.2    Immunofluorescence	10
2.4.2.3    Culture	11
2.4.3    Molecular Method	11
2.4.3.1    PCR and Quantitative PCR	11
2.4.3.2    Loop-mediated isothermal amplification (LAMP)	11
2.4.4    Serological Method	12
2.4.4.1    MAT	12
2.4.4.2    ELISA	12
2.4.4.3    Indirect Hemagglutination Assay (IHA)	13
2.4.4.4    Lateral Flow Assays (LFA)	13
2.5     Rapid Diagnostic Test (RDTs)	14
2.5.1    Challenges with the current rapid test used in Malaysia	15
2.6     Antibiotic Treatment	15

<b>3</b>	<b>METHODOLOGY</b>	16
3.1	Study Design	16
3.1.1	Ethical Approval	16
3.2	Leptospirosis Case Definition	18
3.3	Retrospective Evaluation Approach	18
3.3.1	Sample Size	19
3.3.2	Microscopic Agglutination Test (MAT) Approach (Antibody Detection)	19
3.3.3	Rapid Diagnosis of Acute Leptospirosis Using Immunochromatographic test (Antibody Detection)	21
3.3.3.1	Leptocheck-WB	22
3.3.3.2	ImmuneMed Leptospira IgM Duo Rapid	23
3.4	Prospective Evaluation Approach	25
3.4.1	Sample Size	26
3.4.2	Sample Collection	26
3.4.3	Blood Sample Processing	27
3.4.4	Microscopic Agglutination Test (MAT) Approach (Antibody Detection)	27
3.4.5	<i>LipL32</i> Gene Detection by Real-time PCR (qPCR) Approach ( <i>Leptospira</i> Antigen Detection)	27
3.4.6	Enzyme Linked Immunosorbent Assay (ELISA) Approach (Antibody Detection)	28
3.4.7	Rapid Diagnosis of Acute Leptospirosis Using Immunochromatographic test (Antibody Detection)	29
3.5	Data Analysis	29
<b>4</b>	<b>RESULTS</b>	31
4.1	Retrospective Evaluation Approach	31
4.1.1	Microscopic Agglutination Test Approach (Antibody Detection)	31
4.1.2	Rapid Diagnosis of Acute Leptospirosis Using Immunochromatographic test (Antibody Detection)	32
4.1.2.1	Cross-Reactivity	33
4.2	Prospective Evaluation Approach	33
4.2.1	Microscopic Agglutination Test Approach (Antibody Detection)	33
4.2.2	<i>LipL32</i> Gene Detection By Real-Time PCR (qPCR) (Antigen Detection)	34
4.2.3	Enzyme Linked Immunosorbent Assay (ELISA) Approach (Antibody Detection)	35
4.2.4	Rapid Diagnosis of Acute Leptospirosis Using Immunochromatographic test (Antibody Detection)	36

4.2.4.1	Sensitivity and specificity of Leptocheck-WB and ImmuneMed IgM Duo Rapid	38
4.2.4.2	Time Based Evaluation	38
4.3	Summary of The Diagnostic Performance of the RDTs	38
<b>5</b>	<b>DISCUSSION</b>	<b>40</b>
5.1	Discussion	40
<b>6</b>	<b>SUMMARY AND CONCLUSION, STUDY LIMITATIONS, AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>44</b>
6.1	Summary and Conclusion	44
6.2	Study Limitations	44
6.3	Recommendations for Future Research	44
<b>REFERENCES</b>		<b>45</b>
<b>APPENDICES</b>		<b>53</b>
<b>BIODATA OF STUDENT</b>		<b>66</b>
<b>PUBLICATION</b>		<b>67</b>

## LIST OF TABLES

<b>Table</b>		<b>Page</b>
2.1	Evaluation of diagnostic test	14
3.1	Clinical symptoms of leptospirosis	18
4.1	MAT titer and serogroup of samples patients with leptospirosis. The samples were obtained from Public Health Laboratory Kota Bharu	31
4.2	Positive test of Leptocheck-WB, ImmuneMed IgM Duo Rapid and ELISA at different day post onset of symptoms for all Hospital Serdang samples	38
4.3	Performance of Leptocheck -WB, ImmuneMed IgM Duo Rapid and Serion ELISA <i>classic</i> in detecting leptospirosis	39

## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
2.1	<i>Leptospira</i> demonstrated by Stimson (1907) using Levedatii silver deposition staining technique. (Extracted from the book of Current Topics in Microbiology and Immunology and Leptospirosis, Adler 2015)	5
2.2	Number of leptospirosis cases in Malaysia from the year 2006 to September 2018, Disease Control Division, Ministry of Health 2018	6
2.3	Number of deaths due to leptospirosis from the year 2006 to September 2018, Disease Control Division, Ministry of Health 2018	6
2.4	Top 10 ranking of leptospirosis global incidence from 1996 until 2008	7
2.5	Scanning electron micrograph of serovar patoc Patoc 1	8
2.6	Biphasic nature of leptospirosis and relevant investigations at different stages of the disease	9
2.7	Methods involved in the diagnosis of leptospirosis	10
2.8	Illustration of basis Lateral flow assays LFA	13
3.1	Flow chart of simplified study design for evaluation of immunochromatographic rapid test	17
3.2	Illustration of 96 well microtiter plate for screening of infected serovar	20
3.3	Illustration of 96 well microtiter plate for titration of patient's antibody against <i>Leptospira</i> antigen	21
3.4	Leptocheck WB	22
3.5	Illustration of interpretation results for Leptocheck-WB	23
3.6	ImmuneMed <i>Leptospira</i> IgM Duo Rapid	24
3.7	Illustration of interpretation result of ImmuneMed IgM Duo Rapid	25
3.8	Example of conventional two by two table	30

4.1	Flowchart showing of retrospective samples and rapid diagnostic tests for Public Health Laboratory Kota Bharu, Kelantan	32
4.2	Negative control contain leptospires live culture against patient's serum under DFM (20x magnification)	34
4.3	Positive MAT with 1:800 titer under DFM (20x magnification) live culture 50 % less than negative control in Figure 4.2	34
4.4	Flowchart showing the patients and the results of Serion ELISA <i>classic</i> test	35
4.5	Flowchart showing the prospective samples and the results of rapid diagnostic tests for Hospital Serdang samples	36
4.6	Comparison of RDTs with Reference test (qPCR = Real-time PCR, MAT = Microscopic Agglutination Test, LC = Leptocheck WB, IM = ImmuneMed IgM Duo Rapid)	37

## **LIST OF ABBREVIATIONS**

Buffer AL	DNA Binding Buffer
Buffer AW1	Wash Buffer
Buffer AW2	Desalting Buffer
Buffer AE	Elution Buffer
CFS	Cerebral spinal fluid
DFM	Dark-field Microscope
DNA	Deoxyribonucleic acid
EDTA	Ethylenediaminetetraacid acid
EMJH	Liquid Ellinghausen, Mccullough, Johnson, Harris
ELISA	Enzyme Linked Immunosorbent Assay
GBD	Global Burden Disease
IU/ $\mu$ l	International unit per microliter
LPS	Lipopolysaccharide
MAT	Microscopic Agglutination Test
mg	Milligram
mg/ml	Milligram per milliliter
min	Minute
ml	Milliliter
MOH	Ministry of Health
nm	Nanometer
OD	Optical density
PBS	Phosphate Buffered Saline
PCR	Polymerase chain reaction

pmol	Picmoles
POC	Point of care
pH	Potential of hydrogen
qPCR	Quantitative polymerase chain reaction
RDT's	Rapid diagnostic tests
sec	Second
SPSS	Statistical Package for the Social Sciences
WHO	World Health Organization
$\mu\text{l}$	Microliter
%	Percent
$^{\circ}\text{C}$	Degree Celsius
5-FU	5-Fluorouracil

## CHAPTER 1

### INTRODUCTION

#### 1.1 Study Background

Leptospirosis is one of the emerging zoonotic diseases which are potentially fatal with worldwide distribution (Mathieu Picardeau et al., 2014). It is caused by the pathogenic spirochetes *Leptospira* and affects both human and animals (Haake & Levett, 2015). Classified as a Gram-negative bacterium, *Leptospira* replicates and maintained in nature through chronic renal colonization of rodents and other small mammals. However, leptospirosis in rodents are asymptomatic and the animals appear healthy even when *Leptospira* bacteria are constantly replicating in their kidneys and excreted through (Noguchi, 1917; Stokes et al., 1917). Morphological alteration were reported in a study on bacteria isolated from kidney of rodents (Tucunduva de Faria et al., 2007). The *Leptospira* colonize by forming an amorphous, biofilm-like structure in the renal tubules of the asymptomatic carrier (Thiermann, 1981). Humans acquire the infection through direct or indirect contact with the urine or tissue of an animal infected or colonized with *Leptospira*. The bacteria enter directly into the body through cuts and abrasion on skins, mucous membrane, conjunctivae, or aerosol inhalation of microscopic droplets. However, infection through inhalation of water or aerosol, animal bites or inter-human transmission has been rarely demonstrated (Musso & La Scola, 2013). Thus humans becomes accidental hosts after being infected by *Leptospira* (Sánchez-Montes, Espinosa-Martínez, Ríos-Muñoz, Berzunza-Cruz, & Becker, 2015). Clinical leptospirosis can be divided into “leptospiraemic” phase which happens four to seven days after onset of infection and “leptospiruric” phase that last for 4 to 30 days. Incubation period range from 2 to 20 days while acute symptoms usually develop from 7 to 12 days after infection (Eugene et al., 2015; Levett, 2001). *Leptospira* can be found in the bloodstream during the acute phase and disappear during the convalescent phase. Antibody generally appears during “leptospiruric” phase which begin with the appearance of IgM antibody and usually persist for several months (Chernukha, Shishkina, Baryshev, & Kokovin, 1976; Haake & Levett, 2015).

Leptospirosis cases were highly reported in tropical and subtropical areas due to the suitable condition for the *Leptospira* to spread in the environment (Musso & La Scola, 2013). For example, the Asia Pacific region (Seychelles, India, Sri Lanka, and Thailand) as well as in Latin America and the Caribbean were reported as the highest endemic countries with the incidence rate of over 10 per 1000,000 people (Pappas et al., 2008). From 873,000 annual cases, 48,000 (5.5%) were reported dead by WHO Leptospirosis Burden Epidemiology Reference Group (LERG) (Pappas et al., 2008).

Occupational and outdoor recreational activities that are related to exposure to animals contaminated water and soil emerged as strong risk factors. For example, people who are involved in the occupation such as agriculture, military troops and recreational activities such as fishing and swimming are at high possibility of acquiring leptospirosis (Adler & de la Peña Moctezuma, 2010; Chiat Han Chang, Riazi, Yunus, Osman, & Noordin, 2014). In Malaysia, there is an increasing trend of leptospirosis cases whereby 8291 cases were reported in year 2015 which higher than cases in 2014 (7806) (Wahab, 2015).

## 1.2 Problem Statement

Prevention, control and early detection of the causative agent are key factors to reduce mortality and morbidity. Several factors are important to reduce the severity of the leptospirosis infection such as rapid diagnosis, effective treatment, the awareness and knowledge among the population on diseases prevention. Diagnosis is always a challenge at the early stage of leptospirosis since shows a very common febrile illness symptoms, thus most of the physicians tend to misdiagnose it. In Malaysia, leptospirosis also commonly misdiagnosed with other febrile illness especially dengue. Microscopic agglutination test (MAT) has been used as the gold standard for confirmation of leptospirosis. However, MAT is less reliable as the early detection method since the body does not produce specific antibodies against *Leptospira* as soon as the infection occurs. Polymerase chain reaction (PCR) is a rapid, specific and sensitive method to detect the bacteria in the blood sample at the very early stage. However, PCR is not suitable after the leptospiraemic phase when the body produced antibodies that cleared the bacteria. Even though the PCR method is very sensitive, but not all laboratory can perform this method because of the high cost and technical challenge.

## 1.3 Research Justification

Detailed study on leptospirosis detection methods is still needed, especially rapid diagnosis at early stages of illness to help the physician in starting the right treatment regimen. The method should be easy to handle and affordable. Nowadays there are many rapid test kits available in the market, but not all these rapid test kits are suitable for every country due to different circulating serovars, antibody cut-titer values for MAT confirmation and coinfection with other local febrile illnesses. The present study was aimed at evaluating the diagnostic efficacies of two commercially available immunochromatographic based rapid diagnostic test (RDTs) namely Leptocheck-WB and ImmuneMed IgM Duo Rapid and Serion ELISA *classic* for early detection of leptospirosis. Leptocheck-WB was used in this study because reports from several other countries supported this kit for early detection and evaluation in their hospitals, while ImmuneMed IgM Duo Rapid was recently approved to be used as rapid test in hospitals in Malaysia. Serion ELISA *classic* was selected as it is being used in the common reference laboratory in Malaysia. Evaluation of the diagnostics sensitivity and

specificity of these tests would provide important data to guide Malaysians hospitals to choose the rapid test for the early screening of leptospirosis.

## **1.4 Objectives**

### **1.4.1 General Objective**

The study attempted to evaluate the accuracy of three diagnostic tests on samples for the early screening of leptospirosis.

### **1.4.2 Specific objectives**

1. To determine the diagnostic accuracy of two rapid diagnostic tests for the early detection of leptospirosis through prospective evaluation approach.
2. To evaluate the specificity and the sensitivity of the tests through a retrospective evaluation approach.
3. To compare the diagnostic accuracy of two RDTs with ELISA for early diagnosis of leptospirosis

## REFERENCES

- Adler, B. (2015). History of Leptospirosis and Leptospira. *Curr Top Microbiol Immunol*, 387, 1-9 .
- Adler, B., & de la Peña Moctezuma, A. (2010). Leptospira and leptospirosis. *Veterinary Microbiology*, 140(3–4), 287–296.
- Agampodi, S. (2012). Case definitions in Leptospirosis: a note to Sri Lankan researchers. *Sri Lankan Journal of Infectious Diseases*, 2(2), 55–57.
- Ahmad, S., Shah, S., & Ahmad, F. (2005). Laboratory diagnosis of leptospirosis. *J Postgrad Med*, 51, 195–200.
- Al-Yousif, Y., Anderson, J., Chard-Bergstrom, C., & Kapil, S. (2002). Development, evaluation, and application of lateral-flow immunoassay (immunochromatography) for detection of rotavirus in bovine fecal samples. *Clinical and Diagnostic Laboratory Immunology*, 9(3), 723–5.
- Azizi, S., Kheirandish, R., & Rahimi, E. (2014). Comparison of polymerase chain reaction and Warthin-Starry techniques to detect *Leptospira* spp. in kidneys of slaughtered cattle. *Onderstepoort Journal of Veterinary Research*, 81(1), 1–6.
- Baharudin, B., Napiah, R., Yasin, M., & Othman, A. (2012). Outbreak of Leptospirosis in Pauh, Arau, Perlis. Retrieved from Outbreak of Leptospirosis in Pauh Arau Perlis. Baharudin&publication year 2012
- Baratloo, A., Hosseini, M., Negida, A., & El Ashal, G. (2015). Part 1: Simple Definition and Calculation of Accuracy, Sensitivity and Specificity. *Emergency (Tehran, Iran)*, 3(2), 48–9.
- Benacer, D., Lin, K., Choung, N., Bin, K., Galloway, R. L., Hartskeerl, R. A., et al. (2016). Acta Tropica Epidemiology of human leptospirosis in Malaysia , 2004 – 2012. *Acta Tropica*, 157, 162–168.
- Blacksell, S. D., Smythe, L., Phetsouvanh, R., Dohnt, M., Hartskeerl, R., Symonds, M., et al. (2006). Limited diagnostic capacities of two commercial assays for the detection of *Leptospira* immunoglobulin M antibodies in Laos. *Clinical and Vaccine Immunology*, 13(10), 1166–1169.
- Bossuyt, P. M., Reitsma, J. B., Bruns, D. E., Gatsonis, C. A., Glasziou, P. P., Irwig, L. M., et al. (2003). Towards complete and accurate reporting of studies of diagnostic accuracy : the STARD initiative, 41–44.
- Bourhy, P., Bremont, S., Zinini, F., Giry, C., & Picardeau, M. (2011). Comparison of real-time PCR assays for detection of pathogenic *Leptospira* spp. in blood and identification of variations in target sequences. *Journal of Clinical Microbiology*, 49(6), 2154–2160.

- Bourhy, P., Collet, L., Brisse, S., & Picardeau, M. (2014). *Leptospira mayottensis* sp. nov., a pathogenic species of the genus *Leptospira* isolated from humans. *International Journal of Systematic and Evolutionary Microbiology*, 64, 4061–4067.
- Brandão, a P., Camargo, E. D., da Silva, E. D., Silva, M. V., & Abrão, R. V. (1998). Macroscopic agglutination test for rapid diagnosis of human leptospirosis. *Journal of Clinical Microbiology*, 36(11), 3138–3142.
- Brownlow, T., Kavanagh, O. V., Logan, E. F., Hartskeerl, R. a, Savage, R., Palmer, M. F., et al. (2014). “Leptorapide” - a one-step assay for rapid diagnosis of human leptospirosis. *Epidemiology and Infection*, 142(6), 1182–7.
- Bujang, M. A., & Adnan, T. H. (2016). Requirements for minimum sample size for sensitivity and specificity analysis. *Journal of Clinical and Diagnostic Research*, 10(10), YE01-YE06.
- Cameron, C. E. (2015). Leptospiral Structure, Physiology, and Metabolism. Springer, Berlin, Heidelberg. 21–41.
- Carleton, O., Charon, N. W., Allender, P., & O'Brien, S. (1979). Helix handedness of *Leptospira interrogans* as determined by scanning electron microscopy. *Journal of Bacteriology*, 137(3), 1413–1416.
- Chang, C. H., Amran, F., Riazi, M., Yunus, M. H., Osman, S., & Noordin, R. (2014). Evaluation of two commercial rapid tests for laboratory diagnosis of Leptospirosis in Malaysia. *Asian Pacific Journal of Tropical Disease*, 4(3), 246.
- Chang, C. H., Riazi, M., Yunus, M. H., Osman, S., & Noordin, R. (2014). Limited diagnostic value of two commercial rapid tests for acute leptospirosis detection in Malaysia. *Diagnostic Microbiology and Infectious Disease*, 80(4), 278–281.
- Chernukha, Y. G., Shishkina, Z. S., Baryshev, P. M., & Kokovin, I. L. (1976). The dynamics of IgM- and IgG-antibodies in leptospiral infection in man. *Zentralblatt Fur Bakteriologie, Parasitenkunde, Infektionskrankheiten Und Hygiene. Erste Abteilung Originale. Reihe A: Medizinische Mikrobiologie Und Parasitologie*, 236(2–3), 336–43.
- Cross, R., Ling, C., Day, N. P. J., McGready, R., & Paris, D. H. (2016). Revisiting doxycycline in pregnancy and early childhood--time to rebuild its reputation? *Expert Opinion on Drug Safety*, 15(3), 367–82.
- Cumberland, P., Everard, C. O. R., Wheeler, J. G., & Levett, P. N. (2001). Persistence of Anti-Leptospiral IgM , IgG and Agglutinating Antibodies in Patients Presenting with Acute Febrile Illness in Barbados 1979-1989. Springer, 17(7), 601–608.

- Effler, P. V., Bogard, A. K., Domen, H. Y., Higa, H. Y., Sasaki, D. M., & Katz, A. R. (2002). Evaluation of Eight Rapid Screening Tests for Acute Leptospirosis in Hawaii. *Journal of Clinical Microbiology*, 40(4), 1464–1469.
- Effler, P. V., Domen, H. Y., Bragg, S. L., Aye, T., & Sasaki, D. M. (2000). Evaluation of the indirect hemagglutination assay for diagnosis of acute leptospirosis in Hawaii. *Journal of Clinical Microbiology*, 38(3), 1081–4.
- El-Jalii, I. M., & Bahaman, A. R. (2004). A review of human leptospirosis in Malaysia. *Tropical Biomedicine*, 21(2), 113–119.
- Ellis, W. R., Hovind-Hougen, K., Moller, S. H., & Birch-Andresen, A. (1983). Morphological changes upon subculturing of freshly isolated strains of *Leptospira interrogans* serovar hardjo. *Zentralbl Bakteriol Mikrobiol Hyg A*, 255(2-3), 323–335.
- Eugene, E. J., Handunnetti, S. M., Wickramasinghe, S. A., Kalugalage, T. L., Rodrigo, C., Wickremesinghe, H., et al. (2015). Evaluation of two immunodiagnostic tests for early rapid diagnosis of leptospirosis in Sri Lanka: a preliminary study. *BMC Infectious Diseases*, 15, 319.
- Evangelista, K. V., & Coburn, J. (2010). Leptospira as an emerging pathogen: a review of its biology, pathogenesis and host immune responses. *Future Microbiology*, 5(9), 1413–1425.
- Faine, S. (1994). *Leptospira and leptospirosis*. Boca Raton, USA: CRC Press Inc, 140, 353.
- Fairuz, A., Rani, B. A., Ayu, M., & Hishamshah, I. (2007). Epidemiology of human leptospirosis in Malaysia. *17th European Congress of Clinical Microbiology and Infectious Diseases/25th International Congress of Chemotherapy*, S282–S282.
- Goris, M. G. A., Leeflang, M. M. G., Loden, M., Wagenaar, J. F. P., Klatser, P. R., Hartskeerl, R. A., & Boer, K. R. (2013). Prospective Evaluation of Three Rapid Diagnostic Tests for Diagnosis of Human Leptospirosis, 7(7), e2290.
- Goto, M., Honda, E., Ogura, A., Nomoto, A., & Hanaki, K.-I. (2009). Colorimetric detection of loop-mediated isothermal amplification reaction by using hydroxy naphthol blue. *BioTechniques*, 46(3), 167–172.
- Gussenhoven, G. C., van der Hoorn, M. A., Goris, M. G., Terpstra, W. J., Hartskeerl, R. A., Mol, B. W., et al. (1997). LEPTO dipstick, a dipstick assay for detection of *Leptospira*-specific immunoglobulin M antibodies in human sera. *Journal of Clinical Microbiology*, 35(1), 92–97.
- Gussenhoven, G. C., Van Der Hoorn, M. A. W. G., Goris, M. G. A., Terpstra, W. J., Hartskeerl, R. A., Mol, B. W., et al. (1997). LEPTO dipstick, a dipstick assay for detection of *Leptospira*-specific immunoglobulin M antibodies in human sera. *Journal of Clinical Microbiology*, 35(1), 92–97.

- Haake, D. A., & Levett, P. N. (2015). Leptospirosis in humans. *Curr Top Microbiol Immunol*, 387, 65–97.
- Hartskeerl, R. A., Collares-Pereira, M., & Ellis, W. A. (2011). Emergence, control and re-emerging leptospirosis: Dynamics of infection in the changing world. *Clinical Microbiology and Infection*, 17, 494–501.
- Hospenthal, D. R., & Murray, C. K. (2003). In vitro susceptibilities of seven *Leptospira* species to traditional and newer antibiotics. *Antimicrobial Agents and Chemotherapy*, 47(8), 2646–2648.
- Inada, R., Ido, Y., Hoki, R., Kaneko, R., & Ito, H. (1916). The Etiology, Mode Of Infection, And Specific Therapy Of Weil's Disease (Spirochaetosis Icterohaemorrhagica). *The Journal of Experimental Medicine*, 23(3), 377–402.
- Karuniawati, A., Yasmon, A., & Ningsih, I. (2012). Optimizing real-time PCR method to detect *Leptospira* spp . in human blood and urine specimens, 21(1), 13–17.
- Katz, A. R., Ansdell, V. E., Effler, P. V, Middleton, C. R., & Sasaki, D. M. (2001). Assessment of the clinical presentation and treatment of 353 cases of laboratory-confirmed leptospirosis in Hawaii, 1974-1998. *Clinical Infectious Diseases : An Official Publication of the Infectious Diseases Society of America*, 33(11), 1834–1841.
- Kee, S., Kim, I., Choi, M., & Chang, W. (1994). Detection of Leptospiral DNA by PCR, 32(4), 1035–1039.
- Ko, A. I., Galvão Reis, M., Ribeiro Dourado, C. M., Johnson, W. D., & Riley, L. W. (1999). Urban epidemic of severe leptospirosis in Brazil. *Lancet*, 354(9181), 820–825.
- Kusano, N., Hirashima, K., Kuwahara, M., Narahara, K., Imamura, T., Mimori, et al. (2007). Immunochromatographic assay for simple and rapid detection of Satsuma dwarf virus and related viruses using monoclonal antibodies. *Journal of General Plant Pathology*, 73(1), 66–71.
- Lantz, P. G., Abu al-Soud, W., Knutsson, R., Hahn-Hagerdal, B., & Radstrom, P. (2000). Biotechnical use of polymerase chain reaction for microbiological analysis of biological samples. *Biotechnology Annual Review*, 5, 87–130.
- Levett, P. N. (2001). Leptospirosis. *Clinical Microbiology Reviews*, 14(2), 296–326.
- Limmathurotsakul, D., Turner, E. L., Wuthiekanun, V., Thaipadungpanit, J., Suputtamongkol, Y., Chierakul, W., et al (2012). Fool's Gold: Why Imperfect Reference Tests Are Undermining the Evaluation of Novel Diagnostics: A Reevaluation of 5 Diagnostic Tests for Leptospirosis. *Clinical Infectious Diseases*, 55(3), 322–331.

- Mayer-Scholl, A., Hammerl, J. A., Schmidt, S., Ulrich, R. G., Pfeffer, M., Woll, D., et al. (2014). Leptospira spp. in rodents and shrews in Germany. *International Journal of Environmental Research and Public Health*, 11(8), 7562–7574.
- Miraglia, F., de Moraes, Z. M., Melville, P. A., Dias, R. A., & Vasconcellos, S. A. (2009). EMJH medium with 5-fluorouracil and nalidixic acid associated with serial dilution technique used to recover Leptospira spp from experimentally contaminated bovine semen. *Brazilian Journal of Microbiology : [Publication of the Brazilian Society for Microbiology]*, 40(1), 189–193.
- MOH. (2017). Case definitions for infectious diseases in Malaysia, 03(January), 110.
- Mori, Y., Nagamine, K., Tomita, N., & Notomi, T. (2001). Detection of Loop-Mediated Isothermal Amplification Reaction by Turbidity Derived from Magnesium Pyrophosphate Formation. *Biochemical and Biophysical Research Communications*, 289(1), 150–154.
- Murray, G. L., Srikram, A., Henry, R., Hartskeerl, R. A., Sermswan, R. W., & Adler, B. (2010). Mutations affecting Leptospira interrogans lipopolysaccharide attenuate virulence. *Molecular Microbiology*, 78(3), 701–709.
- Musso, D., & La Scola, B. (2013). Laboratory diagnosis of leptospirosis: A challenge. *Journal of Microbiology, Immunology and Infection*, 46(4), 245–252.
- Nahori, M.-A., Fournié-Amazouz, E., Que-Gewirth, N. S., Balloy, V., Chignard, M., Raetz, C. R. H., et al. (2005). Differential TLR recognition of leptospiral lipid A and lipopolysaccharide in murine and human cells. *Journal of Immunology (Baltimore, Md. : 1950)*, 175(9), 6022–6031.
- Niloofa, R., Fernando, N., De Silva, N. L., Karunananayake, L., Wickramasinghe, H., Dikmadugoda, N., et al. (2015). Diagnosis of leptospirosis: Comparison between microscopic agglutination test, IgM-ELISA and IgM rapid immunochromatography test. *PLoS ONE*, 10(6).
- Niloofa, R., Fernando, N., Silva, N. L. De, & Karunananayake, L. (2015). Diagnosis of Leptospirosis : Comparison between Microscopic Agglutination Test , IgM-ELISA and IgM Rapid Immunochromatography Test, 1–12.
- Noguchi, H. (1917). Spirochaeta Icterohaemorrhagiae In American Wild Rats And Its Relation To The Japanese And European Strains : First Paper. *The Journal of Experimental Medicine*, 25(5), 755–63.
- Notomi. Tsugunori, Hiroto. Okayama, Harumi. Masubuchi, Toshihito. Yonekawa, Keiko.Watanabe, Nobuyuki. Amino, T. H. (2000). Loop Mediated isothermal amplification (LAMP). *Reviews in Medical Virology*, 28(12), e63.

- Panwala, T., Rajdev, S., & Mulla, S. (2015). To Evaluate the Different Rapid Screening Tests for Diagnosis of Leptospirosis, 9(2), DC21-DC24.
- Pappas, G., & Cascio, A. (2006). Optimal treatment of leptospirosis: queries and projections. *International Journal of Antimicrobial Agents*, 28(6), 491–6.
- Pappas, G., Papadimitriou, P., Siozopoulou, V., Christou, L., & Akritidis, N. (2008). The globalization of leptospirosis : worldwide incidence trends.
- Parikh, R., Mathai, A., Parikh, S., Chandra Sekhar, G., & Thomas, R. (2008). Understanding and using sensitivity, specificity and predictive values. *Indian Journal of Ophthalmology*, 56(1), 45–50.
- Picardeau, M. (2013). Diagnosis and epidemiology of leptospirosis. *Medecine et Maladies Infectieuses*, 43(1), 1–9.
- Picardeau, M., Bertherat, E., Jancloes, M., Skouloudis, A. N., Durski, K., & Hartskeerl, R. A. (2014). Rapid tests for diagnosis of leptospirosis: Current tools and emerging technologies. *Diagnostic Microbiology and Infectious Disease*, 78(1), 1–8.
- Posthuma-Trumpie, G. A., Korf, J., & Van Amerongen, A. (2009). Lateral flow (immuno)assay: Its strengths, weaknesses, opportunities and threats. A literature survey. *Analytical and Bioanalytical Chemistry*, 393(2), 569–582.
- Sánchez-Montes, S., Espinosa-Martínez, D. V., Ríos-Muñoz, C. A., Berzunza-Cruz, M., & Becker, I. (2015). Leptospirosis in Mexico: Epidemiology and potential distribution of human cases. *PLoS ONE*, 10(7), 1–16.
- Sayyed Mousavi, M. N., Sadeghi, J., Aghazadeh, M., Asgharzadeh, M., & Samadi Kafil, H. (2017). Current advances in urban leptospirosis diagnosis. *Reviews in Medical Microbiology*, 28(3), 119–123.
- Schreier, S., Doungchawee, G., Chadsuthi, S., Triampo, D., & Triampo, W. (2013). Leptospirosis: current situation and trends of specific laboratory tests. *Expert Review of Clinical Immunology*, 9(3), 263–280.
- Shah, L. J., Mulla, S., Patel, M. G., & Vaghela, G. (2012). Outbreak Surveillance Report on Pulmonary Leptospirosis after a Heavy Floods during 2006 in South Gujarat. *National Journal of Medical Research*, 2(1), 42–46.
- Sharp, C. F., Smith, D. J. W., Tonge, J. I., McComb, D. E., & Chang, R. S. (1957). The Use of Erythrocyte Sensitizing Substance in the Diagnosis of Leptospiroses. *The American Journal of Tropical Medicine and Hygiene*, 6(1), 101-107.
- Smits, H. L., Eapen, C. K., Sugathan, S., Kuriakose, M., Gasem, M. H., Yersin, C., et al. (2001). Lateral-Flow Assay for Rapid Serodiagnosis of Human Leptospirosis, 8(1), 166–169.

- Smits, H. L., Van Der Hoorn, M. A. W. G., Goris, M. G. A., Gussenoven, G. C., Yersin, C., Sasaki, D. M., et al.(2000). Simple latex agglutination assay for rapid serodiagnosis of human leptospirosis. *Journal of Clinical Microbiology*, 38(3), 1272–1275.
- Sonthayanon, P., Chierakul, W., Wuthiekanun, V., Thaipadungpanit, J., Kalambaheti, T., Boonsilp, S., et al. (2011). Accuracy of loop-mediated isothermal amplification for diagnosis of human leptospirosis in Thailand. *The American Journal of Tropical Medicine and Hygiene*, 84(4), 614–20.
- Stimson, A. M. (1907). Note on an Organism Found in Yellow-Fever Tissue. *Public Health Reports (1896-1970)*, 22(18), 541.
- Stockard, J. L., Smadel, J. E., Maisey, C. W., Gleiser, C. A., Robinson, C. R., Levis, D. G., et al. (1957). Leptospirosis in Malaya. *The American Journal of Tropical Medicine and Hygiene*, 6(2), 238–256.
- Stokes, A., Ryle, J. A., & Tytler, W. H. (1917). Weil's Disease (Spirochaetosis Ictero-Haemorrhagica) in the British Army in Flanders. *British medical journal*, 2(2908), 413-417.
- Suppiah, J., Chan, S.-Y., Ng, M.-W., Khaw, Y.-S., Ching, S.-M., Mat-Nor, L. A., et al. (2017). Clinical predictors of dengue fever co-infected with leptospirosis among patients admitted for dengue fever - a pilot study. *Journal of Biomedical Science*, 24(1), 40.
- Suwancharoen, D., Sittiwichewong, B., & Wiratsudakul, A. (2016). Evaluation of loop-mediated isothermal amplification method (LAMP) for pathogenic <i>Leptospira</i> spp. detection with leptospires isolation and real-time PCR. *Journal of Veterinary Medical Science*, 78(8), 1299–1302.
- Thayaparan, S., Robertson, I. D., Fairuz, A., Suut, L., & Abdullah, M. T. (2013). Leptospirosis , an emerging zoonotic disease in Malaysia, 35(2), 123–132.
- Thiermann, A. B. (1981). The Norway rat as a selective chronic carrier of *Leptospira icterohaemorrhagiae*. *Journal of Wildlife Diseases*, 17(1), 39–43.
- Thornley, C. N., Baker, M. G., Weinstein, P., & Maas, E. W. (2002). Changing epidemiology of human leptospirosis in New Zealand. *Epidemiol. Infect.*, 128, 29–36.
- Tomita, N., Mori, Y., Kanda, H., & Notomi, T. (2008). Loop-mediated isothermal amplification (LAMP) of gene sequences and simple visual detection of products. *Nature Protocols*, 3(5), 877–882.
- Torgerson, P. R., Hagan, J. E., Costa, F., Calcagno, J., Kane, M., Martinez-Silveira, M. S., et al. (2015). Global Burden of Leptospirosis: Estimated in Terms of Disability Adjusted Life Years. *PLoS Neglected Tropical Diseases*, 9(10), 1–14.

- Torten, M., Shenberg, E., & Van der Hoeden, J. (1966). The use of immunofluorescence in the diagnosis of human leptospirosis by a genus-specific antigen. *The Journal of Infectious Diseases*, 116(5), 537–543.
- Toyokawa, T., Ohnishi, M., & Koizumi, N. (2011). Diagnosis of acute leptospirosis. *Expert Review of Anti-Infective Therapy*, 9(1), 111–121.
- Tucunduva de Faria, M., Athanazio, D. A., Gonçalves Ramos, E. A., Silva, E. F., Reis, M. G., & Ko, A. I. (2007). Morphological Alterations in the Kidney of Rats with Natural and Experimental Leptospira Infection. *Journal of Comparative Pathology*, 137(4), 231–238.
- Van Thiel, P. H. (1948). Diagnosis and treatment of leptospirosis. *Abstracts. International Congress on Tropical Medicine and Malaria* (4th : 1948 : Washington, D. C.), 56(4th Congr), 21.
- Von Lode, P. (2005). Point-of-care immunotesting: Approaching the analytical performance of central laboratory methods. *Clinical Biochemistry*, 38(7), 591–606.
- Wahab, Z. A. (2015). Epidemiology and Current Situation of Leptospirosis in Malaysia. *Persidangan Kesihatan Persekutuan Pihak Berkuasa Tempatan 2015.*, (8–9 September), 1–67.
- Watt, G., Linda Tuazon, M., Santiago, E., Padre, L. P., Calubaquib, C., Ranoa, C. P., & Laughlin, L. W. (1988). Placebo-Controlled Trial of Intravenous Penicillin for Severe and Late Leptospirosis. *The Lancet*, 331(8583), 433–435.
- Winslow, W. E., Merry, D. J., Pirc, M. L., & Devine, P. L. (1997). Evaluation of a Commercial Enzyme-Linked Immunosorbent Assay for Detection of Immunoglobulin M Antibody in Diagnosis of Human Leptospiral Infection, 35(8), 1938–1942.
- Wisseman, C. L., Traub, R., Gochenour, W. S., Smadel, J. E., & Lancaster, W. E. (1955). Leptospirosis of Man and Animals in Urban, Rural and Jungle Areas of Southeast Asia. *The American Journal of Tropical Medicine and Hygiene*, 4(1), 29–40.
- Wolgemuth, C. W., Charon, N. W., Goldstein, S. F., & Goldstein, R. E. (2006). The Flagellar Cytoskeleton of the Spirochetes. *Journal of Molecular Microbiology and Biotechnology*, 11(3–5), 221–227.
- Zuerner, R. L. (2005). Laboratory Maintenance of Pathogenic *Leptospira*. In *Current Protocols in Microbiology* (Vol. Chapter 12, p. Unit 12E.1). Hoboken, NJ, USA: John Wiley & Sons, Inc.

## **BIODATA OF STUDENT**

Siti Nur Alia binti Ramli was born in small Hospital in Raub on June 1<sup>st</sup>, 1992. She attended her primary school at Sekolah Kebangsaan Ulu Gali from 1999 to 2001, continued at Methodist Girls School Raub from 2002 to 2004. She continued her secondary school at Sekolah Menengah Kebangsaan Seri Raub and took her SPM in Merbok Mara Junior Sciences College. She pursued her Bachelor of Science (Hons) in Microbiology in Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia (UPM) Serdang and graduated with CGPA 3.4. She has been very active in 16<sup>th</sup> College, UPM and university activities. She also actives in the UPM student council election.

She continued to study Master's degree in Medical Microbiology field in 2015 under the supervision of Assoc. Prof. Dr. Vasantha Kumari Neela from the Department of Medical Microbiology and Parasitology, Faculty of Medicine and Health Sciences UPM. She aims to further her PhD soon.

## PUBLICATION

### Journal

Alia, S. N., Joseph, N., Philip, N., Azhari, N. N., Garba, B., Masri, S. N., Sekawi, Z., & Neela, V. K. (2019). Diagnostic accuracy of rapid diagnostic tests for the early detection of leptospirosis. *Journal of infection and public health*, 12(2), 263–269.



## UNIVERSITI PUTRA MALAYSIA

### STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : Second Semester 2018/2019

#### TITLE OF THESIS / PROJECT REPORT :

EVALUATION OF TWO COMMERCIAL IMMUNOCHROMATOGRAPHIC RAPID TESTS FOR  
DETECTION OF IgM ANTIBODIES AGAINST LEPTOSPIRES IN HUMAN SERUM

NAME OF STUDENT: SITI NUR ALIA BINTI RAMLI

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

\*Please tick (✓)

**CONFIDENTIAL**

(Contain confidential information under Official Secret Act 1972).

**RESTRICTED**

(Contains restricted information as specified by the organization/institution where research was done).

**OPEN ACCESS**

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

**PATENT**

Embargo from \_\_\_\_\_ until \_\_\_\_\_  
(date) (date)

**Approved by:**

(Signature of Student)  
New IC No/ Passport No.:

(Signature of Chairman of Supervisory Committee)  
Name:

Date :

Date :

[Note : If the thesis is **CONFIDENTIAL** or **RESTRICTED**, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted.]