



**UNIVERSITI PUTRA MALAYSIA**

**COPY NUMBER VARIATION OF C-C CHEMOKINE LIGAND 3-LIKE 1  
(CCL3L1), C-C CHEMOKINE RECEPTOR TYPE 5 (CCR5) AND CCR2  
POLYMORPHISMS ON THE OUTCOMES OF ANTIRETROVIRAL  
THERAPY AMONG MALAYSIAN HIV PATIENTS**

**IRMA IZANI MOHAMAD ISA**

**FPSK(p) 2020 1**



**COPY NUMBER VARIATION OF C-C CHEMOKINE LIGAND 3-LIKE 1  
(*CCL3L1*), C-C CHEMOKINE RECEPTOR TYPE 5 (*CCR5*) AND *CCR2*  
POLYMORPHISMS ON THE OUTCOMES OF ANTIRETROVIRAL  
THERAPY AMONG MALAYSIAN HIV PATIENTS**

By

**IRMA IZANI MOHAMAD ISA**



**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

**March 2020**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment  
of the requirement for the Degree of Doctor of Philosophy

**COPY NUMBER VARIATION OF C-C CHEMOKINE LIGAND 3-LIKE 1  
(*CCL3L1*), CHEMOKINE RECEPTOR TYPE 5 (*CCR5*) AND *CCR2*  
POLYMORPHISMS ON THE OUTCOMES OF ANTIRETROVIRAL  
THERAPY AMONG MALAYSIAN HIV PATIENTS**

By

**IRMA IZANI MOHAMAD ISA**

**March 2020**

**Chair :Suhaili Abu Bakar @ Jamaludin, PhD**  
**Faculty :Medicine and Health Sciences**

HIV/AIDS is a significant burden in Malaysia, affecting more than 90, 000 of the population. Several host genetic variations have been implicated in the pathogenesis of HIV infection, particularly on HIV susceptibility and disease progression to AIDS. These include *CCR5* and *CCL3L1*, which encode for the *CCR5* receptor and the ligand for the *CCR5* receptor respectively. However, there is a noticeable discordancy in CD4 count recovery and viral load suppression between individuals during the highly active anti-retroviral therapy (HAART). However, the predictive value of these genetic variants on patients' responses to the HAART is largely unknown. Therefore, the main objective of this study is to determine the impact of *CCL3L1* copy number variation and selected *CCR5/CCR2* polymorphisms on HIV susceptibility, CD4 count recovery and viral load suppression among Malaysian HIV patients during early HAART. Besides, the influence of socio-demographic and clinical factors is also considered. This cross-sectional study involved 182 HIV-positive patients of Malay, Chinese and Indian ethnicities who were attending out-patient clinics of three hospitals in Malaysia and 150 non-HIV (comparative) subjects. A subset of 170 HIV subjects who were receiving the standard first-line HAART regimen with available CD4 count and viral load data were selected for analyses on immunological and virological responses for up to 12 months after the initiation of HAART. Typing of *CCL3L1* copy number used parologue ratio test (PRT) followed by the copy number validation by microsatellites analyses. *CCR5-Δ32* was genotyped by using PCR while both *CCR5-R223Q* and *CCR2-V64I* were identified by using PCR-restriction fragment length polymorphism (PCR-RFLP). Logistic regression was used to determine the effect of the predictors on achieving the target outcomes of CD4 count  $\geq 500$  cells/mm<sup>3</sup> and viral load  $\leq 50$  copies/mL. Lower than average *CCL3L1* copy number was associated with an increased risk of acquiring HIV-1 in Malay ethnic. Susceptibility to HIV was also reduced by having the mutant allele of *CCR5-R223Q*. In multivariable analysis after adjustment for socio-demographic and clinical factors, lower than average *CCL3L1* copy number predicted a higher chance of CD4 recovery to  $\geq 500$  cells/mm<sup>3</sup> at 8-12 months treatment with HAART. Furthermore, subjects with pre-treatment CD4 count

$\geq 200$  cells/mm $^3$  were associated with at least five times more likely to achieve optimal CD4 recovery in the first 12 months of HAART. Besides, at 8-12 months of HAART, Chinese and Indian subjects were four and seven times respectively more likely to achieve CD4 count  $\geq 500$  cells/mm $^3$  than Malay subjects. Viral load suppression to  $\leq 50$  copies/mL was not associated with the *CCL3L1* copy number or *CCR5/CCR2* genetic factors. Rather, the viral load suppression was negatively predicted by pre-treatment viral load  $\geq 100,000$  copies/mL and positively predicted by CD4 count  $\geq 200$  cells/mm $^3$  in the first 4-6 months of HAART. In conclusion, both high *CCL3L1* copy number and the mutant *CCR5-R223Q* are the potential factors that prevent HIV infection. The present study also highlights the contribution of low *CCL3L1* copy number, ethnicity and early HAART initiation in order to achieve the optimal CD4 count recovery thus improving the management of HIV treatment.

**Keywords:** *CCL3L1*, *CCR5*, CD4 count, viral load, susceptibility, HAART, Malaysia



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**VARIASI NOMBOR SALINAN C-C KEMOKIN LIGAN 3-SEPERTI 1  
(*CCL3L1*), C-C KEMOKIN RESEPTOR JENIS 5 (*CCR5*) DAN *CCR2*  
TERHADAP KESAN-KESAN TERAPI ANTIRETROVIRAL DI KALANGAN  
PESAKIT HIV DI MALAYSIA**

Oleh

**IRMA IZANI MOHAMAD ISA**

**Mac 2020**

**Pengerusi :Suhaili Abu Bakar @ Jamaludin, PhD**  
**Fakulti :Perubatan dan Sains Kesihatan**

HIV/AIDS merupakan beban yang signifikan di Malaysia, yang menjelaskan lebih daripada 90,000 penduduk. Beberapa variasi genetik pada hos terlibat dalam patogenesis jangkitan HIV, terutamanya terhadap risiko jangkitan dan perkembangan penyakit kepada AIDS. Ini termasuk *CCR5* serta *CCL3L1*, yang masing-masing mengekodkan reseptor *CCR5* dan ligan untuk reseptor *CCR5*. Walau bagaimanapun, terdapat percanggahan yang ketara dalam pemulihan kiraan CD4 dan penurunan beban virus antara individu semasa terapi anti-retroviral sangat aktif (HAART). Walau bagaimanapun, kesan ramalan variasi genetik terhadap tindak balas pesakit terhadap HAART sebahagian besarnya tidak diketahui. Oleh itu, objektif utama kajian ini adalah untuk menentukan kesan variasi nombor salinan pada *CCL3L1* dan polimorfisme *CCR5/CCR2* yang dipilih terhadap risiko jangkitan HIV, pemulihan kiraan CD4 dan penurunan beban virus di kalangan pesakit HIV di Malaysia semasa peringkat awal rawatan HAART. Selain itu, pengaruh faktor sosio-demografi dan klinikal juga dipertimbangkan. Kajian keratan rentas ini melibatkan 182 pesakit HIV-positif daripada etnik Melayu, Cina dan India yang menghadiri klinik pesakit luar dari tiga hospital di Malaysia dan 150 subjek bukan HIV (kawalan). Subset dari 170 subjek HIV yang menerima regimen HAART standard pertama dengan data kiraan CD4 dan data beban virus yang tersedia dipilih untuk dianalisis tentang respon imunologi dan virologi sehingga 12 bulan selepas permulaan HAART. Nombor salinan *CCL3L1* dikira menggunakan ujian nisbah paralogi (PRT) diikuti dengan pengesahan nombor salinan oleh analisis mikrosatellite. Pengesanan genetik *CCR5-Δ32* telah menggunakan tindak balas rantai polimerase (PCR) manakala kedua-dua *CCR5-R223Q* dan *CCR2-V64I* telah dikenal pasti menggunakan polimorfisme panjang pecahan PCR (PCR-RFLP). Regresi logistik digunakan untuk menentukan nilai prediktor untuk mencapai hasil sasaran jumlah  $CD4 \geq 500 \text{ sel/mm}^3$  dan beban virus  $\leq 50 \text{ salinan/mL}$ . Bilangan salinan *CCL3L1* yang lebih rendah daripada purata dikaitkan dengan peningkatan risiko mendapatkan HIV-1 dalam etnik Melayu. Risiko jangkitan terhadap HIV juga dikurangkan dengan mempunyai alel mutan *CCR5-R223Q*. Dalam analisis pelbagai pemboleh ubah selepas penyesuaian untuk faktor

sosio-demografi dan klinikal, bilangan salinan *CCL3L1* lebih rendah daripada purata meramalkan kemungkinan pemulihan CD4 yang lebih tinggi kepada  $\geq 500$  sel/mm<sup>3</sup> pada 8-12 bulan rawatan dengan HAART. Tambahan pula, subjek dengan jumlah CD4 pra-rawatan  $\geq 200$  sel/mm<sup>3</sup> dikaitkan dengan sekurang-kurangnya lima kali lebih tinggi kebolehan untuk mencapai pemulihan CD4 optimum dalam 12 bulan pertama rawatan HAART. Di samping itu, pada 8 hingga 12 bulan rawatan HAART, subjek berbangsa Cina dan India mempunyai empat dan tujuh kali ganda lebih kecenderungan untuk mencapai jumlah CD4 sebanyak  $\geq 500$  sel/mm<sup>3</sup> berbanding subjek berbangsa Melayu. Penurunan beban virus kepada  $\leq 50$  salinan/mL tidak dikaitkan dengan faktor genetik daripada nombor salinan *CCL3L1* dan *CCR5/CCR2*. Sebaliknya penurunan beban virus diramalkan secara negatif oleh beban viral pra-rawatan  $\geq 100,000$  salinan/mL dan diramalkan secara positif oleh jumlah CD4  $\geq 200$  sel/mm<sup>3</sup> dalam 4-6 bulan pertama rawatan HAART. Kesimpulannya, kedua-dua nombor salinan *CCL3L1* yang tinggi dan *CCR5-R223Q* yang mutan adalah faktor yang berpotensi untuk mengelakkan jangkitan HIV. Kajian ini juga menyerlahkan sumbangan nombor salinan *CCL3L1* yang rendah, etnik dan permulaan HAART pada peringkat yang awal untuk mencapai pemulihan kiraan CD4 yang optimal, seterusnya memperbaiki pengurusan rawatan HIV.

**Kata kunci:** *CCL3L1*, *CCR5*, kiraan CD4, beban virus, risiko jangkitan, HAART, Malaysia

## **ACKNOWLEDGEMENTS**

In the name of Allah, the Most Beneficent and the Most Merciful. Alhamdulillah, all the praises to Allah SWT for all the blessings, experiences and memories throughout my journey as a PhD scholar.

First and foremost, I would like to express my special gratitude to my head supervisor, Dr. Suhaili Abu Bakar, for her continuous guidance, motivation and advice throughout the course of my study. I am also thankful for the help, support and suggestions from my co-supervisors, Dr. Ahmad Kashfi, Dr. Zulkefley Othman and Prof. Cheah Yoke Kqueen. This work would also have been impossible without the assistance of Infectious Disease consultants, medical specialists and hospital staff from Hospital Sungai Buloh, Hospital Sultanah Nur Zahirah, Kuala Terengganu and Hospital Kajang especially Dr. Azureen, Dr. Siti Dalila, Mr. Sufian, SN Noraziah@Kak Nor and SN Intan. I also want to thank all my fellow friends, Jalilah, Emira, Hanini, Azmah and Umi as well as all the staff from the Biomedical and Bioinformatic laboratory for lending me a hand whenever I need their help especially during data collection.

To my husband, Abdu Syakur bin Hamid, thank you for your unconditional love and support that make things easier and possible for me. To my parents, Mohamad Isa Sabar and Umi Kalsom Ismail, thank you for your prayers and motivational words. To my children, Abdul Basit, Nusaybah, Abdullah Zuhdi and Ahmad Zubayr, thank you for being my great source of motivation and strength. No words can express how grateful I am to have all of you in my life. Alhamdulillah and thank you so much.

I certify that a Thesis Examination Committee has met on 12 March 2020 to conduct the final examination of Irma Izani Mohamad Isa on her thesis entitled "Copy Number Variation of C-C Chemokine Ligand 3-Like 1 (*CCL3L1*), C-C Chemokine Receptor Type 5 (*CCR5*) and *CCR2* Polymorphisms on the Outcomes of Antiretroviral Therapy among Malaysian HIV Patients" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Huzwah binti Khazaai, PhD**

Senior Lecturer

Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Noorjahan Banu binti Mohammed Alitheen, PhD**

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Rukman bin Awang Hamat, PhD**

Associate Professor

Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Edward John Hollox, PhD**

Senior Lecturer

Department of Genetics and Genome Biology  
University of Leicester  
United Kingdom  
(External Examiner)



A handwritten signature in black ink, appearing to read 'Zuriati Ahmad Zukarnain', is placed over the watermark.

---

**ZURIATI AHMAD ZUKARNAIN, PhD**

Professor Ts. and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 02 June 2020

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Suhaili Abu Bakar @ Jamaludin, PhD**

Senior Lecturer

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Chairman)

**Cheah Yoke Kqueen, PhD**

Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Member)

**Zulkefley Othman, PhD**

Senior Lecturer

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Member)

**Ahmad Kashfi Abd. Rahman, MD, MMed**

Consultant in Infectious Disease

Department of Medicine

Hospital Sultanah Nur Zahirah Terengganu

(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

### **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_

Date: \_\_\_\_\_

Name and Matric No: Irma Izani Mohamad Isa, GS45516

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_

Name of Chairman  
of Supervisory Committee: Dr. Suhaili Abu Bakar @ Jamaludin

Signature: \_\_\_\_\_

Name of Member  
of Supervisory Committee: Professor Dr. Cheah Yoke Kqueen

Signature: \_\_\_\_\_

Name of Member  
of Supervisory Committee: Dr. Zulkefley Othman

Signature: \_\_\_\_\_

Name of Member  
of Supervisory Committee: Dr. Ahmad Kashfi Abd. Rahman

## TABLE OF CONTENT

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF FIGURES</b>	xiii
<b>LIST OF TABLES</b>	xiv
<b>LIST OF APPENDICES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xvi
 <b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	1
1.1 Background of study	1
1.2 Problem statements	4
1.3 Significance of study	4
1.4 Objectives	5
1.5 Hypothesis	5
1.6 Conceptual framework of the study	6
<b>2 LITERATURE REVIEW</b>	7
2.1 The epidemiology of HIV	7
2.1.1 HIV worldwide	7
2.1.2 HIV in Malaysia	11
2.2 Biological properties of HIV virus and the virus life cycle	13
2.2.1 Physical and biological features of HIV	13
2.2.2 The life cycle of HIV	15
2.3 HIV pathogenesis and anti-retroviral therapy	18
2.3.1 Pathogenesis of HIV	18
2.3.2 Anti-retroviral therapy and future prospects of HIV treatment	21
2.4 CD4 count and viral load	25
2.4.1 CD4 count and CD4 recovery during antiretroviral therapy	25
2.4.2 Viral load and virological suppression during antiretroviral therapy	26
2.5 Influence of genetic and other factors on HIV infection	29
2.5.1 Factors associated with HIV susceptibility	29
2.5.2 Factors associated with CD4 count	31
2.5.3 Factors associated with viral load	33
2.6 <i>CCL3L1</i> and the association with HIV	34
2.6.1 <i>CCL3L1</i> and copy number variants	34
2.6.2 Association of <i>CCL3L1</i> copy number with HIV	36
2.6.3 Measurement of CNVs copy number	38

2.7	<i>CCR5</i> and <i>CCR2</i> and the association with HIV	40
2.7.1	<i>CCR5</i> and <i>CCR2</i>	40
2.7.2	Association of <i>CCR5</i> and <i>CCR2</i> with HIV	43
<b>3</b>	<b>GENERAL MATERIALS AND METHODS</b>	<b>46</b>
3.1	Study flow chart	46
3.2	Study population	47
3.2.1	Inclusion criteria for HIV	47
3.2.2	Inclusion criteria for non-HIV (comparative)	47
3.2.3	Exclusion criteria	47
3.3	Ethical clearance	48
3.4	Subjects' intervention	48
3.4.1	Informed consent and proforma filling-up	48
3.4.2	Blood sample collection	49
3.6	Collection of CD4 count, viral load and other clinical data	49
3.7	DNA extraction	50
3.8	Typing of <i>CCL3L1</i> gene copy number	51
3.8.1	Paralogue ratio test (PRT)	51
3.8.2	Microsatellites analysis	52
3.9	Genotyping of <i>CCR5</i> and <i>CCR2</i> polymorphisms	54
3.9.1	Genotyping of <i>CCR5</i> -Δ32 using PCR	54
3.9.2	Genotyping of <i>CCR5-R223Q</i> using PCR-RFLP	54
3.9.3	Genotyping of <i>CCR2-V64I</i> using PCR-RFLP	55
3.10	Electrophoresis	57
3.10.1	Gel electrophoresis	57
3.10.2	Capillary electrophoresis and data analysis	58
3.11	DNA sequencing	59
3.12	Statistical analysis	59
<b>4</b>	<b>INFLUENCE OF SOCIO-DEMOGRAPHIC AND CLINICAL FACTORS ON CD4 RECOVERY AND VIRAL LOAD SUPPRESSION FOLLOWING HAART</b>	<b>60</b>
4.1	Introduction	60
4.2	Materials and methods	61
4.2.1	Study population	61
4.2.2	Data collection	61
4.2.3	Statistical analysis	61
4.3	Results	62
4.3.1	Socio-demographic and clinical profiles	62
4.3.2	Trends of CD4 count recovery and viral load suppression during initial HAART	63
4.3.3	Association of socio-demographic and clinical factors with CD4 count recovery	65
4.3.4	Association of socio-demographic and clinical factors with viral load suppression	72
4.4	Discussion	79

<b>5</b>	<b>INFLUENCE OF <i>CCL3L1</i> COPY NUMBER ON HIV SUSCEPTIBILITY, CD4 RECOVERY AND VIRAL LOAD SUPPRESSION FOLLOWING ANTIRETROVIRAL THERAPY</b>	82
5.1	Introduction	82
5.2	Materials and methods	84
5.2.1	Study population	84
5.2.2	Subjects' intervention and data collection	84
5.2.3	Typing of <i>CCL3L1</i> copy number using PRTs	84
5.2.4	Statistical analysis	85
5.3	Results	86
5.3.1	Internal analysis for quantification of <i>CCL3L1</i> copy number using PRTs	86
5.3.2	Distribution of <i>CCL3L1</i> copy number in HIV and non-HIV	92
5.3.3	Association of <i>CCL3L1</i> copy number with CD4 count recovery	94
5.3.4	Association of <i>CCL3L1</i> copy number with viral load suppression	98
5.4	Discussion	98
<b>6</b>	<b>INFLUENCE OF <i>CCR5</i> AND <i>CCR2</i> POLYMORPHISMS ON HIV SUSCEPTIBILITY, CD4 RECOVERY AND VIRAL LOAD SUPPRESSION FOLLOWING ANTIRETROVIRAL THERAPY</b>	102
6.1	Introduction	102
6.2	Materials and methods	103
6.2.1	Study population	103
6.2.2	Subjects' intervention and data collection	103
6.2.3	Genotyping of <i>CCR5</i> and <i>CCR2</i> polymorphisms	103
6.2.4	Statistical analysis	104
6.3	Results and discussion	105
6.3.1	Gel electrophoresis and sequencing images	105
6.3.2	Distribution of <i>CCR5</i> -Δ32, <i>CCR5</i> -R223Q and <i>CCR2</i> -V64I genotypes in HIV subjects	107
6.3.3	Association of <i>CCR5</i> -Δ32, <i>CCR5</i> -R223Q and <i>CCR2</i> -V64I with CD4 recovery	111
6.3.4	Association of <i>CCR5</i> -Δ32, <i>CCR5</i> -R223Q and <i>CCR2</i> -V64I with viral load suppression	113
<b>7</b>	<b>CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	117
7.1	Conclusion	117
7.2	Recommendation for future studies	119
<b>REFERENCES</b>		120
<b>APPENDICES</b>		146
<b>BIODATA OF STUDENT</b>		164
<b>LIST OF PUBLICATIONS</b>		165

## LIST OF FIGURES

<b>Figure</b>	<b>Page</b>
1.1 Conceptual Framework	6
2.1 Estimated number of people living with HIV/AIDS worldwide	7
2.2 Evolution of the HIV epidemic from 1980 to 2015	8
2.3 Key populations in HIV/AIDS pandemics	9
2.4 Number of new HIV infection in Malaysia from 1986 to 2016	11
2.5 Diagram of HIV a mature virion	13
2.6 The life cycle of HIV	15
2.7 HIV entry processes: i. receptor binding and ii. membrane fusion	16
2.8 Stages of HIV infection	19
2.9 HIV replication cycle as the targets for drug development	21
2.10 Potential approaches of genome editing for anti-HIV therapy	24
2.11 Schematic diagram of <i>CCL3L1</i> PRT assays	39
2.12 Structure of <i>CCR5</i> protein	40
2.13 <i>CCR5</i> polymorphisms and haplotypes	42
2.14 Geographical distribution of <i>CCR5</i> -Δ32 mutation	44
2.15 Schematic diagram of the position of <i>CCR5</i> variants including <i>CCR5-R223Q</i>	44
3.1 Study flow chart	46
4.1 Trend of CD4 count at initiation and during initial treatment with HAART	64
4.2 CD4 recovery and viral load suppression during initial treatment with HAART	64
4.3 Trend CD4 count at initiation and during initial treatment with HAART based on ethnicity	72
5.1 Examples of capillary electrophoresis images from PRTs	86
5.2 Examples of linear regression generated using reference control samples for <i>CCL3A</i> , <i>CCL4A</i> and <i>LTR61A</i> PRT systems	87
5.3 Agreement of unrounded copy number for <i>CCL3A</i> vs <i>CCL4A(A)</i> and <i>CCL3A</i> vs <i>LTR61A (B)</i>	88
5.4 Distribution of unrounded copy number mean of PRT assays	89
5.5 Examples of capillary electrophoresis images from microsatellites amplifications	90
5.6 Summary of <i>CCL3L1</i> copy number typing by PRTs and validation by microsatellites analysis	91
5.7 Distribution of <i>CCL3L1</i> gene copy number in HIV and non-HIV subjects in all ethnicities	92
5.8 Distribution of <i>CCL3L1</i> gene copy number in HIV subjects of Malay, Chinese and Indian ethnicities	93
5.9 Trend of CD4 count based on two categories of <i>CCL3L1</i> copy number	95
6.1 Gel electrophoresis images showing PCR or PCR-RFLP products of <i>CCR5</i> -Δ32 (A), <i>CCR5-R223Q</i> (B) and <i>CCR2-V64I</i> (C)	105
6.2 Images of direct sequencing showing heterozygous <i>CCR5</i> -Δ32 (A), heterozygous <i>CCR5-R223Q</i> (B) and heterozygous <i>CCR2-V64I</i> (C)	106
7.1 Summary of the study results	117

## LIST OF TABLES

Table	Page
2.1 WHO definitions of clinical, immunological and virological failure	27
2.2 Examples of association of host genetic polymorphisms with disease progression and CD4 count `recovery	31
2.3 Association of <i>CCL3L1</i> copy number (CN) with HIV infection	37
3.1 Components of PCR reagents used in duplex PCR for PRTs	52
3.2 Components of PCR reagents used in microsatellites analysis of <i>TTAT17</i> and <i>TATC17</i>	53
3.3 Components of PCR reactions to amplify <i>CCR5-Δ32</i> & <i>CCR5-R223Q</i>	54
3.4 Components of RFLP mixture for <i>CCR5-R223Q</i>	55
3.5 Components of PCR reactions for <i>CCR2-V64I</i>	56
3.6 Components of RFLP mixture for <i>CCR2-V64I</i>	56
4.1 Distribution socio-demographic and clinical characteristics of HIV subjects	62
4.2 Association of socio-demographic and clinical factors with CD4 count	66
4.3 Relationship of socio-demographic and clinical factors with CD4 recovery	69
4.4 Association of socio-demographic and clinical factors with viral load suppression	73
4.5 Relationship of socio-demographic and clinical factors with viral load suppression	76
5.1 Comparison of <i>CCL3L1</i> copy number distribution between HIV and non-HIV subjects according to ethnicity	94
5.2 Association and relationship of <i>CCL3L1</i> copy number with CD4 recovery	95
5.3 Relationship of <i>CCL3L1</i> copy number and other factors with CD4 recovery at 8-12 months of HAART	97
5.4 Association and relationship of <i>CCL3L1</i> copy number with viral load suppression	98
6.1 Association of <i>CCR5-Δ32</i> , <i>CCR5-R223Q</i> and <i>CCR2-V64I</i> genotypes in HIV subjects with ethnicity	108
6.2 Association of <i>CCR5-Δ32</i> , <i>CCR5-R223Q</i> and <i>CCR2-V64I</i> genotypes with HIV susceptibility in different ethnicities	110
6.3 Association and relationship of <i>CCR5-Δ32</i> , <i>CCR5-R223Q</i> and <i>CCR2-V64I</i> with CD4 recovery	112
6.4 Association and impact of <i>CCR5-Δ32</i> , <i>CCR5-R223Q</i> and <i>CCR2-V64I</i> with viral load suppression	114
6.5 Relationship of <i>CCR5-R223Q</i> and other factors with viral load suppression at 8-12 months of HAART	115

## LIST OF APPENDICES

<b>Appendix</b>		<b>Page</b>
A-1	Ethical approval by MREC for HIV subjects	146
A-2	Ethical approval by JKEUPM for non-HIV subjects	147
A-3	Participant Information Sheet and Informed Consent Form	148
A-4	Proforma for HIV subjects	153
A-5	Data Collection Form	155
B-1	Agreement between inferred copy number and known copy number of the reference controls	157
B-2	Complete dataset for PRTs and microsatellites analyses	158



## LIST OF ABBREVIATIONS

3TC	lamivudine
AIDS	acquired immunodeficiency syndrome
APC	antigen-presenting cells
APOBEC3G	apolipoprotein B mRNA-editing enzyme, catalytic polypeptide like 3G
ART	antiretroviral therapy
ARV	antiretroviral
AZT	3'-azido-3'-deoxythymidine / zidovudine
Bim	Bcl-2–interacting molecule
bp	basepair
C4	complement component C4
cART	combination-ART
CCL3	C-C chemokine ligand 3
CCL3L1	C-C chemokine ligand 3-like 1
CCR5	C-C chemokine receptors type 5
CCR5-Δ32	CCR5-32 deletion
CCR2	C-C chemokine receptor type 2
CDC	U.S. Centers for Disease Control and Prevention
CGH	comparative genomic hybridisation
CN	copy number
CNPs	copy number polymorphisms
CNVs	copy number variants
CRISPR	clustered, regularly-interspaced, short palindromic repeats
CRF	circulating recombinant form
CTLs	cytotoxic T lymphocytes
CXCL12	C-X-C motif chemokine type 12
CXCR4	C-X-C chemokine receptor type 4
DCs	dendritic cells
DC-SIGN	dendritic cell-specific intercellular adhesion molecule-3-grabbing non-integrin
DEFB	β-defensins
DNA	deoxyribonucleic acid
dNTP	deoxynucleoside triphosphate
ECACC	European Collection of Cell Cultures
EFV	efavirenz
env	envelope
FCGR3B	Fc gamma receptor 3B
FDC	fixed dose combination
FISH	fluorescence in-situ hybridization
FTC	emtricitabine
gag	group specific antigen
gp120	glycoprotein-120
GRGs	genetic risk groups
GWAS	genome-wide association studies
HAART	highly active anti-retroviral therapy
HCV	hepatitis C
HH	human haplotype

HIV	human immunodeficiency virus
HLA	human leukocyte antigen
HSPC	hematopoietic stem/ progenitor cells
IA-HOD-IA	Investigator's Agreement, Head of Department and Organizational/ Institutional Approval
IBBS	Integrated Bio-Behavioural Survey
IDU	injecting drug user
IFN	interferon
IN	interleukin
INSTI	integrase strand transfer inhibitors
IRIS	immune reconstitution inflammatory syndrome
IVDU	intravenous drug use
JKEUPM	Jawatankuasa Etika Manusia Universiti Putra Malaysia
KIR	killer cell immunoglobulin-like receptors
LTNP	long-term non-progressor
MAPH	multiplex amplifiable probe hybridization
MCP-2	monocyte chemotactic protein 2
MHC	major histocompatibility complex
MIP-1	macrophage-inflammatory protein 1
MLPA	multiplex ligation-dependent probe amplification
MREC	Medical Research and Ethics Committee
mRNA	messenger ribonucleic acid
MSM	men who have sex with men
MVC	maraviroc
NAHR	non-allelic-homologous recombination
NK	natural killer
NMRR	National Medical Research Registry
NNRTI	non-nucleoside reverse transcriptase inhibitor
NRTI	nucleoside reverse transcriptase inhibitor
NSI	non-syncytium-inducing
OIs	opportunistic infections
OR	odd ratio
ORF	open reading frame
PCR	polymerase chain reaction
PI	protease inhibitor
PIC	pre-integration complex
PLWH	people living with HIV
<i>pol</i>	polymerase
PrEP	pre-exposure prophylaxis
PRT	parologue ratio test
PWID	people who inject drug
qPCR	real-time PCR
RANTES	regulated upon activation, normal T-cell expressed and secreted
refSNP	reference SNP
RFLP	restriction fragment length polymorphism
RNA	ribonucleic acid
rpm	revolutions per minute
RT	reverse transcriptase
SDF-1	stromal cell-derived factor 1

SI	syncytium-inducing
SLE	systemic lupus erythematosus
SNP	single nucleotide polymorphism
$t_0$	pre-treatment
$t_1$	at 4-6 months of HAART initiation
$t_2$	at 8-12 months of HAART initiation
TBE	Tris/Borate/EDTA
TDF	tenofovir
Th1	type 1 helper T
TLR	Toll-like receptor
TNF	tumor necrosis factor
TRAIL	TNF-related apoptosis-inducing ligand
TRIM5 $\alpha$	tripartite interaction motif 5 alpha
TSG 101	tumor susceptibility gene 101
UGT	uridine diphosphate glucuronyl transferase
UNAIDS	The Joint United Nations Program on AIDS
VL	viral load
WHO	World Health Organization

# CHAPTER 1

## INTRODUCTION

### 1.1 Background of study

The worldwide trends of human immunodeficiency virus (HIV)/acquired immune deficiency syndromes (AIDS) as well as the unfavourable impact of HIV remain among the major and pressing public health challenges (Cahill & Valadéz, 2013), affecting 36.7 million people around globally (UNAIDS, 2017a). The Asian and the Pacific countries contributed 5.1 million or 14% of the total cases of HIV and AIDS globally. In Malaysia, HIV prevalence is approximately 0.4%, which is equal to 93,089 people (Malaysian AIDS Council, 2016). HIV epidemics in Malaysia involves the most-at-risk or key populations that include men who have sex with men (MSM), people who inject drugs (PWID), sex workers, transgender people, prisoners as well as sexual partners of these key populations (Kilmarx, 2009; UNAIDS, 2017c). Social determinants of health, including being female, poor social-economic status and illiteracy level have been important aspects for control and prevention of HIV/AIDS (Alvarez-Uria, Midde, Pakam, & Naik, 2012; Dean & Fenton, 2010).

A unique feature of HIV viruses is that they replicate themselves using the host machinery system after their genetic material being integrated into host DNA, in the form of proviral DNA (Turner & Summers, 1999). In order to productively infect target cells, the very first step in HIV replicative cycle of HIV involves the binding of HIV to CD4 receptor and to a co-receptor, which is either C-C chemokine receptors type 5 (CCR5) or C-X-C chemokine receptor type 4 (CXCR4) (Sierra, Kupfer, & Kaiser, 2005). CCR5 receptor plays a major role during HIV virus entry to host cells by macrophage-tropic or non-syncytium-inducing (NSI) strain at the early events of HIV-1 infection (Bleul, Wu, Hoxie, Springer, & Mackay, 1997; Choe et al., 1996; Deng et al., 1996; Dragic et al., 1996). One of the current antiretroviral therapy approaches is by blocking the CCR5 co-receptor, known as CCR5 inhibitor such as Maraviroc.

Notably, there is a growing number of people living with HIV (PLWH) as HIV patients can live longer due to the effect of taking anti-retroviral treatment (Wang et al., 2016). However, the current antiretroviral therapy (ART) cannot totally eliminate HIV viruses and has many adverse effects. CCR5 receptor is one of the potential target in the effort to cure HIV, by using hematopoietic stem cell gene therapy, which involves transplantation of genetically modified *CCR5* gene to HIV-infected patient (Passaes & Sáez-Cirián, 2014). The insight for *CCR5* genome editing had been derived from the ‘Berlin patient’, who had underwent a stem cells transplant from human leukocyte antigen (HLA)-matched, homozygous *CCR5* 32 deletion (*CCR5-Δ32*) that leads to undetectable viral load for 20 months post-transplantation without antiretroviral

therapy (Hütter et al., 2009; Wang et al., 2016). Furthermore, the development of new HIV treatment modalities are focusing on vaccines and drugs that target the immune systems (Esté & Cihlar, 2010; Passaes & Sáez-Cirián, 2014).

HIV infection is classified into different stages based on CD4 T-cells count (CDC, 2008; World Health Organization, 2007). HIV virus is the same causative agent that causes AIDS, but the diagnosis of AIDS is made when CD4 cell count < 200 and/or there is the presence of AIDS-defining illnesses (World Health Organization, 2007). The hallmark of HIV-1 infection is the continuous declining of CD4 T cells due to increment in destruction as well as reduction in regeneration of CD4 T cell populations (Douek, Picker, & Koup, 2003). Interestingly, there has been a subset of HIV patients, known as long-term nonprogressors (LTNPs) (Pantaleo et al., 1995), who maintain a normal CD4 count for up to seven years without treatment (Okulicz et al., 2009). CD4 count is an important prognostic factor of individuals starting highly active antiretroviral therapy (HAART) and had been previously used to indicate when to initiate HAART (Egger et al., 2002). With the antiretroviral therapy, CD4 count increases rapidly by approximately 100 cell/mm<sup>3</sup> in the first year of starting HAART (Mocroft et al., 2007).

The entry of HIV virus to host cell leads to a rapid viral replication, known as viral load. The host immune system initially responds to the high viral load by the activation of HIV-1-specific cytotoxic or CD8 T-cells. This HIV-1-specific response leads to an initial decline of viral load few weeks after HIV infection, known as viral set point. However, this HIV-specific CD8 T-cells response is targeted at the dominant viral variant only and is rather limited due to viral immune escape (Goonetilleke et al., 2009). Viral load has been used to monitor response to ART as well as treatment failure (World Health Organization, 2016). During HAART, the viral load is targeted to achieve undetectable level or less than 20 copies/mL (Medical Development Division, 2017). In addition, consistency in viral suppression can minimize risk of viral transmission (Attia, Egger, Müller, Zwahlen, & Low, 2009).

Several host genetic determinants of susceptibility to HIV-1 infection, viral load and disease progression have been identified using classical candidate-gene association studies (O'Brien & Nelson, 2004). One example of how genetic can provide protection to HIV-1 infection is by a deletion mutation on *CCR5* gene, known as *CCR5-Δ32* (Liu et al., 1996). Homozygous *CCR5-Δ32* produces a non-functional CCR5 receptor, thus providing resistance to macrophage-tropic HIV infection (Liu et al., 1996). In addition, the ligand-receptor interaction of C-C chemokine ligand 3-like 1 (*CCL3L1*)-CCR5 complex had also been recognized to influence variability in CD4 count depletion and baseline viral load particularly in the era before HAART (Gonzalez et al., 2005). Nonetheless, a low pre-treatment CD4 count could also be an important factor preventing full recovery of CD4 counts (Kelley et al., 2009).

*CCL3L1* is a ligand to CCR5, which is a HIV co-receptor. The *CCL3L1* gene is known as a multi-allelic gene copy number variant (CNV) because it exhibits a variation in gene copy number (CN) of more than three copies (Girirajan, Campbell, & Eichler,

2011). As one of CNVs gene, it has the potential to alter the gene expression level (Stranger et al., 2007) and have been linked to various types of human diseases including HIV (Usher & McCarroll, 2015; Wain, Armour, & Tobin, 2009). Higher *CCL3L1* CN had been associated with a higher resistance to HIV infection, as this up-regulates the gene expression level and functional chemokine, thereby increasing competition of the HIV virus to the CCR5 receptor (Gonzalez et al., 2005; Townson, Barcellos, & Nibbs, 2002). CNV of *CCL3L1* had also been reported to influence adaptive immune response and immune reconstitution during HAART (Ahuja et al., 2008; Shalekoff et al., 2008). However, some studies had shown no replication or conflicting results in some the disease-association studies, which could be due to the types of genotyping techniques used (Clayton et al., 2005).

CCR5 is a G-protein-coupled receptor that binds a number of C-C chemokines including CCL3 and CCL3L1 (Hughes & Nibbs, 2018). The protein, which is found on immune cells like macrophages and monocytes, is involved in co-stimulatory signal for T-cell activation (Molon et al., 2005). Most importantly, it serves as co-receptor for macrophage-tropic strains during the early HIV infection (Dragic et al., 1996). CCR5 has a 75% similarity in amino acid sequence as C-C chemokine receptor type 2 (CCR2) (Combadiere, Ahuja, Tiffany, & Murphy, 1996). *CCR5-Δ32* (NC\_000003.12:g.46373456\_46373487del) and *CCR2-V64I* (NC\_000003.12:g.46357717G>A), together with other polymorphisms in the *CCR5* promoter region are tightly linked together and have been used to define *CCR5* haplotype (Mummidi et al., 2000). The *CCR5-Δ32* mutation had been demonstrated to reduce susceptibility to HIV infection and to delay development of the AIDS in HIV-1- infected individuals while the phenotypic effect of *CCR2-V64I* is a slower progression to AIDS (Kostrikis et al., 1998; McDermott et al., 1998; Smith et al., 1997; Tang et al., 2002). Another *CCR5* variant, known as *CCR5-R223Q* (NC\_000003.12:g.46373570G>A) is closely associated with *CCR2-V64I* and typically found in the Asian populations (Liu et al., 2007).

## **1.2 Problem statements**

Host genetic variability has been generally accepted to influence the pathogenesis of HIV-1 infection, including susceptibility and disease progression (Fellay, 2009; McLaren & Carrington, 2015). Early study had reported on a small subset of individuals who were resistant to HIV-1 infection regardless of the exposure to the HIV virus (Liu et al., 1996). In addition, the rate of CD4 count depletion can vary among individuals, particularly a small percentage of HIV-infected populations, known as LTNP, who maintain high CD4 T-cell counts for many years and did not progress to AIDS (Pantaleo et al., 1995). Among the most established genetic variants influencing HIV-1 susceptibility and immune response are the mutations on *CCR5/CCR2* and *CCL3L1* genes, which encode the CCR5/CCR2 receptor and the ligand of the CCR5 receptor respectively (Gonzalez et al., 1999, 2005). The *CCR5* haplotypes and *CCL3L1* CN also exhibit race-specific HIV-1 disease-modifying effects, which highlights the need for population or ethnic specific disease association studies (Gonzalez et al., 1999).

However, host genetic determinants of HIV susceptibility are still unknown for the Malaysian population, particularly for the *CCL3L1* CN and the *CCR5/CCR2* polymorphisms. Besides, in this era of HAART, studies on genetic factors that contribute to the variation in the individuals' response to the anti-HIV treatment, in term of immune recovery and viral load suppression are still limited. The phenotypic effects of *CCL3L1* CN and *CCR5/CCR2* polymorphisms on HIV-infected patients of the three major ethnicities in Malaysia are largely unknown. Apart from genetic influence on HIV pathogenesis, there is also a lack of knowledge about the influence of socio-demographic and clinical factors on CD4 recovery and viral load suppression in early response to HAART among Malaysian HIV population.

## **1.3 Significance of study**

First of all, this research will enhance the knowledge on the contribution of genetic diversity on HIV/AIDS pathogenesis among HIV-positive population in Malaysia. The main outcome of this study will add a new element to what is known about CNV of the *CCL3L1* and polymorphisms in *CCR5* and *CCR2* genes as the potential genetic host factors determining HIV susceptibility, immune recovery and viral load suppression following antiretroviral therapy. In addition to the genetic polymorphisms being studied, the impact of other factors including socio-demographic and clinical factors on immune recovery and viral load suppression will also be discovered. Finally, the results of this study can potentially lead to the development of predictive tools or biological markers that would set the stage for personalized HIV medicine, by the generation of genetic risk groups to predict how individuals will respond to the current ART.

## **1.4 Objectives**

To investigate the relationship of *CCL3L1* copy number variation and *CCR5/CCR2* genotypes on HIV susceptibility, CD4 recovery and viral load suppression following HAART.

The specific objectives include

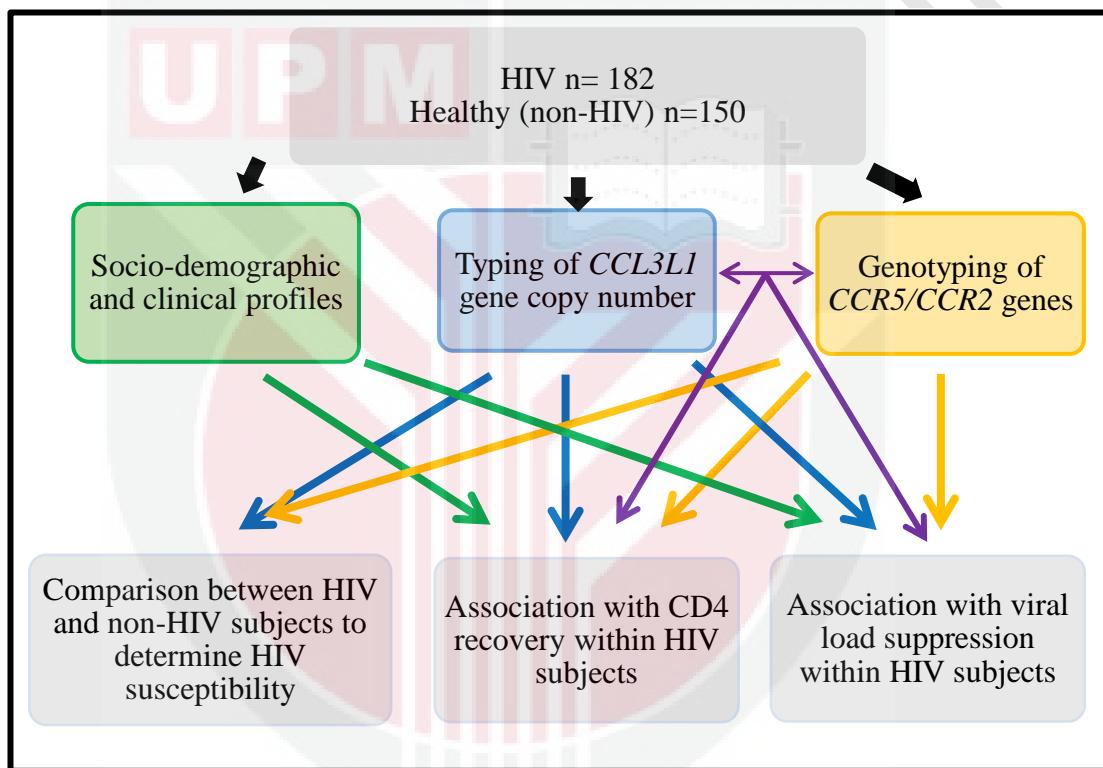
- i. to determine the relationship of socio-demographic and clinical factors on CD4 recovery and viral load suppression following HAART among Malaysian HIV patients (Chapter 4),
- ii. to determine the relationship of *CCL3L1* copy number on HIV susceptibility, and CD4 recovery and viral load suppression following HAART among Malaysian HIV patients (Chapter 5),
- iii. to determine the relationship of *CCR5/CCR2* polymorphisms on HIV susceptibility, and CD4 recovery and viral load suppression following HAART among Malaysian HIV patients (Chapter 6), and
- iv. to generate haplotypes or risk groups based on the relationship of *CCL3L1* copy number and *CCR5/CCR2* polymorphisms on CD4 recovery and viral load suppression following HAART (Chapter 6).

## **1.5 Hypothesis**

- i. Socio-demographic and clinical factors are significantly associated with CD4 recovery and viral load suppression following HAART among Malaysian HIV patients.
- ii. *CCL3L1* copy number is significantly associated with HIV susceptibility, CD4 recovery and viral load suppression following HAART among Malaysian HIV patients.
- iii. *CCR5/CCR2* polymorphisms are significantly associated with HIV susceptibility, CD4 recovery and viral load suppression following HAART among Malaysian HIV patients.

## 1.6 Conceptual framework of the study

This study had four specific objectives, which were separated into three different objective chapters. In the first objective chapter (Chapter 4), the influence of socio-demographic and clinical factors on immune recovery and viral load suppression were discussed. Consequently, the predictive value of *CCL3L1* CN and *CCR5-CCR2* genotypes on HIV susceptibility, and immune recovery and viral load suppression during HAART were analysed in Chapter 5 and Chapter 6 respectively. The last objective, which is the combined effect of *CCR5* and *CCR2* genotypes and *CCL3L1* CN for the prediction of the susceptibility, and viral load and CD4 count during HAART, were also discussed in Chapter 6. **Figure 1.1** summarised the conceptual framework that has been applied in this study.



**Figure 1.1:** Conceptual Framework

## REFERENCES

- Abdulrahman, S. A., Rampal, L., Othman, N., Ibrahim, F., Hayati, K. S., & Radhakrishnan, A. P. (2017). Sociodemographic profile and predictors of outpatient clinic attendance among HIV-positive patients initiating antiretroviral therapy in Selangor, Malaysia. *Patient Preference and Adherence*, 11, 1273–1284. <https://doi.org/10.2147/PPA.S141609>
- Abraha, A., Nankya, I. L., Gibson, R., Demers, K., Tebit, D. M., Johnston, E., ... Arts, E. J. (2009). CCR5- and CXCR4-Tropic Subtype C Human Immunodeficiency Virus Type 1 Isolates Have a Lower Level of Pathogenic Fitness than Other Dominant Group M Subtypes: Implications for the Epidemic. *Journal of Virology*, 83(11), 5592–5605. <https://doi.org/10.1128/JVI.02051-08>
- Abu Bakar, S., Hollox, E. J., & Armour, J. a L. (2009). Allelic recombination between distinct genomic locations generates copy number diversity in human beta-defensins. *Proceedings of the National Academy of Sciences of the United States of America*, 106(3), 853–858. <https://doi.org/10.1073/pnas.0809073106>
- Adamson, C. S., & Freed, E. O. (2010). Novel approaches to inhibiting HIV-1 replication. *Antiviral Research*, 85(1), 119–141. <https://doi.org/10.1016/j.antiviral.2009.09.009>
- Agrawal, L., Lu, X., Qingwen, J., VanHorn-Ali, Z., Nicolescu, I. V., McDermott, D. H., ... Alkhatib, G. (2004). Role for CCR5Delta32 protein in resistance to R5, R5X4, and X4 human immunodeficiency virus type 1 in primary CD4+ cells. *Journal of Virology*, 78(5), 2277–2287. <https://doi.org/10.1128/JVI.78.5.2277>
- Ahuja, S. K., Kulkarni, H., Catano, G., Agan, B. K., F, J., He, W., ... Dolan, M. J. (2008). CCL3L1-CCR5 genotype influences durability of immune recovery during antiretroviral therapy of HIV-1-infected individuals. *Nature Medicine*, 14(4), 413–420. <https://doi.org/10.1038/nm1741.CCL3L1-CCR5>
- Aklillu, E., Odenthal-Hesse, L., Bowdrey, J., Habtewold, A., Ngaimisi, E., Yimer, G., ... Hollox, E. J. (2013). CCL3L1 copy number, HIV load, and immune reconstitution in sub-Saharan Africans. *BMC Infectious Diseases*, 13(1), 536. <https://doi.org/10.1186/1471-2334-13-536>
- Aldhous, M. C., Bakar, S. A., Prescott, N. J., Palla, R., Soo, K., Mansfield, J. C., ... Armour, J. A. L. (2010). Measurement methods and accuracy in copy number variation: Failure to replicate associations of beta-defensin copy number with Crohn's disease. *Human Molecular Genetics*, 19(24), 4930–4938. <https://doi.org/10.1093/hmg/ddq411>
- Alexander, T. S. (2016). Human Immunodeficiency Virus Diagnostic Testing: 30 Years of Evolution. *Clinical and Vaccine Immunology*, 23(4), 249–253. <https://doi.org/10.1128/CVI.00053-16.Editor>
- Alter, G., Heckerman, D., Schneidewind, A., Fadda, L., Kadie, C. M., Carlson, J. M., ... Altfeld, M. (2011). HIV-1 adaptation to NK-cell-mediated immune pressure. *Nature*, 476(7358), 96–101. <https://doi.org/10.1038/nature10237>
- Alvarez-Uria, G., Midde, M., Pakam, R., & Naik, K. (2012). Gender differences, routes of transmission, socio-demographic characteristics and prevalence of HIV related infections of adults and children in an HIV cohort from a rural district of India. *Infectious Disease Reports*, 4:e19. <https://doi.org/10.4081/idr.2012.e19>
- Ammaranond, P., Sanguansitthianan, S., Phaengchomduan, P., Sae-Lee, C., & Mardkhumchan, S. (2013). Impact of CCR2 and SDF1 Polymorphisms on Disease Progression in HIV-Infected Subjects in Thailand. *Journal of Clinical*

- Laboratory Analysis*, 27(1), 38–44. <https://doi.org/10.1002/jcla.21559>
- Ansari-Lari, M. A., Liu, X. M., Metzker, M. L., Rut, A. R., & Gibbs, R. A. (1997). The extent of genetic variation in the CCR5 gene. *Nature Genetics*. <https://doi.org/10.1038/ng0797-221>
- Antiretroviral Therapy Cohort Collaboration. (2010). Causes of Death in HIV-1-Infected Patients Treated with Antiretroviral Therapy, 1996–2006: Collaborative Analysis of 13 HIV Cohort Studies. *Clinical Infectious Diseases*, 50(10), 1387–1396. <https://doi.org/10.1086/652283>
- Appay, V., & Sauce, D. (2008, January 1). Immune activation and inflammation in HIV-1 infection: Causes and consequences. *Journal of Pathology*. John Wiley & Sons, Ltd. <https://doi.org/10.1002/path.2276>
- Aquaro, S., Menten, P., Struyf, S., Proost, P., Damme, J. O. V. A. N., Clercq, E. D. E., & Schols, D. (2001). The LD78beta Isoform of MIP-1alpha Is the Most Potent CC-Chemokine in Inhibiting CCR5-Dependent Human Immunodeficiency Virus Type 1 Replication in Human Macrophages. *Journal of Virology*, 75(9), 4402–4406. <https://doi.org/10.1128/JVI.75.9.4402>
- Arenzana-Seisdedos, F., & Parmentier, M. (2006). Genetics of resistance to HIV infection: Role of co-receptors and co-receptor ligands. *Seminars in Immunology*, 18(6), 387–403. <https://doi.org/10.1016/j.smim.2006.07.007>
- Armour, J. A. L., Palla, R., Zeeuwen, P. L. J. M., Heijer, M. Den, Schalkwijk, J., & Hollox, E. J. (2007). Accurate, high-throughput typing of copy number variation using parologue ratios from dispersed repeats. *Nucleic Acids Research*, 35(3). <https://doi.org/10.1093/nar/gkl1089>
- Arts, E. J., & Hazuda, D. J. (2012). HIV-1 antiretroviral drug therapy. *Cold Spring Harbor Perspectives in Medicine*, 2(4). <https://doi.org/10.1101/cshperspect.a007161>
- Atashili, J., Poole, C., Ndumbe, P. M., Adimora, A. A., & Smith, J. S. (2008). Bacterial vaginosis and HIV acquisition: A meta-analysis of published studies. *AIDS*, 22(12), 1493–1501. <https://doi.org/10.1097/QAD.0b013e3283021a37>
- Attia, S., Egger, M., Müller, M., Zwahlen, M., & Low, N. (2009). Sexual transmission of HIV according to viral load and antiretroviral therapy: Systematic review and meta-analysis. *AIDS*, 23(11), 1397–1404. <https://doi.org/10.1097/QAD.0b013e32832b7dca>
- Auvert, B., Taljaard, D., Lagarde, E., Sobngwi-Tambekou, J., Sitta, R., & Puren, A. (2005). Randomized, controlled intervention trial of male circumcision for reduction of HIV infection risk: The ANRS 1265 trial. *PLoS Medicine*, 2(11), 1112–1122. <https://doi.org/10.1371/journal.pmed.0020298>
- Baker, J. V., Peng, G., Rapkin, J., Abrams, D. I., Silverberg, M. J., MacArthur, R. D., ... Neaton, J. D. (2008). CD4+ count and risk of non-AIDS diseases following initial treatment for HIV infection. *AIDS*, 22(7), 841–848. <https://doi.org/10.1097/QAD.0b013e3282f7cb76>
- Banerjee, A., Chitnis, U., Jadhav, S., Bhawalkar, J., & Chaudhury, S. (2009). Hypothesis testing, type I and type II errors. *Industrial Psychiatry Journal*, 18(2), 127. <https://doi.org/10.4103/0972-6748.62274>
- Bangalore, S., Kamalakkannan, G., Parkar, S., & Messerli, F. H. (2007). Fixed-Dose Combinations Improve Medication Compliance: A Meta-Analysis. *American Journal of Medicine*, 120(8), 713–719. <https://doi.org/10.1016/j.amjmed.2006.08.033>
- Barbaro, G., Scozzafava, A., Mastrolorenzo, A., & Supuran, C. T. (2005). Highly active antiretroviral therapy: Current state of the art, new agents and their

- pharmacological interactions useful for improving therapeutic outcome. *Current Pharmaceutical Design*, 11(14), 1805–1843. <https://doi.org/10.2174/1381612053764869>
- Barmania, F., & Pepper, M. S. (2013). C-C chemokine receptor type five (CCR5): An emerging target for the control of HIV infection. *Applied & Translational Genomics*, 2, 3–16. <https://doi.org/10.1016/j.atg.2013.05.004>
- Barmania, F., Potgieter, M., & Pepper, M. S. (2013). Mutations in C-C chemokine receptor type 5 (CCR5) in South African individuals. *International Journal of Infectious Diseases : IJID : Official Publication of the International Society for Infectious Diseases*, 17(12), e1148-53. <https://doi.org/10.1016/j.ijid.2013.06.009>
- Bashirova, A. A., Bleiber, G., Qi, Y., Hutcheson, H., Yamashita, T., Johnson, R. C., ... Carrington, M. (2006). Consistent Effects of TSG101 Genetic Variability on Multiple Outcomes of Exposure to Human Immunodeficiency Virus Type 1. *Journal of Virology*, 80(14), 6757–6763. <https://doi.org/10.1128/JVI.00094-06>
- Bendel, R. B., & Afifi, A. A. (1977). Comparison of stopping rules in forward “stepwise” regression. *Journal of the American Statistical Association*, 72(357), 46–53. <https://doi.org/10.1080/01621459.1977.10479905>
- Beyrer, C., Baral, S. D., Van Griensven, F., Goodreau, S. M., Chariyalertsak, S., Wirtz, A. L., & Brookmeyer, R. (2012). Global epidemiology of HIV infection in men who have sex with men. *The Lancet*, 380(9839), 367–377. [https://doi.org/10.1016/S0140-6736\(12\)60821-6](https://doi.org/10.1016/S0140-6736(12)60821-6)
- Bhatnagar, I., Singh, M., Mishra, N., Saxena, R., Thangaraj, K., Singh, L., & Sk, S. (2009). The Latitude Wise Prevalence of the CCR5-Δ 32-HIV Resistance Allele in India. *Balk. J. Med. Genet.*, 12(2), 17–27.
- Blaak, H., van't Wout, A. B., Brouwer, M., Hooibrink, B., Hovenkamp, E., & Schuitemaker, H. (2000). In vivo HIV-1 infection of CD45RA(+)CD4(+) T cells is established primarily by syncytium-inducing variants and correlates with the rate of CD4(+) T cell decline. *Proceedings of the National Academy of Sciences of the United States of America*, 97(3), 1269–1274. <https://doi.org/10.1073/pnas.97.3.1269>
- Bleul, C. C., Wu, L., Hoxie, J. A., Springer, T. A., & Mackay, C. R. (1997). The HIV coreceptors CXCR4 and CCR5 are differentially expressed and regulated on human T lymphocytes. *Proceedings of the National Academy of Sciences of the United States of America*, 94(5), 1925–1930. <https://doi.org/10.1073/pnas.94.5.1925>
- Boily, M. C., Baggaley, R. F., Wang, L., Masse, B., White, R. G., Hayes, R. J., & Alary, M. (2009). Heterosexual risk of HIV-1 infection per sexual act: systematic review and meta-analysis of observational studies. *The Lancet Infectious Diseases*, 9(2), 118–129. [https://doi.org/10.1016/S1473-3099\(09\)70021-0](https://doi.org/10.1016/S1473-3099(09)70021-0)
- Botto, L. D., & Khoury, M. J. (2001). Commentary: Facing the challenge of gene-environment interaction: The two-by-four table and beyond. *American Journal of Epidemiology*, 153(10), 1016–1020. <https://doi.org/10.1038/sj.aje.6701849>
- Brien, T. R. O., Winkler, C., Dean, M., Nelson, J. A. E., Carrington, M., Michael, N. L., & Ii, G. C. W. (1997). HIV-1 infection in a man homozygous for CCR5 Δ 32. *The Lancet*, 349, 1219.
- Bucy, R. P., Hockett, R. D., Derdeyn, C. A., Saag, M. S., Squires, K., Sillers, M., ... Kilby, J. M. (1999). Initial increase in blood CD4(+) lymphocytes after HIV antiretroviral therapy reflects redistribution from lymphoid tissues. *The Journal of Clinical Investigation*, 103(10), 1391–1398. <https://doi.org/10.1172/JCI5863>
- Burns, J. C., Shimizu, C., Gonzalez, E., Kulkarni, H., Patel, S., Shike, H., ... Ahuja,

- S. K. (2005). Genetic variations in the receptor-ligand pair CCR5 and CCL3L1 are important determinants of susceptibility to Kawasaki disease. *The Journal of Infectious Diseases*, 192(2), 344–349. <https://doi.org/10.1086/430953>
- Cahill, S., & Valadéz, R. (2013). Growing older with HIV/AIDS: New public health challenges. *American Journal of Public Health*, 103(3), e7–e15. <https://doi.org/10.2105/AJPH.2012.301161>
- Calmy, A., Ford, N., Hirschel, B., Reynolds, S. J., Lynen, L., Goemaere, E., ... Rodriguez, W. (2007). HIV Viral Load Monitoring in Resource-Limited Regions: Optional or Necessary? *Clinical Infectious Diseases*, 44(1), 128–134. <https://doi.org/10.1086/510073>
- Campbell-Yesufu, O. T., & Gandhi, R. T. (2011). Update on human immunodeficiency virus (HIV)-2 infection. *Clinical Infectious Diseases*, 52(6), 780–787. <https://doi.org/10.1093/cid/ciq248>
- Campbell, C. D., Sampas, N., Tsalenko, A., Sudmant, P. H., Kidd, J. M., Malig, M., ... Eichler, E. E. (2011). Population-genetic properties of differentiated human copy-number polymorphisms. *American Journal of Human Genetics*, 88(3), 317–332. <https://doi.org/10.1016/j.ajhg.2011.02.004>
- Cantsilieris, S., Baird, P. N., & White, S. J. (2013). Genomics Molecular methods for genotyping complex copy number polymorphisms. *Genomics*, 101(2), 86–93. <https://doi.org/10.1016/j.ygeno.2012.10.004>
- Cantsilieris, S., & White, S. J. (2013). Correlating Multiallelic Copy Number Polymorphisms with Disease Susceptibility. *Human Mutation*, 34(1), 1–13. <https://doi.org/10.1002/humu.22172>
- Cao, J., McNevin, J., Malhotra, U., & McElrath, M. J. (2003). Evolution of CD8+ T Cell Immunity and Viral Escape Following Acute HIV-1 Infection. *The Journal of Immunology*, 171(7), 3837–3846. <https://doi.org/10.4049/jimmunol.171.7.3837>
- Capoulade-Métay, C., Ma, L., Truong, L. X., Dudoit, Y., Versmisse, P., Nguyen, N. V., ... Theodorou, I. (2004). New CCR5 variants associated with reduced HIV coreceptor function in southeast Asia. *AIDS*, 18(17), 2243–2252. <https://doi.org/10.1097/00002030-200411190-00004>
- Carpenter, D., McIntosh, R. S., Pleass, R. J., & Armour, J. A. L. (2012). Functional effects of CCL3L1 copy number. *Genes and Immunity*, 13(5), 374–379. <https://doi.org/10.1038/gene.2012.5>
- Carpenter, D., Taype, C., Goulding, J., Levin, M., Eley, B., Anderson, S., ... Armour, J. A. (2014). CCL3L1 copy number, CCR5 genotype and susceptibility to tuberculosis. *BMC Med Genet*, 15(1), 5. <https://doi.org/10.1186/1471-2350-15-5>
- Carpenter, Danielle, Färnert, A., Rooth, I., Armour, J. A. L., & Shaw, M. A. (2012). CCL3L1 copy number and susceptibility to malaria. *Infection, Genetics and Evolution*, 12(5), 1147–1154. <https://doi.org/10.1016/j.meegid.2012.03.021>
- Carpenter, Danielle, Walker, S., Prescott, N., Schalkwijk, J., & Armour, J. Al. (2011). Accuracy and differential bias in copy number measurement of CCL3L1 in association studies with three auto-immune disorders. *BMC Genomics*, 12(1), 418. <https://doi.org/10.1186/1471-2164-12-418>
- Carrico, A. W. (2011). Substance use and HIV disease progression in the HAART era: Implications for the primary prevention of HIV. *Life Sciences*, 88(21–22), 940–947. <https://doi.org/10.1016/J.LFS.2010.10.002>
- Carrington, M., Dean, M., Martin, M. P., & O'Brien, S. J. (1999). Genetics of HIV-1 infection: Chemokine receptor CCR5 polymorphism and its consequences. *Human Molecular Genetics*. <https://doi.org/10.1093/hmg/8.10.1939>

- Carrington, M., Kissner, T., Gerrard, B., Ivanov, S., O'Brien, S. J., & Dean, M. (1997). Novel Alleles of the Chemokine-Receptor Gene CCR5. *The American Journal of Human Genetics*, 61(6), 1261–1267. <https://doi.org/10.1016/j.geforum.2008.07.001>
- Carrington, M., & O'Brien, S. J. (2003). The Influence of HLA Genotype on AIDS. *Annual Review of Medicine*, 54(1), 535–551. <https://doi.org/10.1146/annurev.med.54.101601.152346>
- Carter, N. P. (2007). Methods and strategies for analyzing copy number variation using DNA microarrays. *Nat Genet*, 39(JULY), 1–11. <https://doi.org/10.1038/ng2028.Methods>
- Castellino, F., Huang, A. Y., Altan-Bonnet, G., Stoll, S., Scheinecker, C., & Germain, R. N. (2006). Chemokines enhance immunity by guiding naive CD8+T cells to sites of CD4+T cell-dendritic cell interaction. *Nature*, 440(7086), 890–895. <https://doi.org/10.1038/nature04651>
- Catano, G., Chykarenko, Z. A., Mangano, A., Anaya, J. M., He, W., Smith, A., ... Ahuja, S. K. (2011). Concordance of CCR5 genotypes that influence cell-mediated immunity and HIV-1 disease progression rates. *Journal of Infectious Diseases*, 203(2), 263–272. <https://doi.org/10.1093/infdis/jiq023>
- CDC. (2008). Revised Surveillance Case Definitions for HIV Infection Among Adults, Adolescents, and Children Aged <18 Months and for HIV Infection and AIDS Among Children Aged 18 Months to <13 Years — United States, 2008. *Morbidity and Mortality Weekly Report*, 57 (RR-10), 1–12.
- Centers for Disease Control and Prevention. (2014). *Revised surveillance case definition for HIV infection—United States, 2014. Morbidity and Mortality Weekly Report*.
- Choe, H., Farzan, M., Sun, Y., Sullivan, N., Rollins, B., Ponath, P. D., ... Sodroski, J. (1996). The β-chemokine receptors CCR3 and CCR5 facilitate infection by primary HIV-1 isolates. *Cell*. [https://doi.org/10.1016/S0092-8674\(00\)81313-6](https://doi.org/10.1016/S0092-8674(00)81313-6)
- Chow, W. Z., Lim, S. H., Ong, L. Y., Yong, Y. K., Takebe, Y., Kamarulzaman, A., & Tee, K. K. (2015). Impact of HIV-1 subtype on the time to CD4+ T-cell recovery in combination antiretroviral therapy (cART)-experienced patients. *PLoS ONE*, 10(9). <https://doi.org/10.1371/journal.pone.0137281>
- Clayton, D. G., Walker, N. M., Smyth, D. J., Pask, R., Cooper, J. D., Maier, L. M., ... Todd, J. A. (2005). Population structure, differential bias and genomic control in a large-scale, case-control association study. *Nature Genetics*, 37(11), 1243–1246. <https://doi.org/10.1038/ng1653>
- Cock, K. M., Adjourolo, G., Ekpini, E., Sibailly, T., Kouadio, J., Maran, M., ... Gayle, H. D. (1993). Epidemiology and Transmission of HIV-2: Why There Is No HIV-2 Pandemic. *JAMA: The Journal of the American Medical Association*, 270(17), 2083–2086. <https://doi.org/10.1001/jama.1993.03510170073033>
- Cofrancesco, J., Scherzer, R., Tien, P. C., Gibert, C. L., Southwell, H., Sidney, S., ... Grunfeld, C. (2008). Illicit drug use and HIV treatment outcomes in a US cohort. *AIDS*, 22(3), 357–365. <https://doi.org/10.1097/QAD.0b013e3282f3cc21>
- Cohen, M. S., Gay, C. L., Busch, M. P., & Hecht, F. M. (2010). The Detection of Acute HIV Infection. *The Journal of Infectious Diseases*, 202(S2), S270–S277. <https://doi.org/10.1086/655651>
- Colobran, R., Comas, D., Faner, R., Pedrosa, E., Anglada, R., Pujol-Borrell, R., ... Juan, M. (2008). Population structure in copy number variation and SNPs in the CCL4L chemokine gene. *Genes and Immunity*, 9(4), 279–288. <https://doi.org/10.1038/gene.2008.15>

- Combadiere, C., Ahuja, S. K., Tiffany, H. L., & Murphy, P. M. (1996). Cloning and functional expression of CC CKR5, a human monocyte CC chemokine receptor selective for MIP-1 $\alpha$ , MIP-1 $\beta$ , and RANTES. *Journal of Leukocyte Biology*, 60(1), 147–152. <https://doi.org/10.1002/jlb.60.1.147>
- Conrad, D. F., Pinto, D., Redon, R., Feuk, L., Gokcumen, O., Zhang, Y., ... Hurles, M. E. (2010). Origins and functional impact of copy number variation in the human genome. *Nature*, 464(7289), 704–712. <https://doi.org/10.1038/nature08516.Origins>
- Contento, R. L., Molon, B., Boularan, C., Pozzan, T., Manes, S., Marullo, S., & Viola, A. (2008). CXCR4-CCR5: a couple modulating T cell functions. *Proceedings of the National Academy of Sciences of the United States of America*, 105(29), 10101–10106. <https://doi.org/10.1073/pnas.0804286105>
- Conway, B. (2007). The role of adherence to antiretroviral therapy in the management of HIV infection. *Journal of Acquired Immune Deficiency Syndromes*, 45(SUPPL. 1), 14–18. <https://doi.org/10.1097/QAI.0b013e3180600766>
- Corbeau, P., & Reynes, J. (2011). Immune reconstitution under antiretroviral therapy: The new challenge in HIV-1 infection. *Blood*. <https://doi.org/10.1182/blood-2010-12-322453>
- Costin, J. M. (2007). Cytopathic mechanisms of HIV-1. *Virology Journal*, 4, 100. <https://doi.org/10.1186/1743-422X-4-100>
- Crepaz, N., Marks, G., Liau, A., Mullins, M. M., Aupont, L. W., Marshall, K. J., ... Wolitski, R. J. (2009). Prevalence of unprotected anal intercourse among HIV-diagnosed MSM in the United States: A meta-analysis. *AIDS*. <https://doi.org/10.1097/QAD.0b013e32832effae>
- Dahl, V., Josefsson, L., & Palmer, S. (2010). HIV reservoirs, latency, and reactivation: Prospects for eradication. *Antiviral Research*, 85(1), 286–294. <https://doi.org/10.1016/j.antiviral.2009.09.016>
- Dalgleish, A. G., Beverley, P. C. L., Clapham, P. R., Crawford, D. H., Greaves, M. F., & Weiss, R. A. (1984). The CD4 (T4) antigen is an essential component of the receptor for the AIDS retrovirus. *Nature*, 312(5996), 763–767. <https://doi.org/10.1038/312763a0>
- De Silva, E., & Stumpf, M. P. H. (2004). HIV and the CCR5-Δ32 resistance allele. *FEMS Microbiology Letters*, 241(1), 1–12. <https://doi.org/10.1016/j.femsle.2004.09.040>
- de Smith, A. J., Tselenko, A., Sampas, N., Scheffer, A., Yamada, N. A., Tsang, P., ... Blakemore, A. I. F. (2007). Array CGH analysis of copy number variation identifies 1284 new genes variant in healthy white males: implications for association studies of complex diseases. *Human Molecular Genetics*, 16(23), 2783–2794. <https://doi.org/10.1093/hmg/ddm208>
- Dean, H. D., & Fenton, K. A. (2010). Addressing Social Determinants of Health in the Prevention and Control of HIV/AIDS, Viral Hepatitis, Sexually Transmitted Infections, and Tuberculosis. *Public Health Reports*, 125(4\_suppl), 1–5. <https://doi.org/10.1177/00333549101250s401>
- Dean, M., Carrington, M., Winkler, C., Huttley, G. a, Smith, M. W., Allikmets, R., ... O'Brien, S. J. (1996). Genetic restriction of HIV-1 infection and progression to AIDS by a deletion allele of the CKR5 structural gene. *Science*, 273(5283), 1856–1862. <https://doi.org/10.1126/science.273.5283.1856>
- Deng, H. K., Liu, R., Ellmeier, W., Choe, S., Unutmaz, D., Burkhardt, M., ... Landau, N. R. (1996). Identification of a major co-receptor for primary isolates of HIV-1. *Nature*, 381(6584), 661–666. <https://doi.org/10.1038/381661a0>

- Department of Statistics Malaysia. (2016). *Current Population Estimates, Malaysia, 2014 - 2016*.
- Dieye, T. N., Vereecken, C., Diallo, A. A., Ondo, P., Diaw, P. A., Camara, M., ... Kestens, L. (2005). Absolute CD4 T-cell counting in resource-poor settings: Direct volumetric measurements versus bead-based clinical flow cytometry instruments. *Journal of Acquired Immune Deficiency Syndromes*, 39(1), 32–37. <https://doi.org/10.1097/01.qai.0000160515.20581.ad>
- Ding, J., Zhao, J., Zhou, J., Li, X., Wu, Y., Ge, M., & Cen, S. (2018). Association of gene polymorphism of SDF1 (CXCR12) with susceptibility to HIV-1 infection and AIDS disease progression: A meta-analysis. *PLoS ONE*, 13(2), 1–13. <https://doi.org/10.1371/journal.pone.0191930>
- Dolan, M. J., Kulkarni, H., Camargo, J. F., He, W., Smith, A., Anaya, J.-M., ... Ahuja, S. K. S. S. (2007). CCL3L1 and CCR5 influence cell-mediated immunity and affect HIV-AIDS pathogenesis via viral entry-independent mechanisms. *Nature Immunology*, 8(12), 1324–1336. <https://doi.org/10.1038/ni1521>
- Dosekun, O., & Fox, J. (2010). An overview of the relative risks of different sexual behaviours on HIV transmission. *Current Opinion in HIV and AIDS*. <https://doi.org/10.1097/COH.0b013e32833a88a3>
- Douek, D. C., Picker, L. J., & Koup, R. A. (2003). T Cell Dynamics In HIV-1 Infection. *Annual Review of Immunology*, 21(1), 265–304. <https://doi.org/10.1146/annurev.immunol.21.120601.141053>
- Dragic, T., Trkola, A., Thompson, D. A. D., Cormier, E. G., Kajumo, F. A., Maxwell, E., ... Moore, J. P. (2000). A binding pocket for a small molecule inhibitor of HIV-1 entry within the transmembrane helices of CCR5. *Proceedings of the National Academy of Sciences*, 97(10), 5639–5644. <https://doi.org/10.1073/pnas.090576697>
- Dragic, Tatjana, Litwin, V., Allaway, G. P., Martin, S. R., Huang, Y., Nagashima, K. A., ... Paxton, W. A. (1996). HIV-1 entry into CD4+ cells is mediated by the chemokine receptor CC-CKR-5. *Nature*, 381(6584), 667–673. <https://doi.org/10.1038/381667a0>
- Egger, M., May, M., Chêne, G., Phillips, A. N., Ledergerber, B., Dabis, F., ... Sterne, J. A. C. (2002). Prognosis of HIV-1-infected patients starting highly active antiretroviral therapy: A collaborative analysis of prospective studies. *Lancet*, 360(9327), 119–129. [https://doi.org/10.1016/S0140-6736\(02\)09411-4](https://doi.org/10.1016/S0140-6736(02)09411-4)
- Erb, P., Battegay, M., Zimmerli, W., Rickenbach, M., & Egger, M. (2000). Effect of antiretroviral therapy on viral load, CD4 cell count, and progression to acquired immunodeficiency syndrome in a community human immunodeficiency virus-infected cohort. Swiss HIV Cohort Study. *Archives of Internal Medicine*, 160(8), 1134–1140. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/10789606>
- Espeseth, a S., Felock, P., Wolfe, a, Witmer, M., Grobler, J., Anthony, N., ... Hazuda, D. J. (2000). HIV-1 integrase inhibitors that compete with the target DNA substrate define a unique strand transfer conformation for integrase. *Proceedings of the National Academy of Sciences of the United States of America*, 97, 11244–11249. <https://doi.org/10.1073/pnas.200139397>
- Esté, J. A., & Cihlar, T. (2010). Current status and challenges of antiretroviral research and therapy. *Antiviral Research*, 85(1), 25–33. <https://doi.org/10.1016/j.antiviral.2009.10.007>
- Fellay, J. (2009). Host genome influences on HIV-1 disease. *Antiviral Therapy*, 14(6), 731–738. <https://doi.org/10.3851/IMP1253.Host>
- Fellermann, K., Stange, D. E., Schaeffeler, E., Schmalzl, H., Wehkamp, J., Bevins, C.

- L., ... Stange, E. F. (2006). A Chromosome 8 Gene-Cluster Polymorphism with Low Human Beta-Defensin 2 Gene Copy Number Predisposes to Crohn Disease of the Colon. *The American Journal of Human Genetics*, 79(3), 439–448. <https://doi.org/10.1086/505915>
- Feng, Y., Broder, C. C., Kennedy, P. E., & Berger, E. A. (1996). HIV-1 entry cofactor: functional cDNA cloning of a seven-transmembrane, G protein-coupled receptor. *Science*, 272, 872–877. <https://doi.org/10.1126/science.272.5263.872>. www.jimmunol.org
- Fernandez-Jimenez, N., Castellanos-Rubio, A., Plaza-Izurieta, L., Gutierrez, G., Irastorza, I., Castaño, L., ... Bilbao, J. R. (2011). Accuracy in copy number calling by qPCR and PRT: A matter of DNA. *PLoS ONE*, 6(12), e28910. <https://doi.org/10.1371/journal.pone.0028910>
- Feuk, L., Carson, A., & Scherer, S. (2006). Structural variation in the human genome. *Nature Reviews Genetics*, 7(2), 85–97. <https://doi.org/10.1038/nrg1767>
- Fideli, Ü. S., Allen, S. A., Musonda, R., Trask, S., Hahn, B. H., Weiss, H., ... Aldrovandi, G. M. (2001). Virologic and Immunologic Determinants of Heterosexual Transmission of Human Immunodeficiency Virus Type 1 in Africa. *AIDS Research and Human Retroviruses*, 17(10), 901–910. <https://doi.org/10.1089/088922201750290023>
- Field, S. F., Howson, J. M. M., Maier, L. M., Walker, S., M. N., Smyth, D. J., ... Todd, J. A. (2009). Experimental aspects of copy number variant assays at CCL3L1. *Nature Medicine*, 15(10), 1115–1117. <https://doi.org/10.1038/nm.1981>
- Fleming, D. T., & Wasserheit, J. N. (1999). From epidemiological synergy to public health policy and practice : the contribution of other sexually transmitted diseases to sexual transmission of HIV infection From epidemiological synergy to public health policy and practice : the contribution of oth. *Sexually Transmitted Infections*, 75, 3–17. <https://doi.org/10.1136/sti.75.1.3>
- Frade, J. M. R., Llorente, M., Mellado, M., Alcamí, J., Gutiérrez-Ramos, J. C., Zaballos, A., ... Martínez-A., C. (1997). The amino-terminal domain of the CCR2 chemokine receptor acts as coreceptor for HIV-1 infection. *Journal of Clinical Investigation*, 100(3), 497–502. <https://doi.org/10.1172/JCI119558>
- Galvani, A. P., & Novembre, J. (2005). The evolutionary history of the CCR5-Δ32 HIV-resistance mutation. *Microbes and Infection*. <https://doi.org/10.1016/j.micinf.2004.12.006>
- Gao, F., Bailes, E., Robertson, D., & Al., E. (1999). HIV-1 in the chimpanzee Pan troglodytes troglodytes. *Nature*, 397, 436–441.
- Gardner, E. M., Burman, W. J., Steiner, J. F., Anderson, P. L., & Bangsberg, D. R. (2009). Antiretroviral medication adherence and the development of class-specific antiretroviral resistance. *AIDS*, 23(9), 1035–1046. <https://doi.org/10.1097/QAD.0b013e32832ba8ec>
- Geijtenbeek, T. B. H., & van Kooyk, Y. (2003). DC-SIGN: a novel HIV receptor on DCs that mediates HIV-1 transmission. *Current Topics in Microbiology and Immunology*, 276, 31–54. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/12797442>
- Geskus, R. B., Miedema, F. A., Goudsmit, J., Reiss, P., Schuitemaker, H., & Coutinho, R. A. (2003). Prediction of residual time to AIDS and death based on markers and cofactors. *Journal of Acquired Immune Deficiency Syndromes*, 32(5), 514–521. <https://doi.org/10.1097/00126334-200304150-00008>
- Girirajan, S., Campbell, C. D., & Eichler, E. E. (2011). Human Copy Number Variation and Complex Genetic Disease. *Annual Review of Genetics*, 45(1), 203–

226. <https://doi.org/10.1146/annurev-genet-102209-163544>
- Goff, S. P. (2004). Introduction to Retroviruses. In G. P. Wormser (Ed.), *AIDS and Other Manifestations of HIV Infection* (4th ed., pp. 57–93). Elsevier (USA). <https://doi.org/10.1186/1743-422X-4-100>
- Goh, K. L., Chua, C. T., Chiew, I. S., & Soo-Hoo, T. S. (1987). The acquired immune deficiency syndrome: a report of the first case in Malaysia. *The Medical Journal of Malaysia*, 42(1), 58–60. Retrieved from <http://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.433.975&rep=rep1&type=pdf>
- Gonzalez, E., Bamshad, M., Sato, N., Mummidi, S., Dhanda, R., Catano, G., ... Ahuja, S. K. (1999). Race-specific HIV-1 disease-modifying effects associated with CCR5 haplotypes. *Proceedings of the National Academy of Sciences of the United States of America*, 96(21), 12004–12009. <https://doi.org/10.1073/pnas.96.21.12004>
- Gonzalez, E., Kulkarni, H., Bolivar, H., Mangano, A., Sanchez, R., Catano, G., ... Ahuja, S. K. (2005). The influence of CCL3L1 gene-containing segmental duplications on HIV-1/AIDS susceptibility. *Science*, 307(5714), 1434–1440. <https://doi.org/10.1126/science.1101160>
- Goonetilleke, N., Liu, M. K. P., Salazar-Gonzalez, J. F., Ferrari, G., Giorgi, E., Ganusov, V. V., ... McMichael, A. J. (2009). The first T cell response to transmitted/founder virus contributes to the control of acute viremia in HIV-1 infection. *The Journal of Experimental Medicine*, 206(6), 1253–1272. <https://doi.org/10.1084/jem.20090365>
- Gornalusse, G. G., Mummidi, S., Gaitan, A. A., Jimenez, F., Ramsuran, V., Picton, A., ... Ahuja, S. K. (2015). Epigenetic mechanisms, T-cell activation, and CCR5 genetics interact to regulate T-cell expression of CCR5, the major HIV-1 coreceptor. *Proceedings of the National Academy of Sciences of the United States of America*, 112(34), E4762–71. <https://doi.org/10.1073/pnas.1423228112>
- Goto, T., Nakai, M., & Ikuta, K. (1998). The Life-cycle of Human Immunodeficiency Virus Type 1. *Science*, 29(2), 123–138.
- Greub, G., Ledergerber, B., Battegay, M., Grob, P., Perrin, L., Furrer, H., ... Telenti, A. (2000). Clinical progression, survival, and immune recovery during antiretroviral therapy in patients with HIV-1 and hepatitis C virus coinfection: the Swiss HIV Cohort Study. *G. Lancet*, 356, 1800–1805.
- Griffith, J. W., Sokol, C. L., & Luster, A. D. (2014). Chemokines and Chemokine Receptors: Positioning Cells for Host Defense and Immunity. *Annual Review of Immunology*, 32(1), 659–702. <https://doi.org/10.1146/annurev-immunol-032713-120145>
- Grünhage, F., Nattermann, J., Gressner, O. A., Wasmuth, H. E., Hellerbrand, C., Sauerbruch, T., ... Lammert, F. (2010). Lower copy numbers of the chemokine CCL3L1 gene in patients with chronic hepatitis C. *Journal of Hepatology*, 52(2), 153–159. <https://doi.org/10.1016/j.jhep.2009.11.001>
- Gu, J., & Lieber, M. R. (2008). Mechanistic flexibility as a conserved theme across 3 billion years of nonhomologous DNA end-joining. *Genes and Development*. <https://doi.org/10.1101/gad.1646608>
- Gupta, R. K., Gregson, J., Parkin, N., Haile-Selassie, H., Tanuri, A., Andrade Forero, L., ... Bertagnolio, S. (2017). HIV-1 drug resistance before initiation or re-initiation of first-line antiretroviral therapy in low-income and middle-income countries: A systematic review and meta-regression analysis. *The Lancet Infectious Diseases*, 18(3), 346–355. [https://doi.org/10.1016/S1473-3099\(16\)30511-7](https://doi.org/10.1016/S1473-3099(16)30511-7)

- Haas, D. W., Geraghty, D. E., Andersen, J., Mar, J., Motsinger, A. A., D'Aquila, R. T., ... Landay, A. (2006). Immunogenetics of CD4 Lymphocyte Count Recovery during Antiretroviral Therapy: An AIDS Clinical Trials Group Study. *The Journal of Infectious Diseases*, 194(8), 1098–1107. <https://doi.org/10.1086/507313>
- Hajizadeh, M., Onadja, Y., Brewer, T. F., Sia, D., Nandi, A., & Heymann, S. J. (2016). What explains gender inequalities in HIV/AIDS prevalence in sub-Saharan Africa? Evidence from the demographic and health surveys. *BMC Public Health*, 16(1), 1–18. <https://doi.org/10.1186/s12889-016-3783-5>
- Hammer, S. M., Eron, J. J., Reiss, P., Schooley, R. T., Thompson, M. A., Walmsley, S., ... International AIDS Society-USA. (2008). Antiretroviral Treatment of Adult HIV Infection. *JAMA*, 300(5), 555. <https://doi.org/10.1001/jama.300.5.555>
- Hardwick, R. J., Amogne, W., Mugusi, S., Yimer, G., Ngaimisi, E., Habtewold, A., ... Aklillu, E. (2012).  $\beta$ -defensin genomic copy number is associated with HIV load and immune reconstitution in sub-Saharan Africans. *Journal of Infectious Diseases*, 206(7), 1012–1019. <https://doi.org/10.1093/infdis/jis448>
- Haridan, U. S., Mokhtar, U., Machado, L. R., Aziz, A. T. A., Shueb, R. H., Zaid, M., ... Peng, H. B. (2015). A comparison of assays for accurate copy number measurement of the low-affinity FC gamma receptor genes FCGR3A and FCGR3B. *PLoS ONE*, 10(1), 1–13. <https://doi.org/10.1371/journal.pone.0116791>
- Havlir, D. V., Bassett, R., Levitan, D., Gilbert, P., Tebas, P., Collier, A. C., ... Wong, J. K. (2001). Prevalence and predictive value of intermittent viremia with combination HIV therapy. *Journal of the American Medical Association*, 286(2), 171–179. <https://doi.org/10.1001/jama.286.2.171>
- He, L., Pan, X., Dou, Z., Huang, P., Zhou, X., Peng, Z., ... Wang, N. (2016). The factors related to CD4+ T-cell recovery and viral suppression in patients who have low CD4+ T cell counts at the initiation of HAART: A retrospective study of the national HIV treatment sub-database of Zhejiang Province, China, 2014. *PLoS ONE*, 11(2), 1–14. <https://doi.org/10.1371/journal.pone.0148915>
- Hemelaar, J. (2012). The origin and diversity of the HIV-1 pandemic. *Trends in Molecular Medicine*, 18(3), 182–192. <https://doi.org/10.1016/j.molmed.2011.12.001>
- Hendrickson, S. L., Jacobson, L. P., Nelson, G. W., Phair, J. P., Lautenberger, J., Johnson, R. C., ... O'Brien, S. J. (2008). Host genetic influences on highly active antiretroviral therapy efficacy and AIDS-free survival. *Journal of Acquired Immune Deficiency Syndromes*, 48(3), 263–271. <https://doi.org/10.1097/QAI.0b013e31816fdc5f>
- Hendrix, C. W., Collier, A. C., Lederman, M. M., Schols, D., Pollard, R. B., Brown, S., ... Calandra, G. (2004). Safety, Pharmacokinetics, and Antiviral Activity of AMD3100, a Selective CXCR4 Receptor Inhibitor, in HIV-1 Infection. *J Acquir Immune Defic Syndr* • (Vol. 37). Retrieved from <https://insights.ovid.com/pubmed?pmid=15385732>
- HIV/STI Sector, Division of Disease Control, M. of H. M. (2016). *Global AIDS Response Progress Report 2016*.
- HIV/STI Sector Disease Control Division Ministry of Health Malaysia. (2014). *Integrated Biological and Behavioural Surveillance Survey (IBBSS)*. Ministry of Health Malaysia. <https://doi.org/10.2989/16085906.2013.863214>
- Hoffmann, C. (2007). The epidemiology of HIV coreceptor tropism. *European*

- Journal of Medical Research*, 12(9), 385–390. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/17933718>
- Hollingsworth, T. D., Anderson, R. M., & Fraser, C. (2008). HIV-1 transmission, by stage of infection. *The Journal of Infectious Diseases*, 198(5), 687–693. <https://doi.org/10.1086/590501>
- Hollox, E. J., Armour, J. A. L., & Barber, J. C. K. (2003). Extensive Normal Copy Number Variation of a  $\beta$ -Defensin Antimicrobial-Gene Cluster. *The American Journal of Human Genetics*, 73(3), 591–600. <https://doi.org/10.1086/378157>
- Hser, Y.-I., Liang, D., Lan, Y.-C., Vicknasingam, B. K., & Chakrabarti, A. (2016). Drug Abuse, HIV, and HCV in Asian Countries. *Journal of Neuroimmune Pharmacology*, 11(3), 383–393. <https://doi.org/10.1007/s11481-016-9665-x>
- Hsue, P. Y., Scherzer, R., Hunt, P. W., Schnell, A., Bolger, A. F., Kalapus, S. C., ... Deeks, S. G. (2012). Carotid Intima-Media Thickness Progression in HIV-Infected Adults Occurs Preferentially at the Carotid Bifurcation and Is Predicted by Inflammation. *Journal of the American Heart Association*, 1(2), jah3-e000422-jah3-e000422. <https://doi.org/10.1161/JAHA.111.000422>
- Hu, D. J., Dondero, T. J., Rayfield, M. A., George, J. R., Schochetman, G., Jaffe, H. W., ... Curran, J. W. (1996). The emerging genetic diversity of HIV. The importance of global surveillance for diagnostics, research, and prevention. *JAMA*, 275(3), 210–216. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/8604174>
- Huang, M., & Hussein, H. (2004). The HIV/AIDS epidemic country paper: Malaysia. *AIDS Education and Prevention*. <https://doi.org/10.1521/aeap.16.3.5.100.35532>
- Hudgens, M. G., Longini, I. M., Vanichseni, S., Hu, D. J., Kitayaporn, D., Mock, P. A., ... Mastro, T. D. (2002). Subtype-specific transmission probabilities for human immunodeficiency virus type 1 among injecting drug users in Bangkok, Thailand. *American Journal of Epidemiology*, 155(2), 159–168. <https://doi.org/10.1093/aje/155.2.159>
- Hughes, C. E., & Nibbs, R. J. B. (2018). A guide to chemokines and their receptors. *The FEBS Journal*, 285(16), 2944–2971. <https://doi.org/10.1111/febs.14466>
- Hunt, P. W., Brenchley, J., Sinclair, E., McCune, J. M., Roland, M., Page-Shafer, K., ... Deeks, S. G. (2007). Immune activation set point during early HIV infection predicts subsequent CD4 $\leq$  T-cell changes independent of viral load. *The Journal of Infectious Diseases*, 197(1), 126–133. <https://doi.org/10.1086/524143>
- Hütter, G., Nowak, D., Mossner, M., Ganepola, S., Müßig, A., Allers, K., ... Thiel, E. (2009). Long-Term Control of HIV by CCR5 Delta32/Delta32 Stem-Cell Transplantation. *New England Journal of Medicine*, 360(7), 692–698. <https://doi.org/10.1056/NEJMoa0802905>
- Ioannidis, J. P.A., Rosenberg, P. S., Goedert, J. J., Ashton, L. J., Benfield, T. L., Buchbinder, S. P., ... O'Brien, T. R. (2001). Effects of CCR5- $\Delta$ 32, CCR2-64I, and SDF-1 3' a alleles on HIV-1 disease progression: An international meta-analysis of individual-patient data. *Annals of Internal Medicine*, 135(9), 782–795. <https://doi.org/10.7326/0003-4819-135-9-200111060-00008>
- Ioannidis, John P.A. (2003, April 1). Genetic associations: False or true? *Trends in Molecular Medicine*. Elsevier. [https://doi.org/10.1016/S1471-4914\(03\)00030-3](https://doi.org/10.1016/S1471-4914(03)00030-3)
- Ionita-Laza, I., Rogers, A. J., Lange, C., Raby, B. A., & Lee, C. (2009). Genetic association analysis of copy-number variation (CNV) in human disease pathogenesis. *Genomics*, 93(1), 22–26. <https://doi.org/10.1016/j.ygeno.2008.08.012>
- Jarlais, D. C. D., Cooper, H. L. F., Bramson, H., Deren, S., Hatzakis, A., & Hagan, H.

- (2012). Racial and ethnic disparities and implications for the prevention of HIV among persons who inject drugs. *Current Opinion in HIV and AIDS*, 7(4), 354–361. <https://doi.org/10.1097/COH.0b013e328353d990>
- Javor, J., Párnická, Z., Michalik, J., Čopíková-Cudráková, D., Shawkatová, I., Šurmanová, V., ... Buc, M. (2015). The +190 G/A (rs1799864) polymorphism in the C-C chemokine receptor 2 (CCR2) gene is associated with susceptibility to multiple sclerosis in HLA-DRB1\*15:01-negative individuals. *Journal of the Neurological Sciences*, 349(1–2), 138–142. <https://doi.org/10.1016/j.jns.2015.01.002>
- Jiang, Y., Chen, O., Cui, C., Zhao, B., Han, X., Zhang, Z., ... Shang, H. (2013). KIR3DS1/L1 and HLA-Bw4-80I are associated with HIV disease progression among HIV typical progressors and long-term nonprogressors. *BMC Infectious Diseases*, 13(1), 405. <https://doi.org/10.1186/1471-2334-13-405>
- Jurriaans, S., Van Gemen, B., Weverling, G. J., Van Strijp, D., Nara, P., Coutinho, R., ... Goudsmit, J. (1994). The natural history of HIV-1 infection: Virus load and virus phenotype independent determinants of clinical course? *Virology*, 204(1), 223–233. <https://doi.org/10.1006/viro.1994.1526>
- Kahn, J. O., & Walker, B. D. (1998). Acute Human Immunodeficiency Virus Type 1 Infection. *New England Journal of Medicine*, 339(1), 33–39. <https://doi.org/10.1056/NEJM199807023390107>
- Kaslow, R. A., Dorak, T., & Tang, J. J. (2005). Influence of host genetic variation on susceptibility to HIV type 1 infection. *J Infect Dis*, 191 Suppl(Suppl 1), S68–77. <https://doi.org/10.1086/425269>
- Kaslow, R. A., Tang, J. J., & Dorak, M. T. (2004). The Role of Host Genetic Variation in HIV Infection and its Manifestations. <https://doi.org/10.1016/B978-012764051-8/50014-7>
- Kassutto, S., & Rosenberg, E. S. (2004). Primary HIV Type 1 Infection. *Clinical Infectious Diseases*, 38, 1447–1453. <https://doi.org/10.1086/420745>
- Kaur, G., & Mehra, N. (2009). Genetic determinants of HIV-1 infection and progression to AIDS: Susceptibility to HIV infection. *Tissue Antigens*. <https://doi.org/10.1111/j.1399-0039.2009.01220.x>
- Kelley, C. F., Kitchen, C. M. R., Hunt, P. W., Rodriguez, B., Hecht, F. M., Kitahata, M., ... Deeks, S. G. (2009). Incomplete Peripheral CD4 + Cell Count Restoration in HIV-Infected Patients Receiving Long-Term Antiretroviral Treatment. *Clinical Infectious Diseases*, 48(6), 787–794. <https://doi.org/10.1086/597093>
- Kharsany, A. B. M., & Karim, Q. A. (2016). HIV Infection and AIDS in Sub-Saharan Africa: Current Status, Challenges and Opportunities. *The Open AIDS Journal*, 10(1), 34–48. <https://doi.org/10.2174/1874613601610010034>
- Kilmarx, P. H. (2009). Global epidemiology of HIV. *Current Opinion in HIV and AIDS*, 4, 240–246. <https://doi.org/10.1097/COH.0b013e32832c06db>
- Kim, H. E., Kim, J. J., Han, M. K., Lee, K. Y., Song, M. S., Lee, H. D., ... Lee, J. K. (2012). Variations in the number of CCL3L1 gene copies and Kawasaki disease in Korean children. *Pediatric Cardiology*, 33(8), 1259–1263. <https://doi.org/10.1007/s00246-012-0289-5>
- Kohlstaedt, L. A., Wang, J., Friedman, J. M., Rice, P. A., & Steitz, T. A. (1992). Crystal structure at 3.5 Å resolution of HIV-1 reverse transcriptase complexed with an inhibitor. *Science (New York, N.Y.)*, 256(5065), 1783–1790. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/1377403>
- Korenromp, E. L., Williams, B. G., Schmid, G. P., & Dye, C. (2009). Clinical prognostic value of RNA viral load and CD4 cell counts during untreated HIV-1

- infection - A quantitative review. *PLoS ONE*, 4(6). <https://doi.org/10.1371/journal.pone.0005950>
- Kostrikis, L. G., Huang, Y., Moore, J. P., Wolinsky, S. M., Zhang, L., Guo, Y., ... Ho, D. D. (1998). A chemokine receptor CCR2 allele delays HIV-1 disease progression and is associated with a CCR5 promoter mutation. *Nature Medicine*, 4, 350–353. <https://doi.org/10.1038/nm0398-350>
- Koup, R. A., Safrit, J. T., Cao, Y., Andrews, C. A., McLeod, G., Borkowsky, W., ... Ho, D. D. (1994). Temporal association of cellular immune responses with the initial control of viremia in primary human immunodeficiency virus type 1 syndrome. *Journal of Virology*, 68(7), 4650–4655. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/8207839>
- Kulkarni, H., Agan, B. K., Marconi, V. C., O'Connell, R. J., Camargo, J. F., He, W., ... Ahuja, S. K. (2008). CCL3L1-CCR5 genotype improves the assessment of AIDS risk in HIV-1-infected individuals. *PLoS ONE*, 3(9). <https://doi.org/10.1371/journal.pone.0003165>
- Kulkarni, H., Jason, F., Grandits, G., Crum-cianflone, N. F., Landrum, M. L., Hale, B., ... Marconi, V. C. (2011). Early Postseroconversion CD4 Cell Counts Independently Predict CD4 Cell Count Recovery in HIV-1 – Positive Subjects Receiving Antiretroviral Therapy. *Journal of Acquired Immune Deficiency Syndromes*, 57(5), 387–395.
- Lau, K. A., & Wong, J. J. L. (2013). Current trends of HIV recombination worldwide. *Infectious Disease Reports*, 5(1S), 4. <https://doi.org/10.4081/idr.2013.s1.e4>
- Lawn, S. D., Butera, S. T., & Folks, T. M. (2001). Contribution of Immune Activation to the Pathogenesis and Transmission of Human Immunodeficiency Virus Type 1 Infection. *Clinical Microbiology Reviews*, 14(4), 753–777. <https://doi.org/10.1128/CMR.14.4.753-777.2001>
- Lawn, Stephen D., & Meintjes, G. (2011). Pathogenesis and prevention of immune reconstitution disease during antiretroviral therapy. *Expert Review of Anti-Infective Therapy*. <https://doi.org/10.1586/eri.11.21>
- Lee, J. A., Carvalho, C. M. B., & Lupski, J. R. (2007). A DNA Replication Mechanism for Generating Nonrecurrent Rearrangements Associated with Genomic Disorders. *Cell*, 131(7), 1235–1247. <https://doi.org/10.1016/j.cell.2007.11.037>
- Levy, S., Sutton, G., Ng, P. C., Feuk, L., Halpern, A. L., Walenz, B. P., ... Venter, J. C. (2007). The diploid genome sequence of an individual human. *PLoS Biology*, 5(10), e254. <https://doi.org/10.1371/journal.pbio.0050254>
- Li, H., Xie, H. Y., Zhou, L., Feng, X. W., Wang, W. L., Liang, T. B., ... Zheng, S. Sen. (2012). Copy number variation in CCL3L1 gene is associated with susceptibility to acute rejection in patients after liver transplantation. *Clinical Transplantation*, 26(2), 314–321. <https://doi.org/10.1111/j.1399-0012.2011.01486.x>
- Lingappa, J. R., Hughes, J. P., Wang, R. S., Baeten, J. M., Celum, C., Gray, G. E., ... Wald, A. (2010). Estimating the impact of plasma HIV-1 rna reductions on heterosexual HIV-1 transmission risk. *PLoS ONE*, 5(9), 1–7. <https://doi.org/10.1371/journal.pone.0012598>
- Lingappa, J. R., Petrovski, S., Kahle, E., Fellay, J., Shianna, K., McElrath, M. J., ... Goldstein, D. (2011). Genomewide association study for determinants of HIV-1 acquisition and viral set point in HIV-1 serodiscordant couples with quantified virus exposure. *PLoS ONE*, 6(12), 6–13. <https://doi.org/10.1371/journal.pone.0028632>
- Liu, H., Nakayama, E. E., Theodorou, I., Nagai, Y., Likanonsakul, S., Wasi, C., ...

- Shioda, T. (2007). Polymorphisms in CCR5 chemokine receptor gene in Japan. *International Journal of Immunogenetics*, 34(5), 325–335. <https://doi.org/10.1111/j.1744-313X.2007.00694.x>
- Liu, R., Paxton, W. A., Choe, S., Ceradini, D., Martin, S. R., Horuk, R., ... Landau, N. R. (1996). Homozygous defect in HIV-1 coreceptor accounts for resistance of some multiply-exposed individuals to HIV-1 infection. *Cell*, 86(3), 367–377. [https://doi.org/10.1016/S0092-8674\(00\)80110-5](https://doi.org/10.1016/S0092-8674(00)80110-5)
- Liu, S. J., Yao, L., Ding, D. L., & Zhu, H. Z. (2010). CCL3L1 copy number variation and susceptibility to HIV-1 infection: A meta-analysis. *PLoS ONE*, 5(12), 1–7. <https://doi.org/10.1371/journal.pone.0015778>
- Liu, Y., Liu, F.-L. L., He, Y., Li, L., Li, S., Zheng, Y.-T. T., ... Gong, X. (2012). The genetic variation of CCR5, CXCR4 and SDF-1 in three Chinese ethnic populations. *Infection, Genetics and Evolution*, 12(5), 1072–1078. <https://doi.org/10.1016/j.meegid.2012.03.009>
- Lobritz, M. A., Ratcliff, A. N., & Arts, E. J. (2010). HIV-1 entry, inhibitors, and resistance. *Viruses*, 2(5), 1069–1105. <https://doi.org/10.3390/v2051069>
- Long, B. R., Ndhlovu, L. C., Oksenberg, J. R., Lanier, L. L., Hecht, F. M., Nixon, D. F., & Barbour, J. D. (2008). Conferral of Enhanced Natural Killer Cell Function by KIR3DS1 in Early Human Immunodeficiency Virus Type 1 Infection. *Journal of Virology*, 82(10), 4785–4792. <https://doi.org/10.1128/JVI.02449-07>
- Looker, K. J., Elmes, J. A. R., Gottlieb, S. L., Schiffer, J. T., Vickerman, P., Turner, K. M. E., & Boily, M.-C. (2017). Effect of HSV-2 infection on subsequent HIV acquisition: an updated systematic review and meta-analysis. *The Lancet Infectious Diseases*, 17(12), 1303–1316. [https://doi.org/10.1016/S1473-3099\(17\)30405-X](https://doi.org/10.1016/S1473-3099(17)30405-X)
- Lu, J., Sheng, A., Wang, Y., Zhang, L., Wu, J., Song, M., ... Wang, W. (2012). The Genetic Associations and Epistatic Effects of the CCR5 Promoter and CCR2 - V64I Polymorphisms on Susceptibility to HIV-1 Infection in a Northern Han Chinese Population. *Genetic Testing and Molecular Biomarkers*, 16(12), 1369–1375. <https://doi.org/10.1089/gtmb.2012.0235>
- Lupski, J. R. (1998). Genomic disorders: Structural features of the genome can lead to DNA rearrangements and human disease traits. *Trends in Genetics*. [https://doi.org/10.1016/S0168-9525\(98\)01555-8](https://doi.org/10.1016/S0168-9525(98)01555-8)
- Maartens, G., Celum, C., & Lewin, S. R. (2014). HIV infection: epidemiology, pathogenesis, treatment, and prevention. *The Lancet*, 384, 258–271. [https://doi.org/10.1016/S0140-6736\(14\)60164-1](https://doi.org/10.1016/S0140-6736(14)60164-1)
- MacDonald, K. S., Embree, J., Njenga, S., Nagelkerke, N. J. D., Ngatia, I., Mohammed, Z., ... Plummer, F. A. (1998). Mother-Child Class I HLA Concordance Increases Perinatal Human Immunodeficiency Virus Type 1 Transmission. *The Journal of Infectious Diseases*, 177(3), 551–556. <https://doi.org/10.1086/514243>
- Mackelprang, R. D., Carrington, M., Thomas, K. K., Hughes, J. P., Baeten, J. M., Wald, A., ... Lingappa, J. R. (2015). Host Genetic and Viral Determinants of HIV-1 RNA Set Point among HIV-1 Seroconverters from Sub-Saharan Africa. *Journal of Virology*, 89(4), 2104–2111. <https://doi.org/10.1128/JVI.01573-14>
- Magierowska, M., Theodorou, I., Debré, P., Sanson, F., Autran, B., Rivière, Y., ... Costagliola, D. (1999). Combined genotypes of CCR5, CCR2, SDF1, and HLA genes can predict the long-term nonprogressor status in human immunodeficiency virus-1-infected individuals. *Blood*, 93(3), 936–941.
- Malaysian AIDS Council. (2013). *ANNUAL REPORT 2013: Malaysian AIDS Council*

- & Malaysian AIDS Foundation. Malaysian AIDS Council. <https://doi.org/10.1017/S0001972000001765>
- Malaysian AIDS Council. (2016). *Snapshot of HIV & AIDS in Malaysia 2016*.
- Malhotra, D., & Sebat, J. (2012). CNVs: harbingers of a rare variant revolution in psychiatric genetics. *Cell*, 148(6), 1223–1241. <https://doi.org/10.1016/j.cell.2012.02.039>
- Malhotra, R., Hu, L., Song, W., Brill, I., Mulenga, J., Allen, S., ... Kaslow, R. A. (2011). Association of chemokine receptor gene (CCR2-CCR5) haplotypes with acquisition and control of HIV-1 infection in Zambians. *Retrovirology*, 8(1), 22. <https://doi.org/10.1186/1742-4690-8-22>
- Mamtani, M., Rovin, B., Brey, R., Camargo, J. F., Kulkarni, H., Herrera, M., ... Ahuja, S. K. (2008). CCL3L1 gene-containing segmental duplications and polymorphisms in CCR5 affect risk of systemic lupus erythaematosus. *Annals of the Rheumatic Diseases*, 67(8), 1076–1083. <https://doi.org/10.1136/ard.2007.078048>
- Mamtani, Manju, Mummidis, S., Ramsuran, V., Pham, M. H., Maldonado, R., Begum, K., ... Ahuja, S. K. (2011). Influence of Variations in CCL3L1 and CCR5 on Tuberculosis in a Northwestern Colombian Population. *Journal of Infectious Diseases*, 203(11), 1590–1594. <https://doi.org/10.1093/infdis/jir145>
- Mangano, A., Gonzalez, E., Dhanda, R., Catano, G., Bamshad, M., Bock, A., ... Ahuja, S. K. (2001). Concordance between the CC Chemokine Receptor 5 Genetic Determinants That Alter Risks of Transmission and Disease Progression in Children Exposed Perinatally to Human Immunodeficiency Virus. *The Journal of Infectious Diseases*, 183(11), 1574–1585. <https://doi.org/10.1086/320705>
- Marconi, V. C., Grandits, G., Okulicz, J. F., Wortmann, G., Ganesan, A., Crum-Cianflone, N., ... Kulkarni, H. (2011). Cumulative viral load and virologic decay patterns after antiretroviral therapy in HIV-infected subjects influence CD4 recovery and AIDS. *PLoS ONE*, 6(5), 1–9. <https://doi.org/10.1371/journal.pone.0017956>
- Marschner, I. C., Collier, A. C., Coombs, R. W., RT, D. A., DeGruttola, V., Fischl, M. A., ... Saag, M. S. (1998). Use of changes in plasma levels of human immunodeficiency virus type 1 RNA to assess the clinical benefit of antiretroviral therapy. *The Journal of Infectious Diseases*, 177(1), 40–47. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/9419168>
- Martin, M. P., Qi, Y., Gao, X., Yamada, E., Martin, J. N., Pereyra, F., ... Carrington, M. (2007). Innate partnership of HLA-B and KIR3DL1 subtypes against HIV-1. *Nature Genetics*, 39(6), 733–740. <https://doi.org/10.1038/ng2035>
- Martinson, J. J., Chapman, N. H., Rees, D. C., Liu, Y. T., & Clegg, J. B. (1997). Global distribution of the CCR5 gene 32-basepair deletion. *Nature Genetics*, 16(1), 100–103. <https://doi.org/10.1038/ng0597-100>
- Mat Shah, R., Bulgiba, A., Lee, C. K. C., Haniff, J., & Mohamad Ali, M. (2012). Highly Active Antiretroviral Therapy Reduces Mortality and Morbidity in Patients with AIDS in Sungai Buloh Hospital. *Journal of Experimental and Clinical Medicine*, 4(4), 239–244. <https://doi.org/10.1016/j.jecm.2012.06.011>
- Mayer, K. H., & Beyrer, C. (2007). HIV Epidemiology Update and Transmission Factors: Risks and Risk Contexts--16th International AIDS Conference Epidemiology Plenary. *Clinical Infectious Diseases*, 44(7), 981–987. <https://doi.org/10.1086/512371>
- McBride, J. A., & Striker, R. (2017). Imbalance in the game of T cells: What can the CD4/CD8 T-cell ratio tell us about HIV and health? *PLOS Pathogens*, 13(11),

- e1006624. <https://doi.org/10.1371/journal.ppat.1006624>
- McCarroll, S. A. (2008). Copy-number analysis goes more than skin deep. *Nature Genetics*, 40(1), 5–6. <https://doi.org/10.1038/ng0108-5>
- McCarroll, S. A., & Altshuler, D. M. (2007). Copy-number variation and association studies of human disease. *Nature Genetics*, 39(7s), S37–S42. <https://doi.org/10.1038/ng2080>
- McCarroll, S. A., Kuruvilla, F. G., Korn, J. M., Cawley, S., Nemesh, J., Wysoker, A., ... Altshuler, D. (2008). Integrated detection and population-genetic analysis of SNPs and copy number variation. *Nature Genetics*, 40(10), 1166–1174. <https://doi.org/10.1038/ng.238>
- McColl, D. J., & Chen, X. (2010). Strand transfer inhibitors of HIV-1 integrase: Bringing IN a new era of antiretroviral therapy. *Antiviral Research*, 85(1), 101–118. <https://doi.org/10.1016/j.antiviral.2009.11.004>
- McDermott, D. H., Zimmerman, P. A., Guignard, F., Kleeberger, C. A., Leitman, S. F., & Murphy, P. M. (1998). CCR5 promoter polymorphism and HIV-1 disease progression. *The Lancet*, 352(9131), 866–870. [https://doi.org/10.1016/S0140-6736\(98\)04158-0](https://doi.org/10.1016/S0140-6736(98)04158-0)
- McGovern, B. H., Golan, Y., Lopez, M., Pratt, D., Lawton, A., Moore, G., ... Knox, T. A. (2007). The Impact of Cirrhosis on CD4+ T Cell Counts in HIV-Seronegative Patients. *Clinical Infectious Diseases*, 44(3), 431–437. <https://doi.org/10.1086/509580>
- McKinney, C., Merriman, M. E., Chapman, P. T., Gow, P. J., Harrison, A. A., Highton, J., ... Merriman, T. R. (2008). Evidence for an influence of chemokine ligand 3-like 1 (CCL3L1) gene copy number on susceptibility to rheumatoid arthritis. *Annals of the Rheumatic Diseases*, 67(3), 409–413. <https://doi.org/10.1136/ard.2007.075028>
- McLaren, P. J., & Carrington, M. (2015, May 19). The impact of host genetic variation on infection with HIV-1. *Nature Immunology*. Nature Research. <https://doi.org/10.1038/ni.3147>
- McLaren, P. J., Coulonges, C., Bartha, I., Lenz, T. L., Deutsch, A. J., Bashirova, A., ... Fellay, J. (2015). Polymorphisms of large effect explain the majority of the host genetic contribution to variation of HIV-1 virus load. *Proceedings of the National Academy of Sciences*, 112(47), 14658–14663. <https://doi.org/10.1073/pnas.1514867112>
- McMichael, A. J., Borrow, P., Tomaras, G. D., Goonetilleke, N., & Haynes, B. F. (2010). The immune response during acute HIV-1 infection: clues for vaccine development. *Nature Reviews Immunology*, 10(1), 11–23. <https://doi.org/10.1038/nri2674>
- Medical Development Division. (2011). *Guidelines for the management of adult HIV Infection with antiretroviral therapy*. Ministry of Health Malaysia. <https://doi.org/10.1017/CBO9781107415324.004>
- Medical Development Division. (2017). *Malaysian Consensus Guidelines on Antiretroviral Therapy 2017*. Ministry of Health Malaysia. Ministry of Health Malaysia.
- Melikyan, G. B., Markosyan, R. M., Hemmati, H., Delmedico, M. K., Lambert, D. M., & Cohen, F. S. (2000). Evidence that the transition of HIV-1 gp41 into a six-helix bundle, not the bundle configuration, induces membrane fusion. *Journal of Cell Biology*, 151(2), 413–423. <https://doi.org/10.1083/jcb.151.2.413>
- Mellors, J. W. (1997). Plasma Viral Load and CD4+ Lymphocytes as Prognostic Markers of HIV-1 Infection. *Annals of Internal Medicine*, 126(12), 946.

- <https://doi.org/10.7326/0003-4819-126-12-199706150-00003>
- Menten, P., Struyf, S., Schutyser, E., Wuyts, A., De Clercq, E., Schols, D., ... Van Damme, J. (1999). The LD78 $\beta$  isoform of MIP-1 $\alpha$  is the most potent CCR5 agonist and HIV-1- inhibiting chemokine. *Journal of Clinical Investigation*, 104(4), R1-5. <https://doi.org/10.1172/JCI7318>
- Ministry of Health Malaysia. (2017). Malaysian consensus guidelines on antiretroviral therapy 2017, (August), 92.
- Mitsuya, H., Weinhold, K. J., Furman, P. A., St Clair, M. H., Lehrman, S. N., Gallo, R. C., ... Broder, S. (1985). 3'-Azido-3'-deoxythymidine (BW A509U): an antiviral agent that inhibits the infectivity and cytopathic effect of human T-lymphotropic virus type III/lymphadenopathy-associated virus in vitro. *Proceedings of the National Academy of Sciences*, 82(20), 7096–7100. <https://doi.org/10.1073/pnas.82.20.7096>
- Mocroft, A., Phillips, A., Gatell, J., Ledergerber, B., Fisher, M., Clumeck, N., ... Lundgren, J. (2007). Normalisation of CD4 counts in patients with HIV-1 infection and maximum virological suppression who are taking combination antiretroviral therapy: an observational cohort study. *Lancet*, 370(9585), 407–413. [https://doi.org/10.1016/S0140-6736\(07\)60948-9](https://doi.org/10.1016/S0140-6736(07)60948-9)
- Modi, W. S. (2004). CCL3L1 and CCL4L1 chemokine genes are located in a segmental duplication at chromosome 17q12. *Genomics*, 83(4), 735–738. <https://doi.org/10.1016/j.ygeno.2003.09.019>
- Molokhia, M., Fanciulli, M., Petretto, E., Patrick, A. L., McKeigue, P., Roberts, A. L., ... Aitman, T. J. (2011). FCGR3B copy number variation is associated with systemic lupus erythematosus risk in Afro-Caribbeans. *Rheumatology*, 50(7), 1206–1210. <https://doi.org/10.1093/rheumatology/keq456>
- Molon, B., Gri, G., Bettella, M., Gómez-Moutón, C., Lanzavecchia, A., Martínez-A, C., ... Viola, A. (2005). T cell costimulation by chemokine receptors. *Nature Immunology*, 6(5), 465–471. <https://doi.org/10.1038/ni1191>
- Moore, D M, Hogg, R. S., Yip, B., Wood, E., Tyndall, M., Braitstein, P., & Montaner, J. S. G. (2005). Discordant immunologic and virologic responses to highly antiretroviral therapy are associated with increased mortality and poor adherence to therapy. *Journal of Acquired Immune Deficiency Syndromes*, 40(3), 288–293.
- Moore, David M., Hogg, R. S., Yip, B., Craib, K., Wood, E., & Montaner, J. S. G. (2006). CD4 percentage is an independent predictor of survival in patients starting antiretroviral therapy with absolute CD4 cell counts between 200 and 350 cells/??L. *HIV Medicine*, 7(6), 383–388. <https://doi.org/10.1111/j.1468-1293.2006.00397.x>
- Moore, R. D., & Keruly, J. C. (2007). CD4+ Cell Count 6 Years after Commencement of Highly Active Antiretroviral Therapy in Persons with Sustained Virologic Suppression. *Clinical Infectious Diseases*, 44(3), 441–446. <https://doi.org/10.1086/510746>
- Morison, L. (2001). The global epidemiology of HIV/AIDS. *British Medical Bulletin*, 58(1), 7–18. <https://doi.org/10.1093/bmb/58.1.7>
- Moscicki, R., Amento, E., Krane, S., Kurnick, J., & Colvin, R. (1983). Modulation of surface antigens of a human monocyte cell line, U937, during incubation with T-lymphocyte conditioned medium: detection of T4 antigen and its presence on normal blood monocytes. *Journal of Immunology*, 131, 743–748.
- Mugavero, M. J., Amico, K. R., Westfall, A. O., Crane, H. M., Zinski, A., Willig, J. H., ... Saag, M. S. (2012). Early Retention in HIV Care and Viral Load Suppression: Implications for a Test and Treat Approach to HIV Prevention. *J*

- Acquir Immune Defic Syndr*, 59(1), 86–93.  
<https://doi.org/10.1097/QAI.0b013e318236f7d2.Early>
- Mummidi, S., Ahuja, S. S. K., Gonzalez, E., Anderson, S. a, Santiago, E. N., Stephan, K. T., ... Ahuja, S. S. K. (1998). Genealogy of the CCR5 locus and chemokine system gene variants associated with altered rates of HIV-1 disease progression. *Nature Medicine*, 4, 786–793. <https://doi.org/10.1038/nm0798-786>
- Mummidi, Srinivas, Bamshad, M., Ahuja, S. S., Gonzalez, E., Feuillet, P. M., Begum, K., ... Ahuja, S. K. (2000). Evolution of human and non-human primate CC chemokine receptor 5 gene and mRNA: Potential roles for haplotype and mRNA diversity, differential haplotype-specific transcriptional activity, and altered transcription factor binding to polymorphic nucleotides. *Journal of Biological Chemistry*, 275(25), 18946–18961. <https://doi.org/10.1074/jbc.M000169200>
- Murphy, P. M., Baggolini, M., Charo, I. F., Hébert, C. a, Horuk, R., Matsushima, K., ... Power, C. A. (2000). International union of pharmacology. XXII. Nomenclature for chemokine receptors. *Pharmacological Reviews*, 52(1), 145–176. <https://doi.org/10.1124/pr.54.2.265>
- Mustanski, B., Newcomb, M. E., Du Bois, S. N., Garcia, S. C., & Grov, C. (2011). HIV in young men who have sex with men: a review of epidemiology, risk and protective factors, and interventions. *Journal of Sex Research*, 48(2–3), 218–253. <https://doi.org/10.1080/00224499.2011.558645>
- Nakajima, T., Kaur, G., Mehra, N., & Kimura, A. (2009). HIV-1/AIDS susceptibility and copy number variation in CCL3L1, a gene encoding a natural ligand for HIV-1 co-receptor CCR5. *Cytogenetic and Genome Research*. <https://doi.org/10.1159/000184703>
- Nakajima, Toshiaki, Ohtani, H., Naruse, T., Shibata, H., Mimaya, J. I., Terunuma, H., & Kimura, A. (2007). Copy number variations of CCL3L1 and long-term prognosis of HIV-1 infection in asymptomatic HIV-infected Japanese with hemophilia. *Immunogenetics*, 59(10), 793–798. <https://doi.org/10.1007/s00251-007-0252-4>
- Nakayama, E. E., Tanaka, Y., Nagai, Y., Iwamoto, A., & Shiota, T. (2004). A CCR2-V64I polymorphism affects stability of CCR2A isoform. *AIDS*, 18(5), 729–738. <https://doi.org/10.1097/00002030-200403260-00003>
- Nibbs, R. J. B., Yang, J., Landau, N. R., Mao, J. H., & Graham, G. J. (1999). LD78 $\beta$ , a non-allelic variant of human MIP-1 $\alpha$  (LD78 $\alpha$ ), has enhanced receptor interactions and potent HIV suppressive activity. *Journal of Biological Chemistry*, 274(25), 17478–17483. <https://doi.org/10.1074/jbc.274.25.17478>
- Nookhai, S., Ruxrungtham, K., Phanuphak, P., & Oelrichs, R. (2000). Prevalence of CCR2-64I, SDF1-3'A and CCR5- $\Delta$ 32 alleles in healthy Thais. *European Journal of Immunogenetics*, 27(3), 153–157. <https://doi.org/10.1080/00207540500103821>
- Nordang, G. B. N., Carpenter, D., Viken, M. K., Kvien, T. K., Armour, J. A. L., & Lie, B. A. (2012). Association analysis of the CCL3L1 copy number locus by parologue ratio test in Norwegian rheumatoid arthritis patients and healthy controls. *Genes and Immunity*, 13(7), 579–582. <https://doi.org/10.1038/gene.2012.30>
- O'Brien, S. J., & Nelson, G. W. (2004). Human genes that limit AIDS. *Nature Genetics*, 36(6), 565–574. <https://doi.org/10.1038/ng1369>
- O'Brien, T. R., McDermott, D. H., Ioannidis, J. P. A., Carrington, M., Murphy, P. M., Havlir, D. V, & Richman, D. D. (2000). Effect of chemokine receptor gene polymorphisms on the response to potent antiretroviral therapy. *AIDS*, 14(7),

- 821–826. <https://doi.org/10.1097/00002030-200005050-00008>
- Okulicz, J. F., Marconi, V. C., Landrum, M. L., Wegner, S., Weintrob, A., Ganesan, A., ... Dolan, M. J. (2009). Clinical Outcomes of Elite Controllers, Viremic Controllers, and Long-Term Nonprogressors in the US Department of Defense HIV Natural History Study. *The Journal of Infectious Diseases*, 200(11), 1714–1723. <https://doi.org/10.1086/646609>
- Palella, F. J., Delaney, K. M., Moorman, A. C., Loveless, M. O., Fuhrer, J., Satten, G. A., ... Holmberg, S. D. (1998). Declining Morbidity and Mortality among Patients with Advanced Human Immunodeficiency Virus Infection. *New England Journal of Medicine*, 338(13), 853–860. Retrieved from <http://www.embase.com/search/results?subaction=viewrecord&from=export&id=L352186146>
- Pantaleo, G., Menzo, S., Vaccarezza, M., Graziosi, C., Cohen, O. J., Demarest, J. F., ... Fauci, A. S. (1995). Studies in Subjects with Long-Term Nonprogressive Human Immunodeficiency Virus Infection. *New England Journal of Medicine*, 332(4), 209–216. <https://doi.org/10.1056/NEJM199501263320402>
- Pascal, V., Yamada, E., Martin, M. P., Alter, G., Altfeld, M., Metcalf, J. A., ... McVicar, D. W. (2007). Detection of KIR3DS1 on the Cell Surface of Peripheral Blood NK Cells Facilitates Identification of a Novel Null Allele and Assessment of KIR3DS1 Expression during HIV-1 Infection. *The Journal of Immunology*, 179(3), 1625–1633. <https://doi.org/10.4049/jimmunol.179.3.1625>
- Pascual-Pareja, J. F., Martínez-Prats, L., Luczkowiak, J., Fiorante, S., Rubio, R., Pulido, F., ... Delgado, R. (2010). Detection of HIV-1 at between 20 and 49 copies per milliliter by the Cobas TaqMan HIV-1 v2.0 assay is associated with higher pretherapy viral load and less time on antiretroviral therapy. *Journal of Clinical Microbiology*, 48(5), 1911–1912. <https://doi.org/10.1128/JCM.02388-09>
- Passaes, C. P., & Sáez-Cirión, A. (2014). HIV cure research: Advances and prospects. *Virology*, 454–455(1), 340–352. <https://doi.org/10.1016/j.virol.2014.02.021>
- Pawlowski, A., Jansson, M., Sköld, M., Rottenberg, M. E., & Källenius, G. (2012). Tuberculosis and HIV co-infection. *PLoS Pathogens*, 8(2), e1002464. <https://doi.org/10.1371/journal.ppat.1002464>
- Pedraza, M. A., del Romero, J., Roldán, F., García, S., Ayerbe, M. C., Noriega, A. R., & Alcamí, J. (1999). HHeterosexual transmission of HIV-1 is associated with high plasma viral load levels and a positive viral isolation in the infected partner. and a positive viral isolation in the infected partner. *JAIDS*, 21(2), 120–125. <https://doi.org/http://dx.doi.org/10.1016/B978-0-444-63456-6.50046-6>
- Pelak, K., Need, A. C., Fellay, J., Shianna, K. V., Feng, S., Urban, T. J., ... Alter, G. (2011). Copy number variation of KIR genes influences HIV-1 control. *PLoS Biology*, 9(11). <https://doi.org/10.1371/journal.pbio.1001208>
- Persaud, D., Zhou, Y., Siliciano, J. M., & Siliciano, R. F. (2003). Latency in Human Immunodeficiency Virus Type 1 Infection : No Easy Answers. *Journal of Virology*, 77(3), 1659–1665. <https://doi.org/10.1128/JVI.77.3.1659>
- Petrovski, S., Wang, Q., Heinzen, E. L., Allen, A. S., & Goldstein, D. B. (2013). Genic intolerance to functional variation and the interpretation of personal genomes. *PLoS Genetics*, 9(8), e1003709. <https://doi.org/10.1371/journal.pgen.1003709>
- Phillips, A. N., Staszewski, S., Weber, R., Kirk, O., Francioli, P., Miller, V., ... Ledergerber, B. (2001). HIV viral load response to antiretroviral therapy according to the baseline CD4 cell count and viral load. *Journal of the American Medical Association*, 286(20), 2560–2567.

- <https://doi.org/10.1001/jama.286.20.2560>
- Piatak, M., Saag, M. S., Yang, L. C., Clark, S. J., Kappes, J. C., Luk, K. C., ... Lifson, J. D. (1993). High levels of HIV-1 in plasma during all stages of infection determined by competitive PCR. *Science*, 259(5102), 1749–1754. <https://doi.org/10.1126/science.8096089>
- Pinto, D., Pagnamenta, A. T., Klei, L., Anney, R., Merico, D., Regan, R., ... Betancur, C. (2010). Functional impact of global rare copy number variation in autism spectrum disorders. *Nature*, 466(7304), 368–372. <https://doi.org/10.1038/nature09146>
- Quillent, C., Oberlin, E., Braun, J., Rousset, D., Gonzalez-Canali, G., Métais, P., ... Beretta, A. (1998). HIV-1-resistance phenotype conferred by combination of two separate inherited mutations of CCR5 gene. *Lancet*, 351(9095), 14–18. [https://doi.org/10.1016/S0140-6736\(97\)09185-X](https://doi.org/10.1016/S0140-6736(97)09185-X)
- Quinn, T. C., Wawer, M. J., Sewankambo, N., Serwadda, D., Li, C., Wabwire-Mangen, F., ... Gray, R. H. (2000). Viral Load and Heterosexual Transmission of Human Immunodeficiency Virus Type 1. *New England Journal of Medicine*, 342(13), 921–929. <https://doi.org/10.1056/NEJM200003303421303>
- Rajasuriar, R., Gouillou, M., Spelman, T., Read, T., Hoy, J., Law, M., ... Lewin, S. R. (2011). Clinical predictors of immune reconstitution following combination antiretroviral therapy in patients from the australian HIV observational database. *PLoS ONE*, 6(6), e20713. <https://doi.org/10.1371/journal.pone.0020713>
- Rajasuriar, R., Khouri, G., Kamarulzaman, A., French, M. A., Cameron, P. U., & Lewin, S. R. (2013). Persistent immune activation in chronic HIV infection: Do any interventions work? *AIDS*. <https://doi.org/10.1097/QAD.0b013e32835ecb8b>
- Rathore, A., Chatterjee, A., Sivarama, P., Yamamoto, N., Singhal, P. K., & Dhole, T. N. (2009). Association of CCR5-59029 A/G and CCL3L1 Copy number polymorphism with HIV type 1 transmission/progression among HIV type 1-seropositive and repeatedly sexually exposed HIV type 1-seronegative north Indians. *AIDS Research and Human Retroviruses*, 25(11), 1149–1156. <https://doi.org/10.1089/aid.2008.0019>
- Redon, R., Ishikawa, S., Fitch, K. R., Feuk, L., Perry, G. H., Andrews, T. D., ... Hurles, M. E. (2006). Global variation in copy number in the human genome. *Nature*, 444(7118), 444–454. <https://doi.org/10.1038/nature05329>
- Reeves, J. D., & Doms, R. W. (2002). Human immunodeficiency virus type 2. *Journal of General Virology*. <https://doi.org/10.1099/0022-1317-83-6-1253>
- Reynes, J., Portales, P., Segondy, M., Baillat, V., André, P., Réant, B., ... Corbeau, P. (2000). CD4+ T Cell Surface CCR5 Density as a Determining Factor of Virus Load in Persons Infected with Human Immunodeficiency Virus Type 1. *The Journal of Infectious Diseases*, 181(3), 927–932. <https://doi.org/10.1086/315845>
- Richman, D. D., Margolis, D. M., Delaney, M., Greene, W. C., Hazuda, D., & Pomerantz, R. J. (2009). The Challenge of Finding a Cure for HIV Infection. *Science*, 323, 1304–1308. <https://doi.org/10.1126/science.1165706>
- Robbins, G. K., Spritzler, J. G., Chan, E. S., Asmuth, D. M., Gandhi, R. T., Rodriguez, B. A., ... Pollard, R. B. (2009). Incomplete Reconstitution of T Cell Subsets on Combination Antiretroviral Therapy in the AIDS Clinical Trials Group Protocol 384. *Clinical Infectious Diseases*, 48(3), 350–361. <https://doi.org/10.1086/595888>
- Roberts, T., Cohn, J., Bonner, K., & Hargreaves, S. (2016). Scale-up of Routine Viral Load Testing in Resource-Poor Settings: Current and Future Implementation Challenges. *Clinical Infectious Diseases*, 62(8), 1043–1048.

- <https://doi.org/10.1093/cid/ciw001>
- Saag, M. S., Holodniy, M., Kuritzkes, D. R., O'Brien, W. A., Coombs, R., Poscher, M. E., ... Volberding, P. A. (1996, June). HIV viral load markers in clinical practice. *Nature Medicine*. <https://doi.org/10.1038/nm0696-625>
- Sakai, K., Gatanaga, H., Takata, H., Oka, S., & Takiguchi, M. (2010). Comparison of CD4+ T-cell subset distribution in chronically infected HIV+ patients with various CD4 nadir counts. *Microbes and Infection*, 12(5), 374–381. <https://doi.org/10.1016/j.micinf.2010.01.013>
- Sakuma, T., Barry, M. A., & Ikeda, Y. (2012). Lentiviral vectors: basic to translational. *Biochemical Journal*, 443(3), 603–618. <https://doi.org/10.1042/BJ20120146>
- Sallusto, F., & Bagiolini, M. (2008, April 9). Chemokines and leukocyte traffic. *Nature Immunology*. Nature Publishing Group. <https://doi.org/10.1038/ni.f.214>
- Samson, M., Labbe, O., Mollereau, C., Vassart, G., & Parmentier, M. (1996). Molecular cloning and functional expression of a new human CC-chemokine receptor gene. *Biochemistry*, 35(11), 3362–3367. <https://doi.org/10.1021/bi952950g>
- Samson, M., Libert, F., Doranz, B. J., Rucker, J., Liesnard, C., Farber, C.-M., ... Parmentier, M. (1996). Resistance to HIV-1 infection in Caucasian individuals bearing mutant alleles of the CCR-5 chemokine receptor gene. *Nature*, 382(6593), 722–725. <https://doi.org/10.1038/382722a0>
- Samson, M., Soularue, P., Vassart, G., & Parmentier, M. (1996). The genes encoding the human CC-chemokine receptors CC-CKR1 to CC-CKR5 (CMKBR1-CMKBR5) are clustered in the p21.3-p24 region of chromosome 3. *Genomics*, 36(3), 522–526. <https://doi.org/10.1006/geno.1996.0498>
- Schneider, H., Blaauw, D., Gilson, L., Chabikuli, N., & Goudge, J. (2006). Health Systems and Access to Antiretroviral Drugs for HIV in Southern Africa: Service Delivery and Human Resources Challenges. *Reproductive Health Matters*, 14(27), 12–23. [https://doi.org/10.1016/S0968-8080\(06\)27232-X](https://doi.org/10.1016/S0968-8080(06)27232-X)
- Scott, H. M., Vittinghoff, E., Irvin, R., Sachdev, D., Liu, A., Gurwith, M., & Buchbinder, S. P. (2014). Age, race/ethnicity, and behavioral risk factors associated with per contact risk of HIV infection among men who have sex with men in the United States. *Journal of Acquired Immune Deficiency Syndromes* (1999), 65(1), 115–121. <https://doi.org/10.1097/QAI.0b013e3182a98bae>
- Selinger, C., & Katze, M. G. (2013). Mathematical models of viral latency. *Current Opinion in Virology*, 3(4), 402–407. <https://doi.org/10.1016/j.coviro.2013.06.015>
- Shalekoff, S., Meddows-Taylor, S., Schramm, D. B., Donninger, S. L., Gray, G. E., Sherman, G. G., ... Tiemessen, C. T. (2008). Host CCL3L1 gene copy number in relation to HIV-1-specific CD4+ and CD8+ T-cell responses and viral load in South African women. *Journal of Acquired Immune Deficiency Syndromes*, 48(3), 245–254. <https://doi.org/10.1097/QAI.0b013e31816fdc77>
- Shao, W., Tang, J., Song, W., Wang, C., Li, Y., Wilson, C. M., & Kaslow, R. A. (2007). CCL3L1 and CCL4L1: Variable gene copy number in adolescents with and without human immunodeficiency virus type 1 (HIV-1) infection. *Genes and Immunity*, 8(3), 224–231. <https://doi.org/10.1038/sj.gene.6364378>
- Shrestha, S., Nyaku, M., & Edberg, J. C. (2010). Variations in CCL3L gene cluster sequence and non-specific gene copy numbers. *BMC Research Notes*, 3, 74. <https://doi.org/10.1186/1756-0500-3-74>
- Shrestha, S., Strathdee, S. a, Galai, N., Oleksyk, T., Fallin, M. D., Mehta, S., ... Smith, M. W. (2006). Behavioral risk exposure and host genetics of susceptibility to HIV-1 infection. *The Journal of Infectious Diseases*, 193(1), 16–26.

- <https://doi.org/10.1086/498532>
- Sierra, S., Kupfer, B., & Kaiser, R. (2005). Basics of the virology of HIV-1 and its replication. *Journal of Clinical Virology*. <https://doi.org/10.1016/j.jcv.2005.09.004>
- Siliciano, R. F., & Greene, W. C. (2011). HIV latency. *Cold Spring Harbor Perspectives in Medicine*, 1(1), a007096. <https://doi.org/10.1101/cshperspect.a007096>
- Singh, V., Srivastava, N., Srivastava, P., & Mittal, R. D. (2013). Impact of CCL2 and Its receptor CCR2 gene polymorphism in north indian population: A comparative study in different ethnic groups worldwide. *Indian Journal of Clinical Biochemistry*, 28(3), 259–264. <https://doi.org/10.1007/s12291-012-0265-0>
- Smith, C. J., Sabin, C. A., Youle, M. S., Kinloch-de Loes, S., Lampe, F. C., Madge, S., ... Phillips, A. N. (2004). Factors Influencing Increases in CD4 Cell Counts of HIV-Positive Persons Receiving Long-Term Highly Active Antiretroviral Therapy. *The Journal of Infectious Diseases*, 190(10), 1860–1868. <https://doi.org/10.1086/425075>
- Smith, M. W., Dean, M., Carrington, M., Winkler, C., Huttley, G. a, Lomb, D. a, ... O'Brien, S. J. (1997). Contrasting genetic influence of CCR2 and CCR5 variants on HIV-1 infection and disease progression. *Science*, 277(5328), 959–965. <https://doi.org/10.1126/science.277.5328.959>
- Sood, V., Rathore, A., Husain, S., Khan, S., Patra, S., Shankar, V., ... Banerjea, A. C. (2008). Host genes that affect progression of AIDS/HIV in india and novel gene therapeutic approaches against HIV. *Indian Journal of Geo-Marine Sciences*, 45(3), 141–148.
- Stephens, J. C., Reich, D. E., Goldstein, D. B., Shin, H. D., Smith, M. W., Carrington, M., ... Dean, M. (1998). Dating the Origin of the CCR5-Δ32 AIDS-Resistance Allele by the Coalescence of Haplotypes. *The American Journal of Human Genet. Stephens JC, Reich DE, Goldstein DB, Shin HD, Smith MW, Carrington M, et Al. Dating the Origin of the CCR5-Δ32 AIDS-Resistance Allele by the Coalescence of Haplotypes. Am J Hum Genet [Internet].* 1998;62(6):1507–15. Avai, 62(6), 1507–1515. <https://doi.org/10.1086/301867>
- Stirrup, O., Copas, A., Phillips, A., Gill, M., Geskus, R., Touloumi, G., ... Babiker, A. (2017). Predictors of CD4 cell recovery following initiation of antiretroviral therapy among HIV-1 positive patients with well-estimated dates of seroconversion. *HIV Medicine*, (October 2017), 184–194. <https://doi.org/10.1111/hiv.12567>
- Stone, V. E., Jordan, J., Tolson, J., Miller, R., & Pilon, T. (2004). Perspectives on adherence and simplicity for HIV-infected patients on antiretroviral therapy: self-report of the relative importance of multiple attributes of highly active antiretroviral therapy (HAART) regimens in predicting adherence. *Journal of Acquired Immune Deficiency Syndromes* (1999), 36(3), 808–816. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/15213564>
- Stranger, B. E., Forrest, M. S., Dunning, M., Ingle, C. E., Thorne, N., Redon, R., ... Dermitzakis, E. T. (2007). Relative impact of nucleotide and copy number variation on gene expression phenotypes. *Science*, 315(5813), 848–853. <https://doi.org/10.1126/science.1136678.Relative>
- Strathdee, S. A., Hallett, T. B., Bobrova, N., Rhodes, T., Booth, R., Abdool, R., & Hankins, C. A. (2010). HIV and risk environment for injecting drug users: The past, present, and future. *The Lancet*, 376(9737), 268–284. [https://doi.org/10.1016/S0140-6736\(10\)60743-X](https://doi.org/10.1016/S0140-6736(10)60743-X)

- Strauss, K., Hannet, I., Engels, S., Shiba, A., Ward, D. M., Ullery, S., ... Kestens, L. (1996). Performance evaluation of the FACSCount System: A dedicated system for clinical cellular analysis. *Communications in Clinical Cytometry*, 26(1), 52–59. [https://doi.org/10.1002/\(SICI\)1097-0320\(19960315\)26:1<52::AID-CYTO8>3.0.CO;2-I](https://doi.org/10.1002/(SICI)1097-0320(19960315)26:1<52::AID-CYTO8>3.0.CO;2-I)
- Struyf, S., Menten, P., Lenaerts, J. P., Put, W., D'Haese, A., De Clercq, E., ... Van Damme, J. (2001). Diverging binding capacities of natural LD78beta isoforms of macrophage inflammatory protein-1alpha to the CC chemokine receptors 1, 3 and 5 affect their anti-HIV-1 activity and chemotactic potencies for neutrophils and eosinophils. *European Journal of Immunology*, 31(7), 2170–2178.
- Su, B., Jin, L., Hu, F., Xiao, J., Luo, J., Lu, D., ... Chakraborty, R. (1999). Distribution of two HIV-1-resistant polymorphisms (SDF1-3'A and CCR2-64I) in East Asian and world populations and its implication in AIDS epidemiology. *American Journal of Human Genetics*, 65(4), 1047–1053. <https://doi.org/10.1086/302568>
- Suy, A., Castro, P., Nomdedeu, M., García, F., López, A., Fumero, E., ... Plana, M. (2007). Immunological profile of heterosexual highly HIV-exposed uninfected individuals: predominant role of CD4 and CD8 T-cell activation. *The Journal of Infectious Diseases*, 196(8), 1191–1201. <https://doi.org/10.1086/521193>
- Szabo, R., & Short, R. (2000). How does male circumcision protect against HIV infection? *BMJ*, 320(7249), 1592. <https://doi.org/10.1136/bmj.320.7249.1592>
- Tang, J., Li, X., Price, M. A., Sanders, E. J., Anzala, O., Karita, E., ... Gilmour, J. (2015). CD4:CD8 lymphocyte ratio as a quantitative measure of immunologic health in HIV-1 infection: Findings from an African cohort with prospective data. *Frontiers in Microbiology*, 6(JUL), 670. <https://doi.org/10.3389/fmicb.2015.00670>
- Tang, J., Shelton, B., Makhadze, N. J., Zhang, Y., Schaen, M., Louie, L. G., ... Kaslow, R. A. (2002). Distribution of chemokine receptor CCR2 and CCR5 genotypes and their relative contribution to human immunodeficiency virus type 1 (HIV-1) seroconversion, early HIV-1 RNA concentration in plasma, and later disease progression. *Journal of Virology*, 76(2), 662–672. <https://doi.org/10.1128/jvi.76.2.662-672.2002>
- Taub, D., Conlon, K., Lloyd, A., Oppenheim, J., & Kelvin, D. (1993). Preferential migration of activated CD4+ and CD8+ T cells in response to MIP-1 alpha and MIP-1 beta. *Science*, 260(5106), 355–358. <https://doi.org/10.1126/science.7682337>
- Taub, D. D., Turcovski-Corralles, S. M., Key, M. L., Longo, D. L., & Murphy, W. J. (1996). Chemokines and T lymphocyte activation: I. Beta chemokines costimulate human T lymphocyte activation in vitro. *Journal of Immunology (Baltimore, Md. : 1950)*, 156(6), 2095–2103. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/8690897>
- Taylor, J. M. G., Wang, Y., Ahdieh, L., Chmiel, J. S., Detels, R., Giorgi, J. V., ... Margolick, J. (2000). Causal pathways for CCR5 genotype and HIV progression. *Journal of Acquired Immune Deficiency Syndromes*, 23(2), 160–171. <https://doi.org/10.1097/00126334-200002010-00008>
- Taylor, N., Grabmeier-Pfistershammer, K., Egle, A., Greil, R., Rieger, A., Ledergerber, B., & Oberkofler, H. (2013). Cobas Ampliprep/Cobas TaqMan HIV-1 v2.0 Assay: Consequences at the Cohort Level. *PLoS ONE*, 8(8), e74024. <https://doi.org/10.1371/journal.pone.0074024>
- Tee, K. K., Pon, C. K., Kamarulzaman, A., & Ng, K. P. (2005). Emergence of HIV-1 CRF01\_AE/B unique recombinant forms in Kuala Lumpur, Malaysia. *AIDS*,

- 19(2), 119–126. <https://doi.org/10.1097/00002030-200501280-00003>
- Telenti, A., & Ioannidis, J. P. a. (2006). Susceptibility to HIV infection--disentangling host genetics and host behavior. *The Journal of Infectious Diseases*, 193(1), 4–6. <https://doi.org/10.1086/498535>
- The Wellcome Trust Case Control Consortium. (2010). Genome-wide association study of copy number variation in 16 , 000 cases of eight common diseases and 3 , 000 shared controls. *Nature*, 464(7289), 713–720. <https://doi.org/10.1038/nature08979.Genome-wide>
- Townson, J. R., Barcellos, L. F., & Nibbs, R. J. B. (2002). Gene copy number regulates the production of the human chemokine CCL3-L1. *European Journal of Immunology*, 32(10), 3016–3026. [https://doi.org/10.1002/1521-4141\(2002010\)32:10<3016::AID-IMMU3016>3.0.CO;2-D](https://doi.org/10.1002/1521-4141(2002010)32:10<3016::AID-IMMU3016>3.0.CO;2-D)
- Tsamis, F., Gavrilov, S., Kajumo, F., Seibert, C., Kuhmann, S., Ketts, T., ... Dragic, T. (2003). Analysis of the mechanism by which the small-molecule CCR5 antagonists SCH-351125 and SCH-350581 inhibit human immunodeficiency virus type 1 entry. *Journal of Virology*, 77(9), 5201–5208. <https://doi.org/10.1128/JVI.77.9.5201>
- Turner, B. G., & Summers, M. F. (1999). Structural biology of HIV. *Journal of Molecular Biology*, 285(1), 1–32. <https://doi.org/10.1006/jmbi.1998.2354>
- Tyson, J., Majerus, T. M., Walker, S., Armour, J. A. J. J. Al, Iafrate, A., Feuk, L., ... Kent, W. (2009). Quadruplex MAPH: improvement of throughput in high-resolution copy number screening. *BMC Genomics*, 10(1), 453. <https://doi.org/10.1186/1471-2164-10-453>
- UNAIDS. (2017a). *Data 2017*. <https://doi.org/978-92-9173-945-5>
- UNAIDS. (2017b). *Ending AIDS: Progress towards the 90–90–90 targets*.
- UNAIDS. (2017c). *UNAIDS Data 2017*. Retrieved from [http://www.unaids.org/sites/default/files/media\\_asset/20170720\\_Data\\_book\\_2017\\_en.pdf](http://www.unaids.org/sites/default/files/media_asset/20170720_Data_book_2017_en.pdf)
- Urban, T. J., Weintrob, A. C., Fellay, J., Colombo, S., Kevin, V., Gumbs, C., ... Goldstein, D. B. (2009). CCL3L1 and HIV/AIDS susceptibility Thomas. *Nat. Med.*, 15(10), 1110–1112. <https://doi.org/10.1038/nm1009-1110.CCL3L1>
- Usher, C. L., & McCarroll, S. A. (2015). Complex and multi-allelic copy number variation in human disease. *Briefings in Functional Genomics*, 14(5), 329–338. <https://doi.org/10.1093/bfgp/elv028>
- Van de Perre, P., Segondy, M., Foullongne, V., Ouedraogo, A., Konate, I., Huraux, J. M., ... Nagot, N. (2008). Herpes simplex virus and HIV-1: deciphering viral synergy. *The Lancet Infectious Diseases*, 8(8), 490–497. [https://doi.org/10.1016/S1473-3099\(08\)70181-6](https://doi.org/10.1016/S1473-3099(08)70181-6)
- van Manen, D., van 't Wout, A. B., & Schuitemaker, H. (2012). Genome-wide association studies on HIV susceptibility, pathogenesis and pharmacogenomics. *Retrovirology*, 9, 70. <https://doi.org/10.1186/1742-4690-9-70>
- Veal, C. D., Xu, H., Reekie, K., Free, R., Hardwick, R. J., Mcvey, D., ... Talbot, C. J. (2013). Automated design of parologue ratio test assays for the accurate and rapid typing of copy number variation, 29(16), 1997–2003. <https://doi.org/10.1093/bioinformatics/btt330>
- Wain, L. V., Armour, J. A. L., & Tobin, M. D. (2009). Genomic copy number variation, human health, and disease. *Lancet (London, England)*, 374(9686), 340–350. [https://doi.org/10.1016/S0140-6736\(09\)60249-X](https://doi.org/10.1016/S0140-6736(09)60249-X)
- Walker, S., Janyakhantikul, S., & Armour, J. A. L. (2009). Multiplex Parologue Ratio Tests for accurate measurement of multiallelic CNVs. *Genomics*, 93(1), 98–103.

- <https://doi.org/10.1016/j.ygeno.2008.09.004>
- Wang, B., Lau, K. A., Ong, L. Y., Shah, M., Steain, M. C., Foley, B., ... Saksena, N. K. (2007). Complex patterns of the HIV-1 epidemic in Kuala Lumpur, Malaysia: Evidence for expansion of circulating recombinant form CRF33\_01B and detection of multiple other recombinants. *Virology*, 367(2), 288–297. <https://doi.org/10.1016/j.virol.2007.05.033>
- Wang, C. X., Cannon, P. P. M., Chun, T., Carruth, L., Finzi, D., Finzi, D., ... Muth, A. (2016). The clinical applications of genome editing in HIV. *Blood*, 127(21), 2546–2552. <https://doi.org/10.1182/blood-2016-01-678144>
- Wang, H., Wolock, T. M., Carter, A., Nguyen, G., Kyu, H. H., Gakidou, E., ... Zuhlke, L. J. (2016). Estimates of global, regional, and national incidence, prevalence, and mortality of HIV, 1980–2015: the Global Burden of Disease Study 2015. *The Lancet HIV*, 3(8), e361–e387. [https://doi.org/10.1016/S2352-3018\(16\)30087-X](https://doi.org/10.1016/S2352-3018(16)30087-X)
- Ward, H., & Rönn, M. (2010). Contribution of sexually transmitted infections to the sexual transmission of HIV. *Current Opinion in HIV and AIDS*. <https://doi.org/10.1097/COH.0b013e32833a8844>
- Watts, J. M., Dang, K. K., Gorelick, R. J., Leonard, C. W., Bess, J. W., Swanstrom, R., ... Weeks, K. M. (2009). Architecture and secondary structure of an entire HIV-1 RNA genome. *Nature*, 460(7256), 711–716. <https://doi.org/10.1038/nature08237>
- Wensing, A. M. J., Van Maarseveen, N. M., & Nijhuis, M. (2010). Fifteen years of HIV Protease Inhibitors: raising the barrier to resistance. *Antiviral Research*, 85, 59–74. <https://doi.org/10.1016/j.antiviral.2009.10.003>
- Whalen, C., Horsburgh, C. R., Hom, D., Lahart, C., Simberkoff, M., & Ellner, J. (1995). Accelerated course of human immunodeficiency virus infection after tuberculosis. *American Journal of Respiratory and Critical Care Medicine*, 151(1), 129–135. <https://doi.org/10.1164/ajrccm.151.1.7812542>
- WHO. (2016). *Consolidated guidelines on the use of antiretroviral drugs for treating and preventing HIV infection. Recommendations for a public health approach - second edition*. Retrieved from <https://www.fast-trackcities.org/sites/default/files/WHO 2016 Guidelines on The Use of Antiretroviral Drugs for Treating and Preventing HIV Infection.pdf>
- WHO. (2018). *Global Health Estimates 2016: Deaths by Cause, Age, Sex, by Country and by Region, 2000-2016*. WHO. Geneva. <https://doi.org/2013>
- WHO, & Ministry of Health Malaysia. (2001). *Consensus Report on STI, HIV and AIDS Epidemiology*. Retrieved from [http://www.aidsdatahub.org/sites/default/files/documents/Consensus\\_Report\\_M AA\\_2001.pdf](http://www.aidsdatahub.org/sites/default/files/documents/Consensus_Report_M AA_2001.pdf)
- Wichukchinda, N., Nakayama, E. E., Rojanawiwat, A., Pathipvanich, P., Auwanit, W., Vongsheree, S., ... Shioda, T. (2008). Effects of CCR2 and CCR5 polymorphisms on HIV-1 infection in Thai females. *Journal of Acquired Immune Deficiency Syndromes*, 47(3), 293–297. <https://doi.org/10.1097/QAI.0b013e318162caab>
- Willcocks, L. C., Lyons, P. A., Clatworthy, M. R., Robinson, J. I., Yang, W., Newland, S. A., ... Smith, K. G. C. (2008). Copy number of FCGR3B, which is associated with systemic lupus erythematosus, correlates with protein expression and immune complex uptake. *The Journal of Experimental Medicine*, 205(7), 1573–1582. <https://doi.org/10.1084/jem.20072413>
- Wolbers, M., Babiker, A., Sabin, C., Young, J., Dorrucci, M., Chêne, G., ... CASCADE Collaboration Members, on behalf of the C. (2010). Pretreatment

- CD4 cell slope and progression to AIDS or death in HIV-infected patients initiating antiretroviral therapy--the CASCADE collaboration: a collaboration of 23 cohort studies. *PLoS Medicine*, 7(2), e1000239. <https://doi.org/10.1371/journal.pmed.1000239>
- World Health Organization. (2007). *Who Case Definitions of HIV for Surveillance and Revised Clinical Staging and Immunological Classification of HIV-related Disease in Adults and Children*.
- World Health Organization. (2016). *Consolidated guidelines on the use of antiretroviral drugs for treating and preventing HIV infection: recommendations for a public health approach*. World Health Organization. World Health Organization. <https://doi.org/10.1016/j.jped.2014.04.007>
- World Health Organization. (2017). *HIV Drug Resistance Report 2017*. World Health Organization. Retrieved from <http://apps.who.int/iris/bitstream/10665/255896/1/9789241512831-eng.pdf?ua=1>
- Wylie, J. L., Shah, L., & Jolly, A. M. (2006). Demographic, risk behaviour and personal network variables associated with prevalent hepatitis C, hepatitis B, and HIV infection in injection drug users in Winnipeg, Canada. *BMC Public Health*, 6(229). <https://doi.org/10.1186/1471-2458-6-229>
- Yang, Y., Chung, E. K., Wu, Y. L., Savelli, S. L., Nagaraja, H. N., Zhou, B., ... Yung Yu, C. (2007). Gene Copy-Number Variation and Associated Polymorphisms of Complement Component C4 in Human Systemic Lupus Erythematosus (SLE): Low Copy Number Is a Risk Factor for and High Copy Number Is a Protective Factor against SLE Susceptibility in European America. *The American Journal of Human Genetics*, 80(6), 1037–1054. <https://doi.org/10.1086/518257>
- Yazdanpanah, Y. (2004). Clinical efficacy of antiretroviral combination therapy based on protease inhibitors or non-nucleoside analogue reverse transcriptase inhibitors: indirect comparison of controlled trials. *BMJ*, 328(7434), 249–0. <https://doi.org/10.1136/bmj.37995.435787.a6>
- Zhao, X. Y., Lee, S. S., Wong, K. H., Chan, K. C. W., Ng, F., Chan, C. C. S., ... Zheng, B. J. (2005). Functional analysis of naturally occurring mutations in the open reading frame of CCR5 in HIV-infected Chinese patients and healthy controls. *Journal of Acquired Immune Deficiency Syndromes*, 38(5), 509–517. <https://doi.org/10.1097/01.qai.0000151004.19128.4a>
- Zheng, B. J., Zhao, X. Y., Zhu, N. S., Chan, C. P., Wong, K. H., Chi-Wai Chan, K., ... Lee, S. S. (2002). Polymorphisms of CCR5 gene in a southern Chinese population and their effects on disease progression in HIV infections. *AIDS*, 16(18), 2480–2482. <https://doi.org/10.1097/00002030-200212060-00016>
- Zhou, X. J., Cheng, F. J., Lv, J. C., Luo, H., Yu, F., Chen, M., ... Zhang, H. (2012). Higher DEFB4 genomic copy number in SLE and ANCA-associated small vasculitis. *Rheumatology (United Kingdom)*, 51(6), 992–995. <https://doi.org/10.1093/rheumatology/ker419>
- Zlotnik, A., & Yoshie, O. (2000). Chemokines: A new classification system and their role in immunity. *Immunity*, 12(2), 121–127. [https://doi.org/10.1016/S1074-7613\(00\)80165-X](https://doi.org/10.1016/S1074-7613(00)80165-X)

## **BIODATA OF STUDENT**

Dr. Irma Izani Mohamad Isa was born in Ipoh, Perak on 8<sup>th</sup> May 1985. She is married and lives in Semenyih, Selangor with her husband and four children. She had obtained her medical degree (MB BCh BAO) from University College of Dublin, Ireland in 2011 and Master of Science (MSc.) in Pharmacology and Toxicology from Universiti Putra Malaysia in December 2015. Her interest in becoming an academician had brought her to enrol Ph.D programme in Human Genetics since February 2016.

She had previously worked as houseman officer in Hospital Kajang for several months. However, due to her hearing condition at that time, she decided to continue her studies to pursue her ambition as a medical lecturer. While completing her Ph.D, she had worked as a part-time lecturer teaching Pharmacology subject at Royal College of Surgeon Ireland-Perdana University. In the future, she imagines herself as an expert in Pharmacology and Pharmacogenetics field, as combination of her Medical, Pharmacology and Human Genetics qualifications, to contribute to the personalised and precision medicine.

## LIST OF PUBLICATIONS

### **List of article publications**

Mohamad Isa II, Abu Bakar S, AbRahman AK. Ethnicity as predictor of immune reconstitution among Malaysian HIV-positive patients treated with highlyactive antiretroviral therapy. *J Med Virol.* 2020;1–9.<https://doi.org/10.1002/jmv.25680>

Mohamad Isa II, Jamaluddin J, Achim NH, Abubakar S. Population-specific profiling of *CCL3L1* copy number of the three major ethnic groups in Malaysia and the implication on HIV susceptibility [published online ahead of print, 2020 Jun 1]. *Gene.* 2020;754:144821. doi:10.1016/j.gene.2020.144821

Impacts of *CCR5* gene variants on HIV susceptibility and response to antiretroviral therapy among Malaysian HIV patients (submitted)

### **List of proceedings/conferences and others**

Irma Izani Mohamad Isa, Nurfarahin Hanini Achim, Ahmad Kashfi Ab Rahman, Azureen Azmel, Suresh Kumar Chidambaram, Siti Dalila Adnan, Cheah Yoke Kqueen, Zulkefley Othman and Suhaili Abu bakar. *Influence of Socio-Demographic and Viral Transmission Mode on Early CD4 Count and Viral Load among HIV Patients with Anti-Retroviral Therapy.* 4<sup>th</sup> National AIDS Conference, 28-30 September 2018, Swiss Garden Hotel Kuala Lumpur. (Poster presentation)

Irma Izani Mohamad Isa, Nurfhaezah Khairil Wahidin, Nur Sakinah Matnor, Ahmad Kashfi Ab Rahman, Cheah Yoke Kqueen, Zulkefley Othman and Suhaili Abu bakar. *Prevalence of 32 Basepairs Deletion Mutation of CCR5 Gene (CCR5-Δ32) among HIV-1 and Healthy Populations in Malaysia.* 4<sup>th</sup> National AIDS Conference, 28-30 September 2018, Swiss Garden Hotel Kuala Lumpur. (Poster presentation)

Mohamad Isa I and Abu Bakar S (2019). *Influence of polymorphisms in chemokine receptor-5(CCR5) and CCR2 on susceptibility to HIV, viral load and CD4 count with anti-retroviral therapy.* Front. Pharmacol. Conference Abstract: International Conference on Drug Discovery and Translational Medicine 2018 (ICDDTM '18) “Seizing Opportunities and Addressing Challenges of Precision Medicine”. doi: 10.3389/conf.fphar.2018.63.00139. (Oral presentation)

Path to a more tailored HIV treatment, Three-Minute Thesis (3MT) Competition, Faculty of Medicine and Health Sciences, 6<sup>th</sup> March 2019



## UNIVERSITI PUTRA MALAYSIA

### STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : Second Semester 2019/2020

#### TITLE OF THESIS / PROJECT REPORT :

COPY NUMBER VARIATION OF C-C CHEMOKINE LIGAND 3-LIKE 1 (CCL3L1), C-C CHEMOKINE RECEPTOR TYPE 5 (CCR5) AND CCR2 POLYMORPHISMS ON THE OUTCOMES OF ANTIRETROVIRAL THERAPY AMONG MALAYSIAN HIV PATIENTS

NAME OF STUDENT: IRMA IZANI MOHAMAD ISA

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

\*Please tick (v )

**CONFIDENTIAL**

(Contain confidential information under Official Secret Act 1972).

**RESTRICTED**

(Contains restricted information as specified by the organization/institution where research was done).

**OPEN ACCESS**

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

**PATENT**

Embargo from \_\_\_\_\_ until \_\_\_\_\_  
(date) (date)

**Approved by:**

(Signature of Student)  
New IC No/ Passport No.:

(Signature of Chairman of Supervisory Committee)  
Name:

Date :

Date :

[Note : If the thesis is **CONFIDENTIAL** or **RESTRICTED**, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted.]