

# **UNIVERSITI PUTRA MALAYSIA**

# APPLICATION OF ONE FACTOR AT A TIME (OFAT) AND RESPONSE SURFACE METHODOLOGY (RSM) FOR OPTIMIZING PHENOL-DEGRADING PARAMETER BY ISOLATE 1

SEHA ANAK MAMAT

FBSB 2015 76

APPLICATION OF ONE FACTOR AT A TIME (OFAT) AND RESPONSE SURFACE METHODOLOGY (RSM) FOR OPTIMIZING PHENOL-DEGRADING PARAMETER BY ISOLATE 1



SEHA ANAK MAMAT 163910

Dissertation submitted in partial fulfilment for the requirement for the course of BCH 4999 Project in the Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia.

June 2015

### PENGESAHAN

Dengan ini adalah disahkan bahawa projek yang bertajuk "Application of One Factor at a Time (OFAT) and Response Surface Methodology (RSM) for Optimising Phenol-degrading Parameters by Isolate 1" telah disiapkan serta dikemukakan kepada Jabatan Biokimia oleh Seha anak Mamat (163910) sebagai syarat kursus BCH 4999 projek.

Disahkan oleh,



Tarikh : .....

(Dr. Siti Aqlima Ahmad) Penyelia projek Jabatan Biokimia Fakulti Bioteknologi dan Sains Biomolekul Universiti Putra Malaysia

Tarikh : .....

(Prof. Dato' Dr. Abu Bakar Salleh) Ketua Jabatan Biokimia Fakulti Bioteknologi dan Sains Biomolekul Universiti Putra Malaysia

#### ABSTRACT

Global warming has always been a great concern especially in Antarctica due to pollution. Phenol pollution is one of the well-known pollution that highly affects the ecosystem in Antarctic soil and water. Due to it being widely used in various industries, excessive phenol is discharged into rivers and soil which might affect living things. The purpose of this study is to optimise the conditions for phenoldegrading bacteria Isolate 1 to degrade phenol at low temperatures using one factor at a time (OFAT) approach and response surface methodology (RSM). Four major parameters that can affect phenol degradation are pH, nitrogen source, temperature and salinity. From OFAT approach, Isolate 1 was found to have a high capability to degrade phenol with an optimum pH of 7.5 in phosphate buffer, ammonium sulphate as its best nitrogen source with concentration of 0.40 g/L, temperature of 15°C and salinity of 0.10 g/L. Meanwhile, during optimisation using RSM, Isolate 1 shows the highest phenol degradation percentage through interaction between 2 factors which are salinity with optimum sodium chloride (NaCl) concentration of 0.15 g/L and pH of 7.65. The greatest achievement in RSM is that Isolate 1 which is classified as cold-tolerant bacteria can degrade 0.50 g/L phenol up to 98.32% within two days.

#### ABSTRAK

Pemanasan global sentiasa menjadi kebimbangan terutamanya di Antartika disebabkan oleh pencemaran. Pencemaran fenol adalah salah satu pencemaran yang terkenal yang sangat memberi kesan kepada ekosistem tanah dan air Antartik. Oleh kerana ia digunakan secara meluas dalam pelbagai industri, lebihan fenol akan dilepaskan ke sungai dan tanah yg mungkin memberi kesan kepada hidupan. Tujuan kajian ini adalah untuk mengoptimumkan keadaan untuk bakteria pengurai fenol Isolat 1 untuk mengurai fenol pada suhu rendah menggunakan metodologi pendekatan satu faktor pada satu masa (OFAT) dan tindak balas permukaan (RSM). Empat pemboleh ubah utama yg boleh memberi kesan degradasi fenol adalah pH, sumber nitrogen, suhu dan kemasinan. Dari pendekatan OFAT, Isolat 1 mempunyai keupayaan untuk menguraikan fenol dengan optimum pH 7.5 dalam penimbal fosfat, ammonium sulfat sebagai sumber nitrogen yang terbaik dengan kepekatan 0.40 g/L, suhu 15 darjah celsius dan kemasinan 0.1 g/L. Sementara itu, semasa pengoptimuman menggunakan RSM, Isolat 1 menunjukkan peratusan kemerosotan tertinggi fenol melalui interaksi antara 2 faktor iaitu kemasinan dengan optimum kepekatan natrium klorida (NaCl) 0.15 g/L dan pH 7.65. Pencapaian terbesar dalam RSM ialah Isolate 1 yang mengelaskan bakteria bertoleransi sejuk boleh degradasi 0.50 g/L fenol sehingga 98.32% dalam masa dua hari.

#### ACKNOWLEDGEMENT

First of all, I would like to express my appreciation towards my supervisor, Dr Siti Aqlima Ahmad, who gave the permission for me to do this project and also for her endless support, comments and guidance through the process of completing my thesis project.

I would also want to thank my teammates especially Tengku Nur Saleha, Syirhan and Syahir for their consistent help in the experiment, opinion and knowledge with my thesis and also for being very supportive during the completion of my thesis project. Next, I would like to also give my thanks to my seniors in lab 204 for their guidance and always prepare for my every need in thesis work.

Besides, I also want to thanks my academic supervisor, Prof. Mohd Arif Syed and other supervisor, Dr Adeela binti Yasid for their comments and guidance given in my thesis progress presentation.

Lastly, I would like to thanks my family, especially my parents, Mamat anak Angkah and my mother, Maliah anak Jemat for their moral and financial support throughout this project. Not to forget all those who contributed to my thesis directly or indirectly, I want to give my thanks for your support and advices. I am very grateful for everyone who concern about my thesis project.

Seha anak Mamat, 2015

## TABLE OF CONTENTS

PENGESAHAN	i
ABSTRACT	ii
ABSTRAK	iii
ACKNOWLEDGEMENTS	iv
TABLE OF CONTENTS	v
LIST OF TABLES	vii
LIST OF FIGURES	viii
LIST OF ABBREVIATIONS	ix
CHAPTER	

1.0	INT	RODUCTION	1
2.0	LIT	ERATURE REVIEW	3
	2.1	Antarctic pollution	3
	2.2	Phenol	4
		2.2.1 Characteristics of phenol	4
		2.2.2 Application of phenol in industries	4
		2.2.3 Toxicity of phenol	5
	2.3	Biodegradation of phenol	5
		2.3.1 Metabolic pathway for phenol degradation	5
		2.3.2 Factors affecting in phenol degradation	6
		2.3.2.1 pH	6
		2.3.2.2 Temperature	7
		2.3.2.3 Substrate concentration	7
		2.3.2.4 Concentration of oxygen	7
		2.3.2.5 The presence of contaminant	7
		2.3.2.6 Cell immobilisation	8
	2.4	Theoretical aspects of phenol-degrading bacteria,	8
		Acinetobacter sp.	
	2.5	Response surface methodology (RSM)	8
3.0	MA	FERIALS AND METHODS	10
	3.1	Chemicals and equipments	10
	011	3.1.1 Chemical	10
		3.1.2 Apparatus	11
	3.2	Microorganism	11
	3.3	Growth and media condition	11
		3.3.1 Nutrient broth	11
		3.3.2 Minimal salt medium (MSM) broth	12
		3.3.3 Nutrient agar	12
		3.3.4 MSM agar	12
	3.4	Preparation of bacterial stock	13
		3.4.1 Glycerol stock	13
		3.4.2 Nutrient broth	13
	3.5	Reagent preparation for phenol assay	13

		3.5.1 4-aminoantipyrene (4-AAP)	13
		3.5.2 Ammonium chloride solution	13
		3.5.3 Potassium ferric cyanide	14
	3.6	Phenol assay	14
	3.7	Bacteria growth	14
	3.8	Identification of phenol-degrading bacteria	14
		3.8.1 Morphology of phenol-degrading bacteria Isolate 1	14
		3.8.2 Gram staining	15
		3.8.3 Catalase test	15
		3.8.4 Oxidase test	15
	3.9	Optimisation of bacteria growth and phenol degradation	15
		using OFAT	
		3.9.1 pH	16
		3.9.2 Nitrogen sources	16
		3.9.3 Concentration of selected nitrogen source	16
		3.9.4 Temperature	17
		3.9.5 Salinity	17
		3.9.6 Phenol concentration	17
	3.10	Response surface methodology (RSM)	17
		3.10.1 Plackett-Burman design	17
		3.10.2 Central composite design (CCD)	19
			-
4.0	RESU	JLTS AND DISCUSSIONS	21
	4.1	Identification of phenol-degrading bacteria	21
		4.1.1 Morphological characterisation of Isolate 1 using	23
		minimal salt medium (MSM) agar	24
		4.1.2 Gram staining	25
		4.1.3 Biochemical test	25
	4.2	Optimisation condition of Isolate 1 using OFAT	27
		4.2.1 Effect of pH	28
		4.2.2 Effect of temperature	30
		4.2.3 Effect of nitrogen sources	31
		4.2.4 Effect of ammonium sulphate	32
		4.2.5 Effect of salinity	
		4.2.6 Bacteria growth and phenol degradation with	34
		initial concentration of phenol	34
	4.3	Response surface methodology (RSM)	36
		4.3.1 Plackett-Burman design	
		4.3.2 Central composite design (CCD)	41
5.0	CON	CLUSION	42
			45
REF	ERENC	CES	
	ENDIC		

APPENDIC

# LIST OF TABLES

Table		Page
1	List of chemicals.	10
2	List of apparatus.	11
3	Range value of parameters at low and high level of Plackett- Burman design for degradation of phenol by Isolate 1.	18
4	Experiments on phenol degradation by Plackett-Burman design.	18
5	Range value of parameter at low and high level of CCD design for phenol degradation by Isolate 1.	19
6	Experiments on phenol degradation by Isolate 1.	20
7	Experimental design and result of phenol degradation by Plackett-Burman design.	35
8	Analysis of variance (ANOVA) for phenol degradation by Plackett-Burman design.	35
9	Experimental design and result of optimisation of phenol- degrading bacteria (Isolate 1) using CCD.	36
10	Analysis of variance (ANOVA) for CCD.	37

## LIST OF FIGURES

Figure		Page
1	Chemical structure of phenol.	4
2	Flow chart of ortho- and meta- cleavage of phenol.	6
3	Isolate 1 on phenol agar.	22
4	Gram-negative Isolate 1 observed under light microscope using oil immersion (100X).	23
5	Biochemical tests.	25
6	Effect of various pH on bacteria growth and phenol degradation by Isolate 1.	27
7	Effect of temperature on bacteria growth and phenol degradation by Isolate 1.	28
8	Effect of nitrogen sources on bacteria growth and phenol degradation by Isolate 1.	29
9	Effect of ammonium sulphate on bacteria growth and phenol degradation by Isolate 1.	30
10	Effect of salinity on bacteria growth and phenol degradation by Isolate 1.	32
11	Graphs of bacteria growth and phenol degradation with differences initial phenol concentration.	33
12	Contour plot shows effect of pH and salinity on the phenol degradation by Isolate 1.	38
13	3D response surface model for the phenol degradation shows interaction between parameter pH and salinity.	39

### LIST OF ABBREVIATIONS

%  $(NH_4)_2SO_4$ μg μl 4-AAP  $cm^3$ et al., FeSO<sub>4</sub>.H<sub>2</sub>O G  $H_2O$  $H_2O_2$ HCl hrs  $K_2Fe(CN)_6$ K<sub>2</sub>HPO<sub>4</sub> KH<sub>2</sub>PO<sub>4</sub> L mg MgSO<sub>4</sub> min ml mM MnSO<sub>4</sub>.H<sub>2</sub>O mol MSM NaCl NaMoO<sub>4</sub>.2H<sub>2</sub>O NaOH NH<sub>4</sub>Cl nm  $O_2$ °C rpm β

Percent Ammonium sulphate Microgram Microlitre 4-amino antipyrene Centimetre cubes And friends Ferrous sulphate monohydrate Gram Water Hydrogen peroxide Hydrochloric acid Hours Potassium ferric cyanide Di-potassium hydrogen phosphate Potassium dihydrogen phosphate Litre Miligram Magnesium sulphate Minutes Millilitre Milimolar Manganese sulphate Moles Minimal salt medium Sodium chloride Sodium molybdate dehydrate Sodium hydroxide Ammonium chloride Nanometer Oxygen Degree of celcius Revolution per minute Beta

# CHAPTER 1 INTRODUCTION

Antarctic island is one of the last pristine regions of planet (Vodopivez *et al.*, 2015) which facing pollutions issues. In cold climate such as Antarctica, location where there are human habitats or any traces of human activity are mostly increases in chances of soil and water contamination (Litova *et al.*, 2014). Biodegradation and bioremediation by using bacteria are most focusing study in Antarctica during early hydrocarbon pollution (Polmear *et al.*, 2015) due to it safety and harmless rather than chemical clean-up procedure (Luz *et al.*, 2006).

Phenol is an organic compound that is being widely used in agricultural chemicals, pesticides, petrochemicals, pharmaceuticals, textiles and steel industries (Basha *et al.*, 2010; Mahiudddin *et al.*, 2012). The presence of phenol wastes from sewage and industrial discharge have become a great concern because of their toxicity and persistent in environment (Bui *et al.*, 2012). Industrialisation country especially Malaysia is having problems with the unsafe levels of phenol as it can affects the communities' health (Ahmad *et al.*, 2011). Even at low concentration, continuous exposure towards phenol can lead to serious damage on our urinary system especially kidneys and central nervous system with only via inhalation, direct contact or ingestion (Suhaila *et al.*, 2013).

To overcome this situation, a numbers of studies have demonstrated the method of treating phenol wastewater biologically without causing harmful effects (Chen *et al.*, 2002). Several measurements and factors are crucial in the biodegradation of phenol includes temperature, pH, incubation periods, carbon and nitrogen sources (Suhaila *et al.*, 2013). Mesophilic bacteria is usually used for phenol biodegradation in tropical country such as Malaysia in which its temperature range is between 20-37 °C for optimum degrading activity (Ahmad *et al.*, 2011). Thermophilic bacteria usually have the optimum temperature to degrade phenol between 60-65 °C (Margesin *et al.*, 2005). In cold climates, it is important for cold-adapted microorganisms have ability to degrade organic

contaminants under cold condition (Margesin *et al.*, 2005). According to Bergauer *et al.*, (2005), psychrophiles or cold-tolerant phenol degraders are able to degrade phenol at low temperature due to their adaptation to cold environment.

At high concentration of phenol wastewater, bacteria will reach optimal degradation capability by using response surface methodology (RSM). RSM is a statistical and mathematical technique that is used to achieve the optimal response for phenol-degrading bacteria to degrade phenol (Huang *et al.*, 2013). Central composite design is one of the examples of experimental design that used in RSM (Suhaila *et al.*, 2013).

Objectives of this research:

- 1. To identify phenol-degrading bacteria, Isolate 1.
- 2. To characterise the potential of phenol-degrading bacteria using different optimisation methods: one factor at a time approach (OFAT) and response surface methodology (RSM).

#### REFERENCES

- Abd-El-Haleem, D., Beshay, U., Abdelhamid, A.O., Mowad, H. and Zaki, S. 2003. Effects of mixed nitrogen sources on biodegradation of phenol by immobilized Acinetobacter sp. strain W-17. African Journal of Biotechnology, 2: 8-12.
- Ahmad, S.A., Shamaan, N.A., Arif. N.M., Koon, G.B., Shukor, M.Y.A and Syed, M.A. 2012. Enhanced phenol degradation by immobilized *Acinetobacter* sp. Strain AQ5NOL 1. World Journal Microbiology and Biotechnology, 28: 347-352.
- Ahmad, S.A., Syed, M.A., Arif, N.M., Shukor, M.Y.A. and Shamaan, N.A. 2011. Isolation, identification and characterization of elevated phenol degrading *Acinetobacter* sp. strain AQ5NOL 1. *Autralian Journal of Basic and Applied Sciences*, 5: 1035-1045.
- Al-Khalid, T. and El-Naas. 2012. Aerobic biodegradation of phenols: A comprehensive review. *Critical Review in Environmental Science and Technology*, 42: 1631-1690.
- Anwar, F., Hussain, S., Ramzan, S., Hafeez, F., Arshad, M., Imran, M., Maqbool, Z. and Abbas, N. 2014. Characterization of reactive red-120 decolorizing bacterial strain Acinetobacter junii FA10 capable of simultaneous removal of azo dyes and hexavalent chromium. Water Air & Soil Pollution, 225: 2017.
- Arif, N.M., Ahmad, S.A., Syed, M.A. and Shukor, M.Y. 2012. Isolation and characterization of phenol-degrading *Rhodococcus* sp. Strain AQ5NOL 2 KCTC 11961BP. *Journal of Microbiology*, 52: 1-10.
- Basha, K.M., Rajendran, A. and Thangavelu, V. 2010. Recent advances in the biodegradation of phenol: A review. Asian Journal of Experimental Biological Science, 1: 219-234.
- Bergauer, P., Fonteyne, P.A., Nolard, N., Schinner, F. and Margesin, R. 2005. Biodegradation of phenol-related compound by psychrophilic and coldtolerant alphine yeast. *Chemosphere*, 59: 909-918.
- Bergauer, P., Fontyne, P.A., Nolard, N., Schinner, F. and Margesin, R. 2005. Biodegradation of phenol and phenol-related compounds by psychrophilic and cold-tolerant alpine yeast. *Chemosphere*, 59: 909-918.
- Bezerra, M.A., Santelli, R.E., Oliveira, E.P., Villar, L.S. and Escaleira, L.A. 2008. Review Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76: 965-977.
- Bharti, P.K and Niyogi, U.K. 2015. Research article: Assessment of pollution in a freshwater lake at Fisher Island, Larsemann Hills over east Antarctica. *Science International*, 3: 25-30.

- Bui, H.B., Nguyen, L.T and Dang, L.D. 2012. Biodegradation of phenol by native bacteria isolated from dioxin contaminated soils. *Journal of Bioremediation and Biodegradation*, 3: 168.
- Chen, K.C., Lin, Y.H., Chen, W.H. and Liu, Y.C. 2012. Biodegradation of phenol by PAA-immobilised *Candida tropicalis*. *Enzyme and Technology*, 31: 490-497.
- Dahalan, F.A., Johari, W.L.W., Shukor, M.Y., Halmi, M.I.E., Shamaan, N.A. and Syed, M.A. 2013. Growth kinetics of a diesel-degrading bacteria strain from petroleum-contaminated soil. *Journal of Environmental Biology*, 35: 399-406.
- Gami, A.A., Shukor, M.Y., Khalil, K.A., Dahalan, F.A., Khalid, A. and Ahmad, S.A. 2014. Phenol and its toxicity. *Journal of Environmental and Toxicology*, 2: 11-24.
- Huang, L., Xie, J., Lv, B.Y., Shi, X.F., Li, G.Q., Liang, F.L. and Lian, J.Y. 2013. Optimization of nutrient component for diesel oil degradation by *Acinetobacter beijerinckiee* ZRS. *Marine Pollution Bulletin*, 76: 325-332.
- Kiliç, N.K. 2009. Enhancement of phenol biodegradation by Ochrobactrum sp. Isolated from industrial wastewaters. *International Biodeterioration and Biodegradation*, 63: 778-781.
- Krastanov, A., Alexieva, Z., and Yemendzhiev, H. 2013. Review: Microbial degradation of phenol and phenolic derivatives. *Engineering in Life Sciences*, 13: 76-87.
- Litova, K., Gerginova, M., Peneva, N., Manasiev, J. and Alexiera, Z. 2014. Research article: Growth of Antarctic fungal strain on phenol at low temperatures. *Journal of BioScience and Biotechnology*, SE/ONLINE: 43-46.
- Liu, Y.J., Zhang, A.N. and Wang, X.C. 2009. Biodegradation of phenol by using free and immobilized cells of *Acinetobacter* sp. XA05 and *Sphingomonas* sp. FG03. *Biochemical Engineering Journal*, 44: 187-192.
- Luz, A.P., Ciapina, E.M.P., Gamba, R.C., Lauretto, M.S., Farias, E.W.C., Bicego, M.C., Taniguchi, S., Montone, R.C. and Pellizari, V.H 2006. Potential for biodegradation of hydrocarbon polluted soils in the maritime Antarctic. *Antarctic Science*, 18: 335-343.
- Mahiuddin, Md., Fakhruddin, A.N.M. and Abdullah-Al-Mahin. 2012. Research article: Biodegradation of phenol via meta cleavage pathway by Pseudomonas fluorescens PU1. *International Scholarly Research Network*, Article ID: 741820.

- Margesin, R., Fonteyne, P.A. and Redl, B. 2005. Low temperature biodegradation of high amounts of phenol by *Rhodococcus* spp. and basidiomycetous yeasts. *Research in Microbiology*, 156: 68-75.
- Margesin, R., Moertelmaier, C. and Mair, J. 2013. Low temperatures biodegradation of petroleum hydrocarbons (n-alkene, phenol, anthracene, pyrene) by four actinobacterial strains. *International Biodeterioration and Biotechnology*, 84: 185-191.
- Nair, C.I., Jayachandran, K. and Shashidhar, S. 2008. Review: Biodegradation of phenol. *African Journal of Biotechnology*, 7: 4951-4958.
- Nor-Suhaila, Y., Ariff, A., Rosfarizan, M., Abdul Latif, I., Ahmad, S.A., Norazah, M.N. and Shukor, M.Y.A. 2010. Optimization parameters for phenol degradation by *Rhodococcus* UKM-P in shake flask culture. *Proceeding of The World Congress on Engineering*, Volume 1.
- Pandimadevi, M., Venkatesh, P.M. and Vinod, K.V. 2014. Optimization of phenol degradation using *Pseudomonas aeruginosa* (MTCC 7814) by Plackett-Burman design and response surface methodology. *Journal of Bioremediation and Biodegradation*, 5:7.
- Pazarlioglu, N.K. and Telefoncu, A. 2005. Biodegradation of phenol by *Pseudomonas putida* immobilized on activated pumice particles. *Process Biochemistry*, 40: 1807-1814.
- Pham, V.H.T., Kim, J. and Jeong, S.W. 2014. Enhanced isolation and culture of highly efficient psychrophilic oil-degrading bacteria from oil-contaminated soils in South Korea. *Journal of Environmental Biology*, 35: 1145-1149.
- Polmear, R., Stark, J.S., Roberts, D. and McMinn, A. 2015. The effects of oil pollution on Antarctic benthic diatom communities over 5 years. *Marine Pollution Bulletin*, 90: 33-40.
- Schie, P.M.V. and Young, L.Y. 2007. Biodegradation of phenol: Mechanism and application. *Bioremediation Journal*, 4:1-18.
- Shukor, M.Y., Jusoh, A.Z., Perumal, N. Shamaan, N.A., MacCormack, W.P. and Syed, M.A. 2009. Isolation and characterization of *Pseudomonas* dieseldegrading strain from Antartica. *Journal of Environmental Biology*, 30:1-6.
- Sridevi, V., Lakshmi, M.V.V.C., Manasa, M. and Sravani, M. 2012. Metabolic pathways for the biodegradation of phenol. *International Journal of Engineering Science and Advanced Technology*, 2: 695-705.
- Suhaila, Y.N., Ramanan, R.N., Rosfarizan, M., Latif, I.A. and Ariff, A. 2013. Optimization of parameters for improvement of phenol degradation by *Rhodococcus* UKMP-5M using response surface methodology. *Annals of Microbiology*, 63: 513-521.

- Suhaila, Y.N., Rosfarizan, M., Ahmad, S.A., Latif, I.A.and Ariff, A.B. 2013. Nutrient and culture condition requirements for the degradation of phenol by *Rhodococcus* UKMP-5M. *Journal of Environmental Biology*, 34: 635-643.
- Tin, T., Fleming, Z.L., Hughes, K.A., Anley, D.G., Convey, P., Moreno, C.A., Pfeiffer, S., Scott, J and Snape, L. 2009. Review: Impacts of local human activities on the Antarctic environment. *Antarctic Science*, 21: 3-33.
- Tortora, G.J., Funke, B.R. and Case, C.L. 2013. Microbiology: An Introduction, 11<sup>th</sup> Edition. United States, America, pp 60-161.
- Vodopivez, C., Curtosi, A., Villaamil, E., Smichowski, P., Pelletier, E. and Cormack, W.P.M. 2015. Heavy metal in sediments and soft tissues of the Antarctic clam *Laternula elliptica*: More evidence as a ? possible biomonitor of coastal marine pollution at high latitudes? *Science of the Total Environment*, 502: 375-384.
- Yamaga, F., Washio, K. and Morikawa, M. 2010. Sustainable biodegradation of phenol by Acinetobacter calcoaceticus P23 isolated from the rhizophere of duckweed Lemna aoukikusa. Environmental Science and Technology, 44: 6470-6474.
  - Ying, W., Ye, T., Bin, H., Hua-bing, Z. 2007. Biodegradation of phenol by free and immobilized *Acinetobacter* sp. strain PD12. *Journal of Environmental Sciences*, 19: 222-225.