



UNIVERSITI PUTRA MALAYSIA

**ZnO-NPs/rGO/SPCE-MODIFIED ELECTRODE FOR EARLY DETECTION
OF *Ganoderma boninense* IN OIL PALM TREES**

NURHAMIZAH BINTI RAHMAT

ITMA 2019 10



**ZnO-NPs/rGO/SPCE-MODIFIED ELECTRODE FOR EARLY DETECTION OF
Ganoderma boninense IN OIL PALM TREES**

By

NURHAMIZAH BINTI RAHMAT

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

October 2018

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia
in fulfilment of the requirements for the degree of Master of Science

**ZnO-NPs/rGO/SPCE-MODIFIED ELECTRODE FOR EARLY DETECTION OF
Ganoderma boninense IN OIL PALM TREES**

By

NURHAMIZAH BINTI RAHMAT

October 2018

Chairman : Professor Nor Azah Yusof, PhD
Faculty : Institute of Advanced Technology

Ganoderma boninense is a major disease that infects oil palm trees. This problem has caught researchers' attention all around the world, especially those from Asian countries as *Ganoderma boninense* is difficult to trace in its early stage. Undoubtedly, many researchers have lent a hand by suggesting ways to diagnose the disease through the introduction of many methods to detect symptoms related to the *Ganoderma* disease, including tomography, selective media, Indirect Enzyme-linked Immunosorbent Assay (ELISA) and polymerase chain reaction (PCR). However, these diagnostic techniques are less accurate, time-consuming and lead to cost increments. Therefore, another method to detect *Ganoderma boninense* at a faster, lower cost and higher accuracy was introduced which is through the use of electrochemical sensors. Initially, electrode modification was done by reducing graphene oxide (GO) to reduced graphene oxide (rGO). Subsequently, the addition of zinc oxide nanoparticle (ZnO-NPs) was electrodeposited on top of it using the electrodeposition method. Both modifiers were chosen as they have been acknowledged to have large surface areas to interact with analytes, and have great electroconductivity. In this study, this disposable modified SPCE was used as a sensing material in order to detect secondary metabolites acting as possible biomarkers for the *Ganoderma boninense* disease in healthy and infected leaves at the 14 and 30 day periods. It is because secondary metabolites are produced once oil palm trees are infected by *Ganoderma boninense*. The combination of physical and electrochemical characterisation of graphene oxide (GO) before and after reduction together with the deposition of zinc oxide nanoparticles (ZnO-NPs) were observed using field emission scanning electron microscopy (FESEM), energy dispersive x-ray (EDX), Fourier Transform Infrared Spectroscopy (FTIR), Raman spectroscopy and cyclic voltammetry (CV) to reveal the successful modification of ZnO-NPs/rGO/SPCE.

As a consequence, the electrochemical analysis presented a current increment in each modification higher than the bare SPCE due to electrocatalytic activity. Afterwards, the electrochemical signals from secondary metabolites were indicated from differential pulse voltammetry techniques (DPV) under optimum conditions. ZnO-NPs/rGO/SPCE established a good correlation coefficient (R^2) of both healthy and infected oil palm after 14 days and 30 days ($R^2 = 0.9691$, $R^2 = 0.9652$, $R^2 = 0.9905$ and $R^2 = 0.9710$) with detection limits of 1.52 ppm, 1.58 ppm, 3.20 ppm and 3.02 ppm respectively. Consequently, the outstanding sensing performance of ZnO-NPs/rGO/SPCE in the detection of secondary metabolites can be applied to control the *Ganoderma boninense* disease at the earlier stage before spreading out.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**ELEKTROD MODIFIKASI ZnO-NPs/rGO/SPCE UNTUK PENGESANAN
AWAL *Ganoderma boninense* BAGI POKOK KELAPA SAWIT**

Oleh

NURHAMIZAH BINTI RAHMAT

Oktober 2018

Pengerusi : Professor Nor Azah Yusof, PhD
Fakulti : Institut Teknologi Maju

Ganoderma boninense adalah penyakit utama yang menyerang pokok kelapa sawit. Hal ini menjadi tarikan kepada ramai penyelidik di seluruh dunia terutama penyelidik dari negara Asia kerana penyakit *Ganoderma boninense* sukar untuk dikesan pada peringkat awal jangkitan. Tidak dinafikan, ramai penyelidik cuba membantu untuk mengawal penyakit *Ganoderma* ini termasuklah memperkenalkan imbasan tomografi, kultur sel, penetapan kadar imunisorben taut-enzim (ELISA) dan reaksi berantai polimerase (PCR). Akan tetapi, teknik diagnostik tersebut kurang tepat, mengambil masa yang lama untuk bertindak balas dan kos yang agak tinggi. Maka, sebagai satu cara yang lain untuk mengesan penyakit *Ganoderma boninense* dengan tindak balas yang lebih cepat, lebih tepat dan kos yang lebih rendah adalah dengan memperkenalkan pengesanan elektrokimia. Sebagai pemulaan, modifikasi elektrod direncanakan dengan proses penurunan dari grafin oksida (GO) kepada grafin oksida teturunan (rGO). Seterusnya, nanozarah zink oksida (ZnO-NPs) terenap dengan kaedah elektroenantan di atasnya. Kedua-dua penguubah digunakan untuk modifikasi elektrod kerana terkenal dengan keluasan permukaan yang besar untuk bertindak balas dengan analit dan aktiviti elektrokatalitik yang tinggi. Elektrod modifikasi serbaguna SPCE ini dihasilkan bagi mengesan metabolit sekunder sebagai penanda bio dalam penyakit *Ganoderma boninense* diuji keatas daun yang sihat dan yang telah dijangkiti setelah 14 dan 30 hari. Ini kerana, metabolit sekunder akan terhasil secara spontan apabila diserang oleh *Ganoderma boninense*. Gabungan pencirian fizikal dan elektrokimia untuk GO sebelum dan selepas proses penurunan dan terenapan nanozarah zink oksida telah diuji menggunakan mikroskop elektron pengimbas pancaran medan (FE-SEM), spektroskopi tenaga serakan (EDX), fourier spektroskopi inframerah (FTIR), spektroskopi Raman, voltametri berkitar bagi memastikan modifikasi elektrod ZnO-NPs/rGO/SPCE berjaya. Hasilnya, elektrokimia analisis telah menunjukkan peningkatan arus bagi

setiap modifikasi di mana ianya lebih tinggi dari SPCE disebabkan ciri aktiviti elektrokatalitik. Selanjutnya, isyarat elektrokimia dari metabolit sekunder diterjemahkan melalui tindakbalas voltametri denyut dalam keadaan optimum. ZnO-NPs/rGO/SPCE memberi keputusan yang memberangsangkan untuk pekali kolerasi bagi daun sihat dan yang telah dijangkiti selepas 14 dan 30 hari iaitu ($R^2 = 0.9691$, $R^2 = 0.9652$, $R^2 = 0.9905$ and $R^2 = 0.9710$), manakala masing-masing menunjukkan had pengesanan 1.52 ppm, 1.58 ppm, 3.20 ppm and 3.02 ppm. Sebagai hasilnya, ZnO-NPs/rGO/SPCE memberi impak yang bagus dalam prestasi pengesanan berdasarkan metabolit sekunder dalam mengawal *Ganoderma boninense* pada peringkat awal daripada terus merebak.



ACKNOWLEDGEMENTS

Alhamdulillah, thanks to Allah, The Most Beneficent and The Most Merciful, as I have successfully completed my final year project entitled “ZnO-NPs/rGO/SPCE Modified Electrode for the Early Detection of *Ganoderma boninense* in Oil Palm Trees”. Although there were some difficulties faced during this research, I thank the Almighty God that I was able to finish this project with the help and blessings from Him and others.

Firstly, I would like to take this chance to express my gratitude to my main supervisor, Professor Dr. Nor Azah binti Yusof and special thanks to my co-supervisor, Associate Professor Dr. Wong Mui Yun from the Faculty of Agriculture for giving me the opportunity to do my Masters and guiding me in accomplishing all the work through their aspiring guidance, encouraging spirit and invaluable friendly advices which have led me throughout the project.

Besides that, I would like to express my warm thanks to the Director of Institute of Advanced Technology (ITMA), UPM, Associate Professor Dr. Mohd Nizar Hamidon, all members and staffs from ITMA and the Faculty of Science for their help which involved the use of applications, facilities and equipment during my Masters project.

My sincere gratitude goes to my beloved parents, Rahmat bin Othman and Rohaya binti Abd. Rashid, my husband,, Muhammad Afiq bin Misri, my brother, Mohamad Hazim Mustafa bin Rahmat, my sister-in-law, Fatin Shakila binti Mohad Zainuddin and my cute nephew, Muhammad Aisy Dasukee bin Mohammad Hazim Mustafa for their love, encouragement, advice and support which gave me undeniable strength throughout my studies.

Last but not least, I place my heartfelt gratefulness to all my friends, especially Fatimah, Zulaiha, Ainisah, Isshadiba, Ain and my research group members in BASL 103 for being great and wonderful team members. Thank you to those who have directly or indirectly helped me throughout this venture until its successful completion.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Nor Azah Yusof, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Wong Mui Yun, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of
Chairman of
Supervisory
Committee: _____

Signature: _____

Name of
Member of
Supervisory
Committee: _____

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xv
LIST OF APPENDICES	xvi
CHAPTER	
1 INTRODUCTION	
1.1 Background of study	1
1.2 Problem statement and research motivation	2
1.3 Objectives of study	3
1.3.1 General objective	3
1.3.2 Specific objective	3
1.4 Scope and limitation	3
2 LITERATURE REVIEW	
2.1 <i>Ganoderma sp.</i>	4
2.1.1 Phases of <i>Ganoderma</i> disease in oil palm trees	4
2.1.2 Current diagnostic methods to detect <i>Ganoderma boninense</i>	5
2.2 Secondary metabolites in plant	7
2.2.1 Secondary metabolites in oil palm trees	11
2.3 Detection of secondary metabolites	12
2.4 Electrochemical sensor	13
2.5 Modification of electrode using nanomaterial	14
2.5.1 Graphene, graphene oxide (GO) and reduced graphene oxide (rGO)	15
2.5.2 Zinc oxide nanoparticles (ZnO-NPs)	17
2.5.3 Application of zinc oxide nanoparticles as sensing material	18
2.6 The utilisation of ZnO-NPs –rGO as modifier in an electrochemical sensor	18
3 METHODOLOGY	
3.1 Chemicals and reagents	21
3.2 Cleaning of Screen Printed Carbon Electrode (SPCE)	22
3.3 Preparation of modified electrode	22

3.3.1	Preparation of GO/SPCE	22
3.3.2	Preparation of rGO/SPCE	22
3.3.3	Preparation of ZnO-NPs/rGO/SPCE	22
3.4	Preparation of general solutions	23
3.5	Apparatus and instrumentations	23
3.5.1	The electrochemical measurements procedure	23
3.5.2	Other apparatus and instrumentations	24
3.6	General characterisation of modified electrode	24
3.7	Optimisation of the SPCE electrode	25
3.8	Extraction of secondary metabolites from infected leaves	26
3.9	Optimisation of detection of secondary metabolites condition	27
4	RESULT AND DISCUSSION	
4.1	Characterisation of bare and modified SPCE	29
4.1.1	Modification of SPCE	29
4.1.2	The surface morphology of modified electrodes	31
4.1.3	The elemental analyses of modified electrodes	32
4.1.4	Fourier Transform Infrared Spectroscopy (FTIR) analyses in the reduction of graphene oxide	35
4.1.5	Raman Spectroscopy analysis for the reduction of graphene oxide	36
4.1.6	Electrochemical characterisation of modified electrodes	37
4.2	The optimisation of modified electrode	41
4.3	Electrochemical characterisation of modified SPCE for detection of secondary metabolites	45
4.3.1	Lifetime study, repeatability and reproducibility	51
4.3.2	Comparison between healthy and infected leaves	53
4.3.3	Effect of various concentrations of secondary metabolites on current response	54
5	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	
5.1	Summary and conclusions	57
5.2	Recommendation for future work	58
	REFERENCES	59
	APPENDICES	77
	BIODATA OF STUDENT	81
	LIST OF PUBLICATIONS	82

LIST OF TABLES

Table		Page
2.1	Classification of nanomaterial based on their nanomaterial dimensions	15
2.2	The utilisation of ZnO-rGO as modifier in electrochemical sensor	20
3.1	List of chemical and reagents used in this work	21
4.1	Regression value and effective surface area (A) for bare and all modified electrodes	41
4.2	Current change for healthy and infected leaves (14 and 30 days) on various concentrations of secondary metabolites	56

LIST OF FIGURES

Figure	Page	
2.1	Visualisation of the cross-section sonic tomography image by comparing with a real trunk.	5
2.2	The structure of oryzalexin A-F.	8
2.3	The structure of resveratrol.	9
2.4	The structure of GSH.	10
2.5	The structure of (a) linamarin (b) amygdalin.	11
2.6	A schematic diagram of a sensor.	13
2.7	The scale of nanoparticle compared to different types of larger-sized materials.	14
2.8	Reduction of GO to rGO.	16
2.9	Two types of common ZnO crystal structures.	17
3.1	SPCE based on three electrode system.	24
4.1	Modification of SPCE by reduction of GO to rGO.	29
4.2	Voltammogram of electrodeposition process for 0.1 M $\text{Zn}(\text{NO}_3)_2$ and 1.0 M KNO_3 as an electrolyte by chronoamperometry fixed at pH 6 and maintain at temperature 65°C	31
4.3	FE-SEM image of a) bare SPCE (100k magnification) b) GO/SPCE (100k magnification) c) rGO/SPCE (100k magnification) d) ZnO/rGO/SPCE (100k magnification).	32
4.4	EDX elemental analysis of (a) bare SPCE before cleaning (b) bare SPCE after cleaning (c) GO/SPCE (d) rGO/SPCE (e) ZnO-NPs/rGO/SPCE.	34
4.5	FTIR spectra for GO and rGO.	35
4.6	Raman spectra for (a) GO and (b) rGO.	36
4.7	Cyclic voltammetry (CV) response at different modified SPCE electrode; bare SPCE, GO/SPCE, rGO/SPCE and ZnO/rGO/SPCE in 100 mM $[\text{Fe}(\text{CN})_6]^{3-/4-}$ containing 100 mM PBS at scan rate of 100mV/s with a potential range of -0.7 V to 1.0 V.	38
4.8	Different scan rate (0.01 V, 0.02 V, 0.03 V, 0.04 V, 0.05 , 0.06 V, 0.07 V, 0.08 V, 0.09 V, 1.0 V) of (a) bare SPCE (b) GO/SPCE (c) rGO/SPCE (d) ZnO-NPs/rGO/SPCE in 100 mM $[\text{Fe}(\text{CN})_6]^{3-/4-}$ containing 100 mM PBS with a potential range of -0.7 V to 1.0 V and (e) reduction peak current of $[\text{Fe}(\text{CN})_6]^{3-/4-}$ for each SPCE.	40
4.9	Effect of amount of GO on modified electrode by CV response of potassium ferricyanide at scan rate of 100 mV/s.	42

4.10	Effect of $Zn(NO_3)_2$ concentration on modified electrode by CV response of potassium ferricyanide at scan rate of 100 mV/s	43
4.11	Effect of potential on modified electrode by CV response of potassium ferricyanide at scan rate of 100 mV/s.	44
4.12	Effect of deposition time on modified electrode by CV response of potassium ferricyanide at scan rate of 100 mV/s.	44
4.13	DPV analysis for ZnO-NPs/rGO/SPCE for the detection of secondary metabolites using scan rate of 100 mV/s.	45
4.14	The schematic illustration for detection of secondary metabolites.	46
4.15	DPV analysis for bare SPCE, GO/SPCE, rGO/SPCE and ZnO-NPs/rGO/SPCE in 0.1 M citrate buffer and 40 ppm secondary metabolites using scan rate of 100 mV/s.	47
4.16	The effect of supporting electrolyte on DPV analysis towards detection of secondary metabolites detection in various pH using scan rate of 100 mV/s.	49
4.17	The effect of pH on DPV analysis towards detection of secondary metabolites detection in citrate buffer at pH 3 using scan rate of 100 mV/s.	49
4.18	The effect of deposition time on DPV analysis towards detection of secondary metabolites detection in citrate buffer at pH 3 using scan rate of 100 mV/s.	50
4.19	The effect of deposition potential on DPV analysis towards detection of secondary metabolites detection in citrate buffer at pH 3 using scan rate of 100 mV/s.	51
4.20	Lifetime measurement (long term stability) of the electrode after 14 days and 21 days of storage for 40 ppm of secondary metabolites in 0.1 M citrate buffer at pH 3 using scan rate of 100 mV/s.	52
4.21	Current response on repeatability of modified electrode for 40 ppm of secondary metabolites in 0.1 M citrate buffer at pH 3 using scan rate of 100 mV/s.	52
4.22	Reproducibility of seven modified electrodes response for 40 ppm of secondary metabolites in 0.1 M citrate buffer at pH 3 using scan rate of 100 mV/s.	53
4.23	DPV analysis for 10 ppm of secondary metabolites from (a) healthy leaves and (b) infected leaves in 0.1 M citrate buffer at pH 3 using scan rate of 100 mV/s.	54
4.24	Calibration curve at different concentration of secondary metabolites for (a) healthy leaves after 14 days plantation (b) infected leaves after 14 days infection (c) healthy leaves after 30 days plantation and (d) infected leaves after 30 days infection in citrate buffer at pH 3 using scan rate of 100 mV/s.	55

LIST OF ABBREVIATIONS

8-OHdG	8-Hydroxy-2'-deoxyguanosine
A	Effective surface area
ANN	Artificial Neural Network
BSR	Basal stem rot
C/O	Carbon-to-oxygen ratio
CE	Counter electrode
CV	Cyclic voltammetry
CVD	Chemical vapour deposition
DIBA	Dot immunobinding assay
DLC	Double layer charging
DNA	Deoxyribonucleic acid
DPV	Differential pulse voltammetry
EDTA	Ethylenediamine-tetraacetic acid
EDX	Energy Dispersive X-Ray
ELISA	Indirect enzyme-linked immunosorbent assay
e-nose	Electronic nose
FE-SEM	Field Emission Scanning Electron Microscopy
FTIR	Fourier Transform Infrared Spectroscopy
GCE	Glassy carbon electrode
GC-MS	Gas chromatography
GDP	Gross Domestic Product
GO	Graphene oxide
GRAS	Generally Recognised as Safe
GSH	Glutathione
HR	Hypersensitive response
I _D /I _G	Intensity ratio of D and G band
IR	Infrared radiation
ITS	Internal transcribed spacer
LC-MS	Liquid chromatography-mass spectroscopy
LOD	Limit of detection
MEA	Malt extract agar
MWCNT	Multiwall carbon nanotube
NMR	Magnetic resonance spectroscopy
OTA	Ochratoxin A
PCR	Polymerase chain reaction
PDA	Potato dextrose agar
rDNA	Ribosomal RNA gene
RE	Reference electrode
rGO	Reduced graphene oxide
ROS	Reactive oxygen species
RWB	Rubber wood block
s	Seconds
sp.	Species
SPCE	Screen printed carbon electrode
SPGE	Silica paste glass electrode
USR	Upper stem rot
VOC	Volatile organic compounds
WE	Working electrode
ZnO-NPs	Zinc oxide nanoparticles

LIST OF APPENDICES

Appendix		Page
1	Chemical composition of healthy roots (Control of 14 days-post infection)	77
2	Chemical composition <i>Ganoderma boninense</i> infected roots at 14 days-post infection	78
3	Chemical composition of healthy roots (Control of 30 days-post infection)	79
4	Chemical composition of <i>Ganoderma boninense</i> infected roots at 30 days-post infection	80

CHAPTER 1

INTRODUCTION

1.1 Background of Study

Malaysia has been recognised as one of the richest countries in terms of oil palm production. Recently, Malaysia invested about 4.49 million hectares of land for oil palm cultivation which produce a rolling output of approximately 25.64 million tonnes of palm products. Consequently, Malaysia's oil palm industry has become one of the main contributors to Malaysia's Gross Domestic Product (GDP) and foreign exchange earnings, and is the source of many job opportunities. Meanwhile, the export of palm oil has tremendously increased from 1.17 million tonnes in 1975 to 16.05 million tonnes in 2016. Malaysia's exports target market is more than 200 markets worldwide including Africa, West Asia, Europe, the Indian sub-continent and Asia (Nambiappan et al., 2018).

However, a major issue that needs to be accounted when dealing with oil palm trees is the fatality of trees as it can cause a big loss to the palm oil industry. *Ganoderma boninense* is one of the diseases that can infect oil palm trees (Susanto et al., 2005). Mature trees can survive slightly longer, but young trees can live approximately one to two years only. The *Ganoderma boninense* infection commonly occurs at the bottom and upper parts of the stem, and are known as basal stem rot (BSR) and upper stem rot (USR) respectively (Durand-Gasselin et al., 2005).

Over a decade ago, electrochemical sensors have been introduced to detect many elements, including heavy metals (Zhu et al., 2009), gases, biological materials as it can give extensive response to the interested analyte. Basically, the analyte will be diluted with a supporting electrolyte to improve conductivity. Thus, the analyte will interact with the working surface of the electrode to give a current response based on the redox reaction activity.

The nanotechnology field has grown up tremendously all around the world. Nanotechnology applications including nanowire, nanoparticles, carbon nanotubes, and quantum dots are widely utilised in food, electronic devices, cosmetics, and environmental industries. In this context, the employment of nanoparticles in electrodes can enhance the current signal as it provides a high surface area to the electrode.

Graphene oxide (GO) is a nanomaterial that has been used in various applications. Decades ago, GO has been explored due to its variety of applications and advantages. GO has been said to have important qualities such as low cost, non-toxic and having a huge potential to be used as a modifier due to its large surface area (Liu et al., 2013). Along with that, zinc oxide nanoparticles have been applied as one of the famous nanomaterials used as a sensing material. With its great properties such as a versatile semiconductor material (Janotti and Van de Walle, 2009) and a good electrocatalyst (Wayu et al., 2013), a sensor of good conductivity can be made.

1.2 Problem Statement and Research Motivation

The detection of *Ganoderma boninense* at the early stage is significant to researchers as the trees may not show any reactions as soon as they are being attacked. The disease is symptomless at early stage and usually manifested when it has already reached a critical stage, thus imposing challenges on effective disease management. Several foliage symptoms can be appeared at the bottom of the stem where it undergoes decaying process due to the development of basidiocarps. The process caused limitation on water and supplements uptake from the root, making the loss of leaf pigments. At the advanced stage, more fronds shrivel and hang down to a skirt structure. In worst cases, those stems may be crack (Parisa et al, 2017).

Until now, there are many techniques and methods applied to diagnose the disease. In the early studies on BSR, tomography (Idris et al., 2010; Abdullah et al., 2013), selective media (Ariffin et al., 1995), immunoassay and PCR methods have been used to detect the *Ganoderma* disease. Unfortunately, these techniques are time consuming, costly and some of the techniques need expertise to interpret the data, thus limiting the scope of research. Meanwhile, the sensor application is a more sensitive, simple, low cost, and less time-consuming method to diagnose the *Ganoderma boninense* disease.

Therefore, an electrochemical sensor is introduced as one of the alternative ways to detect *Ganoderma boninense*. This method is specifically recommended because it is less time consuming and is low cost. Using this way, the secondary metabolites of an oil palm tree as possible biomarkers of *Ganoderma boninense* can be extracted from the tree, thus giving a more specific response to electrochemical analysis. In addition, modification of bare electrode can be made to enhance a higher surface area of electrode.

Thus, the utilisation of reduced graphene oxide and zinc oxide nanoparticles as electrode modifiers is introduced. Graphene oxide demonstrated a good capability towards electrochemical sensor application based on metal-rGO nanomaterials owing to the unique electrochemical and structural properties of rGO as well as the catalytic characteristics of transition metals. As one of the element in transition metals, zinc oxide nanoparticles can improve

electrochemical performance and elevate electron transfer between the working electrode and secondary metabolites. However, this modified electrode has not been established yet as the capability of zinc oxide nanoparticles to capture secondary metabolites is still doubted. Hence, in this study zinc oxide nanoparticle was tested based on its electrochemical performance towards the behaviour of secondary metabolites in oil palm trees.

1.3 Objectives of Study

1.3.1 General Objective

The main objective of this research is to develop a sensor specifically to detect an oil palm's disease contributed from *Ganoderma boninense* using ZnO-NPs/rGO/SPCE modified electrode.

1.3.2 Specific Objective

- I. To prepare and characterise reduced graphene oxide (rGO) and zinc oxide nanoparticles (ZnO-NPs) using field emission scanning electron microscopy (FESEM), energy dispersive x-ray (EDX), Fourier Transform Infrared Spectroscopy (FTIR), Raman spectroscopy and cyclic voltammetry (CV).
- II. To modify an electrode using drop cast of graphene oxide (GO), reduced graphene oxide (rGO) by CV and zinc oxide nanoparticles (ZnO-NPs) through chronoamperometry technique.
- III. To develop a ZnO-NPs/rGO/SPCE electrochemical sensor for the detection of secondary metabolites as possible biomarkers of *Ganoderma boninense*.

1.4 Scope and Limitation

In this research, crude leaf extract is extracted from oil palm trees at durations of 14 days and 30 days after infection, and 14 days and 30 days after plantation. On account of that, the crude extracts were only able to maintain their performance for about three months in cold conditions. It is because beyond these conditions, the crude extracts' performance may be affected by moist and vapour from the storage condition. In addition, oil palm trees secrete thousand groups of secondary metabolites to defend themselves from diseases. Thus, it is quite difficult to specifically detect the secondary metabolites secreted from oil palm in defence towards *Ganoderma boninense*.

REFERENCES

- Abdolhosseinzadeh, S., Asgharzadeh, H., & Kim, H. S. (2015). Fast and fully-scalable synthesis of reduced graphene oxide, *Scientific Reports*, 5: 10160.
- Abdulgafour, Hassan, H., Z., Al-Hardan, N., & Yam, F. (2010). Growth of zinc oxide nanoflowers by thermal evaporation method, *Physica B: Condensed Matter*, 405(11): 2570–2572.
- Abdullah, A., Adom, A., Shakaff, A., Ahmad, M., Zakaria, A., Saad, F. S. A., Isa, C. M. N. C., Masnan, M. J., & Kamarudin, L. (2012). Intelligent Systems, Modelling and Simulation (ISMS), 2012 Third International Conference IEEE, Feb 8–10: *Hand-held electronic nose sensor selection system for basal stamp rot (BSR) disease detection*. Kota Kinabalu, Malaysia: IEEE.
- Abdullah, A., Shakaff, A. M., Zakaria, A., Saad, F., Shukor, S. A., & Mat, A. (2014). *Application Specific Electronic Nose (ASEN) for Ganoderma boninense detection using artificial neural network*. Electronic Design (ICED), 2014 2nd International Conference on IEEE.
- Abdullah, J., Hassan, H., Shari, M. R., Mohd, S., Mustapha, M., Mahmood, A. A., Jamaludin, S., Ngah, M. R., & Hamid, N. H. (2013). GammaScorpion: Mobile gamma-ray tomography system for early detection of basal stem rot in oil palm plantations. *Optical Engineering*, 52(3): 036502–036502.
- Adhikari, B. & Majumdar, S. (2004). Polymers in sensor applications. *Progress in Polymer Science*, 29(7): 699–766.
- Adom, A. and Shakaff, A. (2006). M. N., Md Ahmad, A. H., Abdullah, M. Z., Daud, M. A, Markom. (Eds.) Proceedings of the IEEE Systems, Man and Cybernetics Society, 5th Conference on AICS: *Electronic olfactory prototype of an automated plant disease detection system for the agricultural industry*.
- Ahmed John, S. & Annadurai, A. (2015). GC-MS screening of active secondary metabolites. Present in the Cleome gynandra. *International Journal of Phytopharmacy*, 5(4): 47 - 52.
- Akanbi, F. S., Yusof, N. A., Abdullah, J., Sulaiman, Y., & Hushiarian, R. (2017). Detection of quinoline in *G. boninense*-infected plants using functionalized multi-walled carbon nanotubes: A field study. *Sensors*, 17(7): 1538.
- Akhavan, O. & Ghaderi, E. (2012). *Escherichia coli* bacteria reduce graphene oxide to bactericidal graphene in a self-limiting manner. *Carbon*, 50(5): 1853–1860.

- Al-Hardan, N., Abdullah, M., Ahmed, N., Yam, F., & Aziz, A. A. (2012). UV Photodetector behavior of 2D ZnO plates prepared by electrochemical deposition. *Superlattices and Microstructures*, 51(6): 765–771.
- Al-Shabib, N. A., Husain, F. M., Ahmed, F., Khan, R. A., Ahmad, I., Alsharaeh, E., Shahnawaz, M. K., Hussain, A. Rehman, M. T., Hassan, I., Khan, J. M., Ashraf, G. M., Alsahme, A., Al-Ami, M. F., Tarasov, V. V., & Aliev, G. (2016). Biogenic synthesis of zinc oxide nanostructures from nigella sativa seed: prospective role as food packaging material inhibiting broad-spectrum quorum sensing and biofilm. *Scientific Reports*, 6: 36761.
- Alexander, A., Sipaut, C. S., Chong, K. P., Lee, P. C., & Dayou, J. (2014). Sensitivity analysis of the detection of *Ganoderma boninense* infection in oil palm using FTIR. *Transactions on Science and Technology*, 1(1): 1–5.
- Aljabali, A. A., Barclay, J. E., Butt, J. N., Lomonosoff, G. P., & Evans, D. J. (2010). Redox-active ferrocene-modified Cowpea mosaic virus nanoparticles. *Dalton Transactions*, 39(32): 7569–7574.
- Allen, M. J., Tung, V. C., & Kaner, R. B. (2009). Honeycomb carbon: A review of graphene. *Chemical Reviews*, 110(1): 132–145.
- Amarnath, C. A., Hong, C. E., Kim, N. H., Ku, B.-C., Kuila, T., & Lee, J. H. (2011). Efficient synthesis of graphene sheets using pyrrole as a reducing agent. *Carbon*, 49(11): 3497–3502.
- Amin, M. T., Alazba, A. A., & Manzoor, U. (2014). A review of removal of pollutants from water/wastewater using different types of nanomaterials. *Advances in Materials Science and Engineering*, 2014: 825910.
- Ansari, A. A., Kaushik, A., Solanki, P. R., & Malhotra, B. (2010). Nanostructured zinc oxide platform for mycotoxin detection. *Bioelectrochemistry*, 77(2): 75–81.
- Arbuzov, A., Tarasov, B., & Muradyan, V. (2012). Synthesis of few-layer graphene sheets via chemical and thermal reduction of graphite oxide. *Proceedings of the International Conference Nanomaterials: Applications and Properties*, 1(1). Ukraine: Sumy State University Publishing.
- Ariffin, D. & Idris, S. (1993). PORIM International Palm Oil Conference: Progress, prospects challenges towards the 21st century (Agriculture), September 9–14, *A selective medium for the isolation of Ganoderma from disease tissues*. Kuala Lumpur, Malaysia: PORIM.
- Ariffin, D., Idris, S., & Khairudin, H. (1995). PORIM International Palm Oil Congress: *Conformation of Ganoderma infected palm by drilling technique*. Kuala Lumpur: PORIM.

- Aristizabal, D., Giraldo, D., Sanchez, S., Taborda, G., & Baeza, A. (2017). Implementation of electrochemical elements for an alternative detection of ochratoxin a. *Journal of Physics: Conference Series*, 786(1): 012040, IOP Publishing.
- Ashrafi, A. & Jagadis, C. H. (2007). Review of zincblende ZnO: Stability of metastable ZnO phases. *Journal of Applied Physics*, 102(7): 4.
- Azizul, I., Nor, A.Y., Rosiah, O., Siti, N. A. A., Wong, M. Y. *Application of GC-MS based metabolomics for metabolite variations in healthy and ganoderma boninense infected oil palm leaf*. Paper presented at the meeting of the Nanomite Annual Symposium, Serdang. November 2017.
- Bai, L., Yuan, R., Chai, Y., Yuan, Y., Wang, Y., & Xie, S. (2012). Direct electrochemistry and electrocatalysis of a glucose oxidase-functionalized bioconjugate as a trace label for ultrasensitive detection of thrombin. *Chemical Communications*, 48(89): 10972–10974.
- Balandrin, M. F., Klocke, J. A., Wurtele, E. S., & Bollinger, W. H. (1985). Natural plant chemicals: Sources of industrial and medicinal materials. *Science*, 228(4704): 1154–1160.
- Bard, A. J. (1983). Chemical modification of electrodes. *Journal of Chemical Education* 60(4): 302–304, ACS Publications.
- Baruah, S. & Dutta, J. (2009). Hydrothermal growth of ZnO nanostructures. *Science and Technology of Advanced Materials*, 10(1): 013001.
- Bassett, K. & Peters, R. N. (2010). *Ganoderma: A significant root pathogen*. Texas: Arborillogical Services Inc. Publication. Retrieved from <http://www.arborillogical.com/tree-knowledgearticlespublications/ganoderma/>
- Beek, W. J., Wienk, M. M., & Janssen, R. A. (2004). Efficient hybrid solar cells from zinc oxide nanoparticles and a conjugated polymer. *Advanced Materials*, 16(12): 1009–1013.
- Bhasha, S., Malik, P., Santosh, S., & Purnima, J. (2015). Synthesis and characterization of nanocrystalline zinc oxide thin films for ethanol vapor sensor. *Journal of Nanomedicine & Nanotechnology*, 6(4): 306 - 309.
- Bhatia, S. (2015). Application of plant biotechnology. In *Modern applications of plant biotechnology in pharmaceutical sciences* (pp. 157–207). London Wall, UK: Elsevier.
- Bhuyan, M. S. A., Uddin, M. N., Islam, M. M., Bipasha, F. A. & Hossain, S. S. (2016). Synthesis of graphene. *International Nano Letters*, 6(2): 65–83.

- Bolarinwa, I. F., Orfila, C. & Morgan, M. R. A. (2014). Amygdalin content of seeds, kernels and food products commercially-available in the UK. *Food Chemistry*, 152: 133-139.
- Bolotin, K. I., Sikes, K., Jiang, Z., Klima, M., Fudenberg, G., Hone, J., Kim, P., & Stormer, H. (2008). Ultrahigh electron mobility in suspended graphene. *Solid State Communications*, 146(9-10): 351–355.
- Bouarab, K., Melton, R., Peart, J., Baulcombe, D. & Osbourn, A. (2002). A saponin-detoxifying enzyme mediates suppression of plant defences. *Nature*, 418 : 889 -892.
- Chan, C. Y. (2016). *Graphene Based Electrical Biosensors for the Detection of Biomolecules*, The Hong Kong Polytechnic University, China.
- Cheng, H., Cheng, J. Zhang, Y., & Wang, Q. M. (2007). Large scale fabrication of ZnO micro and nanostructures by microwave thermal evaporation deposition. *Journal of Crystal Growth*, 299(1): 34–40.
- Cheng, S. Y., Xu, F., & Wang, Y. (2009). Advances in the study of flavonoids in Ginkgo biloba leaves. *Journal of Medicinal Plants Research*, 3(13): 1248–1252.
- Chong, K. P., Atong, M., & Rossall, S. (2012). The role of syringic acid in the interaction between oil palm and *Ganoderma boninense*, the causal agent of basal stem rot. *Plant Pathology*, 61(5): 953–963.
- Cooper, A. J., Wilson, N. R., Kinloch, I. A., & Dryfe, R. A. (2014). Single stage electrochemical exfoliation method for the production of few-layer graphene via intercalation of tetraalkylammonium cations. *Carbon*, 66: 340–350.
- Couto, R., Lima, J., & Quinaz, M. (2016). Recent developments, characteristics and potential applications of screen-printed electrodes in pharmaceutical and biological analysis. *Talanta*, 146: 801–814.
- Creelman, R. A. & Mullet, J. E. (1997). Biosynthesis and action of jasmonates in plants. *Annual Review of Plant Biology*, 48(1): 355–381.
- Cruickshank, A. C., Tay, S. E. R., Illy, B. N., Campo, R. D., Schumann, S., Jones, T. S., Heutz, S., Mclachlan, M. A., McComb, D. W., Riley, D. J., & Ryan, M. P. (2011). *Electrodeposition of ZnO nanostructures on molecular thin films*, *Chemistry of Materials*, 23(17): 3863–3870.
- Dai, S., Li, Y., Du, Z., & Carter, K. R. (2013). Electrochemical deposition of ZnO hierarchical nanostructures from hydrogel coated electrodes. *Journal of The Electrochemical Society*, 160(4): D156–D162.
- Daško, L., Belajová, E., Rauová, D., & Kováč, M. (2005). Determination of ochratoxin A in beer. *Czech Journal of Food Sciences*, 23(2): 69–73.

- Devi, R., Thakur, M., & Pundir, C. (2011). Construction and application of an amperometric xanthine biosensor based on zinc oxide nanoparticles polypyrrole composite film. *Biosensors and Bioelectronics*, 26(8): 3420–3426.
- Diabaté, S., Franqueville, H., Adon, B., Coulibaly, O., & Ake, S. (2009). The role of phenolic compounds in the determination of wilt disease tolerance of oil palm (*Elaeis guineensis* JACQ). *African Journal of Biotechnology*, 8(21): 5679- 5690.
- Ding, J., Zhu, S., Zhu, T., Sun, W., Li, Q., Wei, G., & Su, Z. (2015). Hydrothermal synthesis of zinc oxide-reduced graphene oxide nanocomposites for an electrochemical hydrazine sensor. *RSC Advances*, 5(29): 22935–22942.
- Dixon, R. A. (2001). Natural products and plant disease resistance. *Nature* 411(6839): 843–847.
- Dong, S., & Wang, Y. (1988). Anodic stripping voltammetric determination of trace lead with a nafion/crown-ether film electrode. *Talanta*, 35(10); 819-821.
- Du, D., Liu, J. Zhang, X., Cui, X., & Lin, Y. (2011). One-step electrochemical deposition of a graphene-ZrO₂ nanocomposite: Preparation, characterization and application for detection of organophosphorus agents. *Journal of Materials Chemistry*, 21(22): 8032–8037.
- Durand-Gasselín, T., Asmady, H., Flori, A., Jacquemard, J.-C., Hayun, Z., Breton, F., & De Franqueville, H. (2005). Possible sources of genetic resistance in oil palm (*Elaeis guineensis* Jacq.) to basal stem rot caused by *Ganoderma boninense*—Prospects for future breeding. *Mycopathologia*, 159(1): 93–100.
- Dutse, S. W., Yusof, N. A., Ahmad, H., Hussein, M. Z., & Zainal, Z. (2012). An electrochemical DNA biosensor for *Ganoderma boninense* pathogen of the Oil palm utilizing a new ruthenium complex, [Ru(dppz)₂(qtpy)]Cl₂. *International Journal of Electrochemical Science*, 7: 8105–8115.
- Dutta, R., Morgan, D., Baker, N., Gardner, J. W., & Hines, E. L. (2005). Identification of *Staphylococcus aureus* infections in hospital environment: Electronic nose based approach. *Sensors and Actuators B: Chemical*, 109(2): 355–362.
- Eda, G., Mattevi, C., Yamaguchi, H., Kim, H., & Chhowalla, M. (2009). Insulator to semimetal transition in graphene oxide. *The Journal of Physical Chemistry C*, 113(35): 15768–15771.
- Edmundson, M., Thanh, N. T., & Song, B. (2013). Nanoparticles based stem cell tracking in regenerative medicine. *Theranostics*, 3(8): 573- 582.

- Edwards, R., Blount, J. W., & Dixon, R. A. (1991). Glutathione and elicitation of the phytoalexin response in legume cell cultures. *Planta*, 184(3): 403–409.
- Enculescu, I., Sima, M., Enculescu, M., Enache, M., Vasile, V., & Neumann, R. (2007). Influence of geometrical properties on light emission of ZnO nanowires. *Optical Materials*, 30(1): 72–75.
- Falascioni, M., Gobbi, E., Pardo, M., Della Torre, M., Bresciani, A., & Sberveglieri, G. (2005). Detection of toxigenic strains of *Fusarium verticillioides* in corn by electronic olfactory system. *Sensors and Actuators B: Chemical*, 108(1): 250–257.
- Filipponi, L. & Sutherland, D. (2012). *Nanotechnologies: Principles, applications, implications and hands-on activities*. Luxembourg: Publications Office of the European Union.
- Flood, J., Bridge, P., & Holderness, M. (2000). *Ganoderma* diseases of perennial crops. UK: CABI Bioscience.
- Gao, C., Yu, X.-Y., Xu, R.-X., Liu, J.-H., & Huang, X.-J. (2012). AlOOH-reduced graphene oxide nanocomposites: One-pot hydrothermal synthesis and their enhanced electrochemical activity for heavy metal ions. *ACS Applied Materials & Interfaces*, 4(9): 4672–4682.
- Gardner, J. W., & Bartlett, P. N. (1994). A brief history of electronic noses. *Sensors and Actuators B: Chemical*, 18(1–3), 210–211.
- Gómez-Navarro, C., Weitz, R. T., Bittner, A. M., Scolari, M., Mews, A., Burghard, M., & Kern, K. (2007). Electronic transport properties of individual chemically reduced graphene oxide sheets. *Nano Letters*, 7(11): 3499–3503.
- Goranova, D., Avdeev, G., & Rashkov, R. (2014). Electrodeposition and characterization of Ni–Cu alloys. *Surface and Coatings Technology*, 240: 204–210.
- Grayer, R. J. & Harborne, J. B. (1994). A survey of antifungal compounds from higher plants, 1982–1993. *Phytochemistry*, 37(1): 19–42.
- Grubb, C. D. & Abel, S. (2006). Glucosinolate metabolism and its control. *Trends in Plant Science*, 11(2): 89–100.
- Guan, Y., Ramalingam, S., Nagegowda, D., Taylor, P. W., & Chye, M.-L. (2008). *Brassica juncea* chitinase BjCHI1 inhibits growth of fungal phytopathogens and agglutinates Gram-negative bacteria. *Journal of Experimental Botany*, 59(12): 3475–3484.
- Guex, L. G., Sacchi, B., Peuvot, K. F., Andersson, R. L., Pourrahimi, A. M., Ström, V., Farris, S., & Olsson, R. T. (2017). Experimental review:

Chemical reduction of graphene oxide (GO) to reduced graphene oxide (rGO) by aqueous chemistry. *Nanoscale*, 9(27): 9562–9571.

- Guo H. L., Wang, X. F., Qian, Q. Y., Wang, F. B., & Xia, X. H. (2009). A Green Approach to the Synthesis of Graphene Nanosheets. *ACS Nano*, 3: 2653–2659.
- Guo, Y., Sun, X., Liu, Y., Wang, W., Qiu, H., & Gao, J. (2012). One pot preparation of reduced graphene oxide (RGO) or Au (Ag) nanoparticle-RGO hybrids using chitosan as a reducing and stabilizing agent and their use in methanol electrooxidation. *Carbon*, 50(7): 2513–2523.
- Gupta, P. & Rather, G. (1989). Electrochemical synthesis of quinolinic acid from quinoline. *Asian Journal of Chemistry*, 1(1): 52–56.
- Gurunathan, S., Han, J. W., Kim, E. S., Park, J. H. & Kim, J.-H. (2015). Reduction of graphene oxide by resveratrol: A novel and simple biological method for the synthesis of an effective anticancer nanotherapeutic molecule. *International Journal of Nanomedicine*, 10: 2951 - 2969.
- Hallam, P. M. & Banks, C. E. (2011). Quantifying the electron transfer sites of graphene. *Electrochemistry Communications*, 13(1): 8–11.
- Hamidon, N. A. & Mukhlisin, M. (2014). A review of application of computed tomography on early detection of basal stem rot disease. *Jurnal Teknologi*, 70(3), 45–47.
- Hammond-Kosack, K. E. & Jones, J. (1996). Resistance gene-dependent plant defense responses. *The Plant Cell*, 8(10): 1773 - 1791.
- Hao, J., Wu, K., Wan, C., & Tang, Y. (2018). Reduced graphene oxide-ZnO nanocomposite based electrochemical sensor for sensitive and selective monitoring of 8-hydroxy-2'-deoxyguanosine. *Talanta*, 185: 550–556.
- Hasanuzzaman, M, Nahar, K., T. I., & Fujita, M. (2017). Glutathione in plants: biosynthesis and physiological role in environmental stress tolerance. *Physiology and Molecular Biology of Plant*, 23(2): 249-268.
- Ho, Y. W. & Nawawi, A. (1985). *Ganoderma boninense* Pat. from basal stem rot of oil palm (*Elaeis guineensis*) in Peninsular Malaysia. *Pertanika*, 8(3): 425–428.
- Hsiao, M. C., Liao, S. H., Yen, M. Y., Teng, C. C., Lee, S. H., Pu, N. W., Wang, C. A., Sung, Y., Ger, M. D., Ma, C. C. M. & Hsiao, M. H. (2010). Preparation and properties of a graphene reinforced nanocomposite conducting plate. *Journal of Materials Chemistry*, 20(39): 8496–8505.

- Huang, X., Liu, F., Jiang, P., & Tanaka, T. (2013). IEEE International Conference Solid Dielectrics (ICSD): *Is graphene oxide an insulating material?*, Bologna. June 2013.
- Hummers Jr, W. S. & Offeman, R. E. (1958). Preparation of graphitic oxide. *Journal of the American Chemical Society*, 80(6): 1339.
- Hushiarian, R., Yusof, N. A., & Dutse, S. W. (2013). Detection and control of *Ganoderma boninense*: Strategies and perspectives. *Springer Plus*, 2(1): 1- 12.
- Ibrahim, M. H., Jaafar, H. Z., Karimi, E., & Ghasemzadeh, A. (2012). Primary, secondary metabolites, photosynthetic capacity and antioxidant activity of the Malaysian Herb Kacip Fatimah (*Labisia Pumila Benth*) exposed to potassium fertilization under greenhouse conditions. *International Journal of Molecular Sciences*, 13(11): 15321–15342.
- Idris, A., Kushairi, D., Ariffin, D., & Basri, M. (2006). Technique for inoculation of oil palm germinated seeds with *Ganoderma*. *MPOB Infotmation Series*, 314: 1–4.
- Idris, A., Mazliham, M., Loonis, P., & Wahid, M. (2010). *GanoSken for early detection of Ganoderma infection in oil palm*. MPOB Information Series 442. Selangor: Palm Information Centre MPOB.
- Janata, J. (2008). Introduction: Modern topics in chemical sensing, *Chemical Reviews*, 108(2): 327–328.
- Janotti, A. & Van de Walle, C. G. (2009). Fundamentals of zinc oxide as a semiconductor. *Reports on Progress in Physics*, 72(12): 126501.
- Jian, J. M., Liu, Y. Y., Zhang, Y. L., Guo, X. S., & Cai, Q. (2013). Fast and sensitive detection of Pb²⁺ in foods using disposable screen-printed electrode modified by reduced graphene oxide. *Sensors*, 13(10): 13063–13075.
- Justino, C. I., Gomes, A. R., Freitas, A. C., Duarte, A. C., & Rocha-Santos, T. A. (2017). Graphene based sensors and biosensors. *TrAC Trends in Analytical Chemistry*, 91: 53–66.
- Kang, S. Y. & Kim, Y. C. (2007). Decursinol and decursin protect primary cultured rat cortical cells from glutamate-induced neurotoxicity. *Journal of Pharmacy and Pharmacology*, 59(6): 863–870.
- Karthikeyan, M., Radhika, K., Bhaskaran, R., Mathiyazhagan, S., Samiyappan, R., & Velazhahan, R. (2006). Rapid detection of *Ganoderma* disease of coconut and assessment of inhibition effect of various control measures by immunoassay and PCR. *Plant Protection Science*, 42(2): 49–57.

- Kausar, A., Rafique, I., Anwar, Z., & Muhammad, B. (2016). Perspectives of epoxy/graphene oxide composite: Significant features and technical applications. *Polymer-Plastics Technology and Engineering*, 55(7): 704–722.
- Khelladi, M., Mentar, L., Beniaiche, A., Makhloufi, L., & Azizi, A. (2013). A study on electrodeposited zinc oxide nanostructures. *Journal of Materials Science: Materials in Electronics*, 24(1): 153–159.
- Kim, K. S., Zhao, Y., Jang, H., Lee, S. Y., Kim, J. M., Kim, K. S., Ahn, J. H., Choi, J. Y., & Hong, B. H. (2009). Large scale pattern growth of graphene films for stretchable transparent electrodes. *Nature*, 457(7230): 706 - 710.
- Krishnan, P., Kruger, N., & Ratcliffe, R. (2004). Metabolite fingerprinting and profiling in plants using NMR. *Journal of Experimental Botany*, 56(410): 255–265.
- Kumar, R., Al-Dossary, O., Kumar, G., & Umar, A. (2015). Zinc oxide nanostructures for NO₂ gas–sensor applications: A review. *Nano-Micro Letters*, 7(2): 97–120.
- Kumar, V., Ntwaeaborwa, O., Coetsee, E., & Swart, H. (2016). Role of deposition time on the properties of ZnO: Tb³⁺ thin films prepared by pulsed laser deposition. *Journal of Colloid and Interface Science*, 474: 129–136.
- Lee, C., Wei, X., Kysar, J. W., & Hone, J. (2008). Measurement of the elastic properties and intrinsic strength of monolayer graphene. *Science*, 321(5887): 385–388.
- Liang, Z., Zhang, Q., Jiang, L., Cao, G. (2015). ZnO cathode buffer layers for inverted polymer solar cells, *Energy and Environmental Science*, 8: 3442–3476.
- Liao, F., Han, X., Zhang, Y., Xu, C., & Chen, H. (2017). Hydrothermal synthesis of flower like zinc oxide microstructures with large specific surface area. *Journal of Materials Science: Materials in Electronics*, 28(22): 16855–16860.
- Liu, J., Cui, L., & Losic, D. (2013). Graphene and graphene oxide as new nanocarriers for drug delivery applications. *Acta Biomaterialia*, 9(12): 9243–9257.
- Liu, W. (2017). Preparation of a zinc oxide-reduced graphene oxide nanocomposite for the determination of cadmium (II), lead (II), copper (II), and mercury (II) in water. *International Journal of Electrochemical Science*, 12(6): 5392–5403.

- Liu, X., Qi, X., Zhang, Z., Ren, L., Hao, G., Liu, Y., Wang, Y., Huang, K., Wei, X., Li, J., Huang, Z., & Zhong, J. (2014). Electrochemically reduced graphene oxide with porous structure as a binder-free electrode for high rate supercapacitors. *RSC Advances*, 4(26): 13673–13679.
- Loryuenyong, V., Totepvimarn, K., Eimburanaprat, P., Boonchompoo, W., & Buasri, A. (2013). Preparation and characterization of reduced graphene oxide sheets via water based exfoliation and reduction methods. *Advances in Materials Science and Engineering*, 2013: 1 - 5.
- Lotya, M., Hernandez, Y., King, P. J., Smith, R. J., Nicolosi, V., Karlsson, L. S. Blighe, F. M., De, S., Wang, Z., McGovern, I. Duesberg, G. S., & Coleman, J. N. (2009). Liquid phase production of graphene by exfoliation of graphite in surfactant/water solutions. *Journal of the American Chemical Society*, 131(10): 3611–3620.
- Marcano, D. C., Kosynkin, D. V., Berlin, J. M., Sinitskii, A., Sun, Z., Slesarev, A., Alemany, L. B., Lu, W., & Tour, J. M. (2010). Improved synthesis of graphene oxide. *ACS Nano*, 4(8): 4806–4814.
- Markom, M. A., Shakaff, A. M., Adom, A., Ahmad, M., Hidayat, W., Abdullah, A., & Fikri, N. A. (2009). Intelligent electronic nose system for basal stem rot disease detection. *Computers and Electronics in Agriculture*, 66(2): 140–146.
- Maruthupandy, M., Anand, M., Maduraiveeran, G., Suresh, S., Beevi, A. S. H., & Priya, R. J. (2016). Investigation on the electrical conductivity of ZnO nanoparticles-decorated bacterial nanowires. *Advances in Natural Sciences: Nanoscience and Nanotechnology*, 7(4): 045011 - 045020.
- Mazid, M., Khan, T., & Mohammad, F. (2011). Role of secondary metabolites in defense mechanisms of plants. [Special Issue] *Biology and Medicine*, 3(2): 232–249.
- Mazliham, M. S. U., Loonis, P., & Idris, A. S. (2008). Interpretation of sound tomography image for the recognition of *Ganoderma* infection level in oil palm. *Lecture Notes in Electrical Engineering*, 6: 409- 426.
- Minitha, C. R., & Rajendrakumar, R. T. (2013). Synthesis and characterization of reduced graphene oxide. *Advanced Materials Research*, 678: 56-60.
- Miyamoto, K., Shimizu, T., & Okada, K. (2014). Transcriptional regulation of the biosynthesis of phytoalexin: A lesson from specialized metabolites in rice. *Plant Biotechnology*, 31: 377–388.
- Moncalvo, J. (2000). Systematics of *Ganoderma*. *Ganoderma Diseases of Perennial Crops*, 23–45.
- Munné-Bosch, S. & Alegre, L. (2003). Drought induced changes in the redox state of α -tocopherol, ascorbate, and the diterpene carnosic acid in chloroplasts of Labiatae species differing in carnosic acid contents. *Plant Physiology*, 131(4): 1816-1825.

- Muthulakshmi, A., Margret, R. J., & Mohan, V. (2012). GC-MS analysis of bioactive components of *Feronia elephantum* Correa (Rutaceae). *Journal of Applied Pharmaceutical Science*, 2(2): 69–74
- Naher, L., Yusuf, U. K., Ismail, A., Tan, S. G. & Mondal, M. (2013). Ecological status of '*Ganoderma*' and basal stem rot disease of oil palms ('*Elaeis guineensis*' Jacq.). *Australian Journal of Crop Science*, 7(11): 1723 - 1727.
- Nambiappan, B., Ismail, A., Hashim, N., Ismail, N., Nazrma, D., Hassan, N. A. M., & Kushairi, A. (2018). Malaysia: 100 years of resilient palm oil economic performance. *Journal of Oil Palm Research*, 30(1): 13–25.
- Nath, P., Pandey, R., Das, A., & Mallik, A. (2017). Effect of deposition potential and copper concentration on the phase transformation mechanism and structural distribution during electrodeposition of Ni-Cu magnetic alloy thin films. *Kovove materialy-metallic materials*, 55(4): 255-265.
- Neghmouche, NS. & Lanez, T. (2013). Calculation of diffusion coefficients and layer thickness for oxidation the ferrocene using voltammetry technique. *International Journal of Chemical Sciences*, 1(1): 28-32.
- Nithya, M. (2015). Electrochemical sensing of ascorbic acid on ZnO decorated reduced graphene oxide electrode. *Journal of Biosensors & Bioelectronics*, 6(1): 1.
- Nocito, F. F., Pirovano, L., Cocucci, M., & Sacchi, G. A. (2002). Cadmium induced sulfate uptake in maize roots. *Plant Physiology*, 129(4): 1872–1879.
- Novoselov, K. S., Geim, A. K., Morozov, S. V., Jiang, D. Zhang, Y., Dubonos, S. V., Grignorieva, I. V., & Firsov, A. A. (2004). Electric field effect in atomically thin carbon films. *Science*, 306(5696): 666–669.
- Nusaibah, S., Akmar, A. S. N., Idris, A., Sariah, M., & Pauzi, Z. M. (2016). Involvement of metabolites in early defense mechanism of oil palm (*Elaeis guineensis* Jacq.) against *Ganoderma* disease. *Plant Physiology and Biochemistry*, 109: 156–165.
- Nusaibah, S., Akmar, S. N. A., Pauzi, M. Z., Idris, A., & Sariah, M. (2011). Detection of phytosterols in *Ganoderma boninense*-infected oil palm seedlings through GC-MS analysis. *Journal of Oil Palm Research*, 23: 1069–1077.
- Oh, W. C. & Zhang, F. J. (2011). Preparation and characterization of graphene oxide reduced from a mild chemical method. *Asian Journal of Chemistry*, 23(2): 875–879.

- Pal, S. B. S. K. & Dutta, J. (2002). Nanostructured zinc oxide for water treatment. *Nanoscience and Nanotechnology-Asia*, 2(2): 90–102.
- Palanisamy, S., Chen, S. M., & Sarawathi, R. (2012). A novel nonenzymatic hydrogen peroxide sensor based on reduced graphene oxide/ZnO composite modified electrode. *Sensors and Actuators B: Chemical*, 166: 372–377.
- Pardo, A., Marco, S., Calaza, C., Ortega, A., Perera, A., Sundic, T., & Samitier, J. (2001). Methods for sensors selection in pattern recognition. *Proc. Inst. of Physics Publishing*: 83–88.
- Parisa, A., Farrah, M. M., Khairulmazmi, A., Shattri, M., Idris, A. S. (2017). Early Detection of *Ganoderma* Basal Stem Rot of Oil Palms Using Artificial Neural Network Spectral Analysis. *Plant Disease*, 101(6): 1009 – 1016.
- Park, S. & Ruoff, R. S. (2009). Chemical methods for the production of graphenes. *Nature Nanotechnology*, 4(4): 217–224.
- Park, Y. K., Choi, H. S., Kim, J. H., Kim, J. H., & Hahn, Y.-B. (2011). High performance field effect transistors fabricated with laterally grown ZnO nanorods in solution. *Nanotechnology*, 22(18): 185310.
- Paterson, R. (2007). *Ganoderma* disease of oil palm - A white rot perspective necessary for integrated control. *Crop Protection*, 26(9): 1369–1376.
- Pauli, G. F., Jaki, B. U., & Lankin, D. C. (2005). Quantitative ¹H NMR: Development and potential of a method for natural products analysis. *Journal of Natural Products*, 68(1): 133–149.
- Pearce, G., Strydom, D., Johnson, S., & Ryan, C. A. (1991). A polypeptide from tomato leaves induces wound-inducible proteinase inhibitor proteins. *Science*, 253(5022): 895–897.
- Peters, R. J. (2006). Uncovering the complex metabolic network underlying diterpenoid phytoalexin biosynthesis in rice and other cereal crop plants. *Phytochemistry*, 67(21): 2307–2317.
- Pimenta, M. A., Dresselhaus, G., Dresselhaus, M. S., & Cancado, L. G. (2007). Studying disorder in graphite based systems by Raman Spectroscopy. *Physical Chemistry Chemical Physics*, 9: 1276-1291.
- Protasova, L., Rebrov, E., Choy, K., Pung, S., Engels, V., Cabaj, M., Wheatly, A. E. H., & Schouten, J. (2011). ZnO based nanowires grown by chemical vapour deposition for selective hydrogenation of acetylene alcohols. *Catalysis Science & Technology*, 1(5): 768–777.
- Radić, S., Vujčić, V., Glogoški, M., & Radić-Stojković, M. (2016). Influence of pH and plant growth regulators on secondary metabolite production

and antioxidant activity of *Stevia rebaudiana* (Bert). *Periodicum Biologorum*, 118(1): 9 -19.

- Rajendran, L., Kandan, A., Karthikeyan, G., Raguchander, T., & Samiyappan, R. (2009). Early detection of *Ganoderma* causing basal stem rot disease in coconut plantations. *Journal of Oil Palm Research*, 21(6): 627–635.
- Ray, S. C. & Jana, N. R. (2017). *Carbon Nanomaterials for Biological and Medical Applications*, Elsevier.
- Rees, R., Flood, J., Hasan, Y., Potter, U., & Cooper, R. M. (2009). Basal stem rot of oil palm (*Elaeis guineensis*); Mode of root infection and lower stem invasion by *Ganoderma boninense*. *Plant Pathology*, 58(5): 982–989.
- Rella, S., Giuri, A., Corcione, C. E., Acocella, M. R., Colella, S., Guerra, G., Listrorti, A., Rizzo, A., & Malitesta, C. (2015). X-ray photoelectron spectroscopy of reduced graphene oxide prepared by a novel green method. *Vacuum*, 119: 159–162.
- Rodrigues, S., Shukla, A. K., Munichandraiah, N. (1998). A cyclic voltammetric study of the kinetics and mechanism of electrodeposition of manganese dioxide. *Journal of applied electrochemistry*, 28(11): 1235-1241.
- Rico, C. M., Majumdar, S., Duarte-Gardea, M., Peralta-Videa, J. R., & Gardea-Torresdey, J. L. (2011). Interaction of nanoparticles with edible plants and their possible implications in the food chain. *Journal of Agricultural and Food Chemistry*, 59(8): 3485–3498.
- Ruiz, J. J., Aldaz, A., & Dominguez, M. (1977). Mechanism of L-ascorbic acid oxidation and dehydro-L-ascorbic acid reduction on a mercury electrode. I. Acid medium. *Canadian Journal of Chemistry*, 55(15): 2799–2806.
- Sahebi, M., Hanafi, M. M., van Wijnen, A. J., Akmar, A. S. N. Azizi, P., Idris, A. S., Taheri, A., & Foroughi, M. (2017). Profiling secondary metabolites of plant defence mechanisms and oil palm in response to *Ganoderma boninense* attack. *International Biodeterioration & Biodegradation*, 122: 151–164.
- Saito, K. (2004). Sulfur assimilatory metabolism. The long and smelling road. *Plant Physiology*, 136(1): 2443–2450.
- Salih, E., Mekawy, M., Hassan, R. Y., & El-Sherbiny, I. M. (2016). Synthesis, characterization and electrochemical-sensor applications of zinc oxide/graphene oxide nanocomposite. *Journal of Nanostructure in Chemistry*, 6(2): 137-144.

- Sanderson, F. (2005). An insight into spore dispersal of *Ganoderma boninense* on oil palm. *Mycopathologia*, 159(1): 139–141.
- Sankaran, K., Bridge, P., & Gokulapalan, C. (2005). *Ganoderma* diseases of perennial crops in India—An overview. *Mycopathologia*, 159(1): 143–152.
- Sato F. & Matsui, K. (2012). Engineering the biosynthesis of low molecular weight metabolites for quality traits (essential nutrients, health promoting phytochemicals, volatiles, and aroma compounds). *Plant Biotechnology and Agriculture*, 443–461.
- Sato, S. (2015). Graphene for nanoelectronics. *Japanese Journal of Applied Physics*, 54(4): 1 - 12.
- Schafhaeuti, C. (1840). On the combinations of carbon with silicon and iron, and other metals, forming the different species of cast iron, steel, and malleable iron. *The London, Edinburgh, and Dublin Philosophical Magazine and Journal of Science Series 3*, 16(106): 570–590.
- Scott, S. M., James, D., & Ali, Z. (2006). Data analysis for electronic nose systems. *Microchimica Acta*, 156(3-4): 183–207.
- Seo, G. & Kirk, P. (2000). Ganodermataceae: Nomenclature and classification. *Ganoderma Diseases of Perennial Crops*: 3–22.
- Sha, R., Puttapati, S. K., Srikanth, V. V., & Badhulika, S. (2018). Ultra sensitive non-enzymatic ethanol sensor based on reduced graphene oxide-zinc oxide composite modified electrode. *IEEE Sensors Journal*, 18(5): 1844–1848.
- Shao, H. B., Chu, L. Y., Lu, Z. H., & Kang, C. M. (2008). Primary antioxidant free radical scavenging and redox signaling pathways in higher plant cells. *International Journal of Biological Sciences*, 4(1): 8–14.
- Sharin, S., Rahman, I. A., Ahmad, A. F., Mohd, H. M. K., Mohamed, F., Radiman, S., Yasir, M. S., Sarmani, S., Ayob, M. T. M., & Bastamam, I. S. A. (2015). Reduction of graphene oxide to graphene by using gamma irradiation. *Malaysian Journal of Analytical Sciences* 19(6): 1223–1228.
- Sharma, A., Singh, B., Dhar, S., Gondorf, A., & Spasova, M. (2012). Effect of surface groups on the luminescence property of ZnO nanoparticles synthesized by sol-gel route. *Surface Science*, 606(3–4): 13–17.
- Sharma, N., Sharma, V., Jain, Y., Kumari, M., Gupta, R. Sharma, S., & Sachdev, K. (2017). Synthesis and characterization of graphene oxide (GO) and reduced graphene oxide (rGO) for gas sensing application. [Special Issues] *Macromolecular Symposia*, 376(1): 1700006.

- Shiraz, H. G. & Tavakoli, O. (2017). Investigation of graphene-based systems for hydrogen storage. *Renewable and Sustainable Energy Reviews*, 74: 104–109.
- Shu'ud, M. M., Loonis, P., & Seman, I. A. (2007). Towards automatic recognition and grading of *Ganoderma* infection pattern using fuzzy systems. *International Journal of Medical and Health Sciences*, 1(1), 1–6.
- Si, Y. & Samulski, E. T. (2008). Exfoliated graphene separated by platinum nanoparticles. *Chemistry of Materials*, 20(21): 6792–6797.
- Singh, B. & Sharma, R. A. (2015). Plant terpenes: Defense responses, phylogenetic analysis, regulation and clinical applications. 3 *Biotech*, 5(2): 129–151.
- Singh, S., Arya, S. K., Pandey, P., Malhotra, B. Saha, S., Sreenivas, K., & Gupta, V. (2007). Cholesterol biosensor based on rf sputtered zinc oxide nanoporous thin film. *Applied Physics Letters*, 91(6): 063901.
- Singh, V., Jung, D., Zhai, L., Das, S., Khondaker, S. I., & Seal, S. (2011). Graphene based materials: Past, present and future. *Progress in Materials Science*, 56(8): 1178–1271.
- Smith, B. J. & Sivasithamparam, K. (2003). Morphological studies of *Ganoderma* (*Ganodermataceae*) from the Australasian and Pacific regions. *Australian Systematic Botany*, 16(4): 487–503.
- Sobon, G., Sotor, J., Jagiello, J., Kozinski, R., Zdrojek, M., Holdynski, M., Paletko, P., Boguslawski, J., Lipinska, L., & Abramski, K. M. (2012). Graphene oxide vs. reduced graphene oxide as saturable absorbers for Er-doped passively mode-locked fiber laser. *Optics Express*, 20(17): 19463–19473.
- Stoller, M. D., Park, S., Zhu, Y., An, J., & Ruoff, R. S. (2008). Graphene-based ultracapacitors. *Nano Letters*, 8(10): 3498–3502.
- Su, Y., Gao, X. & Zhao, J. (2014). Reaction mechanisms of graphene oxide chemical reduction by sulfur-containing compounds. *Carbon*, 67: 146–155.
- Šulčiūtė, A. & Valatka, E. (2012). Electrodeposition and photoelectrocatalytic activity of ZnO films on AISI 304 type steel. *Materials Science*, 18(4): 318–324.
- Sun, Y. F., Zhao, L. J. Jiang, T. J., Li, S. S., Yang, M., & Huang, X. J. (2016). Sensitive and selective electrochemical detection of heavy metal ions using amino-functionalized carbon microspheres. *Journal of Electroanalytical Chemistry*, 760: 143–150.

- Susanto, A., Sudharto, P., & Purba, R. (2005). Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia*, 159(1): 153–157.
- Taiz, L. & E. Zeiger (1995). *Plant physiology edition*. New Delhi, Bangalore: Panima Publishing Corporation, Courier companies, Inc.
- Torma, F., Kádár, M., Tóth, K., & Tatár, E. (2008). Nafion®/2, 2'-bipyridyl-modified bismuth film electrode for anodic stripping voltammetry. *Analytica chimica acta*, 619(2): 173-182.
- Unsicker, S. B., Kunert, G., & Gershenson, J. (2009). Protective perfumes: The role of vegetative volatiles in plant defense against herbivores. *Current Opinion in Plant Biology*, 12(4): 479–485.
- Utomo, C. & Niepold, F. (2000). Development of diagnostic methods for detecting *Ganoderma*-infected oil palms. *Journal of Phytopathology*, 148(9–10): 507–514.
- VanEtten, H., Temporini, E., & Wasmann, C. (2001). Phytoalexin (and phytoanticipin) tolerance as a virulence trait: Why is it not required by all pathogens? *Physiological and Molecular Plant Pathology*, 59(2): 83–93.
- Vaseem, M., Umar, A., & Hahn, Y.-B. (2010). ZnO nanoparticles: Growth, properties, and applications. *Metal Oxide Nanostructures and Their Applications*, 5: 1–36.
- Vickers, C. E., Gershenson, J., Lerdau, M. T., & Loreto, F. (2009). A unified mechanism of action for volatile isoprenoids in plant abiotic stress. *Nature Chemical Biology*, 5(5): 283–291.
- Villas-Bôas, S. G., Mas, S., Åkesson, M., Smedsgaard, J., & Nielsen, J. (2005). Mass spectrometry in metabolome analysis. *Mass Spectrometry Reviews*, 24(5): 613–646.
- Wang, B., Akiba, U., & Anzai, J. I. (2017). Recent progress in nanomaterial-based electrochemical biosensors for cancer biomarkers: A review. *Molecules*, 22(7): 1048.
- Wang, G., Wang, B., Park, J., Wang, Y., Sun, B., & Yao, J. (2009). Highly efficient and large-scale synthesis of graphene by electrolytic exfoliation. *Carbon*, 47(14): 3242–3246.
- Wang, J., Manga, K. K., Bao, Q., & Loh, K. P. (2011). High yield synthesis of few-layer graphene flakes through electrochemical expansion of graphite in propylene carbonate electrolyte. *Journal of the American Chemical Society*, 133(23): 8888–8891.

- Wang, J., Pedrero, M., Sakslund, H., Hammerich, O., & Pingarron, J. (1996). Electrochemical activation of screen-printed carbon strips. *Analyst*, 121(3): 345-350.
- Wang, L., Hu, C., & Shao, L. (2017). The antimicrobial activity of nanoparticles: Present situation and prospects for the future. *International Journal of Nanomedicine*, 12: 1227–1249.
- Wang, Z., Yang, Y., Li, J., Shen, G., & Yu, R. (2006). Organic–inorganic matrix for electrochemical immunoassay: Detection of human IgG based on ZnO/chitosan composite. *Talanta*, 69(3): 686–690.
- Wayu, M. B., Spidle, R. T., Devkota, T., Deb, A. K., Delong, R. K., Ghosh, K. C., Wanekaya, A. K., & Chusuei, C. C. (2013). Morphology of hydrothermally synthesized ZnO nanoparticles tethered to carbon nanotubes affects electrocatalytic activity for H₂O₂ detection. *Electrochimica Acta*, 97: 99–104.
- Wen, W. (2016). Introductory chapter: What is chemical sensor?. In *Progresses in chemical sensor*, ed. W. Wang, pp. 3–8. United Kingdom: InTech.
- Wu, Z., Song, L. & Huang, D. (2011). Food grade fungal stress on germinating peanut seeds induced phytoalexins and enhanced polyphenolic antioxidants. *Journal of Agricultural and Food Chemistry*, 59: 5993-6003.
- Wink, M. (1999). *Functions of plant secondary metabolites and their exploitation in biotechnology* (pp. 488 – 488). US: Taylor & Francis.
- Xu, L., Guo, Y., Liao, Q., Zhang, J., & Xu, D. (2005). Morphological control of ZnO nanostructures by electrodeposition. *The Journal of Physical Chemistry B*, 109(28): 13519–13522.
- Ya, Y., Wang, T., Xie, L., Zhu, J., Tang, L., Ning, D., & Yan, F. (2015). Highly sensitive electrochemical sensor based on pyrrolidinium ionic liquid modified ordered mesoporous carbon paste electrode for determination of carbendazim. *Analytical Methods*, 7(4): 1493–1498.
- Yagati, A. K., Park, J., & Cho, S. (2016). Reduced graphene oxide modified the interdigitated chain electrode for an insulin sensor. *Sensors*, 16(1): 109.
- Yang, L., Wang, F., Han, H., Yang, L., Zhang, G., & Fan, Z. (2015). Functionalized graphene oxide as a drug carrier for loading pirfenidone in treatment of subarachnoid hemorrhage. *Colloids and Surfaces B: Biointerfaces*, 129: 21–29.
- Yi, R., Zhang, N., Zhou, H., Shi, R., Qiu, G., & Liu, X. (2008). Selective synthesis and characterization of flower-like ZnO microstructures via a facile hydrothermal route. *Materials Science and Engineering: B*, 153(1–3): 25–30.

- Yoshida, T., Komatsu, D., Shimokawa, N., & Minoura, H. (2004). Mechanism of cathodic electrodeposition of zinc oxide thin films from aqueous zinc nitrate baths. *Thin Solid Films*, 451–452: 166–169.
- Zain, N., Seman, I., Kushairi, A., & Ramli, U. (2013). Metabolite profiling of oil palm towards understanding basal stem rot (BSR) disease. *Journal of Oil Palm Research*, 25(1): 58–71.
- Zamborini, F. P., Bao, L., & Dasari, R. (2011). Nanoparticles in measurement science. *Analytical Chemistry*, 84(2): 541–576.
- Zhang, J., Zhang, F., Yang, H., Huang, X., Liu, H., Zhang, J., & Guo, S. (2010). Graphene oxide as a matrix for enzyme immobilization. *Langmuir*, 26(9): 6083–6085.
- Zhang, Y., Tan, Y. W., Stormer, H. L., & Kim, P. (2005). Experimental observation of the quantum Hall effect and Berry's phase in graphene. *Nature*, 438(7065): 201–204.
- Zheng, A. L. T., Andou, Y., & Zawawi, R. M. (2017). Effects of Deposition Parameters on the Electrochemical Behaviour of ZnO Thin Film. *Journal of Advanced Chemical Sciences*, 3(4): 521-524.
- Zhou, W.J., Zhao, D. D., Xu, M. W., Xu, C. L., & Li, H. L. (2008). Effects of the electrodeposition potential and temperature on the electrochemical capacitance behavior of ordered mesoporous cobalt hydroxide films. *Electrochimica Acta*, 53(24): 7210-7219.
- Zhu, H. & Xu, G. (2017). Electrochemical Determination of Ascorbic Acid Based on Hydrothermal Synthesized ZnO Nanoparticles. *International Journal of Electrochemistry Society*, 12: 3873–3882.
- Zhu, Z., Su, Y., Li, J., Li, D., Zhang, J., Song, S., Zhao, Y., Li, G., & Fan, C. (2009). Highly sensitive electrochemical sensor for mercury (II) ions by using a mercury specific oligonucleotide probe and gold nanoparticle based amplification. *Analytical Chemistry*, 81(18): 7660–7666.
- Zhuang, Y., Sun, J., & Guan, Y. (2015). Electrodeposition of ZnO-RGO thin film for determining Hg (II) in water samples. *International Journal of Electrochemical Science*, 10(7): 5961–5969.

BIODATA OF STUDENT



Nurhamizah binti Rahmat was born on 2nd May 1992 at Hospital Kajang, Selangor. She received her primary education in Sekolah Kebangsaan Ulu Semenyih, Semenyih, Selangor and secondary education in SMKA Maahad Hamidah Kajang, Kajang, Selangor. After that, she went to Universiti Sains Islam Malaysia (USIM) to further her studies. In 2015, she graduated as a bachelor student holding the certificate of Bachelor of Science with Honours, majoring in Industrial Chemistry from the Faculty of Science and Technology. In 2016, she continued her studies in Master of Science at the Institute of Advanced Technology (ITMA), Universiti Putra Malaysia under the supervision of Prof. Dr. Nor Azah Yusof and Assoc. Dr. Wong Mui Yun. During her studies, she became a laboratory demonstrator to assist and monitor undergraduate students taking Physical and Inorganic Chemistry, and Organic 1 subjects.

LIST OF PUBLICATIONS AND CONFERENCE ATTENDED

Publications

Rahmat, N., & Yusof, N. A. (2017). Modification of SPCE by Reduction of Graphene Oxide and Electrodeposition of Zinc Oxide Nanoparticles for Electrochemical Sensor. *Malaysian Journal of Catalysis*, 3(1): 32-35.

Detection of Secondary Metabolites in Oil Palm Trees using ZnO-NPs/rGO/SPCE Modified-Screen Printed Electrode (NMAS 2017 - Submitted).

Conferences

1. NanoMITe Annual Symposium (NMAS 2016) on 28th September 2016 in University Technology Malaysia (UTM KL) as oral presenter.
2. Symposium on Advanced Materials and Nanotechnology (SAMN 2017) on 18th – 19th of July 2017 at Hotel Bangi- Putrajaya, Malaysia as oral presenter.
3. NanoMITe Annual Symposium (NMAS 2017) on 14th – 15th November 2017 in Universiti Putra Malaysia (UPM) Serdang, Malaysia as oral presenter.

Project meeting and others

1. Colloquium On Nanotechnology in Detection and Control of *Ganoderma boninense* (CONGRAB 7) on 28th September 2017 in Cyberview Lodge, Cyberjaya.
2. Nanotechnology in Detection and Control of *Ganoderma boninense* (CONGRAB 8) on 14th – 15th December 2017 at Institute of Nano Electronic Engineering, Universiti Malaysia Perlis and Langkawi Research Centre, Langkawi, Kedah Darul Aman.
3. Summer School Nanosciences: Fundamental and Applications Emphasizing in Nanomedicine on 24th July to 5th August 2017 at Institut Teknologi Bandung (ITB) Bandung, Indonesia.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT
AND COPYRIGHT

ACADEMIC SESSION : Second Semester 2018/2019

TITLE OF THESIS / PROJECT REPORT :

ZnO-NPs/rGO/SPCE-MODIFIED ELECTRODE FOR EARLY DETECTION
OF *Ganoderma boninense* IN OIL PALM TREES

NAME OF STUDENT : NURHAMIZAH BINTI RAHMAT

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as:

*Please tick (✓)

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open acces

This thesis is submitted for:



PATENT

Embargo from _____ until
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

Date :

(Signature of Chairman
of Supervisory Committee)
Name:

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]

© COPY

RIGHT

UPM