

# EFFECTS OF MARINE MICROALGAE AND CYANOBACTERIA TOWARDS THE MORPHOLOGY AND SURVIVAL RATE OF ZEBRAFISH LARVAE, Danio rerio

**TOH EN CHUN** 

FP 2018 22

# EFFECTS OF MARINE MICROALGAE AND CYANOBACTERIA TOWARDS THE MORPHOLOGY AND SURVIVAL RATE OF ZEBRAFISH LARVAE, Danio rerio

TOH EN CHUN 181413

This project is submitted in partial fulfillment of the requirement for the degree of Bachelor of Aquaculture Science

DEPARTMENT OF AQUACULTURE
FACULTY OF AGRICULTURE
UNIVERSITI PUTRA MALAYSIA
SERDANG, SELANGOR

# CERTIFICATION OF APPROVAL DEPARTMENT OF AQUACULTURE FACULTY OF AGRICULTURE UNIVERSITI PUTRA MALAYSIA

Name of student : Toh En Chun

Matric number : 181413

Programme : Bachelor of Agriculture (Aquaculture)

Year : 2018

Name of supervisor : Dr. Natrah Fatin binti Mohd Ikhsan

Title of project : Effects of marine microalgae and

cyanobacteria towards the morphology and

survival rate of zebrafish larvae, Danio rerio

This is to certify that I have examined the final project report and all corrections have been made is recommended by the panel examiners. This project complies with the recommended format stipulated in the AKU4999 project guidelines, Department of Aquaculture, Faculty of Agriculture, Universiti Putra Malaysia.

Signature and official stamp of supervisor:

Supervisor's name: Dr Natrah Fatin binti Mohd Ikhsan

Date:

## **ACKNOWLEDGEMENT**

First and foremost, I am thankful to The Almighty God for giving me the opportunity to complete this research.

I would like to express my deepest thanks to my supervisor Dr. Natrah Fatin binti Mohd Ikhsan for her supervision, guidance and continuous support throughout my study period in completing this thesis.

I also would like to express my gratitude to all lecturers and staffs in Department of Aquaculture that directly and indirectly involved during my study period.

I am grateful to the Master, PhD student and friends that directly and indirectly help and cooperate with me during my experiment. Their continuous support to me is very precious and really helps to motivate me to complete this thesis. They provided me the help and guidance which contribute on my success of finishing my research

I sincerely thanked my parents and family members that always support me in any circumstance and all condition. Without their greatest support and advice, it was difficult for me to accomplish my thesis.

Thank you.

Toh En Chun

#### **ABSTRACT**

The zebrafish (Danio rerio) is becoming internationally recognized as a test model organsims representing animals as well as human. An experiment was conducted to study selected different microalgae and cyanobacteria extracts (Spirulina platensis, Monoraphidium sp., Halamphora sp., Chaetoceros sp. and Chattonella marina) on the D. rerio larvae morphology and survival rate. Samples of microalgae and cyanobacteria at stationary phase were extracted using 95% methanol and were exposed to D. rerio eggs at the concentrations of 0 ppm, 10 ppm, 20 ppm, 50 ppm and 100 ppm. The embryonic development was observed daily until hatched. The results showed that no significant differences between egg morphology were detected through the exposure from 0 ppm until 100 ppm. The experiment was then repeated through an increase of concentration up to 200 ppm. It was shown that a toxic Chattonella marina induced negative impacts towards the zebrafish larvae morphology causing deformation with low survival rate at (46.67%) compared to control treatment (0ppm, sterile distilled water) which have 100% survival rate. This indicated that the Chattonella marina is lethal towards the zebrafish larvae at high concentration.

Keywords: zebrafish, microalgae, cyanobacteria, survival rate

#### **ABSTRAK**

Ikan zebra (Danio rerio) sudah diiktiraf pada peringkat antarabangsa sebagai model untuk mewakili haiwan dan manusia. Eksperimen dilaksanakan untuk mengkaji mikroalga dan sianobakteria ekstrak yang berbeza (Spirulina platensis, Monoraphidium sp., Halamphora sp., Chaetoceros sp. dan Chattonella marina) dalam mempengaruhi morfologi larva dan kadar penetasan larva. Sampel mikroalga dan sianobakteria telah diekstrak pada fasa pegun menggunakan 95% metanol dan didedahkan kepada telur D. rerio pada kepekatan 0 ppm, 10 ppm, 20 ppm, 50 ppm, and 100 ppm. Pembangunan embrio diperhatikan setiap hari sehingga menetas. Keputusan menunjukkan bahawa tiada perbezaan yang ketara antara morfologi telur dari 0 ppm hingga 100 ppm. Eksperimen kemudian diulang melalui peningkatan kepekatan sehingga 200 ppm. Peningkatan rawatan menunjukkan bahawa kawalan positif, Chattonella marina yang beracun telah menyebabkan impak negative terhadap morfologi larva ikan zebra menyebabkan kecacatan bentuk badan dan kadar kelangsungan hidup pada (46.67%) berbanding dengan rawatan kawalan (Oppm, air suling yang steril) yang mempunyai kadar kelangsungan hidup pada 100%. Ini membuktikan bahawa Chattonella marina sangat berbahaya kepada larva zebrafish pada kepekatan yang tinggi.

Keywords: ikan zebra, mikroalga, sianobakteria, kelangsungan hidup

# TABLE OF CONTENTS

Contents		Page	
ACKNOWLEDGEMENT			
ABSTRACT		ii	
ABSTRAK		iii	
TABLE OF CONTENTS		iv	
LIST OF TABLES		vi	
LIST OF F	LIST OF FIGURES		
LIST OF A	BBREVIATIONS AND SYMBOLS	viii	
CHAPTER			
1	INTRODUCTION	1	
-		•	
2	LITERATURE REVIEW	4	
	2.1 Microalgae	4	
	2.2 Cyanobacteria	7	
	2.3 Microalgae and cyanobacteria toxicity	8	
	2.4 Zebrafish	11	
3	METHODOLOGY	12	
	3.1 Culture of microalgae	12	
	3.2 Extraction of microalgae	13	
	3.3 Water parameters for zebrafish breeding	14	
	3.4 Conditioning and breeding of zebrafish	15	
	3.5 Treatment of zebrafish eggs with microalgae extract	15	

	3.6 Observation of embryos development	16
	3.7 Statistical analysis	17
4	RESULTS	18
	4.1 Spawning and fertility rate	18
	4.2 Embryonic development stages of <i>Danio rerio</i>	19
	4.2.1 Effects of microalgae on zebrafish larvae	21
	larvae in five different treatments at day 3	
	4.2.2 Morphological observation of the zebrafish	23
	upon microalgae treatment	
5	DISCUSSION	24
6	CONCLUSION	28
	REFERENCES	29
	APPENDICES	37

LIST OF TABLES		
Table 2.1	Toxicity of microalgae	9
Table 2.2	Toxicity of cyanobacteria	10
Table 3.1	List of extracted microalgae	13
Table 3.2	Water parameters from broodstock conditioning and breeding tanks	14
Table 4.1	Spawning and fertility rate of each tank	18
Table 4.2	Observation of Embryonic Development of zebrafish stage	20
Table 4.3	Comparison on the survival rate of larvae in five different treatments	21
Table 4.4	Images of deform and non-deform larvae treated with 200 ppm microalgae extract	23

LIST OF FIGURES		
Figure 4.1	Image of unfertilized embryo (UN), fertilized embryo (FR) and 1 cell embryo (a)	19
Figure 4.2	Comparison of survival rate of larvae in a higher concentration (200ppm) at day 3	22



# LIST OF ABBREVIATIONS AND SYMBOLS

mL - Milliliter

°C - Degree Celsius

± - Plus minus

< - Lower than

% Percent

ppm - Part per million

rpm - Revolutions per minute

mg/L - Milligram/liter

μl - Microliter

μmol - Micromole

 $m^2s^{-1}$  - Metre squared per second

> - More than

< - Less than

# **CHAPTER 1**

### INTRODUCTION

# 1.0 Introduction

Microalgae is a common live feed in aquaculture. Microalgae are very diverse in species, shape and also size which makes it suitable for variety of fish size and stage to feed. They reproduce by vegetative, asexual and sexual mechanisms (Acreman, 1994). Microalgae are mostly autotrophs at which they use light and carbon dioxide to make their own food, although there are some heterotrophs (cannot produce its own food). In general, microalgae contain lipid, protein, fatty acids and amino acids (Kay, 1991). Micronutrients such as various trace metals and the B-complex vitamins such as thiamin (B1), cyanocobalamin (B12) and biotin (B7) were used in energy production and cell division (Kay, 1991). Although microalgae are commonly regard as beneficial organisms, some species are toxic, also known as harmful algal bloom (HAB) species. These toxin productions are correlated with temperature, age of the culture, light intensity, and pH (Kay, 1991).

Meanwhile, cyanobacteria are bacteria that are capable of producing a wide range of potent toxins as secondary metabolites (Van Apeldoorn et al., 2007). There are several similar characteristics of cyanobacteria and microalgae in terms of pigmentation where both of the organisms possessed chlorophyll a (Van Apeldoorn et al., 2007). In tropical countries such as in Malaysia, cyanobacteria multiply rapidly particularly in ponds that are rich in nutrients. Cyanobacteria has the ability to form thick surface with high cell density. Cyanobacterial blooms happen due to the presence of buoyancy-conferring gas vacuoles (Cook et al., 2004; Chorus, 2000). This phenomenon frequently occurs in shallow littoral areas that are shelter to fish larvae and could poison the fish larvae causing chronic effects or even death (Sivonen & Jones, 1999). Some contains cyanotoxins and are responsible for acute and (sub) chronic poisonings of wild/domestic animals and humans (Van Apeldoorn et al., 2007).

Zebrafish can be used as fish model for toxicity investigations that represents both aquatic and human. Zebrafish (*Danio rerio*) is a type of model fish that originates from South Asia which are India, Bangladesh, Nepal, Myanmar and Pakistan (Lawrence, 2007). Their living requirement is highly related towards aquatic vegetation. They can be found in calm water as in rice field, upper reaches of rivers and irrigation ditches (Lawrence, 2007). The zebrafish normally breeds in groups, they are egg laying fish that have no parental care towards the egg (Lawrence, 2007). It can withstand wide range of temperature from 27°C to 34°C,

pH 6.0-8.0, and depth range from 16 cm to 57 cm (Engeszer, 2007). In natural environment, the zebrafish have to face predator such as the tire-track eel, swamp eel and dragonfly larvae. Although zebrafish is considered as a hardy species, various harmful activities such as strip mining lead to extensive pollution, agricultural practice increase acidity of the aquatic environment, and poisoning entire stream for livelihood are able to cause zebrafish mortality (Engeszer, 2007). Egan (2009) described that the zebrafish is valid and reliable as a model of stress and affective disorders.

The objectives of this study were to evaluate the toxic effects of microalgae and cyanobacteria towards the survivability of zebrafish larvae based on survival rate and overall morphology and to distinguish the toxic and non-toxic of marine microalgae and cyanobacteria species towards the zebrafish larvae.

# **REFERENCES**

- Acreman, J. (1994). Algae and cyanobacteria: isolation, culture and long-term maintenance. *Journal of Industrial Microbiology & Biotechnology*, 13(3), 193-194.
- Amatruda, J. F., Shepard, J. L., Stern, H. M., & Zon, L. I. (2002). Zebrafish as a cancer model system. *Cancer cell*, 1(3), 229-231.
- Ball, A. S., Williams, M., Vincent, D., & Robinson, J. (2001). Algal growth control by a barley straw extract. *Bioresource Technology*, 77(2), 177-181.
- Basti, L., Nagai, K., Go, J., Okano, S., Oda, T., Tanaka, Y., & Nagai, S. (2016). Lethal effects of ichthyotoxic raphidophytes, Chattonella marina, C. antiqua, and Heterosigma akashiwo, on post-embryonic stages of the Japanese pearl oyster, Pinctada fucata martensii. *Harmful algae*, 59, 112-122.
- Becker, E. W. (2007). Micro-algae as a source of protein. *Biotechnology advances*, 25(2), 207-210.
- Belkin, S., & Boussiba, S. (1991). Resistance of Spirulina platensis to ammonia at high pH values. *Plant and cell physiology*, *32*(7), 953-958.
- Berry, J. P., Gantar, M., Gibbs, P. D., & Schmale, M. C. (2007). The zebrafish (Danio rerio) embryo as a model system for identification and characterization of developmental toxins from marine and freshwater microalgae. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 145(1), 61-72.
- Borowitzka, M. A. (1997). Microalgae for aquaculture: opportunities and constraints. *Journal of Applied Phycology*, 9(5), 393.

- Carlsson, G., & Norrgren, L. (2004). Synthetic musk toxicity to early life stages of zebrafish (Danio rerio). *Archives of environmental contamination and toxicology*, 46(1), 102-105.
- Chorus, I., Falconer, I. R., Salas, H. J., & Bartram, J. (2000). Health risks caused by freshwater cyanobacteria in recreational waters. *Journal of Toxicology and Environmental Health Part B: Critical Reviews*, 3(4), 323-347.
- Cook, C. M., Vardaka, E., & Lanaras, T. (2004). Toxic cyanobacteria in Greek freshwaters, 1987—2000: Occurrence, toxicity, and impacts in the Mediterranean region. *CLEAN–Soil, Air, Water*, *32*(2), 107-124.
- Coutteau, P., & Sorgeloos, P. (1992). The use of algal substitutes and the requirement for live algae in the hatchery and nursery rearing of bivalve molluscs: an international survey. *Journal of Shellfish Research*, (2).
- Dave, G., & Xiu, R. (1991). Toxicity of mercury, copper, nickel, lead, and cobalt to embryos and larvae of zebrafish, Brachydanio rerio. *Archives of Environmental Contamination and Toxicology*, 21(1), 126-134.
- Dorantes-Aranda, J. J., Seger, A., Mardones, J. I., Nichols, P. D., & Hallegraeff, G. M. (2015). Progress in understanding algal bloom-mediated fish kills: the role of superoxide radicals, phycotoxins and fatty acids. *PloS one*, 10(7), e0133549.
- Drummond, I. A. (2005). Kidney development and disease in the zebrafish. *Journal of the American Society of Nephrology*, 16(2), 299-304.
- Egan, R. J., Bergner, C. L., Hart, P. C., Cachat, J. M., Canavello, P. R., Elegante, M. F., ... & Mohnot, S. (2009). Understanding behavioral and physiological phenotypes of stress and anxiety in zebrafish. *Behavioural brain research*, 205(1), 38-44.

- Engeszer, R. E., Patterson, L. B., Rao, A. A., & Parichy, D. M. (2007). Zebrafish in the wild: a review of natural history and new notes from the field. *Zebrafish*, *4*(1), 21-40.
- Estrada, J. P., Bescós, P. B., & Del Fresno, A. V. (2001). Antioxidant activity of different fractions of Spirulina platensis protean extract. *Il farmaco*, 56(5-7), 497-500.
- Feuga, A.M (2000). The role of microalgae in aquaculture: situation and trends. *Journal of Applied Phycology*, 12(3-5), 527-534.
- Gallardo, P. P., Alfonso, E., Gaxiola, G., Soto, L. A., & Rosas, C. (1995). Feeding schedule for Penaeus setiferus larvae based on diatoms (Chaetoceros ceratosporum), flagellates (Tetraselmis chuii) and Artemia nauplii. *Aquaculture*, 131(3-4), 239-252.
- Hajiahmadian, M., Vajargah, M. F., Farsani, H. G., & Chorchi, M. M. (2012). Effect of Spirulina platensis meal as feed additive on growth performance and survival rate in golden barb fish, Punius gelius (Hamilton, 1822). *Journal of Fisheries International*, 7(3), 61-64.
- Hill Jr, R. L., & Janz, D. M. (2003). Developmental estrogenic exposure in zebrafish (Danio rerio): I. Effects on sex ratio and breeding success. *Aquatic toxicology*, 63(4), 417-429.
- Hill, A. J., Teraoka, H., Heideman, W., & Peterson, R. E. (2005). Zebrafish as a model vertebrate for investigating chemical toxicity. *Toxicological sciences*, 86(1), 6-19.
- Jiang, H. Y., Hu, C. Q., Yang, H. P., Zhang, L. P., Peng, P. F., Luo, P., ... & Xia, J. J. (2015). Morphology and phylogeny of Halamphora yongxingensis sp. nov.(Bacillariophyta), a new marine benthic diatom isolated from Yongxing Island, South China Sea. *Phytotaxa*, 195(1), 53-64.

- Jopling, C., Sleep, E., Raya, M., Martí, M., Raya, A., & Belmonte, J. C. I. (2010). Zebrafish heart regeneration occurs by cardiomyocyte dedifferentiation and proliferation. *Nature*, 464(7288), 606.
- Kahn, S., Arakawa, O., & Onoue, Y. (1998). Physiological investigations of a neurotoxin-producing phytoflagellate, Chattonella marina (Raphidophyceae). *Aquaculture research*, 29(1), 9-17.
- Kawamura, T., Saido, T., Takami, H., & Yamashita, Y. (1995). Dietary value of benthic diatoms for the growth of post-larval abalone Haliotis discus hannai. Journal of Experimental Marine Biology and Ecology, 194(2), 189-199.
- Kay, R. A., & Barton, L. L. (1991). Microalgae as food and supplement. *Critical reviews in food science & nutrition*, 30(6), 555-573.
- Kimmel, C. B., Ballard, W. W., Kimmel, S. R., Ullmann, B., & Schilling, T. F. (1995). Stages of embryonic development of the zebrafish. *Developmental dynamics*, 203(3), 253-310.
- Lammer, E., Carr, G. J., Wendler, K., Rawlings, J. M., Belanger, S. E., & Braunbeck, T. (2009). Is the fish embryo toxicity test (FET) with the zebrafish (Danio rerio) a potential alternative for the fish acute toxicity test. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 149(2),196-209.
- Lawrence, C. (2007). The husbandry of zebrafish (Danio rerio): a review. *Aquaculture*, 269(1-4), 1-20.
- Lefebvre, K. A., Trainer, V. L., & Scholz, N. L. (2004). Morphological abnormalities and sensorimotor deficits in larval fish exposed to dissolved saxitoxin. *Aquatic Toxicology*, 66(2), 159-170.

- Levkov, Z. (2009). Diatoms of Europe: diatoms of European inland water and comparable habitats. Amphora sensu lato, 5.
- Liu, W., Au, D. W., Anderson, D. M., Lam, P. K., & Wu, R. S. (2007). Effects of nutrients, salinity, pH and light: dark cycle on the production of reactive oxygen species in the alga Chattonella marina. *Journal of experimental marine biology and ecology*, 346(1-2), 76-86.
- MacRae, C. A., & Peterson, R. T. (2003). Zebrafish-based small molecule discovery. Chemistry & biology, 10(10), 901-908.
- Madhumathi, M., & Rengasamy, R. (2011). Antioxidant status of Penaeus monodon fed with Dunaliella salina supplemented diet and resistance against WSSV. *Int J Eng Sci Technol*, *3*(10), 7249-7259.
- Mankiewicz, J., Komárková, J., Izydorczyk, K., Jurczak, T., Tarczynska, M., & Zalewski, M. (2005). Hepatotoxic cyanobacterial blooms in the lakes of northern Poland. *Environmental toxicology*, 20(5), 499-506.
- Marshall, J. A., Nichols, P. D., Hamilton, B., Lewis, R. J., & Hallegraeff, G. M. (2003). Ichthyotoxicity of Chattonella marina (Raphidophyceae) to damselfish (Acanthochromis polycanthus): the synergistic role of reactive oxygen species and free fatty acids. *Harmful Algae*, 2(4), 273-281.
- Massaut, L. (1999). Cyanobacteria management in aquaculture ponds: a review. BULLETIN-INSTITUT OCEANOGRAPHIQUE MONACO-NUMERO SPECIAL-, 579-584.
- Nagel, R. (2002). DarT: the embryo test with the zebrafish Danio rerio—a general model in ecotoxicology and toxicology. *Altex*, *19*(Suppl 1), 38-48.

- Naidu, K. A., Sarada, R., Manoj, G., Khan, M. Y., Swamy, M. M., Viswanatha, S., ... & Srinivas, L. (1999). Toxicity assessment of phycocyanin-A blue colorant from blue green alga Spirulina platensis. *Food Biotechnology*, *13*(1), 51-66.
- Phatarpekar, P. V., Sreepada, R. A., Pednekar, C., & Achuthankutty, C. T. (2000). A comparative study on growth performance and biochemical composition of mixed culture of Isochrysis galbana and Chaetoceros calcitrans with monocultures. *Aquaculture*, 181(1-2), 141-155.
- Roger, P. A., & Kulasooriya, S. A. (1980). *Blue-green algae and rice*. Int. Rice Res. Inst.
- Rougier, F., Menudier, A., Bosgiraud, C., & Nicolas, J. A. (1996). Copper and zinc exposure of zebrafish, Brachydanio rerio (Hamilton–Buchaman): effects in experimental listeria infection. *Ecotoxicology and Environmental safety*, *34*(2), 134-140.
- Sevrin-Reyssac, J., & Pletikosic, M. (1990). Cyanobacteria in fish ponds. *Aquaculture*, 88(1), 1-20.
- Shah, M. R., Lutzu, G. A., Alam, A., Sarker, P., Chowdhury, M. K., Parsaeimehr, A., ... & Daroch, M. (2017). Microalgae in aquafeeds for a sustainable aquaculture industry. *Journal of Applied Phycology*, *30*(1), 197-213.
- Shuba, E. S., & Kifle, D. (2018). Microalgae to biofuels: 'Promising'alternative and renewable energy, review. Renewable and Sustainable Energy Reviews, 81, 743-755.
- Sinden, A., & Sinang, S. C. (2016). Cyanobacteria in aquaculture systems: linking the occurrence, abundance and toxicity with rising temperatures. *International journal of environmental science and technology*, *13*(12), 2855-2862.

- Sivonen, K., & Jones, G. (1999). Cyanobacterial toxins. *Toxic cyanobacteria in water: a guide to their public health consequences, monitoring and management*, 1, 43-112.
- Soares, R. M., Magalhães, V. F., & Azevedo, S. M. (2004). Accumulation and depuration of microcystins (cyanobacteria hepatotoxins) in Tilapia rendalli (Cichlidae) under laboratory conditions. Aquatic Toxicology, 70(1), 1-10.
- Strahle, U., Scholz, S., Geisler, R., Greiner, P., Hollert, H., Rastegar, S., & Braunbeck, T. (2012). Zebrafish embryos as an alternative to animal experiments—a commentary on the definition of the onset of protected life stages in animal welfare regulations. *Reproductive Toxicology*, 33(2), 128-132.
- Sun, X., Chang, Y., Ye, Y., Ma, Z., Liang, Y., Li, T., ... & Luo, L. (2012). The effect of dietary pigments on the coloration of Japanese ornamental carp (koi, Cyprinus carpio L.). *Aquaculture*, 342, 62-68.
- Todd, N. E., & Van Leeuwen, M. (2002). Effects of Sevin (carbaryl insecticide) on early life stages of zebrafish (Danio rerio). *Ecotoxicology and Environmental Safety*, 53(2), 267-272.
- Tornabene, T. G., Bourne, T. F., Raziuddin, S., & Ben-Amotz, A. (1985). Lipid and lipopolysaccharide constituents of cyanobacterium Spirulina platensis (Cyanophyceae, Nostocales). *Marine Ecology Progress Series*, 121-125.
- Usher, K. M., Bergman, B., & Raven, J. A. (2007). Exploring cyanobacterial mutualisms. *Annu. Rev. Ecol. Evol. Syst.*, *38*, 255-273.
- Van Apeldoorn, M. E., Van Egmond, H. P., Speijers, G. J., & Bakker, G. J. (2007). Toxins of cyanobacteria. *Molecular nutrition & food research*, 51(1), 7-60.

- Wiegand, C., Pflugmacher, S., Oberemm, A., Meems, N., Beattie, K. A., Steinberg, C. E., & Codd, G. A. (1999). Uptake and effects of microcystin-LR on detoxication enzymes of early life stages of the zebra fish (Danio rerio). *Environmental Toxicology: An International Journal*, 14(1), 89-95.
- Zhang, D., Xie, P., Liu, Y., & Qiu, T. (2009). Transfer, distribution and bioaccumulation of microcystins in the aquatic food web in Lake Taihu, China, with potential risks to human health. *Science of the total environment*, 407(7), 2191-2199.