



UNIVERSITI PUTRA MALAYSIA

CYTOTOXIC EFFECTS OF METHANOLIC EXTRACTS OF *Psidium guajava* AND *Morinda citrifolia* ON HUMAN OSTEOSARCOMA SAOS-2 CELLS

CHOO JIE YING

FBSB 2015 99

PENGESAHAN

Dengan ini adalah disahkan bahawa projek yang bertajuk “Cytotoxic effects of methanolic extracts of *Psidium guajava* and *Morinda citrifolia* on human osteosarcoma Saos-2 cells” telah disiapkan serta dikemukakan kepada Jabatan Mikrobiologi oleh Choo Jie Ying (165117) sebagai syarat untuk kursus BMY 4999 projek.

Disahkan oleh:

.....

Tarikh:

Prof. Madya Dr Norazizah Shafee

Penyelia

Jabatan Mikrobiologi

Fakulti Bioteknologi dan Sains Biomolekul

Universiti Putra Malaysia

.....

Tarikh:

Prof. Madya Dr. Muhajir Hamid

Ketua

Jabatan Mikrobiologi

Fakulti Bioteknologi dan Sains Biomolekul

Universiti Putra Malaysia

ABSTRACT

Cancer is a major health problem and causes significant morbidity and mortality. Bone cancer is an uncommon cancer that begins in a bone and it mostly occurs in children and teens. Cancer treatment by conventional medicine, surgery, chemotherapy and radiotherapy have been the primary approaches, but they are not always effective. The use of complementary alternative medicine (CAM) has grown rapidly in popularity both among the general population and among cancer patients. Therefore, we hypothesize that plant extracts exert antiproliferative effect on cancer cells. The objective of this study is to evaluate the antiproliferation effects on cancer cells using selected plant extracts. Results showed that *Psidium guajava* extracts showed cytotoxic effect on human Saos-2 cells, however, *Morinda citrifolia* extracts do not exhibit any inhibition on Saos-2 cells. *P.guajava* leaves have more cytotoxic effects than *M.citrifolia* leaves. Further study of *P. guajava* can be applied to determine the optimal concentration of a leaf extract for cancer treatment, while, *M. citrifolia* can be recommended for further study on wound healing effects.

ABSTRAK

Kanser merupakan masalah kesihatan utama dan punca utama morbiditi dan mortaliti. Kanser tulang adalah barah yang luar biasa yang bermula pada tulang dan ia kebanyakannya berlaku pada kanak-kanak dan remaja. Rawatan kanser secara perubatan konvensional seperti pembedahan, kemoterapi dan radioterapi telah menjadi pilihan utama, tetapi tidak sentiasa berkesan. Penggunaan perubatan sampingan dan alternative berkembang dengan pesat dalam populariti kedua-dua di antara penduduk umum dan di kalangan pesakit-pesakit kanser. Oleh itu, kami hipotesis bahawa ekstrak tumbuhan mengenakan antiproliferatif kesan ke atas sel-sel kanser. Objektif kajian ini adalah untuk menilai kesan antiproliferasi ke atas sel-sel kanser oleh tumbuh-tumbuhan penyelidikan. Hasil kajian menunjukkan bahawa *Psidium guajava* ekstrak menunjukkan kesan sitotoksik ke atas sel manusia, Saos-2, Walau bagaimanapun, *Morinda citrifolia* ekstrak tidak mempamerkan apa-apa kesan pada sel-sel Saos-2. Daun *P.guajava* mempunyai lebih banyak kesan sitotoksik daripada daun *M.citrifolia*. Penyelidikan lanjutann daripada *P.guajava* boleh dibuat untuk menentukan kepekatan optimum ekstrak daun untuk rawatan kanser, sementara *M.citrifolia* boleh mengesyorkan kepada satu kajian tentang kesan-kesan penyembuhan luka.

ACKNOWLEDGEMENT

First and foremost, I would like to take this opportunity to thank my project supervisor Assoc. Prof. Dr. Norazizah Shafee for her support and advice through this project.

Besides my project supervisor, I would like to thank my lab senior, Liew Sien Yei for all the guidance and advices for my project. Also, I would like to thank all the members from lab 143 for their kindly assistances along my project.

Furthermore, I would like to thank my friends who involved in this project. I would like to apologize for all the mistakes that I did during my project.

Last but not least, I would like to thank my family members that always support me during the progress of this study. Although there are a lot of obstacles during this project, I was able to handle it with the support from my family.

TABLE OF CONTENTS

PENGESAHAN	i
ABSTRACT	ii
ABSTRAK	iii
ACKNOWLEDGEMENT	iv
LIST OF FIGURES	vii
LIST OF ABBREVIATIONS	viii
CHAPTER 1	1
INTRODUCTION	1
CHAPTER 2	3
LITERATURE REVIEW	3
2.1 Cancer	3
2.1.1 Bone cancer cell lines	4
2.2 Treatment methods	4
2.2.1 Surgery	4
2.3 Chemotherapy and radiotherapy	5
2.4 Complementary and alternative medicines (CAM)	5
2.5 Natural products	6
2.5.1 Plant	6
2.5.2 Research plants used in the present study	7
2.6 Cell viability assays	8
2.6.1 3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assay	8
2.6.2 Lactate dehydrogenase (LDH) assay	9
2.6.3 Sulforhadamine B (SRB) assay	9
2.6.4 Clonogenic assay	9
CHAPTER 3	10
MATERIALS AND METHODS	10
3.1 Chemical and reagents	10
3.2 Cell lines used in the study	10
3.3 Cell culture	10
3.3.1 Cell maintenance	10
3.3.2 Cryopreservation of cells	11
3.3.3 Cell counting	11
3.3.4 MTT assay	12
3.4 Optimization of cell densities	13
3.5 Optimization on diluent, DMSO concentration	13
3.6 Preparation of plant extract	13
3.6.1 Plant collection	13
3.6.2 Plant extraction: Methanol extraction	14
3.6.3 Rotary evaporation	14
3.6.4 Drying of plant extract essence	14
3.7 Plant extract treatment	15
3.8 Statistical analysis	15

CHAPTER 4	16
RESULTS AND DISCUSSION	16
4.1 Cell lines	16
4.2 Optimization of cell density	18
4.3 Optimization of diluent, DMSO concentration	20
4.4 Preparation of plant extracts and determination of yield	23
4.5 Cytotoxic effect of plant extracts on human Saos-2 cells	24
4.5.1 <i>P.guajava</i>	24
4.5.2 <i>M.citrifolia</i>	29
CHAPTER 5	33
CONCLUSION AND RECOMMENDATIONS	33
REFERENCES	34
APPENDIX A	40



LIST OF FIGURES

Figure		Page
1	Morphology of the cells at 100x magnification.	17
2	Optimization of cell densities at 24, 48, 72 and 96 hours.	19
3	Optimization of diluent concentration of human Chang liver cells.	21
4	Optimization of diluent concentration of human Saos-2 cells.	22
5	Selected concentrations of <i>P.guajava</i> extract affect the viability of cells at 24, 48 and 72 hours.	26
6	Morphological changes of human Saos-2 cells at 100x magnification.	27
7	Morphological changes of human Chang liver cells at 100x magnification.	28
8	Selected concentrations of <i>M.citrifolia</i> extract affect the viability of cells at 24, 48 and 72 hours.	30
9	Morphological change of human Saos-2 cells at 100x magnification.	31
10	Morphological change of human Chang liver cells at 100x magnification.	33

LIST OF ABBREVIATIONS

ATCC	American Type Culture Collection
CAM	Complementary alternative medicine
DMEM	Dulbecco's Modified Eagle's Medium
DMSO	Dimethyl Sulfoxide
FBS	Fetal bovine serum
HSC	Hematopoietic stem cell
LLC	Lewis lung carcinoma
MTT	3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide
PBS	Phosphate Buffered Saline
OS	Osteosarcomas
<i>x g</i>	Times gravity

CHAPTER 1

INTRODUCTION

Today, cancer is still one of the key global healthcare challenges claiming millions of lives every year (Arome and Amarachi, 2014). Extensive research has progressed to achieve better understanding of the molecular basis of cancer over the past 30 years (Kundap and Sawade, 2013). Cancer is a condition where cells grow in a specific part of the body and reproduce uncontrollably. The cancerous cells can invade and destroy surrounding healthy tissue, including organs (Saxena et al., 2014). Metastasis is a process which spread and growth of tumor cells at distant organs (Kang et al., 2003). Bone cancer occurs in patients with primary bone tumors and is the most common site of the metastasis in patients with breast cancer, affecting up to 90% of women with advanced disease (Luger et al., 2001 and Body et al., 2003).

Cancer treatment by conventional medicine, surgery, chemotherapy and radiotherapy have been the primary approaches, but they are not always effective (Hsiao and Liu, 2010). Today, the increasing interest in complementary alternative medicines (CAM) among cancer patients may be due to limitations of conventional cancer treatment, increased advertising and media coverage of CAM, or the desire for holistic or natural treatments (Richardson et al., 2000). Medicines which are derived from plants have contributed in human health and well beings (Iwu et al., 1999), on the other hand, herbs were already used as a medicine since ancient times (Calixto, 2000). These medicinal plants can produce a definite physiological action on human body due to some chemical substance in the plants (Edeoga, 2005). Plants act as

reservoirs for a wide variety of secondary metabolites including alkaloids, flavonoids, tannins and terpenoids, which possess therapeutic properties (Khan et al., 2015). Several of herbs have shown antiplatelet, antitumor, hypolipidemic, or immune-stimulating properties that may add on useful in helping reduce the risk of heart disease and cancer (Craig, 1999).

The prevalence of cancer is on a steady increase every year, hence, new therapeutic strategy specifically targeting these cancer cells are needed. We hypothesize that the methanolic plant extracts of *Psidium guajava* and *Morinda citrifolia* can show the cytotoxic effect towards the bone cancer cell line. The main objective of this study is to evaluate the cytotoxic effect of the methanolic plant extracts, *Psidium guajava* and *Morinda citrifolia* on the cancer cells.

To achieve this experiment, three specific aims were planned. They are:

1. To prepare the crude methanolic extract of *P. guajava* and *M. citrifolia*.
2. To optimize the cell density and DMSO concentration.
3. To evaluate the cytotoxic effect of *P. guajava* and *M. citrifolia* toward Saos-2 cell lines.

REFERENCES

- Abate, G., Mshana, R. N., & Miörner, H. (1998). Evaluation of a colorimetric assay based on 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) for rapid detection of rifampicin resistance in *Mycobacterium tuberculosis*. *The International Journal of Tuberculosis and Lung Disease*, 2(12), 1011-1016.
- Al-Hajj, M., Becker, M. W., Wicha, M., Weissman, I., & Clarke, M. F. (2004). Therapeutic implications of cancer stem cells. *Current Opinion in Genetics & Development*, 14(1), 43-47.
- Arnon, J., Meirou, D., Lewis-Roness, H., & Ornoy, A. (2001). Genetic and teratogenic effects of cancer treatments on gametes and embryos. *Human Reproduction Update*, 7(4), 394-403.
- Arome, D., & Amarachi, A. (2014). A Review on Herbal Plants with Anti-Tumour Properties. *Journal of Pharmaceutical, Chemical and Biological Sciences*, 2(2), 43-58.
- Ashikaga, T., Bosompra, K., O'Brien, P., & Nelson, L. (2002). Use of complimentary and alternative medicine by breast cancer patients: prevalence, patterns and communication with physicians. *Supportive Care in Cancer*, 10(7), 542-548.
- Baba, C., Yanagida, K., Kanzaki, T., & Baba, M. (2005). Colorimetric lactate dehydrogenase (LDH) assay for evaluation of antiviral activity against bovine viral diarrhoea virus (BVDV) in vitro. *International Medical Press*, 16(1), 33-39.
- Bertaux, K., Broux, O., Chauveau, C., Jeanfils, J., & Devedjian, J. (2005). Identification of CBFA1-regulated genes on SaOs-2 cells. *Journal of Bone and Mineral Metabolism*, 23(2), 114-122.
- Bhati, W., & Vishwa, A. (2013). Nanotechnology Method Comparison for Early Detection of Cancer. *I.J.Intelligent Systems and Applications*, 5(3), 58-65.
- Body, J., Diel, I. J., Lichinitser, M. R., Kreuser, E. D., Gorbunova, V. A., Buddle, M., & Bergström, B. (2003). Intravenous ibandronate reduces the incidence of skeletal complications in patients with breast cancer and bone metastases. *Annals of Oncology*, 14(9), 1399-1405.
- Bopp, S. K., & Lettieri, T. (2008). Comparison of four different colorimetric and fluorometric cytotoxicity assays in a zebrafish liver cell line. *BMC Pharmacology*, 8(1), 8.

- Burke, R. E., Harris, S. C., & McGuire, W. L. (1978). Lactate Dehydrogenase in Estrogen-responsive Human Breast Cancer Cells. *Cancer Research*, 38(9), 2773-2776.
- Calixto, J. B. (2000). Efficacy, safety, quality control, marketing and regulatory guidelines for herbal medicines (phytotherapeutic agents). *Brazilian Journal of Medical and Biological Research*, 33(2), 179-189.
- Campling, B., Pym, J., Baker, H., Cole, S., & Lam, Y. (1991). Chemosensitivity testing of small cell lung cancer using the MTT assay. *British Journal of Cancer*, 63(1), 75-83.
- Chan-Blanco, Y., Vaillant, F., Perez, A. M., Reynes, M., Brillouet, J., & Brat, P. (2006). The noni fruit (*Morinda citrifolia* L.): A review of agricultural research, nutritional and therapeutic properties. *Journal of Food Composition and Analysis*, 19(6), 645-654.
- Craig, W. J. (1999). Health-promoting properties of common herbs. *American Society for Clinical Nutrition*, 70(3), 491S-499S.
- Cui, Y., Shu, X., Gao, Y., Wen, W., Ruan, Z., Jin, F., & Zheng, W. (2004). Use of complementary and alternative medicine by Chinese women with breast cancer. *Breast Cancer Research and Treatment*, 85(3), 263-270.
- Dai, J., & Mumper, R. J. (2010). Plant Phenolics: Extraction, Analysis and Their Antioxidant and Anticancer Properties. *Molecules*, 15(10), 7313-7352.
- De Graaf, C. A., & Metcalf, D. (2011). Thrombopoietin and hematopoietic stem cells. *Cell Cycle*, 10(10), 1582-1589.
- Edeoga, H. O., Okwu, D. E., & Mbaebie, B. O. (2005). Phytochemical constituents of some Nigerian medicinal plants. *African Journal of Biotechnology*, 4(7), 685-688.
- Fathilah, A. R., Sujata, R., Norhanom, A. W., & Adenan, M. I. (2010). Antiproliferative activity of aqueous extract of *Piper betle* L. and *Psidium guajava* L. on KB and HeLa cell lines. *Journal of Medicinal Plants Research*, 4(11), 987-990.
- Fedr, R., Pernicova, Z., Slabakova, E., Strakova, N., Bouchal, J., Grepl, M., . . . Soucek, K. (2013). Automatic Cell Cloning Assay for Determining the Clonogenic Capacity of Cancer and Cancer Stem-Like Cells. *Cytometry Part A*, 83(5), 472-482.
- Fotakis, G., & Timbrell, J. A. (2006). In vitro cytotoxicity assays: Comparison of LDH, neutral red, MTT and protein assay in hepatoma cell lines following exposure to cadmium chloride. *Toxicology Letters*, 160(2), 171-177.
- FREIMOSER, F. M., JAKOB, C. A., AEBI, M., & TUOR, U. (1999). The MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide) Assay Is a Fast

and Reliable Method for Colorimetric Determination of Fungal Cell Densities. *Applied and Environmental Microbiology*, 65(8), 3727-3729.

- Gangai Abirami, S. K., & Nirmala, P. (2014). A comparative ? invitro study of anticancer effect of mentha piperita, ocimum basilicum and coleus aromaticus against human laryngeal epidermoid carcinoma (HEP-2) cell lines. *Journal of Medicinal Plants Studies*, 2(1), 6-9.
- Ganie, S. A., Dar, T. A., Hamid, R., Zargar, O., Abeer, S. U., Masood, A., & Zargar, M. A. (2014). In Vitro Antioxidant and Cytotoxic Activities of *Arnebia benthamii* (Wall ex. G. Don): A Critically Endangered Medicinal Plant of Kashmir Valley. *Oxidative Medicine and Cellular Longevity*, 2014, 8.
- Gordaliza, M. (2007). Natural products as leads to anticancer drugs. *Clinical & Translational Oncology*, 9(12), 767-776.
- Gutiérrez, R. M., Mitchell, S., & Solis, R. V. (2008). *Psidium guajava*: A review of its traditional uses, phytochemistry and pharmacology. *Journal of Ethnopharmacology*, 117(1), 1-27.
- Hoang, B. H., Kubo, T., Healey, J. H., Yang, R., Nathan, S. S., Kolb, E. A., . . . Gorlick, R. (2004). Dickkopf 3 Inhibits Invasion and Motility of Saos-2 Osteosarcoma Cells by Modulating the Wnt--Catenin Pathway. *Cancer Research*, 64(8), 2734-2739.
- Hsiao, W., & Liu, L. (2010). The Role of Traditional Chinese Herbal Medicines in Cancer Therapy ? from TCM Theory to Mechanistic Insights. *Planta Medica*, 76(11), 1118-1131.
- Huo, L., Liu, K., Pei, J., Yang, Y., Ye, Y., Liu, Y., . . . Guo, Y. (2013). Flouride Promotes Viability and Differentiation of Osteoblast-Like Saos-2 Cells Via BMP/Smads Signaling Pathway. *Biological trace element research*, 155(1), 142-149.
- Iwu, M. M., Duncan, A. R., & Okunji, C. O. (1999). New Antimicrobials of Plant Origin. *Perspectives on new crops and new uses*, 457-462.
- Jones, D. H., Nakashima, T., Sanchez, O. H., Kozieradzki, I., Komarova, S. V., Sarosi, I., . . . Penninger, J. M. (2006). Regulation of cancer cell migration and bone metastasis by RANKL. *Nature*, 440(7084), 692-696.
- K. Rates, S. M. (2001). Plants as source of drugs. *Toxicon*, 39(5), 603-613.
- Kang, Y., Siegel, P. M., Shu, W., Drobnjak, M., Kakonen, S. M., Córdón-Cardo, C., . . . Massagué;J. (2003). A multigenic program mediating breast cancer metastasis to bone. *Cancer Cell*, 3(6), 537-549.
- Kaushik, N. K., Kim, Y. H., Han, Y. G., & Choi, E. H. (2013). Effect of jet plasma on T98G human brain cancer cells. *Current Applied Physics*, 13(1), 176-180.

- Kennedy, J. E. (2005). High-intensity focused ultrasound in the treatment of solid tumours. *Nature Reviews Cancer*, 5(4), 321-327.
- Khan, U. A., Rahman, H., Qasim, M., Hussain, A., Azizllah, A., Murad, W., . . . Adnan, M. (2015). Alkanna tinctoria leaves extracts: a prospective remedy against multidrug resistant human pathogenic bacteria. *BMC Complementary and Alternative Medicine*, 15(1), 127.
- Kundap, U. P., & Sarawade, R. (2013). Recent Potent Molecular Targets for Cancer Treatment - A Review. *International Journal of Pharmaceutical & Biological Archives 2013*, 4(5), 800-818.
- Levy, A. S., & Carley, S. (2012). Cytotoxic Activity of Hexane Extracts of Psidium Guajava L (Myrtaceae) and Cassia Alata L (Caesalpineaceae) in Kasumi-1 and OV2008 Cancer Cell Lines. *Tropical Journal of Pharmaceutical Research*, 11(2), 201-207.
- Luger, N. M., Honore, P., Ann, M., Sabino, C., Schwei, M. J., Rogers, S. D., . . . Mantyh, P. W. (2001). Osteoproteger Diminishes Advanced Bone Cancer Pain. *Cancer Research*, 61(10), 4038-4047.
- McClatchey, W. (2002). From Polynesian Healers to Health Food Stores: Changing Perspectives of Morinda citrifolia(Rubiaceae). *Integrative Cancer Therapies*, 1(2), 110-120.
- Morel, A., Lièvre, M., Thomas, C., Hinkal, G., Ansieau, S., & Puisieux, A. (2008). Generation of Breast Cancer Stem Cells through Epithelial-Mesenchymal Transition. *PLOS One*, 3(8), e2888.
- Nayak, B. S., Sandiford, S., & Maxwell, A. (2009). Evaluation of the Wound-Healing Activity of Ethanolic Extract of Morinda citrifolia L. Leaf. *Evidence-based Complementary and Alternative Medicine*, 6(3), 351-356.
- Ncube, N. S., Afolayan, A. J., & Okoh, A. I. (2008). Assessment techniques of antimicrobial properties of natural compounds of plant origin: current methods and future trends. *African Journal of Biotechnology*, 7(12), 1797-1806.
- Nikzad, S., & Hashemi, B. (2014). MTT assay instead of the clonogenic assay in measuring the response of cells to ionizing radiation. *Journal of Radiobiology*, 1(1), 3-8.
- Nostro, A., Germano, M. P., D'Angelo, V., Marino, A., & Cannatelli, M. A. (2000). Extraction methods and bioautography for evaluation of medicinal plant antimicrobial activity. *Letters in Applied Microbiology*, 30(5), 379-384.
- Odey, M. O., Iwara, I. A., Udiba, U. U., Inetwe, U. V., Asenye, M. E., & Victor, O. (2012). Preparation of plant extracts from indigenous medicinal plants. *Int.J.Sci.Technol*, 1(12), 688-922.

- Pal, R., Mamidi, M. K., Das, A. K., & Bhonde, R. (2011). Diverse effects of dimethyl sulfoxide (DMSO) on the differentiation potential of human embryonic stem cells. *Archives of toxicology*, 86(4), 651-661.
- Park, D., Sykes, D. B., & Scadden, D. T. (2012). The hematopoietic stem cell niche. *Frontiers in bioscience (Landmark edition)*, 17, 30.
- Pautke, C., Schieker, M., Tischer, T., Kolk, A., Neth, P., Mutschler, W., & Milz, S. (2004). Characterization of Osteosarcoma Cell Lines MG-63, Saos-2 and U-2 OS in Comparison to Human Osteoblasts. *Anticancer Research*, 24(6), 3734-3748.
- Pieters, R., Loonen, A. H., Huismans, D. R., Broekema, G., J. Dirven, M. W., Heyenbrok, M. W., . . . P. Veerman, A. J. (1990). In Vitro Drug Sensitivity of Cells From Children With Leukemia Using the MTT Assay With Improved Culture Conditions. *Blood*, 76(11), 2327-2336.
- Prasad, V. G., Kawade, S., Jayashree, B. S., Reddy, N. D., Francis, A., Nayak, P. G., . . . Shenoy, R. R. (2014). Iminoflavones Combat 1,2-Dimethyl Hydrazine-Induced Aberrant Crypt Foci Development in Colon Cancer. *BioMed Research International*, 2014, 7.
- Qi, W., Ding, D., & Salvi, R. J. (2008). Cytotoxic effects of dimethyl sulphoxide (DMSO) on cochlear organotypic cultures. *Hearing Research*, 236(1-2), 52-60.
- Richardson, M. A., Mde, L. C., Nanny, K., & Sanders, C. (2004). Discrepant views of oncologists and cancer patients on complementary/alternative medicine. *Supportive Care in Cancer*, 12(11), 797-804.
- Richardson, M. A., Sanders, T., Palmer, J. L., Greisinger, A., & Singletary, S. E. (2000). Complementary/Alternative Medicine Use in a Comprehensive Cancer Center and the Implications for Oncology. *Journal of Clinical Oncology*, 18(13), 2505-2514.
- Roa, W., Yang, X., Guo, L., Huang, B., Khatibisepehr, S., Gabos, S., . . . Xing, J. (2011). Real-time cell-impedance sensing assay as an alternative to clonogenic assay in evaluating cancer radiotherapy. *Analytical and Bioanalytical Chemistry*, 400(7), 2003-2011.
- Saini, R. K., Chouhan, R., Bagri, L. P., & Bajpai, A. K. (2012). Strategies of Targeting Tumors and Cancers. *Journal of Cancer Research Updates*, 1(1).
- Saleem, A., Husheem, M., Härkönen, P., & Pihlaja, K. (2002). Inhibition of cancer cell growth by crude extract and the phenolics of Terminalia chebula retz. fruit. *Journal of Ethnopharmacology*, 81(3), 327-336.
- Sargent, J., & Taylor, C. (1989). Appraisal of the MTT assay as a rapid test of chemosensitivity in acute myeloid leukaemia. *British Journal of Cancer*, 60(2), 206-210.

- Saxena, V. L., Chaturvedi, P., & Agrahari, N. (2014). Microarray Based Pharmacogenomics Study Of Celecoxib On Breast Cancer Samples. *Journal of Pharmacy and Biological Sciences*, 9(1), 68-71.
- Senaratne, S. G., Pirianov, G., Mansi, J. L., Arnett, T. R., & Colston, K. W. (2000). Bisphosphonates induce apoptosis in human breast cancer cell lines. *British Journal of Cancer*, 82(8), 1459-1468.
- Simmonds, P. C., Primrose, J. N., Colquitt, J. L., Garden, O. J., Poston, G. J., & Rees, M. (2006). Surgical resection of hepatic metastases from colorectal cancer: A systematic review of published studies. *British Journal of Cancer*, 94(7), 982-999.
- Smith, C. (2003). Hematopoietic Stem Cells and Hematopoiesis. *Cancer Control*, 10(1), 9-16.
- Sohafy, S. E., Metwally, A., Omar, A., & Harraz, F. (2010). Phytochemical investigation and antimicrobial activity of *Psidium guajava* L. leaves. *Pharmacognosy Magazine*, 6(23), 212-218.
- Sultana, B., Anwar, F., & Ashraf, M. (2009). Effect of Extraction Solvent/Technique on the Antioxidant Activity of Selected Medicinal Plant Extracts. *Molecules*, 14(6), 2167-2180.
- Thani, W., Luanratana, O., Siripong, P., & Ruangwises, N. (2008). Anti-proliferative activity of Thai Noni (*Morinda citrifolia* linn.) leaf extract. *Planta Medica*, 41(2), 482-489.
- Twentyman, P., & Luscombe, M. (1987). A study of some variables in a tetrazolium dye (MTT) based assay for cell growth and chemosensitivity. *British Journal of Cancer*, 56(3), 279-285.
- Wang, C. R., Hou, D. Y., Feng, H. G., Yang, B. S., Xu, C. S., & Lin, J. T. (2010). Induction of new adam related protein from treated human Chang-liver cells. *Molecular Biology*, 44(5), 748-753.
- Wang, M. Y., & Su, C. (2001). Cancer Preventive Effect of *Morinda citrifolia* (Noni). *Annals of The New York Academy of Sciences*, 952(1), 161-168.
- Woolston, C., & Martin, S. (2011). Analysis of Tumor and Endothelial Cell Viability and Survival Using Sulforhodamine B and Clonogenic Assays. In *Mammalian cell viability: Methods and protocols* (pp. 45-56). New York: Springer.
- Yang, X., Hsieh, K., & Liu, J. (2007). Guajadial: An Unusual Meroterpenoid from Guava Leaves *Psidium guajava*. *Organic Letters*, 9(24), 5135-5138.