

METABOLIC VARIATIONS IN BIOLOGICAL EVALUATION OF Clinacanthus nutans (Burm. f.) Lindau LEAF EXTRACTS ON LIPOPOLYSACCHARIDES-INDUCED NEUROINFLAMMATION IN RATS

AMALINA BINTI AHMAD AZAM

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By

AMALINA BINTI AHMAD AZAM

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February 2020

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DEDICATION

This thesis is dedicated to my lovely parents, family and friends



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

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Chair: Intan Safinar Ismail, PhD Institute: Bioscience

Neuroinflammation is a complex response of injury on any part of the brain resulted in the activation of glial cells, release of inflammatory mediators like cytokines and chemokines, reactive oxygen and nitrogen species, which is a pathological hallmark of many neurological disorders. Therefore, effective control of neuroinflammation is crucial to prevent the related diseases. In this study, the matured leaves of 9-week old *Clinacanthus nutans* (Burm. f.) Lindau (CN) extracted with water and two ethanolic (50% and 100%) phytochemical constituents were profiled by using proton Nuclear Magnetic Resonance (¹H NMR) metabolomics approach. The highest nitric oxide (NO) inhibitory activity, in the *in vitro* neuroinflammation model using the lipopolysaccharides (LPS)-induced BV2 cell line, was observed for aqueous extract with an IC₅₀ value of $336.2 \pm 4.7 \mu$ g/ml. Correlation between NO inhibitory activity and CN constituents by partial least square (PLS) analysis resulted in the identification of most potential metabolites responsible for the activity being schaftoside, acetate, propionate, alanine, and Clinacoside C.

The *in vivo* model of neuroinflammed Sprague Dawley rats induced with LPS was also done via the metabolomics approach. The findings from multivariate data analysis (MVDA) highlighted several similarities and dissimilarities in metabolites concentration in LPS-induced rats (LPS+water) and LPS-induced treated with CN extracts. Although CN doses treated group did not alleviate to ascertain level of cure, continuous 14 days oral administration of aqueous CN extract at 500 (CN500) and 1000mg/kg BW (CN1000) was able to moderately ameliorated the neuroinflammation activity in a similar manner as the positive drug, dextromethorphan hydrobromide (DXM) at 5mg/kg BW. A consistent result has been observed for serum by both analytical platforms of liquid chromatography-mass (LC-MS) and NMR, the physiological sickness behavior and ¹H NMR brain tissue of the neuroinflammed male rats.

The alteration of lipid metabolism; (lysophosphatidic acid (LPA) and 5diphosphomevalonic acid) in sera of multiplatform model, and the changes of metabolites in the brain tissue namely, lactate, pyruvate, phosphorylcholine, glutamine, and α -ketoglutarate in CN500 and DXM exhibited an ameliorating effect when compared to the controlled neuroinflammed rats. CN treatments also significantly reduced IL-1 β , a pro-inflammatory cytokine better than DXM as proven in the quantification of cytokinesby microarray analysis. The physiological sickness behavior such as anxiety, exploration and reduction of locomotion also improved by CN treatments as visualized in the principal component analysis (PCA) model. Hence, herein a comprehensive view of the CN effects in neuroinflammation caused by LPS was successfully profiled, correlated and deciphered between central neuroinflammation, systemic metabolic and physiological disturbance which has potential for future ethnopharmacological and/or nutraceutical studies. Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

VARIASI METABOLIK MELALUI EVALUASI BIOLOGI EKSTRAK DAUN Clinacanthus nutans (Burm. f.) Lindau PADA TIKUS NEUROINFLAMASI TERARUH LIPOPOLISAKARIDA

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Neuroinflamasi adalah tindak balas yang kompleks pada mana-mana bahagian otak yang mengalami kecederaan, serta boleh menyebabkan pengaktifan sel glial, pelepasan pengantara keradangan seperti sitokin dan kimokin, serta reaktifikasi oksigen dan nitrogen spesis, yang merupakan salah satu ciri patologi bagi gangguan neurologi. Oleh itu, kawalan yang berkesan mencegah keradangan saraf adalah penting untuk mencegah penyakit berkaitan. Dalam kajian ini, daun matang Clinacanthus nutans (Burm. f.) Lindau (CN) yang berumur 9 minggu telah diekstrak dengan air dan dua ekstrak ethanolik (50% dan 100%).untuk mengkaii profil juzuk fitokimia mereka menggunakan kaedah resonans magnetik (¹H NMR) yang berasaskan kaedah metabolomik. Nilai tertinggi penghalangan nitrik oksida (NO), pada model neuroinflamasi menggunakan asosiasi titisan (in vitro) sel BV2 yang diaruh dengan lipopolisakarida (LPS), telah diperhatikan pada ekstrak akueus dengan nilai IC_{50} 336.2 ± 4.7 µg/ml. Korelasi antara aktiviti penghalangan NO dengan konstituen CN menggunakan analisa separa persegi (PLS) telah mengakibatkan pengenalpastian metabolit-metabolit yang berkemungkinan bertanggungjawab bagi aktiviti tersebut adalah schaftosida, asetat, propionat, alanin dan Clinacosida C.

Kajian model *in vivo* neuroinflamasi menggunakan tikus Sprague Dawley yang diaruh daripada LPS juga telah dilakukan melalui pendekatan metabolomik. Dapatan multivariat data analisis (MVDA) telah berjaya menyerlahkan beberapa persamaan dan kelainan pada kepekatan metabolit-metabolit tikus yang di aruh LPS (LPS+air) dan tikus LPS yang diaruh serta diberikan CN ekstrak. Walaupun hasil rawatan dengan dos-dos CN tidak memadai sehingga ke tahap penyembuhan, pengambilan secara oral ekstrak akues CN secara berterusan selama 14 hari pada dos 500 (CN500) dan 1000mg/kg berat badan tikus (BB) (CN1000) didapati dapat memperbaiki aktiviti anti-keradangan neuro secara sederhana seperti yang diamati dalam cara ubat kawalan, dextromethorphan hydrobromide pada dos 5mg/kg BB. Hasil keputusan yang konsisten juga telah

diperhatikan dalam serum pada kedua-dua platform analitikal; cecair kromatografi jisim (LC-MS) dan NMR, fisiologi tingkah laku berpenyakit dan ¹H NMR neuroinflammasi tisu otak tikus jantan.

Perubahan metabolism lemak; (lisofosphatidik asid (LPA) dan 5-difoshomevalonik asid) dalam model multiplatform serum, dan perubahan metabolit tisu otak seperti laktik, pyruvida, phosphorylkolin, glutamin, dan alpha-ketoglutarat dalam CN500 dan DXM mempamerkan penambahbaikan apabila dibandingkan dengan tikus rawatan kawalan neuroinflamasi aruhan LPS. Rawatan CN juga, secara ketara telah berjaya mengurangkan ekspresi IL-1*6*, iaitu pro-keradangan sitokin secara lebih baik daripada DXM seperti yang dibuktikan dalam kajian analisa tatasusun kuantitijasad inflamasi tikus. Fisiologi penyakit berkaitan radang otak seperti kebimbangan, penerokaan, serta pengurangan aktiviti lokomotor daripada rawatan CN juga bertambah baik seperti dalam gambaran model analisis komponen utama (PCA). Oleh itu, satu pemerhatian komprehensif bagi kajian kesan CN dalam mengatasi keradangan neuro yang disebabkan oleh LPS telah berjaya diprofil, dikorelasi serta dihuraikan antara radang saraf pusat, gangguan metabolik pada fisiologi, serta keseluruhan sistemik saraf yang mana hasilan ini berpotensi dirujuk bagi kajian etnofarmasi dan/atau pengajian nutraseutikal.

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Who remember Allah while standing, sitting, and lying down on their sides, and contemplate the creation of the heavens and the earth (with the thought) "Our Lord! Not for nothing have You created (all) this. Glory to You! Give us salvation from the suffering of the Fire.

(The Quran, 3:191)

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

%	Percentage
α	Alpha
β	Beta
γ	Gamma
δ	Delta for chemical shift in ppm
°C	Degree centrigrade
μg	Micro gram
μL	Micro litre
μm	Micro meter
13C	Carbon-13
¹ H	Proton
ACUC	Animal Care and Use Committee
ANOVA	Analysis of variance
BBB	Blood-brain barrier
BSA	Bovine serum albumin
BW	Body weight of rat
CN	<i>Clinacanthus nutans</i>
CPMG	Carr-Purcell-Meiboom-Gill
CV	Coefficient of varience
d	Doublet
dd	Doublet of doublets
DXM	Dextromethorphan hydrobromide
ESI-	Electrospray ionization for negative ion
ESI+	Electrospray ionization for positive ion
FDR	False discovery rate
GABA	Gamma-aminobutyric acid
HCA	Hierarchical cluster analysis
HMBC	Heteronuclear Multiple Bond Correlation
HSQC	Heteronuclear Single-Quantum Coherence
ICV	Intracerebroventicular
IFN	Interferon
IL	Interleukin
IMPaLa	Integrated Molecular Pathway Level Analysis
J	Coupling Constant in Hz
LCMS	Liquid chromatography Mass Spectrometry
LPS	Lipopolysaccharides
m	Multiplet
MCP	Monocyte chemoattractant protein
MetPA	Metabolic pathways analysis
MHz	Megahertz
NKEA	National key Economic Area
NMR	Nuclear magnetic resonance
NO	Nitrite oxide
OPLS	Orthogonal partial least square
OPLS-DA	Orthogonal partial least square discriminant analysis
PC	Principal components
PCA	Principal component analysis

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PLS PLS-DA ROS SD SD SEM SIMCA t TCA Th TLR TNF	Partial least square Partial least square- discriminant analysis Reactive oxygen species Singlet Sprague dawley Standard deviation Standard error of the mean Soft independent modeling of class analogy Triplet Tricarboxylic acid T-helper Toll-like receptors Tumor necrosis factor
TNF	
TSP	Trimethylsilanepropionic Acid Sodium Salt
VIP	Variable importance of projection

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CHAPTER 1

INTRODUCTION

1.1 Research background

Neuroinflammation is a key player for severe neurodegenerative diseases. An elevation of neurodegenerative disease that effect mental health brings hindrance to life quality (Beard et al., 2015). A recent report in Malaysian Burden of Disease and Injury Study: 2009 -2014 by the Ministry of Health (MOH), neurological diseases are among the top 12 lead diseases causing morbidity and mortality in Malaysia (MOH, 2017). Hence, there is an alarming interest in the exploration of neuroprotection through the pharmacological application of anti-inflammatory agents either of the synthetic drugs or natural products since both are well documented (Shal et al., 2018).

Herbal medicines or natural products for healthcare have gained popularity due to their safety based on their prolonged usage since the beginning of human civilization. Unfortunately, many of the herbal products for these past decades have been reported with significant side effects. The intentional adulteration and accidental contamination were reported as the primary reasons (Mosihuzzaman, 2012), hence highlights the importance of quality, safety, efficacy, consistency and availability (QSECA) of the claimed herbal medicines.

The assessment of the overall procedures by a conventional method of isolation guided is difficult due to the complexity to ensure the QSECA for the natural products from the original crude extract (WHO, 2008). The quality and safety of an herbal product are practically the outcomes of the standardization of the agronomic steps and material processes (Tripathi et al., 2014). The efficacy of a plant often resulted from the cumulative interaction of a large number of phytoconstituents (Tripathi et al., 2014). Although there were extensive studies on this plant anti-inflammatory properties, to understand the responsible compounds belong to a complex matrix of the plant is not easy.

The metabolomics approach can overcome this problem since it is based on a holistic measurement which possible to relate a collective compound in the whole sample system of a complex unfractionated matrix of crude extract (Verpoorte, 2009) to an activity such as anti-inflammation. The advancement in science and technology of a holistic approach by using modern tools in profiling and fingerprinting the chemical profile of systemic biology of plants and animals has catalyzed and eased the deciphering of the bioactive complex compounds matrix. Metabolomics approach offers a platform to realistically correlate the responsible metabolites to the various components of the biological response, which might be directly linked to the pathological events in the biological system. Hence, a holistic approach enables the identification of multiple compounds from a single extract and their interaction either synergistically or antagonistically in a biological *in vitro*

or *in vivo* system. Also, this approach has the ability to predict the bioactivity of a new set of extracts based on a developed, validated regression model (VRM) of bioactivity-compounds data, which is vital for quality control of a herb preparation. Consequently, the VRM can save time and budget by skipping the assay step of a new set of an extract (Yuliana et al., 2011).

Inflammation is a medical phrase that corresponds to the protective host feedback against infectious or noxious stimuli, where it involves host cells, blood vessel, and protein (Markiewski and Lambris, 2007). One of the primary aims is for defense via the activation of the immune cells to the area by destroying the pathogens and trigger the repair process. Neuroinflammation is a combination of two words, "neuro," which represents brain cells and "inflammation". There are a variety of cues that induce neuroinflammation, such as infection, traumatic injury, toxic metabolites, and/or autoimmunity (Gendelman, 2002). Usually, inflammation is under controlled and self-limited. However, an acute brain injury or infection plunges to be a complex inflammatory response when it involves the activation of microglia, which are the main resident innate immune cells in the central nervous system (CNS). In response to infection, microglia activate the production of cytokines, chemokines, and antibodies, which causes stimulation of other cell types such as astrocytes and T-lymphocytes (Dendrou et al., 2016). Since microglia is a critical player in the immune response of the central nervous system (CNS), it is a suitable choice to be examined for neuroinflammation. Prolong activation of all these are sources of multiple neurotoxic factors, including tumor necrosis factor- α , nitric oxide (NO), interleukin-1 β , and reactive oxygen species (ROS), which leads to progressive neuronal death or chronic neuroinflammation (Lull and Block, 2010). The imbalance of free radicals of ROS and nitrogen species, which is known as oxidative stress, leads to an outset of anti-inflammation and antioxidant exploration to control neuron activation as a promising therapeutic target to combat neuron-based diseases (Arulselvan et al., 2016).

The therapeutic potential of *Clinacanthus nutans* (Burm. f.) Lindau (CN), locally known as Belalai Gajah, is well documented. The most used part of the plant was the leaf, and it was reported that the leaf samples were found to possess more phytochemicals that exhibited antioxidant and a-glucosidase activities when compared to the stem (Khoo et al., 2015). Various other reports credited the CN leaf extracts to have antioxidant, anti-viral, anti-inflammatory and anti-cancer activities (Yong et al., 2013; Alam et al., 2016). To date, the standardization of CN leaf based on the quantification of certain marker compounds such as orientin, isoorientin, vitexin, isovitexin or shaftoside (Sarega et al., 2016) and comprehensive phytochemical analysis and the biological activity of inflammation have been recently conducted (Khoo et al., 2018a). Scientific reports on the pharmacological activities of the selected markers have also been well studied (Khoo et al., 2018d; Le et al., 2017). However, to our best knowledge, no report has highlighted the neuroprotective or anti-neuroinflammatory effects of CN leaf extract. Due to CN potentials in triggering the neuromodulation in the liver, heart, and kidney (Lau et al., 2014), its anti-neuroinflammatory activities are worthwhile to be investigated.

A holistic evaluation of rational, evidence-based CN herbal treatment enables a deeper understanding of the association of metabolic signature with its biological events. It is hypothesized that CN extract will show anti-neuroinflammatory effects in the neuron-BV2 cultured neuroinflammation in *in vitro* model, which in turn will be related to the *in vivo* rat model. This will lead to the discovery and development of effective phytomedical intervention and responses to the immunologic events, hence ensuring better constituents' knowledge and efficacy of herbs used in ethno-studies, nutraceutical, and functional food industries.

1.2 Aims and objectives

This study aimed to identify the possible neuroinflammation and neuroprotective biomarkers in the rat model and associate them with the possible immunologic events via NMR and LCM-MS. The metabolomic tools associated with multivariate analysis were employed to detect and discriminate the modulatory effects of CN on LPS neurotoxicity.

The aims were achieved through the following set of objectives:

- 1. To establish the metabolic fingerprint of aqueous, 50% ethanol and 100% ethanol of dried CN leaves extracts and to correlate their NO inhibition activity with the bioactive compounds via BV2 cells and rat model.
- 2. To identify the neuroinflammatory possible biomarker and the potential of underlying metabolic pathways involved in LPS-induced neuroinflammatory rats.
- 3. To elucidate the overall mechanism between the periphery and central nervous system, while integrating them with physical behavior and cytokine immunological responses.

1.3 Outline of the thesis

This thesis is presented in seven chapters:

Chapter 1 provides a general insight into the research background and aims of the study. The definition of a holistic approach, metabolomic, neuroinflammation, and CN brief background.

Chapter 2 focusses on the comprehensive literature review of the reported data related to the research.

Chapter 3 has been published in the *Records of Natural Products* under the title of "¹H NMR-based metabolomics of *Clinacanthus nutans* leaves extracts in correlation with their anti-neuroinflammation towards LPS-induced BV2 cells". This chapter discusses on the comparative metabolites evaluation of CN extracts from different solvent extractions by application of nuclear magnetic resonance (NMR) spectroscopy and chemometric methods related to bioassay-guided of BV2 cell toxicity and NO inhibition assays.

Chapter 4 has been published in the *Avicenna Journal of Phytomedicine* under the title of "Effects of *Clinacanthus nutans* leaf extract on lipopolysaccharide-induced neuroinflammation in rats: A behavioral and ¹H NMR-based metabolomics study". This chapter emphasizes on the metabolic profile of sera with LPS and CN intervention and their impact on the behavioral studies of physiological interruption of locomotor, anxiety and exploration.

In Chapter 5, evidence of the ¹H NMR brain tissue in the LPS-induced neuroinflammatory rat model showed that metabolomic analysis able to elucidate both metabolites and the genetic perturbation response via cytokines expression integrated OPLS model.

Chapter 6 presents the diverse biomarkers of the LPS-induced neuroinflammatory rat model with CN intervention using the integration application of LC-MS, a more sensitive analytical platform with NMR data. The results highlighted the identified markers from both platforms.

Finally, Chapter 7 summaries the thesis by connecting a network between metabolic profiles, physical behavior, and cytokines expression activation, where it recapitulates general discussion and ends in future applications and recommendations of the results presented.

REFERENCES

- Abbott, N.J, Patabendige, A.A.K., Dolman, D.E.M., Yusof, S.R., Begley, D.J. (2010). Structure and function of the blood-brain barrier. *Neurobiology of Disease*, 37(1), 13–25.
- Abbott, N.J., Ronnback, L. and Hansson, E. (2006) Astrocyte-endothelial interactions at the blood- brain barrier. *Nature Reviews in Neuroscience*. *7*, 41-53.
- Abdul Rahim, M.H., Zakaria, Z.A., Mohd Sani, M.H. et al., (2016). Methanolic extract of *Clinacanthus nutans* exerts antinociceptive activity via the opioid/nitric oxide-mediated, but cGMPindependent, pathways. *Evidence-Based Complementary and Alternative Medicine*. 2016(1494981), pp. 1–11.
- Abdullah, J.M, Hussain, A.M, Tharakan, J., Abdullah, M.R., Saad, R. Kamari., Z. et al. (2006). National response to neurological diseases in Malaysia: Planning for the future. *Southeast Asian Journal of Tropical Medicine And Public Health*, *37*(4), 796-805.
- Abraham, J. and Johnson, R.W. (2009). Consuming a diet supplemented with resveratrol reduced infection-related neuroinflammation and deficits in working memory in aged mice. *Rejuvenation Res.* 12(6):445-53.
- Aderem, A. and Ulevitch, R.J. (2000). Toll-like receptors in the induction of the innate immune response. *Nature*. 406(6797), 782-787
- Ahmad Azam, A., Ismail, I.S., Shaikh, M.F., Shaari, K., Abas, F. (2019). Effects of *Clinacanthus nutans* leaf extract on lipopolysaccharide -induced neuroinflammation in rats: a behavioral and ¹H NMR-based metabolomics study. *Avicenna Journal of Phytomedicine*. 9(2),164-186.
- Ahmad Azam, A., Pariyani, R., Ismail, I.S., Ismail, A., Khatib, A., Abas, F., Shaari, K. (2017). Urinary metabolomics study on the protective role of *Orthosiphon stamineus* in Streptozotocin induced diabetes mellitus in rats via ¹H NMR spectroscopy. *BMC Complementary and Alternative Medicine*, 17(1) 278– 291.
- Ailiah, M.R. (2011). Rawatan Alternatif moden bagi penyakit kanser. [Online] Available from: http://b17nitrilosides.blogspot.com/2011/12/sabah-snakegrass-extract.html?m=0 [Accessed on Dec 2011]
- Akira, S. and Takeda, K. (2004). Toll-like receptor signaling. *Nature Reviews Immunology*, 4, 499 -511
- Alafuzoff, I, Overmyer, M., Helisalmi, S, Soininen, H. (2000). Lower counts of astroglia and activated microglia in patients with Alzheimer's disease with regular use of non-steroidal anti-inflammatory drugs. *Journal of Alzheimer's Disease*. 2(1), 37–46.

- Alam, A., Ferdosh, S., Ghafoor, K., Hakim, A., Juraimi, A., Khatib, A., Sarker, Z. (2016). *Clinacanthus nutans*: A review of the medicinal uses, pharmacology and phytochemistry. *Asian Pacific Journal of Tropical Medicine*, 9(4), 402-409.
- Alam, M. A., Zaidul, I. S. M., Ghafoor K. et al., (2017). Identification of bioactive compounds with GC–Q-TOF–MS in the extracts from *Clinacanthus nutans* using subcritical carbon dioxide extraction. *Separation Science and Technology (Philadelphia)*. 52(5), pp. 852–863.
- Alonso, A., Marsal, S., Julia, A. (2015). Analytical methods in untargeted metabolomics: State of the art in 2015. *Frontiers in Bioengineering and Biotechnology*, 3, 23.
- Ambriz-Perez, D.L., Leyva-Lopez, N., Gutierrez-Grijalva, E.P., Heredia, J.B. (2016). Phenolic compounds: natural alternative in inflammation treatment. A review. Cogent Food and Agriculture, 2(1), 1131412.
- Ameer, F., Munir, R., Zaidi, N. (2019). Lipid metabolism in Encyclopedia of Cancer. *References Module in Biomedical Sciences*. 3, 369-373
- Andrea, P. and Vanderbroek, I. (2007). The ethnobiology and ethnopharmacy of migrations. *New York: Berghahn Books*, p. 112.
- Antharavally, B., Mallia, K., Rangaraj, P., Haney, P., Bell, P. (2009). Quantitation of proteins using a dye-metal-based colorimetric protein assay. *Analytical Biochemistry*. 385(2), 342-345.
- Arakaki, A., Skolnick, J. and McDonald, J. (2008). Marker metabolites can be therapeutic targets as well. *Nature*, 456(7221), pp.443-443.
- Arullappan, S., Rajamanickam, P., Thevar, N. and. Kodimani, C.C. (2014). In vitro screening of cytotoxic, antimicrobial and antioxidant activities of *Clinacanthus nutans* (Acanthaceae) leaf extracts. *Tropical Journal of Pharmaceutical Research*, vol. 13(9), pp. 1455–1461.
- Arulselvan, P., Fard, M. T., Tan, W. S., et al. (2016). Role of antioxidants and natural products in inflammation. Oxidative Medicine and Cellular Longevity, 2016, 5276130.
- Ashraf, T. (2015) *In vitro* and *in vivo* regulation of pro-inflammatory cytokines and drug efflux transporters by signal transduction pathways in glial cells: implications in HIV-1 neuropathogenesis and its treatment. University of Toronto, Canada.
- Auslander, N., Yizhak, K., Weinstock, A., Budhu, A., Tang, W., Wang, X.W. et al. (2016). A joint analysis of transcriptomic and metabolomic data uncovers enhanced enzyme-metabolite coupling in breast cancer. *Scientific Reports*. 6(1).

- Bahar E, Kim JY, Yoon H. (2017). Quercetin attenuates manganese-induced neuroinflammation by alleviating oxidative stress through regulation of apoptosis, iNOS/NF-κB and HO-1/Nrf2 pathways. *International Journal of Molecular Sciences.* 18(9), E1989
- Bain, L., Keren, N., Norris, S. (2018). Biomarkers of neuroinflammation: proceedings of a workshop. *The National Academies Press*
- Balashova, E.E., Maslov, D.L., and Lokhov, P.G. (2018). A metabolomics approach to pharmacotherapy personalization. *Journal of Personalized Medicine*, 8(3), 28. doi: 10.3390/jpm8030028
- Bassi, G.S., Broiz, A.C., Gomes, M.Z., Brand, M.L. (2009). Evidence for mediation of nociception by injection of the NK-3 receptor agonist, senktide, into the dorsal periaqueductal gray of rats. *Psychopharmacology Berl*. 204(1), 13– 24.
- Bassi, G.S., Kanashiro, A., Santin, F.M., de Souza, G.E.P., Nobre, M.J., Coimbra, N.C. (2012). Lipopolysaccharide-induced sickness behaviour evaluated in different models of anxiety and innate fear in rats. *Basic and Clinical Pharmacology and Toxicology*. 110(4), 359-369
- Bayer, A., Yu, A., Adeegbe, D., Malek, T. (2005). Essential role for interleukin-2 for CD4+CD25+T regulatory cell development during the neonatal period. *The Journal of Experimental Medicine*. 201(5), 769-777.
- Bauer, B, Hartz, A.M. and Miller, D.S. (2007) Tumor necrosis factor alpha and endothelin-1 increase p-glycoprotein expression and transport activity at the blood-brain barrier. *Molecular Pharmacology*. **71**, 667-675.
- Beckonert, O., Keun, H., Ebbels, T., Bundy, J., Holmes, E., Lindon, J. et al. (2007). Metabolic profiling, metabolomic and metabonomic procedures for NMR spectroscopy of urine, plasma, serum and tissue extracts. *Nature Protocols*. 2(11), 2692-2703.
- Beg, S., Hasan, H., Hussain, M., Swain, S., Barkat, M. (2011). Systematic review of herbals as potential anti-inflammatory agents: Recent advances, current clinical status and future perspectives. *Pharmacognosy Reviews*, *5*(10), 120. doi: 10.4103/0973-7847.91102
- Begum, A.N., Jones, M.R., Lim, G.P., Morihara, T., Kim, P., Heath, D.D. et al. (2008). Curcumin structure-function, bioavailability, and efficacy in models of neuroinflammation and Alzheimer's disease. *Journal of Pharmacology* and Experimental Therapy. 326(1), 196-208.
- Bélanger, M., Allaman, I., Magistretti, P.J. (2011). Brain energy metabolism: focus on astrocyte-neuron metabolic cooperation. *Cell Metabolism*, 14(16), 724– 738.

- Benaki, D. and Mikros, E. (2018). NMR-based metabolic profiling procedures for biofluids and cell and tissue extracts. *Methods in Molecular Biology*, 1738, 117-131.
- Beutler, B. and Cerami, A. (1988). The common mediator of shock, cachexia, and tumor necrosis. *Advances in Immunology*. 42, 213-231.
- Bixel, M., Shimomura, Y., Hutson, S., Hamprecht, B. (2001). Distribution of key enzymes of branched-chain amino acid metabolism in glial and neuronal cells in culture. *Journal of Histochemistry and Cytochemistry*, 49(3), 407– 418.
- Block, M. (2008). NADPH oxidase as a therapeutic target in Alzheimer's disease. *BMC Neuroscience*. 9, S8.
- Bollard, M.E., Stanley, E.G., Lindon, J.C, Nicholson, J.K. et al. (2005). NMRbased metabonomic approaches for evaluating physiological influences on biofluid composition NMR Biomedicine. 18(3),143-164.
- Boonkerd, G. (1967). The Chemical Constituents of Clinacanthus burmanii, Chulalongkorn University.
- Bouzier, K., Thiaudiere, E., Biran, M., Rouland, R., Canioni, P., Merle, M. (2000). The metabolism of [3-(13) C] lactate in the rat brain is specific of a pyruvate carboxylase-deprived compartment. *Journal of Neurochemistry*, 75(2), 480–486.
- Bradford, J., Shin, J.Y., Roberts, M., Wang, C.E., Li, X.J., Li, S. (2009). Expression of mutant huntingtin in mouse brain astrocytes causes age-dependent neurological symptoms. *Proceedings of the National Academy of Sciences of the United States of America*, 106 (52), 22480–22485.
- Breitner, J.C., Welsh, K.A., Helms, M.J. et al. (1995). Delayed onset of Alzheimer's disease with nonsteroidal anti-Inflammatory and histamine H2 blocking drugs. *Neurobiology of Aging*.16(4), 523–530.
- Broadhurst, D.I and Kell, D.B. (2006). Statistical strategies for avoiding false discoveries in metabolomics and related experiments, *Metabolomics*. 2(4),171–196.
- Brown, M.V., McDunn, J.E., Gunst, P.R., Smith, E.M., Milburn, M.V., Troyer, D.A., Lawton, K.A. (2012). Cancer detection and biopsy classification using concurrent histopathological and metabolomic analysis of core biopsies. *Genome Medicine*.;4, 33.
- Burgess, K., Rankin, N., Weidt, S. (2014). Chapetr 10: Metabolomics. *Handbook* of *Pharmacogenomics and Stratified Medicine*. 181-205. Academic Press.Elsevier, USA.

- Bylesjo, M., Rantalainen, M., Cloarec, O., Nicholson, J.K., et al. (2006). OPLS discriminant analysis: combining the strengths of PLS-DA and SIMCA classification. *Journal of Chemometrics*. 20(8–10), 341–351.
- Calabrese, E. and Mattson, M.P. (2017). How does hormesis impact biology, toxicology, and medicine? *Npj Aging and Mechanisms of Diseases.* 3, 13.
- Carpenter, J.F., Manning, M.C., Randolph, T.W. (2002). Protein stability and storage. *Current Protocols in Protein Science*. doi:10.1002/0471140864.ps0406s27.
- Carson, M.J., Thrash, J.C. and Walter, B. (2006). The cellular response in neuroinflammation: the role of leukocytes, micrglia and astroccytes in neuronal death and survival. *Clinical Neuroscience Research*. 6(5), 237-245.
- Carter, A.J., Feeney, W.E., Marshall, H.H., Cowlishaw, G., Heinsohn, R. (2013). Animal personality: What are behavioural ecologists measuring? *Biology Reviews of the Cambridge Philosophical Society*. 88(2), 465e475.
- Cartmell, T., Ball, C., Bristow, A.F., Mitchell, D., Poole, S. (2003). Endogenous interleukin-10 is required for the defervescence of fever evoked by local lipopolysaccharide-induced and *Staphylococcus aureus*-induced inflammation in rats, *The Journal of Physiology*. 549(2), 653–64.
- Cavaillon, J.M. (2001). Pro- versus anti-inflammatory cytokines: myth or reality. *Cellular and Molecular Biology (Noisy-Le-Grand)*. 47(4), 695-702.
- Chang, L., Munsaka, S.M., Kraft-Terry, S., Ernst, T. (2013). Magnetic resonance spectroscopy to assess neuroinflammation and neuropathic pain. *Journal of Neuroimmune Pharmacology*, 8(3), 576–593.
- Chang, Y., Jia, X., Sun, X., Xu, S., Wu, Y., Zhang, L. et al. (2015). APRIL promotes proliferation, secretion and invasion of fibroblast-like synoviocyte from rats with adjuvant induced arthritis. *Molecular Immunology*. 64(1), 90-98.
- Chavalittumrong, P., Attawish, A., Rugsamon, P., and Chuntapet, P. (1995). Toxicological Study of *Clinacanthus nutans* (Burm. f.) Lindau. *Bulletin of the Department of Medical Science*. 37 (4): 323-337.
- Chelyn, J. L., Omar, M.H., Mohd Yousof, N.S.A., Ranggasamy, R., Wasiman, M.I. and Ismail, Z. (2014). Analysis of flavone *C*-glycosides in the leaves of *Clinacanthus nutans* (Burm. f.) Lindau by HPTLC and HPLC-UV/DAD, *The Scientific World Journal*. 2014, (724267), pp. 1–6.
- Chen, J., Buchanan, J.B., Sparkman, N.L., Godbout, J.P., Freund, G.G., Johnson, R.W. (2008). Neuroinflammation and disruption in working memory in aged mice after acute stimulation of the peripheral innate immune system. *Brain, Behavior and Immunity*. 22(3), 301-311.

- Chen, L., Zhang, B., Shan, S., Zhao, X. (2016). Neuroprotective effects of vitexin against isoflurane-induced neurotoxicity by targeting the TRPV1 and NR2B signaling pathways. *Molecular Medicine Reports*, 14(6), 5607–5613.
- Chen, X., Qi, X., & Duan, L. X. (2015). Overview. In X. Qi, X. Chen, & Y. Wang (Eds.), *Plant Metabolomics: Methods and Applications* (pp. 1-24). Dordrecht: Springer Netherlands.
- Cheng, C., Huang, C., Ma, T., Bian, E., He, Y., Zhang, L., Li, J. (2014). SOCS1 hypermethylation mediated by DNMT1 is associated with lipopolysaccharide-induced inflammatory cytokines in macrophages. *Toxicology Letters*. 225(3), 488-497.
- Cheong, B.E., Ho, S.Y. and Dickens Wong, V.F. (2013). Chemical profiling of Sabah snake grass, *Clinacanthus nutans*, in *Proceedings of the 11th Seminar on Science & Technology*.
- Cherdchu, C., Poopyruchpong, N., Adchariyasucha, R. and Ratanabanangkoon, K. (1977). The absence of antagonism between extracts of *Clinacanthus nutans* Burm. and *Naja naja siamensis* venom. *Southeast Asian Journal of TropicalMedicine and Public Health.* 8(2), 249–254.
- Chiu, F. and Lin, J. (2008). Tomatidine inhibits iNOS and COX-2 through suppression of NF-κB and JNK pathways in LPS-stimulated mouse macrophages. *FEBS Letters*, *582*(16), 2407-2412.
- Chung, K.F. (2003). Cytokines in chronic obstructive pulmonary disease. *European Respiratory Journal*. 50-59.
- Clapp, W.D., Thorne, P.S., Frees, K.L., Zhang, X., Lux, C.R., (1993). The effects of inhalation of grain dust extract and endotoxin on upper and lower airways. *Chest.* 104(3), 825–830
- Clifton, L.A., Skoda, M.W.A., Daulton, E.L., Hughes, A.V., Le Brun, A.P., Lakey, J.H., Holt, S.A. (2013). Asymmetric phospholipid: lipopolysaccharide bilayers; a Gram-negative bacterial outer membrane mimic. *Journal of the Royal Society, Interface*. 10(89), 20130810.
- Conti, B., Tabarean, T., Bartfai. (2004). Cytokines and fever. *Frontiers in Bioscience*. doi: 10.2741/1341
- Cordell, G., and Colvard, M. (2012). Natural products and traditional medicine: Turning on a paradigm. *Journal of Natural Products*, *75*(3), 514-525.
- Crandall, E.A. and Fernstrom, J.D. (1983). Effect of experimental diabetes on the levels of aromatic and branched-chain amino acids in rat blood and brain. *Diabetes*, 32(3), 222–230.
- Crawley, J. and Goodwin, F.K. (1980). Preliminary report of a simple animal behavior model for the anxiolytic effects of benzodiazepines. *Pharmacology, Biochemistry and Behavior.* 13(2), 167–70.

- Cruz, A.P., Frei, F., Graeff, F.G. (1994). Ethopharmacological analysis of rat behavior on the elevated plus-maze. *Pharmacology, Biochemistry and Behavior*. 49(1), 171–176.
- Čuperlović-Culf, M., Barnett, D., Culf, A., and Chute, I. (2010). Cell culture metabolomics: applications and future directions. *Drug Discovery Today*, 15(15-16), 610-621. doi: 10.1016/j.drudis.2010.06.012
- Dampawan, P., Huntrakul, C. and Reutrakul, V. (1977). Constituents of *Clinacanthus nutans* and the crystal structure of lup-20(29)-ene-3-one. *Journal of the Science Society of Thailand*. 3(1), 14–26.
- Dauer, W. and Przedborski, S. (2003). Parkinson's disease: mechanisms and models. *Neuron*. 39(6), 889-909.
- Davis, B. M., Salinas-Navarro, M., Cordeiro, M. F., Moons, L., & De Groef, L. (2017). Characterizing microglia activation: a spatial statistics approach to maximize information extraction. Scientific reports, 7(1), 1576. doi:10.1038/s41598-017-01747-8
- De Luca, C. and Olefsky, J.M.I. (2008). Inflammation and insulin resistance. *FEBS Letters*, 582(1), 97–105.
- Deakin, J.F.W. and Graeff, F.G. (1991). 5-HT and mechanisms of defence. *Journal* of Psychopharmacology (Oxford, England). 5, 305–315.
- Dear, G.J., Ayrton, J., Plumb, R., Sweatman, B.C. et al. (1998) A rapid and efficient approach to metabolite identification using nuclear magnetic resonance spectroscopy, liquid chromatography/mass spectrometry and liquid chromatography/nuclear magnetic resonance spectroscopy/sequential mass spectrometry. *Rapid Communicationn Mass Spectrometry*.12(24), 2023–2030.
- Dendrou, C., McVean, G., and Fugger, L. (2016). Neuroinflammation using big data to inform clinical practice. *Nature Reviews Neurology*, 12(12), 685-698.
- Deshmane, S., Kremlev, S., Amini, S., Sawaya, B. (2009). Monocyte chemoattractant protein-1 (MCP-1): an overview. *Journal of Interferon and Cytokine Research*. 29(6), 313-26.
- Devi, L.A. (2000). G-protein-coupled receptor dimers in the lime light. *Trends in Pharmacological Sciences*. 21(9), 324–6.
- Dillon, W.R. and Goldstein, M. (1984). Multivariate analysis, method and application. xiii+587. New York. Wiley.
- Dinarello, C.A. (1991). The proinflammatory cytokines interleukin-1 and tumor necrosis factor and treatment of the septic shock syndrome. *Journal of Infectious Diseases*. 163(6), 1177-1184.

- Direkbusarakom, S., Ruangpan, L., Ezura, Y. and Yoshimizu, M. (1998). Protective efficacy of *Clinacanthus nutans* on yellow-head disease in black tiger shrimp (*Penaeus monodon*). *Fish Pathology*. 33(4), 401–404.
- DiSabato D, Quan N, Godbout J. (2016). Neuroinflammation: the devil is in the details. *Journal of Neurochemistry*. 139(2),136-153. doi: 10.1111/jnc.13607
- Drake, T.A., Cheng, J., Chang, A., Taylor, F.B. Jr. (1993). Expression of tissue factor, thrombomodulin, and E-selectin in baboons with lethal *Escherichia coli* sepsis. *The American. Journal of. Pathology.* 142(5), 1458–70
- Dubuis, S., Ortmayr, K., & Zampieri, M. (2018). A framework for large-scale metabolome drug profiling links coenzyme A metabolism to the toxicity of anti-cancer drug dichloroacetate. *Communications Biology*, *1*(1). doi: 10.1038/s42003-018-0111-x
- Dumas, M., Maibaum, E., Teague, C., Ueshima, H., Zhou, B., Lindon, J., Nicholson, J., Stamler, J., Elliott, P., Chan, Q. and Holmes, E. (2006). Assessment of analytical reproducibility of ¹H NMR spectroscopy based metabonomics for large-scale epidemiological research: study. *Analytical Chemistry*, 78(7), pp.2199-2208.
- Dunn, W., Broadhurst, D., Begley, P., Zelena, E., Francis-McIntyre, S., Anderson, N. et al. (2011). Procedures for large-scale metabolic profiling of serum and plasma using gas chromatography and liquid chromatography coupled to mass spectrometry. *Nature Protocols*, 6(7), 1060-1083. doi: 10.1038/nprot.2011.335
- Dunn, W., Erban, A., Weber, R., Creek, D., Brown, M., Breitling, R. et al. (2012). Mass appeal: metabolite identification in mass spectrometry-focused untargeted metabolomics. *Metabolomics*, 9(S1), 44-66. doi: 10.1007/s11306-012-0434-4
- Dunn, W.B., Goodacre, R., Neyses, L., Mamas, M. (2011). Integration of metabolomics in heart disease and diabetes research: current achievements and future outlook. *Bioanalysis* 3(19), 2205-2222.
- Duren, W., Weymouth, T., Hull, T., Omenn, G., Athey, B., Burant, C., & Karnovsky,
 A. (2014). MetDisease—connecting metabolites to diseases via literature. *Bioinformatics*, 30(15), 2239-2241.
- Echeverria, V. and Zeitlin, R. (2012). Cotinine: a potential new therapeutic agent against Alzheimer's disease. *CNS Neuroscience and Therapeutics*, 18(7), 517–523.
- El-Ansary, A. and Al-Ayadhi L. (2014). GABAergic/glutamatergic imbalance relative to excessive neuroinflammation in autism spectrum disorders. *Journal of Neuroinflammation*, 19(11), 189-194.

- Emwas, A., Salek, R., Griffin, J. and Merzaban, J. (2013). NMR-based metabolomics in human disease diagnosis: applications, limitations, and recommendations. *Metabolomics*, 9(5), pp.1048-1072.
- Eriksson L, Johansson E, Kettaneh-Wold N, Trygg J, Wikstrom C, Wold S. (2006). *Multi- and Megavariate Data Analysis Part 1: Basic Principles and Applications*. Umea, Sweden: Umetrics, Inc.
- Eriksson, L., Byrne, T., Johansson, E., Trygg, J., Vikström, C. (2013). Multi- and Megavariate Data Analysis Basic Principles and Applications, Volume 1. MKS Umetrics AB.
- Eriksson, L., Trygg, J., Wold, S. (2008). CV-ANOVA for significance testing of PLS and OPLS® models. *Journal of Chemometrics*, 22(11-12), 594-600. doi: 10.1002/cem.1187
- Erridge, C., Bennett-Guerrero, E, Poxton, I.R. (2002). Structure and function of lipopolysaccharides. *Microbes and Infection*. 4(8), 837–851.
- Espinosa-Oliva, A.M., Palos, R.M. and Herrera A. (2013). Intracranial injection of LPS in rat as animal model of neuroinflammation. *Methods in Molecular Biology*. 1041, 295-305.
- Esquivel-Elizondo, S., Ilhan, Z., Garcia-Peña, E., & Krajmalnik-Brown, R. (2017). Insights into butyrate production in a controlled fermentation system via gene predictions. *Msystems*, 2(4). doi: 10.1128/msystems.00051-17
- Executive Summary of the Third Report of the National Cholesterol Education Program (NCEP). 2001. Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III). JAMA, 285: 2486–2497.
- Farsi, E., Abdul Majid, A.S., Abdul Majid, A.M.S. (2016). Clinacanthus nutans, Yesterday's Practice, and Future's Drug: A Comprehensive Review. American Journal of Phytomedicine and Clinical Therapeutics.4(4), 113-126.
- Fernández-Albert, F., Llorach, R., Andrés-Lacueva, C., and Perera, A. (2014). An R package to analyse LC/MS metabolomic data: MAIT (Metabolite Automatic Identification Toolkit). *Bioinformatics (Oxford, England)*, 30(13), 1937–1939. doi:10.1093/bioinformatics/btu136
- Fernstrom, J.D. (2005). Branched-chain amino acids and brain function, *Journal* of *Nutrition*, 135(6 suppl), 1539S–1546S.
- Ferrero-Miliani, L., Nielsen, O.H., Andersen, P.S., Girardin, S.E. (2007). Chronic inflammation: importance of NOD2 and NALP3 in interleukin-1β generation, *Clinical and Experimental Immunology*, 0(0).
- Fiehn, O. (2002) Metabolomics the link between genotypes and phenotypes. *Plant Molecular. Biology.* 48(1-2), 155–171.

- Fiehn, O., Robertson, D., Griffin, J., van der Werf, M., Nikolau, B., Morrison, N. et al. (2007). The metabolomics standard initiative (MSI). Metabolomics. 3(3), 175-178.
- Floral of China Editorial Committee. Flora of China, Vol. 19, (2011). Curcurbitaceae through Valerianaceae with Annonaceae and Berberidaceae. *Beijing: Science Press & Missouri Botanical Garden Press and St. Louis.* p. 1-884.
- Folch, J., Lees, M., Sloane Stanley, G.H. (1957). A simple method for the isolation and purification of total lipides from animal tissues. *The Journal of Biological Chemistry*. 226(1), 497-509.
- Fondi, M., and Liò, P. (2015). Multi -omics and metabolic modelling pipelines: Challenges and tools for systems microbiology. *Microbiological Research*, 171, 52-64. doi: 10.1016/j.micres.2015.01.003
- Fong, S.Y., Piva, T., Urban, S., Huynh, T. (2014). Genetic homogeneity of vegetatively propagated *Clinacanthus nutans* (Acanthaceae). *Journal of Medicinal Plant Research*. 8,903–914.
- Food and Drug Administration (FDA). (1985). Guideline for Inspection technical: Bacterial endotoxins/pyrogens. Retrieved January 12, 2018, from https://www.fda.gov/ICECI/Inspections/InspectionGuides/InspectionTechn icalGuides/ucm072918.htm
- Frede, S., Stockmann, C., Freitag, P., Fandrey, J. (2006). Bacterial lipopolysaccharide induces HIF-1 activation in human monocytes via p44/42 MAPK and NF-κB. *Biochemical Journal*. 396(3), 517-527.
- Fukushima, A. and Kusano, M (2013) Recent progress in the development of metabolome databases for plant systems biology. *Frontiers in Plant Science* 4, 73. (<u>https://doi.org/10.3389/fpls.2013.00073</u>)
- Fünfschilling, U., Supplie, L.M., Mahad, D., Boretius, S., Saab, A.S., Edgar, J et al. (2012). Glycolytic oligodendrocytes maintain myelin and long-term axonal integrity. *Nature*, 485: 517–521.
- Furukawa, S., Yang, L., Sameshima, H. (2014). Galantamine, an acetylcholinesterase inhibitor, reduces brain damage induced by hypoxiaischemia in newborn rats. *International. Journal of. Developmental. Neuroscience*. 37, 52–57. 10.1016/j.ijdevneu.2014.06.011
- Galanos, C., Roppel, J., Weckesser, J., Rietschel, E.T., Mayer, H. (1977). Biological activities of lipopolysaccharides and lipid A from *Rhodospirillaceae. Infection and Immunity.* 16(2), 407-412.
- Galic, M., Riazi, K., Pittman, Q. (2012). Cytokines and brain excitability. *Frontiers in Neuroendocrinology*. 33(1), 116-125.

- Gao, H., Jiang, J., Wilson, B., Zhang, W., Hong, J., and Liu, B. (2002). Microglial activation-mediated delayed and progressive degeneration of rat nigral dopaminergic neurons: relevance to Parkinson's disease. *Journal of Neurochemistry*. 81(6), 1285-1297.
- García-Campos, M., Espinal-Enríquez, J., and Hernández-Lemus, E. (2015). Pathway analysis: State of the Art. *Frontiers in Physiology*, 6. doi: 10.3389/fphys.2015.00383

Garden, G.A. (2002) Microglia in human immunodeficiency virus-associated neurodegeneration. *Glia* 40(2), 240-251.

Garden, G.A. and Moller, T. (2006) Microglia biology in health and disease. *Journal of Neuroimmune Pharmacology.* 1, 127-137.

- Garry, P., Ezra, M., Rowland, M., Westbrook, J., & Pattinson, K. (2015). The role of the nitric oxide pathway in brain injury and its treatment — From bench to bedside. *Experimental Neurology*, 263, 235-243. doi: 10.1016/j.expneurol.2014.10.017
- Gasior, M., Rogawski, M.A, Hartman, A.L. (2006). Neuroprotective and diseasemodifying effects of the ketogenic diet. *Behavioural Pharmacology*, 17(5-6), 431–439.
- Gavaghan, C.L., Wilson, I.D., Nicholson, J.K. (2002). Physiological variation in metabolic phenotyping and functional genomic studies: use of orthogonal signal correction and PLS-DA. *FEBS Letters*. 530(1–3),191–196.
- Gellért, L. and Varga, D. (2016). Locomotion activity measurement in an open field for mice. *eBio-protocol.* 6(13), 1857.
- Gendelman, H. (2002). Neural Immunity: Friend or Foe? *Journal of Neurovirology*, *8*(6), 474-479.
- Ghasemzadeh, A., Nasiri, A., Jaafar, H.Z., Baghdadi, A, Ahmad, I. (2014). Changes in phytochemical synthesis, chalcone synthase activity and pharmaceutical qualities of sabah snake grass (*Clinacanthus nutans* L.) in relation to plant age. *Molecules (Basel, Switzerland*). 19(11), 17632-48. doi:10.3390/molecules191117632.
- Giaume, C., Koulakoff A., Roux, L. et al. (2010). Astroglial networks: a step further in neuroglial and gliovascular interaction. *Nature Reviews in Neuroscience*. 11, 87-99.
- Glascock, J., Osman, E., Coady, T., Rose, F., Shababi, M., Lorson, C. (2011). Delivery of therapeutic agents through intracerebroventricular (ICV) and intravenous (IV) injection in mice. *Journal of Visualized Experiments: JoVe*, 56, pii2968
- Global-disease-burden health grove.com. 2017. Neurological Disorders in Malaysia- Statistics on neurological disorders effect and annual mortality

rates from 1990 to 2013. Available at: http://global-diseaseburden.healthgrove.com/l/54726/Neurological-Disorders-in-Malaysia. (Accessed 12 April 2017).

- Godbout, J.P., Moreau, M., Lestage, J., Chen, J., Sparkman, N.L., Connor, J.O. et al. (2007). Aging exacerbates depressive-like behavior in mice in response to activation of the peripheral innate immune system. *Neuropsychopharmacology*. 33(10), 2341-1351.
- Gordon, P., Moore, D., Miller, R., Florence, J., Verheijde, J., Doorish, C. et al. (2007). Efficacy of minocycline in patients with amyotrophic lateral sclerosis: a phase III randomised trial. *The Lancet Neurology*, 6(12), 1045-1053.
- Goucham, A.Y. and Nicolaïdis S. (1999). Feeding enhances extracellular lactate of local origin in the rostromedial hypothalamus but not in the cerebellum. *Brain Research*, 816(1), 84–91.
- Gould, T.D., Dao, D.T., and Kovacsics, C.E. (2009). Mood and anxiety related phenotypes in mice: The open field test. *Neuromethods*. 42, 1-21.
- Grace, S.C. and Hudson, D.A. (2016). Processing and visualization of metabolomics data using R, in: Metabolomics—Fundamentals and Applications, *InTech*, pp. 63–94.
- Graeber, M.B. and Streit, W.J. (2010). Microglia: biology and pathology. *Acta Neuropathologica*. 119(1), 89-105.
- Grapov, D., Wanichthanarak, K., and Fiehn, O. (2015). MetaMapR: pathway independent metabolomic network analysis incorporating unknowns: Fig. 1. *Bioinformatics*, *31*(16), 2757-2760. doi: 10.1093/bioinformatics/btv194
- Green, L.C., Wagner, D.A., Glogowski, J., Skipper, P.L., Wishnok, J.K., Tannenbaum, S.R. (1982). Analysis of nitrate, nitrite and [15N]nitrate in biological fluids. *Analytical Biochemistry*. 126(1): 131-138.
- Gu, H., Chen, H., Pan, Z., Jackson, A.U. et al. (2007). Monitoring diet effects via biofluids and their implications for metabolomics studies. *Analitical Chemistry*. 79(1), 89–97.
- Gu, H., Pan, Z., Xi, B., Hainline, B.E., Shanaiah, N. et al (2009). ¹H NMR metabolomics study of age profiling in children. *NMR Biomedicine*. 22(8),826–833.
- Guitton, Y., Tremblay-Franco, M., Le Corguillé, G., Martin, J., Pétéra, M., Roger-Mele, P. et al. (2017). Create, run, share, publish, and reference your LC–MS, FIA–MS, GC–MS, and NMR data analysis workflows with the Workflow4Metabolomics 3.0 Galaxy online infrastructure for metabolomics. *The International Journal of Biochemistry and Cell Biology*, 93, 89-101. doi: 10.1016/j.biocel.2017.07.002

- Guzmán, M., Blázquez, C. (2004). Ketone body synthesis in the brain: possible neuroprotective effects. *Prostaglandins Leukotrienes and Essential Fatty Acids*, 70(3), 287–292.
- Haetrakul, T., Dunbar, S. and Chansue, N. (2018). Antiviral activities of *Clinacanthus nutans* (Burm.f.) Lindau extract against Cyprinid herpesvirus 3 in koi (*Cyprinus carpio* koi). *Journal of Fish Diseases*, 41(4), pp.581-587.
- Hall, R.D. (2011). Plant metabolomics in a nutshell: Potential and future challenges. 1-24.
- Hamid, H.A., Yahya, I.H., Yusoff, M.M. and Zareen, S. (2016). Bioassay-guided isolation and antioxidant activity of sulfur containing compounds from *Clinacanthus nutans. Journal of the Chinese Chemical Society*. 63(12), pp. 1033–1037.
- Harris, R.Z., Jang, J.R. and Tsunoda, S. (2003). Dietary effects on drug metabolism and transport. *Clinical pharmacokinetics*. 42(13), 1071-1088.
- Hashim, N. H. N., Latip, J., & Khatib, A. (2016). Metabolite profiling of *Clinacanthus nutans* leaves extracts obtained from different drying methods by ¹H NMR-based metabolomics. In 2016 UKM FST Postgraduate Colloquium: Proceedings of the Universiti Kebangsaan Malaysia, Faculty of Science and Technology 2016 Postgraduate Colloquium (Vol. 1784). [030015] American Institute of Physics Inc., https://doi.org/10.1063/1.4966753
- Hashimoto, T., Hussien, R., Cho, H.S., Kaufer, D., Brooks, G.A. (2008). Evidence for the mitochondrial lactate oxidation complex in rat neurons: demonstration of an essential component of brain lactate shuttles. *PLoS One*, 3(8), e2915.
- Hasin, Y., Seldin, M., Lusis, A. (2017). Multi-omics approaches to disease. *Genome Biology*. 18(1).
- Hemmerle, A.M., Herman, J.P., Seroogy, K.B. (2012). Stress, depression and Parkinson's disease. *Experimental Neurology*, 233(1), 79–86.
- Henry, C.J., Huang, Y., Wynne, A., Hanke, M., Himler, J., Bailey, M.T. et al. (2008). Minocycline attenuates lipopolysaccharide (LPS)-induced neuroinflammation, sickness behavior, and anhedonia. *J Neuroinflammation*. 5, 1-14
- Herrera, A.J, Castaño, A., Venero, J.L., Cano, J., and Machado, A. (2000). The single intranigral injection of LPS as a new model for studying the selective effects of inflammatory reactions on dopaminergic system. *Neurobiology of Disease*. 7(4), 429-447.
- Hirrlinger, J., Moeller, H., Kirchhoff F and Dringen R (2005) Expression of multidrug resistance proteins (Mrps) in astrocytes of the mouse brain: a single cell RT-PCR study. *Neurochemical Research.* 30, 1237-1244.

- Hinzman, J.M., Thomas, T.C., Quintero, J.E., Gerhardt, G.A., Lifshitz, J. (2012). Disruptions in the regulation of extracellular glutamate by neurons and glia in the rat striatum two days after diffuse brain injury. *Journal of Neurotrauma*, 29(6), 1197–1208.
- Hosseini, N.K., Jose, S., Vidyadaran, S., Nordin, A., Syafinaz. A. (2014). Optimization of cell density and LPS concentration for the evaluation of nitric oxide production on BV-2 cells in a Griess assay. *Malaysian Journal* of *Medicine and Health Sciences*. 10(2). 1-8.
- Hou, Y., Adrian-Segarra, J., Richter, M., Kubin, N., Shin, J., Werner, I. et al. (2015). Animal models and "omics" technologies for identification of novel biomarkers and drug targets to prevent heart failure. *BioMed Research International*. 1-10.
- Hsieh, P.F., Chia, L.G., Ni, D.R., Cheng, L.J., Ho, Y.P., Tzeng, S.F. et al. (2002). Behavior, neurochemistry and histology after intranigral lipopolysaccharide injection. *Neuroreport.* 13(3), 277-280
- Huang, D., Li, Y., Cui, F., Chen, J. and Sun, J. (2016). Purification and characterization of a novel polysaccharide-peptide complex from *Clinacanthus nutans* Lindau leaves. *Carbohydrate Polymers*. 137, 701–708.
- Huang, Y., Henry, C.J., Dantzer, R., Johnson, R.W., Godbout, J.P. (2007). Exaggerated sickness behavior and brain proinflammatory cytokine expression in aged mice in response to intracerebroventricular lipopolysaccharide. *Neurobiology of Aging*. 29(11), 1744-1753.
- Human Metabolome Database (HMDB). LysoPA. Access on 24th January 2019 at http://www.hmdb.ca/metabolites/HMDB0007855
- Human Metabolome Database: Showing metabocard for Formic acid (HMDB0000142). <u>http://www.hmdb.ca/metabolites/HMDB0000142;</u> Accessed 12 September 2018.
- Iravani, M.M., Syed, E., Jackson, M.J., Johnston, L.C., Smith, L.A. and Jenner P. (2005). A modified MPTP treatment regime produces reproducible partial nigrostriatal lesions in common marmosets. *European. Journal of. Neuroscience.* 21(44), 841–854.
- Ismail, N., Arsad, H., Samian, M., & Hamdan, M. (2017). Determination of phenolic and flavonoid contents, antioxidant activities and GC-MS analysis of *Clinacanthus nutans* (Acanthaceae) in different locations. *AGRIVITA Journal of Agricultural Science*, *39*(3).
- ISO 10993-5. (2009). Evaluation of medical devices. Part 5: Test for *in vitro* cytotoxicity. International Organization for standardization, Geneva

- Janwitayanuchit, W., Suwanborirux, K., Patarapanich, C., Pummangura, S., Lipipun, V., Vilaivan, T. (2003). Synthesis and anti-herpes simplex viral activity of monoglycosyl diglycerides. *Phytochemistry*. 64(7):1253-64.
- Jayavasu, C., Balachandra, K. and Sangkitporn, S. (1992a). Clinical trial in the treatment of genital herpes patients with *Clinacanthus nutans* extract. *Communicable Disease Journal*. 18(3), pp. 152–161, 1992.
- Jayavasu, C., et al. (1998). Clinical trial in the treatment of genital herpes patients with *Clinacanthus nutants* extract. *In The 9th Ministry of Public Health Symposium*. 57. Bangkok: E.T.O. Press.
- Jayavasu, C., Dechatiwongse, T., Balachandra, K., Chavalittumrong, P. and Jongtrakulsiri, S. (1992b). The virucidal activity of *Clinacanthus nutans* Lindau extracts against herpes simplex virus type-2: An *in vitro* study. *Bulletin* of theDepartment ofMedical Sciences, 34(4), 153–158. 1992.
- Jia, L, Liu, J, Song, Z, Pan, X, Chen, L, Cui, X, Wang, M. (2012). Berberine suppresses amyloid-beta-induced inflammatory response in microglia by inhibiting nuclear factor-kappaB and mitogen-activated protein kinase signalling pathways. *Journal of Pharmacy and Pharmacology*. 64(10), 1510-21.
- Jin, J., Sundararaj, K.P., Samuel, D.J. et al. (2012). Different signaling mechanism regulating IL-6 expression by LPS between gingival fibroblast and mononuclear cells: seeking the common target. *Clinical Immunology*. 143(2), 188-199
- Jingxia, K. and Ming, Z. (2015). *China patent No. CN104547691-A.* Retrieved from https://patents.google.com/patent/CN104547691A/zh
- Johnson, C., Ivanisevic, J. and Siuzdak, G. (2016). Metabolomics: beyond biomarkers and towards mechanisms. *Nature Reviews Molecular Cell Biology*, 17(7), 451-459. doi: 10.1038/nrm.2016.25
- Johnson, C.H., Ivanisevic, J., Benton, H.P., Siuzdak, G. (2015). Bioinformatics: The next frontier of metabolomics. *Analytical Chemistry*. ;87(2), 147-156.
- Johnston, A.P., De, L.M., Parise, G. (2008). Resistance training, sarcopenia, and the mitochondrial theory of aging. *Applied Physiology, Nutrition and Metabolism.* 33(1), 191–199.
- Jonckheere, A.I., Smeitink, J.A.M., Rodenburg, R.J.T. (2012). Mitochondrial ATP synthase: architecture, function and pathology. *Journal of Inherited Metabolic Disease*, 35(2), 211–225.
- Joo, S.S., Yoo, Y.M., Ahn, B.W., Nam, S.Y., Kim, Y.B., Hwang, K.W., et al. (2008). Prevention of inflammation-mediated neurotoxicity by Rg3 and its role in microglial activation. *Biological and Pharmaceutical Bulletin*. 31(7), 1392–1396. 10.1248/bpb.31.1392

- JoVE Science Education Database. (2019). *Neuroscience*. Rodent Stereotaxic Surgery. *Journal of Visualized Experiments: JoVE*, Cambridge, MA.
- Kamburov, A., Cavill, R., Ebbels, T., Herwig, R., Keun, H. (2011). Integrated pathway-level analysis of transcriptomics and metabolomics data with IMPaLA. *Bioinformatics*. 27(20), 2917-2918.
- Kamburov, A., Pentchev, K., Galicka, H., Wierling, C., Lehrach, H., and Herwig, R. (2010). ConsensusPathDB: toward a more complete picture of cell biology. *Nucleic Acids Research*, 39(suppl_1), D712-D717. doi: 10.1093/nar/gkq1156
- Karhausen, J., Haase, V.H., Colgan, S.P. (2005). Inflammatory hypoxia: Role of hypoxia-inducible factor, *Cell Cycle*. 4(2), 256-258.
- Karnovsky, A., Weymouth, T., Hull, T., Tarcea, V.G., Scardoni, G., Laudanna, C. et al. (2012). Metscape 2 bioinformatics tool for the analysis and visualization of metabolomics and gene expression data. *Bioinformatics*. 28(3), 373-380.
- Karpe, A.V, Beale, D.J., Harding, I.H., Palombo, E.A (2016). Chapter 9: Microbial metabolomics in biomass waste management. *Microbial metabolomics: applications in clinical, environmental and industrial microbiology*. Springer International Publishing Switzerland. 261-291.
- Kato, H., Takahashi, S., and Saito, K. (2011). Omics and integrated omics for the promotion of food and nutrition science. *Journal of Traditional and Complementary Medicine*, 1(1), 25-30. doi: 10.1016/s2225-4110(16)30053-0
- Kelley, K.W., Bluthe, R.M., Dantzer, R., Zhou, J.H., Shen, W.H., Johnson, R.W., Broussard, S.R. (2003). Cytokine-induced sickness behavior. *Brain, Behavior and Immunity.* 17(Suppl 1), 112-118.
- Kelly, R., Lasky-Su, J., Yeung, S., Stone, R., Caterino, J., Hagan, S. et al. (2018). Integrative omics to detect bacteremia in patients with febrile neutropenia. *PLos One.* 13(5):e0197049.
- Khatun, R., Hunter, H., Magcalas, W., Sheng, Y., Carpick, B. and Kirkitadze, M. (2017). Nuclear Magnetic Resonance (NMR) study for the detection and quantitation of cholesterol in HSV529 therapeutic vaccine candidate. *Computational and Structural Biotechnology Journal*. 15, pp.14-20.
- Khoo, L., Audrey Kow, S., Lee, M., Tan, C., Shaari, K., Tham, C., & Abas, F. (2018a). A comprehensive review on phytochemistry and pharmacological activities of *Clinacanthus nutans* (Burm.f.) Lindau. *Evidence-Based Complementary and Alternative Medicine*, 2018, 1-39. doi: 10.1155/2018/9276260
- Khoo, L., Audrey Kow, S., Maulidiani, M., Yen Ang, M. et al. (2018b). 1H-HNMR metabolomics for evaluating the protective effect of *Clinacanthus nutans*

(Burm.f.) Lindau water extract against nitric oxide production in LPS-IFN-γ activated RAW 264.7 macrophages. *Phytochemical Analysis*, 2018, 1-16.

- Khoo, L., Audrey Kow, S., Maulidiani, M., Lee, M., Tan, C., & Shaari, K. et al. (2018c). Plasma and urine metabolite profiling reveals the protective effect of *Clinacanthus nutans* in an ovalbumin-induced anaphylaxis model: ¹ H-NMR metabolomics approach. *Journal of Pharmaceutical and Biomedical Analysis*, 158, 438-450. doi: 10.1016/j.jpba.2018.06.038
- Khoo, L., Foong Kow, A., Maulidiani, M., Lee, M., Tan, C., Shaari, K. et al. (2018d). Hematological, biochemical, histopathological and ¹H-NMR metabolomics application in acute toxicity evaluation of *Clinacanthus nutans* water leaf extract. *Molecules*. 23(9), 2172. doi: 10.3390/molecules23092172
- Khoo, L., Mediani, A., Zolkeflee, N., Leong, S., Ismail, I., & Khatib, A. et al. (2015).
 Phytochemical diversity of *Clinacanthus nutans* extracts and their bioactivity correlations elucidated by NMR based metabolomics. *Phytochemistry Letters*, *14*, 123-133. doi: 10.1016/j.phytol.2015.09.015
- Kim, E., Jung, Y.S., Kim, H., Kim, J.S., Park, M., Jeong, J. et al. (2014). Metabolomic signatures in peripheral blood associated with Alzheimer's disease amyloid-β-induced neuroinflammation. *Journal of Alzheimer's Disease*, 42(2), 421–433.
- Kim, H.J., Rowe, M., Ren, M., Hong, J.S., Chen, P.S., Chuang, D.M. (2007). Histone deacetylase inhibitors exhibit anti-inflammatory and neuroprotective effects in a rat permanent Ischemic model of stroke: multiple mechanisms of action. *Journal of Pharmacology and Experimental Therapeutics*, 321(3), 892–901.
- Kim, J.K., Bamba, T., Harada, K., Fukusaki, E., Kobayashi, A. (2006). Time-course metabolic profiling in *Arabidopsis thaliana* cell cultures after salt stress treatment. *Journal of. Experimental. Botany*. 58(3), 415–424
- Kim, S.Y., Gao, J.J., Lee, W.C., Ryu, K.S., Lee, K.R., Kim, Y.C. (1999). Antioxidative flavonoids from the leaves of *Morus alba*. *Archieves of Pharmacal Research*. 22(1):81-5.
- Kim, W.G., Mohney, R.P., Wilson, B., Jeohn, G.H., Liu, B., Hong, J.S. (2000). Regional difference in susceptibility to lipopolysaccharide-induced neurotoxicity in the rat brain: role of microglia. *Journal of Neuroscience*. 20(6), 6309–6316.
- Kim, E., Jung Y.S., Kim, H. et al. (2014). Metabolomic signatures in peripheral blood associated with Alzheimer's disease amyloid-*θ*-induced neuroinflammation. *Journal of Alzheimers Dicoveries*. 42(2), 421-433.
- Kjeldahl, K. and Bro, R. (2010). Some common misunderstandings in chemometrics. *Journal of Chemometrics*. 24(7-8), 558–564.

- Kongkaew, C. and Chaiyakunapruk, N. (2011). Efficacy of *Clinacanthus nutans* extracts in patients with herpes infection: Systematic review and metaanalysis of randomised clinical trials. *Complementary Therapies in Medicine* 19(1), 47–53.
- Kosmides, A.K., Kamisoglu, K., Calvano, S.E. et al. (2013). Metabolomic fingerprinting: challenges and opportunities. *Critical Reviews in Biomedical Engineering*. 41(3), 205-221.
- Kumar, N., Hoque, M., Shahjaman, M., Islam, S. and Mollah, M. (2017). Metabolomic biomarker identification in presence of outliers and missing values. *Biomedical Research International*. 2017, 1-11. doi: 10.1155/2017/2437608
- Kunsorn, P., Ruangrungsi, N., Lipipun, V., Khanboon, A., Rungsihirunrat, K. (2013). The identities and anti-herpes simplex virus activity of *Clinacanthus nutans* and *Clinacanthus* siamensis. Asian Pacific Journal of Tropical Biomedicine, 3(4), 284–290.
- Kurtz, H.J. and Quast, J. (1982). Effects of continuous intravenous infusion of *Escherichia coli* endotoxin into swine. *American Journal of Veterinary Research*. 43(2), 262-268.
- Kuwabara, T., Watanabe, H., Tsuji, S., Yuasa, T. (1995). Lactate rise in the basal ganglia accompanying finger movements: a localized ¹H-MRS study. *Brain Research*, 670(2), 326–328.
- Lau, K., Lee, S., and Chin, J. (2014). Effect of the methanol leaves extract of *Clinacanthus nutans* on the activity of acetylcholinesterase in male mice. *Journal of Acute Disease*, 3(1), 22-25.
- Laura, C. (2011). Fundamentals of inflammation, Yale Journal of Biology and Medicine, 84(1): 64-65.
- Lawson, L.J., Perry, V.H., Dri, P., Gordon, S. (1990). Heterogeneity in the distribution and morphology of microglia in the normal adult mouse brain. *Neuroscience*. 39(I):151–170. doi: 10.1016/0306-4522(90)90229-W.
- Le Guennec, A., Tayyari, F., Edison, A.S. (2017). Alternatives to nuclear overhauser enhancement spectroscopy presat and carr-purcell-meiboom-gill presat for NMR-based metabolomics. *Analytical Chemistry*, 89(17), 8582–8588.
- Le, C., Kailaivasan, T., Chow, S., Abdullah, Z., Ling, S. and Fang, C. (2017). Phytosterols isolated from *Clinacanthus nutans* induce immunosuppressive activity in murine cells. *International Immunopharmacology*, 44, .203-210.
- Lee, J., Lee, B., Chung, J., Hwang, J., Lee, S., Lee, C., & Hong, Y. (2010). Geographical and climatic dependencies of green tea (*Camellia sinensis*) Metabolites: A ¹H NMR-based metabolomics study. *Journal of Agricultural and Food Chemistry*, *58*(19), 10582-10589. doi: 10.1021/jf102415m

- Lee, K.M. (2015). New advances on glial activation in health and disease. *World Journal of Virology*. 4(2), 42.
- Lee, S. Y., Mediani, A., Nur Ashikin, A. H., Azliana, A.B.S., Abas, F., (2014). Antioxidant and α-glucosidase inhibitory activities of the leaf and stem of selected traditional medicinal plants. *International Food Research Journal*, 21(1), pp. 165–172.
- Lee, Y., Morrison, B.M., Li, Y., Lengacher, S., Farah, M.H., Hoffman, P.N et al. (2012). Oligodendroglia metabolically support axons and contribute to neurodegeneration. *Nature*, 487(7408), 443–448.
- Li, X., Kimura, H., Hirota, K., Sugimoto, H., Yoshida, H. (2005). Hypoxia reduces constitutive and TNF-α-induced expression of monocyte chemoattractant protein-1 in human proximal renal tubular cells. *Biochemical and Biophysical Research Communications*. 335(4),1026-1034.
- López, J.C. (2003). Neurodegenerative diseases: a promising therapy for SBMA. *Nature Reviews Neuroscience*, 4: 519–526.
- Lull, M., and Block, M. (2010). Microglial activation and chronic neurodegeneration. *Neurotherapeutics*, 7(4), 354-365.
- Lyman, M., Lloyd, D.G., Ji, X., Vizcaychipi, M.P., Ma, D. (2014). Neuroinflammation: the role and consequences. *Neuroscience Research*, 79, 1–12.
- Ma, S, Chowdhury, S.K., Alton, K.B. (2006). Application of mass spectrometry for metabolite identification. *Current Drug Metabolism* 7(5), 503–523.
- Maddock, R.J., Buonocore, M.H. (2011). MR spectroscopic studies of the brain in psychiatric disorders. In: Carter C., Dalley J. (eds) Brain Imaging in Behavioral Neuroscience. *Current Topics in Behavioral Neurosciences*, pp. 199–251, Berlin, Heidelberg, Springer.
- Mahadevan, S., Shah, S., Marrie, T., and Slupsky, C. (2008). Analysis of metabolomic data using support vector machines. *Analytical Chemistry*, 80(19), 7562-7570. doi: 10.1021/ac800954c
- Mai, C.W., Yap K.S.I., Kho M.T. et al., (2016). Mechanisms underlying the antiinflammatory effects of *Clinacanthus nutans* lindau extracts: Inhibition of cytokine production and toll-like receptor-4 activation," *Frontiers in Pharmacology*. 7(7), 1–11.
- Makino, J., Kato, K. and Maes, F.W. (1991). Temporal structure of open field behavior in inbred strains of mice. *Japanese. Psychological. Research*. 33(4), 145–152.
- Mao, X.-Y., Cao, D.-F., Li X., Yin, J.-Y., Wang, Z.-B., Zhang, Y. et al. (2014). Huperzine A ameliorates cognitive deficits in streptozotocininduced diabetic rats. *International. Journal of. Molecular. Sciences.* 15(5), 7667–7683. 10.3390/ijms15057667

- Maragakis, N.J., Rothstein, J.D. (2001). Glutamate transporters in neurologic disease. *Archieve of. Neurology*, 58(3), 365–370.
- Markel', K. Galaktionov, and V. M. Efimov, (1988). Factor analysis of rat behavior in the open-field test. *Zhurnal Vysshei Nervnoi Deiatelnosti Imeni I P Pavlova.* 38(5), 855-863.
- Markiewski, M., & Lambris, J. (2007). The role of complement in inflammatory diseases from behind the scenes into the spotlight. *The American Journal of Pathology*, *171*(3), 715-727.
- Martins, I. (2016). Bacterial lipopolysaccharides change membrane fluidity with relevance to phospholipid and amyloid beta dynamics in Alzheimer's disease. *Journal of Microbial and Biochemical Technology*, 8(4). doi: 10.4172/1948-5948.1000304
- Mattson, M.P. (2008). Hormesis defined. Ageing Research Reviews. 7(1), 1-7.
- Matsuda, M., Huh, Y., & Ji, R. (2018). Roles of inflammation, neurogenic inflammation, and neuroinflammation in pain. *Journal of Anesthesia*, 33(1), 131-139. doi: 10.1007/s00540-018-2579-4
- Mayer, H., Bhat, U.R., Masoud, H., Radziejewskalebrecht, J., Widemann, C., Krauss, J.H. (1989) Bacterial lipopolysaccharides. *Pure and Applied. Chemistry*. 61(7), 1271–1282.
- Mediani, A., Abas, F., Khatib, A., Maulidiani, H., Shaari, K., Choi, Y., & Lajis, N. (2012). ¹H-NMR-based metabolomics approach to understanding the drying effects on the phytochemicals in *Cosmos caudatus*. *Food Research International*, 49(2), 763-770. doi: 10.1016/j.foodres.2012.09.022
- Medzhitov, R. and Janeway, C. Jr. (2000). Innate immune recognition: mechanisms and pathways. *The New England Journal of Medicine*. 343(5), 338-344
- Metz, G.A., Jadavji, N.M., Smith, L.K. (2005). Modulation of motor function by stress: a novel concept of the effects of stress and corticosterone on behavior. *European Journal of Neuroscience*, 22(5), 1190–1200.
- Milatovic, D., Zaja-Milatovic, S., Montine, K.S., Horner, P.J., Montine, T.J. (2003). Pharmacologic suppression of neuronal oxidative damage and dendritic degeneration following direct activation of glial innate immunity in mouse cerebrum. *Journal of Neurochemistry*, 87(6), 1518–1526.
- Ministry of Public Health, Thailand. National Drug Committee. (1999). *National List of Essential Drugs A.D. 1999 (List of Herbal Medicinal Products).* Bangkok: E.T.O. Press.
- Mochida, K. and Shinozaki, K (2011) Advances in omics and bioinformatics tools for systems analyses of plant functions. *Plant and Cell Physiology* 52(11), 2017–2038.

- Moore, R., Kirwan, J., Doherty, M. and Whitfield, P. (2007). Biomarker discovery in animal health and disease: The application of post-genomic technologies. *Biomarker Insights*. 2, p.117727190700200.
- More, S.V., Kumar, H., Kim, I.S., Song, S.Y. and Choi, D. K. (2013). Cellular and molecular mediators of neuroinflammation in the pathogenesis of Parkinson's disease. *Mediators of Inflammation*. 2013, 952375
- Morrison, D.C. and Rudbach, J. A. (1981). Endotoxin-cell-membrane interactions leading to transmembrane signalling, *Contemporary. Topics in. Molecular. Immunology.* 8, 187.
- Mosihuzzaman, M. (2012). Herbal medicine in healthcare--an overview. *Natural Products Communications*, *7*(6), 807-812.
- Mosmann, T.R. and Coffman, R.L. (1989). TH1 and TH2 Cells: Different patterns of lymphokine secretion lead to different functional properties. *Annual Review of Immunology*. 7(1), 145-173.
- Muller, E.E., Pinel, N., Laczny, C.C., Hoopmann, M.R., Narayanasamy, S., Lebrun, L.A. et al .(2014). Community-integrated omics links dominance of a microbial generalist to fine-tuned resource usage. *Nature Communications.* 5, 5603. (https://doi.org/10.1038/ncomms6603)
- Mumm, R., Hageman, J., Calingacion, M., de Vos, R., Jonker, H., Erban, A. et al. (2016). Multi-platform metabolomics analyses of a broad collection of fragrant and non-fragrant rice varieties reveals the high complexity of grain quality characteristics. *Metabolomics*, 12(2). doi: 10.1007/s11306-015-0925-1
- Munford, R.S., Hall, C.L. and Dietschy, J.M. (1981). Binding of Salmonella typhimurium lipopolysaccharides to rat high-density lipoproteins. Infection and Immunity. 34(3), 835-843.
- Murín, R., Schaer, A., Kowtharapu, B.S., Verleysdonk, S., Hamprecht, B. (2008). Expression of 3-hydroxyisobutyrate dehydrogenase in cultured neural cells. *Journal of Neurochemistry*, 105(4), 1176–1186.
- Murphy, K.M. and Reiner, S.L. (2002). The lineage decisions of helper T cells. *Nature Reviews Immunology*.2(12), 933-944.
- Mustapa, A.N., 'Martin, A., Mato, R.B. and Cocero, M.J. (2015). Extraction of phytocompounds from the medicinal plant *Clinacanthus nutans* Lindau by microwave-assisted extraction and supercritical carbon dioxide extraction. *Industrial Crops and Products*. 74, 83–94.
- Myokai, F., Takashiba, S., Lebo, R and Amar, S. (1999). A novel lipopolysaccharide-induced transcription factor regulating tumor necrosis factor α gene expression: molecular cloning, sequencing, characterization, and chromosomal assignment. 96(8), 4518-4523.

- Nguyen, M.D., Julien, J.P., Rivest, S. (2002). Innate immunity: the missing link in neuroprotection and neurodegeneration? *Nature Reviews. Neuroscience*. 3(3), 216-227.
- Ni, J., Wu, Z., Ling, S., Zhu, X., Lu, P. (2006). Effect of Matrine injection on IL-1beta level and ultrastructural changes of hippocampal neuron in Alzheimer's disease rat. Chinese. Journal of. Anatomy. 29, 608–611.
- Nicholson, J. and Wilson, I. (2003). Understanding 'global' systems biology: Metabonomics and the continuum of metabolism. *Nature Reviews Drug Discovery*, 2(8), 668-676. doi: 10.1038/nrd1157
- Nicholson, J.K. and Lindon, J.C. (2008). System biology: Metabonomics. *Nature*. 455(7216), 1054–1056.
- NIH Eastern Regional Comprehensive Metabolomics Resource Core at Research Triangle Institute procedure for UPLC-TOF MS for HILIC method on broad spectrum analysis (Center Specific Procedure (CSP) no: RTI-RCMRC-LCMS-01 ver.00) (www.rti.org/rcmrc, accessed on 10th April 2017).
- Nishida, K., Ono, K., Kanaya, S., and Takahashi, K. (2014). KEGGscape: a Cytoscape app for pathway data integration. *F1000research*, 3, 144. doi: 10.12688/f1000research.4524.1
- Norouzi, F., Abareshi, A., Anaeigoudari, A., Shafei, N.M., Gholamnezhad, Z., Saeedjalali, M., et al. (2016). The effects of *Nigella sativa* on sickness behavior induced by lipopolysaccharide in male Wistar rats. *Avicenna Journal of Phytomedicine*, 6(1), 104-116.
- Orthmann-Murphy, J.L., Abrams, C.K., Scherer, S.S. (2008). Gap functions couple astrocytes and oligodendrocytes. *Journal of Molecular Nueroscience*. 35, 101-116.
- O'Shea, J., Ma, A., Lipsky, P. (2002). Cytokines and autoimmunity. *Nature Reviews: Immunology*. 2(1), 37-45
- O'Shea, J.J., Gadina, M., Kanno, Y. (2011). Cytokine Signaling: Birth of a pathway. *The Journal of Immunology*. 187 (11), 5475-5478.
- Olajide, O., Bhatia, H., de Oliveira, A., Wright, C. and Fiebich, B. (2013). Inhibition of neuroinflammation in LPS-activated microglia by cryptolepine. *Evidence-Based Complementary and Alternative Medicine*, *2013*, 1-10.
- Paban, V., Loriod, B., Villard, C., Buee, L., Blum, D., Pietropaolo, S. et al. (2017). Omics analysis of mouse brain models of human diseases. *Gene*. 600, 90-100.
- Pannangpetch, P., Laupattarakasem, P., Kukongviriyapan, V., Kukongviriyapan, U., Kongyingyoes, B., and Aromdee. C. (2007). Antioxidant activity and protective effect against oxidative hemolysis of *Clinacanthus nutans*

(Burm.f) Lindau. Songklanakarin Journal of Science and Technology, 29 (1), pp. 1–9.

- Panyakom K. (2006). Structural elucidation of bioactive compounds of *Clinacanthus nutans* (Burm. F.) Lindau leaves. *Thailand: Suranaree University of Technology*.Papaneophytou, C., Georgiou, E. and Kleopa, K.A. (2019). The role of oligodendrocyte gap junctions in neuroinflammation, *Channels*, 13(1), 247-263.
- Pardridge, W.M., Golden, P.L., Kang, Y.S. and Bickel, U. (1997) Brain microvascular and astrocyte localization of p-glycoprotein. *Journal of Neurochemistry.* 68, 1278-1285.
- Pathak, R.R. and Dave, V (2014). Integrating omics technologies to study pulmonary physiology and pathology at the systems level. *Cellular Physiology* and *Biochemistry* 33(5), 1239–1260. (https://doi. org/10.1159/000358693)
- Patti, G.J., Yanes, O., Siuzdak, G. (2012). Innovation: Metabolomics: the apogee of the omics trilogy. *Nature Reviews Molecular Cell Biology*. 13(4), 263-269.
- Petroff, O.A.C. (2002). GABA and glutamate in the human brain. *Neuroscientist*, 8(6), 562–573.
- Pinto, R.C. (2017). Chemometrics methods and strategies in metabolomics. Advances in Experimental Medicine and Biology. 965, 163–190.
- Pluskal, T., Castillo, S., Villar-Briones, A., and Orešič, M. (2010). MZmine 2: Modular framework for processing, visualizing, and analyzing mass spectrometry-based molecular profile data. *BMC Bioinformatics*, 11(1). doi: 10.1186/1471-2105-11-395
- P'ng X.W., Akowuah, G.A., Chin, J.H. (2013). Evaluation of the sub–acute oral toxic effect of methanol extract of *Clinacanthus nutans* leaves in rats. *Journal of Acute Disease*. 2(1), 29-32.
- Polanski, J. (2017). Chemoinformatics: From chemical art to chemistry *in silico*. Reference Module in Chemistry, *Molecular Sciences and Chemical Engineering*. Elsevier, Oxford (2017), 459–506.
- Poller, B., Gutmann, H., Krahenbuhl, S., Weksler, B. et al. (2008) The human brain endothelial cell line HCMEC/D3 as a human blood-brain barrier model for drug transport studies. *Journal of Neurochemistry* 107, 1358-1368.
- Poltorak, A, He, X., Smirnova, I., Liu, M.Y., Van Huffel, C., Du, X., et al. (1998). Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: Mutations in Tlr4 gene. *Science*. 282(5396), 2085–2088.
- Pongphasuk, N., Khunkitti, W. and Chitcharoenthum, M. (2005). Antiinflammatory and analgesic activities of the extract from *Garcinia mangostana* linn. *Acta Horticulturae*, 680, 125–130.

- Purkayastha, S. and Cai, D. (2013). Neuroinflammatory basis of metabolic syndrome. *Molecular Metabolism*, 2(4), 356–363.
- Qian, C., Jiang, X., An, H., Yu, Y., Guo, Z., Liu, S., et al. (2006). TLR agonists promote ERK-mediated preferential IL-10 production of regulatory dendritic cells (diffDCs), leading to NK-cell activation. *Blood*. 108(7), 2307–2315
- Qin, L., Wu, X., Block, M.L., Liu, Y., Breese, G.R., Hong, J.S. et al. (2007). Systemic LPS causes chronic neuroinflammation and progressive neurodegeneration. *Glia*, 55(5), 453–462.
- Raetz, C. R. (1990). Biochemistry of endotoxins. Annual Review of Biochemistry. 59, 129-170.
- Raetz, C.R. and Whitfield, C. (2002). Lipopolysaccharide endotoxin. *Annual Review of Biochemistry*. **7**1, 635 -700.
- Ramesh, G, Maclean, A.G., Philipp, M.T. (2013). Cytokines and chemokines at the crossroads of neuroinflammation, neurodegeneration, and neuropathic pain. *Mediators Inflammation*. 1–20.
- Rathee P, Chaudhary H, Rathee S, Rathee D, Kumar V, Kohli K. (2009). Mechanism of action of flavonoids as antiinflammatory agents. *Inflammation and Allergy Drug Targets*. 8(3): 229-235.
- Reisenauer, C., Bhatt, D., Mitteness, D., Slanczka, E., Gienger, H., Watt, J., & Rosenberger, T. (2011). Acetate supplementation attenuates lipopolysaccharide-induced neuroinflammation. *Journal of Neurochemistry*, 117(2), 264-274.
- Rietschel, E.T., Kirikae, T., Schade, F.U., Mamat, U., Schmidt, G. et al. (1994). Bacterial endotoxin: molecular relationships of structure to activity and function. *FASEB Journal*. 8(2), 217–225
- Reo, N.V. (2002). NMR-based metabolomics. *Drug and Chemical Toxicology*. 25(4):375–82.
- Riske, L., Thomas, R.K., Baker, G.B. et al. (2017). Lactate in the brain: an update on its relevance to brain energy, neurons, glia, and panic order. Therapeutic Advances in Psychopharmacology. 7(2), 85-89.
- Ritchie, M.D., Holzinger, E.R., Li R, Pendergrass, S.A. and Kim, D (2015) Methods of integrating data to uncover genotype-phenotype interactions. *Nature Reviews Genetics*. 16(2), 85–97. (<u>https://doi.org/10.1038/nrg3868</u>)
- Robles, M., Humphrey, S., and Mann, M. (2017). Phosphorylation is a central mechanism for Circadian control of metabolism and physiology. *Cell Metabolism*, 25(1), 118-127. doi: 10.1016/j.cmet.2016.10.004

- Rock, R.B., Gekker, G., Hu, S., Sheng, W.S., Cheeran, M., Lokensgard, J.R. et al (2004). Role of microglia in central nervous system infections role. *Clinical Microbiology Rev*, 17(4), 942–964.
- Rohde, F., Schusser, B., Hron, T., Farkasova, H. et al. (2018). Characterization of chicken tumor necrosis factor-α, a long missed cytokibe in bird. *Frontiers Immunology*.19, 605.
- Ronald, H., Zielke, C., Baab, P. (2009). Direct measurement of oxidative metabolism in the living brain by microdialysis: a review. *Journal of Neurochemistry*. 109, 24-29.
- Roslan, N.A., Kassim, N.I.I., Lim, V. and Jemon, K. (2017). Subacute toxicity study of *Clinacanthus nutans* ethanolic extract *in vivo. Proceeding of International Postgraduate Symposium in Biotechnology 2017. (IPBS 2017), 81-84*
- Roux, A., Lison, D., Junot, C., and Heilier, J. (2011). Applications of liquid chromatography coupled to mass spectrometry-based metabolomics in clinical chemistry and toxicology: A review. *Clinical Biochemistry*, 44(1), 119-135. doi: 10.1016/j.clinbiochem.2010.08.016
- Wold, S., Albano, C., Dunn, W.J. III., Esbensen, K. et al., (1983). Pattern recognition: finding and using regularities in multivariate data. *Martens and Russwurm, Eds*
- Sakdarat, S., Shuyprom, A., Dechatiwongse, N. A. T., Waterman, P. G, .and Karagianis, G. (2006). Chemical composition investigation of the *Clinacanthus nutans* Lindau leaves. *Thai Journal of Phytopharmacy*, 13(2), 13–24.
- Sakdarat, S., Shuyprom, A., Pientong, C., Ekalaksananan, T., Thongchai, S. (2009). Bioactive constituents from the leaves of *Clinacanthus nutans* Lindau. *Bioorganic and Medicinal Chemistry* 17(5), 1857-1860.

Salzet M, Vieau D, Day R (2000). Crosstalk between nervous and immune systems through the animal kingdom: focus on opioids. *Trends in Neuroscience*; 23(11), 550-555.

- Sangkitporn, S., Kroavon, B., Thawatsupha, P., Bunjob, M. and Chavalittumrong, P. (1993). Treatment of recurrent genital herpes simplex virus infection with *Clinacanthus nutans extract. Bulletin of the Department of Medical Service*. 18, 226–231.
- Sarega, N., Imam, M., Esa, N., Zawawi, N., & Ismail, M. (2016). Effects of phenolicrich extracts of Clinacanthus nutans on high fat and high cholesterol dietinduced insulin resistance. *BMC Complementary and Alternative Medicine*, 16(1), 88.
- Saric, J., Li, J., Swann, J., Utzinger, J., Calvert, G., Nicholson, J. et al. (2010). Integrated cytokine and metabolic analysis of pathological responses to

parasite exposure in rodents. *Journal of Proteome Research*. 9(5), 2255-2264.

- Satakhun, S. (2001). Chemical Constituents of *Clinacanthus nutans* leaves, *Chulalongkorn University*.
- Sawasdimongkol, K. (1995). Medicinal Plants and Thai Traditional Medicine. Department of Medical Sciences: Ministry of Public Health, Thailand.
- Schletter, J., Heine, H., Ulmer, A.J. and Rietschel, E.T. (1995). Molecular mechanisms of endotoxin activity. *Archieves of Microbiology*, 164(6), 383– 389
- Schripsema, J. (2010). Application of NMR in plant metabolomics: techniques, problems and prospects. *Phytochemical Analysis*. 21(1), pp.14-21.
- Schurr, A. and Payne, R.S. (2007). Lactate, not pyruvate, is neuronal aerobic glycolysis end product: an *in vitro* electrophysiological study. *Neuroscience*, 147(3), 613–619.
- Scott, A., Khan, K.M., Cook, J.L., Duronio, V. (2004). What is "inflammation"? Are we ready to move beyond Celsus? *British Journal of Sports Medicine*, 38(3), 248–249.
- Sébédio, J., Pujos-Guillot, E., and Ferrara, M. (2009). Metabolomics in evaluation of glucose disorders. *Current Opinion in Clinical Nutrition and Metabolic Care*, 12(4), 412-418. doi: 10.1097/mco.0b013e32832c97c3
- Sedghipour, M.R., and Sadeghi-Bazargani, H. (2012). Applicability of supervised discriminant analysis models to analyze astigmatism clinical trial data. *Clinical Ophthalmology*, 1499. doi: 10.2147/opth.s34907
- Seeliger, S., Janssen, P., & Schink, B. (2002). Energetics and kinetics of lactate fermentation to acetate and propionate via methylmalonyl-CoA or acrylyl-CoA. FEMS Microbiology Letters, 211(1), 65-70. doi: 10.1016/s0378-1097(02)00651-1
- Sengupta, A., Ficker, A.M., Dunn, S.K., Madhu, M., Cancelas, J.A. (2012). Bmi1 reprograms CMLB-lymphoid progenitors to become B-ALL-initiating cells. *Blood*, 119(2), 494–502.
- Serrats, J., Schiltz, J., García-Bueno, B., van Rooijen, N., Reyes, T., Sawchenko, P. (2010). Dual roles for perivascular macrophages in immune-to-brain signaling, *Neuron*. 65(1), 94-106.
- Sethi, S. and Brietzke, E. (2015). Omics-Based Biomarkers: application of metabolomics in neuropsychiatric disorders. *International Journal of Neuropsychopharmacology*, 19(3), 1-13.

- Shah, K., DeSilva, S., Abbruscato, T. (2012). The role of glucose transporters in brain disease: Diabetes and Alzheimer's disease. *International Journal of Molecular Sciences*. 13(12), 12629-12655.
- Shah, K., DeSilva, S., Abbruscato, T. (2012). The role of glucose transporters in brain disease: Diabetes and Alzheimer's disease. *International Journal of Molecular Sciences*. 13(12), 12629-12655.
- Shal, B., Ding, W., Ali, H., Kim, Y., Khan, S. (2018). Anti-neuroinflammatory Potential of Natural Products in Attenuation of Alzheimer's Disease. *Frontiers in Pharmacology*, 9, 548.
- Shanaiah, N., Zhang, S., Desilva, M.A., Raftery, D. (2008). NMR-based metabolomics for biomarker discovery. Biomarker Methods in Drug Discovery and Development Methods in Pharmacology and Toxicology™. 341–68.
- Shuyprom, A. (2004). Chemical composition investigation of the *Clinacanthus nutans* (Burm. F.) Lindau leaves. *Thailand: Suranaree University* of *Technology*
- Siew, Y., Zareisedehizadeh, S., Seetoh, W., Neo, S., Tan, C., and Koh, H. (2014). Ethnobotanical survey of usage of fresh medicinal plants in Singapore. *Journal of Ethnopharmacology*, 155(3), 1450-1466.
- Simon, N., Friedman, J., Hastie, T., Tibshirani, R. (2013). A sparse-group lasso. *Journal of Computational and Graphical Statistics*. 22(2), 231–245.
- Simon, P., Dupuis, R., Costentin, J. (1994). Thigmotaxis as an index of anxiety in mice. Influence of dopaminergic transmissions. *Behavioural Brain Research*. 61(1), 59-64.
- Siriporn Timpawat, .Lo.V. (2013). Clinical evaluation of *Clinacanthus nutans* Lindau in orabase in the treatment of recurrent aphthous stomatitis. Mahidol Dent Journal, 14 (1), pp. 10-16.
- Sittiso, S., Ekalaksananan, T. and Pientong, C. (2010). Effects of compounds from *Clinacanthus nutans* on dengue virus type 2 infection," *Srinagarind Medical Journal.* 25, 272–275.
- Skelly, R.R., Munkenbeck, P. and Morrison, D.C. (1979). Stimulation of Tindependent antibody responses by hapten-lipopolysaccharides without repeating polymeric structure. *Infection and Immunity*. 23(2), 287-293.
- Sleiman, S.F, Henry, J., Al-Haddad, R., El Hayek, L., Abou Haidar, E., Stringer, T. et al. (2016). Exercise promotes the expression of brain derived neurotrophic factor (BDNF) through the action of the ketone body β-hydroxybutyrate. *Elife*, 5; e15092.
- Smith, C., Want, E., O'Maille, G., Abagyan, R., and Siuzdak, G. (2006). XCMS: processing mass spectrometry data for metabolite profiling using

nonlinear peak alignment, matching, and identification. *Analytical Chemistry*. 78(3), 779-787. doi: 10.1021/ac051437y

- Sochocka, M., Diniz, B.S., Leszek, J. (2016). Inflammatory response in the CNS: Friend or foe? *Molecular Neurobiology*;54(10), 8071–8089.
- Solaini, G., Baracca, A., Lenaz, G., Sgarbi, G. (2010). Hypoxia and mitochondrial oxidative metabolism. *Biochimica et Biophysica Acta*, 1797(6-7), 1171–1177.
- Song, X., Fan, X., Li, X., Zhang, W., Gao, J., Zhao, J. et al. (2014). Changes in pro-inflammatory cytokines and body weight during 6-month risperidone treatment in drug naïve, first-episode schizophrenia. *Psychopharmacology*. 231(2), 319-325.
- Sookmai, W., Ekaalaksananan, T., Pientong, C., Sakdarat, S. and Kongyingyoes, B. (2011). The anti-papillomavirus infectivity of *Clincantus nutans* compounds. *Srinagarind Medicinal Journal*, 26(Suppl), 240-242.
- Soomro, S. (2019). Oxidative stress and inflammation. Open Journal of Immunology, 9, 1-20.
- South China botanical garden: *Clinacanthus nutans (Burm.)* Lindau. 2008 [Online] Available from: <u>http://www.efloras.org/</u> florataxon.aspx?flora_id=610&taxon_id=200021997 [Accessed on 22 Feb 2008]
- Sriwanthana, B., Chavalittumrong, P. and Chompuk. L. (1996). Effect of *Clinacanthus nutans* on human cell-mediatedimmune response *in vitro*. *Thailand Journal of Pharmaceutical Science*. 20(4), 261–267.
- Storey, J.D, Tibshirani, R., (2003). Statistical significance for genomewide studies. Proceedings of the National Academy of Sciences.100, 9440–9445.
- Stuehr, D., & Griffith, O. (1992). Advances in Enzymology. Meister A (Ed) (pp. 287-346). New York: Wiley.
- Sun, H., Zhang, A., Wang, X. (2012). Potential role of metabolomic approaches for Chinese medicine syndromes and herbal medicine. *Phytotherapy Research*. 26(10), 1466-1471.
- Sun, Z.K., Yang, H.Q., Chen, S.D. (2013). Traditional chinese medicine: a promising candidate for the treatment of Alzheimer's disease. *Translational Neurodegenerative*, 2(1), 6.
- Tambuyzer, B., Ponsaerts, P. and Nouwen, E. (2009). Microglia: gatekeepers of central nervous system immunology. *Journal of Leukocyte Biology*, 85(3), pp.352-370.
- Tanaka, T., Narazaki, M., T., Kishimoto, T. (2014). IL-6 in inflammation, immunity, and disease. *Cold Spring Harbor Perspective in Biology*. 6(10), a016294.

- Taxonomic Hierarchy of COL-China 2012. "Clinacanthus nutans." Encyclopedia of Life; Available from; http://eol.org/pages/5634686/names/common_names.[Accessed 12 August 2017].
- Teo, C., Chong, W., Tan, E., Basri, N., Low, Z. and Ho, Y. (2015). Advances in sample preparation and analytical techniques for lipidomics study of clinical samples. *TrAC Trends in Analytical Chemistry*. 66, pp.1-18.
- Teshima, K.I., Kaneko, T., Ohtani, K., Kasai, R., Lhieochaiphant, S., Picheansoonthon, C., Yamasaki, K. (1998). Sulphur containing glucosides from *Clinacanthus nutans*. *Phytochemistry*. 48(5), 831-835.
- Thakur, A., Wilcox, M.D., Stapleton, F. (1998). The proinflammatory cytokines and arachidonic acid metabolites in human overnight tears: homeostatic mechanisms. *Journal of Clinical Immunology*. 18(1), 61-70.
- Thamlikitkul, V. (1996). Treatment of herpes genitalis and herpes zoster with *Clinacanthus nutans. Journal of Infectious Diseases and Antimicrobial Agents*, pp. 95-96.
- Theodoridis, G.A., Gika, H.G., Plumb, R., Wilson, D. (2013). Liquid chromatographic methods combined with mass spectrometry in metabolomics. *Proteomic and Metabolomic Approaches to Biomarker Discovery*, 145-161. Academic Press. Elsevier. San Diego, USA.
- Thomson, A.W. and Lotze, M.T. (1993). The cytokine handbook. Immunology today. 13(11): 466-467. doi:10.1016/0167-5699(92)90082-I.
- Thongharb, C. and Tejasen, P. (1977). The effect of Slaed Pang Porn (*Clinacanthus nutans*) on Thailand cobra venom (*Naja naja siamensis*). *Thai Journal of Pharmaceutical Sciences*. 2, 1057–1063, 1977.
- Tieu K. A guide to neurotoxic animal models of Parkinson's disease. *Cold Spring Harbor Perspectives in Medicine*. 2011;1(1):a009316
- Tijani, A.Y., Salawu, O.A., Jaiyeoba, G-I., Anuka, J.A., Hussaini, I.M. (2012). Neuro-pharmacological effects of *Crinum zeylanicum* in mice. *Avicenna Journal of Phytomedicine*, 2(3), 162-168.
- Tinh, T.D.D. (2014). Biological activities of *Clinacanthus nutans* (Burm.F) Lindau Extracts, Vietnam National University in HCMC.
- Tohidpour, A., Morgun, A., Boitsova, E., Malinovskaya, N., Martynova, G., Khilazheva, E. et al. (2017) Neuroinflammation and infection: molecular mechanisms associated with dysfunction of neurovascular unit. *Frontiers in Cellular and Infection Microbiology*. 7. doi:10.3389/fcimb.2017.00276.
- Tomazini, F.M., Reimer, A., Albrechet-Souza, L., Brand, M.L. (2006). Opposite effects of short- and long-duration isolation on ultrasonic vocalization, startle and prepulse inhibition in rats. *Journal of Neuroscience Methods*. 153(1), 114–120.

- Tripathi, Y.C., Anjum, N., Kumar, R, Tewari, D. (2014). Phytochemical Approach to ascertain quality and efficacy of plant drugs. *Journal of Science, Technology and Management,* 7(4), 279-285.
- Trupp, M., Zhu, H., Wikoff ,W.R. et al. (2012). Metabolomics reveals amino acids contribute to variation in response to simvastatin treatment. *PLoS One.* 7(7), e38386. 10.1371/journal.pone.0038386
- Tu, S.-F., Liu,R.H., Y.-B. Cheng, Y.-B. et al., (2014). Chemical constituents and bioactivities of *Clinacanthus nutans* aerial parts, *Molecules*, vol. 19(12), 20382–20390.
- Tulipani, S., Llorach, R., Urpi-Sarda, M. and Andres-Lacueva, C. (2012). Comparative analysis of sample preparation methods to handle the complexity of the blood fluid metabolome: When less is more. *Analytical Chemistry*, 85(1), pp.341-348.
- Tullis, T. and Albert, B. (2013). Chapter 9; special topics. In T. Tullis, & B. Albert (Eds.), *Measuring the user experience (second edition)* (pp. 209-236). Boston: Morgan Kaufmann.
- Tuntiwachwuttikul, P., Pootaeng-On, Y., Phansa, P., Taylor, W.C. (2004). Cerebrosides and a monoacylmonogalactosylglycerol from *Clinacanthus nutans. Chemical and Pharmaceutical Bulletin (Tokyo)*, 52(1), 27–32.
- Tyrtyshnaia, A., Lysenko, L., Madamba, F., Manzhulo, I., Khotimchenko, M., Kleschevnikov, A. (2016). Acute neuroinflammation provokes intracellular acidification in mouse hippocampus. *Journal of Neuroinflammation*. 13(1); 283.
- Uawonggul, N., Chaveerach, A., Thammasirirak, S., Arkaravichien, T., Chuachan, C. and Daduang, S. (2016). Screening of plants acting against *Heterometrus laoticus* scorpion venom activity on fibroblast cell lysis. *Journal of Ethnopharmacology*. 103, pp. 201–207.
- Unsicker, K. and Krieglstein, K. (2002). TGF-betas and their roles in the regulation of neuron survival. Advance in Experimental Medicine and Biology. 513, 353-374.
- Vajrabhaya, L., and Korsuwannawong, S. (2018). Cytotoxicity evaluation of Thai herb usinf tetrazolium (MTT) and sulforhodamine B (SRB) assays. *Journal* of Analytical Science and Technology. 9 (15), 1-8.
- van den Berg, R.A., Hoefsloot, H C.J., Westerhuis, J.A., Smilde, A.K, and van der Werf, M.J. (2006). Centering, scaling, and transformations: improving the biological information content of metabolomics data. *BMC Genomics*, 7, 142.
- van der Kooy, F., Maltese, F., Hae Choi, Y., Kyong Kim, H., Verpoorte, R. (2009). Quality control of herbal material and phytopharmaceuticals with MS and NMR based metabolic fingerprinting. *Planta Medica*, *75*(07), 763-775. doi: 10.1055/s-0029-1185450

- van Meer, G., Voelker, D., Feigenson, G. (2008). Membrane lipids: where they are and how they behave. *Nature Reviews Molecular Cell Biology*, *9*(2), 112-124. doi: 10.1038/nrm2330
- Vargas-Caraveo, A., Sayd, A., Maus, S., Caso, J., Madrigal, J., García-Bueno, B., & Leza, J. (2017). Lipopolysaccharide enters the rat brain by a lipoproteinmediated transport mechanism in physiological conditions. *Scientific Reports*, 7(1), 13113.
- Verma, A., Kumar, I., Verma, N., Aggarwal, P., Ojha, R. (2016). Magnetic resonance spectroscopy- Revisiting the biochemical and molecular milieu of brain tumors. *BBA Clinical*. 5, 170-178.
- Verpoorte, R., Choi, Y., and Choi, H. (2009). Botanicals and NMR-based metabolomics: A perfect holistic match. *Planta Medica*, *75*(04), S-2
- Vibulsreth, S., Hefti, F., Ginsberg, M.D., Dietrich, W.D., Busto, R. (1987). Astrocytes protect cultured neurons from degeneration induced by anoxia. *Brain Research*, 422(2), 303–311.
- Villas-Boas, S.G., Rasmussen, S. and Lane, G.A. (2005). Metabolomics or metabolite profiles? *Trends in Biotechnology*. 23(8), 385-386.
- Walsh, R.N. and Cummins, R.A. (1976). The Open-field test: a critical review. *Psychological Bulletin.* 83(3), 482–504.
- Walz, A., Peveri, P., Aschauer, H., Baggiolini, M. (1987). Purification and amino acid sequencing of NAF, a novel neutrophil-activating factor produced by monocytes. *Biochemical and Biophysical Research Communications*. 149(2), 755–761.
- Walz, W. and Mukerji, S. (1988). Lactate production and release in cultured astrocytes. *Neuroscience Letters*, 86(3), 296–300.
- Wang, H., Vidyadaran, S., Mohd Moklas, M., Baharuldin, M. (2017). Inhibitory activity of *Ficus deltoidea* var. trengganuensis aqueous extract on lipopolysaccharide-induced TNF-α production from microglia. *Evidence-Based Complementary and Alternative Medicine*, 2017, 1-7. doi: 10.1155/2017/2623163
- Wang, M., Lamers, R.J.A., Korthout, H.A., van Nesselrooij, J.H., Witkamp, R.F., van der Hejiden, R. et al. (2005). Metabolomics in the context of system biology: Bridging traditional Chinese medicine and molecular pharmacology. *Phytotherapy Research*. 19, 173-182.
- Wanichthanarak, K., Fahrmann, J., and Grapov, D. (2015). Genomic, proteomic, and metabolomic data integration strategies. *Biomarker Insights*, 10s4, BMI.S29511.
- Wanikiat, P., Panthong, A., Sujayanon, P., Yoosook, C., Rossi, A.G. and Reutrakul, V. (2008). The anti-inflammatory effects and the inhibition of

neutrophil responsiveness by *Barleria lupulina* and *Clinacanthus nutans* extracts. *Journal of Ethnopharmacology*. 116(2), pp. 234–244.

- Watson, R.R. and Preedy, V.R. (2008). Botanical medicine in clinical practice. *Cambridge: CAB International Cambridge*, p. 819.
- Weckwerth, W. (2003). Metabolomics in system biology. *Annual Review of Plant Biology*. 54, 669-689.
- Wehrens R, Franceschi P. (2012). Meta-statistics for variable selection: The R Package. BioMark. J Stat Softw. 51(10), 1-18.
- Weiss, R. and Kim, K. (2011). Metabolomics in the study of kidney diseases. *Nature Reviews Nephrology*, 8(1), 22-33.
- Westerhuis, A., Hoefsloot, H.C.J., Smit, S., Vis, D.J., Smilde, A.K., van Velzen, E.J.J et al. (2008). Assessment of PLSDA cross validation. *Metabolomics*. 4(1), 81–89
- Westerhuis, J., van Velzen, E., Hoefsloot, H., and Smilde, A. (2009). Multivariate paired data analysis: multilevel PLSDA versus OPLSDA. *Metabolomics*, 6(1), 119-128. doi: 10.1007/s11306-009-0185-z
- Wishart, D. (2008). Metabolomics: applications to food science and nutrition research. *Trends in Food Science and Technology*, *19*(9), 482-493. doi: 10.1016/j.tifs.2008.03.003
- Wishart, D., Tzur, D., Knox, C., Eisner, R., Guo, A., and Young, N. et al. (2007). HMDB: the Human Metabolome Database. *Nucleic Acids Research*, *35*(Database), D521-D526.
- Wold, S. (2001). Personal memories of the early PLS development. *Chemometrics* and *Intelligent Laboratory System*, 58(2), 83–84.
- Wolfender, J. L., Bohni, N., Ndjoko-loset, K., and Edison, A. S. (2013). Advanced spectroscopic detectors for identification and quantification: nuclear magnetic resonance. *Liquid Chromatography: Fundamentals and Instrumentation* eds S. Fanali, P. R. Haddad, C. F. Poole, P. Schoenmakers, and D. Lloyd. (Amsterdam: Elsevier), 349–384.
- Wong, F.C., Yong, A.L., Ting, E.P.S et al., (2014). Antioxidant, metal chelating, anti-glucosidase activities and phytochemical analysis of selected tropical medicinal plants. *Iranian Journal of Pharmaceutical Research*. 13(4), pp. 1407–1413.
- World Health Organization (WHO) Dept. of Mental Health and Substance Abuse. (2006). Neurological disorders: a public health approach.19; 27-40.
- World Health Organization (WHO) Essential Medicine and Pharmaceutical Policies Quality Assurance ans Safety: Medicines Health System and Services (2008). Pharmaceuticals: Restrictions in use and availability.

- Wu, C., Kim, H., van Wezel, G., Choi, Y. (2015). Metabolomics in the natural products field – a gateway to novel antibiotics. *Drug Discovery Today: Technologies*, 13, 11-17. doi: 10.1016/j.ddtec.2015.01.004
- Wu, J.-S., Kao, M.-H., Tsai, H.-D. et al., (2017). *Clinacanthus nutans* mitigates neuronal apoptosis and ischemic brain damage through augmenting the C/EBPβ-driven PPAR-γ transcription. *Molecular Neurobiology*, pp. 1–14.
- Xia, J., Sinelnikov, I.V., Han, B., Wishart, D.S. (2015). MetaboAnalyst 3.0- making metabolomics more meaningful. *Nucleic Acids Research*, 43(1), 251-257.
- Xia, J., Psychogios, N., Young, N., Wishart DS. (2009). MetaboAnalyst: a web server for metabolomic data analysis and interpretation. *Nucleic Acids Research*. 37, 652–660.
- Xiao, Q., Wang, C., Li, J., Hou, Q., Li, J., Ma, J., et al. (2010). *Ginkgolide B* protects hippocampal neurons from apoptosis induced by beta-amyloid 25–35 partly via up-regulation of brain-derived neurotrophic factor. *European. Journal of. Pharmacology*. 647(1-3), 48–54. 10.1016/j.ejphar.2010.08.002
- Yahaya, T.A., Okhale, S.E., Adeola, S.O. (2013). Neuropharmacological effects of standardized aqueous stem bark extract of *Parkia biglobossa* in Wistar rats. *Avicenna Journal of Phytomedicine*, 4(1), 59-71
- Yang, J., Karr, J., Watrous, J., and Dorrestein, P. (2011). Integrating '-omics' and natural product discovery platforms to investigate metabolic exchange in microbiomes. *Current Opinion in Chemical Biology*, 15(1), 79-87.
- Yong, Y., Tan, J., Teh, S., Mah, S., Ee, G., Chiong, H. and Ahmad, Z. (2013). *Clinacanthus nutans* extracts are antioxidant with antiproliferative effect on cultured human cancer cell lines. *Evidence-Based Complementary and Alternative Medicine*, 2013, 1-8.
- Yuliana, N., Jahangir, M., Verpoorte, R., & Choi, Y. (2013). Metabolomics for the rapid dereplication of bioactive compounds from natural sources. *Phytochemistry Reviews*, *12*(2), 293-304.
- Yuliana, N., Khatib, A., Choi, Y., and Verpoorte, R. (2011). Metabolomics for bioactivity assessment of natural products. *Phytotherapy Research*. 25(2):157-69.
- Yumi, Z., Hashim, H.Y., Gill C.I.R., Latimer, C. et al (2016). *Colorectal Cancer-From Pathogenesis to Treatment*. Studies of Malaysian plants in prevention and treatment of colorectal cancer. InTech. Rijeka, Croatia, 381-393.
- Zähringer, U., Lindner, B., Rietschel, E.T. (1999). Chemical structure of lipid A. Recent advances in structrural analysis of a biologically active molecules. *Endotoxin in Health and Disease.*, Marcel Dekker, New York, 93-114.

- Zamboni, N., Saghatelian, A., & Patti, G. (2015). Defining the metabolome: size, flux, and regulation. *Molecular Cell*, 58(4), 699-706. doi: 10.1016/j.molcel.2015.04.021
- Zandi, P.P., Anthony, J.C., Hayden, K.M., et al. (2002). Reduced incidence of AD with NSAID but Not H2 Receptor Antagonists: The Cache County Study. *Neurology*. 59(6), 880–886.
- Zhang, A. Sun, H., Wang, P., Han, Ying, Wang, X. (2012). Modern analytical techniques in metabolomics analysis. *The Analyst.* 137(2), 293-300.
- Zhang, A., Sun, H., Yan, G., Yuan, Y., Han, Y., Wang, X. (2013). Metabolomics study of type 2 diabetes using ultra-performance LC-ESI/quadrupole-TOF high-definition MS coupled with pattern recognition methods. *Journal of Physiology and Biochemistry*, 70(1), 117-128.
- Zhang, J., Lei, T., Chen, X., Peng, Y., Long, H., Zhou, L. et al. (2009). Resistin Up-Regulates COX-2 Expression via TAK1-IKK-NF-κB Signaling Pathway. *Inflammation*. 33(1), 25-33.
- Zhang, M., Jiang, S.K., Tian, Z.L., Wang, M., Zhao, R., Wang, L.L. et al. (2015). CB2R orchestrates fibrogenesis through regulation of inflammatory response during the repair of skeletal muscle contusion, *International Journal of Clinical Experimental Pathology*. 8(4), 3491–3502.
- Zhang, W., Wang, T., Qin, L., Gao, H.M., Wilson, B., Ali, S.F. et al. (2004). Neuroprotective effect of dextromethorphan in the MPTP Parkinson's disease model: role of NADPH oxidase. *The FASEB Journal*. 18(3), 589-591.
- Zhu, J., Motejlek, K., Wang, D., Zang, K, Schmidt, A., Reichardt, L.F. (2002). Beta8 integrins are required for vascular morphogenesis in mouse embryos Development. 129(2), 2891-2903
- Zielasek, J. and Hartung, H. (1996). Molecular mechanisms of microglial activation. *Advances in Neuroimmunology*, 6(2), pp.191-222.
- Zulkipli, I., Rajabalaya, R., Idris, A., Sulaiman, N., and David, S. (2017). *Clinacanthus nutans*: a review on ethnomedicinal uses, chemical constituents and pharmacological properties. *Pharmaceutical Biology*, *55*(1), 1093-1113.