



**UNIVERSITI PUTRA MALAYSIA**

**ISOLATION AND CHARACTERIZATION OF BACTERIOPHAGE FROM  
CRAB**

**AMIRA WAHIDA MOHAMAD SAFIEE**

**FBSB 2015 93**

**ISOLATION AND CHARACTERIZATION OF BACTERIOPHAGE  
FROM CRAB**

**AMIRA WAHIDA BINTI MOHAMAD SAFIEE**

**163458**

**Dissertation submitted in partial fulfillment of the requirement for the course  
BMY 4999 Project in the Department of Microbiology  
Universiti Putra Malaysia  
JUNE 2015**

**ISOLATION AND CHARACTERIZATION OF BACTERIOPHAGE  
FROM CRAB**

**AMIRA WAHIDA BINTI MOHAMAD SAFIEE**

**163458**

**DEPARTMENT OF MICROBIOLOGY  
FACULTY BIOTECHNOLOGY AND BIOMOLECULAR SCIENCES  
UNIVERSITI PUTRA MALAYSIA  
2015**

## PENGESAHAN

Dengan ini adalah disahkan bahawa projek yang bertajuk “ISOLATION AND CHARACTERIZATION OF BACTERIOPHAGE FROM CRAB” telah disiapkan serta dikemukakan kepada Jabatan Mikrobiologi oleh AMIRA WAHIDA BINTI MOHAMAD SAFIEE (163458) sebagai syarat untuk kursus BMY 4999 projek.

Disahkan oleh:

.....

Tarikh: .....

Prof. Dr. Tan Wen Siang

Penyelia

Jabatan Mikrobiologi

Fakulti Bioteknologi dan Sains Biomolekul

Universiti Putra Malaysia

.....

Tarikh: .....

Prof. Madya Dr. Muhajir Hamid

Ketua

Jabatan Mikrobiologi

Fakulti Bioteknologi dan Sains Biomolekul

Universiti Putra Malaysia

## ABSTRACT

This project was carried out to isolate, purify and characterize the bacteriophage from the crab (*Callinectes sapidus*) also known as blue crab. The crab was obtained freshly from Jetty Wak Sempo, Bagan Lalang, Sepang. Seven bacteria was isolated from the crab and labelled as A, B, C, D, E, F and G were used as a screening agent for the isolation of the bacteriophage. Besides, another 5 species of known bacteria were used for bacteriophage isolation. The bacteria were *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Escherichia coli*, *Bacillus cereus* and *Staphylococcus aureus*. All 12 species of bacteria were used but only bacteria C was infected by the bacteriophage. The bacteriophage was labelled as C-CC. The bacteriophage infected bacteria was identified by gram staining and biochemical tests. Based on the gram staining, the bacterium is a gram negative bacterium with coccobacillus shape. Then, the isolated bacteriophage was characterized based on their protein, genomic, biology and physical characteristics. At the end of the study, the isolated bacteriophage was confirmed classified as DNA-containing bacteriophage with lytic life cycle. The bacteriophage could be as used in the phage therapy against the bacterial contamination.

## ABSTRAK

Projek ini dijalankan untuk memencilkan, purifikasi dan mengklasifikasikan bakteriofaj daripada ketam (*Callinectes sapidus*) yang juga dikenali sebagai ketam biru. Ketam ini dibeli segar dari Jeti Wak Sempo Bagan Lalang, Sepang. Tujuh bakteria telah dipencilkan daripada ketam dan dilabel sebagai A, B, C, D, E, F dan G dan digunakan sebagai ajen pemeriksaan untuk memencilkan bakteriofaj. Selain itu, terdapat lima bakteria juga yang digunakan untuk memencilkan bakteriofaj. Bakteria-bakteria tersebut ialah *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Escherichia coli*, *Bacillus cereus* dan *Staphylococcus aureus*. Kesemua 12 spesis bakteria yang digunakan untuk memencilkan bakteriofaj, hanya bakteria C yang dijangkiti oleh bakteriofaj. Bakteriofaj yang dipencilkan dilabel sebagai bakteriofaj C-CC. Bakteria yang dijangkiti oleh bakteriofaj dianalisis dengan pewarnaan gram dan ujian biokimia. Berdasarkan pewarnaan gram, bakteria C merupakan gram negatif bakteria dan berbentuk coccobacillus. Selepas itu, bakteriofaj yang dipencilkan dianalisa berdasarkan protein, genomik, dan ciri-ciri biologi dan fizikal. Di akhir eksperimen ini, bakteriofaj yang dipencilkan dipastikan diklasifikasikan sebagai DNA-bakteriofaj dengan kitaran hidup lytic. Bakteriofaj juga boleh digunakan sebagai terapi faj terhadap kontaminasi bakteria.

## ACKNOWLEDGEMENT

Assalamualaikum W.B.T.

First and foremost I am grateful to Prof. Dr. Tan Wen Siang, my project supervisor. I am extremely thankful and indebted to him for sharing expertise, sincere and valuable guidance, advice and encouragement extended to me. I take this opportunity to express gratitude to the Department of Microbiology and Biochemistry members and staffs for their help, support and for providing equipment and services along my final year project.

I wish to express my sincere thanks to my parents Mohamad Safiee bin Selamat and Fariza binti Ali@Fauzi and family who always give encouragement, moral support and attention to me every time even in a far distance. A million thanks you for the supports and loves.

In addition, my sincere thank you to all the lab seniors in the lab 134, especially to Ms. Wong Chuan Loo. Thank you for helping me in my final year project, giving me a kind guidance, patience and supporting me when I was in troubles.

I take this opportunity to express gratitude to my lab mates which are Farah Hanim binti Agusalim, Nurul Natasya binti Zahari and Najatullah Siddiqi bin Lokman. I am so grateful to have them in the lab. A thousand thanks for sharing valuable information and providing excellent teamwork to me in all the times.

I also place on record, my sense of gratitude to one and all, who directly or indirectly, have lent their hand in my final year project.

## TABLE OF CONTENTS

PENGESAHAN	i
ABSTRACT	ii
ABSTRAK	iii
ACKNOWLEDGEMENT	iv
LIST OF TABLES	vi
LIST OF FIGURES	vii
CHAPTER 1	1
INTRODUCTION	1
CHAPTER 2	3
LITERATURE REVIEW	3
2.1 BACTERIAL DISEASES OF CRAB ( <i>Callinectes sapidus</i> )	3
2.2 BACTERIOPHAGE	4
2.2.1 INTRODUCTION TO BACTERIOPHAGE	4
2.2.2 COMPOSITION AND STRUCTURE OF BACTERIOPHAGE	5
2.2.3 CLASSIFICATION OF BACTERIOPHAGE	6
2.2.4 LYTIC AND LYSOGENIC CYCLE	8
2.2.5 INFECTION	9
2.3 APPLICATION OF BACTERIOPHAGE AS PHAGE THERAPY	11
CHAPTER 3	13
MATERIALS AND METHODS	13
CHAPTER 4	25
RESULTS AND DISCUSSION	25
4.1 HOST BACTERIA	25
4.1.1 ISOLATION, PURIFICATION AND IDENTIFICATION OF HOST BACTERIA	25
4.2 BACTERIOPHAGE	32
4.2.1 ISOLATION OF BACTERIOPHAGE	32
4.2.2 AMPLIFICATION AND PURIFICATION OF BACTERIOPHAGE	33
4.2.3 CALCULATION OF BACTERIOPHAGE TITER	34
4.2.4 PROTEIN ANALYSIS AND QUANTIFICATION	35
4.2.5 PHAGE GENOMIC CHARACTERIZATION	36
4.2.6 PHAGE BIOLOGY AND PHYSICAL CHARACTERIZATION	39
CHAPTER 5	43
CONCLUSION AND RECOMMENDATIONS	43
REFERENCES	44
APPENDICES	48



## LIST OF TABLES

<b>Table</b>	<b>Caption</b>	<b>Page</b>
1	The functions of the structural components of bacteriophage	6
2	Classification and basic properties of the bacteriophages	8
3	Result on cultural and biochemical characteristics of isolated bacteria	30
4	Result of cultural and biochemical test of bacteria C	32
5	Bacteriophage titer	34
6	Host range analysis of bacteriophage C-CC	40

## LIST OF FIGURES

Figures	Caption	Page
1	The basic structure of bacteriophage	6
2	The lytic and lysogenic life cycles of a bacteriophage	10
3	Isolated bacteria	26
4	Observation of gram staining of bacteria C under light microscope (oil-immersion 100X)	31
5	Clearing zone of bacteriophage (C-CC) infected bacteria C	33
6	Blue band of purified bacteriophage	34
7	SDS-PAGE analysis of bacteriophage C-CC	36
8	Nucleic acid of bacteriophage C-CC	37
9	DNase and RNase treatment.	38
10	Restriction fragments analysis of phage genomic DNA.	39
11	<i>Pseudomonas aeruginosa</i> infected by bacteriophage C-CC	40
12	Graph on pH versus infection of bacteriophage (pfu/ml)	41
13	Graph on temperature versus infection of bacteriophage (pfu/ml)	42

## CHAPTER 1

### INTRODUCTION

Bacteriophages or phages are viruses that infect bacteria cells and can be found abundantly in environment with an estimation of  $10^{32}$  bacteriophages worldwide (Hanlon, 2007). Bacteriophages are usually found in aqueous environment and have been used as an indicator of bacterial contamination (McLaughlin *et al.*, 2006). The phages' community in the aquatic environment systems was reported by Weinbauer (2004) to be increased in the range of  $10^4$  to  $10^8$  virions per ml. They have been highly exploited in many practical applications due to their high specificity in nature (McLaughlin *et al.*, 2007). Recently, phages have been used as a biological control agent to treat human, plant and animal bacterial infections (McLaughlin *et al.*, 2007; Wong *et al.*, 2014; Shivu *et al.*, 2007; Silva *et al.*, 2014).

Asia contributes more than 90% of the world's aquaculture production. Intensive commercialization results in serious diseases problems in the aquaculture sector (Bondad-Reantaso *et al.*, 2005). Food and Agriculture Organization of the United Nations (FAO) (2004) reported that aquaculture industries are the fastest growing food producing sector in the world. The average annual growth rate is 8.9% since 1970, compared to only 1.2% for capture fisheries and 2.8% for terrestrial farmed meat production systems over the same period. Recent statistics indicated that the aquatic sector have reached the production of 9.4% annual percentage growth rate (APR) compared with meat production such as pigs (3.15%), poultry (5.1%), beef and meal (1.2%) and mutton and lamb (1.0%). In 2002, the total world

aquaculture production including aquatic plants was reported to be 51.4 million tonnes by volume and US\$ 60.0 billion by value (Crespi, 2005).

Like other farming systems, antibiotics have been frequently used in the aquaculture industries have been frequently used to control or treat the bacteria diseases. However, the extreme usage of antibiotics as therapeutics agent in hatcheries and farms has led to the emergences of antibiotics resistant strains of bacteria in the aquatic environment. Thus, the application of bacteriophages in aquaculture is one of the alternative solutions to curb multi drug-resistant bacteria (Karunasagar *et al.*, 1994). The present study was carried out with the aim to reduce the load of bacteria present in the aquaculture environment through the application of bacteriophage isolated from raw crab (*Callinectes sapidus*).

The objectives of this project were:

1. To isolate and purify the bacteriophage from the crab.
2. To characterize the isolated bacteriophage.

## REFERENCES

- Ackermann, H. W. and DuBow, M. S. (1987). Viruses of prokaryotes. I. *General properties of bacteriophages*. Florida: CRC Press.
- Ackerman, H., W. (1998). Tailed bacteriophages: the Caudovirales. *Advance Virus Research* 51:135-201.
- Ackermann, H., W. (2003). Bacteriophage observations and evolution. *Research in Microbiology* 154: 245–251.
- Ackermann H., W. (2007). 5500 phages examined in the electron microscope. *Archives of Virology* 152, 227-243.
- Adams, M., H. (1959). *Bacteriophages*. Interscience Publishers, Incorporation, New York.
- Barrow, P., A. and Soothill, J., S. (1997). Bacteriophage therapy and prophylaxis: rediscovery and renewed assessment of potential. *Trends Microbiology* 5:268–271.
- Bondad-Reantaso, M., G., Subasinghe, R., P., Arthur, J., R., Ogawa, K., Chinabut, S., Adlard, R., Tan, Z. and Shariff, M. (2005). Disease and health management in Asian aquaculture. *Veterinary Parasitology* 132(3-4):249-72.
- Cabello, F., C. (2006). Heavy use of prophylactic antibiotics in aquaculture: a growing problem for human and animal health and for the environment. *Environmental Microbiology* 8: 1137–1144.
- Cappucino, J., G. and Sherman, N. (2011). *Microbiology: A Laboratory Manual*. Pearson Benjamin Cummings Publishers.
- Carlton, R., C. (1999). Phage therapy in the past and future. *Archivum Immunologiae et Therapiae Experimentalis* 47: 267–274.
- Crespi, V., (2005). FAO Fact Sheets on Aquaculture. *FAO Aquacult. Newsl.* 32: 44.
- Davis, J., W. and Sizemore, R., K. (1982). Incidence of *Vibrio* species associated with blue crab (*Callinectes sapidus*) collected from Galveston Bay, Texas. *Appl. Environmental Microbiology* 43: 1092–1097.
- Engelkirk, P., G. and Burton, G., R. (2006). *Burton's Microbiology for The Health Sciences*. Lippincott Williams & Wilkins.
- Fujino, T., Okuno, Y., Nakada, D., Aoyama, A., Fukai, K., Mukai, T. and Uebo, T. (1953). On the bacteriological examination of Shirasu food poisoning. *Medical Journal of Osaka University* 4: 299–304.
- Gorski, A., Dabrowska, K., Switała-Jelen, K., Nowaczyk, M., Weber-Dabrowska, B., Boratynski, J., Wietrzyk, J. and Opolski, A. (2003). New insights into the

possible role of bacteriophages in host defence and disease. *Medical Immunology* 2: 2.

Guttman, B., Raya, R. and Kutter, E. (2005). Basic Phage Biology. In *Bacteriophages: Biology and applications*. CRC Press.

Hanlon, G., W. (2007). Review Bacteriophages: an appraisal of their role in the treatment of bacterial infections. *International Journal of Antimicrobial Agents* 30: 118–128.

Hendrix, R., W. (2002). Bacteriophages: evolution of the majority. *Theoretical Population Biology* 61: 471–480.

Hendrix, R., W., Smith, M., C., M., Burns, R., N., Ford, M., E. and Hatfull, G., F. (1999). Evolutionary relationships among diverse bacteriophages and prophages: All the world's a phage. *Proceedings National Academy of Sciences, USA* 96: 2192–2197.

Huq, A., Huq, S., A., Grimes, D., J., O'brien, M., Chu, K., H. and Capuzzo, J., M. (1986). Colonization of the gut of the blue crab (*Callinectes sapidus*) by *Vibrio cholerae*. *Applied and Environmental Microbiology* 52: 586–588.

Inal, J., M. (2003). Phage therapy: a reappraisal of bacteriophages as antibiotics. *Archivum Immunologiae et Therapia Experimentalis* 51(4):237-244.

Inamdar, M., M., Gelbart, W., M. and Philips, R. (2006). Dynamics of DNA ejection from bacteriophage. *Biophysical Journal* 91: 411-420.

Jensen, E., C., Schrader, H., S., Rieland, B., Thompson, T., L., Lee, K., W., Nickerson, K., W. and Kokjohn, T., A. (1998). Prevalence of broad-host-range lytic bacteriophages of *Sphaerotilus natans*, *Escherichia coli*, and *Pseudomonas aeruginosa*. *Applied and Environmental Microbiology* 64: 575–580.

Kakoma, K (2009). Isolation and characterization of bacteriophage and their potential use for the control of bacterial infections in poultry.

Kalmansom, G. and Bronfenbrenner, J. (1942). Evidence of serological heterogeneity of polyvalent pure line bacteriophages. *Journal Immunology* 45: 13–15.

Karunasagar, I., Pai, R. and Malathi, G., R. (1994). Mass mortality of *Penaeus monodon* larvae due to antibiotic resistant *Vibrio harveyi* infection. *Aquaculture* 128: 203–209.

Kurtböke, I. (2012). Bacteriophages. Publishing Process Manager Maja Bozicevic.

Kutter, E. and Sulakvelidze, A. (2005). Bacteriophages, biology and applications 121–136. CRC Press, USA.

- Matsuzaki, S., Rashel, M., Uchiyama, J., Sakurai, S., Ujihara, T., Kuroda, M., Ikeuchi, M., Tani, T., Fujieda, M., Wakiguchi, H. and Imai S. (2005). Bacteriophage therapy: a revitalized therapy against bacterial infectious diseases. *Journal of Infection and Chemotherapy* 11(5): 211-219.
- McLaughlin, M., R., Balaa, M., F., Sims, J. and King. R. (2006). Isolation of *Salmonella* bacteriophages from swine effluent lagoons. *Journal of Environmental Quality* 35:522-528
- Nakai, T. and Park, S., C. (2002). Bacteriophage therapy of infectious diseases in aquaculture. *Research in Microbiology* 153: 13-18.
- Nester, E., W., Anderson, D., G., Robert, C., E., Pearsall, N., N. and Hurley, D. (2004). *A human prospective of microbiology*. New York: McGraw Hill Book Company Incorporation.
- O'Flaherty, S., Ross, R., P. and Coffey, A. (2009). Bacteriophage and their lysins for elimination of infectious bacteria. *FEMS Microbiology Review* 801-819.
- Olson, M., R, Axler, R., P. and Hicks, R., E. (2004). Effects of freezing and storage temperature on MS2 viability. *Journal of Virological Methods* 122:147-152.
- Rakhuba, D., V., Kolomiets, E., I., SzwajcerDey, E. and Novik, G., I. (2010). Bacteriophage receptors, mechanisms of phage adsorption and penetration into host cell. *Polish Journal of Microbiology* 59(3):145-155.
- Rohwer, F. & Edwards, R. (2002). The phage proteomic tree: Genome-based taxonomy for phage. *Journal Bacteriology* 184: 4529-4535.
- Rosen, B. (1970). Shell disease of aquatic crustaceans. In: A Symposium on Diseases of Fishes and Shellfishes, 409-415. Special Publication No. 5, American Fisheries Society, Washington, DC.
- Shivu, M., M., Rajeeva, B., C., Girisha, S., K., Karunasagar, I., Krohne, G. and Karunasagar, I. (2007). Molecular characterization of *Vibrio harveyi* bacteriophages isolated from aquaculture environments along the coast of India. *Environmental Microbiology* 9(2): 322-331.
- Silva, Y., J., Costa, L., Pereira, C., Mateus, C., Cunha, A., Calado, R., Gomes, N., C., M., Pardo, M., A., Hernandez, I. and Almeida, A. (2014). Phage therapy as an approach to prevent *Vibrio anguillarum* infections in fish larvae production. *Plos ONE* 9(12):e114197.
- Sindermann, C., J. and Lightner, D., F. (1988). *Disease Diagnosis and Control in North American Marine Aquaculture*. Elsevier, New York, NY.
- Summers, W. C. (1999). *Felix d'Herelle and the origins of molecular biology*. Yale University Press New Haven, CT.
- Sulakvelidze, A. (2001). Bacteriophages as therapeutic agents. *Annals of Medicine* 33: 507-509.



- Sulakvelidze, A., Alavidze, Z. and Morris, J. G. Jr. (2001). Bacteriophage therapy. *Antimicrob Agents and Chemotherapy* 45: 649–659.
- Thacker, P., D. (2003). Set a microbe to kill a microbe: drug resistance renews interest in phage therapy. *Journal of the American Medical Association* 290:3183–5.
- Tey, B., T., Ooi, S., T., Yong, K., C., Tan Ng, M., Y., Ling, T., C., Tan, W., S. (2009). Production of fusion m13 phage bearing the disulphide constrained peptide sequence (C-WSFFSNI-C) that interacts with hepatitis B core antigen. *Journal African Biotechnology* 8:268–273.
- Vogan, C., L. and Rowley, A., F. (2002). Effects of shell disease syndrome on the haemocytes and humoral defences of the edible crab, *Cancer pagurus*. *Aquaculture* 205: 237–252.
- Wang, W. (2011). Bacterial diseases of crabs: a review. *Journal of Invertebrate Pathology* 106: 18–26.
- Weinbauer, M.G. (2004). Ecology of prokaryotic viruses. *FEMS Microbiology* 28: 127–181.
- Williams, R., C. and Fraser, D. (1956). Structural and functional differentiation in T2 bacteriophage. *Virology* Volume 2(3):289–307.
- Wong, C., L., Sieo, C., C., Tan, W., S., Abdullah., N., Hair-Bejo, M., Abu, J. and Ho, Y., W. (2014). Evaluation of a lytic bacteriophage,  $\Phi$  st1, for biocontrol of *Salmonella enterica* serovar Typhimurium in chickens. *International Journal of Food Microbiology* 172: 92–101.
- Young, R., Wang, I. -N., & Roof, W. D. (2000). Phages will out: strategies of host cell lysis. *Trends Microbiology* 8:120–128.