

UNIVERSITI PUTRA MALAYSIA

TRANSFORMATION OF TOBACCO (*Nicotiana tabacum*) WITH THE OIL PALM CINNAMYL ALCOHOL DEHYDROGENASE 1 (CAD1) GENE

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Thesis Submitted to the Department of Cell and Molecular Biology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Bachelor of Science

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TRANSFORMATION OF TOBACCO (Nicotiana tabacum) WITH THE OIL PALM CINNAMYL ALCOHOL DEHYDROGENASE 1 (CAD1) GENE

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Lignification in plants occurs via the lignin biosynthetic pathway that involves a series of enzymes; among these enzymes is the cinnamyl alcohol dehydrogenase (CAD). CAD catalyzes the conversion of hydroxycinnamaldehydes into monolignols. In oil palm, two copies of the CAD gene have been identified (EgCAD1 and *EgCAD2*) and their functions are largely undetermined. Studying gene function in oil palm is generally time-consuming and technically difficult. Hence, the study of the CAD1 gene from oil palm was performed via transformation of tobacco (Nicotiana *tabacum*) due to its relatively short life cycle and well-established transformation protocol. Tobacco leaf discs were used as target samples for Agrobacteriummediated transformation of pMDC32-EgCAD1 which carries a T-DNA region consisting of the oil palm CAD1 gene. Phylogeny study of the EgCAD family was also performed to establish the relationship of EgCAD1 with other members of the CAD family across different plant orders. The oil palm CAD1 shares a common ancestor with their counterparts in the angiosperm and gymnosperm lineages. The CAD phylogeny showed that EgCAD1 was clustered together with the genes from rice, pinus and banana, and they share the same recent common ancestor. The oil

palm *CAD1* was successfully integrated into the tobacco genome where positive PCR data showed a genomic region comprising the *EgCAD1* in the p*MDC32-CAD1* construct was amplified in the transformed tobacco plants. Normal phenotype of the transformed plants was observed. This study confirmed the successful integration of the *EgCAD1* into the genome of tobacco and the establishment of *CAD* family phylogeny. The findings serve as a basis for functional characterization of *EgCAD1* in the future.



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TRANSFORMASI TEMBAKAU DENGAN GEN CINNAMYL ALKOHOL DEHIDROGENASE 1 (CAD1) DARIPADA KELAPA SAWIT

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Lignifikasi dalam tumbuhan berlaku melalui tapak jalan biosintesis lignin yang melibatkan satu siri enzim; di antara enzim ini adalah cinnamyl alkohol dehidrogenase (CAD). CAD memangkinkan penukaran dari hidrosicinnamyldehida ke monolignol. Dalam kelapa sawit, dua salinan gen CAD telah dikenal pasti (EgCAD1 dan EgCAD2) dan fungsi gen ini masih belum ditentukan. Penyelidikan fungsi gen dalam kelapa sawit biasanya memakan masa dan sukar dari segi teknikal. Oleh itu, kajian gen CADI dari kelapa sawit dilakukan melalui transformasi tembakau (Nicotiana tabacum) disebabkan oleh kitaran hidupnya yang singkat dan protokol transformasi yang mantap. Kajian filogeni keluarga CAD juga dilakukan untuk mengkaji hubungan EgCAD1 dengan ahli-ahli lain keluarga CAD dan order tumbuhan yang berbeza. Cakera daun tembakau telah digunakan sebagai sampel untuk transformasi Agrobacterium-berantarakan pMDC32-EgCAD1 yang membawa rantau T-DNA terdiri daripada gen CAD1 kelapa sawit. Gen kelapa sawit CAD1 berkongsi moyang yang sama dengan rakan mereka dari keturunan angiosperma dan gimnosperma. Filogeni CAD menunjukkan EgCAD1 telah digolongkan bersama dengan gen daripada padi, Pinus dan pisang; dan mereka berkongsi moyang yang sama. Gen *CAD1* dari kelapa sawit telah berjaya diintegrasikan ke dalam genom tembakau di mana data PCR positif menunjukkan rantau genomik yang terdiri daripada sempadan EgCAD1 telah dapat diamplifikasikan dalam pokok tembakau. Tumbuh-tumbuhan yang telah ditransfomasikan mempunyai fenotip yang normal. Kajian ini mengesahkan kejayaan integrasi EgCAD1 ke dalam genom tembakau dan penubuhan filogeni keluarga gen *CAD*. Penemuan ini menjadi asas bagi pencirian kefungsian EgCAD1 pada masa akan datang.



This thesis was submitted to the Department of Cell and Molecular Biology, Faculty of Biotechnology and Biomolecular Sciences and has been accepted as fulfillment of the requirement for the degree of Bachelor of Science (Hons). The member of the Supervisory Committee was as follows:

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LIST OF ABBREVIATIONS

	α	Alpha
	g	Gravitational acceleration
	μg	Microgram
	μL	Microliter
	A. tumefaciens	Agrobacterium tumefaciens
	BAP	Benzylaminopurine
	BLAST	Basic Local Alignment Search Tool
	Вр	Base Pair
	CAD	Cinnamyl alcohol dehydrogenase
	Cl	Chloride
	cm	Centimeter
	DNA	Deoxyribonucleic acid
	dNTPs	deoxyribonucleotides
	EDTA	Ethylenediaminetetraacetic acid
	LB	Luria Bertani
	М	Molar
	MEGA	Molecular Evolutionary Genetic Analysis
	MgCl ₂	Magnesium chloride
	mL	Milliliter
	mM	Millimolar
	MS	Murashige and Skoog basal Medium
	MUSCLE	Multiple Sequence Comparison by Log-Expectation
	NaOH	Sodium hydroxide
	NCBI	National Center for Biotechnology Information
	ng	Nanogram
	OD	Optical density
	PCR	Polymerase chain reaction

rpm	revolutions per minute
SDS	Sodium dodecyl sulphate
TAE	Tris acetate EDTA
TE	Tris-EDTA
Tm	melting temperature
Tris	Tris (hydroxymethyl) aminomethane
Tris-HCl	Tris-hydrochloride
U	Unit
UV	Ultraviolet
v	Volt
v/v	volume per volume
w/v	weight per volume

CHAPTER 1

INTRODUCTION

Lignification in plants is gaining broadening interests from various agricultural and biotechnology fields due to its importance in garnering economic value especially in terms of recalcitrance of plant biomass. Currently, plant lignification is believed to be part of plant's responses towards biotic and abiotic stresses as well as plant developmental process (Humphreys and Chapple, 2002). Lignification in plants occurs via lignin biosynthetic pathway, where the deposition and polymerization of different types of monolignols form lignin (Whetten and Sederoff, 1995).

The lignin biosynthetic pathway involves several key enzymes; among them is the cinnamyl-alcohol dehydrogenase (CAD). It belongs to the oxidoreductase family and it catalyzes the reversible conversion of p-hydroxycinnamaldehydes to monolignols. In *Arabidopsis*, it is believed that *CAD-c* and *CAD-d* are the primary genes that supply coniferyl and sinapyl alcohols to biosynthesize lignin (Sibout *et. al.*, 2005). Sibout *et al.* (2005) discovered that a *cad-c cad-d* double mutant of *Arabidopsis* has limp floral stem and abnormal pattern of lignin staining, with 40% loss of lignin content in the mutant plant as compared to the wild type. In oil palm, *CAD* was first discovered for its role in the differentiation of the endocarp and mesocarp tissues of the dura variety (Bhasker and Mohankumar, 2001). This revealed the relationship of *CAD* from oil palm with other enzymes in the lignin biosynthetic pathway by comparing the expression levels of *CAD*, *PAL* and *POD* during the development of fruit shell (Bhasker and Mohankumar, 2001).

To date, several classes of *CAD* have been identified across different plant species. In *Camellia sinensis*, deep sequencing of the transcriptome revealed three types of *CAD* with only 20%-54% sequence identities. This showed that the *CAD* genes were of different classes, or exist in a multi-gene family (Deng *et al.*, 2013). However, the function of the each different classes of *CAD* is yet to be determined, and among different species. In *Oryza sativa*, twelve distinct genes were found to encode products that are highly similar to CAD. *OsCAD2*, an orthologue of *CAD* was found to participate in promoter-reporter gene fusions in the onset of lignification (Tobias and Chow, 2005).

Due to the nature of the *CAD* genes that tend to form a multi-gene family, identifying a specific *CAD* gene responsible for lignifications in oil palm is difficult. Previously, two copies of *CAD* gene namely *EgCAD1* and *EgCAD2* have been isolated from oil palm. The aim to single out the specific *CAD* gene for lignification in oil palm is made especially difficult when no prior information about the gene is available in the plant. One approach in which the gene function can be predicted is through phylogenetic analysis of the gene families from different groups of plant. Preliminary expression analysis of *EgCAD1* indicated that the gene is expressed in the oil palm root, leaf and stem. This information helps to narrow down the possible functional characteristics of *EgCAD1*. Since functional analysis of genes in oil palm is technically difficult and time-consuming; thus, tobacco is a good plant candidate for studying the basic function of *EgCAD1* in plant lignification. The current study encompasses the transformation of tobacco with *EgCAD1* and phylogenetic analysis of the *CAD* gene family specific objectives:

i. To establish the phylogeny of the oil palm *CAD1* gene via maximum likelihood method;

ii. To transform tobacco leaf discs with the oil palm *CAD1* gene via *Agrobacterium*-mediated transformation.



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