

UNIVERSITI PUTRA MALAYSIA

CELLULASE ACTIVITY ON THERMOPHILIC FUNGI

MUHAMMAD ADIB AZHAN

FBSB 2015 113

CELLULASE ACTIVITY ON THERMOPHILIC FUNGI



MUHAMMAD ADIB AMIN BIN AZHAN

THESIS

Department of Microbiology Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia

2015

ACKNOWLEDGEMENTS

In the name of Allah swt, the Most Merciful and Most Compassionate.

First and foremost, I would like to thank my supervisor Dr. WanZuhainis Saad for her invaluable supervision and support throughout the year.

I also would like to express my deepest gratitude and appreciation to Muhammad Hariadi for his valuable guidance, advice, suggestion and discussion throughout the completion of this project while I knew very little.

My sincere appreciation to all my seniors in the Mycology lab especially Liyin Wong and all the staffs in the Department Microbiology for their help.

Last but not least, I would like to convey my very special gratitude and love to my parents and my brother and sister for their endless love, care and support.

> Muhammad Adib Amin bin Azhan June 2015

LIST FIGURES

FIGURES

 \bigcirc

2a:	A 3D illustration of the endoglucanase complex					
	(quaternary structure) with cellulose in its active site7					
4a:	Hotspring area where the first sample isolated and processed	3				
4b:	Hotspring area where the second sample isolated and processed1	8				
4c:	Front view and back view growth of Aspergillus fumigatus					
	isolate 1 on Potato Dextrose Agar plates	0				
4d:	Front view and back view growth of Aspergillus fumigatus					
	isolate 2 on Potato Dextrose Agar plates	1				
4e:	Aspergillus fumigatus isolate 1 subculture 1 under microscope2	2				
4f:	Aspergillus fumigatus isolate 1 subculture 2 under microscope2	3				
4g:	Aspergillus fumigatus isolate 1 subculture 3 under microscope2	4				
4h:	Aspergillus fumigatus isolate 2 subculture 1 under microscope2	5				
4i:	Aspergillus fumigatus isolate 2 subculture 2 under microscope2	7				
4j:	Aspergillus fumigatus isolate 2 subculture 3 under microscope2	8				
4k:	Clear zone of first isolate appear on CMC agar plates)				
41:	Clear zone of second isolate appear on CMC agar plates	1				
4m:	Fungi are incubated in the incubator under					

	Solid State Fermentation	32
4n:	Filtration for fungi crude enzyme	33
40:	Graph of absorbance versus glucose	35
4p:	Graph of absorbance versus ρ -Nitrophenol	40

LIST OF TABLES

TABLES

5

PAGE

4a:	Morphological feature and cellulolytic index	
	of cellulolytic thermophilic isolates	29
4b:	Absorbance reading of glucose standard	34
4c:	Absorbance reading of endoglucanase activity	36
4d:	Absorbance reading of total cellulase activity	38
4e:	Absorbance reading of nitrophenol standard	39
4f:	Absorbance reading of β -glucosidase activity	40

LIST OF ABBREVIATIONS

SSF	solid state fermentation	hoNPG	ρ -nitrophenol- β -
			glucoside
М	molar	mm	millilitre
GH	glycosidase hydrolase	mg	miligram
°C	degree centrigrade	μl	microliter
PDA	potato dextrose agar	IU/ml	international unit / mililitre
СМС	carboxymethyl cellulose	FPU	filter paper unit
nm	nanometre		
$NH_4H_2PO_4$	ammonium hydrogen phosphat	te	
KCl	potassium chloride		
$MgSO_4.7H_20$	magnesium sulphate heptahydr	rate	
NaCl	sodium chloride		
CI	cellulolytic index		
ml	millilitre		
w/v	weight per volume		
$(NH_4)_2PO_4$	diammonium phosphate		
g	gram		
KH ₂ PO ₄	potassium dihydrogen phospha	ate	
$MgS0_4$	magnesium sulphate		
psi	per square inch		
rpm	revolutions per minute		
xg	centrifugal force		
Min	minute		
mM	mili molar		
DNS	3,5-dinitrosalicylic acid		
μmol	micro mole		

ABSTRACT

Thermophilic fungi produce thermostable enzymes which are very stable at extreme and elevated temperature. One of the enzyme produced by thermophilic fungi is cellulase. Interaction of three components of cellulase enzyme in degrading cellulose into its small subunits make cellulase became popular in industrial processes. The sample of thermophilic fungi isolated from hotspring area diluted using serial dilution method grow on potato dextrose agar after incubated at 50°C. The function grow of fungi on potato dextrose agar is to get the single colony of thermophilic fungi. The single colony of thermophilic fungi subcultured into carboxymethyl cellulose agar (CMC) to determine its cellulase production. The fungi identified through macroscopic and microscopic method in order to know the species of fungi. Cellulase comprise of three components which are endoglucanase, exoglucanase and β glucosidase. The three components of cellulase tested in each specific assay such as filter paper assay, endoglucanase assay and β -glucosidase assay. Filter paper assay determine the total cellulase activity, endoglucanase assay determine endoglucanase activity and β -glucosidase assay determine β -glucosidase activity.

ABSTRAK

Kulat thermophilic menghasilkan enzim tahan panas yang sangat stabil pada suhu yang melampau dan tinggi. Salah satu enzim yang dihasilkan oleh kulat thermophilic adalah selulase. Interaksi tiga komponen enzim selulase dalam meleraikan selulosa ke subunit kecil membuat selulase menjadi popular dalam proses perindustrian. Sampel kulat thermophilic diasingkan dari kawasan kolam air panas dicairkan menggunakan kaedah pencairan bersiri tumbuh di kentang dekstrosa agar selepas dieram pada 50 °C. Fungsi tumbuh kulat diatas kentang dekstrosa agar adalah untuk mendapatkan satu koloni kulat thermophilic. Satu koloni tunggal kulat thermophilic disubkulturkan ke carboxymethyl selulosa agar (CMC) untuk menentukan pengeluaran selulasenya. Kulat yang dikenal pasti melalui kaedah makroskopik dan mikroskopik untuk mengetahui spesies kulat. Selulase terdiri daripada tiga komponen iaitu endoglucanase, exoglucanase dan β -glucosidase. Ketiga-tiga komponen selulase diuji dalam setiap assay tertentu seperti penapis assay kertas, assay endoglucanase dan β glucosidase assay. Penapis kertas assay menentukan keseluruhan aktiviti selulase, endoglucanase assay menentukan aktiviti endoglucanase dan β -glucosidase assay menentukan aktiviti β-glucosidase.

TABLE OF CONTENT

CHAPTER		TITLE	PAGE
TABLE OF CONTENT			i
CHAPTER 1		INTRODUCTION	1
CHAPTER 2		LITERATURE REVIEW	5
	2.1	Cellulase Enzyme	5
	2.2	Endo-1,4- β -glucanase	7
	2.3	Exo- β -1,3-glucanase	8
	2.4	β -glucosidase	8
	2.5	Cellulose	9
	2.6	Enzymatic Reaction of Cellulase	9
	2.7	Cellulolytic Potential	10
	2.8	The Production of Bio-fuels from Lignocellulosic Biomass	11
CHAPTER 3		MATERIALS AND METHODS	13
	3.1	Sample Collection and Processing	13
	3.2	Qualitative Screening of Thermophilic Fungi	13
	3.3	Identification of Thermophilic Fungi	14
	3.4	Quantitative Determination of Thermop Fungi by Solid State Fermentation	hilic 14
	3.5	Enzyme Assay	15
		3.5.1 Endoglucanase Activity : Carboxymethyl Cellulose Activity	y 15
		3.5.2 Filter Paper Assay : Total Cellulase Activity	16
		3.5.3 β -Glucosidase assay	16

CHAPTER	TITLE	PAGE
CHAPTER 4	RESULTS AND DISCUSSION	18
	4.1 Sample Collection and Processing	18
	4.2 Identification of Thermophilic Fungi	19
	4.3 Qualitative Screening of Thermophilic Fungi	29
	4.4 Quantitative Determination of Thermoph Fungi by Solid State Fermentation	ilic 32
	 4.5 Enzyme Assay 4.6 Endoglucanase Activity : Carboxymethyl Cellulose Activity 	33
	4.7 Filter Paper Assay : Total Cellulase Activity	38
	4.8 β -Glucosidase assay	39
CHAPTER 5	5.0 CONCLUSION	42
REFERENCES		43

C

 \bigcirc

1.0 INTRODUCTION

Fungi is an organisms called the decomposers grow in the soil or on dead plant matter where they play an important role in the cycling of carbon and other elements. Fungi also one of important organisms in biodiversity mainly in degrading plant materials such as cellulose, hemicellulose and lignin. Fungi degrade plant materials by secreting crucial enzymes that are stable in extreme environment. Thus fungi become popular in industrial use due to their useful enzymes. For instance, cellulase enzyme secreted by fungi used for exchange of biomass to fermentable sugars in biorefineries and in textiles, paper, and detergent industries (Karmakar and Ray, 2011; Kuhad *et al.*, 2011).

The only one organism in eukaryotic group that have capability to survive at the elevated temperature was called thermophilic fungi. Thermophilic fungi are species that grow at a lowest temperature of 20°C or above, and a highest temperature of 50°C or above (Maheshwari *et al.*, 2000). Therefore they are the only representatives of mycoflora that can grow at high temperature above 45°C. Despite thermophilic fungi are able to grow at elevated temperature, but they offer faster growth rates as compared with mesophilic fungi, an organism that grows best in moderate temperature (Ashraf *et al.*, 2007). Nevertheless, thermophilic fungi are not as extreme as in eubacteria and archaea, which are characterized by their ability to tolerate extreme temperature at 100°C and live in hydrothermal vents (Brock, 1995). The known orders of thermophilic fungi are Sordariales, Eurotiales, and Onygenales from phylum ascomycetes and order Mucorales from phylum zygomycetes (Berka *et al.*, 2011; Morgenstern *et al.*, 2012).

A lot of information about the location of thermophilic fungi from various types of soils and in habitats where decomposition of plant materials takes place whether in tropical as well as temperate regions. In nature, thermophilic fungi are usually found in composts, piles of hays, wood chip piles, stored grains, animal dung, nesting material of birds and animals, snuff, municipal refuse, other environments that are selfheating due to degradation of plant materials, and other accumulations of organic matter where in the warm, humid, and aerobic environment that provides the basic physiological conditions to their development (Johri et al., 1999). In these habitats, thermophilic fungi may occur either as resting propagules or as active mycelia rely on the presence of nutrients and favourable environmental conditions. Thermophilic fungi can be measured based on their cellulolytic potential whether they are good cellulosedegraders or poor cellulose-degraders. Since such material often contains high concentrations of cellulose some good cellulose-degraders will degrade cellulose easily, but some poor cellulose-degraders seem to utilize sugars released by cellulolytic species in the biotope (Maheshwari et al., 2000). Therefore, thermophilic fungi can differ greatly in their cellulolytic potential.

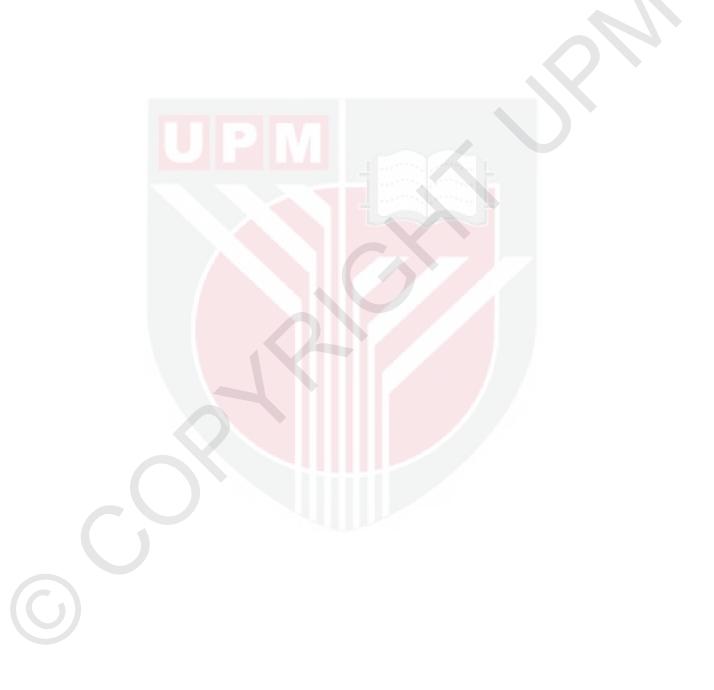
Thermophilic fungi have a great capability to break down plant materials and polysaccharides constituents of biomass like cellulose and become the potential source of cellulolytic enzyme with scientific and commercial importance. Mesophilic fungi produce enzymes that are usually effective at temperature below than 50°C whereas thermophilic fungi produce more thermostable enzymes that able to function at temperatures up to 70°C (Murray *et al.*, 2004; Parry *et al.*, 2002; Venturi *et al.*, 2002; Voutilainen *et al.*, 2008). Several factors of thermophilic fungi that make the industrial process more economical such as their thermostable enzyme, ability to saccharify

under non-aseptic conditions, and high rate of cellulolysis (Haggerdal *et al.*, 1980; Merchant *et al.*, 1988; Maheshwari *et al.*, 2000; Sohail *et al.*, 2009). Other than that, thermostable enzymes which have been isolated from these fungi, have found a number of commercial applications because of their overall inherent stability (Demirijan *et al.*, 2001). Examples of thermophilic fungi that produce thermostable cellulases are Talaromyces emersonii (Murray *et al.*, 2004; Voutilainen *et al.*, 2010), Myceliophthora thermophila (Roy *et al.*, 1990), *Chaetomium thermophilum* and *Acremonium thermophilum* (Voutilainen *et al.*, 2008).

There are several problem statements arise in this study. Firstly, to solve problem of inadequate sources of industrially relevant thermostable enzymes. Secondly, stimulated isolation of a number of microbes from thermal environments in order to access enzymes that could significantly increase the window for enzymatic bioprocess operations. Another problem has to solve in technical products and processes, often in a very large scale using industrial enzymes from thermophilic fungi. There are also some hypothesis about this study. Firstly, thermophile cellulase are key enzymes for efficient biomass degradation. Their importance stem from the fact that cellulose swells at higher temperatures, thereby becoming easier to break down. Secondly, thermophilic fungi and their proteins are able to function at elevated temperatures (high temperatures, above 55°C). Thirdly, thermophilic enzyme rarely require toxic metal ions for functionality, hence creating the possibility to use more environmentally friendly processing. Lastly, thermostable enzymes offer robust catalyst alternatives, able to withstand the often relatively harsh conditions of industrial processing.

Therefore, the specific objectives of this study were :

- 1. To Isolate and identify thermophilic fungi from hotspring,
- 2. To screen the cellulolytic enzymes from the fungal isolates.



REFERENCES

- Adsul M. G., K.B. Bastawde, A.J. Varma, D.V. Gokhale., (2007). Strain improvement of Penicillium janthinellum NCIM 1171 for increased cellulase production. Bioresour. Technol., 98, pp. 1467–1473.
- Andric P, Meyer AS, Jensen PA, Johansen KD., (2010) Reactor design for minimizing product inhibition during enzymatic lignocellulose hydrolysis: I. Significance and mechanism of cellobiose and glucose inhibition on cellulolytic enzymes. Biotechnol Adv 28:308–324.
- Ashraf, R., F. Shahid and T.A. Ali., (2007). Association of fungi, bacteria and actinomycetes with different composts. Pak. J. Bot., 39(6): 2141-2151.
- Balat M., (2007). An overview of biofuels and policies in the European Union countries. Energy Sources Part B, 2, pp. 167–181.
- Banerjee G, Car S, Scott-Craig JS, Borrusch MS, Aslam N, Walton JD., (2010). Synthetic enzyme mixtures for biomass deconstruction: production and optimization of a core set. Biotechnol Bioeng. 3:707–20. doi: 10.1002/bit.22741.
- Beg Q. K., B. Bhushan, M. Kapoo, G.S. Hoondal., (2000). Enhanced production of a thermostable xylanase from Streptomyces sp. QG-11-3 and its application in biobleaching of eucalyptus kraft pulp. Enzyme Microbiol. Technol., 27, pp. 459–466.
- Benko Z, Drahos E, Szengyel Z, Puranen T, Vehmaanpera J, Reczey K.,(2007). Thermoascus aurantiacus CBHI/Cel7A production in Trichoderma reesei on alternative carbon sources. Applied Biochemistry and Biotechnology. 137–140(1–12):195–204.
- Bergmeyer, H.U., (1983). Methods of Enzymatic Analysis, 3rded. Verlag Chemie, Weinheim.

Bisswanger, H., (2011). Practical Enzymology, 2nded. Wiley- Blackwell, Weinheim.

Bradford M. M., (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Analytical Biochemistry, 72, pp. 248–254.

Brenda database (www.brenda-enzymes.org).

- Brock T. D. (1995) The road to Yellowstone—and beyond. Annu. Rev. Microbiol. 49:1–28.
- Chandra M, Kalra A, Sharma PK, Sangwan RS. (2009). Cellulase production by six Trichoderma spp. fermented on medicinal plant processings. J Ind Microbiol Biotechnol 36: 605–609.

Charitha Dev M, et al., (2012). J. Microbiol. Biotech. Res., 2 (1): 120-128.

Collins CM, Murray PG, Denman S, et al., (2007). Molecular cloning and expression analysis of two distinct β-glucosidase genes, bg1 and aven1, with very

different biological roles from the thermophilic, saprophytic fungus Talaromyces emersonii. Mycological Research. 111(7):840–849.

- Dashtban M, Schraft H, Qin W., (2009). Fungal bioconversion of lignocellulosic residues; opportunities & perspectives. Int J Biol Sci. 3:578–595.
- Demirbas A., (2008). Biomethanol production from organic waste materials. Energy Sources Part A, 30, pp. 565–572.
- Demirbas A., (2008). The importance of bioethanol and biodiesel from biomass. Energy Sources Part B, 3, pp. 177–185.
- Demirijan D., F. Moris-Varas, C. Cassidy., (2001). Enzymes from extremophiles. Curr. Opin. Chem. Boil., 5, pp. 144–151.
- Du F, Wolger E, Wallace L, Liu A, Kaper T, Kelemen B (2010). Determination of product inhibition of CBH1, CBH2, and EG1 using a novel cellulase activity assay. Appl Biochem Biotechnol 161:313–317.
- Duenas R., R. Tengerdy, M. Gutierrez-corea., (1995). Cellulase production by mixed fungi in solid–substrate fermentation of bagasse. W. J. Microbiol. Biotechnol., 11, pp. 333–337.
- Emerson R., (1968). Thermophiles in The fungi, an advanced treatise, eds Ainsworth G. C., Sussman A. S. (Academic Press, Inc. New York, N.Y), 3:105–128.
- ExPASydatabase (www.expasy.org/enzymes).
- Gerday, C.,(2007). Physiology and Biochemistry of Extremophiles. ASM Press,Washington.
- Ghose TK., (1987) Measurement of cellulase activities. Pure Appl. Chem. 59:257–268.
- Gomes I, Gomes J, Gomes DJ, Steiner W., (2000). Simultaneous production of high activities of thermostable endoglucanase and β-glucosidase by the wild thermophilic fungus Thermoascus aurantiacus. Appl Microbiol Biotechnol. 3:461–468.
- Grassick A, Murray PG, Thompson R, et al., (2004). Three-dimensional structure of a thermostable native cellobiohydrolase, CBH IB. molecular and characterization of the cel7 gene from the filamentous fungus, Talaromyces European Journal Biochemistry. emersonii. of 271(22):4495-4506.
- Haggerdal, B., F.D. Ferchak and E.K. Pye. (1980). Saccharification of the enzyme system of Thermomonospora sp: Stability of cellulolytic activity with respect to time, temperature and pH. Biotechnol. Bioeng., 22: 1515-1526.
- Hammond G. M., S. Kallu, M.C. McManus., (2009). Development of biofuels for the UK automotive market. Appl Energy, 86, pp. 506–515.
- Harikrishna S., K.C.S. Rao, J.S. Babu, D.S. Reddy., (2000). Studies on the production and application of cellulase from Trichoderma reesei QM9414. Bioprocess Eng., 22, pp. 467–470.

- Harris PV, Welner D, McFarland KC, Re E, Navarro Poulsen JC, Brown K, Salbo R, Ding H, Vlasenko E, Merino S, Xu F, Cherry J, Larsen S, Lo Leggio L., (2010). Stimulation of lignocellulosic biomass hydrolysis by proteins of glycoside hydrolase family 61: structure and function of a large, enigmatic family. Biochemistry.
- Henrissat B, Davies G., (1997). Structural and sequence-based classification of glycoside hydrolases. Curr Opin Struct Biol. 3:637–644. doi: 10.1016/S0959-440X(97)80072-3.
- Holker U., M. Hofer, J. Lenz., (2004). Biotechnological advantages of laboratory-scale solid- state fermentation with fungi. Appl. Microbiol. Biotechnol., 64, pp. 175–186.
- Hong J, H. Tamaki, S. Akiba, K. Yamamoto, H. Kumagai, (2001). Cloning of a gene encoding a highly stable endo-β-1,4-glucanase from Aspergillus niger and its expression in yeast. J. Biosci. Bioeng., 92, pp. 434–441.
- Hong J, Tamaki H, Yamamoto K, Kumagai H., (2003). Cloning of a gene encoding thermostable cellobiohydrolase from Thermoascus aurantiacus and its expression in yeast. Appl Microbiol Biotechnol. 3:42–50.
- Hong J, Tamaki H, Kumagai H., (2007). Cloning and functional expression of thermostable β-glucosidase gene from Thermoascus aurantiacus. Appl Microbiol Biotechnol. 3:1331–1339.
- James, G. C. and Natalie, S. (2001). Microbiology. A laboratory Manual (ed.). Pp. 211-223.
- Johri BN, Satyanarayana T, Olsen J. (1999). Thermophilic moulds in biotechnology. 1. Dordrecht, The Netherlands : Kluwer Academic Publishers.
- Juhasz T., K. Kozma, Z. Szengyel, K. Reczey., (2003). Production of β-glucosidase in mixed culture of Aspergillus niger BKMF 1305 and Trichoderma reesei RUT C30. Food Technol. Biotechnol., 41 (1), pp. 49–53.
- Juhasz T., Z. Szengyel, N. Szijarto, K. Reczey., (2004). Effect of pH on cellulase production of Trichoderma reesei RUT C30. Appl. Biochem. Biotechnol., 113, pp. 201–211.
- Karmakar M, Ray RR (2011). Current trends in research and application of microbial cellulases. Res J Microbiol 6:41-53.
- Kaur J, B.S. Chadha, B.A. Kumar, H.S. Saini. (2007). Purification and characterization of two endoglucanases from Melanocarpus sp. MTCC 3922. Bioresour. Technol., 98, pp. 74–81.
- Khandke KM, Vithayathil PJ, Murthy SK., (1989). Purification of xylanase, β -glucosidase, endocellulase, and exocellulase from a thermophilic fungus, Thermoascus aurantiacus. Arch Biochem Biophys. 3:491–500.
- Khokhar I., Haider M. S., Mushtaq S. & Mukhtar I. (2012). Isolation and screening of highly cellulolytic filamentous fungi. Journal of Applied Sciences Environmental Management, 16, 223-226.

- Kotchoni S. O., Shonukan O. O., (2002). Regulatory mutations affecting the synthesis of cellulase in Bacillus pumilus. World J. Microbiol. Biotechnol., 18, pp. 487–491.
- Kuhad RC, Gupta R, Singh A (2011). Microbial cellulases and their industrial applications. Enzyme Res 2011:1-10.
- Kumar, R., Sing, S., Singh, O.V. (2008). Bioconversion of lignocellulosic biomass: biochemical and molecular perspectives. J Ind Microbiol Biotechnol. 35: 377-391.
- Langston JA, Shaghasi T, Abbate E, Xu F, Vlasenko E, Sweeney MD. (2011). Oxidoreductive cellulose depolymerisation by the enzymes cellobiose dehydrogenase and glycoside hydrolase 61. Appl Environ Microbiol. 3:7007–7015.
- Li Y H, M. Ding, J. Wang, G.J. Xu, F. Zhao. (2006). A novel thermoacidophilic endoglucanase, Ba-EGA, from a new cellulose-degrading bacterium, Bacillus sp. AC-1. Appl. Microbiol. Biotechnol., 70, pp. 430–436.
- Maheshwari, R., G. Bharadwaj and M.K. Bhat. (2000). Thermophilic Fungi: Their physiology and enzymes. Microbiol. Mol. Biol. Rev., 63(3): 461-488.
- Mandels. M, J. Weber. (1969). The production of cellulases Advances in Chemistry: American Chemical Society, pp. 391–414.
- Mandels, M., R. Andreotti and R. Roche. (1976). Biotech. Bioeng. Symp., 6:17-37.
- Merchant, R., F. Merchant and A. Margaritis. (1988). Production of xylanase by thermophilic fungus Thielavia terrestris. Biotechnol. Lett., 10: 513-516.
- Miehe H. (1907). Die Selbsterhitzung des Heus. Eine biologische Studie. (Gustav Fischer Verlag, Jena, Germany).
- Miller G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. Analytical Chemistry, 31, pp. 426–428.
- Murray P, Aro N, Collins C, Penttilä M, Saloheimo M, Tuohy M., (2004). Expression in Trichoderma reesei and characterisation of a thermostable family 3 β -glucosidase from the moderately thermophilic fungus Talaromyces emersonii. Protein Expr Purif. 3:248–257.
- Murray PG, Collins CM, Grassick A, Tuohy MG. (2003). Molecular cloning, transcriptional, and expression analysis of the first cellulase gene (cbh2), encoding cellobiohydrolase II, from the moderately thermophilic fungus Talaromyces emersonii and structure prediction of the gene product. Biochemical and Biophysical Research Communications. 301(2):280–286.
- Muthuvelayudham R., T. Viruthagiri, T. Selvapandian. (2005). Biosynthesis of cellulose protein on substrates like cellulose, xylose and lactose using Trichoderma reesei, Annamalai University. J. Eng. Technol. pp. 118–121.
- Nimlos, M.R., Matthews, J.F., Crowley, M.F., Walker, R.C., Chukkapalli, G., Brady, J.W., Adney, W.S., Cleary, J.M., Zhong, L., Himmel, M.E. (2007). Molecular modeling suggests indcued fit of Family 1 carbohydrate-binding

modules with a broken-chain cellulose. Protein engineering Design and Selection. 20(4):179-187.

- Olsson L., T.M.I.E. Christensen, K.P. Hansen, E.A. Palmqvist. (2003). Influence of the carbon source on production of cellulases, hemicellulases and pectinases by Trichoderma reesei RUT C-30. Enzyme Microbiol. Technol., 33 (5), pp. 612–619.
- Parry NJ, Beever DE, Owen E, Vandenberghe I, Van Beeumen J, Bhat MK. (2001). Biochemical characterization and mechanism of action of a thermostable βglucosidase purified from Thermoascus aurantiacus. Biochem J. 3:117–127.
- Parry NJ, Beever DE, Owen E, Nerinckx W, Claeyssens M, Van Beeumen J, Bhat MK. (2002). Biochemical characterization and mode of action of a thermostable endoglucanase purified from Thermoascus aurantiacus. Arch Biochem Biophys. 3:243–253.
- Pocas-Fonseca MJ, Silva-Pereira I, Rocha BB, Azevedo MDO. (2000). Substratedependent differential expression of humicola grisea var. thermoidea cellobiohydrolase genes. Canadian Journal of Microbiology. 46(8):749– 752.
- Quinlan RJ, Sweeney MD, Lo Leggio L, Otten H, Poulsen JC, Johansen KS, Krogh KB, Jørgensen CI, Tovborg M, Anthonsen A, Tryfona T, Walter CP, Dupree P, Xu F, Davies GJ, Walton PH. (2011). Insights into the oxidative degradation of cellulose by a copper metalloenzyme that exploits biomass components. Proc Natl Acad Sci USA. 3:15079–15084.
- Rainey, F.A., Oren, A., (2006). Methods Microbiol.35,1-25.
- Romanelli RA, Houston CW, Barnett SM. (1975). Studies on thermophilic cellulolytic fungi. Appl Microbiol. 3:276–281.
- Roy SK, Dey SK, Raha SK, Chakrabarty SL. (1990). Purification and properties of an extracellular endoglucanase from Myceliophthora thermophila D-14 (ATCC 48104) J Gen Microbiol. 3:1967–1971.
- Schomburg, D., Schomburg, I. (Eds.), (2009). Springer Handbook of Enzymes. Second Edition. Springer, Berlin.
- Senthilkumar S. R., B. Ashokkumar, K.C. Raj, P. Gunasekaran. (2005). Optimization of medium composition for alkali-stable xylanase production by Aspergillus fischeri Fxn 1 in solid-state fermentation using central composite rotary design. Bioresour. Technol., 96, pp. 1380–1386.
- Sohail, M., S. Naseeb, S.K. Sherwani, S. Sultana, S. Aftab, S. Shahzad, A. Ahmad and S.A. Khan. (2009). Distribution of hydrolytic enzymes among native fungi: Aspergillus the pre-dominant genus of hydrolase producer. Pak. J. Bot., 41(5): 2567-2582.
- Soma Mrudula and Rangasamy Murugammal. (2011). Production of cellulase by Aspergillus niger under submerged and solid state fermentation using coir waste as a substrate. Braz. J. Microbiol. vol.42 no.3 São Paulo July/Sept.

- Sukumaran RK, Surender VJ, Sindhu R, et al. (2010). Lignocellulosic ethanol in India prospects, challenges and feedstock availability. Bioresource Technology. 101(13):4826–4833.
- Suto M, Tomita F. (2001). Induction and catabolite repression mechanisms of cellulase in fungi. Journal of Bioscience and Bioengineering. 92(4):305–311.
- Venturi LL, de Polizeli M, Terenzi L, Furriel HF, Rdos P, Jorge JA. (2002). Extracellular β-D-glucosidase from Chaetomium thermophilum var. coprophilum: production, purification and some biochemical properties. J Basic Microbiol. 3:55–66.
- Vieille, C., Zeikus, J.G., (2001). Microbiol. Mol. Biol. Rev. 65, 1-43.
- Vlasenko E, Schülein M, Cherry J, Xu F. (2010). Substrate specificity of family 5, 6, 7, 9, 12, and 45 endoglucanases. Bioresource Technology. 101(7):2405–2411.
- Voutilainen SP, Puranen T, Siika-Aho M, Lappalainen A, Alapuranen M, Kallio J, Hooman S, Viikari L, Vehmaanperä J, Koivula A. (2008). Cloning, expression, and characterization of novel thermostable family 7 cellobiohydrolases. Biotechnol Bioeng. 3:515–528.
- Voutilainen SP, Murray PG, Tuohy MG, Koivula A. (2010). Expression of Talaromyces emersonii cellobiohydrolase Cel7A in Saccharomyces cerevisiae and rational mutagenesis to improve its thermostability and activity. Protein Eng Des Sel. 3:69–79.
- Wilson DB. (2009). Cellulases and biofuels. Current Opinion in Biotechnology. 20(3):295–299.
- Westereng B, Ishida T, Vaaje-Kolstad G, Wu M, Eijsink VG, Igarashi K, Samejima M, Ståhlberg J, Horn SJ, Sandgren M. (2011). The putative endoglucanase PcGH61D from Phanerochaete chrysosporium is a metal-dependent oxidative enzyme that cleaves cellulose. PLoS ONE.
- Xia L., P.L. Cen. (1999). Cellulase production by solid state fermentation on lignocellulosic waste from the xylose industry. Process Biochem., 34, pp. 909–912.
- Xiao Z, Zhang X, Gregg DJ, Saddler JN (2004). Effects of sugar inhibition on cellulases and β-glucosidase during enzymatic hydrolysis of softwood substrates. Appl Biochem Biotechnol 113–116:1115–1126.
- Yang S. Q., Q.J. Yan, Z.Q. Jiang, L.T. Li, H.M. Tian, Y.Z. Wang. (2006). High-level of xylanase production by the thermophilic Paecilomyces themophila J18 on wheat straw in solid-state fermentation. Bioresour. Technol., 97, pp. 1794– 1800.
- Zhong, L., Matthews, J.F., Hansen, P.I., Crowley, M.F., Cleary, J.M., Walker, R.C., Nimlos, M.R., Brooks III, C.L., Adney, W.S., Himmel, M.E., Brady, J.W. (2009). Computational simulations of the Trichoderma reesei cellobiohydrolase I acting on microcrystalline cellulose 1beta: the enzymesubstrate complex. Carbohydrate Research. 344(15):1984-1992.