



UNIVERSITI PUTRA MALAYSIA

**ANTIBACTERIAL ACTIVITY OF LOCALLY ISOLATED THERMOPHILIC
FUNGI**

AKMAL HAYAT ABDUL KARIM

FBSB 2015 91

**ANTIBACTERIAL ACTIVITY OF LOCALLY ISOLATED
THERMOPHILIC FUNGI**

AKMAL HAYAT BIN ABDUL KARIM

162579

Dissertation submitted in partially fulfilment of the
Requirement for the course BMY 4999 Project in the
Department of Microbiology
Universiti Putra Malaysia

June 2015

PENGESAHAN

Dengan ini adalah disahkan bahawa tesis projek yang bertajuk “Antibacterial Activity Of Locally Isolated Thermophilic Fungi” telah disiapkan serta dikemukakan kepada Jabatan Mikrobiologi oleh Akmal Hayat Bin Abdul Karim (162579) sebagai syarat untuk kursus BMY 4999 Projek.

Disahkan oleh:

(Dr. Wan Zuhainis Binti Saad)

Penyelia Projek

Tarikh:

(Prof. Madya Dr. Muhajir Bin Hamid)

Ketua Jabatan Mikrobiologi,
Fakulti Bioteknologi dan Sains Biomolekul,
Universiti Putra Malaysia.

Tarikh:

ACKNOWLEDGEMENTS

In the name of Allah, The Most Gracious, The Most Merciful.

All praises to Allah Almighty, for the strengths and guidance in completing my Final Year Project BMY 4999. Without His help, ones cannot reach their destination.

First and foremost, I would like to express my deepest gratitude to my supervisor, Dr. Wan Zuhainis Saad for her support, guidance and encouragement throughout the completion of this project. I would also express my deepest appreciation to Mohd Khairil Radzali for all the guidance and knowledge in assisting me throughout the year and also to all postgraduate students in Mycology laboratory.

Extend deepest gratitude and devotion to my family for their understanding and support, and also to Coordination Officer PALAPES UPM (Laut), Lt. Kdr. Mohd Syahirudin Bin Hamsin TLDM for the understanding my duty and responsibility as final year student instead of being a part of the Royal Malaysian Navy (RMN) officers.

Lastly, I wish to express my thankful to all those who have been involved in completing this project.

TABLE OF CONTENTS

	PAGE
ACKNOWLEDGEMENTS	i
LIST OF CONTENTS	ii
LIST OF TABLES	v
LIST OF FIGURES	vi
LIST OF ABBREVIATIONS	vii
ABSTRACT	viii
ABSTRAK	ix
CHAPTERS:	
1. Introduction	1
1.1 Research Background	1
1.2 Problem Statement	3
1.3 Hypothesis	3
1.4 Objectives	3
2. Literature review	4
2.1 Introduction to Fungi	4
2.2 Introduction to Thermophilic Fungi	5
2.3 History of Antimicrobial Discovery	7
2.4 Introduction to Fungal Metabolites	8
2.5 Isolation of Fungal Secondary Metabolites	9
2.5.1 Solid State Fermentation (SSF)	10

2.5.2	Submerged Fermentation (SmF)	12
3.	Methodology	14
3.1	Sampling	14
3.2	Cultivation and Isolation of Pure Culture	14
3.3	Growth of Thermophilic Fungi	14
3.4	Morphology Study on isolated thermophilic fungi	15
3.4.1	Macroscopic	15
3.4.2	Microscopic	15
3.5	Dry Cell Weight	15
3.5.1	Potato Dextrose Broth Culture	15
3.5.2	Determination of Dry Cell Weight	16
3.6	Extraction of Fungal Metabolites	16
3.6.1	Potato Dextrose Broth Culture	16
3.6.2	Methanolic Crude Extraction	16
3.7	Antibiotic Disc Preparation	17
3.8	Susceptibility Test	17
3.8.1	Preparation of Inoculums	17
3.8.2	Disc Diffusion Method	17
4.	Results and discussion	19
4.1	Sampling, Isolation and Cultivation of Pure Culture	19
4.2	Growth of Thermophilic Fungi	19
4.3	Macroscopic and Microscopic Study of CP1 and CP2	22
4.4	Dry Cell Weight	23
4.5	Disc Diffusion Method	25
5.	Conclusions	33

6. References	34
7. Appendices	38



LIST OF TABLES

	PAGE
Table 1: Growth of CP1 at 45°C, 50°C, 55°C	20
Table 2: Growth of CP2 at 45°C, 50°C, 55°C	21
Table 3: Antibacterial activity against targeted microorganisms	25
Table 4: Antibacterial activity of CP1 and CP2 against <i>Escherichia coli</i> ATCC 8739	27
Table 5: Antibacterial activity of CP1 and CP2 against <i>Escherichia coli</i> H7 E187	28
Table 6: Antibacterial activity of CP1 and CP2 against <i>Salmonella typhimurium</i> S836	29
Table 7: Antibacterial activity of CP1 and CP2 against Methicillin-resistant <i>Staphylococcus aureus</i> S547	30
Table 8: Antibacterial activity of CP1 and CP2 against <i>Listeria monocytogenes</i> L10	31
Table 9: The soil depth measurement, temperature and pH of the soil samples	40
Table 10: Dry cell weight of CP1 (g/L)	41
Table 11: Dry cell weight of CP2 (g/L)	42

LIST OF FIGURES

	PAGE
Figure 1: Microscopic view of mycelium	4
Figure 2: Relationships between the phyla of Kingdom Fungi	5
Figure 3: Typical fermenter of Submerged Fermentation	13
Figure 4: CP1 pure culture	19
Figure 5: CP2 pure culture	19
Figure 7: Macroscopic and microscopic (1000x) view of CP1	22
Figure 8: Macroscopic and microscopic (1000x) view of CP2	22
Figure 9: Dry cell weight of CP1 (g/L)	24
Figure 10: Dry cell weight of CP2 (g/L)	24

LIST OF ABBREVIATIONS

°C - degree Celcius

µg - microgram

g - gram

L - Litre

mg - milligram

mL - millimeter

sp - species

SSF - Solid State Fermentation

SmF - Submerged Fermentation

PDA - Potato Dextrose Agar

PDB - Potato Dextrose Broth

MHA - Muller-Hinton Agar

TSB - Trypticase Soy Broth

ABSTRACT

Sampling of thermophilic fungi was done at Cherana Puteh Hot Springs in Alor Gajah, Melaka. Isolation was done and two pure cultures were obtained named CP1 and CP2. Then, it was incubated at three different ranges of temperature to determine the best temperature of fungal growth. After that, the growth profiles of isolated thermophilic fungi was measured based on their dry cell weight. The stationary phase growths of thermophilic fungi were determined. For the screening of antibacterial properties, the secondary metabolites of thermophilic fungal secondary metabolites were extracted using methanol. The methanolic mixtures were filtered to separate the filtrate and mycelium. The crude extracts were screened for antibacterial activity against some clinical strains of test microorganisms. The test microorganisms were *Escherichia coli* ATCC 8739, *Escherichia coli* H7 ATCC E187, *Staphylococcus aureus* ATCC 6538, *Pseudomonas aeruginosa* ATCC 15442, *Bacillus subtilis* ATCC 6633, *Salmonella typhimurium* S836, Methicillin-resistant *Staphylococcus aureus* S547, and *Listeria monocytogenes* L10. Disc diffusion method was conducted to screen for the antibacterial properties that produced by the isolates against stated test microorganisms.

ABSTRAK

Sampel *thermophilic fungi* telah berjaya diambil dari Kolam Air Panas Cherana Putih di Alor Gajah, Melaka. Pengasingan sampel telah selesai dilakukan dan mendapat dua kultur tulen yang dinamakan sebagai CP1 dan CP2. Kemudian ia telah dikulturkan di tiga suhu yang berbeza untuk mengenal pasti suhu terbaik untuk pembesarannya. Selepas itu, pembesaran kultur *thermophilic fungi* diukur di melalui peningkatan berat kering sel dengan. Fasa tumbesaran pegun *thermophilic fungi* dikenal pasti. Untuk saringan keupayaan antimikrob, metabolit sekunder *thermophilic fungi* diekstrak dengan menggunakan metanol dan kandungannya telah ditapis bagi mengasingkan miselium. Ekstrak mentah telah disaring keupayaan antimikrobnya melalui ujian terhadap mikroorganisma sasaran. Kesemua mikroorganisma sasaran adalah *Escherichia coli* ATCC 8739, *Escherichia coli* H7 E187, *Staphylococcus aureus* ATCC 6538, *Pseudomonas aeruginosa* ATCC 15442, *Bacillus subtilis* ATCC 6633, *Salmonella typhimurium* S836, Methicillin-resistant *Staphylococcus aureus* S547, and *Listeria monocytogenes* L10. Kaedah resapan cakera telah diaplikasi untuk mengenal pasti keupayaan antimikrob yang dihasilkan oleh *thermophilic fungi*.

CHAPTER 1

INTRODUCTION

1.1 Research Background

According to Cooney & Emerson (1964), thermophilic fungi are the fungi that have the ability to grow at temperatures between 20°C and 50°C. The distribution of thermophilic fungi occurs at warmer sites. Thermophilic fungi are often distributed in large scales in natural geothermal sites such as hot springs, geothermal soils and hot springs effluent channels (Ellis, 1980).

Thermophilic fungi are special microorganisms with their extremely stable cellular component together with unique metabolic capabilities. This is a great interest to microbiologists and biotechnologists, as the organisms isolated from these environments are a good source of thermostable enzymes and antimicrobial properties offer considerable promise for biotechnological application (Sharma *et al.*, 2013).

Thermophilic fungi are productive and talented producers of structurally diverse bioactive metabolites and have yielded some of the most important products for the pharmaceutical industry (Ola, 2014). According to Bérdy (2005), eukaryotic fungi are the most frequent and most versatile producers for bioactive metabolites. Fungal secondary metabolites have revolutionized the yielding of antimicrobial drugs (Aly *et al.*, 2011).

Antibiotics are the most important category of secondary metabolites extracted from microorganisms using fermentation (Subramaniyam and Vimala, 2012). The first antibiotics, penicillin G from *Penicillium notatum* was discovered by

Alexander Fleming in 1928 (Aly *et al*, 2011). Antibiotics are products of fermentation and are produced in the stationary phase, often in response to stress conditions (Robinson *et al.*, 2001).

The 21st Century opened a new era of antibiotic research. The value of biopharmaceuticals was estimated to be 41 billion dollars in the global market, with a growth rate of 21% over the period from 2004 to 2008. This is due to urgent clinical needs and the increasing serious health problem such as HIV, multi-resistant strains, reappearing mycobacteria thus required development of new technologies which are more effective (Aly *et al*, 2011).

1.2 Problem Statement

Serious infections caused by bacteria that have become resistant to commonly used antibiotics have become a major global healthcare problem in the 21st century. They are not only more severe, require longer and complex treatments, but they are also significantly more expensive to diagnose and to treat (Alanis, 2005).

To date, certain antibacterial agent or antibiotics available in market are irrelevant to be used. The morbidity and mortality has increased including the cost of healthcare due to emergence of antibiotics resistance microorganisms.

Antibiotic resistance, initially a problem of the hospital setting associated with an increased number of hospital acquired infections usually in critically ill and immunosuppressed patients, has now extended out to the community causing severe infections difficult to diagnose and treat (Alanis, 2005).

1.3 Hypothesis.

1. Thermophilic fungi isolated locally from the hot springs may contain antibacterial properties.

1.4 Objectives

1. To isolate local thermophilic fungi
2. To screen the antibacterial properties of thermophilic fungi.

REFERENCES

- Adimpong, D. B., Sørensen, K. I., Thorsen, L., Stuer-Lauridsen, B., Abdelgadir, W. S., Nielsen, D. S., & Jespersen, L. (2012). Antimicrobial susceptibility of *Bacillus* strains isolated from primary starters for African traditional bread production and characterization of the bacitracin operon and bacitracin biosynthesis. *Applied and environmental microbiology*, 78(22), 7903-7914.
- Alanis, A. J. (2005). Resistance to antibiotics: are we in the post-antibiotic era?. *Archives of medical research*, 36(6), 697-705.
- Alderman, D. J., & Smith, P. (2001). Development of draft protocols of standard reference methods for antimicrobial agent susceptibility testing of bacteria associated with fish diseases. *Aquaculture*, 196(3), 211-243.
- Aly, A. H., Debbab, A., & Proksch, P. (2011). Fifty years of drug discovery from fungi. *Fungal Diversity*, 50(1), 3-19.
- Arunsasi, ManthiriKani, S., Jegadeesh, G. & Ravikumar, M. (2010). Submerged fermentation of amylase enzyme by *Aspergillus flavus* using *Cocos nucifera* meal. *Kathmandu University Journal of Science, Engineering and Technology*. 6(11): 75-87.
- Babu, K.R. & Satyanarayan, T. (1996). Production of Bacterial Enzymes by Solid State Fermentation. *Journal of Scientific and Industrial Research*, 55: 464-467.
- Bassetti, M., Ginocchio, F., & Mikulska, M. (2011). New treatment options against gram-negative organisms. *Critical Care*, 15(2), 215.
- Blackwell, M. (2011). The Fungi: 1, 2, 3... 5.1 million species?. *American Journal of Botany*, 98(3), 426-438.
- Brakhage, A. A. (2013). "Regulation of fungal secondary metabolism." *Nature Reviews Microbiology* (2012).
- Berdy, J. (2005). Bioactive microbial metabolites. *The Journal of antibiotics*, 58(1), 1-26.
- Carlile, M. J., Watkinson, S. C., & Gooday, G. W. (2001). *The Fungi*, second edition. London: Academic press.
- Callan, B. E., & Carris, L. M. (2004). Fungi on living plant substrata, including fruits. Chap. 7 in: GM Mueller, GF Bills, and MS Foster, eds. Biodiversity of Fungi. *Inventory and Monitoring Methods*.
- Carris, L. M., C. R. Little & C. M. Stiles. (2012). Introduction to Fungi. *The Plant Health Instructor*.
- Cooney, D. G., & Emerson, R. (1964). Thermophilic Fungi. *An Account of Their Biology, Activities & Classification*. W.H. Freeman and Co., San Francisco.

- de Barros, B. S., da Silva, J. P., de Souza Ferro, J. N., Agra, I. K. R., de Almeida Brito, F., Albuquerque, É. D., Caetano, L. C., & Barreto, E. (2011). Methanol extract from mycelium of endophytic fungus *Rhizoctonia* sp. induces antinociceptive and anti-inflammatory activities in mice. *Journal of natural medicines*, 65(3-4), 526-531.
- Ellis, D. H. (1980). Thermophilic fungi isolated from a heated aquatic habitat. *Mycologia*, 1030-1033.
- Fox, E. M., & Howlett, B. J. (2008). Secondary metabolism: regulation and role in fungal biology. *Current opinion in microbiology*, 11(6), 481-487.
- Haddadin, A. S., Fappiano, S. A., & Lipsett, P. A. (2002). Methicillin resistant *Staphylococcus aureus* (MRSA) in the intensive care unit. *Postgraduate medical journal*, 78(921), 385-392.
- Hedger, J. N. (1975). The ecology of thermophilic fungi in Indonesia. In *Biodegradation et Humification*, I (ed. G. Kilbertus, O. Reisinger, A. Mourey & j. A. Cancela de Fonseca), pp. 59-65. Pierron: Nancy, France.
- Hirose, K., Tamura, K., Sagara, H., & Watanabe, H. (2001). Antibiotic Susceptibilities of *Salmonella enterica* Seroovar Typhi and *S. enterica* Seroovar Paratyphi A Isolated from Patients in Japan. *Antimicrobial agents and chemotherapy*, 45(3), 956-958.
- Hölker, U., Höfer, M. & Lenz, J. (2004). Biotechnological advantages of laboratory-scale solid-state fermentation with fungi. *Applied and environmental microbiology*, 64: 175-186.
- Kavanagh, K. (2011). *Fungi: Biology and Application*, Second Edition (pp. 125-146). Chichester: John Wiley & Son, Ltd.
- Lee, H., Lee, Y. M., Jang, Y., Lee, S., Lee, H., Ahn, B. J., K, G. H. & Kim, J. J. (2014). Isolation & Analysis of the Enzymatic Properties of Thermophilic Fungi from Compost. *Mycobiology*, 42(2), 181-184.
- Lister, P. D., Wolter, D. J., & Hanson, N. D. (2009). Antibacterial-resistant *Pseudomonas aeruginosa*: clinical impact and complex regulation of chromosomally encoded resistance mechanisms. *Clinical microbiology reviews*, 22(4), 582-610.
- Maheshwari, R., Bharadwaj, G., & Bhat, M. K. (2000). Thermophilic fungi: their physiology and enzymes. *Microbiology and molecular biology reviews*, 64(3), 461-488.
- Melgar, Z., De Assis, F. V. S., & Da Rocha, L. C. (2013). Growth Curves of Filamentous Fungi for Utilization in Biocatalytic Reduction of Cyclohexanones. *Global Journal of Science Frontier Research*, 13(5).
- Morgenstern, I., Powlowski, J., Ishmael, N., Darmond, C., Marqueteau, S., Moisan, M. C., Quenneville, G. & Tsang, A. (2012). A molecular phylogeny of thermophilic fungi. *Fungal biology*, 116(4), 489-502.

- Morvan, A., Moubareck, C., Leclercq, A., Hervé-Bazin, M., Bremont, S., Lecuit, M., & Le Monnier, A. (2010). Antimicrobial resistance of *Listeria monocytogenes* strains isolated from humans in France. *Antimicrobial agents and chemotherapy*, 54(6), 2728-2731.
- Ola, A. R. (2014). Natural Products from Endophytic Fungi: Approaches for Activation Silent Biosynthetic Pathways, Structure Elucidation and Bioactivity.
- Pandey, A. (1992). Recent process developments in solid-state fermentation. *Process Biochemistry*. 27: 109-117.
- Pandey, A. (2003). Solid-state fermentation. *Biochemical Engineering Journal*. 13: 81-84.
- Pandey, A., Selvakumar, P., Soccol, C.R. & Nigam, P. (1999). Solid state fermentation for the production of industrial enzymes. *Current Science*, 77 (1): 149-162.
- Pantosti, A., Sanchini, A., & Monaco, M. (2007). Mechanisms of antibiotic resistance in *Staphylococcus aureus*.
- Redman, R. S., Litvintseva, A., Sheehan, K. B., Henson, J. M., & Rodriguez, R. J. (1999). Fungi from geothermal soils in Yellowstone National Park. *Applied and environmental microbiology*, 65(12), 5193-5197.
- Renge, V.C., Khedkar, S.V. & Nandurkar, N.R. (2012). Enzyme synthesis by fermentation method: *Scientific Reviews & Chemical Communications* 2(4): 585-590.
- Robinson, T., Singh, D., & Nigam, P. (2001). Solid-state fermentation: a promising microbial technology for secondary metabolite production. *Applied microbiology and biotechnology*, 55(3), 284-289.
- Sharma, N., Vyas, G., & Pathania, S. (2013). Research Article Culturable Diversity of Thermophilic Microorganisms Found in Hot Springs of Northern Himalayas and to Explore Their Potential for Production of Industrially Important Enzymes.
- Silver, L. L. (2012). Rational approaches to antibacterial discovery: pre-genomic directed and phenotypic screening. In *Antibiotic Discovery and Development* (pp. 33-75). Springer US.
- Subramaniam, R., & Vimala, R. (2012). Solid state and submerged fermentation for the production of bioactive substances: a comparative study. *International Journal of Science and Nature*. 3(3): 480-486.
- Tendencia, E. A. (2004). . Disk diffusion method. In *Laboratory manual of standardized methods for antimicrobial sensitivity tests for bacteria isolated from aquatic animals and environment* (pp. 13-29). SEAFDEC Aquaculture Department.
- Webster, J. & R.W.S. Weber. (2007). *Introduction to Fungi* , Third Edition (pp. 1-2). New York: Cambridge University Press.

Yu, J. H., & Keller, N. (2005). Regulation of secondary metabolism in filamentous fungi. *Annual Review of Phytopathology* , 43, 437-45.

