



***SYNTHESIS AND CHARACTERIZATION OF GRAPHENE OXIDE-
MOLECULARLY IMPRINTED POLYMER AND ITS ADSORPTION
PROPERTIES FOR NEOPTERIN FROM AQUEOUS SOLUTION***

KHOO WAI CHAT

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By

KHOO WAI CHAT

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements of the Degree of Master of
Science**

August 2019

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

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KHOO WAI CHAT

August 2019

Chair : Sazlinda Binti Kamaruzaman, PhD
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Neopterin is a useful biomarker for detection of malignant diseases. However, there are only a few study that has been done on the adsorption of neopterin using a graphene oxide-polymer hybrid material such as grapheme oxide-molecularly imprinted polymer (GO-MIP). The aim of this research was to synthesize and characterize GO-MIP, and the details on adsorption study toward neopterin have been thoroughly discussed. GO-MIP with neopterin as the template was synthesized via free radical polymerization method, methacrylic acid (MAA) as the monomer, ethylene glycol dimethacrylate (EGDMA) as the cross-linker, ammonium persulfate (APS) as the initiator, and 8/2 v/v ratio of dimethylsulfoxide/acetonitrile (DMSO/ACN) solution as porogen solvent. The molar ratio of target template, monomer, and cross-linker used was 1:4:16 respectively. Reflux was performed at 50 °C for 24 hours under inert nitrogen atmosphere. Neopterin binding sites were obtained by removing neopterin template from the synthesized GO-MIP via acid washing. A graphene oxide-non imprinted polymer (GO-NIP) without neopterin imprints was prepared with the same methodology. Fourier-transform infrared spectroscopy (FTIR) result for GO-MIP showed a less intense hydroxyl –OH stretching peak and a more intense carboxylic CO- stretching peak compared to GO. Elemental CHNS analysis for GO-MIP showed that the carbon wt% was 4.37% lower and the hydrogen wt% was 3.702% higher compared to GO. A thermal decomposition peak for GO-MIP can be observed at 400 °C with thermogravimetric analysis (TGA) due to the thermal degradation of MAA. Field emission scanning electron microscopy (FESEM) and transmission electron microscopy (TEM) images showed the presence of neopterin binding sites and the structural morphology of GO-MIP hybrid. Neopterin adsorption study was carried out with a mixture of 10 mg GO-MIP and 1 mL neopterin 10 mg/L standard solution. The mixture was stirred at 300 rpm for 60 minutes, then filtered using nylon syringe filter and the supernatant was analyzed using high performance liquid chromatography coupled with fluorescence detector (HPLC-FLD). GO-MIP adsorbed 45.13 % of

neopterin while GO-NIP adsorbed just 23.48 %, about twice the amount of neopterin compared to its non-imprinted counterpart. GO-MIP also showed good neopterin selectivity when under the effect of analog compound 6-biopterin, where GO-MIP adsorbed 34.63 % of neopterin but just 18.64 % 6-biopterin. The static adsorption mechanism was studied using Langmuir and Freundlich isotherms, while the adsorption kinetics was studied using Lagergren pseudo-first-order and pseudo-second-order kinetics models. The adsorption mechanism and kinetics were best described using Freundlich isotherm ($R^2=0.9917$) and Lagergren pseudo-second-order ($R^2=0.9874$), respectively. The adsorption capacity at equilibrium was found to be 0.4749 mg/g with the adsorption parameters as described (10 mg GO-MIP, 1 mL neopterin 10 mg/L). The neopterin adsorption method validation was performed via a non-matrix-matched calibration method. The limit of detection (LOD) and the limit of quantitation (LOQ) of the proposed adsorption method were determined to be 0.9126 mg/L and 3.042 mg/L respectively with linearity range (LR) of 1-10 mg/L. The synthesized GO-MIP showed promising adsorption performance towards neopterin and could be developed into a neopterin sensor in the future.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
Sebagai memenuhi keperluan untuk ijazah Master Sains

**SINTESIS DAN PENCIRIAN GRAFIN OKSIDA-POLIMER MOLEKUL
PENCETAKAN DAN PENJERAPAN TERHADAP NEOPTERIN DALAM
LARUTAN AKUEUS**

Oleh

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Neopterin adalah sejenis biopenanda yang berguna untuk pengesanan penyakit malignan. Sungguhpun begitu, hanya sedikit kajian telah dijalankan ke atas penyerapan neopterin menggunakan bahan hibrid grafin oksida-polimer seperti grafin oksida-polimer molekul pencetakan (GO-MIP). Tujuan kajian ini adalah untuk mensintesis dan mencirikan GO-MIP dan butiran lengkap ke atas kajian penyerapan terhadap neopterin telah dibincangkan dengan teliti. GO-MIP dengan neopterin sebagai templat telah disintesis melalui kaedah radikal pempolimeran bebas, asid metakrilik (MAA) sebagai monomer, etelina glikol dimetakrilik (EGDMA) sebagai pemaut silang, ammonium persulfat (APS) sebagai pemula, dan nisbah 8/2 v/v larutan dimetilsulfoksida/asetonitril (DMSO/ACN) sebagai pelarut porogen. Nisbah molar sasaran templat, monomer, dan pemaut silang yang telah digunakan adalah 1:4:16 masing-masing. Reflux telah dijalankan pada 50 °C untuk 24 jam di bawah atmosfera nitrogen lengai. Tapak pengikat neopterin telah diperolehi dengan merobohkan templat neopterin daripada GO-MIP yang telah disintesis melalui pembasuhan asid. Grafin oksida tiada peneraan polimer (GO-NIP) tanpa peneraan neopterin telah disediakan dengan kaedah yang sama. Keputusan spektroskopi transformasi Fourier inframerah (FTIR) untuk GO-MIP telah menunjukkan puncak regangan hidroksida -OH yang kurang amat dan puncak regangan yang lebih amat untuk karboksilik CO- berbanding dengan GO. Analisis unsur CHNS untuk GO-MIP telah menunjukkan bahawa peratusan karbon wt% adalah 4.37 % lebih rendah dan peratusan hidrogen wt% adalah 3.702 % lebih tinggi berbanding dengan GO. Puncak penguraian terma untuk GO-MIP boleh dicerap pada 400 °C dengan analisis gravimetrik terma (TGA) disebabkan oleh penguraian terma MAA. Imej-imej mikroskopi imbasan elektron pancaran medan (FESEM) dan mikroskopi penghantaran elektron (TEM) telah menunjukkan kehadiran tapak pengikatan neopterin dan morfologi berstruktur hibrid GO-MIP. Kajian penyerapan neopterin telah dijalankan dengan campuran 10 mg GO-MIP dan 1 mL 10 mg/L larutan standard neopterin. Campuran telah dikacau pada

300 rpm selama 60 minit, kemudian telah dituras menggunakan turas picagari nilon dan supernatant telah dianalisa menggunakan kromatografi cecair prestasi tinggi dengan pengesanan pendafluor (HPLC-FLD). GO-MIP telah menjerap 45.13 % neopterin sementara GO-NIP telah menjerap hanya 23.48 %, kira-kira dua kali jumlah neopterin berbanding dengan kaunterpart tiada peneraan. GO-MIP juga telah menunjukkan kepilihan neopterin yang bagus di bawah kesan sebatian analog 6-biopterin, di mana GO-MIP telah menjerap 34.63 % neopterin dan 18.64 % 6-biopterin. Mekanisma penjerapan statik telah dikaji menggunakan Isoterm Langmuir dan Freundlich, dan penjerapan kinetik telah dikaji menggunakan model kinetik Lagergren pseudo-tertib pertama dan pseudo-tertib kedua. Mekanisma penjerapan dan kinetik telah diperihalkan menggunakan Isoterm Freundlich ($R^2=0.9917$) dan Lagergren pseudo-tertib kedua ($R^2=0.9874$) masing-masing. Keseimbangan muatan penjerap telah ditemui pada 0.4749 mg/g dengan parameter penjerap seperti yang diperihalkan (10 mg GO-MIP, 1 mL neopterin 10 mg/L). Kaedah pengesanan penjerapan neopterin telah dilakukan melalui kaedah kalibrasi bukan matrik yang sama. Had pengesanan (LOD) dan had kuantitatif (LOQ) kaedah penjerap yang dibangunkan adalah 0.9126 mg/L dan 3.042 mg/L masing-masing dengan julat kelinearan (LR) 1-10 mg/L. GO-MIP yang telah disintesis telah menunjukkan prestasi penjerapan yang berprestasi terhadap neopterin dan boleh dibangunkan sebagai sensor neopterin pada masa akan datang.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

MIT	Molecular Imprinting Technology
GO-MIP	Graphene Oxide-Molecularly Imprinted Polymer
MAA	Methacrylic Acid
EGDMA	Ethylene Glycol Dimethacrylate
APS	Ammonium Persulfate
GO-NIP	Graphene Oxide-Non Imprinted Polymer
FTIR	Fourier-Transform Infrared Spectroscopy
GO	Graphene Oxide
TGA	Thermogravimetric Analysis
FESEM	Field Emission Scanning Electron Spectroscopy
TEM	Transmission Electron Spectroscopy
HPLC-FLD	High Performance Liquid Chromatography-Fluorescent Detector
LOD	Limit of Detection
LOQ	Limit of Quantitation
LR	Linearity Range
GTP	Guanosine Triphosphate
RIA	Radioimmunoassay
ELISA	Enzyme-Linked Immunosorbent Assay
HPLC	High Performance Liquid Chromatography
MIP	Molecularly Imprinted Polymer
SPE	Solid Phase Extraction
RAFT	Reversible Addition and Fragmentation Chain Transfer
UV-Vis	Ultraviolet-Visible Spectrometer
ATR	Attenuated Total Reflectance
q_e	Adsorption Capacity at Equilibrium
C_o	Initial Concentration
C_e	Equilibrium Concentration
q_m	Maximum Adsorption Capacity

K_L	Langmuir Constant
K_F	Freundlich Constant
n	Freundlich Adsorption Favorability
q_t	Adsorption Capacity at Time t
k_1	Pseudo-First-Order Rate Constant
k_2	Pseudo-Second-Order Rate Constant
FLD	Fluorescent Detector
σ	Standard Error of Estimate
s	Slope of Calibration
DTA	Differential Thermal Analysis
R^2	Correlation of Coefficient

CHAPTER 1

INTRODUCTION

1.1 Research background

Molecular imprinting technology (MIT) involves the development of a polymer with molecularly imprinted binding sites of specific functional group, shape and size distribution which are highly selective toward the target template molecule, namely a MIP (Chen, Wang, Lu, Wu, & Li, 2016). The fundamental working principle of MIP is very akin to the lock-and-key mechanism famously found in enzymatic reactions. By introducing the desired template molecule into the polymer during polymerization process, the template molecule would leave behind empty cavities upon template removal which serves as highly selective molecular binding sites for the template (Haupt, Linares, Bompert, & Tse, 2013). Due to the ease of preparation, flexibility, and effectiveness of MIP, they are widely researched and utilized in various molecular imprinting studies, adsorption studies of specific compounds, molecular detection studies, and more (Altintas et al., 2015; Boulanouar, Mezzache, Combès, & Pichon, 2018; Daoud Attieh, Zhao, Elkak, Falcimaigne-Cordin, & Haupt, 2017; Erdőssy, Horváth, Yarman, Scheller, & Gyurcsányi, 2016; Zamora-Gálvez et al., 2016).

One of the most common MIP synthesis method is the free radical polymerization method. The method is based on Mosbach's idea of a "biochemist's" approach where most biological molecular recognition are non-covalent in nature (Arshady & Mosbach, 1981; Wackerlig & Lieberzeit, 2015). Thus, a self-assembled MIP synthesis method which promotes non-covalent interaction between functional monomer and template molecule is desired. Due to its simplicity and flexibility, MIP synthesized using free radical polymerization method allows for the use of various functional monomers as well as template molecules.

In order to further improve and enhance the performance of MIP, modification or functionalization of MIP has been heavily explored in recent years. One of the most common modification of MIP is the addition of GO into the polymer, forming a GO-MIP hybrid material as a result. Consisting of sp^2 hybridized carbon atoms arranged in a honeycomb lattice structure, GO is a monolayer graphitic nanomaterial which possesses a range of highly labile oxygen functional groups with high specific surface area and high Young's modulus measurement (Tan et al., 2017). Due to its high specific surface area, the introduction of monolayer GO allows the fabrication of MIP across the surface of the nanomaterials, bypassing the general limitations of bulk polymerization (Luo et al., 2017). This results in the optimization of the MIP structure, where the template molecules reside at the surface or near the surface of the material instead of being trapped inside a typical bulky MIP (Anirudhan, Deepa, & Stanly, 2019).

In this study, a GO-MIP would be synthesized with neopterin selected as the desired template molecule. Neopterin is a chemical compound which belongs to the chemical class known as pteridines. It is a product from the catabolic reaction of guanosine triphosphate (GTP), a type of purine nucleotide, occurring in human macrophages. The production of neopterin is stimulated by cytokine interferon-gamma, a type of cytokine which are produced by T helper cells. Since the primary function of T helper cells revolve around cellular or adaptive immune response activation, elevated concentrations of neopterin are usually detected in patients suffering from but not limited to diseases such as HIV-1, coronary atherosclerotic heart disease, chronic obstructive pulmonary disease, renal allograft rejections, and tuberculosis (Abdel, Adawy, & Sayed, 2016; Carey et al., 2013; Cesur et al., 2014; Lyu, Jiang, & Dai, 2015). Owing to its unique biosynthesis mechanism involving cytokine interferon-gamma stimulation, neopterin has become a vital biomarker for laboratory detection and diagnosis of various malignant diseases (Sucher et al., 2010).

1.2 Problem statement

Although free radical polymerization is easy to execute and the synthesized MIP is suitable to a wide range of functional monomers and template molecules, MIP synthesized with free radical polymerization method suffers from a few major drawbacks. Due to the uncontrollable nature of free radical polymerization, the binding site distribution of the synthesized MIP is heterogeneous, with varying affinities towards the target template. This results in nonhomogeneous molecular cavities, thus leading to slower response time due to the difference in affinities of binding sites, and low reproducibility of binding performance (Zeng, Wang, Liu, Kong, & Nie, 2012). Besides, MIP synthesized with conventional free radical polymerization suffers from incomplete template removal and poor binding site accessibility due to the imprinted sites located deep within the bulky polymer structure (Luo, Gao, Tan, Wei, & Liu, 2016).

Another major problem regarding this research is the lack of neopterin adsorption study with the use of GO-MIP or any other functionalized variants of MIP in general. One of the related research is the work published by Del Sole and co. in 2013, in which it detailed the determination of neopterin with the use of MIPs synthesized using various monomers (Del Sole et al., 2013). Another research of interest is done by Sharma and co. in 2016, where they developed a potentiometric chemosensor for neopterin detection using electrochemically synthesized MIP as the sensing unit (Sharma et al., 2016). However, no report regarding GO-MIP synthesis and adsorption study with neopterin as the template molecule is published at the time of concluding the laboratory work and writing this thesis.

1.3 Research objectives

The overall aim of this research is to synthesize and characterize a GO-MIP using neopterin as the template molecule for neopterin adsorption study. The research objectives are as follows:

- a) To synthesize GO-MIP using neopterin as the template molecule and characterize GO-MIP with various characterization instruments.
- b) To study the neopterin adsorption behavior on GO-MIP using Langmuir and Freundlich adsorption isotherms with HPLC-FLD as the method of detection.
- c) To determine the neopterin adsorption kinetics on GO-MIP using Lagergren pseudo-first-order and pseudo-second-order kinetic models with HPLC-FLD as the method of detection.

1.4 Scope of research

Synthesis and preparation of a GO-MIP using selected materials was studied. The synthesized GO-MIP was characterized using FT-IR, CHNS elemental analysis, TGA, FESEM, and TEM. A neopterin adsorption method was developed using the synthesized GO-MIP as the adsorbent. A simple HPLC-FLD method for the detection of neopterin was developed. The adsorption performance of GO-MIP towards neopterin was studied via comparison with an analogue compound, 6-biopterin. The adsorption mechanism and kinetics of neopterin onto GO-MIP was studied with the use of Langmuir isotherm, Freundlich isotherm, and Lagergren kinetic models.

1.5 Significance of research

In recent years, the use of MIP or functionalized MIPs to determine various organic compounds has been widely discussed. However, there has yet to be a publication detailing the synthesis of a GO-MIP for neopterin adsorption study. In a similar fashion, multiple assay studies of neopterin have been published utilizing various sample preparation and HPLC methods but not GO-MIP. Thus, a research about neopterin adsorption on GO-MIP seems feasible.

In this study, a GO-MIP using neopterin as the target template was synthesized with free radical polymerization method, where the disadvantages of MIP synthesized using free radical polymerization would be solved with the addition of GO, and adsorption study of neopterin onto GO-MIP were studied extensively with the use of HPLC-FLD. This research will showcase a simple free radical synthesis method for GO-MIP preparation with the use of neopterin as the template molecule. The adsorption study of neopterin onto GO-MIP will provide a significant insight into the adsorption performance, as well as the adsorption behavior and kinetics of the process. In short, this research will open the path to future adsorption studies of GO-MIP with neopterin as the template molecule.

1.6 Thesis outline

This thesis consists of 5 chapters. Chapter 1 covers research background, problem statement, research objectives, scope of research, and significant of research. Chapter 2 covers the literature review of this thesis, with detailed discussion on neopterin and its role as a biomarker in diagnosis of various diseases, detection methods of neopterin including immunoassay and liquid chromatography methods, and detailed discussion on MIP as well as GO-MIP hybrid material. Chapter 3 describes the preparation of GO powder, synthesis of GO-MIP, preparation of neopterin standard solution, adsorption method of neopterin using GO-MIP as the adsorbent, and adsorption study with Langmuir, Freundlich, and Lagergren kinetics. Chapter 4 details the experimental result and data of GO-MIP synthesis and characterizations, the adsorption study including adsorption selectivity and contact time optimization, adsorption mechanism analysis from Langmuir, Freundlich, and Lagergren kinetics model, as well as method validation data including LOD and LOQ. Chapter 5 summarizes the thesis by presenting the overall conclusions and also suggestions for future research directions.

REFERENCES

- Abdel, A., Adawy, Z., & Sayed, M. (2016). Role of neopterin among COPD patients. *Egyptian Journal of Chest Diseases and Tuberculosis*, 65(1), 23–27. <https://doi.org/10.1016/j.ejcdt.2015.11.010>
- Altintas, Z., Gittens, M., Guerreiro, A., Thompson, K.-A., Walker, J., Piletsky, S., & Tothill, I. E. (2015). Detection of Waterborne Viruses Using High Affinity Molecularly Imprinted Polymers. *Analytical Chemistry*, 87(13), 6801–6807. <https://doi.org/10.1021/acs.analchem.5b00989>
- Anirudhan, T. S., Deepa, J. R., & Stanly, N. (2019). Fabrication of a molecularly imprinted silylated graphene oxide polymer for sensing and quantification of creatinine in blood and urine samples. *Applied Surface Science*, 466, 28–39. <https://doi.org/https://doi.org/10.1016/j.apsusc.2018.10.001>
- Arshady, R., & Mosbach, K. (1981). Synthesis of substrate-selective polymers by host-guest polymerization. *Die Makromolekulare Chemie*, 182(2), 687–692. <https://doi.org/10.1002/macp.1981.021820240>
- Aulitzky, W. E., Tilg, H., Niederwieser, D., Riccabona, G., Obendorf, L., Margreiter, R., Pfaller, W., & Huber, C. (1988). Comparison of serum neopterin levels and urinary neopterin excretion in renal allograft recipients. *Clinical Nephrology*, 29(5), 248–252. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/3293855>
- Boulanouar, S., Mezzache, S., Combès, A., & Pichon, V. (2018). Molecularly imprinted polymers for the determination of organophosphorus pesticides in complex samples. *Talanta*, 176, 465–478. <https://doi.org/https://doi.org/10.1016/j.talanta.2017.08.067>
- Brodie, B. C. (1860). Sur le poids atomique du graphite. *Ann. Chim. Phys*, 59(466), e472.
- Carey, B. S., Jain, R., Adams, C. L., Wong, K. Y., Shaw, S., Tse, W. Y., & Kaminski, E. R. (2013). Serum neopterin as an indicator of increased risk of renal allograft rejection. *Transplant Immunology*, 28(2–3), 81–85. <https://doi.org/10.1016/j.trim.2013.02.001>
- Cesur, S., Aslan, T., Hoca, N. T., Çimen, F., Tarhan, G., Çifçi, A., Ceyhan, I., & Şipit, T. (2014). Clinical importance of serum neopterin level in patients with pulmonary tuberculosis. *International Journal of Mycobacteriology*, 3(1), 5–8. <https://doi.org/10.1016/j.ijmyco.2014.01.002>
- Ceylan, M. F., Uneri, O. S., Guney, E., Ergin, M., Alisik, M., Goker, Z., Senses Dinc, G., Karaca Kara, F., & Erel, O. (2014). Increased levels of serum neopterin in attention deficit/hyperactivity disorder (ADHD). *Journal of Neuroimmunology*, 273(1), 111–114. <https://doi.org/https://doi.org/10.1016/j.jneuroim.2014.06.002>
- Chan, C. P. Y., Choi, J. W. Y., Cao, K.-Y., Wang, M., Gao, Y., Zhou, D.-H., Di, B., Xu, H.-F., Leung, M.-F., Bergmann, A., Lehmann, M., Nie, Y.-M., Cautherley, G. W. H., Fuchs, D., Renneberg, R., & Zheng, B.-J. (2006).

- Detection of serum neopterin for early assessment of dengue virus infection. *Journal of Infection*, 53(3), 152–158. <https://doi.org/10.1016/J.JINF.2005.11.008>
- Chang, L., Wu, S., Chen, S., & Li, X. (2011). Preparation of graphene oxide-molecularly imprinted polymer composites via atom transfer radical polymerization. *Journal of Materials Science*, 46(7), 2024–2029. <https://doi.org/10.1007/s10853-010-5033-z>
- Chen, L., Wang, X., Lu, W., Wu, X., & Li, J. (2016). Molecular imprinting: perspectives and applications. *Chemical Society Reviews*, 45(8), 2137–2211. <https://doi.org/10.1039/C6CS00061D>
- Cui, P., Lee, J., Hwang, E., & Lee, H. (2011). One-pot reduction of graphene oxide at subzero temperatures. *Chemical Communications*, 47(45), 12370. <https://doi.org/10.1039/c1cc15569e>
- Daoud Attieh, M., Zhao, Y., Elkak, A., Falcimaigne-Cordin, A., & Haupt, K. (2017). Enzyme-Initiated Free-Radical Polymerization of Molecularly Imprinted Polymer Nanogels on a Solid Phase with an Immobilized Radical Source. *Angewandte Chemie*, 129(12), 3387–3391. <https://doi.org/10.1002/ange.201612667>
- Del Sole, R., Scardino, A., Lazoi, M. R., Mergola, L., Scorrano, S., & Vasapollo, G. (2013). A molecularly imprinted polymer for the determination of neopterin. *Microchimica Acta*, 180(15–16), 1401–1409.
- Dickey, F. H. (1949). The Preparation of Specific Adsorbents. *Proceedings of the National Academy of Sciences* (Vol. 35). Retrieved from <http://www.pnas.org/content/pnas/35/5/227.full.pdf>
- Duan, F., Chen, C., Wang, G., Yang, Y., Liu, X., & Qin, Y. (2014). Efficient adsorptive removal of dibenzothiophene by graphene oxide-based surface molecularly imprinted polymer. *RSC Advances*. Retrieved from <http://pubs.rsc.org/en/content/articlehtml/2014/ra/c3ra45354e>
- Elbarbary, N. S., Ismail, E. A. R., El-Hilaly, R. A., & Ahmed, F. S. (2018). Role of neopterin as a biochemical marker for peripheral neuropathy in pediatric patients with type 1 diabetes: Relation to nerve conduction studies. *International Immunopharmacology*, 59, 68–75. <https://doi.org/https://doi.org/10.1016/j.intimp.2018.03.026>
- Erdőssy, J., Horváth, V., Yarman, A., Scheller, F. W., & Gyurcsányi, R. E. (2016). Electrosynthesized molecularly imprinted polymers for protein recognition. *TrAC Trends in Analytical Chemistry*, 79, 179–190. <https://doi.org/https://doi.org/10.1016/j.trac.2015.12.018>
- Ettre, L. S., & Sakodinskii, K. I. (1993). M. S. Tswett and the discovery of chromatography II: Completion of the development of chromatography (1903–1910). *Chromatographia*, 35(5–6), 329–338. <https://doi.org/10.1007/BF02277520>
- Foo, K. Y., & Hameed, B. H. (2010). Insights into the modeling of adsorption isotherm systems. *Chemical Engineering Journal*, 156(2).

<https://doi.org/10.1016/j.cej.2009.09.013>

- Fuchs, D., Murr, C., Widner, B., Wirleitner, B., & Fuchs, D. (2002). Neopterin as a Marker for Immune System Activation. *Current Drug Metabolism*, 3, 175–187. <https://doi.org/10.2174/1389200024605082>
- Geisler, S., Peter, M., Kathrin, B., Harald, S., Dietmar, F., & M, G. J. (2015). Serum tryptophan, kynurenine, phenylalanine, tyrosine and neopterin concentrations in 100 healthy blood donors. *Pteridines*. <https://doi.org/10.1515/pterid-2014-0015>
- Hamerlinck, F. F. V. (1999). Neopterin: a review. *Experimental Dermatology*, 8(3), 167–176. <https://doi.org/10.1111/j.1600-0625.1999.tb00367.x>
- Haupt, K., Linares, A. V., Bompert, M., & Tse, B. (2013). Molecularly imprinted polymers. *TripleC*, 11(1), 13–35. <https://doi.org/10.1007/128>
- Hausen, A., Fuchs, D., Grünewald, K., Huber, H., König, K., & Wachter, H. (1981). Urinary neopterin as marker for haematological neoplasias. *Clinica Chimica Acta*, 117(3), 297–305.
- Hausen, A., Fuchs, D., König, K., & Wachter, H. (1982). Determination of neopterin in human urine by reversed-phase high-performance liquid chromatography, 227, 61–70.
- Hopkins, F. G. (1889). Note on a yellow pigment in butterflies. *Nature*, 40, 335.
- Huber, C., Batchelor, J. R., Fuchs, D., Hausen, A., Lang, A., Niederwieser, D., Reibnegger, G., Swetly, P., Troppmair, J., & Wachter, H. (1984). Immune response-associated production of neopterin. Release from macrophages primarily under control of interferon-gamma, 160(1), 310–316.
- Humbert, M., Delattre, R. M., Cerrina, J., Dartevelle, P., Simonneau, G., & Emilie, D. (1993). Serum Neopterin After Lung Transplantation. *Chest*, 103(2), 449–454. <https://doi.org/https://doi.org/10.1378/chest.103.2.449>
- Hummers, W. S., & Offeman, R. E. (1958). Preparation of Graphitic Oxide. *Journal of the American Chemical Society*, 80(6), 1339. <https://doi.org/10.1021/ja01539a017>
- Hyun Jung, K., & Spivak, D. A. (2003). New Insight into Modeling Non-Covalently Imprinted Polymers. <https://doi.org/10.1021/JA0361502>
- Ilhan, F., Akbulut, H., Karaca, I., Godekmerdan, A., Ilkay, E., & Bulut, V. (2005). Procalcitonin, c-reactive protein and neopterin levels in patients with coronary atherosclerosis. *Acta Cardiologica*, 60(4), 361–365. <https://doi.org/10.2143/AC.60.4.2004983>
- Kappock, T. J., & Caradonna, J. P. (1996). Pterin-Dependent Amino Acid Hydroxylases. *Chemical Reviews*, 96(7), 2659–2756. <https://doi.org/10.1021/cr9402034>
- Khan, S., Bhatia, T., Trivedi, P., Satyanarayana, G. N. V., Mandrah, K., Saxena, P. N., Mudiam, M. K. R., & Roy, S. K. (2016). Selective solid-phase extraction using molecularly imprinted polymer as a sorbent for the analysis

- of fenarimol in food samples. *Food Chemistry*, 199, 870–875. <https://doi.org/10.1016/j.foodchem.2015.12.091>
- Krcmova, L. K., Cervinkova, B., Solichova, D., Sobotka, L., Hansmanova, L., Melichar, B., & Solich, P. (2015). Fast and sensitive HPLC method for the determination of neopterin, kynurenine and tryptophan in amniotic fluid, malignant effusions and wound exudates. *Bioanalysis*, 7(21), 2751–2762. <https://doi.org/10.4155/bio.15.175>
- Li, W., Tang, X. Z., Zhang, H. Bin, Jiang, Z. G., Yu, Z. Z., Du, X. S., & Mai, Y. W. (2011). Simultaneous surface functionalization and reduction of graphene oxide with octadecylamine for electrically conductive polystyrene composites. *Carbon*, 49(14), 4724–4730. <https://doi.org/10.1016/j.carbon.2011.06.077>
- Li, Y., Li, X., Dong, C., Qi, J., & Han, X. (2010). A graphene oxide-based molecularly imprinted polymer platform for detecting endocrine disrupting chemicals. *Carbon*. Retrieved from <http://www.sciencedirect.com/science/article/pii/S0008622310003751>
- Lim, F. K., & Holdsworth, I. C. (2018). Effect of Formulation on the Binding Efficiency and Selectivity of Precipitation Molecularly Imprinted Polymers. *Molecules*. <https://doi.org/10.3390/molecules23112996>
- Linus Pauling, B., & Campbell, D. H. (1942). The Manufacture of Antibodies In Vitro. *Journal of Experimental Medicine*. <https://doi.org/10.1084/jem.76.2.211>
- Luo, J., Gao, Y., Tan, K., Wei, W., & Liu, X. (2016). Preparation of a Magnetic Molecularly Imprinted Graphene Composite Highly Adsorbent for 4-Nitrophenol in Aqueous Medium. *ACS Sustainable Chemistry & Engineering*, 4(6), 3316–3326. <https://doi.org/10.1021/acssuschemeng.6b00367>
- Luo, J., Huang, J., Wu, Y., Sun, J., Wei, W., & Liu, X. (2017). Synthesis of hydrophilic and conductive molecularly imprinted polyaniline particles for the sensitive and selective protein detection. *Biosensors and Bioelectronics*, 94, 39–46. <https://doi.org/https://doi.org/10.1016/j.bios.2017.02.035>
- Lyu, Y., Jiang, X., & Dai, W. (2015). The roles of a novel inflammatory neopterin in subjects with coronary atherosclerotic heart disease. *International Immunopharmacology*, 24(2), 169–172. <https://doi.org/10.1016/j.intimp.2014.11.013>
- Martin, A. J., & Synge, R. L. (1941). A new form of chromatogram employing two liquid phases: A theory of chromatography. 2. Application to the micro-determination of the higher monoamino-acids in proteins. *The Biochemical Journal*, 35(12), 1358–1368. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/16747422>
- Mehmet, A., Halil, Y., Tuncer, C., Irfan, S., Muzaffer, O., Ibrahim, A., Fevzi, N. A., Yasemin, G. K., Erdinc, C., & Emin, O. A. (2012). Comparison of two different HPLC methods and elisa method for measurement of serum neopterin. *Journal of Investigational Biochemistry*, 1(1), 43–47.

<https://doi.org/10.5455/jib.20120815013746>

- Melichar, B., Gregor, J., Solichová, D., Lukes, J., Tichý, M., & Pidman, V. (1994). Increased urinary neopterin in acute myocardial infarction. *Clinical Chemistry*, 40(2). Retrieved from <http://clinchem.aaccjnls.org/content/40/2/338.short>
- Melmed, R. N., Taylor, J. M., Detels, R., Bozorgmehri, M., & Fahey, J. L. (1989). Serum neopterin changes in HIV-infected subjects: indicator of significant pathology, CD4 T cell changes, and the development of AIDS. *Journal of Acquired Immune Deficiency Syndromes*, 2(1), 70–76. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/2783972>
- Nagatsu, T., Sawada, M., Yamaguchi, T., Sugimoto, T., Matsuura, S., Akino, M., Nakazawa, N., & Ogawa, H. (1984). Radioimmunoassay for neopterin in body fluids and tissues. *Analytical Biochemistry*, 141(2), 472–480. [https://doi.org/10.1016/0003-2697\(84\)90073-3](https://doi.org/10.1016/0003-2697(84)90073-3)
- Ogiwara, S., Kiuchi, K., Nagatsu, T., Teradaira, R., Nagatsu, I., Fujita, K., & Sugimoto, T. (1992). Highly sensitive, specific enzyme-linked immunosorbent assay of neopterin and biopterin in biological samples. *Clinical Chemistry*, 38(10). Retrieved from <http://clinchem.aaccjnls.org/content/38/10/1954.short>
- Pauling, L. (1940). A Theory of the Structure and Process of Formation of Antibodies. *Journal of the American Chemical Society*, 62(10), 2643–2657. <https://doi.org/10.1021/ja01867a018>
- Polyakov, M. V. (1931). Adsorption properties and structure of silica gel. *Zhur Fiz Khim*, 2, 799–805.
- Pourakbari, B., Mamishi, S., Zafari, J., Khairkhah, H., Ashtiani, M. H., Abedini, M., Afsharpaiman, S., & Rad, S. S. (2010). Evaluation of procalcitonin and neopterin level in serum of patients with acute bacterial infection. *Brazilian Journal of Infectious Diseases*, 14(3), 252–255. <https://doi.org/10.1590/S1413-86702010000300009>
- Prommegger, R., Widner, B., Murr, C., Unger, A., Fuchs, D., & Salzer, G. M. (2000). Neopterin: a prognostic variable in operations for lung cancer. *The Annals of Thoracic Surgery*, 70(6), 1861–1864. [https://doi.org/10.1016/S0003-4975\(00\)01840-3](https://doi.org/10.1016/S0003-4975(00)01840-3)
- Purmann, R. (1941). Die Synthese des Xanthopterins. Über die Flügelpigmente der Schmetterlinge. X3. *Justus Liebigs Annalen Der Chemie*, 546(1-2), 98–102.
- Razak, M. R., Yusof, N. A., Haron, M. J., Ibrahim, N., Mohammad, F., Kamaruzaman, S., & Al-Lohedan, H. A. (2018). Iminodiacetic acid modified kenaf fiber for waste water treatment. *International Journal of Biological Macromolecules*, 112, 754–760. <https://doi.org/https://doi.org/10.1016/j.jbiomac.2018.02.035>
- Reibnegger, G., Hetzel, H., Fuchs, D., Fuith, L. C., Hausen, A., Werner, E. R., & Wachter, H. (1987). Clinical significance of neopterin for prognosis and

- follow-up in ovarian cancer. *Cancer Research*, 47(18), 4977–4981. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/3621185>
- Reibnegger, Gilbert, Egg, D., Fuchs, D., Günther, R., Hausen, A., Werner, E. R., & Wachter, H. (1986). Urinary neopterin reflects clinical activity in patients with rheumatoid arthritis. *Arthritis & Rheumatism*, 29(9), 1063–1070. <https://doi.org/10.1002/art.1780290902>
- Rembold, H., & Buschmann, L. (1963a). Struktur und synthese des neopterin. *Chemische Berichte*, 96(5), 1406–1410.
- Rembold, H., & Buschmann, L. (1963b). Untersuchungen über die Pteridine der Bienenpuppe (*Apis mellifica*). *Justus Liebigs Annalen Der Chemie*, 662(1), 72–82.
- Robert, J. U. II., Sarah, C. B., Yizhao, C., Ripal, N. S., & Ken D. S. (2001). Characterization of Molecularly Imprinted Polymers with the Langmuir–Freundlich Isotherm. <https://doi.org/10.1021/AC0105686>
- Sakurai, A., & Goto, M. (1967, January). Neopterin: Isolation from human urine.
- Schild, H. G. (1993). Thermal Degradation of Poly (methacrylic acid): Further Studies Applying TGA/ FTIR. *Journal of Polymer Science Part A: Polymer Chemistry*, 31(9), 2403–2405.
- Schniepp, H. C., Li, J., Mcallister, M. J., Sai, H., Herrera-alonso, M., Adamson, D. H., Prud, R. K., Car, R., Saville, D. A., & Aksay, I. A. (2006). Functionalized Single Graphene Sheets Derived from Splitting Graphite Oxide. *ACS Publications*, 2, 8535–8539. <https://doi.org/10.1021/jp060936f>
- Sellergren, B. (1989). Molecular imprinting by noncovalent interactions. Enantioselectivity and binding capacity of polymers prepared under conditions favoring the formation of template complexes. *Die Makromolekulare Chemie*, 190(11), 2703–2711. <https://doi.org/10.1002/macp.1989.021901104>
- Sharma, P. S., Wojnarowicz, A., Sosnowska, M., Benincori, T., Noworyta, K., D'Souza, F., & Kutner, W. (2016). Potentiometric chemosensor for neopterin, a cancer biomarker, using an electrochemically synthesized molecularly imprinted polymer as the recognition unit. *Biosensors and Bioelectronics*, 77, 565–572. <https://doi.org/10.1016/j.bios.2015.10.013>
- Shrivastava, A., & Gupta, V. B. (2011). Methods for the determination of limit of detection and limit of quantitation of the analytical methods. *Chronicles of Young Scientists*, 2(1), 21–25. <https://doi.org/10.4103/2229-5186.79345>
- Slazyk, W., & Spierto, F. (1990). Liquid-chromatographic measurement of biopterin and neopterin in serum and urine. *Clinical Chemistry*. Retrieved from <http://clinchem.aaccjnls.org/content/36/7/1364.short>
- Spivak, D. A. (2005). Optimization, evaluation, and characterization of molecularly imprinted polymers. *Advanced Drug Delivery Reviews*, 57(12), 1779–1794. <https://doi.org/10.1016/j.addr.2005.07.012>

- Staudenmaier, L. (1898). Verfahren zur Darstellung der Graphitsäure. *Berichte Der Deutschen Chemischen Gesellschaft*, 31(2), 1481–1487. <https://doi.org/10.1002/cber.18980310237>
- Strohmaier, W., Redl, H., Schlag, G., & Inthorn, D. (1987). D-erythro-neopterin plasma levels in intensive care patients with and without septic complications. *Critical Care Medicine*, 15(8), 757–760. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/3497012>
- Sucher, R., Schroecksadel, K., Weiss, G., Margreiter, R., Fuchs, D., & Brandacher, G. (2010). Neopterin, a prognostic marker in human malignancies. *Cancer Letters*, 287(1), 13–22. <https://doi.org/10.1016/j.canlet.2009.05.008>
- Tan, C., Cao, X., Wu, X.-J., He, Q., Yang, J., Zhang, X., Chen, J., Zhao, W., Han, S., Nam, G.-H., Sindoro, M., & Zhang, H. (2017). Recent Advances in Ultrathin Two-Dimensional Nanomaterials. *Chemical Reviews*, 117(9), 6225–6331. <https://doi.org/10.1021/acs.chemrev.6b00558>
- Turgan, N., Habif, S., Parildar, Z., Özmen, D., Mutaf, I., Erdener, D., & Bayindir, O. (2001). Association between homocysteine and neopterin in healthy subjects measured by a simple HPLC-fluorometric method. *Clinical Biochemistry*, 34(4), 271–275. [https://doi.org/https://doi.org/10.1016/S0009-9120\(01\)00226-0](https://doi.org/https://doi.org/10.1016/S0009-9120(01)00226-0)
- Wachter, H., Hausen, A., & Grassmayr, K. (1979). Erhöhte Ausscheidung von Neopterin im Harn von Patienten mit malignen Tumoren und mit Viruserkrankungen. *Hoppe Seylers Z Physiol Chem*, 360(12), 1957–1960.
- Wackerlig, J., & Lieberzeit, P. A. (2015). Molecularly imprinted polymer nanoparticles in chemical sensing – Synthesis, characterisation and application. *Sensors and Actuators B: Chemical*, 207, 144–157. <https://doi.org/https://doi.org/10.1016/j.snb.2014.09.094>
- Weiss, G., Willeit, J., Kiechl, S., Fuchs, D., Jarosch, E., Oberhollenzer, F., Reibnegger, G., Tilz, G. P., Gerstenbrand, F., & Wachter, H. (1994). Increased concentrations of neopterin in carotid atherosclerosis. *Atherosclerosis*, 106(2), 263–271. [https://doi.org/10.1016/0021-9150\(94\)90131-7](https://doi.org/10.1016/0021-9150(94)90131-7)
- Werner, E., Fuchs, D., & Hausen, A. (1987). Simultaneous determination of neopterin and creatinine in serum with solid-phase extraction and on-line elution liquid chromatography. *Clinical*. Retrieved from <http://www.clinchem.org/content/33/11/2028.short>
- Wieland, H., & Schöpf, C. (1925). Über den gelben Flügelfarbstoff des Citronenfalters (*Gonepteryx rhamni*). *Berichte Der Deutschen Chemischen Gesellschaft (A and B Series)*, 58(9), 2178–2183.
- Wulff, G., Kemmerer, R., Vietmeier, J., & Poll, H. G. (1982). Chirality of vinyl-polymers-the preparation of chiral cavities in synthetic-polymers. *Nouveau Journal De Chimie-New Journal Of Chemistry*, 6(12), 681–687.
- Zamora-Gálvez, A., Ait-Lahcen, A., Mercante, L. A., Morales-Narváez, E., Amine,

A., & Merkoçi, A. (2016). Molecularly Imprinted Polymer-Decorated Magnetite Nanoparticles for Selective Sulfonamide Detection. *Analytical Chemistry*, 88(7), 3578–3584. <https://doi.org/10.1021/acs.analchem.5b04092>

Zeng, H., Wang, Y., Liu, X., Kong, J., & Nie, C. (2012). Preparation of molecular imprinted polymers using bi-functional monomer and bi-crosslinker for solid-phase extraction of rutin. *Talanta*, 93, 172–181. <https://doi.org/10.1016/j.talanta.2012.02.008>

