



UNIVERSITI PUTRA MALAYSIA

**CHEMICAL MODIFICATION OF LIPASE AND ITS IMMOBILIZATION ON
POLYMER BEADS FOR USE IN ORGANIC SYNTHESIS**

MAHIRAN BASRI

FSAS 1993 2

**CHEMICAL MODIFICATION OF LIPASE AND ITS IMMobilization ON
POLYMER BEADS FOR USE IN ORGANIC SYNTHESIS**

By

MAHIRAN BASRI

**Thesis Presented in Fulfillment of the
Requirements for the Degree of Doctor of Philosophy
in the Faculty of Science & Environmental Studies,
Universiti Pertanian Malaysia.**

1993



DEDICATIONS

To my beloved grandmother....

**who has taught me all there is about beauty,
love and human grace.**

To abah and mak....

who have made me feel very special.

To adik-adik....

who have always given me the challenge to work harder.

To Jamal....

who has given me all the patience, love and support.

To Gaja and Jordie....

who make living a joyous happening.

ACKNOWLEDGEMENTS

Special Acknowledgement

I would like to express my sincere appreciation to Dr. Kamaruzaman Ampon for his compassion and optimism, Dr. Abu Bakar Salleh for his vision and persistence, Dr. Wan Zin Wan Yunus for his understanding and patience and Dr. Che Nyonya Abd. Razak for her constant support. Their contributions have been an inspiration in the completion of this thesis.

Acknowledgement

Special thanks are also extended to my colleagues, friends and department staff whose help, advice, encouragement and humour were invaluable during the course of the research:

Puan Raja Nor Zaliha Raja Abd. Rahman, En. Yusof Samad, Puan Zainoha Zakaria, Puan Rosiah Musani, Puan Normayati, Cik Rohaidah.

En. Atan Sharif, Dr. Sidik Silong, Puan Kamaliah Sirat, En. Nazari Ahmad, En. Buharan Nordin, En. Kamal Margona.

Dr. Khatijah Yusof, Dr. Juzu Hayati.

Members of Enzyme and Microbial Technology Lab.

Members of Jabatan Biokimia & Mikrobiologi.

Members of Jabatan Kimia.

The Public Service Department Malaysia, The Universiti Pertanian Malaysia, The Southeast Asian Ministers of Education Organization (SEAMEO) and Southeast Asia Regional Center for Graduate Studies and Research in Agriculture (SEARCA) and the Ministry of Science, Technology and Environment, Malaysia (IRPA Project no. 1 07 05 086) for their continued supports.

TABLE OF CONTENTS

	Page
ACKNOWLEDGEMENTS	ii
LIST OF TABLES	x
LIST OF FIGURES	xii
LIST OF PLATES	xvi
LIST OF ABBREVIATIONS	xvii
ABSTRACT	xviii
ABSTRAK	xxi
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	7
Enzymes as Biocatalysts	7
Lipases	17
Reactions Catalyzed by Lipases	17
Lipase Selectivity	20
Importance of Esters	26
Lipase Catalysis in Organic Solvents	32
Chemical Modification of Enzyme	34
Modification to Alter Properties	
of Enzymes	35

	Page
Modification of Lipase with PEG	36
Amidination of Lipase with Imidoesters . . .	38
Reductive Alkylation of Lipase	
with Aldehydes	40
 Immobilization of Enzyme	41
Immobilization Techniques	42
Immobilization by Adsorption	44
 3 MATERIALS AND METHODS	52
Purification of Lipase	56
Protein Determination	56
Activity Determination	57
 Synthetic Activity	58
 Hydrolytic Activity	58
Chemical Modification Reagents	60
 Activation of Monomethoxypolyethylene	
 Glycol	60
 Synthesis of Imidoesters	61
 Chemical Modification of Lipase	63
 Modification with Activated PEG	63
 Amidination with Imidoesters	64
 Reductive Alkylation with Aldehydes	65

	Page
Analysis of Modified Lipases using	
Hydrophobic Chromatography	66
Immobilization of Lipase	66
Adsorption of Native and	
Modified Lipases	68
Effect of Temperature on Enzyme	
Adsorption	68
Effect of pH on Enzyme Adsorption	68
Effect of Bead Sizes on	
Enzyme Adsorption	69
Operational Stability	69
Glutaraldehyde Crosslinking	70
Characteristics of Lipases	70
Effect of Temperature on the	
Esterification Activity	70
Catalytic Activity in the Presence of	
Organic Solvents	71
Patty Acid Selectivity in the	
Esterification Reaction	71
Thermostability	71
Stability in Organic Solvents	72
Storage Stability	72
Kinetic Studies	72

	Page
4 RESULTS	75
Modification of Lipase with Activated PEG . . .	75
Properties of Activated	
Monomethoxypolyethylene Glycol	75
Reaction of Lipase with Activated PEG . . .	78
Amidination of Lipase with Imidoesters	78
Properties of Imidoesters	78
Amidination of Lipase	86
Reductive Alkylation of Lipase with Aldehydes. .	86
Reductive Alkylation of Lipase	86
Characteristics of Modified Lipases	87
Catalytic Activity of PEG-Lipases	87
Catalytic Activity of Amidinated Lipases . .	87
Catalytic Activity of Alkylated Lipases . .	90
The Effect of Temperature on	
Esterification Activity	94
Catalytic Activity in the	
Presence of Organic Solvents	100
Patty Acid Selectivity in the	
Esterification Reaction	104
Thermostability	108
Stability in Organic Solvents	115
Storage Stability	120

	Page
Hydrophobicity of Modified Lipases	123
Immobilization of Modified Lipases	126
 Adsorption of Native and	
 Modified Lipases	126
 Adsorption Temperature	131
 Adsorption pH	131
 Effect of Bead Sizes on Adsorption	131
Characteristics of Immobilized Lipases	135
 Catalytic Activity of Immobilized	
 Lipases	135
 The Effect of Temperature on	
 Esterification Activity	137
 Catalytic Activity in the Presence	
 Organic Solvents	137
 Patty Acid Selectivity in the	
 Esterification Reaction	142
 Thermostability	142
 Stability in Organic Solvents	145
 Storage Stability	149
 Operational Stability	149
Kinetic Studies	152
5 DISCUSSION	155
 Modification of Lipase	155
 Characteristics of Modified Lipases	158

	Page
Immobilization of Modified Lipases	165
Characteristics of Immobilized Lipases	170
Kinetic Studies	177
6 SUMMARY	179
Recommendations for Further Studies	184
BIBLIOGRAPHY	186
APPENDIX	203
VITA	205

LIST OF TABLES

Table		Page
1	Properties of the Different Polymer Beads	67
2	Characteristics of Imidoesters	80
3	Catalytic Activity of PEG-lipases	88
4	Catalytic Activity of Amidinated Lipases	89
5	Catalytic Activity of Alkylated Lipases	93
6	Catalytic Activity of PEG-lipases in Various Organic Solvents	101
7	Catalytic Activity of Imidoester Lipases in Various Organic Solvents	102
8	Catalytic Activity of Alkylated Lipases in Various Organic Solvents	103
9	Storage Stability of PEG-lipase after 60 days	121
10	Storage Stability of Amidinated Lipase after 60 days	122
11	Storage Stability of Alkylated Lipase after 60 days	124
12	Hydrophobicity of Modified Lipases	125
13	Immobilized Activity of Modified Lipases on Different Types of Polymer Beads	136
14	Effect of Increasing the Degree of Lipase Modification on the Immobilized Activity	138
15	a: Specific Esterification Activities of XAD7 Immobilized Lipases in Various Organic Solvents	140
15	b: Specific Esterification Activities of PL1900 Immobilized onto Several Types of Polymer Beads in Various Organic Solvents	141

Table		Page
16	Stability of XAD7 Immobilized Lipase Preparations in Different Storage Conditions after 60 days	150
17	Kinetic Parameters of Lipases	154

LIST OF FIGURES

Figure		Page
1	Reactions Catalyzed by Lipases	18
2	A Gas Chromatogram Showing the Synthesis of Propyl Oleate from Oleic Acid and Propanol in the Presence of Lipase	74
3	a: IR Spectrum of Activated PEG1900	76
	b: IR Spectrum of Unactivated PEG1900	77
4	Modification of Lipase with Different Amounts of Activated PEG1900 (APEG)	79
5	a: IR Spectrum of Methyl 4-phenylbutyrimidate (Imidoester (VI))	81
	b: IR Spectrum of 4-phenyl-butyronitrile, the Corresponding Nitrile of Methyl 4-phenylbutyrimidate	82
6	a: H-NMR Spectrum of Methyl 4-phenylbutyrimidate (Imidoester (VI))	84
	b: H-NMR Spectrum of 4-phenyl-butyronitrile, the Corresponding Nitrile of Methyl 4-phenylbutyrimidate	85
7	a: The Effect of Increasing the Degree of Modification on the Esterification Activity of Lipase Modified with Methyl Acetimidate (Imidoester I)	91
7	b: The Effect of Increasing the Degree of Modification on the Esterification Activity of Lipase Modified with Methyl 4-phenyl- butyrimidate (Imidoester VI)	92
8	a: The Effect of Increasing the Degree of Modification on the Esterification Activity of Lipase Modified with Acetaldehyde	95

	Page
8 b: The Effect of Increasing the Degree of Modification on the Esterification Activity of Lipase Modified with Dodecyldehyde	96
9 The Effect of Temperature on the Esterification Reaction by PEG-lipases	97
10 The Effect of Temperature on the Esterification Reaction by Imidoester Lipases	98
11 The Effect of Temperature on the Esterification Reaction by Alkylated lipases	99
12 The Effect of Chainlength of Fatty Acids as Acyl Donor in the Esterification Reaction by PEG-lipase	105
13 The Effect of Chainlength of Fatty Acids as Acyl Donor in the Esterification Reaction by Imidoester Lipase	106
14 The Effect of Chainlength of Fatty Acids as Acyl Donor in the Esterification Reaction by Alkylated Lipase	107
15 a: Thermostability of PEG-lipases Incubated for One Hour in Benzene	109
b: Stability PEG1900-lipase with Respect to Time at 40°C and 50°C	111
16 a: Thermostability of Imidoester Lipases Incubated for One Hour in Benzene	112
b: Stability of Imidoester Lipase (VI) with Respect to Time at 40°C and 50°C	113
17 a: Thermostability of Alkylated Lipases Incubated for One Hour in Benzene	114
b: Stability of Dodecyl Lipase with Respect to Time at 40°C and 50°C	116

	Page
18 Stability of PEG-lipases Incubated in Benzene for Ten Days at Room Temperature	117
19 Stability of Imidoester Lipases Incubated in Benzene for Ten Days at Room Temperature	118
20 Stability of Alkylated Lipases Incubated in Benzene for Ten Days at Room Temperature	119
21 Protein Adsorption Profile of Native and PEG-lipase onto Several Types of Polymer Beads	127
22 Activity Adsorption Profile of Native and PEG-lipase onto Several Types of Polymer Beads (Intact Beads)	128
23 Activity Adsorption Profile of Native and PEG-lipase onto Several Types of Polymer Beads (Crushed Beads)	129
24 The Effect of Different Temperature on the Immobilization of Lipase onto Several Types of Polymer Beads	132
25 The Effect of pH on the Immobilization of Lipase onto Several Types of Polymer Beads	133
26 The Effect of XAD7 Bead Sizes on the Immobilization of PEG-lipase	134
27 The Effect of Temperature on the Esterification Reaction by Immobilized Lipase	139
28 The Effect of Chainlength of Fatty Acids as Acyl Donor in the Esterification Reaction by Immobilized Lipase	143
29 Thermostability PEG-lipase Immobilized onto Several Types of Polymer Beads Incubated for One Hour in Benzene	144

	Page
30 Thermostability of XAD2 Immobilized, Modified Lipases Incubated for One Hour in Benzene	146
31 Stability of Immobilized PEG-lipases Incubated in Benzene for Ten Days at Room Temperature	147
32 Stability of XAD2 Immobilized, Modified Lipases Incubated in Benzene for Ten Days at Room Temperature	148
33 Operational Stability of Immobilized Lipases Before and After Crosslinking with Different Glutaraldehyde Concentrations	151
34 A Lineweaver-Burk Plot Showing a Typical Graph of $1/V$ against $1/[S]$	153

LIST OF PLATES

Plate		Page
1	A Thin-layer Chromatogram Showing the Synthesis of Propyl Oleate from Oleic Acid and Propanol in the Presence of Native and Modified Lipases	59

LIST OF ABBREVIATIONS

PEG	monomethoxypolyethylene glycol
PEG1900	monomethoxypolyethylene glycol (MW=1900)
PEG5000	monomethoxypolyethylene glycol (MW=5000)
APEG	activated PEG
TNBS	trinitrobenzene sulfonate
RCOOH	polycarboxylic acid
RCN	polyacrylonitrile
DVB	divinyl benzene
PPLC	Fast performance liquid chromatography
IR	infra red
NMR	nuclear magnetic resonance
NL	native lipase
Imidoester I	methyl acetimidate
Imidoester II	methyl benzimidate
Imidoester III	methyl 4-biphenyl benzimidate
Imidoester IV	methyl n-dodecanimide
Imidoester V	methyl 3-phenylpropionimidate
Imidoester VI	methyl 4-phenylbutyrimidate
PL1900	lipase modified with PEG1900
PL5000	lipase modified with PEG5000
IL(I)	lipase modified with imidoester I
IL(II)	lipase modified with imidoester II
IL(III)	lipase modified with imidoester III
IL(IV)	lipase modified with imidoester IV
IL(V)	lipase modified with imidoester V
IL(VI)	lipase modified with imidoester VI
AL	lipase modified with acetaldehyde (ethyl lipase)
DL	lipase modified with dodecyldehyde
XAD2NL	native lipase immobilized on XAD2
XAD2PL	PEG-lipase immobilized on XAD2
XAD7NL	native lipase immobilized on XAD7
XAD7PL	PEG-lipase immobilized on XAD7
XAD7IL	imidoester lipase immobilized on XAD7
XAD7AL	ethyl lipase immobilized on XAD7
RCOOHNL	native lipase immobilized on RCOOH
RCOOHPL	PEG-lipase immobilized on RCOOH

Abstract of the thesis presented to the Senate of Universiti Pertanian Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy.

CHEMICAL MODIFICATION OF LIPASE AND ITS IMMOBILIZATION ON POLYMER BEADS FOR USE IN ORGANIC SYNTHESIS

By

Mahiran Basri

April 1993

Chairman: Associate Professor Dr. Kamaruzaman Ampon

Faculty: Science & Environmental Studies

A simple and effective method to produce a more active, stable and practical lipase preparation was identified. Soluble lipase from *Candida rugosa* was modified with different types of hydrophobic chemical modifying reagents.

The esterification activities of the modified lipases were enhanced following their modification. The degree of activity enhancement depends on the type and molecular weight of the modifiers used and the degree of modification of the enzyme. A lower degree of enzyme derivatization was required for modification with the high molecular weight modifiers to attain maximal activities. In the case of monomethoxypolyethylene glycol (PEG), however, maximal activity was attained only after exhaustive modification.

The optimum esterification temperature and preference of fatty acids as acyl donors of the modified lipases were very similar to those of the native enzyme. Both were more active in non-polar solvents than in polar solvents. The modified lipases showed higher thermostability, solvent stability and storage stability compared to the native lipase. The lipase modified with PEG1900 was the most thermostable, and that modified with methyl 4-phenylbutyrimidate (imidoester VI) was the most stable when incubated in benzene for ten days. The best storage condition was at low temperature and in the lyophilized form.

When porous polymer beads were added to a solution of the modified lipase at room temperature and stirred gently for 0.5 to 2 hours, the enzyme was strongly adsorbed onto them. Native lipase was only weakly adsorbed onto the supports.

The immobilized activity of modified lipases were higher compared with the native enzyme. The magnitude of the difference depended on the type of modifiers, the degree of modification of the enzyme and the type of polymers used for immobilization. Lipase that was modified with the more hydrophobic modifiers showed higher immobilized activity compared to those modified with the less hydrophobic modifiers (except for PEG5000). The highest immobilized activity was exhibited by lipase modified with PEG1900 (95% modification).

Amberlite XAD7, XAD8 and Polycarboxylic acid beads (RCOOH) were good supports for enzyme immobilization by the above methods.

The optimum esterification temperature of lipase immobilized in this manner was not changed. However, its preference for certain fatty acids as acyl donors was altered. Medium chain length fatty acids were favoured over longer chain fatty acids when the enzymes were immobilized. Their activities were increased even in the presence of polar solvents. The immobilized lipases also showed higher thermostability, solvent stability and storage stability compared with native and modified lipases.

Abstrak tesis yang dikemukakan kepada Senat Universiti Pertanian Malaysia bagi memenuhi keperluan Ijazah Doktor Falsafah.

PENGUBAHSUAIAN LIPASE SECARA KIMIA DAN SEKATGERAKANYA KEPADA BEBERAPA POLYMER UNTUK KEGUNAAN DALAM SINTESIS ORGANIK.

Oleh

Mahiran Basri

April 1993

Pengerusi : Prof. Madya Dr. Kamaruzaman Ampon

Pakulti : Sains & Pengajian Alam Sekitar

Satu kaedah yang mudah dan berkesan telah dikenalpasti untuk menghasilkan lipase yang lebih aktif, stabil dan praktikal. Lipase dari *Candida rugosa* yang terlarut telah diubahsuai dengan beberapa bahan pengubahsuai hidrofobik.

Lipase terubahsuai telah menunjukkan kelebihan aktiviti esterifikasi. Darjah pertambahan aktiviti bergantung kapada jenis dan berat molekul bahan pengubahsuai yang digunakan dan darjah ubahsuai enzim itu sendiri. Darjah yang lebih rendah adalah diperlukan bagi mengubahsuai enzim dengan bahan yang mempunyai berat molekul yang tinggi bagi mendapatkan aktiviti maksimum. Untuk monomethoxypolyethylene glycol (PEG), aktiviti maksimum didapati dengan mengubahsuai lipase habis-habisan.

Suhu optimum esterifikasi dan pemilihan terhadap asid lemak sebagai penderma kumpulan asil untuk lipase terubahsuai adalah sama dengan lipase natif. Selain dari itu, kedua-dua jenis lipase didapati lebih aktif di dalam pelarut organik yang tidak berikutub berbanding dengan di dalam pelarut organik yang lebih berikutub. Lipase terubahsuai menunjukkan kestabilan terma, kestabilan dalam pelarut organik dan kestabilan penyimpanan yang lebih tinggi. Lipase yang diubahsuai dengan PEG1900 menunjukkan kestabilan terma yang tertinggi dan lipase yang dimodifikasi dengan metil 4-fenilbutirimidat (imidoester VI) adalah yang paling stabil apabila dieram dalam benzena selama sepuluh hari. Keadaan penyimpanan terbaik adalah pada suhu rendah dan keadaan beku-kering.

Apabila butir-butir polimer yang berliang telah dimasukkan ke dalam larutan lipase terubahsuai pada suhu bilik dan digoncang selama 0.5-2 jam, enzim didapati terjerap lebih kuat. Lipase natif hanya terjerap dengan lemah kepada butir-butir polimer.

Lipase terubahsuai menunjukkan aktiviti tersekatgerak yang lebih tinggi berbanding dengan lipase natif. Penambahan aktiviti bergantung kepada jenis bahan pengubahsuai, darjah ubahsuai dan jenis polimer yang digunakan. Lipase yang diubahsuai dengan bahan yang lebih hidrofobik menunjukkan aktiviti tersekatgerak yang lebih tinggi berbanding dengan

lipase diubahsuai dengan bahan yang kurang hidrofobik (melainkan PEG5000). Lipase diubahsuai dengan PEG1900 (95% ubahsuai) memberikan aktiviti tersekatgerak paling tinggi. Amberlite XAD7, XAD8 dan polikarboksilik asid (RCOOH) adalah polimer yang baik untuk menyekatgerakan enzim dalam kajian ini.

Suhu optimum lipase disekatgerak seperti ini didapati tidak bertukar. Walau bagaimanapun, pilihan terhadap asid lemak sebagai penderma kumpulan asil didapati bertukar. Asid lemak rantai sederhana lebih disukai berbanding dengan asid lemak rantai panjang apabila enzim disekatgerak. Aktivitinya juga tinggi di dalam pelarut organik yang lebih polar. Lipase tersekatgerak menunjukkan kestabilan terma, kestabilan dalam pelarut organik dan kestabilan penyimpanan yang lebih tinggi lagi daripada lipase natif dan lipase terubahsuai.