



***ANTIOXIDANT AND ANTI-AGEING ACTIVITIES OF COCOA BEAN
EXTRACT FOR COSMECEUTICAL USE***

NORLIZA BINTI ABDUL WAHAB

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By

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**Thesis Submitted to the School of Graduate Studies, Universiti
Putra Malaysia, in Fulfilment of the Requirements for the Degree of
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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

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September 2019

Chairman : Professor Russly Abdul Rahman, PhD
Faculty : Halal Products Research Institute

Theobroma cacao which is best-known as 'Food of the God' by an ancient Greek, belongs to *Sterculiaceae* family. It is an indigenous plant from the tropical region of America which culture was further extended to equatorial areas of Africa and Asia. The goodness of *T.cacao* bean against cancer, cardiovascular-related diseases, certain neurodegenerative disorders and premature ageing have been often reported. However, information pertaining to antioxidant capacity and anti-ageing activity of potential Malaysian cocoa bean extract (CBE) for cosmeceutical use is limited.

Therefore, the main objective was to determine the antioxidant capacity and anti-ageing activity of two types of cocoa beans' clones, known as PBC140 and PBC123. Response surface methodology employing a central composite design throughout twenty experiments was conducted for the optimization of extraction condition, focusing on ethanol (30-80%, v/v), temperature (30-60°C) and extraction time (60-240 min). Optimal ethanol concentration (70% [v/v]) and extraction temperature (50°C) have resulted in the highest total phenolic (377 and 231 mg GAE/g DW, respectively) and total flavonoid content (73 and 63 mg RE/g DW, respectively). For antioxidant capacity, PBC140 was significantly higher ($p>0.05$) than PBC123 in term of 2,2-Diphenyl-1-picrylhydrazyl scavenging whereas ferric reducing antioxidant power and β -carotene leanoleate bleaching assays showed no significant different ($p>0.05$). PBC140 was higher than PBC123 but with no significant different ($p>0.05$) in both anti-collagenase and anti-elastase when conducted on Ac-PLG-[2-mercapto-4-methyl-pentanoyl]-LG-OC₂H₅ and MeOSuc-Ala-Ala-Pro-Val-pNA substrates, respectively. PBC140 was cytotoxic at IC₅₀=1100 μ g/mL on 1×10^5 human dermal fibroblast (HDFa) cells/well at sixth passage or the highest average HDFa cell viability level. The extracted ribonucleic acid of HDFa treated with 1×10^3 μ g/mL CBE significantly ($p<0.05$) downregulated the

ultraviolet-induced matrix metalloproteinase-1 by 25-fold relative to calibrator. Ultraviolet A ratio for *in vitro* photoprotective effect of 0.1% (w/v) CBE formulation indicated 0.553 from the Boots Star Rating (Good). Intervention of approximately 500 mg 0.1% (w/v) CBE formulation on 20 human subjects aged between 30 to 46 years for 2 months duration in the *in vivo* skin efficacy studies recorded significant ($p < 0.05$) percent changes of skin texture parameters, namely volume (-40%), energy (46%), contrast (-18%) and variance (-21%). The skin elasticity parameter for CBE formulation recorded significant ($p < 0.05$) increment of ten times compared to placebo group. To conclude, Malaysian CBE is a potential active material due to the encouraging results of antioxidant capacity, *in vitro* anti-ageing, safety, and *in vivo* skin efficacy that meet the primary objective of producing harmless yet natural cosmeceutical with significant skin improvement.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

ANTIOKSIDAN DAN AKTIVITI ANTI-PENUAAN EKSTRAK BIJI KOKO UNTUK KEGUNAAN KOSMESEUTIKAL

Oleh

NORLIZA BINTI ABDUL WAHAB

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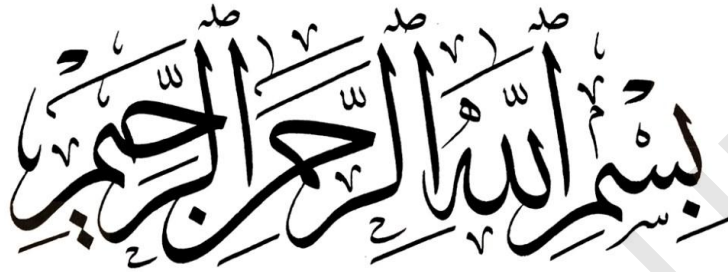
Pengerusi : Professor Russly Abdul Rahman, PhD
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Theobroma cacao tergolong dalam keluarga *Sterculiaceae* dan dikenali sebagai 'Food of the God' oleh Yunani kuno. Ia adalah tumbuhan asli dari rantau tropika Amerika dan penanamannya telah diperluaskan ke kawasan khatulistiwa benua Afrika dan Asia. Kebaikan *T.cacao* dalam pencegahan penyakit kanser, kardiovaskular, gangguan *neurodegenerative*, dan penuaan pramatang telah sering dilaporkan. Walau bagaimanapun, maklumat berkaitan dengan kapasiti antioksidan dan aktiviti anti-penuaan ekstrak biji koko klon terpilih Malaysia (CBE) untuk kegunaan kosmeseutikal adalah terhad.

Oleh itu, objektif utama penyelidikan ialah bagi menentukan kapasiti antioksidan dan aktiviti anti-penuaan bagi dua jenis klon koko iaitu PBC140 dan PBC123. Kaedah *response surface methodology* menggunakan *central composite design* yang melibatkan dua puluh eksperimen telah dijalankan bagi mengoptimumkan keadaan pengekstrakan iaitu peratusan etanol (30-80 %, i/i), suhu (30-60°C) dan masa (60-240 min). Kandungan fenolik yang tertinggi bagi PBC140 dan PBC123 masing-masing sebanyak 377 dan 231 mg GAE/g DW manakala kandungan flavonoid iaitu 73 dan 63 mg RE/g DW telah dihasilkan daripada peratusan etanol (70% [i/i]) dan suhu pengekstrakan (50°C) yang optimum. Bagi kapasiti antioksidan, PBC140 lebih tinggi daripada PBC123 untuk ujian *2,2-Diphenyl-1-picrylhydrazyl* manakala masing-masing klon tidak menunjukkan perbezaan signifikan ($p > 0.05$) untuk ujian *ferric reducing antioxidant power* serta *β -carotene leanoleate bleaching*. PBC140 lebih tinggi daripada PBC123 namun tiada perbezaan signifikan ($p > 0.05$) bagi kedua-dua ujian anti-kolagenase ke atas substrat *Ac-PLG-[2-mercapto-4-methyl-pentanoyl]-LG-OC₂H₅* dan anti-elastase ke atas substrat *MeOSuc-Ala-Ala-Pro-Val-pNA*. PBC140 bersifat sitotoksik pada $IC_{50} = 1100 \mu\text{g/mL}$ ke atas 1×10^5 sel *human dermal fibroblast* (HDFa)/well iaitu pada transaksi sel yang keenam di mana sel HDFa berada pada tahap yang paling progresif. Ekstrak asid

ribonukleik daripada HDFa pada dos 1×10^3 $\mu\text{g/mL}$ CBE telah menunjukkan rencatan ekspresi ke atas gen *matrix metalloproteinase 1* daripada rangsangan sinar ultraungu, yang signifikan ($p < 0.05$) iaitu 25 kali ganda relatif kepada calibrator. Penarafan *Boots Star* formulasi 0.1% (b/i) CBE bagi kesan fotoprotektif daripada sinar ultraungu A ialah 0.553 (Baik). Sejumlah 500 mg 0.1% (b/i) formulasi CBE yang diaplikasikan ke atas 20 subjek manusia antara 30 hingga 46 tahun selama 2 bulan bagi ujian *in vivo*, telah merekodkan peratus perubahan tekstur kulit yang signifikan ($p < 0.05$) dari segi *volume* (-40%), *energy* (46%), *contrast* (-18%) dan *variance* (-21%) serta purata kenaikan yang signifikan ($p < 0.05$) bagi parameter keanjalan kulit iaitu sepuluh kali ganda berbanding kumpulan plasebo. Kesimpulannya, hasil ujian keupayaan antioksidan, anti-penuaan secara *in vitro*, aspek keselamatan, dan keberkesanan kulit secara *in vivo* memenuhi objektif utama kajian iaitu potensi CBE klon Malaysia untuk dibangunkan sebagai bahan aktif semulajadi di dalam penghasilan kosmeseutikal yang selamat serta berkebolehan memperbaiki tekstur kulit dengan berkesan.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Russly bin Abdul Rahman, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Amin bin Ismail, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Shuhaimi bin Mustafa, PhD

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Puziah binti Hashim, PhD

Senior Lecturer
Halal Products Research Institute
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

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Committee: _____

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Committee: _____

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LIST OF ABBREVIATIONS

A	Water to Skin Ratio Coefficient
Ac	Mean OD of Control
At	Mean OD of Tested Extract
ACD	ASEAN Cosmetic Directive
AOC	Antioxidant Capacity
APC	Aerobic Plate Count
AR	Allergic Reactions
b	Estimated Regression Coefficient
BHA	tert-butyl-4-hydroxy anisole
BHT	2,6-di-tert-butyl-p-hydroxy-toluene
Brij™ 35	Nonionic Polyoxyethylene Surfactant
CAGR	Compound Annual Growth Rate
CBE	Cocoa Bean Extract
CCD	Central Composite Design
CPB	Cocoa Pod Borer
d0	Variation in Concentration
d1	Space Travelled
DPPH	2,2-Diphenyl-1-picrylhydrazyl
E	PCR Efficiency
FD&C	Food, Drugs, and Cosmetic Act
FRAP	Ferric Reducing Antioxidant Power
GAE	Gallic Acid Equivalents
HD	High Dose of CBE Sample
HDFa	Human Dermal Fibroblasts, adult
JAKIM	Jabatan Kemajuan Islam Malaysia

JKEUPM Malaysia	Jawatankuasa Etika Universiti Putra
/	Path Length of Light Through the Sample in cm
MCB	Malaysian Cocoa Board
MMP	Matrix Metalloproteinase
MMPLO	Ac-PLG-[2-Mercapto-4-Methyl- Pentanoyl]-LG-OC ₂ H ₅
MMP-1	Matrix Metalloproteinase-1
MMP-12	Matrix Metalloproteinase-12
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5- diphenyl tetrazoliumbromide MTT
MYR	Malaysian Ringgit
M/V	Solid to Liquid Ratio
NDGA	Nordihydroguaiaretic acid
NPRA	National Pharmaceutical Regulatory Agency
PIF	Product Information File
R	Residual Deformation at the End of Measuring Cycle
RE	Rutin Equivalents
RSM	Response Surface Methodology
SC	Stratum Corneum
SELS	Surface Evaluation of Living Skin
SEr	Skin Roughness
SEsc	Skin Scaliness
SEsm	Skin Smoothness
SEw	Skin Wrinkles
<i>T.cacao</i>	<i>Theobroma cacao</i>
TBHQ	Tertiary Butylhydroquinone

TPC	Total Phenolic Content
TFC	Total Flavonoid Content
TEWL	Transepidermal Water Loss
UV	Ultraviolet
VSD	Vascular-streak Dieback
YMC	Yeast and Mould Count
β -CB	β -carotene-linoleate bleaching
IIR	Immediate Irritation Reactions
IR	Irritation Reactions
3D	Three Dimensional

CHAPTER 1

GENERAL INTRODUCTION

1.1 Background of the Study

Cosmeceuticals are topical cosmetic-pharmaceutical hybrids mainly to promote the beauty through permeation of active ingredients into the dermis skin layer. Cosmeceuticals constitute of materials that have significant effect on skin's biological function. The global cosmeceuticals market which was estimated at USD46.93 billion in 2017 has been forecasted to reach USD80.36 billion by 2023 at 9.83% compound annual growth rate (CAGR) within the predicted period (2018 – 2023). Incorporation of polyphenol antioxidants has boosted the sales of anti-ageing cosmeceuticals in the overall cosmetic product market. Moreover, emergence of organic and halal ingredients has significantly affected the demand for cosmeceutical products. This is to indicate the huge margin of sales, thus offering endless opportunity in the exploration of natural-based cosmeceutical.

In Malaysia, assurance of cosmetic/cosmeceutical product's safety and non-hazardous to consumers as required by the Act 368 (Sales of Drug Act, 1952), are subjected to the regulations of National Pharmaceutical Regulatory Agency (NPRA) which is indispensable in the application of Malaysian Standard MS 2200-1:2008-Islamic Consumer Goods Part 1: Cosmetic and Personal Care-General Guidelines (MS 2200-1:2008, 2008). However, despite of the rule, occurrences of adulteration in cosmetics due to utilization of toxic synthetic chemicals, i.e., mercury, hydroquinone, tretinoin, methylene chloride and synthetic antioxidants such as 2,6-di-tert-butyl-p-hydroxy-toluene (BHT) and tert-butyl-4-hydroxy anisole (BHA), among others, have been relentlessly reported as the main cause of adverse health risks and severe skin injuries (Atta et al., 2017; Moein et al., 2008; Utusan Malaysia, 25th March 2017; News Straits Times, 25th Disember 2016; Harian Metro, 29th May 2010). The danger lurked due to various hazardous cosmetic products which were freely marketed in Malaysia is shown in Figure 1.1. In addition, a recent report entitled *NPRA senaraikan 14 produk kosmetik beraacun* which was published in Berita Harian dated on 9th October 2018, indicating serious deficiency with regard to safety enforcement of cosmetic products in Malaysia.



Figure 1.1: Inadequate enforcement pertaining to the cosmetics safety regulation in Malaysia (Source: *Harian Metro*, 29th May 2010)

Nowadays, incorporation of plant materials in a cosmetic formulation is considerably desirable due to public fears over various hazardous synthetic chemicals present in anti-ageing cosmeceuticals. There have been several reports concerning application of natural phenolics for anti-ageing effects as an alternative antioxidant against oxidative damages. In doing so, recoveries of bioactives at the highest yield and quality have been regularly conducted in the optimization on extraction conditions of plant materials. Many studies showed that response surface methodology (RSM) which design of experiment accounts for possible interaction effects between variables, have significantly maximized the extraction yield with targeted components and antioxidant power at optimized variables condition (Spigno et al., 2007; Tan et al., 2014). Moreover, for this tool, the mathematical model helps in providing an apparent visual regarding the effects of various extraction factors and also to view the region wherein the extraction is optimized (Bezerra et al., 2008).

Due to chronic UV exposure, denaturation of collagen and elastin have often attributed to excessive formation of collagenase and elastase in the dermis skin layer. This has given rise to the formation of wrinkles whereas severe hyperpigmentation which is another type of skin photoageing due to over production of melanin in animal tissues, can be effectively prevented through anti-tyrosinase catalytic treatment. Significant correlations between antioxidant capacity and anti-ageing properties in terms of anti-collagenase, anti-elastase and anti-tyrosinase activities, have been found in many phytochemical studies that reflected the reactive responses of antioxidant phenolics (Dudonné et al., 2009). Overall, collagenase, elastase and tyrosinase activities can be

suppressed through preventive action against oxidative damages, which are driven by antioxidant capacity property of the extract material. It helps by preventing the initiation and extension of oxidizing chain reaction metabolic processes disorder (Chanda and Nagani, 2010).

Phenolic compounds demonstrate diverse range of medicinal and pharmacological benefits particularly on antioxidant, anti-microbial, anti-inflammatory, anti-cancer and cardioprotective effects. It is thus suggested that these constituents can eliminate the shelf life of cancer cells as well as healthy cells due to cytotoxicity effect of many phenolic compounds (Balasundram et al., 2006). Fibroblast cell lines have been regularly conducted in the *in vitro* cytotoxicity test due to their presences which can be found mainly in the dermis skin layer. Lower doses of phenolic compounds are sufficient for the antioxidant attribute whereas increasing doses may cause pro-oxidant effects (Amudha et al., 2011). For this reason, cytotoxicity assessment in a healthy fibroblast cell has been conducted particularly to avoid toxicological risks that were often caused by the plant concentrates (Dufrane et al., 2001). Moreover, *in vitro* inhibition of matrix metalloproteinase-1 (MMP-1) expression in UV-irradiated fibroblast cell portrays the substantial function of phenolic components for the prevention of accelerated ageing due to external insults in the dermis skin layer.

Potential results of antioxidant compounds showed from photoprotective and inhibition of destructive tyrosinase, collagenase and elastase of plant materials including beans/seeds, are the desirable anti-ageing property own by cosmeceutical products. Plant material of *Theobroma cacao* bean which is classified under the genus *Theobroma* and belongs to the *Sterculiaceae* family, has been blessed with major bioactive components such as epicatechin, catechin, and methylxanthines (Scapagnini et al., 2014). In Malaysia, potential cocoa clones with premium quality, namely MCBC1, MCBC4, MCBC5, MCBC6, KKM1, KKM5, KKM22, PBC140, PBC123 and QH1003, have been developed in Malaysian Cocoa Board research and development stations at Jengka, Pahang and Bagan Datoh, Perak. Antioxidant benefits of cocoa bean have been considerably reported in consumable products, however, data on polyphenolic components of the Malaysian potential clone extracts for cosmeceutical use have not been thoroughly investigated. Furthermore, development of phenolics-rich cocoa extract cosmeceutical may contribute to the merit in assessing the product's feasibility study.

1.2 Problem Statement

According to Jahanban-Esfahlan et al. (2019), Truong et al. (2019), Plaza et al. (2017) and Altemimi et al. (2017), solvents for phenolic compounds extractions from plant materials are mainly ethanol, water, acetone, methanol, and their water mixtures, either with or without acid. With regard to safety concern, water and ethanol are more desirable due to their non-toxic property compared to other diluents (Liyana-Pathirana and Shahidi, 2005). However, extraction

efficiency informations of water and ethanol mediums on recoveries of total phenolic content (TPC) and total flavonoid content (TFC) of the ten (10) potential Malaysian clones are limited. In addition, it is questionable whether the amount of phenolic compounds extracted could be also affected by the different type of clones. Having said that, according to the Two-way ANOVA, it is hypothesized that; H_0 = Extraction medium and cocoa clone do not affect the total phenolic (TPC) and total flavonoid content (TFC) in mg GAE/g DW and mg RE/g DW, respectively ($p>0.05$), whereas H_1 = At least one of the factors gives significant amount of the TPC and TFC ($p<0.05$).

Experimental design suggested by Response Surface Methodology (RSM) in many researches has been proven as an effective appliance in providing the highest yield of bioactive compounds in diverse plant extracts (Raissi, 2009; Tan et al., 2014). Many types of extraction solvents with several concentrations were proposed to obtain the required bioactives. However, informations pertaining to optimization of extraction conditions by using RSM on *Theobroma cacao* bean extracts of Malaysian potential clones are still limited. Moreover, subsequent analyses involving characterization of their cosmeceutical properties, namely antioxidant capacity such as DPPH radical scavenging, ferric reducing power, and β -carotene-linoleate bleaching along with anti-ageing activity in terms of tyrosinase, collagenase, and elastase inhibition, leads to this investigation since no wide research has ever been done on the optimized cocoa bean crude extracts of Malaysian potential clones.

There were several reports regarding natural products to play as a substantial character on the downregulation of MMPs in UV-irradiated cells (Chae et al., 2011; Zhang et al., 2009; Mohan et al., 2016). Since focus of Malaysian potential clones have been on the development of food-based products, thus cocoa-based cosmeceutical-related study in terms of evaluation of their harmful effects or cytotoxicity degree as well as photoprotective property against MMP-1 production on UV-irradiated human dermal fibroblasts cells for anti-ageing property, are yet to discover. Even though many researchers have worked on antioxidants and anti-ageing of *Theobroma cacao*, very few researches were conducted on skin benefits pertaining to intervention of a cosmeceutical product formulated from bean extract of potential Malaysian cocoa clone that reflects the limitation data of natural cosmeceutical product development incorporated with such extract. In term of novelty, though there are similar works, but the present bean extracts are specifically derived from potential Malaysian cocoa clones with superior quality that has never been before assessed for their antioxidant and anti-ageing activities for cosmeceutical use. This study aims to investigate overall potential of Malaysian cocoa bean extracts for their antioxidant capacity and anti-ageing activities in term of photoprotective effect against matrix metalloproteinase-1 (MMP-1) gene expression of human dermal fibroblast cells *in vitro*. Furthermore, *in vivo* application of cocoa bean extract formulation involving twenty (20) human volunteers between 30 to 46 years of age for 2 months of assessment period, are expected to gain skin benefits particularly in a prevention of destructive ultraviolet (UV) light over skin's connective tissues such as collagen and elastin fibres which lie in the deeper layer of skin dermis.

1.3 Objectives

The objectives of this research are:

1. To determine the extraction conditions of optimized ethanol concentrations, extraction time and temperature on total phenolic content (TPC) and total flavonoid content (TFC) of CBEs.
2. To determine antioxidant capacity of CBEs, namely radical scavenging ability, ferric reducing power, and neutralization of linoleate-free radical, whereas *in vitro* anti-ageing activity include anti-collagenase, anti-elastase and anti-tyrosinase activities.
3. To determine the cytotoxicity of CBE towards human dermal fibroblast cell (HDF) and its protective effect on the expression of MMP-1.
4. To develop a stable, safe and efficient CBE formulation with functional *in vitro* photoprotective and non-invasive *in vivo* assessment of skin structure for anti-ageing.

REFERENCES

- Abramovic, H., & Abram, V. (2006). Effect of added rosemary extract on oxidative stability of *Camelina sativa* oil. *Acta Agriculturae Slovenica*, 87, 255-261.
- Act 368 – Sale of Drugs Act 1952 (Revised 1989). Incorporating Latest Amendment – Act A1084/2000.
- Adamson, G. E., Lazarus, S. A., Mitchell, A. E., Prior, R. L., Cao, G., Jacobs, P. H., Kremers, B. G., Hammerstone, J. F., Rucker, R. B., & Ritter, K. A. (1999). HPLC method for the quantification of procyanidins in cocoa and chocolate samples and correlation to total antioxidant capacity. *Journal of Agricultural and Food Chemistry*, 47, 4184-4188.
- Adhami, V. M., & Mukhtar, H. (2006). Polyphenols from green tea and pomegranate for prevention of prostate cancer. *Free Radical Research*, 40, 1095-1104.
- Adriana, C., & Cornelia, P. (2013). The influence of the extraction solvent on the polyphenol content determination in cocoa products. *Natural Resources and Sustainable Development*, pp. 115-120.
- Agati, G., Azzarello, E., Pollastri, S., & Tattini, M. (2012). Flavonoids as antioxidants in plants: location and functional significance. *Plant Science*, 196, 67-76.
- Akhtar, N., Zaman, S. U., Khan, B. A., Amir, M. N., & Ebrahimzadeh, M. A. (2011). Calendula extract: Effects on mechanical parameters of human skin. *Acta Poloniae Pharmaceutica – Drug Research*, 68, 693-701.
- Al-Olayan, E. M., El-Khadragy, M. F., Aref, A. M., Othman, M. S., Kassab, R. B., & Moneim, A. E. A. (2014). The potential protective effect of *Physalis peruviana* L. against carbon tetrachloride-induced hepatotoxicity in rats is mediated by suppression of oxidative stress and downregulation of MMP-9 expression. *Oxidative Medicine and Cellular Longevity*, 2014, 1-12.
- Aldag, C., Teixeira, D. N., Leventhal, P. S. (2016). Skin rejuvenation using cosmetic products containing growth factors, cytokines, and matrikines: a review of the literature. *Clinical, Cosmetic and Investigational Dermatology*, 9, 411-419.
- Ali, A., Akhtar, N., Khan, B. A., Khan, M. S., Rasul, A., Khalid, N., Waseem, K., Mahmood, T., & Ali, L. (2012). *Acacia nilotica*: A plant of multipurpose medicinal uses. *Journal of Medicinal Plants Research*, 6, 1492-1496.
- Ali, A., Naveed, A., & Khan, M. S. (2013). Enhancement of human cheek skin texture by *Acacia nilotica* extract cream. *Tropical Journal of Pharmaceutical Research*, 12, 323-327.
- Ali, S. M., & Yosipovitch, G. (2013). Skin pH: from basic science to basic skin care. *Acta Dermato-Venereologica*, 93, 261-267.

- Ali, A., Akhtar, N., & Chowdhary, F. (2014). Enhancement of human skin facial revitalization by moringa leaf extract cream. *Postepy Dermatologii I Alergologii*, 2, 71-76.
- Ali, F., Ranneh, Y., Ismail, A., & Esa, N. M. (2015). Identification of phenolic compounds in polyphenols-rich extract of Malaysian cocoa powder using the HPLC-UV-ESI-MS/MS and probing their antioxidant properties. *Journal of Food Science and Technology*, 52, 2103-2111.
- Altemimi, A., Lakhssassi, N., Baharlouei, A., Watson, D. G., & Lightfoot, D. A. (2017). Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant extracts. *Plants*, 6, 1-23.
- Amarowicz, R., Pegg, R. B., Rahimi-Moghaddam, P., Barl, B., & Weil, J. A. (2004). Free radical scavenging capacity and antioxidant activity of selected plant species from the Canadian prairies. *Food Chemistry*, 84, 551-562.
- Amin, I., Koh, B. K., & Asmah, R. (2004). Effect of cacao liquor extract on tumor marker enzymes during chemical hepatocarcinogenesis in rats. *Journal of Medicinal Food*, 7, 7-12.
- Amir, M., Khan, A., Mujeeb, M., Ahmad, M. A., & Siddiqui, N. A. (2011). Phytochemical screening and *in vitro* antioxidant activity of *Jawarish amla* – a poly herbal formulation. *Pharmacognosy Journal*, 3, 54-60.
- Amit, K., & Priyadarsini, K. I. (2011). Free radicals, oxidative stress and importance of antioxidants in human health. *Journal of the History of Medicine and Allied Science*, 1, 53-60.
- Amudha, J., Balasubramani, G., Malathi, V. G., Monga, D., Kranthi, K. R. (2011). Cotton leaf curl virus resistance transgenic with the antisense coat protein gene (AV1). *Current Science*, 101, 300-307.
- An, B. J., Kwak, J. H., Park, J. M., Lee, J. Y., Park, T. S., & Lee, J. T. (2005). Inhibition of enzyme activities and the antiwrinkle effect of polyphenol isolated from the persimmon leaf (*Diospyros kakifolium*) on human skin. *Dermatologic Surgery*, 31, 848-854.
- Anbudhasan, P., Alagarsamy, S., Sivanandham, K., & Satishkumaran, S. (2014). Natural antioxidants and its benefits. *International Journal of Food and Nutritional Sciences*, 3, 225-232.
- Andujar, I., Recio, M. C., Giner, R. M., & Rios, J. L. (2012). Cocoa polyphenols and their potential benefits for human health. *Oxidative Medicine Cell Longevity*, 2012, 1-23.
- Angkatavanich, J., Dahlan, W., Nimmannit, V., Sriprasert, V., & Sulongkood, N. (2009). Development of clay liquid detergent for Islamic cleansing and the stability study. *International Journal of Cosmetic Science*, 31, 131-141.
- Arlorio, M., Bottini, C., Travaglia, F., Locatelli, M., Bordiga, M., Coisson, J. D., Martelli, A., & Tessitore, L. (2009). Protective activity of *Theobroma cacao* L. phenolic extract on AML12 and MLP29 liver cells by preventing apoptosis and inducing autophagy. *Journal of Agricultural*

- and Food Chemistry, 57, 10612-10618.
- Artaria, C., Maramaldi, G., Bonfigli, A., & Rigano, L. (2011). Lifting properties of the alkamide fraction from the fruit husks of *Zanthoxylum bungeanum*. *International Journal of Cosmetic Science*, 33, 328-333.
- Atikah, A. R., Budi, H. S., & Kusumaningsih, T. (2017). Antibacterial effects of 70% ethanol and water extract of cacao beans (*Theobroma cacao* L.) on *Aggregatibacter actinomycetemcomitans*. *Dental Journal (Majalah Kedokteran Gigi)*, 49, 104-109.
- Atta, E. M., Mohamed, N. H., & Abdelgawad, A. A. M. (2017). Antioxidants: An overview on the natural and synthetic types. *European Chemical Bulletin*, 6, 365-375.
- Aware, C., Patil, R., Gaikwad, S., Yadav, S., Bapat, V., & Jadhav, J. (2017). Evaluation of L-dopa, proximate composition with *in vitro* anti-inflammatory and antioxidant activity of *Mucuna macrocarpa* beans: A future drug for Parkinson treatment. *Asian Pacific Journal of Tropical Biomedicine*, 7, 1097-1106.
- Azizah, O., Abbe, M. M. J., Kong, K. W., Amin, I., Nawalyah, A. G., & Ilham, A. (2010). Epicatechin content and antioxidant capacity of cocoa beans from four different countries. *African Journal of Biotechnology*, 9, 1052-1059.
- Azizah, O., Amin, I., Nawalyah, A. G., & Ilham, A. (2007). Antioxidant capacity and phenolic content of cocoa beans. *Food Chemistry*, 100, 1523-1530.
- Azizkhani, M., & Zandi, P. (2009). Effects of some natural antioxidants mixtures on margarine stability. *World Academy of Science Engineering and Technology*, 49, 93-96.
- Badalica-Petrescu, M., Dragan, S., Ranga, F., Fetea, F., & Socaciu, C. (2014). Comparative HPLC-DAD-ESI(+) ms fingerprint and quantification of phenolic and flavonoid composition of aqueous leaf extracts of *Cornusmas* and *Crataegus monogyna*, in relation to their cardiotoxic potential. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 42, 9-18.
- Badrie, N., Bekele, F., Sikora, E., & Sikora, M. (2014). Cocoa agronomy, quality, nutritional, and health aspects. *Critical Reviews in Food Science and Nutrition*, pp. 620-659.
- Bae, J., Choi, J. -S.; Choi, Y. -J., & Kang, Y. -H. (2008). (-)Epigallocatechin gallate hampers collagen destruction and collagenase activation in ultraviolet-B-irradiated human dermal fibroblasts: Involvement of mitogen-activated protein kinase. *Food and Chemical Toxicology*, 46, 1298-1307.
- Balasundram, N., Sundram, K., & Samman, S. (2006). Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. *Food Chemistry*, 99, 191-203.
- Bando, N., Hayashi, H., Wakamatsu, S., Inakuma, T., Miyoshi, M., Nagao, A., Yamauchi, R., & Terao, J. (2004). Participation of singlet oxygen in

- ultraviolet-a-induced lipid peroxidation in mouse skin and its inhibition by dietary β -carotene: an *ex vivo* study. *Free Radical Biology and Medicine*, 37, 1854-1863.
- Basavaraj, K. N. (2017). Development of cosmeceuticals. *World Journal of Pharmacy and Pharmaceutical Sciences*, 6, 643-691.
- Basketter, D. A., Scholes, E. W., & Kimber, I. (1994). The performance of the local lymph node assay with chemicals identified as contact allergens in the human maximization test. *Food and Chemical Toxicology*, 32, 543-547.
- Basketter, D. A., Chamberlain, M., Griffiths, H. A., Rowson, M., Whittle, E., & York, M. (1997). The classification of skin irritants by human patch test. *Food and Chemical Toxicology*, 35, 845-852.
- Basketter, D., Jirova, D., & Kandárova, H. (2012). Review of skin irritation/corrosion hazards on the basis of human data: a regulatory perspective. *Interdisciplinary Toxicology*, 5, 98-104.
- Bassani, D. C., Nunes, D. S., & Granato, D. (2014). Optimization of phenolics and flavonoids extraction conditions and antioxidant activity of roasted yerba-mate leaves (*Ilex paraguariensis* A. St.-Hil., Aquifoliaceae) using response surface methodology). *Annals of the Brazilian Academy of Sciences*, 86, 1-11.
- Batubara, I., Darusman, L. K., Mitsunaga, T., Aoki, H., Rahminiwati, M., Djauhari, E., & Yamauchi, K. (2011). Flavonoid from *Intsia palembanica* as skin whitening agent. *Journal of Biological Sciences*, 11, 475-480.
- Baumann, L. (2007). Botanical ingredients in cosmeceuticals. *Journal of Drugs in Dermatology*, 6, 1084-1088.
- Baumann, L. S. (2007). Less-known Cosmeceuticals. *Dermatologic Therapy*, 20, 339-342.
- Beelik, A. (1956). Kojic acid. *Advances in Carbohydrate Chemistry*, 11, 145-183.
- Bella, J., Eaton, M., Brodsky, B., & Berman, H. M. (1994). Crystal and molecular structure of a collagen-like peptide at 1.9 Å resolution. *Science*, 266, 75-81.
- Behera, S. K., Meena, H., Chakraborty, S., & Meikap, B. C. (2018). Application of response surface methodology (RSM) for optimization of leaching parameters for ash reduction from low-grade coal. *International Journal of Mining Science and Technology*, 28, 621-629.
- Bellad, K. A., Nanjwade, B. K., Kamble, M. S., Srichana, T., & Idris, N. F. (2017). Development of Cosmeceuticals. *World Journal of Pharmacy and Pharmaceutical Sciences*, 6, 643-691.
- Benites, R. S. R., Formagio, A. S. N., Argandoña, E. J. S., & Volobuff, C. R. F. (2015). Contents of constituents and antioxidant activity of seed and pulp extracts of *Annona coriacea* and *Annona sylvatica*. *Brazilian Journal of Biology*, 75, 685-691.

- Benzie, I. F., & Stezo, Y. T. (1999). Total antioxidant capacity of teas by the ferric reducing/ antioxidant power assay. *Journal of Agricultural and Food Chemistry*, 47, 633-636.
- Benzie, I. F. F., & Strain, J. J. (1996). Ferric reducing ability of plasma (FRAP) as a measure of antioxidant power: the FRAP assay. *Analytical Biochemistry*, 239, 70-76.
- Benzie, I. F. F., & Strain, J. J. (1999). Ferric reducing/antioxidant power assay: Direct measure of total antioxidant activity of biological fluids and modified version for simultaneous measurement of total antioxidant power and ascorbic acid concentration. *Methods in Enzymology*, 299, 15-27.
- Beringhs, A. O., Dalmina, M., Creczynski-Pasa, T. B., & Sonaglio, D. (2015). Response surface methodology IV-optimal design applied to the performance improvement of an RP-HPLC-UV method for the quantification of phenolic acids in *Cecropia glaziovii* products. *Revista Brasileira de Farmacognosia*, 25, 513-521.
- Betton, C. I (2007). The REACH Regulation of the European Union. *Global Regulatory Issues for the Cosmetics Industry*.
- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., & Escalera, L.A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76, 965-977.
- Bondet, V., Brand-Williams, W., & Berset, C. (1997). Kinetics and mechanisms of antioxidant activity using the DPPH• free radical method. *LWT-Food Science and Technology*, 30, 609-615.
- Botterweck, A. A. M., Goldbohm, R. A., Verhagen, H., & Kleinjans, J. (2000). Intake of butylated hydroxyanisole and butylated hydroxytoluene and stomach cancer risk: Results from analyses in the Netherlands Cohort Study. *Food and Chemical Toxicology*, 38, 599-605.
- Brand-Williams, W., Cuvelier, M. E., & Berset, C. (1995). Use of free radical method to evaluate antioxidant activity. *Lebensmittel-Wissenschaft und- Technologie*, 28, 25-30.
- Brunetto, M. R., Gutiérrez, L., Delgado, Y., Galignani, M., Zambrano, A., Gómez, A., Ramos, G., & Romero, G. (2007). Determination of theobromine, theophylline and caffeine in cocoa samples by a high-performance liquid chromatographic method with on-line sample clean-up in a switching-column system. *Food Chemistry*, 100, 459-467.
- Burkill, I. H. (1966). *A dictionary of the economic products of the Malay Peninsular*. Ministry of Agriculture and Cooperatives, Kuala Lumpur.
- Cai, Y., Luo, Q., Sun, M., & Corke, H. (2004). Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. *Life Sciences*, 74, 2157-2184.
- Casagrande, R., Georgetti, S. R., Verri Jr., W. A., Dorta, D. J., dos Santos, A. C., & Fonseca, M. J. V. (2006). Protective effect of topical

- formulations containing quercetin against UVB-induced oxidative stress in hairless mice. *Journal of Photochemistry and Photobiology B*, 84, 21-27.
- Chanda, S., & Nagani, K. (2010). Antioxidant capacity of *Manilkara zapota* L. leaves extracts evaluated by four *in vitro* methods. *Nature and Science of Sleep*, 8, 260-266.
- Chang, C., Yang, M., Wen, H., & Chern, J. (2002) Estimation of total flavonoid content in propolis by two complementary colorimetric methods. *Journal of Food Drug Analysis*, 10, 178-182.
- Chang, T. S. (2009). An updated review of tyrosinase inhibitors. *International Journal of Molecular Sciences*, 10, 2440-2475.
- Chae, S., Piao, M. J., Kang, K. A., Zhang, R., Kim, K. C., Youn, U. J., Nam, K-W., Lee, J. H., & Hyun, J. W. (2011). Inhibition of matrix metalloproteinase-1 induced by oxidative stress in human keratinocytes by Mangiferin isolated from *Anemarrhena asphodeloides*. *Bioscience, Biotechnology and Biochemistry*, 75, 2321-2325.
- Che Man, Y., & Sazili, A. Q. (2010). Food Production from the Halal Perspective. In: Guerrero- Legarreta, I., Alarcón-Rojo, A. D., Hui, Y. H., & Alavarado, C. *Handbook of Poultry Science and Technology*, 183-216.
- Che Man, Y., & Mustafa, S. (2010). Updates of halal products authentication. Slides presentation at World Halal Research Summit 2010. Malaysia: Kuala Lumpur.
- Che Sulaiman, I. S., Basri, M., Masoumi, H. R. F., Chee, W. J., Ashari, S. E., & Ismail, M. (2017). Effects of temperature, time, and solvent ratio on the extraction of phenolic compounds and the anti-radical activity of *Clinacanthus nutans* Lindau leaves by response surface methodology. *Chemistry Central Journal*, DOI 10.1186/s13065-017-0285-1
- Chen, C. -Y., Kuo, P. -L., Chen, Y. -H., Huang, J. -C., Ho, M. -L., Lin, R. -J., Chang, J. -S., & Wang, H. -M. (2010). Tyrosinase inhibition, free radical scavenging, antimicroorganism and anticancer proliferation activities of *Sapindus mukorossi* extracts. *Journal of the Taiwan Institute of Chemical Engineers*, 41, 129-135.
- Cheng, Y. -C., & Prusoff, W. H. (1973). Relationship between the inhibition constant (KI) and the concentration of inhibitor which causes 50 per cent inhibition (I_{50}) of an enzymic reaction. *Biochemical Pharmacology*, 22, 3099-3108.
- Chen, Q. X., & Kubo, I. (2002). Kinetics of mushroom tyrosinase inhibition by quercetin. *Journal of Agricultural and Food Chemistry*, 50, 4108-4112.
- Cheng, Z., Moore, J., & Yu, L. (2006). High-throughput relative DPPH• radical assay. *Journal of Agriculture and Food Chemistry*, 54, 7429-7436.

- Cheung, L. M., Cheung, P. C. K., & Ooi, V. E. C. (2003). Antioxidant activity and total phenolics of edible mushroom extract. *Food Chemistry*, 81, 249-255.
- Chiang, H. M., Lin, T. J., & Chiu, C. Y. (2011). *Coffea arabica* extract and its constituents prevent photoaging by suppressing MMPs expression and MAP kinase pathway. *Food and Chemical Toxicology*, 49, 309-318.
- Chiari, B. G., Trovatti, E., Pecoraro, É, Corr ea, M. A., Cicarelli, R. M. B., Ribeiro, S. J. L., & Isaac, V. L. B. (2014). Synergistic effect of green coffee oil and synthetic sunscreen for health care application. *Industrial Crops and Products*, 52, 389-393.
- Chidambaram, U., Pachamuthu, V., Natarajan, S., Elango, B., Narayanan, S., Ramkumar, K. M. (2013). *In vitro* evaluation of free radical scavenging activity of *Codario calyxmotorius* root extract. *Asian Pacific Journal of Tropical Medicine*, 6, 188-194.
- Chiang, H. -M., Lin, T. -J., Chiu, C. -Y., Chang, C. -W., Hsu, K. -C., Fan, P. -C., & Wen, K. -C. (2011). *Coffea arabica* extract and its constituents prevent photoaging by suppressing MMPs expression and MAP kinase pathway. *Food and Chemical Toxicology*, 49, 309-318.
- Choi, K. C., Lee, K. D., Song, C. E., Ilavenil, S., Sriganpalram, S., Arasu, M. V., & Kuppusamy, P. (2018). Quantification of major phenolic and flavonoid markers in forage crop *Lolium multiflorum* using HPLC-DAD. *Revista Brasileira de Farmacognosia*, 28, 282-288.
- Chueahongthong, F., Ampasavate, C., Okonogi, S., Tima, S., & Anuchapreeda, S., (2011). Cytotoxic effects of crude kaffir lime (*Citrus hystrix*, DC.) leaf fractional extracts on leukemic cell lines. *Journal of Medicinal Plants Research*, 5, 3097-3105.
- COLIPA – The European Cosmetics Association (2008). *Cosmetics Europe: Guidelines for the evaluation of the efficacy of cosmetic products*, www.cosmeticseurope.eu, 1-16.
- Contreras, J., Calder n-Jaimes, L., Guerra-Hern andez, E., & Garcia-Villanova, B. (2011). Antioxidant capacity, phenolic content and vitamin C in pulp, peel and seed from 24 exotic fruits from Colombia. *Food Research International*, 44, 2047-2053.
- Cooper, K. A., Campos-Gimenez, E., Alvarez, D. J., Nagy, K., Donovan, J. L., & Williamson, G. (2007). Rapid reverse-phase ultra-performance liquid chromatography analysis of the major cocoa polyphenols and inter-relationship of their concentration in chocolate. *Journal of Agricultural and Food Chemistry*, 55, 2841-2847.
- Coppo, E., & Marchese, A. (2014). Antibacterial activity of polyphenols. *Current Pharmaceutical Biotechnology*, 15, 380-390.
- Corazza, M., Virgili, A., Toni, G., Minghetti, S., & Tiengo, S. (2016). Level of use and safety of botanical products for itching vulvar dermatoses. Are patch tests useful? *Contact Dermatitis* 74, 289-294.

- Cornacchione, S., Sadick, N. S., Neveu, M., & Talbourdet, S. (2007). *In vivo* skin antioxidant effect of a new combination based on a specific *Vitis vinifera* shoot extract and a biotechnological extract. *Journal of Drugs in Dermatology*, 6, 8-13.
- Cowan, M. M. (1999). Plant products as antimicrobial agents. *Clinical Microbiology Reviews*, 12, 564-582.
- Cuatrecasas, J. (1964). Cacao and its allies. A taxonomic revision of the genus *Theobroma*. *Contributions from the United States National Herbarium*, 36, 379-614.
- D'Alessandro, L. G., Kriaa, K., Nikov, I., & Dimitrov, K. (2012). Ultrasound assisted extraction of polyphenols from black chokeberry. *Separation Science and Technology*, 93, 42-47.
- D'Orazio, J., Jarrett, S., Amaro-Ortiz, A., & Scott, T. (2013). UV radiation and the skin. *International Journal of Molecular Sciences*, 14, 12222-12248.
- Dal'Belo, S. E., Gaspar, L. R., & Maia Campos, P. M. (2006). Moisturizing effect of cosmetic formulations containing Aloe vera extract in different concentrations assessed by skin bioengineering techniques. *Skin Research and Technology*, 12, 241-246.
- Darlenski, R., Sassning, S., Tsankov, N., & Fluhr, J. (2009). Non-invasive *in vivo* methods for investigation of the skin barrier. *European Journal of Pharmaceutics and Biopharmaceutics*, 72, 295-303.
- Darvin, M. E., Fluhr, J. W., Schanzer, S., Richter, H., Patzelt, A., Meinke, M. C., Zastrow, L., Golz, K., Doucet, O., Sterry, W., & Lademann, J. (2011). Dermal carotenoid level and kinetics after topical and systemic administration of antioxidants: Enrichment strategies in a controlled *in vivo* study. *Journal of Dermatological Science*, 64, 53-58.
- De Paepe, K., Lagarde, J. M., Gall, Y., Roseeuw, D., & Rogiers, V. (2000). Microrelief of the skin using a light transmission method. *Archives of Dermatological Research*, 292, 500-510.
- Debowska, R., Rogiewicz, K., & Iwanenko, T. (2005). Folic acid (Folacin) – new application of a cosmetic ingredient. *Kosmetische Medizin*, 3, 16-22.
- Demiate, I. M., Zielinski, A., Wojeicchowski, J. P., & Nogueira, A. (2018). Extraction optimization of phenolic extracts from carioca bean (*Phaseolus vulgaris* L.) using response surface methodology. *Food Analytical Methods*, 12, 148-159.
- Demina, N. S., & Lysenko, S. V. (1996). Collagenolytic enzymes synthesized by microorganisms. *Mikrobiologija*, 65, 293-304.
- Deus, V. L., Cerqueira E Silva, M. B., Maciel, L. F., Miranda, L. C. R., Hirooka, E. Y., Soares, S. E., Ferreira, E. S., & Silva Bispo, E. (2018). Influence of drying methods on cocoa (*Theobroma cacao* L.): antioxidant activity and presence of ochratoxin A. *Food Science and Technology*, 38, 278-285.

- Di Giovanni, C., Arcoraci, V., Gambardella, L., & Sautebin, L. (2006). Cosmetovigilance survey: are cosmetics considered safe by consumers? *Pharmacological Research*, 53, 16-21.
- Diffey, B., & Robson, J. (1989). A new substrate to measure sunscreen protection factors throughout the ultraviolet spectrum. *Journal of the Society of Cosmetic Chemists*, 40, 127-133.
- Dobrev, H. (2007). Clinical and instrumental study of the efficacy of a new sebum control cream. *Journal of Cosmetic Dermatology*, 6, 113-118.
- Department of Statistic Malaysia (2019). Agriculture statistics-time series. Department of Statistics, Malaysia, December 2019.
- Draelos, Z. D. (2014). Facial skin care products and cosmetics. *Clinics in Dermatology*, 32, 809-812.
- Dreosti, I. E. (2000). Antioxidant polyphenols in tea, cocoa and wine. *Nutrition*, 16, 7-8.
- Dudonné, S., Vitrac, X., Coutiere, P., Woillez, M., & Merillon, J. M. (2009). Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. *Journal of Agriculture and Food Chemistry*, 57, 1768-1774.
- Dufrane, D., Cornu, O., & Verraes, T. (2001). In vitro evaluation of acute cytotoxicity of human chemically treated allografts. *European Cells and Materials*, 1, 52-58.
- Dyke, T. M., & Sams, R. A. (1998). Detection and determination of theobromine and caffeine in urine after administration of chocolate-coated peanuts to horses. *Journal of Analytical Toxicology*, 22, 112-116.
- Elias, P. M. (2005). Stratum corneum defensive functions: an integrated view. *Journal of Investigative Dermatology*, 125, 183-200.
- Elias, P. M. (2012). Structure and function of the stratum corneum extracellular matrix. *Journal of Investigative Dermatology*, 132, 2131-2133.
- Emelda, A., Rusli, & Nurlina (2015). Polyphenol content in powder and purified extract of unfermented cocoa beans from enrekang regency of South Sulawesi Indonesia. *Journal of Chemical and Pharmaceutical Research*, 7, 972-974.
- European Union (2006). 2006/647/EC: Commission Recommendation of 22 September 2006 on the efficacy of sunscreen products and the claims made relating thereto (notified under document number C(2006) 4089. *Official Journal – European Union Legislation L*, 49, 39-43.
- Evans, J. A., & Johnson, E. J. (2010). The role of phytonutrients in skin health. *Nutrients*, 2, 903-928.
- Facino, R. M., Carini, M., Aldini, G., & Calloni, M. T. (1998). Sparing effect of procyanidins from *Vitis vinifera* on vitamin E: *in vitro* studies. *Planta Medica*, 64, 343-347.
- Farag, R. S., Badei, A. Z. M. A., & El-Baroty, G. S. A. (1989). Antioxidant activity of some spice essential oils on linoleic acid oxidation in

- aqueous media. *Journal of the American Oil Chemists' Society*, 66, 800-804.
- Farris, P., Krutmann, J., Li, Y. -H., McDaniel, D., & Krol, Y. (2013). Resveratrol: A unique antioxidant offering a multi-mechanism approach for treating aging skin. *Journal of Drugs in Dermatology*, 12, 1389-1394.
- Fentem, J. H., & Botham, P. A. (2002). ECVAM's activities in validating alternative tests for skin corrosion and irritation. *Alternatives to Laboratory Animals*, 30, 61-67.
- Fernandez, D. P., Goodwin, A. R. H., Lemmon, E. W., Levelt-sengers, J. M. H., & Williams, R. C. (1997). A formulation for the static permittivity of water and steam at temperature from 238 to 873 at pressures up to 1200 MPa, including derivatives and Debye-Hückel coefficients. *Journal of Physical Chemistry*, 26, 1125-1166.
- Fiore, M., Zanier, R., & Degrassi, F. (2002). Reversible G(1) arrest by dimethyl sulfoxide as a new method to synchronize Chinese hamster cells. *Mutagenesis*, 17, 419-424.
- Fisher, R. A. (1935). The logic of inductive inference. *Journal of the Royal Statistical Society A*, 98, 39-54.
- Fisher, G. J., Wang, Z. Q., Datta, S. C., Varani, J., Kang, S., & Voorhees, J. J. (1997). Pathophysiology of premature skin aging induced by ultraviolet light. *The New England Journal of Medicine*, 337, 1419-1428.
- Forsyth, W. G. C., Quesnel, V. C., & Roberts, J. B. (1958). Interaction of polyphenols and proteins during cacao curing. *Journal of Science of Food and Agriculture*, 9, 181-184.
- Fujii, T., Wakaizumi, M., Ikami, T., & Saito, M. (2008). Amla (*Emblca officinalis* Gaertn.) extract promotes procollagen production and inhibits matrix metalloproteinase-1 in human skin fibroblasts. *Journal of Ethnopharmacology*, 119, 53-57.
- Fujikake, K., Tago, S., Plasson, R., Nakazawa, R., Okano, K., Maezawa, D., Mukawa, T., Kuroda, A., & Asakura, K. (2014). Problems of *in vitro* SPF measurements brought about by viscous fingering generated during sunscreen applications. *Skin Pharmacology and Physiology*, 27, 254-262.
- Gaikwad, M., & Kale, S. (2011). Formulation and *in vitro* evaluation for sun protection factor of *Moringa oleifera* Lam (Family-Moringaceae) oil sunscreen cream. *International Journal of Pharmacy and Pharmaceutical Sciences*, 3, 371-375.
- Garcia-Castello, E. M., Rodriguez-Lopez, A. D., Mayor, L., Ballesteros, R., Conidi, C., & Cassano, A. (2015). Optimization of conventional and ultrasound assisted extraction of flavonoids from grapefruit (*Citrus paradise* L.) solid wastes. *LWT – Food Science and Technology*, 64, 1114-1122.

- Garcia-Salas, P., Morales-Soto, A., Segura-Carretero, A., & Fernandez-Gutierrez, A. (2010). Phenolic-compound-extraction systems for fruit and vegetable samples: Review. *Molecules*, 15, 8813-8826.
- Garg, C., Khurana, P., & Garg, M. (2017). Molecular mechanisms of skin photoaging and plant inhibitors. *International Journal of Green Pharmacy*, 11, 217-232.
- Gasser, P., Lati, E., Peno-Mazzarino, L., Bouzoud, D., Allegaert, L., & Bernaert, H. (2008). Cocoa polyphenols and their influence on parameters involved in *ex vivo* skin restructuring. *International Journal of Cosmetic Science*, 30, 339-345.
- Gęgotek, A., Bielawska, K., Biernacki, M., Dobrzyńska, I., & Skrzydlewska, E. (2017). Time-dependent effect of rutin on skin fibroblasts membrane disruption following UV radiation. *Redox Biology*, 12, 733-744.
- Gertenbach, D. D. (2001). Solid-liquid extraction technologies for manufacturing nutraceuticals from botanicals. In: Shi, J., Mazza, G. & Maguer, M.L. (Eds.), *Functional Foods Biochemical and Processing Aspects*, 2. CRC, Boca Raton, pp. 331–366.
- Ghuman, S., Ncube, B., Finnie, J. F., McGaw, L. J., Coopoosamy, R. M., & Staden, J. V. (2016). Antimicrobial activity, phenolic content, and cytotoxicity of medicinal plant extracts used for treating dermatological diseases and wound healing in KwaZulu-Natal, South Africa. *Frontiers in Pharmacology*, 7, 1-9.
- Giulietti, A., Overbergh, L., Valckx, D., & Decallonne, B. (2001). An overview of real-time quantitative PCR: Applications to quantify cytokine gene expression. *Methods*, 25, 386-401.
- Gotti, R., Furlanetto, S., Pinzauti, S., & Cavrini, V. (2006). Analysis of catechins in *Theobroma cacao* beans by cyclodextrin-modified micellar electrokinetic chromatography. *Journal of Chromatography A*, 1112, 345-352.
- Greer, F., Hudson, R., Ross, R., & Graham, T. (2001). Caffeine ingestion decrease glucose disposal during a hyperinsulinemic-euglycemic clamp in sedentary humans. *Diabetes*, 50, 2349-2354.
- Griffiths, H. A., Wilhelm, K. -P., Robinson, M. K., York, M., & Basketter, D. A. (1996). Inter-laboratory evaluation of a human 4h patch test for the evaluation of skin irritation potential/hazard. *Food and Chemical Toxicology*, 35, 255-260.
- Grosso, C., Valentão, P., Ferreres, F., & Andrade, P. B. (2015). Alternative and efficient extraction methods for marine-derived compounds. *Marine Drugs*, 13, 3182-3230.
- Gu, L., Kelm, M., Hammerstone, J. F., Beecher, G., Cunningham, D., Vannozi, S., & Prior, R. L. (2002). Fractionation of polymeric procyanidins from lowbush blueberry and quantification of procyanidins in selected foods with an optimized normal-phase HPLC-MS fluorescent detection method. *Journal of Agriculture and Food Chemistry*, 50, 4852-4860.

- Gu, L., House, S. E., Wu, X., Ou, B., & Prior R. L. (2006). Procyanidin and catechin contents and antioxidant capacity of cocoa and chocolate products. *Journal of Agricultural and Food Chemistry*, 54, 4057-4061.
- Gunes, S., Tamburaci, S., Dalay, M. C., & Gurhan, I. D. (2017). *In vitro* evaluation of *Spirulina platensis* extract incorporated skin cream with its wound healing and antioxidant activities. *Pharmaceutical Biology*, 55, 1824-1832.
- Guo, C., Yang, J., Wei, J., Li, Y., Xu, J., & Jiang, Y. (2003). Antioxidant activities of peel, pulp and seed fractions of common fruits as determined by FRAP assay. *Nutrition Research*, 23, 1719-1726.
- Hachem, J. P., Crumrine, D., Fluhr, J., Brown, B. E., Feingold, K. R., & Elias, P. M. (2003). pH directly regulates epidermal permeability barrier homeostasis, and stratum corneum integrity/cohesion. *Journal of Investigative Dermatology*, 121, 345-353.
- Hachem, J., -P., Behne, M., Aronchik, I., Demerjian, M., Feingold, K. R., Elias, P. M., & Mauro, T. M. (2005). Extracellular pH controls NHE1 expression in epidermis and keratinocytes: implications for barrier repair. *Journal of Investigative Dermatology*, 125, 790-797.
- Hagerman, A. E., & Butler, L. G. (1981). The specificity of the proanthocyanins-protein interaction. *Journal of Biological Chemistry*, 256, 4494-4497.
- Halal Industry Development (2014). Available from <http://www.hdcglobal.com>.
- Hammerstone, J. F., Sheryl, A. L., Alyson, E. M., Robert, R., & Harold, H. S. (1999). Identification of procyanidins in cocoa (*Theobroma cacao*) and chocolate using high-performance liquid chromatography/ Mass spectrometry. *Journal of Agricultural and Food Chemistry*, 47, 490-496.
- Hashim, P., & Mat Hashim, D. (2013). A review of cosmetic and personal care products: Halal perspective and detection of ingredient. *Pertanika Journals of Science and Technology*, 21, 281-292.
- Hashim, P., Shahab, N., Masilamani, T., Baharom, R., & Ibrahim, R. (2009). A cosmetic analysis in compliance with the legislative requirements, halal and quality control. *Malaysian Journal of Chemistry*, 11, 1081-1087.
- Haya, R., Nuraziawati, M. Y., Mohd. Jaafar, H., & Mohd. Qahar, M. (2012). *Book of Malaysia Cocoa Clones*. Malaysian Cocoa Board.
- Heinrich, U., Neukam, K., Tronnier, H., Sies, H., & Stahl, W. (2006). Long-term ingestion of high flavanol cocoa provides photoprotection against UV-induced erythema and improves skin condition in women. *The Journal of Nutrition*, 136, 1565-1569.
- Helfrich, Y. R., Sachs, D., & Voorhees, J. J. (2008). Overview of skin aging and photoaging. *Dermatology Nursing*, 20, 177-183.
- Hernandez, C. H., Viera, I., Morales-Siliero, A., & Fernandez-Bolanos, J. (2018). Bioactive compounds in Mexican genotypes of cocoa cotyledon and husk. *Food Chemistry*, 240, 831-839.

- Hii, C. L., Law, C. L., Cloke, M., & Suzannah, S. (2009). Thin layer drying kinetics of cocoa and dried product quality. *Biosystems Engineering*, 102, 153-161.
- Hok, H. C., Ramli, N., Yatim, A. M., & Said, M. (2016). HPLC determination of methylxanthines and polyphenols levels in cocoa and chocolate products. *Malaysian Journal of Analytical Sciences*, 7, 377-386.
- Horváth, S. (1980). Cytotoxicity of drugs and diverse chemical agents to cell cultures. *Toxicology*, 16, 59-66.
- Hsu, C. -K., Chang, C. -T., Lu, H. -Y., & Chung, Y. -C. (2007). Inhibitory effects of the water extracts of *Lavendula* sp. on mushroom tyrosinase activity. *Food Chemistry*, 105, 1099-1105.
- Huda-Faujan, N., Noriham, A., Norrakiah, A. S., & Babji, A. S. (2009). Antioxidant activity of plants methanolic extracts containing phenolic compounds. *African Journal of Biotechnology*, 8, 484-489.
- Husain, R., Abd, G. I., Mohammad, A., & Mehad, S. (2012). Current practices among halal cosmetics manufacturer in Malaysia. *Journal of Statistical Modelling and Analytic*, 3, 46-51.
- Hussin, S. R., Hashim, H., Yusof, R. N., & Alias, N. N. (2013). Relationship between product factors, advertising, and purchase intention of halal cosmetics. *Pertanika Journals of Social Sciences & Humanities*, 21, 85-100.
- Ichihashi, M., Ando, H., Yoshida, M., Niki, Y., & Matsui, M. (2009). Photoaging of the skin. *Journal of Anti-Aging Medicine*, 6, 46-59.
- Ingrid, G., Barbara, P., Juan, G., Victor, C., Renato, N., Miguel Angel, M., Natalio, G., Julia, B., Antonio, C., & Miguel, A. (2012). Evaluation of the cell viability of human Wharton's jelly stem cells for use in cell therapy. *Tissue Engineering: Part C*, 18, 408-419.
- International Organization for Standardization (2010) ISO 24444: cosmetics – sun protection test methods – *in vivo* determination of the sun protection factor (SPF), Geneva.
- Ishii, H., Kitamura, T., & Todo, H. (2008). Combined effect of sodium chondroitin sulfate and sodium hyaluronate on skin moisturization following single and repeated application. *Asian Journal of Pharmaceutical Sciences*, 3, 94-101.
- Jacek, A., & Katarzyna, P. (2008). Flavonoids as components of biologically active cosmeceuticals. *Clinics in Dermatology*, 26, 347-357.
- Jahanban-Esfahlan, A., Ostadrahimi, A., Tabibiazar, M., & Amarowicz, R. (2019). A comparative review on the extraction, antioxidant content and antioxidant potential of different parts of walnut (*Juglans regia* L.) fruit and tree. *Molecules*, 24, 1-40.
- Jarzycka, A., Lewińska, A., Gancarz, R., & Wilk, K. A. (2013). Assessment of extracts of *Helichrysum arenarium*, *Crataegus monogyna*, *Sambac nigra* in photoprotective UVA and UVB; photostability in cosmetic

- emulsions. *Journal of Photochemistry and Photobiology B: Biology*, 128, 50-57.
- Jayaprakasha, G. K., Singh, R. P., & Sakariah, K.K. (2001). Antioxidant activity of grape seed (*Vitisvinifera*) extracts on peroxidation models *in vitro*. *Food Chemistry*, 73, 285-290.
- Jinap, S., Dimick, P. S., & Hollender, R. (1995). Flavour evaluation of chocolate formulated from cocoa beans from different countries. *Journal of Food Control*, 6, 105-110.
- Jonfia-Essien, W. A., West, G., Alderson, P. G., & Tucker, G. (2008). Phenolic content and antioxidant capacity of hybrid variety cocoa beans. *Food Chemistry*, 108, 1155-1159.
- Jovanovic, S. V., Steenhen, S., Simic, M. O., & Hara, Y. (1998). Antioxidant properties of flavonoids. In: Rice-Evans C.A. & Packer, L. (Eds.), *Flavonoids in health and disease*. New York: Marcel Dekker, 1998. P. 137-162.
- Judge, M. R., Griffiths, H. A., & Basketter, D. A. (1996). Variation in response of human skin to irritant challenge. *Contact Dermatitis*, 34, 115-117.
- Kanashiro, A., Souza, J. G., Kabeya, L. M., & Azzolini, A. E. C. S. (2007). Elastase release by stimulated neutrophils inhibited by flavonoids: importance of the catechol group. *Zeitschrift fur Naturforschung*, 62, 357-361.
- Kamali, M. H. (2000). Principles of Islamic Jurisprudence. *Journal of Law and Religion*, 15, 385-387.
- Kamath, V., & Rajini, P. S. (2007). The efficacy of cashew nut (*Anacardium occidentale* L.) skin extract as a free radical scavenger. *Food Chemistry*, 103, 428-433.
- Kameyama, K., Sakai, C., Kuge, S., Nishiyama, S., Tomita, Y., Ito, S., Wakamatsu, K., & Hearing, V. J. (1995). The expression of tyrosinase, tyrosinase-related proteins 1 and 2 (TRP1 and TRP2), the silver protein, and a melanogenic inhibitor in human melanoma cells of differing melanogenic activities. *Pigment Cell Research*, 8, 97-104.
- Kampf, G., & Ennen, J. (2006). Regular use of a hand cream can attenuate skin dryness and roughness caused by frequent hand washing. *BMC Dermatology*, 6, 1.
- Karadag, A., Ozcelik, B., & Saner, S. (2009). Review of methods to determine antioxidant capacities. *Food Analytical Methods*, 2, 41-60.
- Katz, D. L., Doughty, K., & Ali, A. (2011). Cocoa and chocolate in human health and disease. *Antioxidants & Redox Signaling*, 15, 2779-2811.
- Kelm, M., Robbins, R. J., Johnson, J. C., & Hammerstone, J. (2006). High-performance liquid chromatography separation and purification of cacao (*Theobroma cacao* L.) procyanidins according to degree of polymerization using a diol stationary phase. *Journal of Agricultural and Food Chemistry*, 54, 1571-1576.

- Khan, B. A., Akhtar, N., Mahmood, T., Qayum, M., & Zaman, S. U. (2010). Formulation and pharmaceutical evaluation of a W/O emulsion of *Hippophae ramnoides* fruit extract. *Journal of Pharmacy Research*, 3, 1342-1344.
- Khazaka, G. (2000). CK Electronic Cologne; 2000. Information and operating instructions for the Visioscan VC 98 and the software SELS (Surface evaluation of living skin) pp. 34-36.
- Khoddami, A., Wilkes, M. A., & Roberts, T. H. (2013). Techniques for analysis of plant phenolic compounds. *Molecules*, 18, 2328-2375.
- Khopde, S. M., Priyadarsini, K. I., Mukherjee, T., Kulkarni, P. B., Satav, J. G., & Bhattacharya, R. K. (1998). Does β -carotene protect membrane lipids from nitrogen dioxide? *Free Radical Biology & Medicine*, 25, 66-71.
- Khurana, S., Venkataraman, K., Hollingsworth, A., Piche, M., & Tai, T. C. (2013). Polyphenols: Benefits to the cardiovascular system in health and in aging. *Nutrients*, 5, 3779-3827.
- Kielty, C. M., Sheratt, M. J., & Shuttleworth, C. A. (2002). Elastic fibres. *Journal of Cell Science*, 115, 2817-2828.
- Kim, E., Kim, S., Nam, G. W., Lee, H., Moon, S., & Chang, I. (2009). The alkaline pH-adapted skin barrier is disrupted severely by SLS-induced irritation. *International Journal of Cosmetic Science*, 31, 263-269.
- Kim, H., & Keeney, P. G. (1984). (-)-Epicatechin content in fermented and unfermented cocoa beans. *Journal of Food Science*, 49, 1090-1092.
- Kim, H. J., Seo, S. H., Lee, B. G., & Lee, Y. S. (2005). Identification of tyrosinase inhibitors from *Glycyrrhiza uralensis*. *Planta Medica*, 71, 785-787.
- Kim, Y., Uyama, H., & Kobayashi, S. (2004). Inhibition effects of (+)-catechin aldehyde polycondensates on proteinases causing proteolytic degradation of extracellular matrix. *Biochemical Biophysical Research Communications*, 320, 256-261.
- Kim, J. S., Kwon, Y. S., Sa, Y. J., & Kim, M. J. (2011). Isolation and identification of Sea Buckthorn (*Hippophae rhamnoides*) phenolics with antioxidant activity and α -glucosidase inhibitory effect. *Journal of Agricultural and Food Chemistry*, 59, 138-144.
- Kim, Y. -S., & Joh, T. H. (2012). Matrix metalloproteinases, new insights into the understanding of neurodegenerative disorders. *Biomolecules & Therapeutics*, 20, 133-143.
- Kim, J.- E., Song, D., Kim, J., Choi, J., Kim, J. R., & Yoon, H.- S., (2016). Oral supplementation with cocoa extract reduces UVB-induced wrinkles in hairless mouse skin. *Journal of Investigative Dermatology*, 136, 1012-1021.
- Kimmelman, J., Weijer, C., & Meslin, E. (2009). Helsinki discords: FDA, ethics, and international drug trials. *The Lancet*, 373, 13-14.
- Kofink, M., Papagiannopoulos, M., & Galensa, R. (2007). Enantioseparation of catechin and epicatechin in plant food by chiral capillary

- electrophoresis. *European Food Research and Technology*, 225, 569-577.
- Kohen, R., & Gati, I. (2000). Skin low molecular weight antioxidants and their role in aging and in oxidative stress. *Toxicology*, 148, 149-157.
- Kohl, E., Steinbauer, J., Landthaler, M., & Szeimes, R. M. (2011). Skin Aging. *Journal of the European Academy of Dermatology and Venereology*, 25, 873-884.
- Kondo, T., & Hearing, V. J. (2011). Update on the regulation of mammalian melanocyte function and skin pigmentation. *Expert Review of Dermatology*, 6, 97-108.
- Korac, R. R., & Khambholja, K. M. (2011). Potential of herbs in skin protection from ultraviolet radiation. *Pharmacognosy Reviews*, 5, 164-173.
- Kowalski, R. (2007). GC analysis of changes in the fatty acid composition of sunflower and olive oils heated with quercetin, caffeic acid, protocatechuic acid and butylated hydroxyanisole. *Acta Chromatographica*, 18, 15-23.
- Krysiak, W. (2006). Influence of roasting conditions on coloration of roasted cocoa beans. *Journal of Food Engineering*, 77, 449-453.
- Kubista, M., Andrade, J. M., Bengtsson, M., Forootan, A., Jonák, J., Lind, K., Sindelka, R., Sjöback, R., Sjögreen, B., Strömbom, L., Ståhlberg, A., & Zoric, N. (2006). The real-time polymerase chain reaction. *Molecular Aspects of Medicine*, 27, 95-125.
- Kubo, I., & Kinoshita, I. (1999). Flavonols from saffron flower: tyrosinase inhibitory activity and inhibition mechanism. *Journal of Agricultural and Food Chemistry*, 47, 4121-4125.
- Kubo, I., Kinoshita, I., & Yokokawa, Y. J. (1994). Tyrosinase Inhibitors from *Anacardium occidentale* Fruits. *Journal of Natural Product*, 57, 545-551.
- Kumar, S., & Pandey, A. K. (2013). Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*, 1-16.
- Kumaran, A., & Karunakaran, J. R. (2006). Antioxidant and free radical scavenging activity of an aqueous extract of *Coleus aromaticus*. *Food Chemistry*, 97, 109-114.
- Kwist, K., Bridges, W. C., & Burg, K. J. L. (2016). The effect of cell passage number on osteogenic and adipogenic characteristics of D1 cells. *Cytotechnology*, 68, 1661-1667.
- Lademann, J., Martina, C. M., Wolfram, S., & Maxim, E. D. (2011). Carotenoids in human skin. *Experimental Dermatology*, 20, 377-382.
- Lamuela-Raventós, R. M., Romero-Pérez, A. I., Andrés-Lacueva, C., & Tornero, A. (2005). Review: Health effects of cocoa flavonoids. *Food Science and Technology International*, 11, 159-176.
- Lazarus, S. A., Adamson, G. E., Hammerstone, J. F., & Schmitz, H. H. (1999). High-performance liquid chromatography/mass spectrometry analysis

- of procyanidins in food and beverages. *Journal of Agriculture and Food Chemistry*, 47, 3693-3701.
- Le Coz, C. J., & Schneider, G. A. (1987). Contact dermatitis from tertiary butylhydroquinone in a hair dye with cross sensitivity to BHA and BHT. *Contact Dermatitis*, 17, 257-258.
- Lee, J. -S., Park, K. -Y., Min, H. -G.; Lee, S. J., Kim, J. -J., Choi, J. -S., Kim, W. -S., & Cha, H. -J. (2010). Negative regulation of stress-induced matrix metalloproteinase-9 by SIRT 1 in skin tissue. *Experimental Dermatology*, 19, 1060-1066.
- Levine, R. L., & Stadtman, E. R. (2001). Oxidative modification of proteins during ageing. *Experimental Gerontology*, 36, 1495-1502.
- Liang, C., Lim, J. H., Kim, S. H., & Kim, D. S. (2012). Dioscin: a synergistic tyrosinase inhibitor from the roots of *Smilax china*. *Food Chemistry*, 134, 1146-1148.
- Lieberei, R., Kadow, D., & Seigler, D. (2013). Cocoa cultivation, directed breeding and polyphenolics. In: Ramawat, K., & Mérillon, J. M. (eds.) *Natural Products*. Springer, Berlin, Heidelberg.
- Lin, J. Y., & Tang, C. Y. (2007). Determination of total phenolic and flavonoid contents in selected fruits and vegetables, as well as their stimulatory effects on mouse splenocyte proliferation. *Food Chemistry*, 101, 140-147.
- Liu, J., Cao, R., Yi, W., Ma, C., Wan, Y., Zhou, B., Ma, L., & Song, H. (2009). A class of potent tyrosinase inhibitors: alkylidenethiosemicarbazide compounds. *European Journal of Medicinal Chemistry*, 44, 1773-1778.
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta C_T}$ method. *Methods*, 25, 402-408.
- Liyana-Pathirana, C., & Shahidi, F. (2005). Optimization of extraction of phenolic compounds from wheat using response surface methodology. *Food Chemistry*, 93, 47-56.
- Lodyga-Chruścińska, E., Sykula, A., & Więdocha, M. (2018). Hidden metals in several brands of lipstick and face powder present on Polish market. *Cosmetics*, 5, 1-8.
- Lohani, A., Verma, A., Joshi, H., Yadav, N., & Karki, N. (2014). Nano technology- based cosmeceuticals. *ISRN Dermatology*, 22, 1-14.
- Lourith, N., Kanlayavattanukul, M., Chaikul, P., Chansriniyom, C., & Bunwatcharaphansakun, P. (2017). *In vitro* and cellular activities of the selected fruits residues for skin aging treatment. *The Anais da Academia Brasileira de Ciências*, 89, 577-589.
- Machnik, M., Kaiser, S., Koppe, S., Kietzmann, M., Schenk, I., Düe, M., Thevis, M., Schänzer, W., & Toutain, P. -L. (2016). Control of methylxanthines in the competition horse: pharmacokinetic/pharmacodynamics studies

- on caffeine, theobromine and theophylline for the assessment of irrelevant concentrations. *Drug Testing and Analysis*, 9, 1372-1384.
- Madani, W., Kermasha, S., & Bisakowaki, B. (1999). Inhibition of tyrosinase activity by a polyphenol esterase using selected phenolic substrates. *Phytochemistry*, 52, 1001-1008.
- Madhan, B., Krishnamoorthy, G., Rao, J. R., & Nair, B. U. (2007). Role of green tea polyphenols in the inhibition of collagenolytic activity by collagenase. *International Journal of Biological Macromolecules*, 41, 16-22.
- Mahmood, T., Akhtar, N., Khan, B. A., Khan, M. S., & Saeed, T. (2011). Changes in skin mechanical properties after long-term application of cream containing green tea extract. *Aging Clinical and Experimental Research*, 23, 333-336.
- Maity, K. K., Patra, S., Dey, B., & Bhunia, S. K. (2011). A heteropolysaccharide from aqueous extract of an edible mushroom, *Pleurotus ostreatus* cultivar: structural and biological studies. *Carbohydrate Research*, 346, 366-372.
- Majeed, M., Bhat, B., Jadhav, A. N., Srivastava, J. S., & Nagadhushanam, K. (2009). Ascorbic acid and tannins from *Embllica officinalis* Gaertn. Fruits-a revisit. *Journal of Agricultural and Food Chemistry*, 57, 220-225.
- Mäkelä, M. (2017). Experimental design and response surface methodology in energy applications: A tutorial review. *Energy Conversion and Management*, 151, 630-640.
- Mandal, J., Acharya, S., & Parija, S. C. (2011). Ethics in human research. *Tropical Parasitology*, 1, 2-3.
- Maria, P., Edoardo, Z., Piergiorgio, N., Andrea, P., Denis, G., & Mauro, A. (2012). *In vitro* evaluation of sunscreens: An update for the clinicians. *ISRN Dermatology*, 2012, 1-4.
- Martín, M. A., Goya, L., & Ramos, S. (2013). Potential for preventive effects of cocoa and cocoa polyphenols in cancer. *Food Chemistry Toxicology*, 56, 336-351.
- Mathen, C., Thergaonkar, R., Teredesai, M., Soman, G, & Peter, S. (2014). Evaluation of anti-elastase and antioxidant activity in antiaging formulations containing *Terminalia* extracts. *International Journal of Herbal Medicine*, 2, 95-99.
- Matousek, J. L., Campbell, K. L., Kakoma, I., Solter, P. F., & Schaeffer, D. J. (2003). Evaluation of the effect of pH on *in vitro* growth of *Malassezia pachydermatis*. *Canadian Journal of Veterinary Research*, 67, 56-59.
- McCook, J. P. (2016). Topical products for the aging face. *Clinics in Plastic Surgery*, 43, 597-604.
- McCullough, J. L., & Kelly, K. M. (2006). Prevention and treatment of skin aging. *Annals of the New York Academy of Sciences*, 1067, 323-331.

- McGaw, L. J., Steenkamp, V., & Eloff, J. N. (2007). Evaluation of *Athrixia* bush tea for cytotoxicity, antioxidant activity, caffeine content and presence of pyrrolizidine alkaloids. *Journal of Ethnopharmacology*, 110, 16-22.
- Meda, A., Lamien, C. E., Romito, M., Millogo, J., & Nacoulma, O. G. (2005). Determination of the total phenolic, flavonoid and proline contents in Burkina Fasan Honey, as well as their radical scavenging activity. *Food Chemistry*, 91, 571-577.
- Mehta, C. R., & Patel, N. R. (1983). A network algorithm for performing Fisher's exact test in rxc contingency tables. *Journal of the American Statistical Association*, 78, 424-434.
- Meyer, W., Neurand, K., & Radke, B. (1981). Elastic fibre arrangement in the skin of the pig. *Archives of Dermatological Research*, 270, 391-401.
- Miao, Y. P., Wen, R., Aoshima, H., & Zhou, P. G. (2004). Synthesis and antioxidative activity of 2-substitutedphenyl-5-(3-indolyl)-oxazole derivatives. *Yao Xue Xue Bao*, 39, 37-40.
- Milam, E. C., & Rieder, E. A. (2016). An approach to cosmeceuticals. *Journal of Drugs in Dermatology*, 15, 452-456.
- Misnawi, Jinap, S., Nazamid, S., & Jamilah, B. (2002). Activation of remaining key enzymes in dried under-fermented cocoa beans and its effect on aroma precursor formation. *Food Chemistry*, 78, 407-417.
- Mitani, H., Ryu, A., Suzuki, T., Yamashita, M., Arakane, K., & Koide, C. (2007). Topical application of plant extracts containing xanthine derivatives can prevent UV induced wrinkle formation in hairless mice. *Photodermatology, Photoimmunology & Photomedicine*, 23, 86-94.
- Moein, M. R., Moein, S., & Ahmadizadeh, S. (2008). Radical scavenging and reducing power of *Salvia mirzayanii* subfractions. *Molecules*, 13, 2804-2813.
- Mohan, V., Talmi-Frank, D., Arkadash, V., Papo, N., & Sagi, I. (2016). Matrix metalloproteinase protein inhibitors: highlighting a new beginning for metalloproteinases in medicine, 3, 31-47.
- Montgomery, D. C. (2001). *Design and Analysis of Experiments*, fifth ed., Wiley, New York.
- Moradi, M. T., Karimi, A., Alidadi, S., & Ghasemi-Dehkordi, P. (2016). Cytotoxicity and *in vitro* antioxidant potential of *Quercus Brantii* acorn extract and the corresponding fractions. *International Journal of Pharmacognosy and Phytochemical Research*, 8, 558-562.
- Mordi, R. C., Walton, J. C., Burton, G. W., Hughes, L., Ingold, K. U., & Lindsay, D. A. (1991). Exploratory study of β -carotene autoxidation. *Tetrahedron Letters*, 32, 4203-4206.
- Morobe, I. C., Mthethwa, N. S., & Bisi-Johnson, M. A. (2012). Cytotoxic effects and safety profiles of extracts of active medicinal plants from South Africa. *Journal of Microbiology Research*, 2, 176-182.

- Mossman, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65, 55-63.
- MS 2200-1:2008 (2008). Islamic Consumer Goods-Part 1: Cosmetic and Personal Care-General Guidelines. Department of Standards Malaysia, Ministry Energy, Science, Technology, Environment and Climate Change (MESTECC), Malaysia.
- Mukta, S., & Adam, F. (2010) Cosmeceuticals in day-to-day clinical practice. *Journal of Drugs in Dermatology*, 9, 62-66.
- Mukherjee, P. K., Maity, N., Nema, N. K., & Sarkar, B. K. (2011). Bioactive compounds from natural resources against skin aging. *Phytomedicine*, 19, 64-73.
- Muñiz-Márquez, D. B., Martínez, G. C., Wong-Paz, J. E., & Belmares, R. (2013). Ultrasound-assisted extraction of phenolic compounds from *Laurus nobilis* L. and their antioxidant activity. *Ultrasonics Sonochemistry*, 20, 1149-1154.
- Muñiz-Márquez, D. B., Rodríguez, R., Balagurusamy, N., Carrillo, M. L., Belmares, R., Contreras, J. C., Nevárez, G. V., & Aguilar, C. N. (2013). Phenolic content and antioxidant capacity of extracts of *Laurus nobilis* L., *Coriandrum sativum* L. and *Amaranthus hybridus* L. *CyTA – Journal of Food*, pp. 1-6.
- Murakami, Y., Kawata, A., Katayama, T., & Fujisawa, S. (2015). Anti-inflammatory activity of the artificial antioxidants 2-tert-butyl-4-methoxyphenol (BHA), 2,6-di-tert-butyl-4-methylphenol (BHT) and 2,4,6-tri-tert-butylphenol (TBP), and their various combinations. *International Journal of Experimental and Clinical Pathophysiology and Drug Research*, 29, 197-206.
- Mustafar, M., Ismail, R. M., Othman, S. N., & Abdullah, R. (2018). A study on halal cosmetic awareness among Malaysian cosmetic manufacturers. *International Journal of Supply Chain Management*, 7, 492-496.
- Naczki, M., & Shahidi, F. (2006). Phenolics in cereals, fruits and vegetables: Occurrence, extraction and analysis. *Journal of Pharmaceutical and Biomedical Analysis*, 41, 1523-1542.
- Naik, J. P. (2001). Improved high-performance liquid chromatography method to determine theobromine and caffeine in cocoa and cocoa products. *Journal of Agriculture and Food Chemistry*, 49, 3579-3583.
- Narayana, K. R., Reddy, M. S., Chaluvadi, M., & Devarakonda, K. (2001). Bioflavonoids classification, pharmacological, biochemical effects and therapeutic potential. *Indian Journal of Pharmacology*, 33, 2-16.
- Nash, J. F., & Tanner, P. R. (2014). Relevance of UV filter/sunscreen product photostability to human safety. *Photodermatology, Photoimmunology & Photomedicine*, 30, 88-95.
- National Pharmaceutical Regulatory Agency (2016). Guidelines for control of cosmetics products in Malaysia, Ministry of Health, Malaysia.

- Natsume, M., Osakabe, N., Yamagishi, M., Takizawa, T., Nakamura, T., Miyatake, H., Hatano, T., & Yoshida, T. (2000). Analyses of polyphenols in cacao liquor, cocoa, and chocolate by normal-phase and reversed-phase HPLC. *Bioscience Biotechnology, and Biochemistry*, 64, 2581-2587.
- Nazaruddin, R., Seng, L. K., Hassan, O., & Said, M. (2006). Effect of pulp preconditioning on the content of polyphenols in cocoa beans (*Theobroma cacao*) during fermentation. *Industrial Crops and Products*, 24, 87-94.
- Ndiaye, M., Philippe, C., Mukhtar, H., & Ahmad, N. (2011). The grape antioxidant resveratrol for skin disorders: Promise, prospects and challenges. *Archives of Biochemistry and Biophysics*, 508, 164-170.
- Nenandis, N., Zhang, H. Y., & Tsimidou, M. Z. (2003). Structure-antioxidant activity relationship of ferulic acid derivatives: effect of carbon side chain characteristic group. *Journal of Agricultural and Food Chemistry*, 51, 1874-1879.
- Neukam, K., Stahl, W., Tronnier, H., Sies, H., & Heinrich, U. (2007). Consumption of flavanol-rich cocoa acutely increases microcirculation in human skin. *European Journal of Nutrition*, 46, 53-56.
- Ngoh, Y. -Y., & Gan, C. -Y. (2016). Enzyme-assisted extraction and identification of antioxidative and α -amylase inhibitory peptides from pinto beans (*Phaseolus vulgaris* cv. Pinto). *Food Chemistry*, 190, 331-337.
- Nichols, J., & Katiyar, S. (2010). Skin photoprotection by natural polyphenols: anti-inflammatory, antioxidant and DNA repair mechanisms. *Archives for Dermatological Research*, 302, 71-83.
- Niemnak, N., Rohsius, C., Elwers, S., Omokolo, D., & Lieberei, R. (2006). Comparative study of different cocoa (*Theobroma cacao* L.) clones in terms of their phenolics and anthocyanins contents. *Journal of Food Composition and Analysis*, 19, 612-619.
- Nixon, G. A., Tyson, C. A., & Wertz, W. C. (1975). Interspecies comparisons of skin irritancy. *Toxicology and Applied Pharmacology*, 31, 481-490.
- Noblesse, E., Cenizo, V., Bouez, C., & Damour, O. (2004). Lysyl oxidase-like and lysyl oxidase are present in the dermis and epidermis of a skin equivalent and in human skin and are associated to elastic fibers. *Journal of Investigative Dermatology*, 122, 621-630.
- Norshazila, S., Koy, C. N., Rashidi, O., Ho, L. H., Azrina, I., Nurul Zaizuliana, R. A., & Zarinah, Z. (2017). The effect of time, temperature and solid to solvent ratio on pumpkin carotenoids extracted using food grade solvents. *Sains Malaysiana*, 46, 231-237.
- Nunu, M. Louisa, N., Ketevan, J., Marine, D., & Eduard, C. (2008). Identification of free radicals induced by UV irradiation in collagen water solutions. *Journal of Photochemistry and Photobiology B*, 93, 61-65.

- Oboh, H. (2011). Total phenolics and antioxidant capacity of some Nigerian beverages. *Nigerian Journal of Basic and Applied Sciences*, 19, 68-75.
- Ohguchi, K., Itoh, T., Akao, Y., Inoue, H., Nozawa, Y., & Ito, M. (2010). SIRT 1 modulates expression of matrix metalloproteinases in human dermal fibroblasts. *British Journal of Dermatology*, 163, 689-694.
- Oliveira, A. C., Valentim, I. B., Silva, C. A., Berchara, E. J. H., Barros, M. P., Mano, C. M., & Goulart, O. F. (2009). Total phenolic content and free radical scavenging activities of methanolic extract powders of tropical fruit residues. *Food Chemistry*, 115, 469-475.
- Ozhogina, O. A., & Kasaikina, O. T. (1995). B-carotene as an interceptor of free radicals. *Free Radical Biology and Medicine*, 19, 575-581.
- Panche, A. N., Diwan, A. D., & Chandra, S. R. (2016). Flavonoids: an overview. *Journal of Nutritional Science*, 5, 1-15.
- Parvez, S., Kang, M., Chung, H. S., Cho, C., Hong, M. C., Shin, M. K., & Bae, H. (2006). Survey and mechanism of skin depigmenting and lightening agents. *Phytotherapy Research*, 20, 921-934.
- Patel, V. R., Patel, P. P., & Kujal, S. S. (2010). Antioxidant activity of some selected medicinal plants in western Region of India. *Advances in Biological Research*, 4, 23-26.
- Patwardhan, J., & Bhatt, P. (2015). Ultraviolet-B protective effect of flavonoids from *Eugenia caryophyllata* on human dermal fibroblast cells. *Pharmacognosy Magazine*, 11, 397-406.
- Pauwels, M., & Rogiers, V. (2004). Safety evaluation of cosmetics in the EU reality and challenges for the toxicologist. *Toxicology Letters*, 151, 7-17.
- Peng, W. L., Khanafi, M. A., Setapar, S. H. M., Idham, Z., Yunus, M. A. C., & Abbas, M. Z. A. (2014). Development of emulsification containing natural colorant from local plant (Roselle). *Jurnal Teknologi (Sciences & Engineering)*, 69, 15-17.
- Phillips, L., Steinbert, M., Maibach, H. I., & Akers, W. A. (1972). A comparison of rabbit and human skin responses to certain irritants. *Toxicology and Applied Pharmacology*, 21, 369-382.
- Pinelo, M., Rubilar, M., Jerez, M., Sineiro, J., & Nunez, M. J. (2005). Effect of solvent, temperature, and solvent-to-solid ratio on the total phenolic content and antiradical activity of extracts from different components of grape pomace. *Journal of Agricultural and Food Chemistry*, 53, 2111-2117.
- Pittayapruek, P., Meehansan, J., Prapapan, O., Komine, M., & Ohtsuki, M. (2016). Role of matrix metalloproteinases in photoaging and photocarcinogenesis. *International Journal of Molecular Sciences*, 17, 1-20.

- Plaza, M., Oliveira, D., Nilsson, A., & Turner, C. (2017). Green and efficient extraction method to determine polyphenols in cocoa and cocoa products. *Food Analytical Methods*, 10, 2677-2691.
- Poljšak, B., & Dahmane, R. (2012). Free radicals and extrinsic skin aging. *Dermatology Research and Practice*, 2012, 1-4.
- Potekaev, N., & Zhukova, O. (2013). Evaluation of safety and efficacy of the Maximus™ system for facial wrinkles. *Journal of Cosmetics, Dermatological Sciences and Applications*, 3, 151-156.
- Prasad, M. P. (2014). *In vitro* phytochemical analysis and antioxidant activity of seeds belonging to *Cucurbitaceae* family. *Indian Journal of Advances in Plant Research*, 1, 13-18.
- Pulido, R., Bravo, L., & Saura-Calixto, F. (2000). Antioxidant activity of dietary polyphenols as determined by a modified ferric reducing/antioxidant power assay. *Journal of Agricultural and Food Chemistry*, 48, 3396-3402.
- Pura N. J. (2001). Improved high-performance liquid chromatography method to determine theobromine and caffeine in cocoa and cocoa products. *Journal of Agricultural and Food Chemistry*, 49, 3579-3583.
- Quan, T., Wang, F., Shao, Y., Rittie, L., Xia, W., Orringer, J. S., Voorhees, J. J., & Fisher, G. J. (2013). Enhancing structural support of the dermal microenvironment activates fibroblasts, endothelial cells and keratinocytes in aged human skin *in vivo*. *Journal of Investigative Dermatology*, 658-667.
- Raissi, S. (2009). Multivariate process capability indices on the presence of priority for quality characteristics. *Journal of Industrial Engineering International*, 5, 27-36.
- Raj, D., Brash, D. E., & Grossman, D. (2006). Keratinocyte apoptosis in epidermal development and disease. *Journal of Investigative Dermatology*, 126, 243-257.
- Ramachandran, G. N., & Kartha, G. (1955). Structure of collagen. *Nature*, 176, 593-595.
- Ramli, N. (2006). Influence of roasting conditions on volatile flavour of roasted Malaysian cocoa beans. *Journal of Food Processing and Preservation*, 78, 281-298.
- Ramos, L., Kristenson, E. M., & Brinkman, U. A. T. (2002). Current use of pressurized liquid extraction and subcritical water extraction in environmental analysis. *Journal of Chromatography A*, 975, 3-29.
- Rasul, A., & Akhtar, N. (2011). Formulation and *in vitro* evaluation for anti-aging effects of an emulsion containing basil extract using non-invasive biophysical techniques. *Journal of Pharmaceutical Sciences*, 19, 344-350.
- Rice-Evans, C. A., Miller, N. T., & Paganga, G. (1997). Antioxidant properties of phenolic compounds. *Trends in Plant Science*, 4, 304-309.

- Richelle, M., Tavazzi, I., Enslin, M., & Offord, E. A. (1999). Plasma kinetics in man of epicatechin from black chocolate. *European Journal of Clinical Nutrition*, 53, 22-26.
- Richter, B. E. (2000). Extraction of hydrocarbon contamination from soils using accelerated solvent extraction. *Journal of Chromatography A*, 874, 217-224.
- Rios, L. Y., Gonthier, M. P., Remesy, C., Mila, I., Lapiere, C., Lazarus, S. A., Williamson, G., & Scalbert, A. (2003). Chocolate intake increases urinary excretion of polyphenol-derived phenolic acids in healthy human subjects. *The American Journal of Clinical Nutrition*, 77, 912-918.
- Robinson, M. K., Cohen, C., de Brugerolle, de Fraissinette, Ponc, M., Whittle, E., & Fentem, J. H. (2002). Non-animal testing strategies for assessment of the skin corrosion and skin irritation potential of ingredients and finished products. *Food and Chemical Toxicology*, 40, 573-592.
- Robinson, M. K., McFadden, J. P., & Basketter, D. A. (2001). Validity and ethics of the human 4-h patch test as an alternative method to assess acute skin irritation potential. *Contact Dermatitis*, 45, 1-12.
- Roselló-Soto, E., Martí-Quijal, F. J., Cilla, A., Muneke, P. E. S., Lorenzo, J. M., Remize, F., & Barba, F. J. (2019). Influence of temperature, solvent and pH on the selective extraction of phenolic compounds from Tiger Nuts by-products: Triple-TOF-LC-MS-MS characterization. *Molecules*, doi:10.3390/molecules 24040797.
- Ryu, H. S., Joo, Y. H., Kim, S. O., & Youn, S. W. (2008). Influence of age and regional differences on skin elasticity as measured by cutometer (R). *Skin Research and Technology*, 14, 354-358.
- Rzepka, Z., Buszman, E., Beberok, A., & Wrześniak, D. (2016). From tyrosine to melanin: Signaling pathways and factors regulating melanogenesis. *Postępy higieny i medycyny doświadczalnej*, 70, 695-708.
- Saadiah, S. (2009). Safety assessment: available local expertise. ASEAN Cosmetic Directive Fostering Industry Compliance Seminar, 19 Oct, National Pharmaceutical Control Bureau and The Cosmetic Toiletries and Fragrance Association of Malaysia.
- Sabahannur, S., Alimuddin, S., & Rahmawati (2018). Changes in phenol level and antioxidant activity of cocoa beans during fermentation and roasting. *Journal of Food Research*, 7, 23-29.
- Sahu, M., Verma, D., & Haris, K. (2014). Phytochemical analysis of the leaf, stem and seed extracts of *Cajanus cajan* L. (dicotyledoneae: Fabaceae).
- Salamanca, C. H., Barrera-Ocampo, A., Lasso, J. C., Camacho, N., & Yarce, C. J. (2018). Franz Diffusion Cell approach for pre-formulation characterisation of ketoprofen semi-solid dosage forms. *Pharmaceutics*, 148, 1-10.

- Sánchez-Ferrer, A., Rodríguez-López, J. N., García-Cánovas, F., & García-Carmona, F. (1995). Tyrosinase: a comprehensive review of its mechanism. *Biochimica et Biophysica Acta*, 1247, 1-11.
- Sanchez-Rabaneda, F., Jauregui, O., Casals, I., Andres-Lacueva, C., Izquierdo-Pulido, M., & Lamuela-Raventos, R.M. (2003). Liquid chromatographic/ electro spray ionization tandem mass spectrometric study of the phenolic composition of cocoa (*Theobroma cacao*). *Journal of Mass Spectrometry*, 38, 35-42.
- Sari, A. B. T., Mahriani, Tiningrum, G. A. P., Wahyudi, T., & Misnawi (2015). Cocoa extract indicated has activity on selectively killing breast cancer cells. *The Journal of Tropical Life Science*, 5, 128-132.
- Scapagnini, G., Davinelli, S., Di Renzo, L., Olarte, H. H., Micali, G., Cicero, A. F., & Gonzalez, S. (2014). Cocoa bioactive compounds: significance and potential for the maintenance of skin health. *Nutrients*, 6, 3202-3213.
- Schmid-Wendtner, M. -H., & Korting, H. C. (2006). The pH of the skin surface and its impact on the barrier function. *Skin Pharmacology and Physiology*, 19, 296-302.
- Schurer, N. Y., & Elias, P. M. (1991). The biochemistry and function of stratum corneum lipids. *Advances in Lipid Research*, 24, 27-56.
- Séhédic, D., Hardy-Boismartel, A., Couteau, C., & Coiffard, L. J. M. (2009). Are cosmetic products which include an SPF appropriate for daily use? *Archives of Dermatological Research*, 301, 603-608.
- Seo, S. Y., Sharma, V. K., & Sharma, N. (2003). Mushroom tyrosinase: Recent prospects. *Journal of Agricultural and Food Chemistry*, 51, 2837-2853.
- Shai, L. J., Mcgaw, L. J., Masoko, P., & Eloff, J. N. (2008). Antifungal and antibacterial activity of seven traditionally used South African plant species active against *Candida albicans*. *South African Journal of Botany*, 74, 677-684.
- Shibayama, H., Hisama, M., Matsuda, S., Kawase, A., Ohtsuki, M., Hanada, K., & Iwaki, M. (2008). Effect of a novel ascorbic derivative, disodium isostearyl 2-O-L-Ascorbyl Phosphate, on normal human dermal fibroblasts against reactive oxygen species. *Bioscience, Biotechnology, and Biochemistry*, 72, 1015-1022.
- Sieniawska, E., Los, R., Baj, T., & Malm, A. (2013). Antimicrobial efficacy of *Mutellina purpurea* essential oil and α -pinene against *Staphylococcus epidermis* grown in planktonic and biofilm cultures. *Industrial Crops and Products*, 51, 152-157.
- Silva, E. M., Rogez, H., & Larondelle, Y. (2007). Optimization of extraction of phenolics from *Inga edulis* leaves using response surface methodology. *Separation and Purification Technology*, 55, 381-387.
- Solano, F., Briganti, S., Picardo, M., & Ghanem, G. (2006). Hypopigmenting agents: an updated review on biological, chemical and clinical aspects. *Pigment Cell Research*, 19, 550-571.

- Souza, P. M., Sales, P. M., Simeoni, L. A., Silva, E. C., Silveira, D., & Magalhães, P. O. (2012). Inhibitory activity of α -amylase and α -glucosidase by plant extracts from the Brazilian cerrado. *Planta Medica*, 78, 393-399.
- Spigno, G., & De Faveri, D. M. (2007). Antioxidants from grape stalks and marc: influence of extraction procedure on yield, purity and antioxidant power of the extracts. *Journal of Food Engineering*, 78, 793-801.
- Spigno, G., Tramelli, L., & Dante Marco, D. F. (2007). Effects of extraction time, temperature and solvent on concentration and antioxidant activity of grape marc phenolics. *Journal of Food Engineering*, 81, 200-208.
- Spindola, D. G., Hinsberger, A., Antunes, V. M. de S., Michelin, L. F. G., Bincoletto, C., & Oliveira, C. R. (2018). *In vitro* cytotoxicity of chemical preservatives on human fibroblast cells. *Brazilian Journal of Pharmaceutical Sciences*, 54, 1-9.
- Statista (2019). Available from www.statista.com/statistics/750779/natural-organic-beauty-market-worldwide
- Sun, T., & Ho, C. T. (2005). Antioxidant activities of buckwheat extracts. *Food Chemistry*, 90, 743-749.
- Sunarintyas, S., Siswomihardjo, W., & Tontowi, A. E. (2012). Cytotoxicity of *Cricula trifenestrata* cocoon extract on human fibroblasts. *International Journal of Biomaterials*, doi:10.1155/2012/493075.
- Sutivisedsak, N., Cheng, H. N., Willett, J. L., & Lesch, W. C. (2010). Microwave-assisted extraction of phenolics from bean (*Phaseolus vulgaris* L.). *Food Research International*, 43, 516-519.
- Svobodová, A., Psotová, J., & Walterová, D. (2003). Natural phenolics in the prevention of UV-induced skin damage. A review. *Biomedical Papers*, 147, 137-145.
- Tabart, J., Kevers, C., Sipel, A., Pincemail, J., Defraigne, J. O., & Dommes, J. (2007). Optimisation of extraction of phenolics and antioxidants from black currant leaves and buds and of stability during storage. *Food Chemistry*, 105, 1268-1275.
- Talib, W. H., & Mahasneh, A. M. (2010). Antiproliferative activity of plant extracts used against cancer in traditional medicine. *Scientia Pharmaceutica*, 78, 33-45.
- Tan, M. C., Tan, C. P., Khoo, H. E., & Ho, C. W. (2014). Optimization for extraction on total phenolic content and radical scavenging capacity of Henna (*Lawsonia inermis*) stems using response surface methodology. *International Food Research Journal*, 21, 789-794.
- Tan, S. P., Stathopoulos, C., Parks, S., & Roach, P. (2014). An optimised aqueous extract of phenolic compounds from bitter melon with high antioxidant capacity. *Antioxidants*, 3, 814-829.
- Tansirikongkol, A., Pientaweeratch, S., & Panapisal, V. (2016). Antioxidant, anti-collagenase and anti-elastase activities of *Phyllanthus emblica*,

- Manilkara zapota* and silymarin: an in vitro comparative study for anti-aging applications. *Pharmaceutical Biology*, 54, 1865-1872.
- Tapas, A. R., Sakarkar, D. M., & Kakde, R. B. (2008). Flavonoids as nutraceuticals: a review. *Tropical Journal of Pharmaceutical Research*, 7, 1089-1099.
- Tarahovsky, Y. S., Kim, Y. A., Yagolnik, E. A., & Muzafarov, E. N. (2014). Flavonoid-membrane interactions: Involvement of flavonoid-metal complexes in raft signaling. *Biochimica et Biophysica Acta*, 1838, 1235-1246.
- Tebbe, B. (2001). Relevance of oral supplementation with antioxidants for prevention and treatment of skin disorders. *Skin Pharmacology and Applied Skin Physiology*, 14, 296-302.
- Thomas, J. B., James, J. H., Schantz, M. M., Porter, B. J., & Sharpless, K.E. (2004). Determination of caffeine, theobromine, and theophylline in standard reference material 2384, baking chocolate, using reversed-phase liquid chromatography. *Journal of Agricultural and Food Chemistry*, 52, 3259-3263.
- Thring, T. S. A., Hili, P., & Naughton, D. P. (2009). Anti-collagenase, anti-elastase and anti-oxidant activities of extracts from 21 plants. *BMC Complementary and Alternative Medicine*, 9, 1-11.
- Timbie, D. J., Sechrist, L., & Kenney, P. G. (1978). Application of HPLC to the study of variables affecting theobromine and caffeine concentrations in cocoa beans. *Journal of Food Science*, 43, 560-565.
- Tomas-Barberan, F., Cienfuegos-Jovellanos, E., Marin, A., Mugarza, B., Gil-Izquierdo, A., & Cerda, B. (2007). A new process to develop a cocoa powder with higher flavonoid monomer content and enhanced bioavailability in healthy human. *Journal of Agricultural Food and Chemistry*, 55, 3926-3935.
- Tomšik, A., Pavlic, B., Vlastic, J., & Ramić, M. (2015). Optimization of ultrasound-assisted extraction of bioactive compounds from wild garlic (*Allium ursinum* L.). *Ultrasonics Sonochemistry*, 29, 502-511.
- Truong, D. -H., Nguyen, D. H., Ta, N. T. A., Bui, A. V., Do, T. H., & Nguyen, H. C. (2019). Evaluation of the use of different solvents for phytochemical constituents, antioxidants, and *in vitro* anti-inflammatory activities of *Severinia buxifolia*. *Journal of Food Quality*, <https://doi.org/10.1155/2019/8178294>
- Turan, B., Tuncay, E., & Vassort, G. (2012). Resveratrol and diabetic cardiac function: Focus on recent *in vitro* and *in vivo* studies. *Journal of Bioenergetics and Biomembranes*, 44, 281-296.
- Turker, N., & Erdogdu, F. (2006). Effects of pH and temperature of extraction medium on effective diffusion coefficient of anthocyanin pigments of black carrot (*Daucus carota* var. L.). *Journal of Food Engineering*, 76, 579-583.

- Tyug, T. S., Prasad, K. N., & Amin, I. (2010). Antioxidant capacity, phenolics and isoflavones in soybean by-products. *Food Chemistry*, 123, 583-589.
- Udompataikul, M., Sripiroj, P., & Palungwachira, P. (2009). An oral nutraceutical containing antioxidants, minerals and glycosaminoglycans improves skin roughness and fine wrinkles. *International Journal of Cosmetic Science*, 31, 427-435.
- Uitto, J. (2008). The role of elastin and collagen in cutaneous aging: Intrinsic aging versus photoexposure. *Journal of Drugs in Dermatology*, 7, 12-16.
- Valdez-Ortiz, A., German-Baez, L. J., Valdez-Flores, M., Figueroa-Perez, M. G., Garduno-Felix, K. G., Valdez-Ortiz, R., & Meza-Ayala, K. A. (2017). Anti-aging and nutraceutical characterization of plant infusions used in traditional medicine. *Pakistan Journal of Nutrition*, 16, 285-292.
- Varma, S., Mishra, A., Vijayakumar, M., & Paramesh, R. (2017). Anti-skin ageing phytochemicals in cosmetics: an appraisal. *Cosmetic Active Ingredients*, 12, 20-23.
- Vasisht, K., Sharma, N., & Karan, M. (2016) Current Perspective in the International Trade of Medicinal Plants Material: an Update. *Current Pharmaceutical Design*, 22, 4288-4336.
- Vatai, T., Škerget, M., & Knez, Z. (2009). Extraction of phenolic compounds from elder berry and different grape marc varieties using organic solvents and/or supercritical carbon dioxide. *Journal of Food Engineering*, 90, 246-254.
- Velioglu, Y. S., Mazza, G., Gao, L., & Oomah, B. D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables and grain products. *Journal of Agricultural and Food Chemistry*, 46, 4113-4117.
- Vig, K., Chaudhari, A., Tripathi, S., Dixit, S., Sahu, R., Pillai, S., Dennis, V. A., & Singh, S. R. (2017). Advances in skin regeneration using tissue engineering. *International Journal of Molecular Sciences*, 18, 1-19.
- Vijayakumar, R., Ghani, S. S. A., & Mokhtar, N. F. M. (2017). Anti-elastase, anti-collagenase and antimicrobial activities of the underutilized red pitaya peel: an *in vitro* study for anti-aging applications. *Asian Journal of Pharmaceutical and Clinical Research*, 10, 251-255.
- Visscher, M., Robinson, M., & Wickett, R. (2011). Stratum corneum free amino acids following barrier perturbation and repair. *International Journal of Cosmetic Science*, 33, 80-89.
- Visse, R., & Nagase, H. (2003). Matrix metalloproteinases and tissue inhibitors of metalloproteinases: structure, function and biochemistry. *Circulation Research*, 92, 827-839.
- Wang, B. -E., Liu, C. -T., Tseng, C. -Y., & Yu, Z. -R. (2005). Antioxidant activity of *Bupleurum kanoi* Liu (Chao et Chuang) fractions fractionated

- by supercritical CO₂. LWT – Food Science and Technology, 38, 281-287.
- Wang, Y., Deng, L., & Fan, Y. (2018). Preparation of soy-based adhesive enhanced by waterborne polyurethane: optimization by response surface methodology. *Advances in Materials Science and Engineering*, doi.org/10.1155/2018/9253670.
- Wen, K. C., Chiu, H. H., Fan, P. C., Chen, C. W., Wu, S. M., Chang, J.H., & Chiang, H. M. (2011). Antioxidant activity of *Ixora parviflora* in a cell/cell-free system and in UV-exposed human fibroblasts. *Molecules*, 16, 5732-5752.
- Wollgast, J., & Anklam, E. (2000). Review on polyphenols in *Theobroma cacao*: changes in composition during the manufacture of chocolate and methodology for identification and quantification. *Food Research International*, 33, 423-447.
- Wollgast, J. (2004). The contents and effects of polyphenols in chocolate. Dissertation for obtaining the degree of doctor of The Faculty of Agricultural and Nutritional Sciences, Home Economics and Environmental Management at The University of GieBen, Germany.
- Wong, W. S., & McLean, A. E. M. (2000). Effects of phenolic antioxidants and flavonoids on DNA synthesis in rat liver, spleen, and testis *in vitro*. *Toxicology*, 139, 243-253.
- Xiao, W., Han, L., & Shi, B. (2008). Optimization of microwave-assisted extraction of flavonoid from *Radix astragali* using response surface methodology. *Separation Science and Technology*, 43, 671-681.
- Xie, L-P., Chen, Q-X., Huang, H., Wang, H-Z., & Zhang, R-Q. (2003). Inhibitory effects of some flavonoids on the activity of mushroom tyrosinase. *Biochemistry (Moscow)*, 68, 487-491.
- Xu, Y., & Fisher, G. J. (2005). Ultraviolet (UV) light irradiation induced signal transduction in skin photoaging. *Journal of Dermatological Science Supplement*, 1, 1-8.
- Yeum, K. J., Smith, A. F., Krinsky, N. I., Russell, R. M., & Dos, A. L. (2000). The effect of α -tocopherol on the oxidative cleavage of β -carotene. *Free Radical Biology and Medicine*, 29, 105-114.
- Yen, G. -C., & Chen, H. -Y. (1995). Antioxidant activity of various tea extracts in relation to their antimutagenicity. *Journal of Agricultural and Food Chemistry*, 43, 27-32.
- Yilmaz, Y., & Toledo, R. T. (2006). Oxygen radical absorbance capacities of grape/ wine industry by-products and effect of solvent type on extraction of grape seed polyphenols. *Journal of Food Composition and Analysis*, 19, 41-48.
- Yokozawa, T., Kim, H. Y., Kim, H. J., Tanaka, T., Sugino, H., Okubo, T., Chu, D. C., & Juneja, L. R. (2007). Amla (*Embllica officinalis* Gaertn.) attenuates age-related renal dysfunction by oxidative stress. *Journal of Agricultural and Food Chemistry*, 55, 7744-7752.

- Yoshimatsu, H. (2006). Collective action problems and regional integration in ASEAN. *Contemporary Southeast Asia A Journal of International and Strategic Affairs*, 28, 115-140.
- Zakaria, Z. (2015). Regulation of cosmetics: what has Malaysia learnt from the European system? *Journal of Consumer Policy*, 38, 39-59.
- Zion Market Research (2018). Available from <https://zionmarketresearch.com>
- Zhao, C. F., Li, S., Li, S. J., Song, G. H., Yu, L. J., & Zhang, H. (2013). Extraction optimization approach to improve accessibility of functional fraction based on combination of total polyphenol, chromatographic profiling and antioxidant activity evaluation: *Pyracantha fortuneana* fruit as an example. *Journal of Functional Foods*, 5, 715-728.
- Zhang, C., & Kim, S-K. (2009). Matrix metalloproteinase inhibitors (MMPi)s from marine natural products: the current situation and future prospects. *Marine Drugs*, 7, 71-84.
- Zhang, P., Chen, J., Li, T., & Zhu, Y. Y. (2013). Use of small RNA as anti-ageing cosmeceuticals. *Journal of Cosmetic Science*, 64, 455-468.
- Zumbe, A. (1998). Polyphenols in cocoa: Are there health benefits? *British National Formulary. Nutrition Bulletin*, 23, 94-102.