

UNIVERSITI PUTRA MALAYSIA

HPLC SUGAR PROFILES OF MALAYSIAN HONEYS

NURUL HUSNA QAIYIMAH IBRAHIM

FBSB 2015 71

HPLC SUGAR PROFILES OF MALAYSIAN HONEYS



NURUL HUSNA QAIYIMAH BINTI IBRAHIM

162512

DEPARTMENT OF BIOCHEMISTRY

FACULTY OF BIOTECHNOLOGY AND BIOMOLECULAR SCIENCES

(FBSB)

UNIVERSITI PUTRA MALAYSIA

2015

HPLC SUGAR PROFILES OF MALAYSIAN HONEYS



NURUL HUSNA QAIYIMAH BINTI IBRAHIM

162512

Thesis Submitted in Partial Fulfilment of the Requirement For the course BCM 4999 (Project) in the Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

June 2015

PENGESAHAN

Dengan ini disahkan bahawa laporan bertajuk profil gula daripada madu Malaysia telah disiapkan serta dikemukan kepada Jabatan Biokimia, Fakulti Bioteknologi dan Sains Biomolekul, Universiti Putra Malaysia oleh Nurul Husna Qaiyimah Binti Ibrahim (162512) sebagai syarat untuk kursus BCM 4999 (Projek).

Disahkan oleh:	
	Tarikh:
Dr. Amir Syahir Bin Amir Hamzah	
Penyelia Projek	
Jabatan Biokimia	
Fakulti Bioteknologi dan Sains Biomolekul	
Universiti Putra Malaysia	
	Tarikh:
Prof. Dato' Abu Bakar Salleh	
Ketua	
Jabatan Biokimia	
Fakulti Bioteknologi dan Sains Biomolekul	
Universiti Putra Malaysia	
i	

ACKNOWLEDGEMENT

الله بسم الرحمن الرحيم

In the name of Allah, who is Beneficent and Merciful

First of all, I would like to give my deepest gratitude to my supervisor, Dr. Amir Syahir Bin Amir Hamzah for his supervision and suggestions that guide and help me to complete this project successfully. My warmest gratitude also to Dr. Aqlima Binti Ahmad for her permission that allows me to use Lab 204. I also would like to thank to Prof. Dr. Mohd Arif Syed for his assistance as the coordinator for this BCM 4999.

Furthermore, I would like to thank to all my members of Enzymology and Bioremediation Lab (115 and 204) especially to Mr. Khalizan for all knowledge and sharing of experiences that have been learned along with technical assistance throughout the study. A very special gratitude to my dearest labmates, for helping me in this project. I also would like to thank my classmates for all their help during lab works and writing thesis.

Finally, I would like to thank the authority of Faculty of Biotechnology and Biomolecular Sciences that provided a good environment and facilities. Not forgettable, I especially would like to thank my family members, especially my husband and my parents for their support and advices throughout the study. This research project will be impossible if all these people above were not lending their help to me. Thank you so much!

TABLE OF CONTENTS

	Page
PENGESAHAN	i
ACKNOWLEDGEMENTS	ii
TABLE OF CONTENTS	iii
LIST OF TABLES	v
LIST OF FIGURES	vi
LIST OF ABBREVIATIONS	vii
ABSTRACT	ix
ABSTRAK	Х
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	
2.1 Honey	4
2.1.1 Production of honey	4
2.1.2 Composition of honey	5
2.1.3 Uses and product of honey	7
2.1.4 Honey authenticity	7
2.2 Profiling	10
2.2.1 Carbohydrates analysis	10
2.2.2 High performance liquid chromatography (HPLC)	11
2.2.3 Column	11
2.2.4 Detector	12
2.3 Statistical analysis	12

3 MATERIALS AND METHOD

6

	3.1	Material	ls	14
		3.1.1	Honey samples	14
		3.1.2	Chemicals and standards	15
		3.1.3	Equipments	15
	3.2	Method	l	
		3.2.1	Sample preparation	16
		3.2.2	Sample analysis by HPLC-UV	16
		3.2.3	Statistical analysis	17
4	RESUI	LTS AND	DISCUSSION	
	4.1	Sugar pi	rofiling	18
	4.2	Statistic	al analysis of HPLC sugar analysis	
		4 <mark>.2.1</mark>	Hierarchical clustering analysis	25
		<mark>4.2.2</mark>	Principal components analysis	29
5	CONCI	LUS <mark>ION</mark>	AND RECOMMENDATION	32
REFERENCES 33				
APPENDICES 38				

LIST OF TABLES

TABLE		Page
Table 1:	The characteristic and composition of honey.	6
Table 2:	Composition criteria for honey according to Codex Standard.	9
Table 3:	List of honey samples according to their types and production statue.	14
Table 4:	Results of sugar values (mg/ml) in honey samples.	23-24
Table 5:	Factors of Principal component analysis (PCA).	31

C

LIST OF FIGURES

FIGURE		Page
Figure 1:	Structures of some common sugars found in the nectar and honey.	4
Figure 2:	HPLC-UV chromatographic profiles of 5 standard sugars.	18
Figure 3:	Average distribution of 5 sugars in honey sampels.	19
Figure 4:	Average distribution of 5 sugars in honey samples based on type of honeys.	21
Figure 5:	Hierarchical clustering analysis of sugars honeys samples obtained by Euclidean distances.	27
Figure 6:	Hierarchical clustering analysis of sugars honeys samples obtained by Cosine distances.	28
Figure 7:	Distribution of honey samples on score plot of 5 data sugars.	30
Figure 8:	Distribution of honey samples on score plot based 3 sugars.	30

C

LIST OF ABBREVIATIONS

HPLC	High Performance Liquid Chromatoragphy
RID	Refractive Index Detector
GC	Gas Chromatography
MS	Mass Spectroscopy
HPAEC	High-Performance Anion-Exchange Chromatography
PAD	Pulsed Amperometric Detection
NMR	Nuclear Magnetic Resonance
UV	Ultraviolet
RI	Refractive Index
ELSD	Evaporating Light Scattering Detector
CAD	Charge Aerosol Detector
НСА	Hierachical Clustering Analysis
PCA	Principal Components Analysis
NIR	Near-Infrared Spectroscopy
PC	Principal Components
F/G	Fructose to Glucose Ratio
C8	Carbon-8
C18	Carbon-18
NH ₂	Amino group
HMF	Hydroxymethylfurfural
%	Percentage
mg	Milligram
ml	Millilitre

6

nm	Nanometer
°C	Degree Celcius
Cal	Calorie Unit
ms	Millisecond
cm	Centimeter
g	Gram
v/v	Volume per volume
mm	Millimeter
μl	Microliter
mAU.s	Milliabsorbance unit per second
μm	Micrometer
et al.,	and friends

C

ABSTRACT

Sugar profiles of 34 honey samples from different regions of Malaysia which coming from Perlis, Kedah, Kelantan, Perak, Terengganu, Kuala Lumpur, Selangor, Pahang, Melaka, Negeri Sembilan, Johor, Sabah and Sarawak are analysed by HPLC with UV detector at 195 nm. These samples consist of 10 Kelulut honeys, 2 Acacia honeys, 3 Hutan honeys, 9 Tualang honeys and 10 unknown honeys (unknown types). Five sugars are identified which are three monosaccharides and two dissacharides. Fructose (32.66±1.19%) is the major sugar in all samples followed by glucose (31.57±1.20%), galactose (2.62±1.94%), maltose (2.06±0.35%) and sucrose (2.95±0.39%). Fructose and glucose are the main monosaccharides in all samples. Galactose, sucrose and maltose are only present in certain honey samples. Hierarchical clustering analysis (HCA) and principal component analysis (PCA) used to classify honeys correctly based on sugar composition. Euclidean distance shows the concentration of each sugars presence in the samples which differentiate low and high concentration of total sugar. Cosine distance study recognized the pattern of sugar in honey samples from different region of Malaysia. Based on principal component analysis (PCA), honey samples are scattered according to their PC1 and PC2, thus allows the authenticity prediction.

ABSTRAK

Profil gula 34 sampel madu dari kawasan-kawasan yang berlainan di Malaysia yang terdiri dari Perlis, Kedah, Kelantan, Perak, Terengganu, Kuala Lumpur, Selangor, Pahang, Melaka, Negeri Sembilan, Johor, Sabah dan Sarawak dianalisis oleh HPLC dengan menggunakan pengesan UV pada 195 nm. Sampel ini terdiri daripada 10 madu Kelulut, 2 madu Acacia, 3 madu Hutan, 9 madu Tualang dan 10 madu Unknown (jenis yang tidak diketahui). Lima gula dikenal pasti iaitu tiga monosakarida dan dua disakarida. Fruktosa ($32.66 \pm 1.19\%$) adalah gula utama dalam semua sampel diikuti dengan glukosa (31.57 ± 1.20%), galaktosa (2.62 ± 1.94%), maltosa ($2.06 \pm 0.35\%$) dan sukrosa ($2.95 \pm 0.39\%$). Fruktosa dan glukosa adalah monosakarida utama dalam semua sampel. Galaktosa, sukrosa dan maltosa hanya hadir dalam sampel madu tertentu. Analisis hierarki kelompok (HCA) dan analisis komponen utama (PCA) yang digunakan untuk mengkelaskan madu dengan betul berdasarkan komposisi gula. Jarak Euclidean menunjukkan kepekatan setiap kehadiran gula dalam sampel yang membezakan kepekatan rendah dan tinggi daripada jumlah kesuluruhan gula. Kajian jarak Cosine mengenalpasti corak gula dalam sampel madu dari kawasan yang berlainan di Malaysia. Berdasarkan analisis komponen utama (PCA), sampel madu bertaburan mengikut data PC1 dan PC2, sekali gus membolehkan ramalan keaslian.

CHAPTER 1

INTRODUCTION

According to Codex Standard for honey, honey is defined as the natural sweet substance produced by honey bees from the nectar of plants or from secretions of living parts of plants or excretions of plant sucking insects on the living parts of plants, which the bees collect, transform by combining with specific substances of their own, deposit, dehydrate, store and leave in the honeycomb to ripen and mature (Codex Standard., 2011).

Honey mainly composes of 80-85% of carbohydrates and other minor substances such as protein and amino acids, minerals, vitamins, enzymes and organic acids (Arvanitoyannis *et al.*, 2005). Sugars in honey are obtained from several reactions of enzymes on the nectar sources. The enzymes involved are: invertase, diastase and glucose oxidase (Crane, 1980). The main enzyme responsible to sugar content in honey is invertase which converts sucrose to subunits fructose and glucose by breaking their glycosidic bonds.

Malaysia is a country that rich in biodiversity. There are several type of honey found in Malaysia which are Tualang, Kelulut, Hutan, Acacia and so on which highly influenced by type of honey bees. Honey bees come from *Apidae* with single genus, *Apis* comprises four species: *Apis dorsata* (Tualang), *Apis florea, Apis mellifera* (Acacia) and *Apis cerana* (Yong and Othman, 2007). One major problem faces by the Malaysian public is the question of honey purity and quality. The adulteration of honey slightly increased in honey products by adding the natural carbohydrates content of honey with cheaper or artificial sweeteners to gain more profit. Many adulterants such as starch and dextrin (Wang *et al.*, 2015), isoglucose syrups and acid-inverted syrups (Crane, 1980), corn syrups (Ribeiro *et al.*, 2014), and beet sugars (Canabero *et al.*, 2006) are reported in past research.

Therefore, the methods of quality control to identify the authenticity of honey are required. Several methods have been developed to detect adulteration of honey. Many analytical methods for sugar analysis have been reported: high-performance liquid chromatography coupled with refractive index detector (HPLC-RID) (Can *et al.*, 2015) or gas chromatography-mass spectroscopy (GC-MS) (Pasini *et al.*, 2013), high-performance anion-exchange chromatography with pulsed amperometric detection (HPAEC-PAD) (Ouchemoukh *et al.*, 2010) and nuclear magnetic resonance (NMR) (Consonni *et al.*, 2013).

Sugar profiling of different types of honey has been reported by many scientists to detect adulteration of honey (Arvanitoyannis *et al.*, 2005). The profiling showed that total sugar content in honey samples generally composed of monosaccharides, disaccharides and oligosaccharides (Fuente *et al.*, 2011). As stated in Codex Standard, the sum of both fructose and glucose not less than 60/100 g and sucrose not more than 5/100 g has been used as a standard to measure sugar composition. The sugar profiles were used determined in multifloral and unifloral honeys from Algeria (Ouchemoukh *et al.*, 2010).

In this study, 34 honey samples were analysed by using high-performance liquid chromatography with ultraviolet detector (HPLC-UV) at 195 nm. Sugars profiling are expected to be capable to detect the adulterated honey samples, detect sugar composition and identify pattern of various type of honeys. Multivariate statistical analysis is carried out to obtain graphic representation which shows best summary of the information. Hierarchical clustering analysis (Euclidean distance and Cosine distance) and principal component analysis are used to characterize the information of honey.

Thus, in this study, there are three major objectives that will be accomplished which are:

- 1. To identify different types of sugars in honey samples.
- 2. To analyse sugar pattern of Malaysia honey based on sugar composition.
- 3. To predict authenticity of honeys by using statistical methods and based on Codex Standard.

REFERENCES

- Abdel, M. E. S., Abdelhmied, B. A. and Salih, Z. A. 2013. Quality evaluation of honey obtained from different sources. *Food and Public Health 3*: 137 – 141.
- Arvanitoyanis, I. S., Chalhoub, C., Gotsiou, P., Simantiris, N. L. and Kefalas. P. 2005. Novel quality control methods in conjunction with chemometrics (Multivariate Analysis) for detecting honey authenticity. A critical reviews in Food Science and Nutrition 45: 193 203.
- Berthod, A., Chang, S. S. C., Kullman, J. P. S. and Armstrong, D. W. 1998. Practice and mechanism of HPLC oligosaccharide separation with a cyclodextrin bonded phase. *Talanta* 47: 1001 – 1012.
- Bhargava, H. R. and Mothilal, M. 2014. A comparative study on the chemical composition, chemical characterization and floral studies of raw and processed honey samples of *Apis* Species. *World Applied Science Journal 32*: 302 308.
- Bobis, O., Marghitas, L. A., Bonta, V., Dezmirean, D. and Maghear, O. 2007. Free phenolics acids, flavonoids, and abscisic acid related to HPLC sugar profile in Acacia honey. *Bullentin USAMV-CN*: 63 64.
- Canabero, A. I., Recio, J. L. and Ruperez, M. 2006. Liquid Chromatography Coupled to Isotope Ratio Mass Spectrometry: A new perspective on honey adulteration detection. *Journal of Agriculutural and Food Chemistry 54*: 9719–9727.

- Can, Z., Yildiz, O., Sahin, H., Turumtay, E. A., Silici, S. and Kolayli, S. 2015. An investigation of Turkish honeys: Their physic-chemical properties, antioxidant capacities and phenolic profiles. *Food Biochemistry 180*: 133 – 141.
- Cengriz, M. F., Durak, M. Z. and Ozturk, M. 2014. In- house validation for the determination of honey adulteration with plant sugars (C₄) by Isotope Ratio Mass Spectrometry (IR-MS). *Food Science and Technology* 57: 9 – 15.
- Chakir, A., Romane, A., Marcazzan, G. L. and Ferrazi, P. 2011. Physicochemical properties of some honeys produced from different plants in Morocco. *Arabian Journal of Chemistry*: 1 - 9.

Codex Standard, 2011.

Cosonni, R., Cagliani, L. R. and Cogliati, C. 2013. Geographical discrimination of honeys by saccharides analysis. *Food Control 32*: 543 – 548.

Crane, E., 1980. A book of honey. Oxford University Press 1980.

Fuente, E. D. L., Matute, A. I. R., Barrera, R. M. V., Sanz, J. and Castro, I. M. 2011.
Carbohydrate composition of Spanish unifloral honeys. *Food Chemistry* 129: 1483 – 1489.

Guler, A., Kocaokutgen, H., Garipoglu, A. V., Onder, H., Ekinci, D. and Biyik, S. 2014. Detection of adulterated honey produced by honeybee (*Apis Mellifera L.*) colonies fed with different level of commercial industrial sugar (C₃ and C₄ plants) syrups by carbon isotope ratio analysis. *Food Chemistry 155*: 155 – 160.

- Guo, W., Zhu, X., Liu, Y. and Zhuang, H. 2010. Sugar and water contents of honey with dielectric property sensing. *Journal of Food Engineering* 97: 275 – 281.
- Karimov, E., Xalilzad, Z., Hobbi, P. and Alekperov, J. 2014. Quality evaluation of honey from the different region of Azerbaijan. *Journal of Food Biochemistry and Nutrition* 2: 71 – 79.
- Karkacier, M., Erbas, M., Uslu, M. K. and Aksu, M. 2003. Comparison of different extraction and detection methods for sugar using amino-bonded phase HPLC. *Journal of Chromatographic Science* 4: 331 – 333.
- Kowalski, S. 2013. Changes of antioxidant activity and formation of 5hydroxymethylfurfural in honey during thermal and microwave processing. *Food Chemistry 141*: 1378 – 1382.
- Manzares, A. B., Garcia, H. G., Galdon. B. R., Rodriguez, E. R. and Romero, C. D.
 2011. Differentaition of blossom and honeydew honey using multivariate analysis on the physiochemical parameters and sugar composition. *Food Biochemistry* 126: 664 667.
- Manzares, A. B., Garcia, Z. H., Galdon, B. R., Rodriguez, E. R. and Romero, C.D.
 2014. Physicochemical characteristics of minor monofloral honeys from Tenerife, Spain. *Food Science and Technology* 55: 572 – 578.
- Niculina, M., Marghitas, L. A., Dezmirean, D., Bonta, V. and Bobis, O. 2012. Botanical origin authentication of Black Locust (*Robina Pseudoacacia*) honey, by means of sugar spectrum determination. *Agriculture 3-4*: 95 – 100.

- Ouchemoukh, S., Schweiter, P., Bey, M. B., Kadji, H. D. and Louaileche, H. 2010. HPLC sugar profiles of Algerian honeys. *Food Biochemistry 121*: 561 – 568.
- Pasini, F., Gardini, S., Marcazzan, G. L. and Caboni, M. F. 2013. Buckwheat honeys: Screening of composition and properties. *Food Chemistry* 141: 2802-2811.
- Poyrazoglu, E. S., Haroun, M. I., Konar, N., Hospolat, I. and Artik, N. 2012. Sugar profiles and contents of Turkish honeydew and floral honeys. *International Conference on Environment, Agriculture and Food Sciences (ICEAFS'2012):* 59 - 62.
- Rahman, N. A., Hasan, M., Hussain, M. A. and Jahim, J. 2008. Determination of glucose and fructose from glucose isomerization process by highperformance liquid chromatography with UV detection. *Modern Applied Science Vol.* 2: 151 – 154.
- Ribeiro, R. O. R., Marsico, E. T., Carneiro, C. S., Monteiro, M. L. G., Junior, C. C. and Jesus, E. F. O. 2014. Detection of honey adulteration of high fructose corn syrup by Low Nuclear Magnetic Resonance (LH ¹H NMR). *Journal of Food Engineering 135*: 39 – 43.

Salonen, A., Hiltunen, J. and Titto, R. J. 2011. Composition of unique unifloral honeys. *Journal of Apiproduct and Apimedical Science 3*: 128 – 136.

Thakkar, A., Ramni, K., Navneet, K., Ashutosh, U. and Suri, O. P. 2011. High performance liquid chromatography detectors. *International Research Journal of Pharmacy* 2: 1 – 7.

- Wang, S., Gou, Q., Wang, L., Lin, L., Shi, H., Cao, H. and Cao, B. 2015. Detection of honey adulteration with starch syrup by high performance liquid chromatography. *Food Chemistry* 172: 669 – 674.
- Yong, P. H. and Othman, M. S. H. 2007. Economical Values of Honey Bees, Peninsular Malaysia. Forestry Department Peninsular Malaysia (FDPM).
- Yucel, Y. and Sultanoglu, P. 2013. Characterization of honeys from Hatay Region by their physicochemical properties combined with chemometrics. *Food Bioscience 1*: 16-25.
- Zhu, X., Li, S., Zhang, Z., Li, G., Su, D. and Liu, F. 2010. Detection of adulterants such as sweeteners materials in honey using near-infrared spectroscopy and chemometrics. *Journal of Food Engineering 101*: 92 – 97.