



UNIVERSITI PUTRA MALAYSIA

**EFFECTS OF ROOT RESTRICTION AND MYCORRHIZA INOCULATION
ON GROWTH, PHYSIOLOGICAL AND YIELD RESPONSES OF CHILLI
GROWN IN SOILLESS CULTURE**

NURUL IDAYU ZAKARIA

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IN SOILLESS CULTURE**

By

NURUL IDAYU ZAKARIA

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Philosophy**

July 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

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By

NURUL IDAYU ZAKARIA

July 2018

Chairman : Professor Mohd Razi Ismail, PhD
Institute : Institute of Tropical Agriculture and Food Security

Chilli (*Capsicum annum* L.) that is used in food preparation for its flavour, aroma, and colour is a fruit vegetable with a high economic value that is cultivated in soilless culture in Malaysia. In soilless culture, the use of high substrate volume of about 6 to 8 liters per plant is closely related to increases in water usage, fertilizer and manpower. Meanwhile, the rising price of substrate and fertilizers coupled with pollution threat due to the disposal of used substrate in the large scale commercial production area results in higher production cost. Therefore, an approach to optimize the volume of substrate and concentration of fertilizer in small container could potentially reduce the production cost in soilless culture meanwhile by understanding physiological mechanism of the growth reduction, root system manipulation by using beneficial microbes such as mycorrhiza could potentially be adopted to improve yield under restricted root condition. Thus, the growth and yield responses of chilli plants grown in small containers were tested using six sizes of polybags. The result from this study showed reductions of stem diameter and total plant biomass of chilli grown in horizontal polybag of 17 cm in length with 32% yield reduction which is associated with root restriction compared to 31.7% yield increment in horizontal polybag of 27 cm in length. The next experiment was conducted to determine the optimum fertilizer concentration in different container sizes chosen from the previous study. Here the effects of 1.5 and 2.5 dS/m fertilizer concentration subjected to different container sizes of 2805 cm³, 6831 cm³ and 10557 cm³ on growth, physiological response and yield of chilli were determined. The result showed reduction of vegetative growth with 24% and 17% reduction of fruit fresh weight in 1.5 dS/m fertilizer concentration and

in 2805 cm³ container, respectively. Similarly, photosynthetic rate and stomatal conductance were reduced in 1.5 dS/m fertilizer concentration in 2805 cm³ container. Based on the aforementioned trials, 6831 cm³ container and 2.5 dS/m fertilizer concentration were found to be the optimum container size and fertilizer concentration for chilli production in soilless culture. The mechanism of growth reduction of plants subjected to root restriction remained unclear, therefore the following study was conducted to determine the physiological mechanism of growth reduction of root restricted chilli plants grown in small PVC column. The result from this study showed sucrose accumulation in the stem with changes in gas exchange parameters while chlorophyll fluorescence was not affected by root restriction. About 50% volume of soilless substrate can be saved with 23% of yield reduction for each plant subjected to root restriction. Based on the potential benefit of soilless substrate saving in small container, the following trial was conducted to minimize the yield gap of root restricted plants through manipulation of the root system to improve the root development by inoculation of beneficial microbes such as mycorrhiza. The result showed that inoculation of mycorrhiza in root restricted plants increased sucrose accumulation in the leaves, stem and fruit at flowering stage yet there were no net increase in yield compared to root restricted plants without mycorrhiza. Manipulation of the root system by mycorrhiza inoculation did not improve the growth and yield of chilli plants grown under restricted root condition.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah.

KESAN PEMBATASAN AKAR DAN INOKULASI MIKORIZA TERHADAP TINDAK BALAS TUMBESARAN, FISILOGI DAN HASIL CILI YANG DITANAM DALAM KULTUR TANPA TANAH

Oleh

NURUL IDAYU ZAKARIA

Julai 2018

Pengerusi : Profesor Mohd Razi Ismail, PhD
Institut : Institut Pertanian Tropika dan Sekuriti Makanan

Cili (*Capsicum annum* L.) yang digunakan untuk rasa, aroma dan warna di dalam penyediaan makanan merupakan sayur-sayuran buah bernilai tinggi yang ditanam menggunakan kultur tanpa tanah di Malaysia. Dalam kultur tanpa tanah, keperluan jumlah substrat yang tinggi kira-kira 6 hingga 8 liter bagi setiap tanaman adalah berkait rapat dengan penggunaan air, baja dan tenaga manusia yang tinggi. Di samping itu, kenaikan harga substrat dan baja ditambah dengan ancaman pencemaran kerana pelupusan substrat terpakai di kawasan pengeluaran komersial berskala besar membawa kepada kos pengeluaran yang lebih tinggi. Oleh itu, pendekatan untuk mengoptimalkan jumlah substrat dan kepekatan baja di dalam bekas kecil berpotensi untuk mengurangkan kos pengeluaran dalam kultur tanpa tanah, manakala dengan memahami mekanisme pengurangan tumbesaran, pengubahsuaian sistem akar dengan menggunakan mikrob bermanfaat seperti mikoriza berpotensi untuk digunakan bagi meningkatkan hasil di dalam keadaan sekatan akar. Oleh itu, tindak balas pertumbuhan dan hasil tanaman cili yang ditanam di dalam bekas kecil telah diuji dengan menggunakan enam saiz polibeg. Keputusan kajian ini menunjukkan pengurangan diameter batang dan jumlah biomas tumbuhan cili yang ditanam di dalam polibeg horizontal dengan panjang 17 cm dengan pengurangan hasil sebanyak 32% yang dikaitkan dengan sekatan akar berbanding kenaikan hasil sebanyak 31.7% di dalam polibeg horizontal dengan panjang 27 cm. Kajian berikutnya telah dijalankan untuk menentukan kepekatan optimum baja di dalam saiz bekas berbeza yang dipilih daripada kajian terdahulu. Di sini, kesan kepekatan baja 1.5 dan 2.5 dS/m di dalam saiz bekas berbeza iaitu 2805 cm³, 6831 cm³ dan 10557 cm³ telah ditentukan terhadap tumbesaran, tindak balas fisiologi dan hasil cili. Keputusannya menunjukkan penurunan pertumbuhan vegetatif dengan 24% dan 17% pengurangan berat buah segar dalam kepekatan baja 1.5 dS/m dan

di dalam bekas 2805 cm³, masing-masing. Begitu juga, kadar fotosintesis dan konduksi stomata berkurang dalam kepekatan baja 1.5 dS/m di dalam bekas 2805 cm³. Berdasarkan ujikaji tersebut, bekas 6831 cm³ dan kepekatan baja 2.5 dS/m didapati sebagai ukuran bekas dan kepekatan baja yang optimum bagi pengeluaran cili dalam kultur tanpa tanah. Mekanisme pengurangan tumbesaran tanaman yang ditanam dengan sekatan akar adalah tidak jelas, oleh itu, kajian seterusnya telah dijalankan untuk menentukan mekanisme fisiologi pengurangan tumbesaran tanaman cili dengan sekatan akar yang ditanam di dalam kolom PVC kecil. Keputusan kajian ini menunjukkan pengumpulan sukrosa di dalam batang dengan perubahan parameter pertukaran gas sementara fluorezen klorofil tidak terjejas oleh sekatan akar. Sebanyak 50% jumlah substrat tanpa tanah boleh dijimatkan dengan 23% pengurangan hasil bagi setiap tanaman yang ditanam dengan sekatan akar. Berdasarkan manfaat potensi penjimatan substrat tanpa tanah di dalam bekas kecil, kajian berikutnya dilakukan untuk meminimumkan jurang hasil tanaman dengan sekatan akar melalui manipulasi sistem akar untuk memperbaiki perkembangan akar dengan inokulasi mikrob yang bermanfaat seperti mikoriza. Keputusannya menunjukkan bahawa inokulasi mikoriza ke dalam tanaman dengan sekatan akar meningkatkan pengumpulan sukrosa di dalam daun, batang dan buah pada peringkat berbunga tetapi tidak ada kenaikan bersih hasil berbanding tanaman dengan sekatan akar tanpa inokulasi mikoriza. Pengubahsuaian sistem akar melalui inokulasi mikoriza tidak dapat meningkatkan tumbesaran dan hasil tanaman cili yang ditanam di dalam keadaan sekatan akar.

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I certify that a Thesis Examination Committee has met on 6 July 2018 to conduct the final examination of Nurul Idayu binti Zakaria on her thesis entitled "Effects of Root Restriction and Mycorrhiza Inoculation on Growth, Physiological and Yield Responses of Chilli Grown in Soilless Culture" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Halimi b Mohd Saud, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Mahmud b Tengku Muda Mohamed, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Osumanu Haruna Ahmed, PhD

Professor
Faculty of Agriculture and Food Science (Bintulu Campus)
Universiti Putra Malaysia
(Internal Examiner)

Takashi Muraoka, PhD

Professor
Center for Nuclear Energy in Agriculture
University of Sao Paulo
Brazil
(External Examiner)



ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 4 September 2019

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohd Razi Ismail, PhD

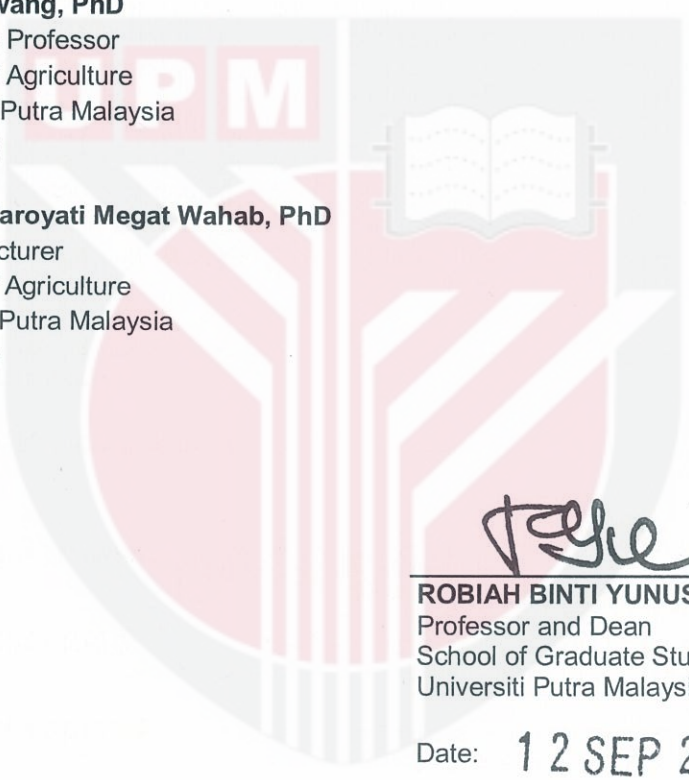
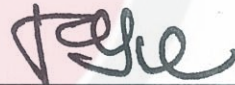
Professor
Institute of Tropical Agriculture and Food Security
Universiti Putra Malaysia
(Chairman)

Yahya Awang, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Puteri Edaroyati Megat Wahab, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)



ROBIAH BINTI YUNUS, PhD
Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 12 SEP 2019

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Signature: _____
Name of Chairman
of Supervisory
Committee: Professor Dr. Mohd Razi Ismail

Signature: _____
Name of Member of
Supervisory
Committee: Associate Professor Dr. Yahya Awang

Signature: _____
Name of Member of
Supervisory
Committee: Dr. Puteri Edaroyati Megat Wahab

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LIST OF ABBREVIATIONS

%	Percentage
<	Less than
=	Equal to
>	Greater than
≤	Less than and equal to
*	Significantly different at P≤0.05
**	Significantly different at P≤0.01
°C	Degree celcius
µg	Microgram
µmol	micromol
A	Net CO ₂ assimilation rate
AAS	Atomic Absorption spectroscopy
ABA	Abscisic acid
AI	Acid invertase
ANOVA	Analysis of variance
ATP	Adenine TriPhosphate
C/N ratio	Carbon/nitrogen ratio
Ca ²⁺	Calcium
CCD	Coconut coir dust
cm	Centimetre
cm ²	Centimeter square
CO ₂	Carbon dioxide
DAT	Day after transplant
dS/m	desiSiemen per meter
EC	Electrical conductivity
EFB	Empty fruit bunch
F _m	Maximal fluorescence
F _N	Nitrogen uptake rate per root unit volume
F _o	Minimal fluorescence
F _v /F _m	Quantum yield of PSII
F _w	Transpiration rate per root unit volume
g	gram
H ₂ O ₂	Hydrogen peroxide
K	Potassium
kg	kilogram
L	Litre
LSD	Least Significant Different
m ²	Meter square
m ³	Metre cubic
Mg	Magnesium
mg	miligram
ml	mililitre
mmol	milimole
mS/cm	MiliSiemen per centimetre
N	Nitrogen

n
NFT
ns/NS
P
pH
PVC
RCBD
SE
SuSy
UV
V:v

Number of observation
Nutrient Film Technique
Not significant
Phosphorus
Measurement of acidity/alkalinity
Polyvinyl-chloride
Randomized Complete Block Design
Standard error
Sucrose synthase
Ultraviolet
Volume per volume



CHAPTER 1

INTRODUCTION

In Malaysia, vegetables are crucial food crops as they contribute towards development of agricultural sector with a planted area of about 63,569 hectares and total annual production of about 1,195,647 tonnes in 2016 (Department of Agriculture, 2016). Chilli (*Capsicum annum* L.) is among the most important high value vegetables in Malaysia. It is of economical importance due to its pungency and high level of provitamin A, vitamin C and E, and carotenoids (Perucka and Materska, 2007; Shaha *et al.*, 2013). It is either marketed as fresh chilli or processed products such as chilli sauces and chilli pastes. From 2012 to 2016, the chilli production area increased from 2.5 to 3.1 thousand hectares while annual production increased from 40 to 43.7 thousand tonnes (Department of Agriculture, 2016). It is necessary to improve domestic production of chilli to attain the level of self sufficiency. Currently, the self sufficiency level of chilli is only at 51.4% which contributes significantly to the high food import bill (Department of Statistics, 2015).

The adoption of modern cultivation technique can improve plant growth, yield production and income. Substrate culture or fertigation system under protected structure has been used sporadically for commercial vegetables production in Malaysia. The major paradigm shift from the use of soil into container with soilless substrate is to control problems in agriculture, such as soil borne pest and diseases, lack of fertile soil, pollution of freshwater reservoir by discharge of nutritional ion and pesticide residue into the soil, water scarcity and high soil salinity (Raviv and Lieth, 2008). The widespread adoption of soilless culture under protected structures or rain shelters is primarily to replace uncertainty in crop production due to climatic changes such as excessive rainfall, to provide a favorable microclimatic condition for the growth of shoot and root whilst enabling precision crop management to provide a year round of vegetable (Ismail, 2000). Compared to conventional soil based cultivation, this growing system is more cost effective due to efficient and accurate control of water and nutrients and enhanced yield production and quality (Grafiadellis *et al.*, 2000; Nicola *et al.*, 2005; Nejad and Ismaili, 2014).

Presently, polyethylene bags or polybag filled with coconut coir dust as growing substrate is a crucial input and commonly use in soilless culture production to provide plant support and retain water and nutrient. Container filled with substrate is important variable cost items in soilless culture (Barreto and Jagtap, 2006; Rodriguez *et al.*, 2006; Cantliffe *et al.*, 2007; Barrett *et al.*, 2016). Beside substrate, fertilizers are also crucial inputs and are highest variable cost item in a soilless culture production (Engindeniz and Gul, 2009). Plants in substrate culture are related with root restriction since it has small root system compared to soil cultivation (Ismail and Davies, 1998; Raviv *et al.*, 2009).

Growers tend to use large container with higher requirement of substrate to promote root growth and avoid root restriction. However, this creates problem of poor substrate management due to oversupply of substrate which is closely related with higher requirement of water, fertilizer as well as manpower and pollution threat due to the disposal of used substrate in the large scale commercial production area that result in increased cost of production. Besides the problem of poor management of substrate, there are other problems for the growers, which is limited supply of substrate coupled with rising prices of substrate and fertilizer that affect the production cost. Hence, optimization of substrate volume and fertilizer concentration via production of plant in small container size is becoming important issue for the farmers to potentially reduce the high cost of production.

To address these problems, it is obviously important to find a proper management approach that promotes efficient use of substrate to reduce the production cost. Root restriction is important in soilless culture for production of container grown plant with limited substrate such as potted culture. It is a feasible solution to save substrate usage while minimizing the cost of production and controlling environment of root system (Shi *et al.*, 2008b). Varying container sizes resulted in variable degree of root restriction (Alsadon, 2000). Previous studies have shown that root restriction did markedly reduced plant growth and total plant yield (Ismail and Dalia, 1995; Xu *et al.*, 2001; Saito *et al.*, 2008; Massetani *et al.*, 2017). Although there was a slight decrease in total yield, but fruit quality and yield efficiency tends to be higher than unrestricted plant (Mandre *et al.*, 1995; Atkinson *et al.*, 1997; Mataa and Tominaga, 1998; Xie *et al.*, 2009).

In a soilless culture, plants are supplied with water including fertilizer by drip irrigation system in container filled with soilless substrate. Small root system of the root restricted plant may be beneficial in increasing efficiency of fertilizer use by targeting applications to plant roots (Mandre *et al.*, 1995). Container grown plant requires careful management of fertilizers as the limited root substrate and concentration of essential plant nutrients in small substrate are frequently insufficient to sustain plant growth (Falovo *et al.*, 2009). Fertilizer application in fertigation system is not specific to the optimal electrical conductivity (EC) but a certain range is used. Root restriction usually caused reduction of morphology and physiological process in plant (Ronchi *et al.*, 2006; Shi *et al.*, 2008a) but does not affect photosynthesis rate (Krizek *et al.*, 1985; Kharkina *et al.*, 1999). Low EC level of nutrient solution significantly reduced yield of lettuce (Falovo *et al.*, 2009) and rockmelon (Zulkarami *et al.*, 2012) however it did not affect the yield of cucumber plant (Al-Harbi, 1994). The extreme stress due to the combination of low nutrient concentration and small container significantly reduced total fruit yield of sweet pepper but increased the translocation of assimilates into the fruits (Xu *et al.*, 2001).

These studies demonstrate the potential benefits of root restriction for chilli production in soilless culture. The physiological mechanism of growth reduction remains unclear where there are contradictory findings of nutrient and water resources involvement in various plants under root restriction. Hameed *et al.* (1987) and Tschaplinski and Blake (1985) suggested that water stress led to the reduction of leaf growth and stomatal conductance of tomato and alder seedlings plants, respectively, grown in confined root space of solution culture. However, Krizek *et al.* (1985), Rieger and Marra (1994) and Ismail and Davies (1998) ruled out the involvement of water stress in growth reduction of root restricted soybean, young peach tree and pepper, respectively. Furthermore, Rieger and Marra (1994) and Shi *et al.* (2008a) suggested that factors other than photosynthesis and carbohydrate level were involved in controlling reduction of growth in the root restricted plant.

Several measures are available to manage chilli plant under root restriction particularly by manipulation of root development to enhance the yield gap. Previously, Shi *et al.* (2007) found that reduction of growth in root restricted plant was alleviated by aeration in the root zone of tomato plant grown under hydroponic system. Besides that, the exogenous application of hormones such as gibberellins and cytokinin can partially overcome reduction of growth in root restricted plant (Richards and Rowe, 1977; Carmi and Heuer, 1981). Other techniques of root manipulation such as by boosting more fertilizer, frequent irrigation and inoculation of beneficial microbes can be applied to root restriction. The role of beneficial microbes has been widely investigated in stress plant since the use of biological methods to alleviate the stress effect is more useful, economically and environmentally to the plant (Miransari *et al.*, 2008; Al-Khalil, 2010; Asrar and Elhindi, 2011).

Chilli plants require appropriate amount of substrate hence suitable level of root restriction for chilli plants need to be determined. The information on the appropriate EC of nutrient solution on chilli plants subjected to root restriction will enable growers not only to manage substrate and fertilizer wisely but also enable them to save substrate and fertilizer cost, however the information was limited. Complete understandings of the mechanism of growth reduction in root restriction are still not fully understood therefore to manage plant survival and enhancing yield under root restriction, physiological mechanism and source and sink relationship effect on growth and yield of root restricted chilli need to be determined. Information on improvement of plants yield under root restricted condition are still limited therefore manipulation of the root system by using mycorrhiza which is easy to apply and applicable to the farmer could potentially be adopted to improve yield under root restriction.

The main objective of this study is to determine the effect of root restriction and fertilizer concentration in root restricted chilli plants as well as to understand physiological mechanism of the growth reduction and yield improvement of root restricted chilli plants through inoculation of mycorrhiza.

The main hypotheses to be tested in the present study were:

- 1) Growing plant in small container size cause root restriction that will reduce plant growth and yield while growing chilli in optimum container size will improve plant growth and yield.
- 2) Appropriate application of nutrient solution EC level will improve plant growth, physiological response and yield of chilli subjected to root restriction.
- 3) Selected beneficial microbes will increase plant growth, physiological response and yield of chilli subjected to root restriction.

Looking the economic important of chilli in soilless culture and potential of adopting root restriction, this research was carried out to explore the following objectives:

- 1) To determine the effects of root restriction on growth and yield of chilli.
- 2) To determine the effect of low EC of nutrient solution on growth, physiological response and yield of chilli subjected to root restriction.
- 3) To understand the physiological mechanism and source and sink relationship on the reduction of growth and yield of chilli subjected to root restriction.
- 4) To determine the effect of manipulation of the root system by beneficial microbes on growth and yield of chilli subjected to root restriction.

REFERENCES

- A'fifah, A.R., Mohd Razi, I., Mohd Fauzihan, K., Puteri Edaroyati, M.W., Siti Norakmar, A. and Kausar, H. (2013). Changes in leaf gas exchange, biochemical properties, growth and yield of chilli grown under soilless culture subjected to deficit fertigation. *Australian Journal of Crop Science*. 7(10): 1582-1589.
- Abad, M., Noguera, P., Puchades, R., Maquieira, A., Noguera, V. (2002). Physico-chemical and chemical properties of some coconut coir dusts for use as a peat substitute for containerised ornamental plants. *Bioresource Technology* 82: 241-245.
- Abak, K., Dasgan, H.Y., Rehber, Y., Ortas, I. (2010). Effect of vesicular arbuscular mycorrhizas on plant growth of soilless grown muskmelon. *Acta Hort.* 871: 301-306.
- Abdul Aziz, R.A. (1994). The effect of nutrient solution conductivity on the growth of cucumber plants. *J. King Saud Univ. Vol 6, Agric Sci (1)*: 125-132.
- Abugre, S. and Oti-Boateng, C. (2011). Seed source variation and polybag size on early growth of *Jatropha curcas*. *Journal of Agricultural and Biological Science*. 6(4): 39-45.
- Ahmet, E. and Sakine, B. (2016). Growth and yield of pepper (*Capsicum annum* L.) under root zone restriction. *Agricultural Research & Technology: Open Access Journal*. 2(5): 1-6.
- Albaho, M., Bhat, N., Abo-Rezq, H., Thomas, B. (2009). Effect of three different substrates on growth and yield of two cultivars of *Capsicum annum*. *European Journal of Scientific Research*. 28(2): 227-233.
- Al-Harbi, A.R. (1994). The effect of nutrient solution conductivity on the growth of cucumber plants. *J. King Saud Univ.* 6(1): 125-132.
- Al-Khaliel, A.S. (2010). Effect of salinity stress on mycorrhizal association and growth response of peanut infected by *Glomus mosseae*. *Plant Soil Environ.*, 56(7): 318–324.
- Alsadon, A.A. (2000). Container size influences growth and yield of transplanted tomato. *Arab Univ. J. Agric. Sci. Shams Univ., Cairo*. 8(3): 705-718.
- Araki, A., Rattin, J., Di Benedetto, A., Mirave, P. (2007). Temperature and cytokinin relationships on lettuce (*Lactuca sativa* L.) and celery (*Apium graveolens* L.) nursery growth and yield. *International Journal of Agricultural Research*. 2(8):725-730.

- Arenas, M., Vavrina, C.S., Cornell, J.A., Hanlon, E.A., Hochmuth, G.J. (2002). Coir as an alternative to peat in media for tomato transplant production. *HortScience*. 37 (2): 309-312.
- Arnon, D.I. (1949). Copper enzymes in isolated chloroplasts. *Plant Physiology*. 24: 1–15.
- Arp, W.J. (1991). Effects of source-sink relations on photosynthetic acclimation to elevated CO₂. *Plant, Cell & Environment*. 14(8): 869–875.
- Artetxe, A., Teres, V., Beunza, A.I. (1997). Effects of container size and substrates on *Hydrangea macrophylla* growth. *Acta Hort*. 450: 419-424.
- Asaduzzaman, M., Saifullah, M., Salim Reza Mollick, A.K.M., Mokter Hossain, M., Halim, G.M.A., Asao, T. (2015). Soilless Culture - Use of Substrates for the Production of Quality Horticultural Crops. <http://dx.doi.org/10.5772/59708>.
- Ashraf, M. and Harris, P.J.C. (2013). Photosynthesis under stressful environments: An overview. *Photosynthetica*. 51(2): 163-190.
- Asiah, A., Mohd Razi, I., Mohd Khanif, Y., Marziah, M., Shaharuddin, M. (2004). Physical and chemical properties of coconut coir dust and oil palm empty fruit bunch and the growth of hybrid heat tolerant cauliflower plant. *Pertanika J. Trop. Agric. Sci*. 27(2): 121-133.
- Asrar, A-W. A. and Elhindi, K.M. (2011). Alleviation of drought stress of marigold (*Tagetes erecta*) plants by using arbuscular mycorrhizal fungi. *Saudi Journal of Biological Sciences*. 18: 93-98.
- Atkinson, C.J., Webster, A.D., Vaughan, S., Lucas, A.S. (1997). Effects of root restriction on the physiology of apple tree growth. *Acta Horticulturae*. 451: 587-598.
- Atwell, B.J. (1988). Physiological responses of lupin roots to soil compaction. *Plant and Soil*. 111: 277-281.
- Audet, P. and Charest, C. (2006). Effects of AM colonization on “wild tobacco” plants grown in zinc-contaminated soil. *Mycorrhiza*. 16: 277–283.
- Audet, P. and Charest, C. (2010). Identification of constraining experimental-design factors in mycorrhizal pot-growth studies. *Journal of Botany*, 2010. doi:10.1155/2010/718013.

- Awang, Y. and Ismail, M.R. (1997). The growth and flowering of some annual ornamentals on coconut dust. Proceedings of international symposium on growing media and plant nutrition in horticulturae, (Roeber, R.U., ed.) Acta Horticulturae 450: 31-38.
- Barreto, M.S. and Jagtap, K.B. (2006). Assessment of substrates for economical production of gerbera (*Gerbera jamesonii* Bolus ex Hooker F.) flowers under protected cultivation. Journal of Ornamental Horticulture. 9(2): 136-138.
- Barrett, G.E., Alexander, P.D., Robinson, J.S., Bragg, N.C. (2016). Achieving environmentally sustainable growing media for soilless plant cultivation systems – A review. Scientia Horticulturae. 212: 220–234.
- Bar-Tal, A. (1990). The significance of root size for plant nutrition in intensive horticulture. In Z. Rengel (Ed.) *Mineral Nutrition of Crops: Fundamental Mechanisms and Implications*. Florida: CRC Press.
- Bar-Tal, A. and Pressman, E. (1996). Root restriction and potassium and calcium solution concentrations affect dry-matter production, cation uptake, and blossom-end rot in greenhouse tomato. J. Amer. Soc. Hort. Sci. 121(4): 649-655.
- Bar-Tal, A. and Pressman, E. (1996). Root restriction and potassium and calcium solution concentrations affect dry-matter production, cation uptake, and blossom-end rot in greenhouse tomato. J. Amer. Soc. Hort. Sci. 121(4): 649–655.
- Bar-Tal, A., Feigin, A., Rylski, I., and Pressman, E. (1994a). Effects of root pruning and N-NO₃ solution concentration on tomato plant growth and fruit yield. Scientia Horticulturae. 58: 91-103.
- Bar-Tal, A., Feigin, A., Rylski, I., and Pressman, E. (1994b). Effects of root pruning and N-NO₃ solution concentration on nutrient uptake and transpiration of tomato plants. Scientia Horticulturae. 58: 77-90.
- Bar-Tal, A., Feigin, A., Sheinfeld, S., Rosenberg, R., Sternbaum, B., Rylski, I., Pressman, E. (1995). Root restriction and N-NO₃ solution concentration effects on nutrient uptake, transpiration and dry matter production of tomato. Scientia Horticulturae. 63: 195-208.
- Bar-Yosef, B., Schwartz, S., Markovich, T., Lucas, B., Assaf, R. (1988). Effect of root volume and nitrate solution concentration on growth, fruit yield and temporal N and water uptake rates by apple trees. Plant and Soil. 107: 49-56.

- Basu, S.K. and De, A.K. (2003). *Capsicum*: Historical and botanical perspectives. In: A.K. De (Ed.) *Capsicum: The genus Capsicum*. (pp. 1–15). London: Taylor & Francis Ltd.
- Bath, B., Kristensen, H.L., Thorup-Kristensen, K. (2008). Root pruning reduces root competition and increases crop growth in a living mulch cropping system. *Journal of Plant Interaction*. 3(3): 211-221.
- Bauerle, W.L., 1984. Bag culture production of greenhouse tomatoes. Special Circular 108, Ohio Agricultural Research and Development Center, Ohio State University, Wooster, OH.
- Beltrano, J., Ruscitti, M., Arango, M.C., Ronco, M. (2013). Effects of arbuscular mycorrhiza inoculation on plant growth, biological and physiological parameters and mineral nutrition in pepper grown under different salinity and P levels. *Journal of Soil Science and Plant Nutrition*. 13(1): 123-141.
- Benedetto, A.D. (2007). Alternative substrates for potted ornamental plants based on Argentinean peat and Argentinean river waste: A review. *Floriculture and Ornamental Biotechnology*. 1(2): 90-101.
- Bennie, A.T.P. (1991). Growth and mechanical impedance. In: Y. Waisel, A. Eshel, U. Kalkafi (Eds), *Plant Roots: The Hidden Half*. (pp. 393-414). New York: Marcel Dekker Inc.
- Ben-Porath, A., Baker, D.N. (1990). Taproot restriction effects on growth, earliness and dry weight partitioning of cotton. *Crop Sci*. 30: 809–814.
- Bhandari, S.R., Jung, B.D., Baek, H.Y., Lee, Y.S. (2013). Ripening-dependent changes in phytonutrients and antioxidant activity of red pepper (*Capsicum annuum* L.) fruits cultivated under open-field conditions. *HortScience*. 48:1275-1282.
- Bhat, R.M. and Rao, S.N.K. (1997). Growth and photosynthesis in bell-pepper as affected by sink manipulation. *Biologia Plantarum*. 39(3): 437-439.
- Bilderback, T.E. and Fonteno, W.C. (1987). Effects of container geometry and media physical properties on air and water volumes in containers. *J. Environ. Hort*. 5(4): 180-182.
- Boland, A.M., Jerie, P.H., Mitchell, P.D., Goodwin, I., Connor, D.J. (2000). Long-term effects of restricted root volume and regulated deficit irrigation on peach: I. Growth and mineral nutrition. *J. Amer. Soc. Hort. Sci*. 125 (1): 135-142.
- Brien, E.E. and Brown, J.S. (2008). Games roots play: effects of soil volume and nutrients. *Journal of Ecology*. 96: 438-446.

- Broschat, T.K. and Klock-Moore K.A. (2000). Root and shoot growth responses to phosphate fertilization in container-grown plants. *HortTechnology*. 10(4): 765-767.
- Brouwer, R. (1963). Some aspects of the equilibrium between overground and underground plant parts. *Meeded. Inst. Biol. Scheikd. Onderzoek Landbouwgewassen*. 213: 31-39.
- Bussell, W.T. and Mckennie, S. (2004). Rockwool in horticulture, and its importance and sustainable use in New Zealand. *New Zealand Journal of Crop and Horticultural Science*. 32: 29-37.
- Calori, A.H., Factor, T.L., Junior, S.L., Moraes, L.A.S., Barbosa, P.J.R., Tivelli, S.W., Purquerio, L.F.V. (2014). Electrical conductivity and plant spacing on baby leaf table beet and lettuce production. *Horticultura Brasileira*. 32: 426-433.
- Campostrini, E. and Yamanishi, O.K. (2001). Influence of mechanical root restriction on gas-exchange of four papaya genotypes. *Revista Brasileira de Fisiologia Vegetal*. 13(2): 129-138.
- Campostrini, E., Yamanishi, O.K., Maldonado, J.S., Marin, S.L.D. (2002). Influence of root restriction on chlorophyll and carotenoids concentrations in leaves of four papaya (*Carica papaya* L.) genotypes. *Agronomia*. 36: 1-6.
- Cantliffe, D. J. (1993). Pre - and postharvest practices for improved vegetable transplant quality. *HortTechnology*. 3(4): 415-418.
- Cantliffe, D. J., Paranjpe, A.V., Stoffella, P.J., Lamb, E.M., Powell, C.A. (2007). Influence of soilless media, growing containers, and plug transplants on vegetative growth and fruit yield of 'sweet charlie' strawberry grown under protected culture. *Proc. Fla. State Hort. Soc.* 120: 142-150.
- Carlile, W.R. (1997). The requirement of growing media. In peat in Horticulture- Its use and sustainability. *Proc. International Peat Conference*, ed. G. Schmilewski, pp. 17-23. Amsterdam.
- Carlson, W.C. (1986). Root system considerations in the quality of loblolly pine seedlings. *Southern Journal of Applied Forestry*. 10(2): 87-92.
- Carmi, A. (1986). Effects of root zone volume and plant density on the vegetative and reproductive development of cotton. *Field Crops Research*. 13: 25-32.
- Carmi, A. (1993). Effects of root zone restriction on amino acid status and bean plant growth. *J. Exp. Bot.* 44: 1161-1166.

- Carmi, A. and Heuer, B. (1981). The role of roots in control of bean shoot growth. *Ann. Bot.* 48: 519-527.
- Carmi, A., Hesketh, J.D., Enos, W.T., Peters, D.B. (1983). Interrelationships between shoot growth and photosynthesis as affected by root growth restriction. *Photosynthetica.* 17: 240-245.
- Carmi, A., Plaut, Z., Heuer, B., Grava, A. (1992). Establishment of shallow and restricted root systems in cotton and its impact on plant response to irrigation. *Irrigation Science.* 13: 87-91.
- Cekic, F.O., Unyayar, S., Ortas, I. (2012). Effects of arbuscular mycorrhizal inoculation on biochemical parameters in *Capsicum annuum* grown under long term salt stress. *Turkish Journal of Botany.* 36: 63-72.
- Chirino, E., Vilagrosa, A., Hernandez, E.I., Matos, A., Vallejo, V.R. (2008). Effects of a deep container on morpho-functional characteristics and root colonization in *Quercus suber* L. seedlings for reforestation in Mediterranean climate. *Forest Ecology and Management.* 256: 779-785.
- Choi, J.H., Chung, G.C., Suh, S.R., Yu, J.A., Sung, J.H., Choi, K.J. (1997). Suppression of calcium transport to shoots by root restriction in tomato plants. *Plant Cell Physiol.* 38(4): 495-498.
- Chong, C.W. and Chu, L.M. (2007). Growth of vetivergrass for cutslope landscaping: Effects of container size and watering rate. *Urban Forestry & Urban Greening.* 6: 135-141.
- Clark, R.B. and Zeto, S.K. (2000). Mineral acquisition by arbuscular mycorrhizal plants. *Journal of Plant Nutrition.* 23: 867-902.
- Cooper, A.J. (1979). *The ABC of NFT.* London: Grower Books.
- Dakora, F.D. (2003). Defining new roles for plant and rhizobial molecules in sole and mixed plant cultures involving symbiotic legumes. *New Phytologist.* 158: 39-49.
- Davies, F.T. Olalde-Portugal, V., Aguilera-Gomez, L., Alvarado, M.J., Ferrera-Cerrato, R.C., Boutton, T.W. (2002). Alleviation of drought stress of Chile ancho pepper (*Capsicum annuum* L. cv. San Luis) with arbuscular mycorrhiza indigenous to Mexico. *Scientia Horticulturae.* 92: 347-359.
- Delatorre, J., Pinto, M., Cardemil, L. (2008). Effects of water stress and high temperature on photosynthetic rates of two species of *Prosopis*. *Journal of Photochemistry and Photobiology B: Biology.* 92: 67-76.

- Demir, S. (2004). Influence of arbuscular mycorrhiza on some physiological growth parameters of pepper. *Turk. J. Biol.* 28: 85-90.
- Department of Agriculture. (2016). Statistik Tanaman Sayur-sayuran dan Tanaman Ladang) 2016. Accessed on 25 June 2017 at www.doa.gov.my. pp: 145-146.
- Department of Statistics, Malaysia. (2015). Supply and utilization accounts selected agricultural commodities, 2010-2014. Accessed on 25 June 2017 at www.statistic.gov.my. pp: 34.
- Dolph, C.A. and Proebstring, E.L. (1989) Root confinement affects growth and physiology of sweet cherry and apple. ASHS Annual Meeting, Tulsa, Oklahoma, Prog. & Abstract. pp: 99.
- Dubik, S.P., Krizek, D.T., Stimart, D.P. (1990). Influence of root zone restriction on mineral element concentration, water potential, chlorophyll concentration, and partitioning of assimilate in spreading euonymus (*Euonymus kiautschovica* Loes. 'Sieboldiana'). *Journal of Plant Nutrition.* 13(6): 677-699.
- Dubois, M. , Gilles, K. A., Hamilton, J. K., Rebers, P. A., and Smith, F. (1956) Colorimetric method for determination of sugars and related substances. *Anal. Chem.*, 1956, 28 (3): 350–356.
- Elmstrom, G.W. (1973). Watermelon root development affected by direct seeding and transplanting. *HortScience.* 8: 134-136.
- Engindeniz, S. and Gul, A. (2009). Economic analysis of soilless and soil-based greenhouse cucumber production in Turkey. *Sci. Agric. (Piracicaba, Braz.)*. 66(5): 606-614.
- Eshel, A., Rao, C.S., Benzioni, A., Waisel, Y. (2001). Allometric relationships in young seedlings of faba bean (*Vicia faba* L.) following removal of certain root types. *Plant and Soil.* 233: 161-166.
- Falovo, C., Roupael, Y., Rea, E., Battistelli, A., Colla, G. (2009). Nutrient solution concentration and growing season affect yield and quality of *Lactuca sativa* L. var. acephala in floating raft culture. *J. Sci. Food Agric.* 89: 1682-1689.
- Fandi, M., Al- Muhtaseb, J. A., Hussein, M. A. (2008). Yield and fruit quality of tomato as affected by the substrate in an open soilless culture. *Jordan Journal of Agricultural Sciences.* 4: 65-72.
- Fenneman, Dan, Michael Sweat, George Hochmuth, and Robert Hochmuth. 2012. Production Systems—Florida Greenhouse Vegetable Production

Handbook, Vol 3. University of Florida, University of Florida/Institute of Food and Agricultural Sciences Extension Service: Gainesville, Florida. Publication HS785. <http://edis.ifas.ufl.edu>.

- Ferdousee, N., Jabbar, F., Hossain, M.K., Rafiqul Hoque, T.M. (2010). Comparative growth performance of *Leucaena leucocephala* seedlings raised in nursery bed, polybag and root trainers. Proc. of International Conference on Environmental Aspects of Bangladesh. 10: 65-68.
- Flexas, J. and Medrano, H. (2002). Drought-inhibition of photosynthesis in C3 plants: Stomatal and non-stomatal limitations revisited. *Annals of Botany*. 89: 183-189.
- Franco, J.A., Banon, S., Vicente, M.J., Miralles, J., Martinez-Sanchez, J.J. (2011). Root development in horticultural plants grown under abiotic stress conditions- a review. *Journal of Horticultural Science and Biotechnology*. 86 (6): 543-556.
- Geigenberger, P. (2003). Response of plant metabolism to too little oxygen. *Current Opinion in Plant Biology*. 6: 247–256.
- Goto, T., Matsuno, T., Yoshida, Y., Kageyama, Y. (2002). Photosynthetic, evapotranspiratory and leaf morphological properties of chrysanthemum grown under root restriction as affected by fertigation frequency. *J. Jpn. Soc. Hortic. Sci.* 71: 277–283.
- Grafiadellis, I., Mattas, K., Maloupa, E., Tzouramani, I., Galanopoulos, K. (2000). An economic analysis of soilless culture in gerbera production. *Hortscience*. 35(2):300-303.
- Gruda, N. (2009). Do soilless culture systems have an influence on product quality of vegetables?. *Journal of Applied Botany and Food Quality*. 82: 141 – 147.
- Guang-cheng, S., Rui-qi, G., Na, L., Shuang-en, Y., Weng-gang, X. (2011). Photosynthetic, chlorophyll fluorescence and growth changes in hot pepper under deficit irrigation and partial root zone drying. *African Journal of Agricultural Research*. 6(19): 4671-4679.
- Guerfel, M., Baccouri, O., Boujnah, D., Chaibi, W., Zarrouk, M. (2009). Impacts of water stress on gas exchange, water relations, chlorophyll content and leaf structure in the two main Tunisian olive (*Olea europaea* L.) cultivars. *Scientia Horticulturae*. 119: 257-263.

- Haldankar, P.M., Parulekar, Y.R., Kulkarni, M.M., Lawande, K.E. (2014). Effect of size of polybag on survival and growth of mango grafts. *Journal of Plant Studies*. 3(1): 91-95.
- Hameed, M.A. (1988). The effects of root confinement on the growth, assimilate partitioning, mineral nutrition, nitrogen assimilation and water relations of tomato plants (*Lycopersicon esculentum* Mill.) Ph.D Thesis, Lincoln University, Lincoln, New Zealand.
- Hameed, M.A., Reid, J.B., Rowe, R.N. (1987). Root confinement and its effects on the water relations, growth and assimilate partitioning of tomato (*Lycopersicon esculentum* Mill.). *Annals of Botany*. 59: 685-692.
- Hannan, J.J. (1998). *Greenhouse: Advanced technology for protected Horticulture* (pp. 661). Washington, USA: CRC Press.
- Hazzoumi, Z., Moustakime, Y., Elharchli, E-H, Joutei, K. A. (2015). Effect of arbuscular mycorrhizal fungi (AMF) and water stress on growth, phenolic compounds, glandular hairs, and yield of essential oil in basil (*Ocimum gratissimum* L). *Chemical and Biological Technologies in Agriculture*. 2: 10.
- Hess, L. and Kroon, H.D. (2007). Effects of rooting volume and nutrient availability as an alternative explanation for root self/non-self discrimination. *Journal of Ecology*. 95: 241-251.
- Heuvelink, E. and Buiscool, R.P.M. (1995). Influence of sink-source interaction on dry matter production in tomato. *Annals of Botany*. 75: 381-389.
- Hildebrandt, U., Kaldorf, M., Bothe, H. (1999). The zinc violet and its colonization by arbuscular mycorrhizal fungi. *Plant Physiol*. 154: 709-717.
- Hunt, R. (1978). *Plant growth analysis*. London: Edward Alnord.
- Hunt, R. (1982). *Plant growth curves: the functional approach to plant growth analysis*. London: Edward Arnold.
- Hurley, M.B. and Rowarth, J.S. (1999). Resistance to root growth and changes in the concentrations of ABA within the root and xylem sap during root-restriction stress. *J. Exp. Bot*. 50: 799-804.
- Hussain, A., Iqbal, K., Aziem, S., Mahato, P., Negi, A.K. (2014). A review on the science of growing crops without soil (soilless culture) – a novel alternative for growing crops. *International Journal of Agriculture and Crop Sciences*. 7(11): 833-842.

- Iersel, V.M. (1997). Root restriction effects on growth and development of salvia (*Salvia splendens*). HortScience. 32(7): 1186-1190.
- Ikiz, O., Abak, K., Dasgan, H.Y., Ortas, I. (2009). Effects of mycorrhizal inoculation in soilless culture on pepper plant growth. Acta Hort. 807: 533-540.
- Ismail, M.R. (2000). Plant Nutrition in Hydroponics. In *Hydroponics production in tropics*. Jofri Print. Sdn. Bhd.
- Ismail, M.R. and Dalia, S. (1995). Growth, physiological processes and yield of tomato grown in different root zone volumes using sand culture. Pertanika Journal Tropical Agriculture Science, 18: 141-147.
- Ismail, M.R. and Davies, W.J. (1998). Root restriction affects leaf growth and stomatal response: the role of xylem sap ABA. Scientia Horticultura. 74: 257-268.
- Ismail, M.R. and Noor, K.M. (1996). Growth, water relations and physiological processes of starfruit (*Averrhoa carambola* L) plants under root growth restriction.
- Ismail, M.R., Sze, L.Y., Poulus, P., Ibrahim, H. (2004). The use of empty oil palm bunch (EFB) compost as additive in coconut dust soilless system for vegetable crop production. Proceedings of the international symposium on growing media and hydroponics, Alnarp, Sweden, (Alsanius, B., Jensen, P., and Asp, H., eds.). Acta Hort 644: 193-198.
- Ismail, S.M., Ozawa, K., Khondaker, N.A. (2008). Influence of single and multiple water application timings on yield and water use efficiency in tomato (variety First power). Agricultural Water Management. 95: 116-122.
- Jaenicke, H. (1999). *Good tree nursery practices - Practical guidelines for community nurseries* (pp. 23). Nairobi, Kenya: Majestic Printing Work.
- Jayasinghe, G. Y. (2014). Utilization of agricultural waste compost as an alternative potting media component with coir dust for leafy vegetable *Ipomoea aquatica*. Journal of Plant Nutrition. 37: 1601-1611.
- Jensen, M.H. (1997). Hydroponics. HortScience. 32(6): 1018-1021.
- Jovicich, E., Cantliffe, D.J., Sargent, S.A., Osborne, L.S. (2004). Production of greenhouse-grown peppers in Florida. University of Florida. Institute of Food and Agricultural Sciences Department of Horticultural Sciences Extension Service. Publication HS979. <http://edis.ifas.ufl.edu>.

- Judd, L.A., Jackson, B.E., Fonteno, W.C. (2015). Advancements in root growth measurement technologies and observation capabilities for container-grown plants. *Plants*. 4: 369-392.
- Kagi, B.F. (1998). Vigour control in grapevines for wine production: The effects of root restriction on the vegetative growth, fruitfulness and fruit maturity of *Vitis vinifera* cv. Cabernet sauvignon. Master Thesis, Lincoln University, Lincoln, New Zealand.
- Kalaji, H.M., Govindjee, Bosa, K., Koscielniak, J., Zuk-Golaszewska, K. (2011). Effects of salt stress on photosystem II efficiency and CO₂ assimilation of two Syrian barley landraces. *Environmental and Experimental Botany*. 73: 64-72.
- Kang J.G. and Chon, S.U. (2010). Combined effects of container volume and fertilizer level on plant growth, physiological characteristics and nutrient uptake of vinca plant (*Catharanthus roseus*). *Korean J. Crop Sci.* 55(3): 268-274.
- Karaki, G. N. (2000). Growth of mycorrhizal tomato and mineral acquisition under salt stress. *Mycorrhiza*. 10: 51–54.
- Karimi, F., Arunkumar, B., Asif, M., Murthy, B.N.S., Venkatesha, K.T. (2013). Effect of different soilless culture systems on growth, yield and quality of strawberry cv. Strawberry Festival. *International Journal of Agricultural Sciences*. 9(1): 366-372.
- Karni, L., Aloni, B., Bar-Tal, A., Moreshet, S., Keinan, M., Yao, C. (2000). The effect of root restriction on the incidence of blossom-end rot in bell-pepper (*Capsicum annuum* L.). *Journal Hort. Sci. and Biotech.*, 75: 364-369.
- Kasai, M., Koide, K., Ichikawa, Y. (2012). Effect of pot size on various characteristics related to photosynthetic matter production in soybean plants. *International Journal of Agronomy*. Article ID 751731, 7 pages. doi:10.1155/2012/751731.
- Keever, G.J., Cobb, G.S., Reed, R.B. (1985). Effect of container dimension and volume on growth of three woody ornamentals. *Hortscience*, 20: 276-278.
- Kharkina, T.G., Ottosen, C.O., Rosenqvist, E. (1999). Effect of root restriction on growth and physiology of cucumber plants. *Physiologia Plantarum*, 105: 434-441.
- Klaring, H.P. (2001). Strategies to control water and nutrient supplies to greenhouse crops. A review. *Agronomie*. 21: 311-321.

- Korner, C., Menendezriedl, S.P., John, P.C.L. (1989). Why are bonsai plants small? A consideration of cell size. *Australian Journal of Plant Physiology*. 16(5): 443-448.
- Kratky, B.A., Wang, J.K., Kubojiri, K. (1982). Effect of container size, transplant age, and plant spacing on Chinese cabbage. *Journal of the American Society for Horticultural Science*. 107: 345-347.
- Kreij, C.D. and Van Den Berg, TH.J.M (1990). Nutrient uptake, production and quality of *Rosa hybrida* in rockwool as affected by electrical conductivity of the nutrient solution. In: M.L. van Beusichem (Ed.), *Plant Nutrition - Physiology and Applications. Developments in Plant and Soil Sciences* (pp. 519-523). Dordrecht: Springer.
- Krikun, J., Haas, J.H., Bar-Yosef, B. (1987). Use of VA mycorrhizal-fungus inoculum in soils in arid and semi-arid climates: a field study with bell pepper and transplants. *Angew. Bot.*, 61: 97-105.
- Krizek, D.T. and Dubik, S.P. (1987). Influence of water stress and restricted root volume on growth and development of urban trees. *Journal of Arboriculture*, 13(2): 47-55.
- Krizek, D.T., Carmi, A., Mirecki, R.M., Snyder, F.W., Bunce, J.A. (1985). Comparative effects of soil moisture stress and restricted root zone volume on morphogenetic and physiological responses of soybean (*Glycine max* L. Merr). *Journal of Experimental Botany*. 36 (162): 25-38.
- Kurunc, A. and Unlukara, A. (2010). Growth, yield, and water use of okra (*Abelmoschus esculentus*) and eggplant (*Solanum melongena*) as influenced by rooting volume. *New Zealand Journal of Crop and Horticultural Science*. 37(3): 201-210.
- Latimer, J.G. (1991). Container size and shape influence growth and landscape performance of marigold seedlings. *HortScience*. 26(2): 124-126.
- Lawlor, D.W. and Cornic, G. (2002). Photosynthetic carbon assimilation and associated metabolism in relation to water deficits in higher plants. *Plant, Cell and Environment*. 25: 275-294.
- Leiva, M.J. and Fernandez-Ales, R. (1998). Variability in seedling water status during drought within a *Quercus ilex* subsp. *ballota* population, and its relation to seedling morphology. *Forest Ecology and Management*. 111: 147-156.

- Liu, A. and Latimer, J.G. (1995). Water relations and abscisic acid levels of watermelon as affected by rooting volume restriction. *Journal of Experimental Botany*. 46: 1011-1015.
- Liu, T., Sheng, M., Wang, C.Y., Chen, H., Li, Z., Tang, M. (2015). Impact of arbuscular mycorrhizal fungi on the growth, water status, and photosynthesis of hybrid poplar under drought stress and recovery. *Photosynthetica*. 53(2): 250-258.
- Long, R.L., Walsh, K.B., Rogers, G., Midmore, D.J. (2004). Source-sink manipulation to increase melon (*Cucumis melo* L.) fruit biomass and soluble sugar content. *Australian Journal of Agricultural Research*. 55: 1241-1251.
- Lorenzo, R.D., Pisciotta, A., Santamaria, P., Scariot, V. (2013). From soil to soil-less in horticulture: quality and typicity. *Italian Journal of Agronomy*. 8(30): 255-260.
- Luo, H.H., Tao, X. P., Hu, Y.Y., Zhang, Y. L., Zhang, W. F. (2015). Response of cotton root growth and yield to root restriction under various water and nitrogen regimes. *J. Plant Nutr. Soil Sci*. 178: 384–392.
- MacKay, B.R. (1995). Root restriction and root-shoot relationships in tomato (*Lycopersicon esculentum* Mill.). Ph.D Thesis, Massey University, New Zealand.
- Mafakheri, A., Siosemardeh, A., Bahramnejad, B., Struik, P.C., Sohrabi, Y. (2010). Effects of drought stress on yield, proline and chlorophyll contents in three chickpea cultivars. *Australian Journal of Crop Science*. 4(8): 580-585.
- Mandre, O., Rieger, M., Myers, S.C., Seversen, R., Regnard, J.L. (1995). Interaction of root confinement and fruiting in peach. *J. American Society of Horticultural Science*. 120(2): 228-234.
- Marihal, A.K., Pradeep, S.M., Jagadeesh, K.S. (2011). Effect of commercial mycorrhizal inoculant on growth and yield of chilli (*Capsicum annum* L.) in field conditions. *Karnataka J. Agric. Sci*. 24(4): 589-590.
- Markham III, J.W., Bremer, D.J., Boyer, C.R., Schroeder, K.R. (2011). Effect of container color on substrate temperatures and growth of red maple and redbud. *HortScience*. 46(5): 721–726.
- Marschner, H. and Dell, B. (1994). Nutrient uptake in mycorrhizal symbiosis. *Plant and Soil*. 159: 89-102.

- Massetani, F., Savini, G., Neri, D. (2017). Effect of substrate and container type in the strawberry soilless cultivation. *Acta Horticulturae*. 1156: 295-300.
- Mataa, M. and Tominaga, S. (1998). Effects of root restriction on tree development in Ponkan mandarin (*Citrus reticulata* Blanco). *J. Amer. Soc. Hort. Sci.* 123(4): 651-655.
- Mathers, H.M., Lowe, S.B., Scagel, C., Struve, D.K., Case, L.T. (2007). Abiotic factors influencing root growth of woody nursery plants in containers. *HortTechnology*. 17(2): 151-162.
- Maynard, E.T., Vavrina, C.S., Scott, W.D. (1996). Containerized muskmelon transplants: Cell volume effects on pretransplant development and subsequent yield. *HortScience*. 31(1): 58-61.
- McConnaughay, K.D.M. and Bazzaz, F.A. (1991). Is physical space a soil resource?. *Ecology*. 72(1): 94-103.
- Mena-Violante, H.G., Ocampo-Jiménez, O., Dendooven L., Martinez-Soto, G., Gonzalez-Castaneda, J., Davies Jr., F.T., Olalde-Portugal, V. (2006). Arbuscular mycorrhizal fungi enhance fruit growth and quality of chile ancho (*Capsicum annuum* L. cv San Luis) plants exposed to drought. *Mycorrhiza*. 16: 261-267.
- Miransari, M. (2010). Contribution of arbuscular mycorrhizal symbiosis to plant growth under different types of soil stress. *Plant Biology*. 12: 563–569.
- Miransari, M. (2013). Corn (*Zea mays* L.) growth as affected by soil compaction and arbuscular mycorrhizal fungi. *Journal of Plant Nutrition*. 36(12): 1853-1867.
- Miransari, M., Bahrami, H.A., Rejali, F., Malakouti M.J. (2008). Using arbuscular mycorrhiza to alleviate the stress of soil compaction on wheat (*Triticum aestivum* L.) growth. *Soil Biology and Biochemistry*. 40: 1197–1206.
- Miransari, M., Bahrami, H.A., Rejali, F., Malakouti, M.J., Torabi, H. (2007). Using arbuscular mycorrhiza to reduce the stressful effects of soil compaction on corn (*Zea mays* L.) growth. *Soil Biology and Biochemistry*. 39(8): 2014–2026.
- Mobini, S.H., Ismail, M.R., Arouiee, H. (2009). Influence of ventilation and media on potato (*Solanum tuberosum* L.) tuberization and its growth characteristics. *African Journal of Biotechnology*. 8(10): 2232-2241.
- Mohd Razi, I. (1994). *Pengeluaran Tanaman Hidroponik*. Kuala Lumpur: Dewan Bahasa dan Pustaka.

- Mokhtari, S., Ismail, M.R., Kausar, H., Musa, M.H., Wahab, P.E.M., Berahim, Z., Omar, M.H., Habib, S.H. (2013). Use of organic enrichment as additives in coconut coir dust on development of tomato in soilless culture. *Compost Science and Utilization*. 21(1): 16-21.
- Molahoseini, H., Sepahvand, H., Banaei, A., Abdolbaghi, M. (2013). Effects of different nutrient solution concentrations on yield and cut flower quality of gerbera grown in soilless culture. *International Journal of Traditional and Herbal Medicine*. 1(3): 59-64.
- Morano, G., Amalfitano, C., Sellitto, M., Cuciniello, A., Maiello, R., Caruso, G. (2017). Effects of nutritive solution electrical conductivity and plant density on growth, yield and quality of sweet basil grown in gullies by subirrigation. *Adv. Hort. Sci.* 31(1): 25-30.
- Murphy, G.P., File, A.L., Dudley, S.A. (2013). Differentiating the effects of pot size and nutrient availability on plant biomass and allocation. *Botany*. 91(11): 799-803.
- Myers, S.C. (1992). Root restriction of apple and peach with in-ground fabric containers. *Acta Horticulturae*. 322: 215-220.
- Nadian, H., Smith, S.E., Alston, A.M., Murray, R.S. (1997). Effects of soil compaction on plant growth, phosphorus uptake and morphological characteristics of vesicular-arbuscular mycorrhizal colonization of *Trifolium subterraneum*. *New Phytol.* 135: 303-311.
- Navindra, B., Gianjeet, D., Joyce, G-S. (2011). Influence of soilless growing media, pot size and sieved media on the production of *Hibiscus sabdariffa* L. seedlings. *Australian Journal of Agricultural Engineering*. 2(5): 147-154.
- Nazari, F., Farahmand, H., Khosh-Kui, M., Salehi, H. (2011). Effects of coir as a component of potting media on growth, flowering and physiological characteristics of hyacinth (*Hyacinthus orientalis* L. cv. Sonbol-e-Irani). *International Journal of Agricultural and Food Science*, 1(2): 34-35.
- Neales, T.F. and Incoll, L.D. (1968). The control of leaf photosynthesis rate by the level of assimilate concentration in the leaf: A review of hypothesis-*Bot. Rev.* 34: 107-125.
- Nejad, A.R. and Ismaili, A. (2014). Changes in growth, essential oil yield and composition of geranium (*Pelargonium graveolens* L.) as affected by growing media. *J. Sci. Food Agric.* 94: 905–910.
- Nemali, K.S. and Iersel, V.M.W. (2004). Light intensity and fertilizer concentration: II. Optimal fertilizer solution concentration for species

- differing in light requirement and growth rate. HortScience. 39(6): 1293-1297.
- NeSmith, D.S., Bridges, D.C., Barbour, J.C. (1992). Bell pepper responses to root restriction. J. Plant Nutr. 15: 2763-2776.
- NeSmith, D.S. (1993). Summer squash response to root restriction under different light regimes. Journal of Plant Nutrition. 16(5): 765-780.
- NeSmith, D.S. and Duval, J.R. (1998). The effect of container size. HortTechnology October-December. 8(4).
- Neuman, N.S. (1993). Shoot responses to root stress - A resource gathering point of view. Journal of Arboriculture. 19(2): 118-123.
- Nicola, S., Hoeberechts, J., Fontana, E. (2005). Comparison between traditional and soilless culture systems to produce rocket (*Eruca sativa*) with low nitrate content. Acta Hort. 697: 549-555.
- Niinemets, U. (2002). Stomatal conductance alone does not explain the decline in foliar photosynthetic rates with increasing tree age and size in *Picea abies* and *Pinus sylvestris*. Tree Physiology. 22: 515-535.
- Nishizawa, T. and Saito, K. (1998). Effects of rooting volume restriction on the growth and carbohydrate concentration in tomato plants. J. Amer. Soc. Hort. Sci. 123(4): 581-585.
- Oddiraju, V.G., Beyl, C.A., Barker, P.A., Stutte, G.W. (1994). Container size alters root growth of western black cherry as measured via image analysis. HortScience. 29(8): 910-913.
- Onguso, J.M., Mizutani, F., Sharif Hossain, A.B.M., El-Shereif, A.R., Rutto, K.L. (2006). Effect of vertical root restriction with corrugated plastic sheets on growth and flower bud formation in young peach trees. Bull. Exp. Farm Fac. Agr. Ehime Univ. 28: 1-7.
- Oseni, T.O., Shongwe, N.S., Masarirambi, M.T. (2010). Effect of arbuscular mycorrhiza (AM) inoculation on the performance of tomato nursery seedlings in vermiculite. International Journal of Agriculture & Biology. 12: 789-792.
- Pardossi, A., Carmassi, G., Diara C., Incrocci, L., Maggini, R., Massa, D. (2011). Fertigation and substrate management in closed soilless culture.
- Park, S-Y., Oh, S-B., Kim, S-M., Cho, Y-Y., Oh, M-M. (2016). Evaluating the effects of a newly developed nutrient solution on growth, antioxidants, and

- chicoric acid contents in *Crepidiastrum denticulatum*. Hortic. Environ. Biotechnol. 57(5): 478-486.
- Paul, M.J. and Pellny, T.K. (2003) Carbon metabolite feedback regulation of leaf photosynthesis and development. Journal of Experimental Botany. 54: 539–547.
- Perucka, I. and Materska, M. (2007). Antioxidant vitamin contents of *Capsicum annuum* fruit extracts as affected by processing and varietal factors. Acta Sci. Pol. Technol. Aliment. 6(4): 67-74.
- Peterson, T.A., Reinsel, M.D., Krizek, D.T. (1991a). Tomato (*Lycopersicon esculentum* Mill cv Better Bush) plant response to root restriction. I. Alteration of plant morphology. J. Exp. Bot. 42: 1233-1240.
- Peterson, T.A., Reinsel, M.D., Krizek, D.T. (1991b). Tomato (*Lycopersicon esculentum* Mill cv. Better Bush) plant response to root restriction. II. Root respiration and ethylene generation. J. Exp. Bot. 42: 1241-1249.
- Pezeshki, S.R. and Santos, M.I. (1998). Relationships among rhizosphere oxygen deficiency, root restriction, photosynthesis, and growth in baldcypress (*Taxodium distichum* L.) seedlings. Photosynthetica. 35(3): 381-390.
- Pigeaire, A., Pate, J.S., Atkins, C.A. (1990). Contrasting effects of rooting volume and aerial spacing on shoot morphology and reproductive performance of *Lupinus angustifolius* L cultivar danja. Australian Journal of Agricultural Research. 41(2): 339-350.
- Pires, R.C.M., Furlani P.R., Ribeiro, R.V., Junior, D.B., Sakai, E., Lourencao, A.L., Neto, A.T. (2011). Irrigation frequency and substrate volume effects in the growth and yield of tomato plants under greenhouse conditions. Scientia Agricola. 68 (4): 400-405.
- Poorter, H., Buhler, J., Dusschoten, D.V., Climent, J. (2012b). Pot size matters: a meta-analysis of the effects of rooting volume on plant growth. Functional Plant Biology. 39: 839–850.
- Poorter, H., Fiorani, F., Stitt, M., Schurr, U., Finck, A., Gibon, Y., Usadel, B., Munns, R., Atkin, O.K., Tardieu, F., Pons, T.L. (2012a). The art of growing plants for experimental purposes: a practical guide for the plant biologist. Functional Plant Biology. 39: 821–838.
- Potter, J.R. and Jones, J.W. (1977). Leaf area partitioning as an important factor in growth. Plant Physiology. 59: 10-14.

- Rahimi, Z., Aboutalebi, A., Hasanzadeh, H. (2013). Effect of various culture media on tomato transplant production. *International Research Journal of Applied and Basic Sciences*. 4(2): 326-328.
- Rakow, D.A. (1987). Containerised trees in urban environments. *Journal of Arboriculture*. 13 (12): 294-298.
- Ramanjulu, S., Sreenivasalu, N., Kumar, S.G., Sudhakar, C. (1998). Photosynthetic characteristics in mulberry during water stress and rewatering. *Photosynthetica*. 35(2): 259-263.
- Ran, Y., Bar-Yosef, B., Erez, A. (1992). Root volume influence on dry matter production and partitioning as related to nitrogen and water uptake rates by peach trees. *Journal of Plant Nutrition*. 15: 713-726.
- Rapparini, F. and Penuelas, J. (2014). *Use of microbes for the alleviation of soil stresses*. (pp. 21-42). New York: Springer.
- Raviv, M. and Lieth, J.H. (2008). *Soilless Culture Theory and Practices*. London: Elsevier.
- Richards, D. (1986). Tree growth and productivity- the role of the roots. *Acta Horticulturae*. 175: 27-35.
- Richards, D. and Rowe, R.N. (1977). Effects of root restriction, root pruning and 6-benzylaminopurine on the growth of peach seedlings. *Ann. Bot.* 41: 729-740.
- Rieger, M. and Marra, F. (1994). Responses of young peach trees to root confinement. *Journal of American Society for Horticultural Science*. 119(2): 223-228.
- Robbins, N. S. and Pharr, D.M. (1988). Effect of restricted root growth on carbohydrate metabolism and whole plant growth of *Cucumis sativus* L. *Plant Physiology*. 87: 409-413.
- Rodriguez, J.C., Cantliffe, D.J., Shaw, N.L., Karchi, Z. (2006). Soilless media and containers for greenhouse production of Galia type muskmelon. *Hortscience*. 41(5): 1200-1205.
- Ronchi, C.P., DaMatta, F.M., Batista, K.D., Moraes, G.A.B.K., Loureiro, M.E., Ducatti, C. (2006). Growth and photosynthetic down-regulation in *Coffea arabica* in response to restricted root zone volume. *Functional Plant Biology*. 33: 1013-1023.

- Roper, T.R., Keller, J.D., Loescher, W.H., Rom, C.R. (1988). Photosynthesis and carbohydrate partitioning in sweet cherry: Fruiting effects. *Physiologia Plantarum*. 72(1): 42-47.
- Rosadi, R.A.B., Senge, M., Suhandy, D., Tusi, A. (2014). The effect of EC levels of nutrient solution on the growth, yield, and quality of tomatoes (*Solanum lycopersicum*) under the hydroponic system. *Journal of Agricultural Engineering and Biotechnology*. 2(1): 7-12.
- Ruff, M.S., Krizek, R.M., Inouye, D.W. (1987). Restricted root volume: Influence on growth and development of tomato. *J. Amer. Soc. Hort. Sci.* 112: 763-769.
- Saito, S.T., Fukuda, N., Iikubo, T., Inai, S., Fujii, T., Konishi, C., Ezura, H. (2008). *J. Japan. Soc. Hort. Sci.* 77 (2): 165–172.
- Samarakoon, U.C., Weerasinghe, P.A., Weerakkody, W.A.P. (2006). Effect of electrical conductivity (EC) of the nutrient solution on nutrient uptake, growth and yield of leaf lettuce (*Lactuca sativa* L.) in stationary culture. *Tropical Agricultural Research*. 18: 13-21.
- Sarooshi, R.A. and Cresswell, G.C. (1994). Effects of hydroponic solution composition, electrical conductivity and plant spacing on yield and quality of strawberries. *Australian Journal of Experimental Agriculture*. 34: 529-35.
- SAS Institute Inc. (2004). SAS user's guide statistics, version 9 SAS Institute, Cary, NC.
- Sato, F., Yoshioka, H., Fujiwara, T. (2001). Effect of root volume restriction on growth and carbohydrate status of cabbage seedlings. *Environ. Control in Biol.* 39(4): 235-243.
- Schaffer, B., Whiley, A.W., Searle, C., Nissen, R.J. (1997). Leaf gas exchange, dry matter partitioning, and mineral element concentrations in mango as influenced by elevated atmospheric carbon dioxide and root restriction. *Journal of American Society for Horticultural Science*. 122(6): 849-855.
- Schwarz, D., Klaring, H.P., Iersel, V.M.W., Ingram, K.T. (2002). Growth and photosynthetic response of tomato to nutrient solution concentration at two light levels. *J. Amer. Soc. Hort. Sci.* 127(6): 984-990.
- Selvakumar, G. and Thamizhiniyan, P. (2011). The effect of the arbuscular mycorrhizal (am) fungus *Glomus intraradices* on the growth and yield of chilli (*Capsicum annuum* L.) under salinity stress. *World Applied Sciences Journal*. 14(8): 1209-1214.

- Sengupta, A. and Banerjee, H. (2012). Soilless culture in modern agriculture. *World Journal of Science and Technology* 2012, 2(7): 103-108.
- Sensoy, S., Demir, S., Turkmen, O., Erdinc, C., Savur, O.B. (2007). Responses of some different pepper (*Capsicum annuum* L.) genotypes to inoculation with two different arbuscular mycorrhizal fungi. *Scientia Horticulturae*. 113: 92–95.
- Shaha, R.K., Rahman, S., Asrul, A. (2013). Bioactive compounds in chilli peppers (*Capsicum annuum* L.) at various ripening (green, yellow and red) stages. *Annals of Biological Research*. 4(8): 27-34.
- Sharkey, T.D. (1985). O₂-insensitive photosynthesis in C₃ plants. Its occurrence and a possible explanation. *Plant Physiol*. 78: 71–75.
- Shen, J., Li, C., Mi, G., Li, L., Yuan, L., Jiang, R., Zhang, F. (2013). Maximizing root/rhizosphere efficiency to improve crop productivity and nutrient use efficiency in intensive agriculture of China. *Journal of Experimental Botany*, 64(5): 1181–1192.
- Shi, K., Ding, X.T., Dong, D.K., Zhou, Y.H., Yu, J.Q. (2008a). Root restriction-induced limitation to photosynthesis in tomato (*Lycopersicon esculentum* Mill.) leaves. *Scientia Horticulturae*. 117: 197-202.
- Shi, K., Fu, L.J., Dong, D.K., Zhou, Y.H., Yu, J.Q. (2008b). Decreased energy synthesis is partially compensated by a switch to sucrose synthase pathway of sucrose degradation in restricted root of tomato plants. *Plant Physiology and Biochemistry*. 46: 1040-1044.
- Shi, K., Hu, W.H., Dong, D.K., Zhou, Y.H., Yu, J.Q. (2007). Low O₂ supply is involved in the poor growth in root-restricted plants of tomato (*Lycopersicon esculentum* Mill.). *Environmental and Experimental Botany*. 61: 181-189.
- Singh, A. (2003). *Fruit Physiology and Production*. (pp. 88-92). Ludhiana: Kalyani Publishers,
- Siti Zaharah, S. and Razi, I.M. (2009). Growth, stomata aperture, biochemical changes and branch anatomy in mango (*Mangifera indica*) cv. Chokanan in response to root restriction and water stress. *Scientia Horticulturae*. 123(1): 58-67.
- Smith, S.E. and Read, D.J. (1997). *Mycorrhizal symbiosis*. San Diego, California: Academic Press.
- Sosu, G. (2014). Growth of cocoa seedlings as affected by different growth media and different polybag sizes. Master Thesis, University of Ghana.

- Spomer, L.A. (1975). Small soil containers as experimental tools: Soil water relations. *Commun. Soil Sci. Plant Anal.* 6: 121-126.
- Spomer, L.A. (1981). The effect of container soil volume on plant growth. *HortScience.* 17: 680-681.
- Spomer, L.A. (1990). Evaluating 'drainage' in container and other shallow-drained horticultural soils. *Commun. Soil Sci. Plant Anal.* 21(3&4): 221-235.
- Spomer, L.A., Berry, W.L., Tibbitts, T.W. (1997). Plant culture in solid media. In R.W. Langhans, and T.W. Tibbitts (Eds.) *Plant Growth Chamber Handbook* (pp. 105–118). Ames, IA: Iowa State University.
- Subramanian, K.S., Santhanakrishnan, P., Balasubramanian, P. (2006). Responses of field grown tomato plants to arbuscular mycorrhizal fungal colonization under varying intensities of drought stress. *Scientia Horticulturae.* 107: 245–253.
- Suhaimi, M.Y., Mohamad, A.M., Mahamud, S., Khadzir, D. (2012). Effects of substrates on growth and yield of ginger cultivated using soilless culture. *J. Trop. Agric. and Fd. Sc.* 40(2): 159-168.
- Tadesse, T., Nichols, M.A., Fisher, K.J. (1999). Nutrient conductivity effects on sweet pepper plants grown using a nutrient film technique. *New Zealand Journal of Crop and Horticultural Science.* 27(3): 229-237.
- Ternesí, M.A., Andrade, A.P., Jorriñ, J., and Benlloch, M. (1994). Root-shoot signalling in sunflower plants with confined root systems. *Plant Soil* 166: 31-36.
- Tezara, W., Martínez, D., Rengifo, E., Herrera, A. (2003). Photosynthetic responses of the tropical spiny shrub *Lycium nodosum* (Solanaceae) to drought, soil salinity and saline spray. *Annals of Botany.* 92: 757-765.
- Thomas, R.B. and Strain, B.R. (1991). Root restriction as a factor in photosynthetic acclimation of cotton seedlings grown in elevated carbon dioxide. *Plant Physiology.* 96: 627-634.
- Torres-Quezada, E.A., Santos, B.M., Zotarelli, L., Treadwell, D.A. (2015). Soilless media and containers for bell pepper production. *International Journal of Vegetable Science.* 21(2): 177-187.
- Townend, J. and Dickinson, A. L. (1995). A comparison of rooting environment in containers of different sizes. *Plant and Soil.* 175: 139-146.

- Tsakaldimi, M., Zagas, T., Tsitsoni, T., Ganatsas, P. (2005). Root morphology, stem growth and field performance of seedlings of two Mediterranean evergreen oak species raised in different container types. *Plant and Soil*. 278: 85-93.
- Tschaplinski, T. J. and Blake, T.J. (1985). Effects of root restriction on growth correlations, water relations and senescence of alder seedlings. *Physiologia Plantarum*. 64(2): 167-176.
- Tyson, R.V., Hochmuth, R.C., Lamb, E.M., Hochmuth, G.J., Sweat, M.S. (2001). A decade of change in Florida's greenhouse vegetable industry:1991-2001. *Proc. Fla. State Hort. Soc.* 114: 280-283.
- Voogt, W. (1981). "What electrical conductivity (EC) for cucumber in rockwool?". *Groenten en Fruit*. 36(34): 26-27.
- Whiley, A.W., Searle, C., Schaffer, B., Wolstenholme, B.N. (1999). Cool orchard temperatures or growing trees in containers can inhibit leaf gas exchange of avocado and mango. *J. Amer. Soc. Hort. Sci.* 124(1): 46–51.
- Will, R.E. and Teskey, R.O. (1997). Effect of elevated carbon dioxide concentration and root restriction on net photosynthesis, water relations and foliar carbohydrate status of loblolly pine seedlings. *Tree Physiology*. 17(10): 655-661.
- Williamson, J.G. and D.C. Coston. (1990). Planting method and irrigation rate influence vegetative and reproductive growth of peach planted at high density. *J. Amer. Soc. Hort. Sci.* 115: 207-212.
- Wira, A.B., Mohd Razi, I., Abd Jamil, Z. (2011). Composts as additive in coconut coir dust culture for growing rockmelon (*Cucumis melo* L.). *J. Trop. Agric. and Fd. Sc.* 39(2): 229-237.
- Wu, H-H., Zou, Y-N., Rahman, M.M., Ni, Q-D., Wu, Q-S. (2017). Mycorrhizas alter sucrose and proline metabolism in trifoliolate orange exposed to drought stress. *Scientific Reports*. 7: 42389.
- Wu, M. and Kubota, C. (2008). Effects of electrical conductivity of hydroponic nutrient solution on leaf gas exchange of five greenhouse tomato cultivars. *HortTechnology*. 18(2): 271-277.
- Wu, Q-S., Xia, R-X., Zou, Y-N. (2008). Improved soil structure and citrus growth after inoculation with three arbuscular mycorrhizal fungi under drought stress. *European Journal of Soil Biology*. 44: 122–128.

- Xie, Z.S., Cao, H., Li, B., Forney, C.F., Wang, B., Xu, W., Wang, S. (2011). Effect of root restriction on the ultrastructure of phloem in grape leaves. *African Journal of Biotechnology*. 10(85): 19737-19744.
- Xie, Z.S., Guo, X., Cao, H. (2013). Effect of root restriction on vegetative growth and leaf anatomy of 'Kyoho' grapevines cultivar. *African Journal of Agricultural Research*. 8(15): 1304-1309.
- Xie, Z.S., Li, B., Forney, C. F., Xu, W.P., Wang, S.P. (2009). Changes in sugar content and relative enzyme activity in grape berry in response to root restriction. *Scientia Horticulturae*. 123: 39-45.
- Xiong, J., Tian, Y., Chen, Q. (2017). Comparison of coconut coir, rockwool, and peat cultivations for tomato production: nutrient balance, plant growth and fruit quality. *Frontiers in Plant Science*. 8: 1327.
- Xu, G., Wolf, S., Kafkafi, U. (2001). Interactive effect of nutrient concentration and container volume on flowering, fruiting and nutrient uptake of sweet pepper. *Journal of Plant Nutrition*. 24(3): 479-501.
- Xu, H-L., Gauthier, L., Gosselin, A. (1995). Effects of fertigation management on growth and photosynthesis of tomato plants grown in peat, rockwool and NFT. *Scientia Horticulturae* 63: 11-20.
- Yang, Z., Hammer, G., Oosterom, E.V., Rochais, D., Deifel, K. (2010). Effects of pot size on growth of maize and sorghum plants. *Proceedings of the 1st Australian Summer Grains Conference, Gold Coast, Australia, 21st– 24th June 2010*.
- Yeh, D.M. and Chiang, H.H. (2001). Growth and flower initiation in hydrangea as affected by root restriction and defoliation. *Scientia Horticulturae*. 91: 123-132.
- Yin, C.Y. Berninger, F., Li, C.Y. (2006). Photosynthetic responses of *Populus przewalski* subjected to drought stress. *Photosynthetica*. 44(1): 62-68.
- Yong Jean, W. H., Stuart Letham, D., Chin Wong, S., Farquhar, G. D. (2010). Effects of root restriction on growth and associated cytokinin levels in cotton (*Gossypium hirsutum*). *Functional Plant Biology*. 37: 974-984.
- Zainudin, M. (2001). Root restriction for growth control and precocity in starfruit (*Averrhoa carambola*). Ph.D Thesis, Universiti Putra Malaysia, Serdang, Selangor.
- Zainudin, M. (2006). Effects of root restriction on growth, flowering and water uptake of starfruit. *J.Trop. Agric. and Fd. Sc.* 34(1): 27-36.

- Zangaro, W., Torezan, J.M.D., Rostirola, L.V., Souza, D.P.B., Nogueira, M.A. (2015). Influence of mycorrhizas, organic substrates and container volumes on the growth of *Heliocarpus popayanensis* kunth. *Cerne*. 21(3): 395-403.
- Zhang, R., Shen, Z., Guan, H. (1997). Influences of root zone restriction and nitrogen nutrition on hybrid wheat growth. *Pedosphere*. 7(2):171-176.
- Zhu, L., Wang, S., Yang, T., Zhang, C., Xu, W. (2006). Vine growth and nitrogen metabolism of 'Fujiminori' grapevines in response to root restriction. *Scientia Horticulturae*. 107: 143-149.
- Zulkarami, B., Tajul, M.I., Fariz, A., Husni, M.O., Norazrin, A., Radziah, O., Mohd Razi, I., Rafii, M.Y. (2012). Effects of bacteria and arbuscular mycorrhizae inoculation at different electrical conductivity level on growth and yield of rockmelon (*Cucumis melo*) under soilless culture. *Australian Journal of Crop Science*. 6(11): 1494-1501.

BIODATA OF STUDENT

Nurul Idayu Zakaria was born in Pekan, Pahang, Malaysia in 1986. She is the third sister of six siblings. Her father was passed away in 2014 and her mother was retired government officer. She graduated secondary school in Jerantut, Pahang in 2003. She finished matriculation level in life science in Pahang Matriculation College, Malaysia in 2004. She got her Bachelor of Agricultural Science with first class honors in University Putra Malaysia in 2009. She started her job in Faculty of Agriculture as a research assistant from May 2009 till July 2009. Based on interest and opportunity, she decided to extend her study to a higher level in Doctor of Philosophy in Environmental Plant Physiology since September 2009 till now.

LIST OF PUBLICATIONS

- Nurul Idayu, Z.,** Mohd Razi, I., Yahya, A., Puteri Edaroyati, M.W., Zulkarami, B. (2019). Physiological responses, root morphology and yield of chilli plant (*Capsicum annuum* L.) in response to container size and nutrient solution concentration. *Asian Journal of Crop Science*. Accepted on 26th April 2019.
- Nurul Idayu, Z.,** Mohd Razi, I., Yahya, A., Puteri Edaroyati, M.W., Zulkarami, B. (2020). Effect of root restriction on the growth, photosynthesis rate, and source and sink relationship of chilli (*Capsicum annuum* L.) grown in soilless culture. *BioMed Research International*. Vol. 2020. Article ID 2706937, 14 pages. <https://doi.org/10.1155/2020/2706937>
- Nurul Idayu, Z.,** Mohd Razi, I., Yahya, A., Puteri Edaroyati, M.W., Deivaseeno, D. (2019). Effect of polybag size on growth, root morphology and yield of chilli (*Capsicum annuum* L.) grown in soilless culture. *The Journal of Animal and Plant Sciences* (Submitted).



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