



**UNIVERSITI PUTRA MALAYSIA**

**ANTIBACTERIAL ACTIVITIES OF LACTIC ACID BACTERIA ISOLATED  
FROM ROTTEN VEGETABLES**

**LEE PEI ZHEN**

**FBSB 2015 107**

**ANTIBACTERIAL ACTIVITIES OF LACTIC ACID BACTERIA  
ISOLATED FROM ROTTEN VEGETABLES**

**LEE PEI ZHEN**

**162958**

**DEPARTMENT OF MICROBIOLOGY  
FACULTY BIOTECHNOLOGY AND BIOMOLECULAR SCIENCES  
UNIVERSITI PUTRA MALAYSIA  
2015**

**ANTIBACTERIAL ACTIVITIES OF LACTIC ACID BACTERIA  
ISOLATED FROM ROTTEN VEGETABLES**

**LEE PEI ZHEN**

**162958**

**Dissertation submitted in partial fulfillment of the requirement for the course  
BMY 4999 Project in the Department of Microbiology  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
JUNE 2015**

## PENGESAHAN

Dengan ini adalah disahkan bahawa projek yang bertajuk “**Antibacterial Activities of Lactic Acid Bacteria Isolated from Rotten Vegetables**” telah disiapkan serta dikemukakan kepada Jabatan Mikrobiologi oleh **Lee Pei Zhen (162958)** sebagai syarat untuk kursus BMY 4999 projek.

Disahkan oleh:

Tarikh: .....

Prof Dr. Shuhaimi Mustafa

Penyelia

Jabatan Mikrobiologi

Fakulti Bioteknologi dan Sains Biomolekul

Universiti Putra Malaysia

Tarikh: .....

Prof. Madya Dr. Muhajir Hamid

Ketua

Jabatan Mikrobiologi

Fakulti Bioteknologi dan Sains Biomolekul

Universiti Putra Malaysia

## ABSTRACT

Rising trend of foodborne illness and emergence of antimicrobial resistance in Malaysia pose serious threat to the public and thus a new and effective antibacterial agent is required to address the situation. The non-pathogenic lactic acid bacteria (LAB) with high antibacterial potential can be isolated from rotten vegetables and showed inhibitory activity toward foodborne pathogens. The objective of this study was to isolate and identify LAB, also to determine the inhibitory activity from rotten vegetables against foodborne pathogens. A total of eight LAB isolates were isolated from seven types of rotten vegetables. The antibacterial activities of isolates against *Escherichia coli* ATCC 8739, *Staphylococcus aureus* ATCC 6538, *Listeria monocytogenes* L10 were determined using the agar-well diffusion method. *E. coli* ATCC 8739 appeared to be the most susceptible test organisms to LAB isolates as the inhibition zone of all LAB isolates against *E. coli* ATCC 8739 were obviously larger than the other test organisms. The antibacterial agents were stable within limit range of pH values from pH 2.0 to 5.0. Antibacterial activity was not observed when the pH of the supernatant was adjusted to above pH 6.0. Inhibitory activity of isolates can be observed even at high temperature from 50°C to 100°C, yet the zone of inhibition showed a decrease trend in all isolates as the temperature goes up.

## ABSTRAK

Peningkatan trend penyakit bawaan makanan dan kemunculan rintangan antimikrob di Malaysia menimbulkan ancaman yang serius kepada umum. Justeru, agen antibakteria baru dan efektif diperlukan untuk menangani situasi tersebut. Bakteria asid laktik (LAB) tidak patogenik boleh diisolasi daripada sayur-sayuran busuk dan ia berpotensi merencat aktiviti patogen bawaan makanan. Objektif kajian ini adalah untuk mengasingkan dan mengenalpasti LAB, juga untuk menentukan aktiviti perencatan bakteria terhadap patogen bawaan makanan. Sebanyak lapan isolat LAB telah diasingkan daripada tujuh jenis sayur-sayuran busuk. Aktiviti perencatan isolat LAB terhadap *Escherichia coli* ATCC 8739, *Staphylococcus aureus* ATCC 6538, *Listeria monocytogenes* L10 telah ditentukan dengan menggunakan kaedah peresapan agar-perigi. *E. coli* ATCC 8739 muncul sebagai organisma ujian yang paling mudah terdedah kepada isolat LAB yang diasingkan sebab zon perencatan semua isolat LAB terhadap *E. coli* ATCC 8739 ternyata lebih besar daripada organisma ujian lain. Agen antibakteria stabil dalam julat had nilai pH antara 2.0 sehingga 5.0. Aktiviti antibakteria tidak dapat diperhati apabila pH supernatan telah diselaraskan kepada pH 6.0 dan ke atas. Aktiviti antibakteria masih boleh diperhatikan walaupun telah terdedah kepada suhu yang tinggi dari 50°C hingga 100°C, namun zon perencatan menunjukkan trend penurunan dalam semua isolat terhadap bawaan makanan patogen apabila suhu meningkat.

## ACKNOWLEDGEMENT

I would like to express utmost gratitude to my supervisor, Prof. Dr. Shuhaimi Mustafa for his patient encouragement, valuable advice and support throughout this project. His inspiring guidance helped me a lot in carrying out the research and writing of the thesis.

Also, I would like to show my appreciation to all the staff from Microbiology Department, especially Puan Sharipah and Encik Hussein Ibrahim who assisted me to get the materials and equipment needed.

My thanks extended to the post-graduate seniors especially Nur Afiqah Hunain and Nur Diana Ismail, as well as my lab members, for all their countless hours of guiding, great deal of suggestions and help for completion of my project in proposal preparation, methods reviewing and trouble shooting. I could never finish my project promptly and efficiently without their help.

Finally, I would also like to acknowledge the complete support of my family and friends for being encourage, understanding and supportive from the beginning until the end of the project and the fulfilling of my degree. Thank you, for always being there for me.

## TABLE OF CONTENTS

	<b>Page</b>
<b>PENGESAHAN</b>	<b>i</b>
<b>ABSTRACT</b>	<b>ii</b>
<b>ABSTRAK</b>	<b>iii</b>
<b>ACKNOWLEDGEMENT</b>	<b>iv</b>
<b>LIST OF TABLES</b>	<b>vii</b>
<b>LIST OF FIGURES</b>	<b>viii</b>
<b>LIST OF ABBREVIATIONS</b>	<b>ix</b>
<b>CHAPTER 1 INTRODUCTION</b>	<b>1</b>
1.1 Introduction	1
1.2 Objectives	3
<b>CHAPTER 2 LITERATURE REVIEW</b>	<b>4</b>
2.1 Vegetables and microorganisms	4
2.2 Lactic acid bacteria (LAB)	6
2.2.1 Carbohydrates fermentation pattern of LAB	7
2.3 Antibacterial potential of lactic acid bacteria	10
2.3.1 Bacteriocin	10
2.3.2 Organic acids	13
2.3.3 Hydrogen peroxide	14
2.4 Antagonistic effects of LAB towards foodborne pathogen	14
<b>CHAPTER 3 MATERIALS AND METHODOLOGY</b>	<b>16</b>
3.1 Materials and apparatus	16
3.2 Sample collection	17
3.3 Media and chemicals preparation	17
3.4 Enumeration, isolation and identification method	18
3.4.1 Gram staining	18
3.4.2 Biochemical tests	18

3.4.3	Physical characterization	19
3.5	Maintenance of isolates	19
3.6	Growth curve	19
3.7	Antibacterial activity assays	20
3.7.1	Preparation of test organisms	20
3.7.2	Preparation of LAB cell-free filtrate	20
3.8	Determination of minimum inhibitory dilution and minimum bactericidal dilution	21
3.9	Determination of pH	21
3.10	Effect of pH on antibacterial activities of LAB isolates	21
3.11	Effect of neutralization of LAB isolates on antibacterial activities	22
3.12	Effect of temperature on antibacterial activities of LAB isolates	22
<b>CHAPTER 4</b>	<b>RESULTS AND DISCUSSION</b>	<b>23</b>
4.1	Isolation and identification of LAB	23
4.2	Growth curve	27
4.3	Antibacterial activity assay of LAB isolates	30
4.4	Effect of pH on antibacterial activity of LAB isolates	33
4.5	Effect of neutralization of LAB isolates on antibacterial activities	37
4.6	Effect of temperature on antibacterial activity of LAB isolates	38
<b>CHAPTER 5</b>	<b>CONCLUSION AND RECOMMENDATIONS</b>	<b>41</b>
	<b>REFERENCES</b>	<b>42</b>
	<b>APPENDIX</b>	<b>49</b>

## LIST OF TABLES

Tables	Caption	Page
1	Classification, characteristics and examples of bacteriocins.	12
2	List of materials used in experiment.	16
3	Morphological characteristics of LAB isolates.	26
4	Biochemical test and carbohydrate utilization of LAB isolates.	26
5	Preliminary identification of LAB isolates.	26
6	Means of inhibition zones diameter of LAB isolates against different test microorganisms.	31
7	Minimum inhibitory dilution and minimum bactericidal dilution of LAB isolates against various test organisms.	32
8	Means of inhibition zones diameter of LAB isolates against <i>Listeria monocytogenes</i> L10 under various pH.	34
9	Means of inhibition zones diameter of LAB isolates against <i>Staphylococcus aureus</i> ATCC 6538 under various pH.	35
10	Means of inhibition zones diameter of LAB isolates against <i>Escherichia coli</i> ATCC 8739 under various pH.	36
11	Means of inhibition zones diameter of LAB isolates against different test microorganisms after neutralization.	37
12	Means of inhibition zones diameter of LAB isolates against <i>Listeria monocytogenes</i> L10 under different temperature treatment.	39
13	Means of inhibition zones diameter of LAB isolates against <i>Staphylococcus aureus</i> ATCC 6538 under different temperature treatment.	39
14	Means of inhibition zones diameter of LAB isolates against <i>Escherichia coli</i> ATCC 8739 under different temperature treatment.	40
15	The absorbance readings of LAB isolates for 24 hours at 600 nm	49

## LIST OF FIGURES

Figures	Caption	Page
1	Embden-Meyerhof-Parnas pathway adapted from Fugelsang & Edwards (2007).	8
2	Phosphoketolase pathway adapted from Fugelsang & Edwards (2007).	9
3	Colonies of isolates on MRS agar (left) and its respective gram stain microscopic observation under 1000X magnification.	25
4	Growth curve of all eight LAB isolates.	29
5	Formation of inhibition zone for eight isolates against <i>Escherichia coli</i> ATCC 8739.	30

## LIST OF ABBREVIATIONS

%	Percentage
µl	Microliter
µm	Micrometer
CFF	Cell-free filtrate
g	Gram
h	Hour
HCl	Hydrochloric acid
HPLC	High performance liquid chromatography
LAB	Lactic acid bacteria
M	Molarity
min	Minute
mL	Milliliter
mL/min	Milliliter per minute
mmol/L	Millimole per litre
MRS	de Man, Rogosa and Sharpe
NaOH	Sodium hydroxide
nm	Nanometer
°C	Degree celcius
OD	Optical density
rpm	Rotation per minute
sp	Species

## CHAPTER 1

### INTRODUCTION

#### 1.1 Introduction

Gram-positive, non-spore forming lactic acid bacteria (LAB) is well-known as probiotic strains and has been involved for long history in fermentations and food preservations. One of the significant properties of LAB that contributes to preservative effect is the capability of LAB to produce a variety of antimicrobial compounds through homo-fermentative or hetero-fermentative pathway (Abbott *et al.*, 2009). The noteworthy antimicrobial compound is bacteriocin which is biologically active protein compounds produced in some LAB strains (Reis *et al.*, 2012). Bacteriocin such as nisin has been used as food preservatives commercially. Moreover, organic acids such as lactic acid, acetic acid and propionic acid are responsible for the acidification of environment in which unfavorable for most of the spoilage and pathogenic microorganisms, thus contributes to bio-preservative. In addition, hydrogen peroxide and reuterin are also important metabolites of LAB that exert antimicrobial properties (Yang *et al.*, 2012).

In the recent years, food borne diseases have shown a rising trend in Malaysia primarily due to the rapid population growth, consumption habits and hygienic issues (Sharifa Ezat *et al.*, 2013). A serious threat poses to the public as some of the food borne bacteria showed antibacterial resistance such as the methicilin-resistant *Staphylococcus aureus*. Thus, a new and effective antibacterial agent through a convenient method

should be studied to address the situation. The generally regarded as safe organism (GRAS), LAB is targeted because of its antibacterial potential against common food borne bacteria namely *Escherichia coli*, *Staphylococcus aureus*, *Salmonella typhi*, *Bacillus cereus*, *Shigella* sp., and *Listeria monocytogenes* (Kazemipoor *et al.*, 2012; Senthilkumar *et al.*, 2012; Galvez *et al.*, 2010). Source and culture of LAB from fermented food, dairy product, meat, fresh fruits and vegetables are frequently published (Hajar & Hamid, 2013; Yusra *et al.*, 2013; Yang *et al.*, 2012; Oliveira *et al.*, 2008; Trias *et al.*, 2008). However, the micro-flora on rotten vegetable and the biological activities of LAB isolated from rotten vegetable are not well studied.

Rotten vegetables are often associated with microbial spoilage due to high polysaccharide cellulose, pectin and hemicellulose content. Their neutral pH as well as high water activity makes them an ideal niche for several bacteria (Barth *et al.*, 2009; Doyle, 2007). Some of the normal flora of vegetables such as LAB is one of the contributors in rotting of vegetables (Dris & Jain, 2004). *Leuconostoc mesenteroides* had been found in decayed tomatoes and reported to cause sour rot type decay of tomatoes (Bartz *et al.*, 1995). *Lactobacillus* sp. and *Leuconostoc* sp. also were observed in a complex segment in spoiled segment of fresh cut celery (Robbs *et al.*, 1996). Thus, the possibility to isolate LAB from high availability, abundant and cheap rotten vegetables is relatively high and this makes the rotten vegetables a potential new source and culture for LAB.

## 1.2 Objectives

This study was carried out with the main objectives to investigate the antibacterial activities of LAB from rotten vegetables against food borne pathogens. The specific objectives of present study were:-

- a) To isolate and identify the LAB from rotten vegetables
- b) To characterize the antibacterial activities of the isolated LAB against food borne pathogens

## REFERENCES

- ICMSF, I. C. 2006. *Microorganisms in Foods 6: Microbial Ecology of Food Commodities*. Springer Science & Business Media.
- Abbott, D. A., van den Brink, J., Minneboo, I. M., Pronk, J. T. and van Maris, A. J. 2009. Anaerobic homolactate fermentation with *Saccharomyces cerevisiae* results in depletion of ATP and impaired metabolic activity. *Federation of European Microbiological Societies Yeast Research* 9: 349-357.
- Adams, M. R. and Nicolaidis, L. 1997. Review of the sensitivity of different foodborne pathogens to fermentation. *Food Control* 8 227-239.
- Afolabi, O. R., Bankole, M. O. and Minz, A. 2007. A rapid and simple protocol for extraction and purification of genomic DNA from *Lactobacillus* species isolated from Ogi, a Nigerian fermented food. *Journal of Molecular Cell Biology* 3:121-123.
- Ammor, S., Tauveron, G., Dufour, E. and Chevallier, I. 2006. Antibacterial activity of lactic acid bacteria against spoilage and pathogenic bacteria isolated from the same meat small-scale facility 1—Screening and characterization of the antibacterial compounds. *Food Control* 17 454-461.
- Assefa, E., Beyene, F. and Santhanam, A. 2008. Effect of temperature and pH on the antimicrobial activity of inhibitory substances produced by lactic acid bacteria isolated from Ergo, an Ethiopian traditional fermented milk. *African Journal of Microbiology Research* 2: 229-234.
- Axelsson, L. (1998). Lactic acid bacteria: classification and physiology. In *Lactic Acid Bacteria: Microbiology and Functional Aspects*, eds. S. Salminen, and A. VonWright, pp 1-72. New York. Marcel Dekker.
- Barth, M., Hankinson, T. R., Zhuang, H. and Breidt, F. (2009). Microbiological spoilage of fruits and vegetables. In *Compendium of the Microbiological Spoilage of Foods and Beverages*, eds. W. H. Sperber and M. P. Doyle, pp 135-183. Berlin. Springer Science Business Media.
- Bartz, J. A., Hodge, N. C., Perez, Y. and Concelm, D. 1995. Two different bacteria cause a decay in tomatoes that is similar to sour rot. *Phytopathology* 85: 1123.
- Beshkova, D. and Frengova, G. 2012. Bacteriocins from lactic acid bacteria: Microorganisms of potential biotechnological importance for the dairy industry. *Engineering in Life Sciences* 12: 419-432.
- Blom, H. and Moetvedt, C. 1991. Antimicrobial substances produced by food-associated microorganisms. *Biochemical Society Transactions* 19: 694-698.

- Brashears, M. M., Amezquita, A. and Jaroni, D. (2005). Control of foodborne bacterial pathogens in animals and animals products through microbial antagonism. In *Food Biotechnology*, eds. A. Pometto, K. Shetty, G. Paliyath, and R. E. Levin, pp 1359-1381. Boca Raton. CRC Press.
- Brocklehurst, T. F., Zaman-Wong, C. M. and Lund, B. M. 1987. A note on the microbiology of retail packs of prepared salad vegetables. *Journal of Applied Bacteriology* 63: 409-415.
- Castro, M. P., Palavecino., N. Z., Herman, C., Garro, O. A. and Campos, C. A. 2011. Lactic acid bacteria isolated from artisanal dry sausages: Characterization of antibacterial compounds and study of the factors affecting bacteriocin production. *Meat Science* 87: 321-329.
- Coetzer, E. (2006). Microbiological risk in produce from the field to packing. In *Microbial Hazard Identification in Fresh Fruits and Vegetables*, ed. J. James, pp 73-94. New Jersey. A John Wiley & Sons, Inc.
- Conn, K. E., Ogawa, J. M., Manji, B. T. and Adaskaveg, J. E. 1995. *Leuconostoc mesenteroides* subsp. *mesenteroides*, the first report of a coccoid bacterium causing a plant disease. *Phytopathology* 85: 593-599.
- Dahiya, R. S. and Speck, M. L. 1968. Hydrogen peroxide formation by lactobacilli and its effect on *Staphylococcus aureus*. *Journal of Dairy Science* 51: 1568-1572.
- Doyle, M. E. (2007). *Microbial Food Spoilage—Losses and Control Strategies: A Brief Review of the Literature*.  
[http://fri.wisc.edu/docs/pdf/FRI\\_Brief\\_Microbial\\_Food\\_Spoilage\\_7\\_07.pdf](http://fri.wisc.edu/docs/pdf/FRI_Brief_Microbial_Food_Spoilage_7_07.pdf)  
 [accessed 05 August 2014]
- Dris, R. and Jain, S. M. 2004. *Quality Handling and Evaluation*. Springer Science & Business Media.
- Eklund, T. (1989). Organic acids and esters. In *Mechanisms of Action of Food Preservation Procedures*, ed. G. W. Gould, pp 161-200. New York. Elsevier.
- Eschenbach, D. A., Davick, P. R., Williams, B. L., Klebanoff, S. J., Young-Smith, K., Critchlow, C. M. and Holmes, K. K. 1989. Prevalence of hydrogen peroxide-producing *Lactobacillus* species in normal women and women with bacterial vaginosis. *Journal of Clinical Microbiology* 27: 251.
- Françoisea, L. 2010. Occurrence and role of lactic acid bacteria in seafood products. *Food Microbiology* 27: 698-709.
- Freese, E., Sheu, C. W. and Galliers, E. 1973. Function of lipophilic acids as antimicrobial food additives. *Nature* 241: 321-325.
- Freitas, W. C., Souza, E. L., Sousa, C. P. and Travassos, A. E. 2008. Anti-staphylococcal effectiveness of nisaplin in refrigerated pizza doughs. *Brazilian Archives of Biology and Technology* 51: 595-599.

- Galvez, A., Abriouel, H., Benomar, N. and Lucas, R. 2010. Microbial antagonists to food-borne pathogens and biocontrol. *Current Opinion in Biotechnology* 21: 142–148.
- Gram, L., Ravn, L., Rasch, M., Bruhn, J. B., Christensen, A. B. and Givskov, M. 2002. Food spoilage—interactions between food spoilage bacteria. *International Journal of Food Microbiology* 78: 79-97.
- Girum, T., Eden, E. and Mogese, A. 2010. Assessment of the antimicrobial activity of lactic acid bacteria isolated from borde and shameta, traditional Ethiopian fermented beverage, on some food borne pathogen and affected of growth medium on the inhibitor activities. *International Journal of Food Safety* 5:13-20.
- Goodliffe, J. P. and Heale, J. B. 1975. Incipient infections caused by *Botrytis cinerea* in carrots entering storage. *Annals of Applied Biology* 80: 243-246.
- Hajar, S. and Hamid, T. 2013. Isolation of lactic acid bacteria strain *Staphylococcus piscifermentans* from Malaysian traditional fermented shrimp cinaluk. *International Food Research Journal* 20: 125-129.
- Holzappel, W. H., Haberer, P., Geisen, R., Bjorkroth, J. and Schillinger, U. 2001. Taxonomy and important features of probiotics microorganisms in food and nutrition. *The American Journal of Clinical Nutrition* 73: 365S-373S.
- Kashket, E. R. 1987. Bioenergetics of lactic acid bacteria: cytoplasmic pH and osmotolerance. *FEMS Microbiology Reviews* 46: 233-244.
- Kazemipoor, M., Radzi, C., Begum, K. and Yaze, I. 2012. Screening of antibacterial activity of lactic acid bacteria isolated from fermented vegetables against foodborne pathogens. *Archives Des Sciences* 65: 453-466.
- Khalid, K. 2011. An overview of lactic acid bacteria. *International Journal of Biosciences* 1: 1-13.
- Klaenhammer, T. R. 1988. Bacteriocins of lactic acid bacteria. *Biochimie* 70: 337-349.
- Konig, H. and Frohlich, J. (2009). Lactic acid bacteria. In *Biology of Microorganisms on Grapes in Must and in Wine*, eds. H. Konig, G. Unden, and J. Frohlich, pp 3-30. Berlin. Springer.
- Lahtinen, S., Ouwehand, A. C., Salminen, S. and Wright, A. 2011. *Lactic Acid Bacteria: Microbiological and Functional Aspects*. CRC Press.
- Lamikanra, O. 2002. *Fresh-cut Fruits and Vegetables: Science, Technology and Market*. CRC Press.
- Lewus, C. B., Kaiser A. and Montville T. J. 1991. Inhibition of foodborne bacterial pathogens by bacteriocins from lactic acid bacteria isolated from meat. *Applied and Environmental Microbiology* 57: 1683-1688.
- Liao, C. H. and Wells, J. M. 1989. Diversity of pectolytic, fluorescent Pseudomonas

causing soft rots of fresh vegetables at produce markets. *Phytopathology* 77: 673-677.

- Lopez-Pedemonte, T., Roig-Sagues, A. X., Trujillo, A. J., Capellas, M. and Guamis, B. 2003. Inactivation of spores of *Bacillus cereus* in cheese by high hydrostatic pressure with the addition of nisin or lysozyme. *Journal of Dairy Science* 86: 3075-3081.
- Lorca, G. L. and de Valdez, G. F. (2009). Lactobacillus stress responses. In *Lactobacillus Molecular Biology: From Genomics to Probiotics*, eds. A. Ljungh, and T. Wadstrom, pp 115-138. Norfolk. Caister Academic Press.
- Lund, B. M. (1983). Bacterial spoilage. In *Post-Harvest Pathology of Fruits and Vegetables*, ed. C. Dennis, pp 219-257. London. Academic Press.
- Magnusson, J., Strom, K., Roos, S., Sjogren, J. and Schnurer, J. 2003. Broad and complex antifungal activity among environmental isolates of lactic acid bacteria. *FEMS Microbiology Letters* 219: 129-135.
- Mahon, C. R., Lehman, D. C., and Manuseli, G. 2014. *Textbook of Diagnostic Microbiology*. Elsevier Health Sciences.
- Maragkoudakis, P. A., Mountzouris, K. C., Psyrras, D., Cremonese, S., Fischer, J., Canter, M. D. and Tsakalidou, E. 2009. Functional properties of novel protective lactic acid bacteria and application in raw chicken meat against *Listeria monocytogenes* and *Salmonella enteridis*. *International Journal of Food Microbiology* 130: 219-226.
- Mattu, B. and Chauhan, A. 2013. Lactic acid bacteria and its use in probiotics. *Journal of Bioremediation and Biodegradation* 4: 8.
- Morsi El Soda, Ahmed, N., Omran, N., Osman, G. and Morsi, A. 2003. Isolation, identification and selection of lactic acid bacteria cultures for cheesemaking. *Emirates Journal of Agricultural Science* 15: 51-71.
- Muynck, C. D., Leroy, A. I., Maeseneire, S. D., Arnaut, F., Soetaert, W. and Vandamme, E. J. 2004. Potential of selected lactic acid bacteria to produce food compatible antifungal metabolites. *Microbiological Research* 159: 339-346.
- Noordiana, N., Fatimah, A. B. and Mun, A. S. 2013. Antibacterial agents produced by lactic acid bacteria isolated from threadfin salmon and grass shrimp. *International Food Research Journal* 20:117-124.
- Oliveira, R. B., Oliveira, A. L. and Gloria, M. B. 2008. Screening of lactic acid bacteria from vacuum packaged beef for antimicrobial activity. *Brazilian Journal of Microbiology* 39: 368-374.
- Orla-Jensen, S. 1919. *The Lactic Acid Bacteria*. Andre Fred Host & Son.

- Padaga, M., Heard, G. M., Paton, J. E. and Fleet, G. H. 2000. Microbial species associated with different sections of broccoli harvested from three regions in Australia. *International Journal of Food Microbiology* 60: 15-24.
- Parada, J. L., Caron, C. R., Medeiros, A. B. and Soccol, C. R. 2007. Bacteriocins from lactic acid bacteria: purification, properties and use as biopreservatives. *Brazilian Archives of Biology and Technology* 50: 521-542.
- Pederson, C. S. and Breed, R. S. (1929). Control of spoilage in tomato products. <https://dspace.library.cornell.edu/bitstream/1813/4662/1/bulletin570.pdf> [accessed 08 August 2014]
- Pendergrass, A. and Isenberg, F. 1974. The effect of relative humidity on the quality of stored cabbage. *Horticultural Science* 9: 226-227.
- Podolak, P. K., Zayas, J. F., Kastner, C. L. and Fung, D. Y. 1996. Inhibition of *Listeria monocytogenes* and *Escherichia coli* O157:H7 on beef by application of organic acids. *Journal of Food Protection* 59: 370-373.
- Pohronezny, K., Larsen, P. O. and Leben, P. O. 1978. Observations on cucumber fruit invasion by *Pseudomonas lachrymans*. *Plant Disease Reporter Supplements* 62: 306-309.
- Ponde, A. G., Moreira, M. R., Valle, C. E. and Roura, S. I. 2008. Preliminary characterization of bacteriocin-like substances from lactic acid bacteria isolated from organic leafy vegetables. *LWT—Food Science and Technology* 41: 432-441
- Pouwels, P. H., Leer, R. J., Shaw, M., Heijne den Bak-Glashouwer, M. J., Tielen, F. D., Smit, E., Martinez, B., Jore, J. and Conway, P. L. 1998. Lactic acid bacteria as antigen delivery vehicles for oral immunization purposes. *International Journal of Food Microbiology* 41: 155-167.
- Rattanachaikunsopon, P. and Phumkhachorn, P. 2010. Lactic acid bacteria: their antimicrobial compounds and their uses in food production. *Annals of Biological Research* 1: 218-228.
- Ray, B. 2003. *Fundamental Food Microbiology*. CRC Press.
- Reis, J. A., Paula, A. T., Casarotti, S. N. and Penna, A. L. 2012. Lactic acid bacteria antimicrobial compounds: characteristics and applications. *Food Engineering Reviews* 4: 124-140.
- Robbs, P. G., Bartz, J. A., McFie, G. and Hodge, N. C. 1996. Causes of decay of fresh-cut celery. *Journal of Food Science* 61: 444-448.
- Robert, C. M. and Hoover, D. G. 1996. Sensitivity of *Bacillus coagulans* spores to combinations of high hydrostatic pressure, heat, acidity and nisni. *Journal of Applied Bacteriology* 81: 363-368.

- Schillinger, U., Becker, B., Vignolo G. and Holzappel, W. H. 2001. Efficacy of nisin in combination with protective cultures against *Listeria monocytogenes* Scott A in tofu. *International Journal of Food Microbiology* 71: 159-168.
- Senthilkumar, P. K., Reetha, D., Ramya, N. and Stella, D. 2012. Antibacterial potential of lactic acid bacteria and its metabolites against foodborne pathogens. *International Journal of Pharmaceutical and Biological Archives* 3: 342-347.
- Sharifa Ezat, W. P., Netty, D. and Sangaran, G. 2013. Paper review of factors, surveillance and burden of foodborne disease outbreak in Malaysia. *Malaysian Journal of Public Health Medicine* 13: 98-105.
- Snijders, J. M., van Logtestijn, J. G., Mossel, D. A. and Smulders, F. J. 1985. Lactic acid as a decontaminant in slaughter and processing procedures. *Veterinary Quarterly* 7: 277-282.
- Staszewski, M. and Jagus, R. J. 2008. Natural antimicrobials: effect of microgard and nisin against *Listeria innocua* in liquid cheese whey. *International Dairy Journal* 18: 255-259.
- Suganthi, V., Selvarajan, E., Subathradevi, C. and Mohanasrinivasan, V. 2012. Lantibiotic nisin: natural preservative from *Lactococcus lactis*. *International Research Journal of Pharmacy* 3: 13-19.
- Šušković, J., Kos, B., Beganović, J., Pavunc, A. L., Habjanić, K. and Matošić, S. 2010. Antimicrobial activity: the most important property of probiotic and starter lactic acid bacteria. *Food Technology and Biotechnology* 48: 296-307.
- Tagg, J. R., Dajani, A. S. and Wannamaker, L. W. 1976. Bacteriocins of gram-positive bacteria. *Bacteriology Reviews* 40: 722.
- Thirabunyanon, M., Boonprason, P. and Niamsup, P. 2009. Probiotic potential of lactic acid bacterial isolated from fermented dairy milks on anti-proliferation of colon cancer cells. *Biotechnology Letters* 31: 571-576.
- Trias, R., Bañeras, L., Montesinos, E. and Badosa, E. 2008. Lactic acid bacteria from fresh fruit and vegetables as biocontrol agents of phytopathogenic bacteria and fungi. *International Microbiology* 11: 231-236.
- Valgas, C., Souza, S. M., Smania, E. F. and Smânia Jr., A. 2007. Screening methods to determine antibacterial activity of natural products. *Brazilian Journal of Microbiology* 38: 369-380.
- Vaughan, A., Eijsink, V. G. and VanSinderen, D. 2003. Functional characterization of a composite bacteriocin locus from malt isolate *Lactobacillus sakei* 5. *Applied and Environmental Microbiology* 69: 7194-7203.
- Vinderola, C. G. and Reinheimer, J. A. 2002. Interactions among lactic acid starter and probiotic bacteria used for fermented dairy products. *Journal of Dairy Science* 85: 721-729.

- Vollenweider, S., Grassi, G., Konig, I. and Puhon, Z. 2003. Purification and structural characterization of 3-hydroxypropionaldehyde and its derivatives. *Journal of Agricultural and Food Chemistry* 51: 3287-3293.
- Vuyst, L. D. and Leroy, F. 2007. Bacteriocins from lactic acid bacteria: production, purification, and food applications. *Journal of Molecular Microbiology and Biotechnology* 13: 194-199.
- Wheater, D. M., Hirsch, A. and Mattick, A. T. 1952. Possible identity of 'lactobacillin' with hydrogen peroxide produced by lactobacilli. *Nature* 170: 623-624.
- Wiles, A. B. and Walker, J. C. 1951. The relation of *Pseudomonas lachrymans* to cucumber fruits and seeds. *Phytopathology* 41: 1059-64.
- Wu, C. M., Koehler, P. E. and Ayres, J. C. 1972. Isolation and identification of xanthotoxin (8-methoxypsoralen) and bergapten (5-methoxypsoralen) from celery infected with *Sclerotinia sclerotiorum*. *Journal of Applied Microbiology* 23: 852-856.
- Yang, E., Fan, L., Jiang, Y., Doucette, C. and Fillmore, S. 2012. Antimicrobial activity of bacteriocin-producing lactic acid bacteria isolated from cheeses and yogurts. *AMB Express* 2: 1-12.
- Yang, J., Cao, Y., Cai, Y. and Terada, F. 2010. Natural populations of lactic acid bacteria isolated from vegetable residues and silage fermentation. *Journal of Dairy Science* 93: 3136-3145.
- Yelnetty, A., Purnomo, H., Purwadi and Mirah, A. 2014. Biochemical characteristics of lactic acid bacteria with proteolytic activity and capability as starter culture isolated from spontaneous fermented local goat milk. *Journal of Natural Sciences Research* 4: 137-146.
- York, G. K., Heil, J. R., Marsh, G. L., Ansar, A., Merson, R. L., Wolcott, T. and Leonard, S. 1975. Thermobacteriology of canned, whole, peeled tomatoes. *Journal of Food Science* 40: 764-769.
- Yusra, Azima, F., Novelina and Periadnadi. 2013. Antimicrobial activity of lactic acid bacteria isolated from budu of west sumatera to food biopreservatives. *Pakistan Journal of Nutrition* 12: 628-635.