

FLORAL STRUCTURE AND POLLEN LOAD IN RELATION TO FRUIT DEVELOPMENT AND PHYSICO-CHEMICAL CHARACTERISTICS OF DRAGON FRUIT [Hylocereus polyrhizus (F.A.C Weber) Britton & Rose] AT HARVEST

JOANNA CHO LEE YING

FP 2011 57



FLORAL STRUCTURE AND POLLEN LOAD IN RELATION TO FRUIT DEVELOPMENT AND PHYSICO-CHEMICAL CHARACTERISTICS OF DRAGON FRUIT [*Hylocereus polyrhizus* (F.A.C Weber) Britton & Rose] AT HARVEST



JOANNA CHO LEE YING

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

April 2011

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

FLORAL STRUCTURE AND POLLEN LOAD IN RELATION TO FRUIT DEVELOPMENT AND PHYSICO-CHEMICAL CHARACTERISTICS OF DRAGON FRUIT [*Hylocereus polyrhizus* (F.A.C Weber) Britton & Rose] AT HARVEST

By

JOANNA CHO LEE YING

April 2011

Chairman Faculty : Associate Professor Phebe Ding, PhD : Agriculture

In Malaysia, dragon fruits are graded and priced according to their fruit weight. As dragon fruit is a type of berry, its fruit weight depends on the number of developing seeds. In order to obtain better quality and yield in term of fruit weight, a study was conducted on the floral structure and pollen load on fruit development of red-fleshed dragon fruit (Hylocereus polyrhizus). The first objective was to determine the pollination events of H. polyrhizus. Light and scanning electron microscopy (SEM) were used to examine pollen and stigma morphology and pollen-stigma interaction of red-fleshed dragon fruit at 0, 1, 2, 3 and 4 day after pollination (DAP). Fluorescence microscopy was used to detect pollen tube growth stained with aniline blue in the style at 0, 2 and 4 DAP. The flowers of *H. polyrhizus* bloomed at night in an upright position with a spatial different between the anthers and the stigma suggesting cross-pollination. The pollen grains were categorized as large with polar axis (P) to equatorial diameter (E) ratio of 0.97, oblate spheroidal-shaped and trizonocolpate with the exine covered with echinates. The stigma had multiple lobes ($n = 28 \pm 3$) and belonged to the dry type. The style is long $(27 \pm 2.54 \text{ cm})$ and solid with one hollow canal. Pollen grains germinated within 2 h after landing on the receptive stigma and pollen tubes grow within the transmitting tract located beneath the hollow canal of the style to the ovary. Starch granules within the style tissue depleted as DAP progressed. Findings indicated that a few pollen tubes had started to arrive in the ovary cavity after 2 DAP. Successful fertilization of male gametes (from pollen tube) with female gamete (ovule) led to fruit set. The second objective was to evaluate the changes in cellular structure using light microscopy and SEM during fruit development of H. polyrhizus at every 5 DAP. Starch pattern during fruit development was examined using iodine-potassium iodide (I₂/KI). After successful fertilization, microscopic studies revealed that fruit development occurred in two distinct period of growth of cell division and cell expansion especially in the pulp region. Initially, parenchyma cells were small and rigid but loosen as DAP progressed. The early phase of cell division occurred in the first 20 DAP followed by cell enlargement thereafter. These cellular structure changes occurred concomitantly with the decrease of starch granules and colour change in the pulp. Starch granules accumulated within the pulp parenchyma cells and hydrolysed to sugar once the pulp turned red at 25 DAP as indicated by iodine stain clearing. For the third objective, effects of different pollen load size on H. polyrhizus fruits in term of fruit size and postharvest qualities were studied. Freshly dehisced pollen grains collected during anthesis were weighed at 0.001, 0.05, 0.10, 0.15 and 0.20 g (representing pollen load of 0.01, 0.5, 1.0, 1.5 and 2.0 x, respectively). The usual pollen load applied by the grower was used as control (C). Fruits were harvested at every 5 DAP until day 35. Results indicated that pollen load of 0.01 x produced the smallest fruit in term of fruit weight, length and diameter, followed by 0.5 x. Fruits of other treatments produced similar fruit size as control. There were no significant differences in peel and pulp colour, respiration rate and chemical characteristic among treatments except for soluble solids concentration (SSC). Smaller fruit tends to vield higher SSC. A positive correlation between seed number and fruit weight was determined (r = 0.931, P ≤ 0.001). In conclusion, flowers of H. polyrhizus are nocturnal bloomers with large pollen grains that are covered with echinates suggested zoophilous pollination. Fruits of H. polyrhizus are considered as fast crop as it took 30 days suggested by cellular structure and starch pattern change during fruit development. Findings indicated that fruit weight of H. polyrhizus could be manipulated by the amount of pollen grains as the pulp derived from the funiculli of developing seed.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

STRUKTUR BUNGA DAN KESAN DOS DEBUNGA KE ATAS PEMBENTUKAN BUAH DAN CIRI FISIKO-KIMIA BUAH NAGA [*Hylocereus polyrhizus* (F.A.C Weber) Britton & Rose]

Oleh

JOANNA CHO LEE YING

April 2011

Pengerusi : Profesor Madya Phebe Ding, PhD Fakulti : Pertanian

Di Malaysia, harga dan gred buah naga ditentukan berdasarkan jisim buah. Seperti buah beri yang lain, jisim buah naga dipengaruhi dengan bilangan biji yang terbentuk. Bagi mendapatkan hasil buah yang berkualiti tinggi terutamanya dari segi saiz buah, suatu kajian telah dijalankan ke atas struktur bunga dan dos debunga ke atas pembentukan buah naga isi merah (Hylocereus polyrhizus). Objektif kajian pertama jalah untuk mengenalpasti aktiviti pendebungaan H. polyrhizus. Kajian morfologi debunga dan stigma serta interaksi debunga dengan stigma semasa pendebungaan dikaji dengan menggunakan mikroskop cahaya dan mikroskop pengimbas elektron (SEM) pada 0, 1, 2, 3 dan 4 hari selepas pendebungaan (HSP). Mikroskop pendafluor digunakan untuk mengkaji pertumbuhan tiub debunga di dalam stil bunga yang diwarnai dengan anilin biru pada 0, 2 dan 4 hari selepas pendebungaan (HSP). Bunga H. polyrhizus mekar pada waktu malam dalam keadaan tegak. Penempatan cepu debunga dan stigma bunga adalah jarang dan ciri ini mencadangkan pendebungaan silang. Saiz debunga adalah besar dengan nisbah paksi polar (P) kepada diameter equatorial (E) bersamaan dengan 0.97, berbentuk sfera buntal (oblate) dan trizonokolpat dengan permukaan exine debunga diselaputi dengan deduri (echinates). Stigma tergolong dalam kategori kering (dry) dan terdiri daripada 28 ± 3 jejari yang bertempat di atas stil bunga yang panjang (27 ± 2.54 cm). Stil bunga bercirikan pejal dan mempunyai satu saluran kosong (hollow canal). Dua jam selepas pendebungaan, tiub debunga dari debunga kelihatan tumbuh menerusi tisu-tisu transmisi pada stigma dan bahagian bawah saluran menuju ke ovari. Granul-granul kanji yang terdapat di dalam sel-sel parenkima stil berkurangan apabila bilangan HSP berlaku. Interaksi antara debunga dan stigma mengambil masa 1 HSP. Segelintir tiub debunga tiba di ovari bunga pada 2 HSP. Hasil fertilisasi antara gamet jantan (dari tiub debunga) dan gamet betina (ovul) mengakibatkan pembentukan buah. Objektif kedua ialah mengkaji perubahan struktur sel yang berlaku semasa pembentukan buah H. polyrhizus. Buah dituai pada setiap 5 HSP. Perubahan struktur sel pada setiap peringkat dikaji dengan menggunakan mikroskop cahaya dan SEM. Kehadiran kanji di dalam buah juga diuji dengan menggunakan iodin/kalium iodida (I₂/KI). Selepas fertilisasi, pembentukan buah terdiri daripada dua fasa iaitu divisi dan pembesaran saiz sel terutamanya pada isi buah. Pada peringkat awal, sel parenkima adalah kecil dan mempunyai dinding sel yang teguh tetapi bertambah besar dan longgar apabila buah semakin matang. Divisi sel berlaku pada 20 HSP yang pertama diikuti dengan pembesaran saiz sel pada HSP seterusnya. Perubahan struktur sel berlaku seiring dengan pengurangan kanji dan perubahan warna isi buah pada 25 HSP. Granul-granul kanji yang didapati dalam sel parenkima pada bahagian isi buah menghasilkan kawasan biru hitam apabila diuji dengan larutan I₂/KI pada 5 - 20 HSP. Pelunturan I₂/KI bermula pada bahagian isi buah yang menjadi merah pada 25 HSP. Objektif ketiga ialah untuk mengkaji kesan dos debunga ke atas buah H. polyrhizus dari segi pembentukan buah, saiz buah dan kualiti buah selepas tuai. Butiran debunga yang segar dikumpul dari bunga mekar dan dos debunga ditentukan dengan menimbang debunga yang dikumpul kepada 0.001, 0.05, 0.10, 0.15 dan 0.20 g. Setiap rawatan mewakili dos debunga yang terdiri daripada 0.01, 0.5, 1.0, 1.5 dan 2.0 x. Dos debunga yang biasa digunakan oleh peladang dijadikan sebagai kawalan (K). Buah dituai secara rawak pada 5 HSP sehingga hari ke-35. Keputusan kajian menunjukkan rawatan 0.01 x menghasilkan buah yang paling kecil dari segi jisim, panjang dan diameter, diikuti dengan 0.05 x, berbanding dengan rawatan yang lain. Rawatan lain menghasilkan buah yang sama seperti kawalan dari semua aspek yang dikaji. Tiada perbezaan bererti ditemui dalam kadar respirasi dan fizikal-kimia antara rawatan kecuali bacaan jumlah kandungan pepejal terlarut (KPT). Buah bersaiz kecil dari rawatan 0.01 x didapati mempunyai nilai KPT yang tinggi berbanding dengan rawatan yang lain. Jisim buah berkolerasi positif dengan bilangan biji yang terhasil di dalam buah (r = 0.931, P ≤ 0.001). Kesimpulannya, bunga H. polyrhizus mekar pada waktu malam serta ciri debunga besar yang diselaputi dengan duri merupai ciri pendebungaan zoofili. Buah H. polyrhizus mencapai tahap pemakanan yang optimal pada 30 HSP. Kenyataan ini disokong dengan perubahan struktur sel buah dan corak kanji semasa pembentukan buah. Isi buah H. polyrhizus dibentuk oleh funikuli biji yang berkembang menunjukkan bahawa jisim buah naga isi merah dapat dimanipulasi dengan mengawal dos debunga yang diguna semasa pendebungaan.

ACKNOWLEDGEMENTS

First and foremost, I would like to express my gratitude to my project supervisor Assoc. Prof Dr. Phebe Ding for her advice guidance, comments and encouragement in my Master's study. I would like to offer my sincere appreciation to Prof. Abdul Rahman bin Abdul Razak for his expertise and to Assoc. Prof. Dr. Zakaria bin Wahab for his constructive comments to improve in my research.

I would like to offer my sincere appreciation to Mr. Yap Tee and his family for sharing his knowledge in dragon fruit and also his generosity in lending me his farm to carry out my experiments. I would also like to thank my fellow pollinators; Ms Yap Moh Yuin, Ms Tee Yei Kheng, Ms Jackie Chua, Ms Isabel Freddy, Ms Pong Vui Mei, Pn Nur Adilla Jamaluddin, Pn Nur Shairah and Mr Boon for their kind assistance in this research. With their support, assistance and tolerance during the whole duration of this project had made it an enjoyable and memorable experience for me.

Other than that, I would like to extend my gratitude to En. Daud Mustam from Plant Microstructure and Anatomy Laboratory for his technical assistance in using the scanning electron microscopy and histology techniques. And also to Pn Zahida Muhammed and En Rafiuz Zaman Hassan from Microscopy Unit, Institute of Bioscience for their guidance in the use of light and fluorescence microscopy.

My heartfelt gratitude and appreciation go to my family and friends for their constant support, faith, understanding and encouragement towards me. Thank you Pa, Ma and Jie Jie for always being there for me.

Last but not least, I would like to include a favourite quote of mine from Steve Jobs in which he said, "You have to trust that the dots will somehow connect in your future, You have to trust in something – your gut, destiny, life, karma, whatever. This approach has never let me down, and it has made all the difference in my life."

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Phebe Ding, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Zakaria bin Wahab, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

Abdul Rahman bin Abd. Razak, PhD

Professor Faculty of Agrotechnology and Food Science Universiti Malaysia Terengganu (Member)

BUJANG KIM HUAT, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 8 Sept 2011

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:

Date:

Name and Matric No.: Joanna Cho Lee Ying (GS19887)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	Associate Professor Dr. Phebe Ding
Signature:	
Name of Member	
of Supervisory	Professor
Committee:	Dr. Zakaria bin Wahab
Signature: Name of Member of Supervisory Committee:	Professor Dr. Abdul Rahman bin Abd. Razak

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	V
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xvii

CH	AP	T	ER

PTER				
1	INTE	RODUCT	FION	1
2	2.12.22.32.4	Introdu 2.1.1 Flower 2.2.1 2.2.2 Fruit ch Pollen	Cacti as fruit crops Flower types and pollinators Pollination naracteristic load rvest qualities changes during fruit	3 3 4 7 8 9 11
		2.5.1 2.5.2 2.5.3 2.5.4	Fruit size Colour Respiration rate and ethylene production Texture Taste	11 12 13 14 15
3	FLE		ON PROCESS OF RED RAGON FRUIT (<i>Hylocereus</i>	17
		Introdu	ation	17
	3.1		als and Methods	17
	J.Z		Plant material	17
	3.3		Statistical analysis s and discussion	20
	ა.ა	3.3.1		20 20
		3.3.1	The flower of red-fleshed dragon fruit	20
		3.3.2		29
		3.3.3	Stigma morphology of red- fleshed dragon fruit at different time intervals	31

		3.3.4	Pollination process of red-fleshed dragon fruit	34
	3.4	Conclu	•	43
4			ELOPMENT OF RED-FLESHED RUIT (<i>Hylocereus polyrhizus</i>)	45
	4.1	Introdu		45
	4.2	Materi	als and Methods	46
		4.2.1	Plant material	46
		4.2.2	Statistical analysis	47
	4.3	Result	s and discussion	47
		4.3.1	Fruit development	47
		4.3.2	Presence of starch granules	59
			during fruit development	
	4.4	Conclu	ision	61
5			F POLLEN LOAD AND QUALITY DEVELOPMENT OF RED-	63
			RAGON FRUIT (Hylocereus	
		rhizus)		
	5.1		uction	63
	5.2		als and Methods	63
		5.2.1	Determination of postharvest	66
			qualities	
		5.2.2	Statistical analysis	66
	5.3	Result	s and discussion	
		5.3.1	Fruit weight, length and diameter	66
		5.3.2	Carbon dioxide and ethylene production	73
		5.3.3	Colour development of peel and	76
		5.5.5	pulp	70
		5.3.4	Postharvest qualities	79
	5.4	Conclu	ision	82
6	CO	NCLUSI	ONS AND RECOMENDATIONS	
		Conclu		83
	6.2	Recorr	nmendations	84
REFEREN	CES			85
	APPENDICES			95
BIODATA	OF S	TUDENT	Г	99
LIST OF P	UBLIC	CATION		100

xi

 \mathbf{G}

LIST OF TABLES

Table		Page
3.1	Mean ± S.D of number, length, diameter, ratio and shape of petaloid and sepaloid of <i>Hylocereus polyrhizhus</i> flower	22
3.2	Mean ± S.D of number of stamens, filament length and anther length of <i>Hylocereus polyrhizhus</i>	24
3.3	Mean ± S.D of number of stigma lobe, stigma length, style length and diameter and distance between stigma and anther of <i>Hylocereus polyrhizhus</i>	24
3.4	Mean ± S.D of pollen size of <i>Hylocereus polyrhizhus</i>	29
4.1	Peel and pulp measurements during fruit development of <i>Hylocereus polyrh<mark>i</mark>zhus</i>	49
5.1	Effects of pollen load and day after pollination on weight, length and diameter of red-fleshed dragon fruit (<i>Hylocereus polyrhizhus</i>)	67
5.2	Correlation coeffiencents of weight, length and diameter of red-fleshed dragon fruit (<i>Hylocereus polyrhizhus</i>)	72
5.3	Effects of pollen load and day after pollination on carbon dioxide and ethylene production of red-fleshed dragon fruit (<i>Hylocereus polyrhizhus</i>)	74
5.4	Effects of pollen load and day after pollination on colour development of peel and pulp of red-fleshed dragon fruit (<i>Hylocereus polyrhizhus</i>)	77
5.5	Effects of pollen load and day after pollination on firmness, soluble solids concentration (SSC), pH and citric acid of red-fleshed dragon fruit (<i>Hylocereus polyrhizhus</i>)	79
5.6	Correlation coefficients of firmness, soluble solids concentration (SSC), pH and citric acid of red-fleshed dragon fruit (<i>Hylocereus polyrhizhus</i>)	81

LIST OF FIGURES

Figure		Page
3.1	Flower of Hylocereus polyrhizus at athesis (2200 h)	21
3.2	Androecium of Hylocereus polyrhizus	23
3.3	Flower parts of the Hylocereus polyrhizus	25
3.4	The blooming sequence of <i>Hylocereus polyrhizus</i> flower at 1900 (A), 0000 (B), and 0900 h (C) with accompanying stigma (D, E and F respectively)	27
3.5	The blooming sequence of <i>Hylocereus polyrhizus</i> flower at 2 (A), 3 (B) and 4 (C) day after pollination (DAP) with accompanying stigma (D, E and F) respectively	28
3.6	Features of the <i>Hylocereus polyrhizus</i> pollen grain under scanning electron microscope (A and B)	29
3.7	Features of the <i>Hylocereus polyrhizus</i> pollen grain under light microscope (A and B)	30
3.8	Pollen tube growing from one of the three aperture after 2 h of germination	30
3.9	Stigma of <i>Hylocereus polyrhizus</i> during athesis at 2200 h	32
3.10	Scanning electron micrographs of the stigmatic surface of <i>Hylocereus polyrhizus</i> at 1900, 0000 and 0900 h the following morning	33
3.11	Pollen-stigma interactions at 0 day after pollination (DAP)	35
3.12	Fluorescence microscopy of pollen tube growth of <i>Hylocereus polyrhizus</i> at 0 DAP	35
3.13	Tranverse section of the style of <i>Hylocereus polyrhizus</i> at 0 DAP as seen under the light microscope. All sections were stained with Toluidine Blue O	36
3.14	Longitudinal section of the style of <i>Hylocereus polyrhizus</i> at 0 DAP as seen under the light microscope. All sections were stained with Toluidine Blue O	37

C

3.15	Pollen-stigma interactions at 1 day after pollination (DAP)	38
3.16	Pollen-stigma interactions at 2 day after pollination (DAP)	39
3.17	Fluorescence microscopy of pollen tube growth of <i>Hylocereus polyrhizus</i> at 2 DAP	39
3.18	Fluorescence microscopy of pollen tube growth of <i>Hylocereus polyrhizus</i> at 4 DAP	40
3.19	Transverse section of the middle part of the style (10 cm below the stigma) at 4 DAP	40
3.20	Transverse section of the style at 4 DAP as seen under the light microscope. All sections were stained with Toluidine Blue O	42
3.21	Longitudinal section of the style of <i>Hylocereus</i> <i>polyrhizus</i> at 4 DAP as seen under the light microscope. All sections were stained with Toluidine Blue O	43
4.1	Fruit development of Hylocereus polyrhizus at 5 DAP	48
4.2	The fruit of <i>Hylocereus polyrhizus</i> at 5 DAP	50
4.3	Cellular structure in the peel, peel-to-pulp transition and pulp region of fruit of <i>Hylocereus polyrhizus</i> at 10 DAP	51
4.4	Cellular structure in the peel, peel-to-pulp transition and pulp region of fruit of <i>Hylocereus polyrhizus</i> at 15 DAP	53
4.5	Cellular structure in the peel, peel-to-pulp transition and pulp region of fruit of <i>Hylocereus polyrhizus</i> at 20 DAP	54
4.6	Cellular structure in the peel, peel-to-pulp transition and pulp region of fruit of <i>Hylocereus polyrhizus</i> at 25 DAP	56
4.7	Cellular structure in the peel, peel-to-pulp transition and pulp region of fruit of <i>Hylocereus polyrhizus</i> at 30 DAP	58

Ś

59

60

68

69

70

Starch pattern of *Hylocereus polyrhizus* during fruit development using I₂/KI staining

Cellular structure in the peel, peel-to-pulp transition

and pulp region of fruit of Hylocereus polyrhizus at

5.1 Effects of different pollen load and day after pollination on fruit weight of red-fleshed dragon fruit (*Hylocereus polyrhizus*). Each point represented the mean value (n=9). First colour change from green to red was observed at 25 DAP whereas full colour change to fully red were observed at 30 DAP for all treatments.

4.8

4.9

35 DAP

- 5.2 Effects of pollen load x day after pollination on fruit weight of red-fleshed dragon fruit (*Hylocereus polyrhizus*). Means separation pertaining to each day after pollination is by LSD at $P \le 0.05$.
- 5.3 Effects of different pollen load and day after pollination on fruit length of red-fleshed dragon fruit (*Hylocereus polyrhizus*). Each point represented the mean value (n=9). First colour change from green to red was observed at 25 DAP whereas full colour change to fully red were observed at 30 DAP for all treatments.
- 5.4 Effects of different pollen load and day after 70 pollination on fruit diameter of red-fleshed dragon fruit (*Hylocereus polyrhizus*). Each point represented the mean value (n=9). First colour change from green to red was observed at 25 DAP whereas full colour change to fully red were observed at 30 DAP for all treatments.
- 5.5 Effects of pollen load x day after pollination on fruit 71 length of red-fleshed dragon fruit (*Hylocereus polyrhizus*). Means separation pertaining to each day after pollination is by LSD at $P \le 0.05$.
- 5.6 Effects of pollen load x day after pollination on fruit 72 diameter of red-fleshed dragon fruit (*Hylocereus polyrhizus*). Means separation pertaining to each day after pollination is by LSD at $P \le 0.05$.
- 5.7 Correlation between number of seeds with fruit 73 weight of red-fleshed dragon fruit (*Hylocereus polyrhizus*).

- 5.8 Effects of pollen load x day after pollination on carbon dioxide production of red-fleshed dragon fruit (*Hylocereus polyrhizus*). Means separation pertaining to each day after pollination is by LSD at $P \le 0.05$.
- 5.9 Effects of pollen load x day after pollination on hue value of red-fleshed dragon fruit (*Hylocereus polyrhizus*). Means separation pertaining to each day after pollination is by LSD at $P \le 0.05$.
- 5.10 Effects of pollen load x day after pollination on soluble solids concentration (SSC) of red-fleshed dragon fruit (*Hylocereus polyrhizus*). Means separation pertaining to each day after pollination is by LSD at $P \le 0.05$.

75

78

80

LIST OF ABBREVIATIONS

	°C	celcius
	C*	chroma
	C_2H_4	ethylene
	CO ₂	carbon dioxide
	cm	centimeter
	CPD	critical point dryer
	DAP	day after pollination
	FAA	formaldehye-acetic acid-alcohol
	FID	flame ionization detector
	h	hour
	h°	hue
	g	gram
	L*	lightness
	L	liter
	K ₃ PO ₄	potassium phosphate
	LSD	least significant difference
	mL	milliliter
	n	Newton
	NaOH	sodium hydroxide
	r ²	correlation coefficient
(C)	SAS	statistical analysis sytem
	SEM	scanning electron microscope
	SSC	soluble solids concentration
	TTA	titratable acidity

- TBO toluidine blue O
- TCD thermal conductivity detector
- UV ultraviolet
- µm micrometer



CHAPTER 1

GENERAL INTRODUCTION

Pitaya or dragon fruit belongs to the Cactaceae family which is a native of the tropical regions of Mexico and Central and South America (Anderson, 2001). In South East Asia, the fruit is commonly known as dragon fruit as the peel of the fruit is covered with bracts which resembles the 'scales' of the mythological dragon (Wu et al., 2006). Dragon fruit is also known as thanh long, pitaya, pitahaya, pitajaya, pitajuia or pitalia in different parts of the world (Le Bellec et al., 2006). The plants are characterised by elongated stems (usually three winged) with aerials roots, large white monoecious flowers and spineless edible berry fruit with many edible seeds (Tel-Zur et al., 2004). The peel colour varies from pinkish to red while the pulp colour varies from white to red or purplish, depending on varieties. There are only few varieties of dragon fruits currently found cultivated in Malaysia such as the red fleshed varieties; *Hylocereus polyrhizus, H. costaricensis* and the white fleshed; *H. undatus*.

Fruit weight of dragon fruit is positively correlated with the number of viable seeds (Weiss et al., 1994). Hence, achievements of uniform and profitable yields depend entirely on hand pollination. As a result, variation of weight and size are produced. The optimal time to harvest dragon fruit is within 28 to 30 days after flower anthesis at full colour development (Nerd et al., 1999). The average weight for a mature fruit ranged between 350 – 900 g. The price of dragon fruit is paid according to fruit weight and varieties.

Fruit weight, like fruit colour, plays an important role in the consumer purchase process. For example, in order to gain access to Europe market, the preferred average weight for white-fleshed dragon fruits in Vietnam is 300 g (Hoa, 2008). In Malaysia, the Department of Agriculture graded and priced the dragon fruits into different catergories of: Grade A = more than 450 g, Grade B = 300-450 g and Grade C = less than 300 g (Ministry of Agriculture, 2006).

The fruit is unique since it contains betacyanins pigments with antioxidant properties; a rare properties which is hard to find in other fruits. Thorough examination of the floral structure and its relation to fruit development would provide useful data for the plantation industry. Moreover, researches done for dragon fruits are mostly carried out in Israel where the environment and planting conditions were different in Malaysia. In addition to that, as pollen load was found to affect the fruit size and weight of cherimoya (Gonzalez et al., 2006) and cranberries (Cane and Schiffhauer, 2003), a study on the pollen load effect on dragon fruits were also carried out in order to optimize the fruit yield.

Therefore, the objectives of this study were (i) to investigate the pollination process of red-fleshed dragon fruit, (ii) to elucidate the changes in cellular structure during fruit development and (iii) to evaluate the effects of pollen load size on fruit set, size and postharvest qualities of the red-fleshed dragon fruit.



REFERENCES

- Abbott, J. A. 1999. Quality measurement of fruits and vegetables. *Postharvest Biology and Technology* 15 (3): 207-225.
- Aliasgharpour, M., Hekmatshoar, H., Hosseyni, M. and Some-eh, F. 2004. Lipids in the stigmatic secretion of *Datur stramonium* L (Solanaceae). *Iranian Journal of Science & Technology* 28 (A1): 19-31.

Anderson, E.F. 2001. The Cactus Family. USA: Timber Press.

- Artes, F., Minguez, M.I. and Hornero, D. 2002. Analysing changes in fruit pigments. In *Colour in food: Improving quality*. ed. D.B. MacDougall, pp. 248-277. England: Woodhead Publishing Limited.
- Bassetto, E., Jacomino, A.P., Pinheiro, A.L. and Kluge, R.A. 2005. Delay of ripening of 'Pedro Sato' guava with 1-methylcyclopropene. *Postharvest Biology and Technology* 35: 303-308.
- Beeckman, T., De Rycke, R., Mane, R. and Inze, D. 2000. Histological study of seed coat development in *Arabidopsis thaliana*. *Journal of Plant Research* 113: 139-148.
- Belitz, H.D., Grosch, W. and Schieberle, P. 2004. *Food chemistry*, 3rd edition. Germany: Springer-Verlag Berlin Heidelberg.
- Briggs, C.L., Morris, E.C. and Ashford, A.E. 2005. Investigations into seed dormancy in *Grevillea linearifolia*, *G. buxifolia* and *G. sericea*: anatomy and histochemistry of the seed coat. *Annals of Botany* 96: 965-980.
- Boke, N. H. 1964. The cactus gynoecium: a new interpretation. *American Journal* of Botany 51:598-610.
- Boke, N.H. 1980. Developmental morphology and anatomy in Cactaceae. *BioScience* 30: 605–610.
- Bourne, M.C. 1979. Texture of temperate fruits. *Journal of Texture Studies* 10: 25-44.
- Boyle, T.H., Menalled, F.D. and O'Leary, M.C. 1994. Occurrence and physiological breakdown of self-incompatibility in Easter Cactus. *Journal* of the American Society for Horticultural Science 119 (5): 1060-1067.
- Boyle, T.H., Karle, R. and Han, S.S. 1995. Pollen germination, pollen tube growth, fruit set, and seed development in *Schlumbergera truncata and S. x buckleyi* (Cactaceae). *Journal of the American Society for Horticultural Science* 120: 313-317.

- Cadwallader, K.R. 2005. Flavor and volatile metabolism in produce. In *Produce degradation: Pathways and prevention*. ed. O. Lamikanra, S.H. Imam and D.O. Ukuku. pp. 155-190. USA: Taylor and Francis.
- Cane, J.H. and Schiffhauer, D. 2003. Dose-response relationships between pollination and fruiting refine pollinator comparisons for cranberry (*Vaccinium macrocarpon* [Ericaceae]). *American Journal of Botany* 90 (10): 1425-1432.
- Carrington, C.M.S. and King, R.A.G. 2002. Fruit development and ripening in Barbados cherry, *Malpighia emarginata* DC. *Scientia Horticulturae* 92 (1): 1-7.
- Casas, A. and Barbera, G. 2002. Mesoamerican domestication and diffusion. In *Cacti: Biology and uses.* ed. P.S. Nobel. USA: University of California Press.
- Cheah, L.S. and Zulkarnain, W. M. 2008. In: Seminar on Pitaya Production, Market and Export-Challenges and Prospects. Palm Garden Hotel, IOI Resort, Putrajaya, Malaysia. 20 October 2008.
- Considine, J.A. and Knox, R.B. 1979. Development and histochemistry of the cells, cell walls, and cuticle of the dermal system of fruit of the grape, *Vitis vinifera* L. *Protoplasma* 99: 347-365.
- Coombe, B.G. 1976. The development of fleshy fruit. Annual Reviews of Plant Physiology 27: 507-528.
- Cosgrove, D.J. 2005. Growth of the plant cell wall. *Molecular Cell Biology* 6 (11): 850-861.
- Dag, A. and Mizrahi, Y. 2005. Effect of pollination method on fruit set and fruit characteristics in the vine cacti Selenicereus megalanthus (yellow pitaya). *Journal of Horticultural Science and Biotechnology* 80 (5): 618-622.
- Daun, H. 2005. Produce color and appearance. In *Produce degradation: Pathways and prevention.* ed. O. Lamikanra, S.H. Imam and D.O. Ukuku. pp. 191-222. USA: Taylor and Francis.
- Defilippi, B.G., Manriquez, D., Luengwilai, K. and Gonzalez-Aguero, M. 2009. Aroma volatiles: Biosynthesis and mechanisms of modulation during fruit ripening (Abstract). *Advances in Botanical Research* 50: 1-37.

Denney, J.O. 1992. Xenia includes metaxenia. *HortScience* 27:722–728.

Ding, P., Ahmad, S. H. and Ghazali, H. M. 2007. Changes in selected quality characteristics of minimally processed carambola (*Averrhoa carambola* L.) when treated with ascorbic acid. *Journal of the Science of Food and Agricultural* 87: 702- 709.

- Erdelska, O. and Ovecska, M. 2004. Senescence of unfertilised flowers in *Epiphyllum* hybrids. *Biologia Plantarum* 48 (3): 381-388.
- Erdtman G. 1971. *Pollen morphology and plant taxonomy*. USA: Harner Publishing Company Inc.
- Esau, K. 1960. *Plant Anatomy*. New York: Wiley.
- Evangelista, R.M., Chitarra, A.B. and Chitarra, M.I.F., 2002. Ultrastructural changes in the cell wall of mango 'Tommy Atkins' treated with calcium chloride at pre-harvest time. (Abstract). *Revista Brasileria de Fruticultura* 24: 254–257.
- Feugang, J.M., Konarski, P., Zou, D., Stintzing, F.C. and Zou, C. 2006. Nutritional and medicinal use of cactus pear (*Opuntia* spp.) cladodes and fruits. *Frontiers in Bioscience* 11: 2574–2589.
- Fonseca, S., Hackler, L., Zvara, A., Ferreira, S. Balde, A., Dudits, D., Pais, M.S. and Puskas, L.G. 2004. Monitoring gene expression along pear fruit development, ripening and senescence using cDNA microarrays. *Plant Science* 167: 457-469.
- Francis, F.J. 2002. Food colorings. In *Colour in food: Improving quality*. ed. D.B. MacDougall. pp. 297-327. England: Woodhead Publishing Limited.
- Garcia, E. and Lajolo, F.M. 1998. Starch transformation during banana ripening: the amylase and glucosidase behavior. *Journal of Food Science* 53(4): 1181-1186.
- Gillaspy, G., Ben-David, H. and Gruissem, W. 1993. Fruits: A developmental perspective. *The Plant Cell* 5: 1439-1451.
- Giovannoni, J. 2001. Molecular biology of fruit maturation and ripening. Annual Reviews of Plant Physiology and Plant Molecular Biology 52: 725-749.
- Gonzalez, M.V., Coque, M. and Herrero, M. 1995. Papillar integrity as an indicator of stigmatic receptivity in kiwifruit (*Actinidia deliciosa*). *Journal of Experimental Botany* 46 (2): 263-269.
- Gonzalez, M.V., Coque, M. and Herrero, M. 1998. Influence of pollination systems on fruit set and fruit quality in kiwifruit (*Actinidia deliciosa*). *Annals of Applied Biology* 132 (2): 349-355.
- Gonzalez, M., Baeza, E., Lao, J. L. and Cuevas, J. 2006. Pollen load affects fruit set, size, and shape in cherimoya. *Scientia Horticulturae* 110: 51-56.
- Goulao, L.F. and Oliviera, C.M. 2008. Cell wall modifications during fruit ripening: when a fruit is not the fruit. *Trends in Food Science and Technology* 19: 4-25.

- Grant, V. and Grant, K.A. 1979. The pollination spectrum in the Southwestern American cactus flora. *Plant Systematics and Evolution* 133: 29-37.
- Hampson, C. R. and Azarenko, A.N. 1993. Pollen-stigma interactions following compatible and incompatible pollinations in hazelnut. *Journal of the American Society for Horticultural Science* 118 (6): 814-819.
- Hernadez, M.S., Martinez, O. and Fernandez-Trujillo, J.P. 2006. Behavior of araza (*Eugenia stipitata* Mc Vaugh) fruit quality traits during growth, development and ripening. *Scientia Horticulturae* 111 (3): 220-227.
- Heslop-Harrison, Y. and Shivanna, K.R. 1977. The receptive stigma of the angiosperm stigma. *Annals of Botany* 41: 1233-1258.
- Heyes, J.A. and Sealey, D.F. 1996. Textural changes during nectarine (*Prunus persica*) development and ripening. *Scientia Horticulturae* 65: 49-58.
- Hiscock, S.J., Hoedemaekers, K., Friedman, W.E. and Dickinson, H.G. 2002. The stigma surface and pollen-stigma interactions in *Senecio squalidus* L. (Asteraceae) following cross (compatible) and self (incompatible) pollinations. *International Journal of Plant Science* 163 (1): 1-16.
- Hoa, N. V. 2008. In: Seminar on Pitaya Production, Market and Export-Challenges and Prospects. Palm Garden Hotel, IOI Resort, Putrajaya, Malaysia. 20 October 2008.
- Hopkins, W.G. 1999. *Introduction to plant physiology*, 2nd edition. USA: John Wiley & Sons.
- Huang, Z., Zhu, J., Mu, X. and Lin, J., 2004. Pollen dispersion, pollen viability and pistil receptivity in *Leymus chinensis*. *Annals of Botany* 93: 295-301.
- Iglesias, I. and Echeverria, G. 2009. Differential effect of cultivar and harvest date on nectarine colour, quality and consumer acceptance. *Scientia Horticulturae* 120: 41-50.

Jabatan Pertanian Malaysia, 2006. Pakej Teknologi Tanaman Pitaya.

- Janssen, B.J., Thodey, K., Schafferi, R.J., Alba, R., Balakrishnan, L., Bishop, R., Bowen, J.H., Crowhurst, R.N., Gleave, A.P, Ledger, S., McArtney, S., Pichler, F.B., Snowden, K.C. and Ward, S. 2008. Global gene expression analysis of apple fruit development from the floral bud to ripe fruit. *BMC Plant Biology* 8 (16): 1-29.
- Jha, S.N., Kingsly, A.R.P. and Chopra, S. 2006. Physical and mechanical properties of mango during growth and storage for determination of quality. *Journal of Food Engineering* 72: 73-76.
- Johri, B.M., Ambegaokar, K.B. and Srivastava, P.S. 1992. *Comparative embryology of Angiosperms I.* New York: Springer-Verlag.

- Kays, S.J. 1991. Postharvest physiology of perishable plant products. New York: AVI Publishing, Van Nostrand Reinhold.
- Kelly, K.M., Van Staden, J. and Bell, W.E. 1992. Seed coat structure and dormancy. *Plant Growth Regulation* 11: 201-209.
- Kim, J.G., Takami, Y., Mizugami, T., Beppu, K., Fukuda, T. and Kataoka, I. 2005. CPPU application on size and quality of hardy kiwifruit. *Scientia Horticulturae* 110 (2): 219-222.
- Knox, R.B. 1971. Pollen and allergy. Great Britain: The Camelot Press.
- Kreitschitz, A. 2009. Biological properties of fruit and seed slime envelope: How to live, fly and not die. In *Functional surfaces in biology*, ed. S.N. Gorb, pp. 11-30. Netherlands: Springer Netherlands.
- Lalel, H.J.D., Singh, Z. and Tan, S.C. 2003. Glycosidically-bound aroma volatile compounds in the skin and pulp of 'Kensington Pride' mango fruit at different stages of maturity. *Postharvest Biology and Technology* 29 (2): 205-218.
- Lam, P.F. and Wan, C.K. 1983. Climateric nature of the carambola (Averrhoa carambola L.) fruit. *Pertanika* 6 (3): 44-47.
- Le Bellec, F., Vaillant, F. and Imbert, E. 2006. Pitahaya (*Hylocereus* spp.): a new fruit crop, a market with a future. *Fruits* 61: 237-250.
- Lersten, N.R. 2004. Flowering plant embryology. USA: Blackwell Publishing.
- Lewallen, K.S. and Marini, R.P. 2003. Relationship between flesh firmness and ground colour in peach as influenced by light and canopy position. *Journal of the American Society for Horticultural Science* 128:163-170.
- Lichtenzveig, J., Abbo, S., Nerd, A., Tel-Zur, N. and Mizrahi, Y. 2000. Cytology and mating systems in the climbing cacti *Hylocereus* and *Selenicereus*. *American Journal of Botany* 87(7): 1058-1065.
- Mandujano, M.C., Carrillo-Angeles, I., Martinez-Peralta, C. and Golubov, J. 2010. Reproductive Biology of Cactaceae. In *Desert Plants: Biology and Biotechnology*. ed. K.G. Ramawat. pp. 197-230. Germany: Springer-Verlag Berlin Heidelberg.
- Marcelis, L.F.M and Baan Hofman-Eijer, L.R. 1997. Effects of seed number on competition and dominance among fruits in *Capsicum annuum* L. *Annals of Botany* 79: 687-693.
- Martin, F. W. 1959. Staining and observing pollen tubes in the style by means of fluorescence. *Stain Technology* 34: 125-128.
- Martin, M. J., Chapman., P.R. and Auger. H.A. 1971. *Cacti and their cultivation.* London: Faber and Faber.

- Masyahit, M., Sijam, K. Awang, Y. and Mohd Ghazali., M.S. 2009. First report on bacterial soft rot disease on dragon fruit (*Hylocereus* spp.) caused by *Enterobacter cloacae* in Peninsular Malaysia. *International Journal of Agriculture and Biology* 11(6): 659-666.
- Mohammad, P. and Shiraishi, M. 1998. Anatomical observations on the accumulation and utilization of storage starch and epidermis-related development in roots of vigorous and non-vigorous Satsuma mandarin trees from early winter to early summer. *Journal of the Japan Society for Horticultural Science* 67(5): 660-670.
- Mejia, A. and Cantwell, M. 2003. Prickly pear fruit development and quality in relation to gibberellic acid applications to intact and emasculated flower buds. *Journal of the Professional Association for Cactus Development* 5: 72-85.
- Mercado-Silva, E., Bautista, P.B. and Garcia-Velasco, M.A. 1998. Fruit development, harvest index ripening changes of guavas produced in central Mexico. *Postharvest Biology and Technology* 13: 143-150.
- Merten, S. 2003. A Review of *Hylocereus* Production in the United States. Journal of the Professional Association for Cactus Development 5: 98-105.
- Mitcham, B., Cantwell, M. and Kader, A. 1996. Methods for determining quality of fresh commodities. *Perishables Handling Newsletter* 85: 1-5.
- Mizrahi, Y., Nerd, A. and Nobel, P.S. 1996. Cacti as crops. *Horticultural Reviews* 18: 291- 320.
- Mizrahi, Y. and Nerd, A. 1999. Climbing and columnar cacti: New Arid Land Fruit Crops. In *Perspectives on New Crops and New Uses*, ed. J. Janick, pp. 358-366. Alexandaria, USA: ASHS Press.
- Mulas, M., Agabbio, M. and Nieddu, G. 1989. Ultrastructural morphology of 'Nonpareil' almond pollen. *HortScience* 24(5): 816-818.
- Munasque, V.S. and Mendoza, D.B. 1990. Development physiology and ripening behaviour of Senorita banana (*Musa* sp L.) fruits. *ASEAN Food Journal* 5: 152-157.
- Nagar, P.K. and Rao, R. 1986. Early changes in growth regulator content of pollinated guava fruits. *Scientia Horticulturae* 29: 139-146.
- Nerd, A. and Mizrahi, Y. 1997. Reproductive biology of cactus fruit crops. *Horticultural Reviews* 18: 321-346.
- Nerd, A. and Y. Mizrahi. 1998. Fruit development and ripening in yellow pitaya. Journal of the American Society for Horticultural Science 123: 560–562.

- Nerd, A., Gutman, F. and Mizrahi, Y. 1999. Ripening and postharvest behaviour of fruits of two *Hylocereus* species (Cactaceae). *Postharvest Biology and Technology* 17: 39-45.
- Nerd, A., Sitrit, Y., Kaushik, R.A. and Mizrahi, Y. 2002. High summer temperatures inhibit flowering in vine pitaya crops (*Hylocereus* spp.). *Scientia Horticulturae* 96: 343-350.
- Nieddu, G. and Spano, D. 1992. Flowering and growth in *Opuntia ficus-indica*. *Acta Horticulturae* 296: 153–159.
- Nobel, P.S., Cavelier, J. and Andrade, J.L. 1992. Mucilage in cacti: its apoplastic capacitance, associated solutes and influence on tissue water relations. *Journal of Experimental Botany* 43: 641-648.
- Ohmiya, A. and Kakiuchi, N. 1990. Quantitative and morphological studies on starch of apple fruit during development. *Journal of the Japan Society for Horticultural Science* 59:417-423.
- Ong, B.T., Nazimah, S.A.H, Osman, A., Quek, S.Y., Yoon, Y.Y., Mat Hashim, D., Chew, P.M. and Kong, Y.W. 2006. Chemical and flavour changes in jackfruit (*Artocarpus heterophyllus* Lam.) cultivar J3 during ripening. *Postharvest Biology and Technology* 40 (3): 279-286.
- Osborn, M.M., Kevan, P.G., Lane, M.A. 1988. Pollination biology of *Opuntia* polyacantha and *Opuntia* phaeacantha (Cactaceae) in southern Colorado. *Plant Systematics and Evolution* 159: 85–94.
- Opara, L.U. 2000. Fruit growth measurement and analysis. *Horticultural reviews* 24: 373-432.
- Ozgen, M., Palta, J.P. and Smith, J.D. 2002. Ripeness stage at harvest influences postharvest life of cranberry fruit: physiological and anatomical explanations. *Postharvest Biology and Technology* 24 (3): 291-299.
- Palapol, Y., Ketsa, S., Stevenson, D., Cooney, J.M., Allan, A.C. and Ferguson, I.B. 2009. Colour development and quality of mangosteen (*Garcinia mangostana* L.) fruit during ripening and after harvest. *Postharvest Biology and Technology* 51: 349-353.
- Palmer, J.K. 1971. The banana. *The biochemistry of fruits and their products.* Volume 2, London: Academic Press.
- Payasi, A., Mishta, N.N., Chaves, A.L.S. and Singh, R. 2009. Biochemistry of fruit softening: an overview. *Physiology and Molecular Biology of Plants* 15(2): 103-113.
- Pereira, T., de Almeida, P.S.G., de Azevedo, I.G., da Cunha, M., de Oliveira, J.G., da Silva, M.G. and Vargas, H. 2009. Gas diffusion in 'Golden' papaya fruit at different maturity stages. *Postharvest Biology and Technology* 54: 123-130.

- Perkins-Veazie, P. 1995. Growth and ripening of strawberry fruit. *Horticultural Reviews* 17: 267-297.
- Pimienta-Barrios, E.U., Nobel, P.S., Robles-Murguía, C., Méndez-Morán, L., Pimienta-Barrios, E.N. and Yepez-González, E. 1997. Ethnobotany, productivity, and ecophysiology of pitaya (*Stenocereus queretaroensis*). *Journal of the Professional Association for Cactus Development* 2: 29–47.
- Pimienta-Barrios, E. and del Castillo, R.F. 2002. Reproductive biology. In *Cacti: Biology and uses.* ed. P.S. Nobel. pp. 75- 90. USA: University of California Press.
- Ranggana, S. 1977. *Manual of analysis of fruits and vegetable products*. New Delhi: Tata McGraw-Hill Pub. Co. Ltd.
- Reaves, R. M. 1959. Histological and histochemical changes in the developing and ripening peaches. *American Journal of Botany* 46: 214-248.
- Rocculi, P., Romani, S. and Rosa, M.D. 2004. Evaluation of physico-chemical parameters of minimally processed apples packed in non-conventional modified atmosphere. *Food Research International* 37(4): 329-335.
- Rodrigo, J., Herrero, M. and Hormaza, J. J. 2009. Pistil traits and flower fate in apricot (*Prunus armeniaca*). Annals of Applied Biology 154(3): 365-375.
- Roth, I. 1977. *Fruits of angiosperms*. Germany: Gebruder Borntraege Berlin Stuttgart.
- Salvador, A., Arnal, L., Besada, C., Larrea, V., Quiles, A. and Perez-Munuera, I., 2007. Physiological and structural changes during ripening and deastringency treatment of persimmon fruit cv. 'Rojo Brillante'. *Postharvest Biology and Technology* 46, 181–188.
- Salveit, M.E. 1999. Effect of ethylene on quality of fresh fruits and vegetables. *Postharvest Biology and Technology* 15: 279–292.

Simpson, M.G. 2006. Plant systematics. USA: Academic Press.

- Spayd, S.E., Morris, J.R., Balingen, W.E. and Himelrick, D.J. 1990. Maturity standards, harvesting, postharvest handling and storage. In *Small fruit management*. ed. G.J. Galleta and D.J. Himelrick. pp. 504-528. New Jersey: Prentice Hall.
- Stafford, H.A. 1994. Anthocyanins and betalains: evolution of the mutually exclusive pathways. *Plant Science* 110: 91-98.
- Stephenson, A.G., Devlin, B. and Horton, J.B. 1988. The effects of seed number and prior fruit dominance on the pattern of fruit production in *Cucurbita pepo* (Zucchini squash). *Annals of Botany* 62: 653-661.

- Stuppy, W., Kesseler, R. and Papadakis, A. 2008. *Fruit: Edible, inedible, incredible.* China: Firefly books.
- Tanaka, N., Uehara, K. and Murata, J. 2004. Correlation between pollen morphology and pollination mechanisms in the Hydrochariteceae. *Journal* of *Plant Research* 117, 265-276.
- Tangmitcharoen, S. and Owens, J.N. 1997. Floral biology, pollination, pistil receptivity, and pollen tube growth of teak (*Tectona grandis* Linn f.). *Annals of Botany* 79: 227-241.
- Tel-Zur, N., Abbo, S., Bar-Zvi, D. and Mizrahi, Y. 2004. Genetic relationship among *Hylocereus* and *Selenicereus* vine cacti (Cactaceae): Evidence from hybridization and cytological studies. *Annals of Botany* 94: 527-534.
- To, L.V., Ngu, N., Duc, N.D. and Huong, H.T.T. 2002. Dragon fruit quality and storage life: effect of harvesting time, use of plant growth regulators and modified atmosphere packaging. *Acta Horticulturae* 575: 611-612.
- Toivonen, P.M.A. and Beveridge, T.H.J. 2005. Maturity, ripening and quality relationships. In *Produce degradation: Pathways and prevention.* ed. O. Lamikanra, S.H. Imam and D.O. Ukuku. pp. 55-78. USA: Taylor and Francis.
- Valiente-Banuet, A., Santos Gally, R., Arizmendi, M.C. and Casas, A. 2007. Pollination biology of the hemiepiphytic cactus *Hylocereus undatus* in the Tehuacán Valley, Mexico. *Journal of Arid Environments* 68(1): 1-8.
- Varoquaux, F., Blanvillain, R., Delseny, M. and Gallois, P. 2000. Less is better: new approaches for seedless fruit production. *Trends in Biotechnology* 18: 233-242.
- Varoquaux, P. and Ozdemir, I.S. 2005. Packaging and produce degradation. In *Produce degradation: Pathways and prevention.* ed. O. Lamikanra, S.H. Imam and D.O. Ukuku. pp. 117-154. USA: Taylor and Francis.
- Vendramini, A.L. and Trugo, L.C. 2000. Chemical composition of acerola fruit (*Malpighia punicifolia* L.) at three stages of maturity. *Food chemistry* 72 (2): 195-198.
- Ward, G. and Nussovitch, A. 1996. Peel gloss as a potential indicator of banana ripeness. *Lebensmittel-Wissenschaft und-Technologie* 29(3): 289-294.
- Weatherspoon, L., Mosha, T. and Nnyepi, M. 2005. Nutrient loss. In *Produce degradation: Pathways and prevention.* ed. O. Lamikanra, S.H. Imam and D.O. Ukuku. pp. 223-266. USA: Taylor and Francis.
- Weiss, J., Nerd, A. and Mizrahi, Y. 1994. Flowering behavior and pollination requirements in climbing cacti with fruit crop potential. *HortScience* 29(12): 1487-1492.

- Western, T.L., Skinner, D.J and Haughn, G.W. 2000. Differentiation of mucilage secretory cells of the Arabidopsis seed coat. *Plant Physiology* 122: 345-355.
- Wetzstein, H.Y. and Sparks, D. 1985. Structure and *in vitro* germination of the pollen of pecan. *Journal of the American Society for Horticultural Science* 110(6): 778-781.
- Williams, G. and Adam, P. 1999. Pollen sculpture in subtropical rain forest plants: Is wind pollination more common than previously suspected? *Biotropica* 31(3): 520-524.
- Wills, R.B.H., Lee, T.H., Graham, D., McGlasson, W.B. and Hall, E.G. 2007. *Postharvest: An introduction to the physiology and handling of fruit and vegetables.* 5th edition. Australia: New South Wales University Press Limited.
- Wu, L.C., Hsu, H.W., Chen, Y.C., Chiu, C.C., Lin, Y.I. and Ho, J.A. 2006. Antioxidant and antiproliferative activities of red pitaya. *Food Chemistry* 95: 319-327.
- Wybraniec, S., Nowak-Wydra, B., Mitka, K., Kowalski, P. and Mizrahi, Y. 2007. Minor betalains in fruits of *Hylocereus* species. *Phytochemistry* 68: 252-259.
- Yadav, P.K. 2007. *Fruit production technology*. New Delhi: International Book Distributing.
- Yeung, E.C. and Cavey, M.J. 1990. Developmental changes in the inner epidermis of the bean seed coat. *Protoplasma* 154: 45-52.
- Yonemoto, Y. Nomura, K. Ide, M., Inoue, H., Majikina, M. and Okuda, H. 2006. Index for harvesting time of white sapote (*Casimiroa edulis* Llave & Lex.) cv. 'Cuccio'. *Journal of Horticultural Science and Biotechnology* 81 (1): 18-22.
- Zulkarnain, M. 2007. Dragon fruit cultivation in Malaysia. *Tropical Fruit Network* 8: 16-17.