

UNIVERSITI PUTRA MALAYSIA

ANTIHYPERGLYCEMIC AND ANTIOXIDATIVE POTENTIALS OF MALAYSIAN BANANA (*Musa sp.*) FLOWER EXTRACTS

TAN SHIH JEN

FBSB 2015 81

ANTIHYPERGLYCEMIC AND ANTIOXIDATIVE POTENTIALS OF MALAYSIAN BANANA (*Musa* sp.) FLOWER EXTRACTS



TAN SHIH JEN

163598

DEPARTMENT OF BIOCHEMISTRY

FACULTY OF BIOTECHNOLOGY AND BIOMOLECULAR SCIENCES

UNIVERSITI PUTRA MALAYSIA

2015

ANTIHYPERGLYCEMIC AND ANTIOXIDATIVE POTENTIALS OF MALAYSIAN BANANA (*Musa* sp.) FLOWER EXTRACTS



TAN SHIH JEN

163598

Dissertation submitted in partial fulfillment of the requirement for the course of BCH 4999

Project in the Department of Biochemistry Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia

June 2015

PENGESAHAN

Dengan ini adalah disahkan bahawa laporan projek yang bertajuk "ANTIHYPERGLYCEMIC AND **ANTIOXIDATIVE** POTENTIALS OF MALAYSIAN BANANA (Musa sp.) FLOWER EXTRACTS" telah disiapkan serta dikemukakan kepada Jabatan Biokimia oleh TAN SHIH JEN sebagai memenuhi syarat untuk kursus BCH 4999.

Dr. Mohammed Nazrim Marikkar Penyelia Projek Jabatan Biokimia Fakulti Bioteknologi dan Sains Biomolekul Universiti Putra Malaysia

Disahkan oleh:

Tarikh:

Tarikh:

Prof. Dato' Dr. Abu Bakar Salleh

Ketua Jabatan Jabatan Biokimia Fakulti Bioteknologi dan Sains Biomolekul Universiti Putra Malaysia

AKNOWLEDGEMENT

Foremost, I would like to express my sincere appreciation and deepest gratitude to my project advisor, Dr. Mohammed Nazrim Marikkar for the continuous support of my degree study and research, for his patience, motivation, caring, enthusiasm, and immense knowledge. I might be unable to finish my degree study without his relentless support, tremendous encouragement and constructive comments. I was so glad and lucky to have a responsible and motivating advisor for my project.

I would also like to take this opportunity to thank all the academic staffs in Faculty of Biotechnology and Biomolecular Sciences at University Putra Malaysia (UPM) for the past four years of teaching, guiding and helping me to build up and developed a strong background and foundation in biochemistry and other related fields.

Special thanks to Dr. Mohd Shukuri Mohamad Ali for his generous help and permission to access his laboratory with the using of microplate reader machine to complete my analysis study. Also, I would like to thank all the laboratory assistants from the Biochemistry Department, Faculty of Biotechnology and Biomolecular Sciences, UPM for the guiding and training on using of the laboratory equipments.

Not forgetting Mr. Tanko Abubakar Saadiq, thanks for his willingness to share his ideas throughout the process of my study. I would also thank all of my wonderful friends and coursemates for their supports and encouragements. Last but not least, my heartfelt gratitude goes to my dearest family members. They were always supporting me and encouraging me with their best wishes.

ABSTRACT

Banana (Musa sp.) is one of the important agricultural products in Malaysia. Its flower commonly referring as 'banana heart' is a blossom which is usually red or dark red in color attached at the end of the bunch of banana fruits. Banana flower consumption is popular among the countries of Southeast Asia such as Malaysia, Philippines, Indonesia, and Si Lanka. In Malaysia, banana flowers are served as vegetables in preparing different types of cuisines. Previously, several studies were conducted to analyze the antioxidant capacity of different part of banana such as pulp, pseudostem and flower. However. the peel. leaves. antioxidant and antihyperglycemic properties of the Malaysian banana flowers have not been reported previously. In this study, six different cultivars of banana flower (Abu, Berangan, Nipah, Susu, Mas and Rastali) were investigated. The objectives of this study were to evaluate the antihyperglycemic and antioxidant properties of the Malaysian banana flowers. The potential of banana flowers as a dietary antihyperglyceamic agent was studied through screening of their anti-amylase and anti-glucosidase activity (%) at the concentration of 200.0 µg / ml. Overall, the antiamylase and anti-glucosidase activity of the Malaysian banana flowers was found in the range of 47.31 - 62.58 % and 74.98 - 91.62 %, respectively. All banana flower extracts inhibited the activity of α -glucosidase better than α -amylase at the same concentration. The total phenolic and flavonoid contents were evaluated through Folin-Ciocalteu colorimetric method and aluminium chloride method, respectively. The reducing power of the banana flower was determined by ferric reducing antioxidant power (FRAP) assay. The antioxidant capacity of the banana flower was measured through the scavenging activities of the ABTS⁺ and DPPH radical. The Susu cultivar with the highest amount of phenolic compounds $(80.13 \pm 4.64 \text{ mg of})$ Gallic acid equivalent / g of extract) was found to possess the highest and DPPH radical scavenging activity (24.73 \pm 0.04 and 25.10 \pm 0.15 µmole of Trolox equivalent [TE] / g of extract). This study concluded that the Malaysian banana flower was a potential postprandial hyperglycemia regulator and a good natural source of antioxidant.

Key Words: Antioxidant activity, total phenolic content, antihyperglycemic, banana flower

ABSTRAK

Pisang (Musa sp.) adalah salah satu produk pertanian yang penting di Malaysia. Bunganya ataupun biasa dikenali sebagai 'jantung pisang' adalah bunga berwarna merah atau merah gelap bercantung pada akhir tandan buah pisang. Penggunaan bunga pisang adalah popular di kalangan negara-negara Asia Tenggara seperti Malaysia, Filipina, Indonesia, dan Si Lanka. Di Malaysia, bunga pisang dihidangkan sebagai sayur-sayuran dalam penyediaan pelbagai jenis masakan. Sebelum ini, beberapa kajian telah dijalankan untuk menganalisis kapasiti antioksidan bahagian yang berbeza daripada pokok pisang seperti pulpa, kulit, daun, batang semu dan bunga. Walau bagaimanapun, sifat-sifat antioksidan dan antihyperglycemic bunga pisang yang terdapat di Malaysia belum dikaji sebelum ini. Dalam kajian ini, enam kultivar bunga pisang yang berbeza (Abu, Berangan, Nipah, Susu, Mas dan Rastali) telah dikaji. Objektif kajian ini adalah untuk menilai dan menganalisis sifat-sifat antihiperglisemia dan antioksidan bunga pisang yang terdapat di Malaysia. Potensi bunga pisang sebagai ejen antihiperglisemia dikaji melalui saringan aktiviti antiamilase dan anti-glucosidase (%) pada kepekatan 200.0 µg / ml. Secara keseluruhan, aktiviti anti-amilase dan anti-glucosidase bunga pisang Malaysia didapati dalam lingkungan 47.31 – 62.58 % dan 74.98 – 91.62 % masing-masing. Ekstrak bunga pisang Malaysia menghalang aktiviti α -glucosidase lebih efektif daripada α -amilase pada kepekatan yang sama. Kandungan fenol telah dinilai melalui kaedah Folin-Ciocalteu manakala kandungan flavonoid dinilai dengan kaedah aluminium klorida. Kuasa penurunan ekstrak bunga pisang telah diuji dengan kaedah ferric reducing antioxidant power (FRAP). Kapasiti antioksidan bunga pisang dianalisis dengan mengukur aktiviti pemerangkapan radikal bebas ABTS⁺ dan DPPH. Bunga pisang Susu mengandungi kandungan fenol yang tertinggi (80.13 ± 4.64 mg GAE / g ekstrak) telah didapati memiliki aktiviti pemerangkapan radikal bebas 2,2'-azinobis(3-ethylbenzothiazoline-6-sulfonic acid) diammonium salt (ABTS) dan 1,1diphenyl-2-picrylhydrazine (DPPH) yang tertinggi (24.73 ± 0.04 dan 25.10 ± 0.15 µmole daripada Trolox equivalent [TE] / g ekstrak). Kajian ini menyimpulkan bahawa bunga pisang yang terdapat di Malaysia adalah satu ejen yang berpotensi untuk mengawal gula darah dan juga sumber antioksidan baik yang boleh didapati daripada makanan.

Kata kunci : Aktiviti antioksidan, kandungan fenol, bunga pisang

TABLE OF CONTENT

		PAGE
	AKNOWLEDGEMENT	i
	ABSTRAK	ii
	ABSTRAK	iii
	TABLE OF CONTENT	iv
	LIST OF TABLES	vii
	LIST OF FIGURES	viii
	LIST OF SYMBOLS AND ABBREVIATIONS	ix
CHAI	PTER	
1.	INTRODUCTION	1
	1.1 Research Background	1
	1.2 Research Hypothesis	4
	1.3 Research Objectives	4
2.	LITERATURE REVIEW	5
	2.1 Diabetes mellitus	5
	2.2 Diabetes and overproduction of reactive oxygen species (ROS)	6
	2.3 Enzyme α -amylase and α -glucosidase	7
	2.4 Banana Plant	9
	2.5 Antioxidants	11
	2.6 Plant phenolic compounds	13
	2.7 Flavanoid	14

 \overline{O}

3.	METHODS AND MATERIALS	17
	3.1 Collection of banana flower samples	17
	3.2 List of chemicals used in the experiments	17
	3.3 Extraction of banana flowers active compounds	18
	3.4 Anti-amylase activity (%) of Malaysian banana flowers	19
	3.5 Anti-glucosidase activity (%) of Malaysian banana flowers	19
	3.6 Determination of phenolic content in Malaysian banana flowers	20
	3.7 Determination of flavonoid content in Malaysian banana flowers	21
	3.8 Determination of ABTS ⁺ radical scavenging activity of Malaysian	21
	banana flowers	
	3.9 Determination of DPPH radical scavenging activity of Malaysian	22
	banana flowers	
	3.10 Determination of reducing power of Malaysian banana flowers by	23
	FRAP	
	3.11 Statistical analysis	23
4.	RESULTS AND DISCUSSIONS	24
	4.1 Yield (%) of crude extract	24
	4.2 Determination of the antihyperglycemic activity of crude extracts of	26
	banana flowers	
	4.2.1 α-Amylase inhibiton assay	26
	4.2.2 α -Glucosidase inhibition assay	26
	4.3 Determination of phenolic content in Malaysian banana flowers	31
	4.4 Determination of flavonoid content in Malaysian banana flowers	33

v

	4.5 ABTS ⁺ radical scavenging activity of Malaysian banana flowers	35
	4.6 DPPH radical scavenging activity of Malaysian banana flowers	37
	4.7 Determination of reducing power of Malaysian banana flowers by	38
	FRAP	
5.	CONCLUSION	40
	REFERENCES	42
		51

LIST OF TABLES

Table		Page
1	The four main classes of flavonoid and the example of members in each group and their dietary sources	16
2	Yield (%) of extract from 100 g of six different Malaysian banana flowers	25
3	The antihyperglycemic properties of six different Malaysian banana flowers	27
4	The antioxidant properties of six different Malaysian banana flowers	32

LIST OF FIGURES

Figure		Page
1	The picture of banana (Musa sp.) flower	3
2	The structural formula of four common types of flavonoids	15
3	The inhibitory effect (%) of six cultivars of Malaysian banana flowers on the enzyme activities of α -amylase and α -glucosidase.	29
4	Total phenolic content of six Malaysian banana flower expressed in mg of Gallic acid equivalent [GAE] / g of extract	32
5	Total phenolic and flavonoid content of six Malaysian banana flowers	34
6	The ABTS ⁺ free radical scavenging activity of six Malaysian banana flowers.	36
7	DPPH radical scavenging activity of six Malaysian banana flowers	38
8	The reducing power of six Malaysian banana flowers equivalent to	39
	µmole of FeSO4 / g of extract	

LIST OF SYMBOLS AND ABBREVIATIONS

2,2'-azino-bis(3-ethylbenzothiazoline-

ABTS

6-sulfonic acid) diammonium salt AICI₃ Aluminium trichloride α-amylase Alpha amylase α-glucosidase Alpha glucosidase OD Absorbance value And others et al BRG Banana cultivar Berangan AB Banana cultivar Abu NPH Banana cultivar Nipah SS Banana cultivar Susu MS Banana cultivar Mas RAS Banana cultivar Rastali 1,1-diphenyl-2-picrylhydrazine DPPH °C **Degree of Celcius** FRAP Ferric reducing antioxidant power g Gram GAE Gallic acid equivalents Trolox 6-hydroxy-2,5,7,8-tetramethylchroman-2 carboxylic acid L Liter μ Micro mg Miligram Microgram μg mΜ Milimolar μΜ Micromolar Mililiter ml μ Microliter Nanometer nm Percentage % QE Quercetin equivalents R2 **R-square** ROS Reactive oxygen species NAOH Sodium hydroxide Na2CO3 Sodium bicarbonate NaNo2 Sodium nitrite w:w Weight over weight ratio Weight over volume ratio w:v

CHAPTER 1

INTRODUCTION

1.1 Research Background

It cannot be denied that agricultural sector plays an important role in boosting the economic and GDP growth of Malaysia. Since Malaysia is located on the peninsula of Southeast Asia, it has an ideal climate and condition to nourish various types of tropical fruits such as papayas, bananas, guavas, pineapples and so on. Meanwhile, the antioxidant properties of pineapples (Yuris & Siow, 2014), papaya (Maisarah *et al.*, 2013), guava (Siow & Hui, 2013) and banana (Sulaiman *et al.*, 2011) has been studied. Banana (*Musa* sp.) plant belongs to the genus of *Musa* fom the family of *Musaceae*. The banana plant is a large, perennial monocotyledonous herb. It can grow from 2 meter up to 9 meter tall at maturity (Nelson *et al.*, 2006). There are various cultivars of banana plant can be found in Malaysia. Most of these cultivars are derived from the two wild species, namely *Musa acuminata* and *Musa balbisiana*. The *Musa* species can be classified as edible and ornamental banana. Up to date, there is more than 50 edible banana cultivars available in Malaysia (Sulaiman *et al.*, 2011).

The banana fruit is the dominant product of the banana plant. It was ranked as the fourth most important food crops in the diet of millions of people especially in Southeast Asia (Darvari *et al.*, 2010). A 100 g of banana fruit provides roughly 116 kCal of energy (Kumar *et al.*, 2012). Banana fruit, which is high in nutritional value can be eaten raw or cooked. It is known for its high content of the potassium, magnesium and various vitamins. According to Kumar *et al.* (2012), a single banana can provide up to 23 % of the potassium that we need on a daily basis. In addition, due the presence of tryptophan (the precursor of serotonin), banana fruit was also known as a good natural source of antidepressant. In facts, several pharmacological properties such as antidiarrhoeal, antiulcerative, antimicrobial, antihypertensive, hypocholesterolaemic and hypoglycemic activities of banana were discussed previously (Imam & Akter, 2011). Apart from the banana fruit, other parts of the banana tree are beneficial to human as well. For example, the leaves are commonly used for wrapping food such as the local food *nasi lemak*. The fiber extracted from the banana pseudostem can be used for different purposes. The plant sap can be applied externally to stings and bites. The roots, leaves and seed mucilage also use for medicinal purposes in certain countries (Kumar *et al.*, 2012).

Banana flower or sometime refers as 'banana heart' is a blossom or inflorescence which usually in red or dark red color that is attached to the end of the bunch of banana fruits. In the red or dark red bracts, there are lots of small whitish flowers which would turn into the mature edible banana fruit. Banana flower consumption is popular among the countries in Southeast Asia such as Malaysia, Philippines, Indonesia, and Si Lanka. Its taste is a little starchy and bitter. In Malaysia, banana flowers are served as vegetables in preparing different types of cuisines. In China, banana flower is traditionally used to treat certain illness such as heart pain, diarrhea, asthma and stomach cramps (Sumathy *et al.*, 2011). Besides, the cooked banana flower was reported as a great food for diabetic patients and it can be used for the treatment of bronchitis, dysentery and ulcers (Kumar *et al.*, 2012). Both *in vivo* and *in vitro* studies have shown the health benefits for consumption of banana

2

flowers especially of its antioxidant and anti-diabetic properties (Bhaskar *et al.*, 2011; China *et al.*, 2011; Jamuna & Nandini, 2014; Sheng *et al.*, 2010).

Several studies on different parts of Malaysian banana plant were conducted previously. For example, the antioxidant property and mineral contents of several cultivar of banana fruit (Sulaiman *et al.*, 2011) and the relationship between extracting solvents and antioxidant properties of three Malysian banana (Berangan, Mas and Raja) (Shian *et al.*, 2012). Meanwhile, three cultivars of native banana namely Berangan, Mas and Nipah were also reported to possess antibacterial activity against gram negative bacteria (Fadhilah *et al.*, 2014). Apart from the banana fruit, a previous study on animal model also revealed that oral administration of both green and yellow banana peel extract at different dose (200 and 400 mg / kg) can be used to treat depression (Tee & Hassan, 2011). Although Sumathy *et al.* (2011) was done the phytochemical screening of banana flowers and reported the antimicrobial and antioxidant properties (through DPPH radical scavenging activity), there is hardly any study available which discuss the other pharmacological properties of Malaysian banana flowers. Thus, the current study was carried out to compare the antioxidant and hypoglycemic properties of different Malaysian banana flowers.



Figure 1. The picture of banana (Musa sp.) flower.

1.2 Research Hypothesis

After adverse effects of synthetic antioxidants on human health reported, the source of naturally occurring antioxidants has been targeted in recent research. Plant based foods, which are known for its phytochemical content, especially phenolic compounds which contribute to the antioxidant activity. Different cultivars of banana flowers commonly found in Malaysia has been selected in this study as they are consumed as vegetable among Malaysians. Due to the different polyphenols content, different Malaysian banana flowers may exert different antioxidant and anti-hyperglycemia properties. Increase consumption of plant based foods rich in phenolic compounds is a good way to enhance anti-oxidative defense system in the living body as well as facilitate the postprandial blood sugar regulation.

1.3 Research Objectives

- 1. To study the inhibitory effect of ethanolic extracts of banana flower on the enzyme activities of α -amylase and α -glucosidase.
- 2. To determine the antioxidant activities of ethanolic extract of banana flower with various antioxidant assays.

REFERENCES

- Adil, Z., Chabuck, G., Al-charrakh, A. H., Khazal, N., Hindi, K., & Khazal, S. (2013). Antimicrobial effect of aqueous banana peel extract, Iraq. *Pharmaceutical Sciences*, 1, 73–75.
- Ahmed, O. M., Moneim, A. A., Yazid, I. A., & Mahmoud, A. M. (2010). Antihyperglecemic, antihyperlipidemic and antioxidant effects and the probable mechanisms of action of *Ruta Graveolens* infusion and rutin in nicotinamidestreptozotocin-induced diabetes rats. *Diabetologia Croatica*, 39(1), 15–35.
- Alam, M. N., Bristi, N. J., & Rafiquzzaman, M. (2013). Review on in vivo and in vitro methods evaluation of antioxidant activity. *Saudi Pharmaceutical Journal*, 21(2), 143–152.
- Alothman, M., Bhat, R., & Karim, A. A. (2009). Antioxidant capacity and phenolic content of selected tropical fruits from Malaysia, extracted with different solvents. *Food Chemistry*, 115(3), 785–788.
- American Diabetes Association. (2012). Diagnosis and classification of diabetes mellitus. *Diabetes Care*, 35(1), 64–71.
- Anwar, F., Kalsoom, U., Sultana, B., Mushtaq, M., Mehmood, T., And, & Arshad, H.
 A. (2013). Effect of drying method and extraction solvent on the total phenolics and antioxidant activity of cauliflower (*Brassica oleracea* L.) extracts. *International Food Research Journal*, 20(2), 653–659.
- Asghar, M. N., Khan, I. U., Zia, I., Ahmad, M., & Qureshi, F. A. (2008). Modified 2,2'-azinobis(3-ethylbenzo thiazoline)-6-sulphonic acid radical cation decolorization assay for antioxidant activity of human plasma and extracts of traditional medicinal plants. *Acta Chimica Slovenica*, 55, 408–418.
- Ashok Kumar, B. S., Lakshman, K., Nandeesh, R., Arun Kumar, P. a, Manoj, B., Kumar, V., & Sheshadri Shekar, D. (2011). *In vitro* alpha-amylase inhibition and *in vivo* antioxidant potential of *Amaranthus spinosus* in alloxan-induced oxidative stress in diabetic rats. *Saudi Journal of Biological Sciences*, 18(1), 1– 5.
- Babu, P. V. A., Liu, D., & Gilbert, E. R. (2013). Recent advances in understanding the anti-diabetic actions of dietary flavonoids. *The Journal of Nutritional Biochemistry*, 24(11), 1777–1789.

Bernfeld, P. (1955). Amylases, alpha and beta. Methods in Enzymology, 1, 149-158.

Bhaskar, J. J., Shobha, M. S., Sambaiah, K., & Salimath, P. V. (2011). Beneficial effects of banana (*Musa* sp. var. elakki bale) flower and pseudostem on hyperglycemia and advanced glycation end-products (AGEs) in streptozotocininduced diabetic rats. *Journal of Physiology and Biochemistry*, 67(3), 415–25.

- Bouayed, J., & Bohn, T. (2010). Exogenous antioxidants—double-edged swords in cellular redox state : health beneficial effects at physiologic doses versus deleterious effects at high doses. Oxidative Medicine and Cellular Longevity, 3(4), 228–237.
- Cano, A., Acosta, M., & Arnao, M. B. (2000). A method to measure antioxidant activity in organic media application to lipophilic vitamins. *Redox Report*, 5(6), 365–370.
- Carlsen, M. H., Halvorsen, B. L., Holte, K., Bøhn, S. K., Dragland, S., Sampson, L., ... Blomhoff, R. (2010). The total antioxidant content of more than 3100 foods, beverages, spices, herbs and supplements used worldwide. *Nutrition Journal*, 9(3) 1–11.
- Celep, E., Aydın, A., & Yesilada, E. (2012). A comparative study on the *in vitro* antioxidant potentials of three edible fruits: Cornelian cherry, Japanese persimmon and cherry laurel. *Food and Chemical Toxicology*, 50(9), 3329–3335.
- China, R., Dutta, S., Sen, S., Chakrabarti, R., Bhowmik, D., Ghosh, S., & Dhar, P. (2011). *In vitro* antioxidant activity of different cultivars of banana flower (*Musa paradicicus* L.) extracts available in India. *Journal of Food Science*, 76(9), 1292–1299.
- Darsini, D. T. P., Maheshu, V., Vishnupriya, M., & Sasikumar, J. M. (2012). *In vitro* antioxidant activity of banana (*Musa* spp. ABB cv. Pisang Awak). *Indian Journal of Biochemistry and Biophysics*, 49, 124–129.
- Darvari, F. M., Sariah, M., Puad, M. P., & Maziah, M. (2010). Micropropagation of some Malaysian banana and plantain (*Musa* sp.) cultivars using male flowers. *African Journal of Biotchnology*, 9(16), 2360–2366.
- Ebrahinzadeh MA, Pourmorad F, Bekhradnia AR. (2008). Iron chelating activity screening, phenol and flavanoid content of some medicinal plants from Iran. *African Journal of Biotechnology*, 32, 43–49.
- Fadhilah, F., Jalani, M., Mohamad, S., Nazatul, W., & Shahidan, S. (2014). Antibacterial effects of banana pulp extracts based on different extraction methods against selected microorganisms. *Asian Journal of Biomedical and Pharmaceutical Sciences*, 4(36), 14–19.
- Fatiha, B., Khodir, M., Farid, D., Tiziri, R., Karima, B., Sonia, O., & Mohamed, C. (2012). Optimisation of solvent extraction of antioxidants (phenolic compounds) from Algerian mint (*Mentha spicata* L.). *Pharmacognosy Communications*, 2(4), 72–86.
- Ford, B. L., Bai, J., Manthey, J., & Baldwin, E. A. (2010). Improved removal of ascorbate interference in the Folin-Ciocalteu assay of "total phenolic content." *Proceedings of the Florida State Horticultural Society*, 123, 220–222.

- Forbes, J. M. (2003). Role of advanced glycation end products in diabetic nephropathy. *Journal of the American Society of Nephrology*, 14, 254–258.
- Fowler, M. J. (2007). Diabetes treatment, part 2: oral agents for glycemic management. *Clinical Diabetes*, 25(4), 131–134.
- Giacco, F., & Brownlee, M. (2010). Oxidative stress and diabetic complications. *Circulation Research*, 107(9), 1058–1070.
- Gladine, C., Morand, C., Rock, E., Bauchart, D., & Durand, D. (2007). Plant extracts rich in polyphenols (PERP) are efficient antioxidants to prevent lipoperoxidation in plasma lipids from animals fed n–3 PUFA supplemented diets. *Animal Feed Science and Technology*, 136(2007), 281–296.
- González-Montelongo, R., Lobo, M. G., & González, M. (2010). Antioxidant activity in banana peel extracts: Testing extraction conditions and related bioactive compounds. *Food Chemistry*, 119(3), 1030–1039.
- Gupta, R., Gigras, P., Mohapatra, H., Goswami, V. K., & Chauhan, B. (2003). Microbial α-amylases : a biotechnological perspective. *Process Biochemistry*, 38(11), 1599–1616.
- Halliwell, B. (2001). Free radicals and other reactive species in disease. *Encyclopedia of Life Science*, 1–7.
- Hamauzu, Y., Irie, M., Kondo, M., & Fujita, T. (2008). Antiulcerative properties of crude polyphenols and juice of apple, and Chinese quince extracts. *Food Chemistry*, 108(2), 488–495.
- Hanhineva, K., Törrönen, R., Bondia-Pons, I., Pekkinen, J., Kolehmainen, M., Mykkänen, H., & Poutanen, K. (2010). Impact of dietary polyphenols on carbohydrate metabolism. *International Journal of Molecular Sciences*, 11(4), 1365–1402.
- Haripyaree, A., Guneshwor, K., & Damayanti, M. (2010). Evaluation of Antioxidant
 Properties of Phenolics Extracted from Ananas comosus L. Notulae Scientia Biologicae, 2(2), 68–71.
- Hashim, A., Khan, M. S., Khan, M. S., Baig, M. H., & Ahmad, S. (2013). Antioxidant and alpha-amylase inhibitory property of *Phyllanthus virgatus* L.: An *in vitro* and molecular interaction study. *Biomed Research International*, 2013, 1–12.
- Haytowitz, D. B., Bhagwat, S., & Holden, J. M. (2013). Sources of variability in the flavonoid content of foods. *Procedia Food Science*, 2, 46–51.
- Hodgson, J. M., & Croft, K. D. (2010). Tea flavonoids and cardiovascular health. *Molecular Aspects of Medicine*, 31(6), 495–502.

- Hossain, M. S., Alam, M. B., Asadujjaman, M., & Zahan, R. (2011). Antidiarrheal, antioxidant and antimicrobial activities of the *Musa sapientum* seed. *Avicenna Journal of Medical Biotechnology*, 3(2), 95–105.
- Horváthová, V., Janeček, Š., & Šturdík, E. (2000). Amylolytic enzymes: their specificities, origins and properties. *Biologia*, 55(6), 605–615.
- Hue, S., Boyce, A. N., & Somasundram, C. (2012). Antioxidant activity, phenolic and flavonoid contents in the leaves of different varieties of sweet potato (*Ipomoea batatas*). Australian Journal of Crop Science, 6(3), 375–380.
- Hussain, S. A., Ahmed, Z. A., Mahwi, T. O., & Aziz, T. A. (2012). Effect of quercetin on postprandial glucose excursion after mono- and disaccharides challenge in normal and diabetic rats. *Journal of Diabetes Mellitus*, 2(1), 82–87.
- Ikawa, M., Schaper, T. D., Dollard, C. A., & Sasner, J. J. (2003). Utilization of Folin-Ciocalteu phenol reagent for the detection of certain nitrogen compounds. *Journal of Agricultural and Food Chemistry*, 51, 1811–1815.
- Imam, M. Z., & Akter, S. (2011). *Musa paradisiaca* L. and *Musa sapientum*. : A phytochemical and pharmacological review. *Journal of Applied Pharmaceutical Science*, 1(5), 14–20.
- Irshad, M., Zafaryab, M., Singh, M., & Rizvi, M. M. A. (2012). Comparative analysis of the antioxidant activity of Cassia fistula extracts. *International Journal of Medicinal Chemistry*, 2012, 1–7.
- Jahan, M., Khatoon, F., Islamia, J. M., & Delhi, N. (2010). Concentration influence on antimicrobial activity of banana blossom extract-incorporated chitosanpolyethylene glycol (CS-PEG) blended film. *Journal of Chemical and Pharmaceutical Research*, 2(5), 373–378.
- Jain, P., Bhuiyan, M. H., Hossain, K. R., & Bachar, S. C. (2011). Antibacterial and antioxidant activities of local seeded banana fruits. *African Journal of Pharmacy and Pharmacology*, 5(11), 1398–1403.
- Jamuna, J. B., & Nandini, C. D. (2014). Feeding of banana flower and pseudostem to diabetic rats results in modulation of renal GLUTs, TGFβ, PKC and extracellular matrix components. *Nutrition, Metabolism, and Cardiovascular Diseases*, 24(6), 623–631.
- Jawla, S., Kumar, Y., & Khan, M. (2012). Antimicrobial and antihyperglycemic activities of *Musa paradisiaca* flowers. *Asian Pacific Journal of Tropical Biomedicine*, 2(2), 914–918.
- Junqueira, V. B. C., Barros, S. B. M., Chan, S. S., Rodrigues, L., Giavarotti, L., Abud, R. L., & Deucher, G. P. (2004). Aging and oxidative stress. *Molecular Aspects of Medicine*, 25, 5–16.

- Kandasamy, N., & Ashokkumar, N. (2012). Myricetin, a natural flavonoid, normalizes hyperglycemia in streptozotocin-cadmium-induced experimental diabetic nephrotoxic rats. *Biomedicine & Preventive Nutrition*, 2(4), 246–251.
- Kappel, V. D., Cazarolli, L. H., Pereira, D. F., Postal, B. G., Madoglio, F. a., Buss, Z. D. S., ... B. Silva, F. R. M. (2013). Beneficial effects of banana leaves (*Musa x paradisiaca*) on glucose homeostasis: Multiple sites of action. *Brazilian Journal of Pharmacognosy*, 23(4), 706–715.
- Kazeem, M. I., Ogunbiyi, J. V, & Ashafa, A. O. T. (2013). In vitro studies on the inhibition of α-amylase and α-glucosidase by leaf extracts of *Picralima nitida* (Stapf). Tropical Journal of Pharmaceutical Research, 12, 719–725.
- Kim, J.-S., Kwon, C.-S., & Son, K. H. (2000). Inhibition of α -glucosidase and α -amylase by luteolin, a flavanoids. *Bioscience, Biotechnology, and Biochemistry*, 64(11), 2458–2461.
- Kolluru, G. K., Bir, S. C., & Kevil, C. G. (2012). Endothelial dysfunction and diabetes : Effects on angiogenesis, vascular remodeling, and wound healing. *International Journal of Vascular Medicine*, 1–30.
- Kris-etherton, P. M., & Keen, C. L. (2002). Evidence that the antioxidant flavonoids in tea and cocoa are beneficial for cardiovascular health. *Nutrition and Metabolism*, 13, 41–49.
- Krishna, V., Girish Kumar, K., & Pradeepa, K. (2013). Antibacterial activity of ethanol extract of *Musa paradisiaca* cv. Puttable and *Musa acuminate* cv. Grand naine. *Asian Journal of Pharmaceutical and Clinical Research*, 6(2), 169–172.
- Kumar, K. P. S., Bhowmik, D., Duraivel, S., & Umadevi, M. (2012). Traditional and medicinal uses of banana. *Journal of Pharmacognosy and Phytochemistry*, 1(3), 51–63.
- Kumar, S., & Pandey, A. K. (2013). Chemistry and biological activities of flavonoids : An overview. *The Scientific World Journal*, 1–16.
- Kwon, Y., Apostolidis, E., & Shetty, K. (2008). Inhibitory potential of wine and tea against alpha amylase and alpha glucosidase for management of hyperglycemia linked to type 2 diabetes. *Journal of Food Biochemistry*, 32(1), 15–31.
- Le Bras, M., Clément, M.-V., Pervaiz, S., & Brenner, C. (2005). Reactive oxygen species and the mitochondrial signaling pathway of cell death. *Histology and Histopathology*, 20(1), 205–19.
- Leonard, S. S., Cutler, D., Ding, M., Vallyathan, V., Castranova, V., & Shi, X. (2002). Antioxidant properties of fruit and vegetable juices : More to the story than ascorbic acid. *Annals of Clinical & Laboratory Science*, 32(2), 193–200.
- Leufkens, A. M., van Duijnhoven, F. J. B., Woudt, S. H. S., Siersema, P. D., Jenab, M., Jansen, E. H. J. M., ... Bueno-de-Mesquita, H. B. (2012). Biomarkers of

oxidative stress and risk of developing colorectal cancer: a cohort-nested casecontrol study in the European prospective investigation into cancer and nutrition. *American Journal of Epidemiology*, 175(7), 653–663.

- Liang, F.-Q., & Godley, B. F. (2003). Oxidative stress-induced mitochondrial DNA damage in human retinal pigment epithelial cells: a possible mechanism for RPE aging and age-related macular degeneration. *Experimental Eye Research*, 76(4), 397–403.
- Maisarah, A. ., Nurul Amira, B., Asmah, R., & Fauziah, O. (2013). Antioxidant analysis of different parts of Carica papaya. *International Food Research Journal*, 20(3), 1043–1048.
- Maisuthisakul, P., Pasuk, S., & Ritthiruangdej, P. (2008). Relationship between antioxidant properties and chemical composition of some Thai plants. *Journal* of Food Composition and Analysis, 21(3), 229–240.
- Malesev, D., & Kuntic, V. (2007). Investigation of metal-flavonoid chelates and the determination of flavonoids via metal-flavonoid complexing reactions. *Journal* of the Serbian Chemical Society, 72(10), 921–939.
- Manaharan, T., Palanisamy, U. D., & Ming, C. H. (2012). Tropical plant extracts as potential antihyperglycemic agents. *Molecules*, 17(5), 5915–5923.
- Marinova, G., & Batchvarov, V. (2011). Evaluation of the methods for determination of the free radical scavenging activity by DPPH. Bulgarian Journal of Agricultural Science, 17(1), 11–24.
- Martysiak-Żurowska, D., & Wenta, W. (2012). A comparison of ABTS and DPPH methods for assessing. *Food Science and Human Nutrition*, 11(1), 83–89.
- Mohamed Sham Shihabudeen, H., Hansi Priscilla, D., & Thirumurugan, K. (2011). Cinnamon extract inhibits α-glucosidase activity and dampens postprandial glucose excursion in diabetic rats. *Nutrition & Metabolism*, 8(46), 1–11.
- Molyneux, P. (2004). The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. *Songklanakarin Journal of Science and Technology*, 26(2), 211–219.
- Nair, S. S., Kavrekar, V., & Mishra, A. (2013). *In vitro* studies on alpha amylase and alpha glucosidase inhibitory activities of selected plant extracts. *European Journal of Experimental Biology*, 3(1), 128–132.
- Nelson, S. C., Ploetz, R. C., & Kepler, A. K. (2006). *Musa* species (banana and plantain). *Species Profiles for Pacific Island Agroforestry*, 1–33.
- Nijveldt, R. J., Nood, E. van, Hoorn, D. E. van, Boelens, P. G., Norren, K. van, & Leeuwen, P. A. van. (2001). Flavonoids: a review of probable mechanisms of action and potential applications. *The American Journal of Clinical Nutrition*, 74, 418–425.

- Palsamy, P., & Subramanian, S. (2011). Resveratrol protects diabetic kidney by attenuating hyperglycemia-mediated oxidative stress and renal inflammatory cytokines via Nrf2-Keap1 signaling. *Biochimica et Biophysica Acta*, 1812(7), 719–731.
- Pandey, K. B., & Rizvi, S. I. (2009). Plant polyphenols as dietary antioxidants in human health and disease. Oxidative Medicine and Cellular Longevity, 2(5), 270–278.
- Pari, L., & Umamaheswari, J. (2000). Antihyperglycaemic activity of *Musa* sapientum flowers: effect on lipid peroxidation in alloxan diabetic rats. *Phytotherapy Research*, 14(2), 136–138.
- Premakumara, G. a. S., Abeysekera, W. K. S. M., Ratnasooriya, W. D., Chandrasekharan, N. V., & Bentota, a. P. (2013). Antioxidant, anti-amylase and anti-glycation potential of brans of some Sri Lankan traditional and improved rice (*Oryza sativa* L.) varieties. *Journal of Cereal Science*, 58(3), 451–456.
- Prior, R. L., Wu, X., & Schaich, K. (2005). Standardized methods for the determination of antioxidant capacity and phenolics in foods and dietary supplements. *Journal of Agricultural and Food Chemistry*, 53(10), 4290–4302.
- Pulido, R., Bravo, L., & Saura-Calixto, F. (2000). Antioxidant activity of dietary polyphenols as determined by a modified ferric reducing/antioxidant power assay. *Journal of Agricultural and Food Chemistry*, 48(8), 3396–3402.
- Re, R., Pellegrini, N., Proteggente, A., Pannala, A., Yang, M., & Rice-Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biology and Medicine*, 26(98), 1231–1237.
- Rufo S. Calixtro, J., Malalay, A. P., Epino, P. B., & Avelino, L. E. (2014). Wound healing potential of the ethanolic extract of banana. *International Journal of Pharmacy*, 4(2), 33–37.
- Sahaa, R. K., Acharyaa, S., Shovon, S. S. H., & Royb, P. (2013). Medicinal activities of the leaves of *Musa sapientum* var. sylvesteris *in vitro*. *Asian Pacific Journal* of Tropical Bomedicine, 3(6), 476–482.
- Sama, K., Murugesan, K., & Sivaraj, R. (2012). *In vitro* alpha amylase and alpha glucosidase inhibition activity of crude ethanol extract of *Cissus arnottiana*. *Asian Journal of Plant Science and Research*, 2(4), 550–553.
- Sánchez-Rangel, J. C., Benavides, J., Heredia, J. B., Cisneros-Zevallos, L., & Jacobo-Velázquez, D. a. (2013). The Folin–Ciocalteu assay revisited: improvement of its specificity for total phenolic content determination. *Analytical Methods*, 5, 5990–5999.
- Saravanan, K., & Aradhya, S. M. (2011). Polyphenols of pseudostem of different banana cultivars and their antioxidant activities. *Journal of Agricultural and Food Chemistry*, 59(8), 3613–3623.

- Sattanathan, K., Dhanapal, C. K., Umarani, R., & Manavalan, R. (2011). Antihyperglycemic effect of rutin supplementation on diabetic mellitus patients. *International Journal of Research in Pharmaceutical and Biomedical Sciences*, 2(4), 1725–1730.
- Scalbert, A., Johnson, I. T., & Saltmarsh, M. (2005). Polyphenols: antioxidants and beyond. *The American Journal of Clinical Nutrition*, 81(1), 215–217.
- Sekar, M., Zulhilmi, M., Abdullah, B. I. N., Yasser, A., Bin, H., Azlan, N. O. R., ... Syafiq, M. (2014). Ten commonly available medicinal plants in Malaysia used for the treatment of diabetes - A review. *Asian Journal of Pharmaceutical and Clinical Research*, 7(1), 1–5.
- Shai, L. J., Masoko, P., Mokgotho, M. P., Magano, S. R., Mogale, a. M., Boaduo, N., & Eloff, J. N. (2010). Yeast alpha glucosidase inhibitory and antioxidant activities of six medicinal plants collected in Phalaborwa, South Africa. South African Journal of Botany, 76(3), 465–470.
- Sheng, Z., Ma, W., Gao, J., Bi, Y., Zhang, W., Dou, H., & Jin, Q. (2011). Antioxidant properties of banana flower of two cultivars in China using 2 ,2diphenyl-1-picrylhydrazyl (DPPH), sulphonate (ABTS) and inhibition of lipid peroxidation assays. *African Journal of Biotechnology*, 10(21), 4470–4477.
- Sheng, Z., Ma, W., Jin, Z., Bi, Y., Sun, Z., Dou, H., ... Han, L. (2010). Investigation of dietary fiber, protein, vitamin E and other nutritional compounds of banana flower of two cultivars grown in China. *African Journal of Biotechnology*, 9(25), 3888–3895.
- Shian, T. E., Abdullah, A., Musa, K. H., Maskat, M. Y., & Ghani, M. A. (2012). Antioxidant properties of three banana cultivars (*Musa acuminata* "Berangan", "Mas" and "Raja") extracts. *Sains Malaysiana*, 41(3), 319–324.
- Singal, P. (1998). The role of oxidative stress in the genesis of heart disease. *Cardiovascular Research*, 40(3), 426–432.
- Someya, S., Yoshiki, Y., & Okubo, K. (2002). Antioxidant compounds from bananas (*Musa Cavendish*). Food Chemistry, 79(3), 351–354.
- Sulaiman, S. F., Yusoff, N. A. M., Eldeen, I. M., Seow, E. M., Sajak, A. A. B., & Ooi, K. L. (2011). Correlation between total phenolic and mineral contents with antioxidant activity of eight Malaysian bananas (*Musa* sp.). *Journal of Food Composition and Analysis*, 24(1), 1–10.
- Sumathy, V., Lachumy, S. J., Zakaria, Z., & Sasidharan, S. (2011). *In vitro* bioactivity and phytochemical screening of *Musa acuminata* flower. *Pharmacologyonline 2*, 127, 118–127.
- Tee, T. P., & Hassan, H. (2011). Antidepressant-like activity of banana peel extract in mice. *American Medical Journal*, 2(2), 59–64.

- Tsao, R. (2010). Chemistry and biochemistry of dietary polyphenols. *Nutrients*, 2(12), 1231–1246.
- Tundis, R., Loizzo, M. R., & Menichini, F. (2010). Natural products as alphaamylase and alpha-glucosidase inhibitors and their hypoglycaemic potential in the treatment of diabetes : An update. *Medicinal Chemistry*, 10(4), 315–331.
- Valavanidis, A., Vlachogianni, T., Fiotakis, K., & Loridas, S. (2013). Pulmonary oxidative stress, inflammation and cancer: respirable particulate matter, fibrous dusts and ozone as major causes of lung carcinogenesis through reactive oxygen species mechanisms. *International Journal of Environmental Research and Public Health*, 10(9), 3886–3907.
- Wall, M. M. (2006). Ascorbic acid, vitamin A, and mineral composition of banana (Musa sp.) and papaya (*Carica papaya*) cultivars grown in Hawaii. Journal of Food Composition and Analysis, 19(5), 434–445.
- Wan Nazaimoon, W. M., Md Isa, S. H., Wan Mohamad, W. B., Khir, a S., Kamaruddin, N. a, Kamarul, I. M., ... Khalid, B. a K. (2013). Prevalence of diabetes in Malaysia and usefulness of HbA1c as a diagnostic criterion. *Diabetic Medicine*, 30(7), 825–828.
- Wang, H., Cao, G., & Prior, R. L. (1996). Total antioxidant capacity of fruits. Journal of Agricultural and Food Chemistry, 44(3), 701–705.
- Whitcomb, D. C., & Lowe, M. E. (2007). Human pancreatic digestive enzymes. *Digestive Diseases and Sciences*, 52(1), 1–17.
- Wickramarachchi, K. S., & Ranamukhaarachchi, S. L. (2005). Preservation of fiber rich banana blossom as a dehydrated vegetable. *Science Asia*, 31, 265–271.
- Wild, S., Roglic, G., Sicree, R., & Green, A. (2001). Global burden of diabetes mellitus in the year 2000. *Global Burden of Disease* 2000, 1–28.
- Yu, R., Mandlekar, S., & Kong, A. T. (2000). Molecular mechanisms of butylated hydroxylanisole-induced toxicity : induction of apoptosis through direct release of cytochrome c. *Molecular Pharmacology*, 6(3), 431–437.
- Yuris, A., & Siow, L.-F. (2014). A comparative study of the antioxidant properties of three pineapple (*Ananas comosus* L.) varieties. *Journal of Food Studies*, 3(1), 40–56.
- Zainol, M. ., Abd-Hamid, A., Yusof, S., & Muse, R. (2003). Antioxidative activity and total phenolic compounds of leaf, root and petiole of four accessions of *Centella asiatica* (L.) Urban. *Food Chemistry*, 81(4), 575–581.