

DEVELOPMENT OF HPLC-MS/MS METHOD FOR SIMULTANEOUS QUANTIFICATION OF VETERINARY ANTIBIOTICS AND HORMONES IN SOIL AND BIOSOLIDS

HO YU BIN

FPAS 2012 3

DEVELOPMENT OF HPLC-MS/MS METHOD FOR SIMULTANEOUS QUANTIFICATION OF VETERINARY ANTIBIOTICS AND HORMONES IN SOIL AND BIOSOLIDS



HO YU BIN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilments of the Requirements for the Degree of Doctor of Philosophy

July 2012

DEDICATION

This thesis is dedicated to my parents

Ho Chee Weng and Yeo Bee Chuen

who brought me into this world, taught me the best knowledge, guided and supported me along the way to successful.

To my brother Ho Yu En and my fiancé Thai Hoeng

who encouraged and supported me during my study.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

DEVELOPMENT OF HPLC-MS/MS METHOD FOR SIMULTANEOUS QUANTIFICATION OF VETERINARY ANTIBIOTICS AND HORMONES IN SOIL AND BIOSOLIDS

By

HO YU BIN

July 2012

Chair: Professor Mohamad Pauzi Zakaria, PhD Faculty: Faculty of Environmental Studies

Repeated applications of animal manure as fertilizer are normal agricultural practices that may release veterinary antibiotics and hormone into the environment from treated animals. The occurrence of veterinary antibiotics and hormone residues in the environment is of concern because of the development of antibiotic-resistant bacteria and its ecotoxicological behavior to plants and animals. To date, the occurrence of veterinary antibiotics and hormone in the Malaysian agricultural field has never been reported. The lack of data could be attributed to the absence of universal method for the analysis of veterinary pharmaceutical residues in complex solid environmental matrices. Therefore, a reliable, simple, fast and cost-effective method was developed in order to quantify nine antibiotics (doxycycline, enrofloxacin, erythromycin, flumequine, norfloxacin, sulfadiazine, tilmicosin, trimethoprim, tylosin) and one hormone (progesterone) in soil, broiler manure and manure compost. Generally, this method has shorter time of analysis, lower cost of solid phase extraction (SPE), better recoveries, lower method detection limit (MDL) and method quantification limit (MQL) values as compared to USEPA Method 1694, Kim and Carlson (2007) and Blackwell et al, (2004). The current method was compared to the USEPA Method 1694 where the method has the closest target analytes and sample matrices. The time of extraction has reduced from >125 mins to >63 mins, the time of liquid chromatography analysis has reduced from 61 mins to 25 min and the cost of SPE cartridge has reduced from RM70.00 to RM10.30 per sample when compared to the USEPA Method 1694. The developed method was based on ultrasonic extraction with MeOH:ACN:EDTA:McIlvaine buffer (pH 4) (30:20:25:25), SPE using HLB (3cc/60mg) cartridge, followed by instrumental analysis using liquid chromatography-tandem mass spectrometry (LC-MS/MS) with 25 mins total run time. The developed method was validated and tested on soil, broiler manure and manure compost samples and showed that the method is able to simultaneously detect and quantify the target analytes with good selectivity and sensitivity. The overall method performance was good for the majority of the analytes, with recoveries greater than 80% for most of the analytes in each type of sample matrix. MDL and MQL were achieved at as low as 0.5 and 2 μ g/kg dry weight (DW) for trimethoprim. Tilmicosin was first time reported in environmental matrices. The developed method was then applied on broiler manure samples and its relative manure amended agricultural soil samples collected in Selangor, Negeri Sembilan and Melaka to identify and quantify veterinary antibiotic and hormone residues in the environment. The broiler manure samples were found to be contaminated with at least seven target analytes, which are doxycyline, enrofloxacin, flumequine, norfloxacin, trimethoprim, tylosin and progesterone. The maximum concentration of antibiotic detected in broiler manure was 78516.1 µg/kg DW for doxycycline. For manure amended agricultural soil samples, doxycycline, enrofloxacin, flumequine and trimethoprim residues were detected in every soil samples. The maximum concentration of antibiotic detected in soil was 1331.4 µg/kg DW for flumequine. The results showed that the method can potentially be adopted for the analysis of veterinary antibiotic and hormone wastes in solid environmental matrices.

Keywords: Veterinary antibiotics, hormone, HPLC-MS/MS, soil, manure

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PEMBANGUNAN KAEDAH HPLC-MS/MS UNTUK KUANTIFIKASI SERENTAK ANTIBIOTIK DAN HORMON VETERINAR DALAM TANAH DAN BIO-PEPEJAL

Oleh

HO YU BIN

Julai 2012

Pengerusi: Profesor Mohamad Pauzi bin Zakaria, PhD

Fakulti: Fakulti Pengajian Alam Sekitar

Aplikasi berulangan najis haiwan sebagai baja di tanah pertanian adalah amalan biasa pertanian yang boleh membebaskan sisa farmaseutikal veterinar ke alam sekitar melalui haiwan yang dirawat. Kejadian sisa antibiotik dan hormon veterinar di alam sekitar amat membimbangkan kerana pembangunan bakteria tahan antibiotik dan tingkah laku eko-tosikologikal kepada tumbuhtumbuhan dan haiwan. Sehingga kini, kejadian antibiotik veterinar dan hormon di tanah pertanian Malaysia tidak pernah dilaporkan. Kekurangan data mungkin disebabkan oleh tiada kaedah yang sejagat untuk analisis sisa antibiotik dan hormon dalam matriks pepejal alam sekitar yang kompleks. Oleh itu, satu kaedah yang boleh dipercayai, mudah, cepat dan kos efektif telah dibangunkan untuk kuantifikasi sembilan antibiotik (doxycycline, enrofloxacin, erythromycin, flumequine, norfloxacin, sulfadiazine, tilmicosin, trimethoprim, tylosin) dan satu hormon (progesteron) dalam najis haiwan, tanah, dan kompos najis. Secara umumnya, kaedah ini mempunyai masa analisis yang lebih singkat, kos pengekstrakan fasa pepejal (PFP) yang lebih rendah, kadar perolehan yang lebih baik, had pengesanan kaedah (HPK) dan had kuantifikasi kaedah (HKK) yang lebih rendah berbanding dengan Kaedah USEPA 1694, Kim dan Carlson (2007) dan Blackwell et al, (2004). Kaedah ini mempunyai analit sasaran dan matriks sampel yang paling dekat dengan Kaedah USEPA 1694. Masa pengekstrakan telah dikurangkan daripada> 125 minit kepada > 63 minit, masa analisis kromatografi cair telah dikurangkan daripada 61 minit kepada 25 minit dan kos PFP kartrij telah dikurangkan daripada RM70.00 kepada RM10.30 per sampel berbanding dengan Kaedah USEPA 1694. Kaedah yang dibangunkan adalah berdasarkan pengekstrakan ultrasonik dengan MeOH: ACN: EDTA: McIlvaine penampan (pH 4) (30:20:25:25), PFP menggunakan kartrij HLB (3cc/60mg) diikuti oleh analisis menggunakan kromatografi cair berprestasi tinggi-tandem jisim spektrometri (KCBT-JS/JS) dengan 25 minit jumlah masa analisis. Kaedah ini telah disahkan dan diuji ke atas tanah, najis dan sampel kompos najis dan ia menunjukkan bahawa kaedah ini mampu untuk mengesan dan kuantifikasi analit sasaran secara serentak dengan pemilihan dan kepekaan yang baik. Prestasi keseluruhan kaedah adalah baik untuk analisis majoriti analit, dengan kadar perolehan lebih daripada 80% untuk majority analit dalam setiap jenis sampel matrik. HPK dan HKK telah dicapai µg/kg serendah 0.5 dan 2 berat kering (BK) untuk

trimethoprim. Tilmicosin adalah kali pertama dilaporkan dalam matriks alam sekitar. Kaedah ini kemudiannya diaplikasikan pada sampel najis ayam daging dan tanah pertanian yang diambil di Selangor, Negeri Sembilan dan Melaka untuk mengenal pasti dan menentukan sisa antibiotik dan hormon veterinar dalam alam sekitar. Sampel najis ayam daging didapati tercemar dengan sekurang-kurangnya tujuh analit sasaran, iaitu doxycyline, enrofloxacin, flumequine, norfloxacin, trimethoprim, tylosin dan progesteron. Kepekatan maksimum antibiotik yang dikesan dalam najis ayam daging ialah doxycycline iaitu sebanyak 78516.1 µg/kg BK. Untuk sampel tanah pertanian, sisa doxycycline, enrofloxacin, flumequine dan trimethoprim telah dikesan dalam setiap sampel tanah. Kepekatan maksimum yang dikesan di dalam tanah adalah flumequine iaitu sebanyak 1331.4 µg/kg BK. Hasil kajian menunjukkan bahawa kaedah ini berpotensi untuk analisis sisa antibiotik dan hormon veterinar dalam matriks pepejal alam sekitar.

Kata kunci: antibiotik veterinar, hormon, KCBT-JS/JS, najis, tanah

ACKNOWLEDGEMENTS

I would like to express my sincere of appreciation to my main supervisor Prof. Dr. Mohamad Pauzi Zakaria and co-supervisors Associate Prof. Dr. Puziah Abdul Latif and Prof. Dr. Nazamid Saari for advising and guiding me throughout my Ph.D study.

This study was financially supported by Fundamental Research Grant Scheme (project number 5524013) from Ministry of Higher Education Malaysia. I hereby acknowledge the financial support from University Putra Malaysia for awarding the Graduate Research Fellowship. I would like to thank Chromatography Laboratory, Faculty Food Science and Technology, Universiti Putra Malaysia for their support on LC-MS/MS operation.

Special thanks to Mr. Thai Hoeng for his support and help during the field sampling. I am deeply appreciated to my course mates and friends for their moral support and help throughout my study. In particular, Madam Eugenie Tan Sin Sing, Mr. Muhammad Raza, Dr. Najat Ahmed Al-Odaini, Mr. Yap Ken Choy, Miss Khairunnisa Zainuddin, Miss Norazida Manan, Miss Nurul Afiqah, Madam Nur Hazirah, Mr. Mohd Fahmi, Mr. Mohd Saiful, Mr. Mohammad Roshide, Miss Zakiah Ponrahono and all others I fail to mention. I am grateful to the staffs in Faculty of Environmental Studies for providing me their analytical equipments and help in my research. Finally, I would like to thank my parents and family members for their encouragement and love.

TABLE OF CONTENTS

DEDICATIO ABSTRACT ABSTRAK ACKNOWL APPROVAI DECLARAT LIST OF TA LIST OF FIO LIST OF AB	ON EDGE TION BLES GURES BREV	MENTS G IATIONS/NOTATIONS/GLOSSARY OF TERMS	Page iii iv vii x xi xii xviii xviii xx xxiii
CHAPTER			
1	1.1 1.2 1.3 1.4 1.6	INTRODUCTION Background of the Study Problem Statement Significance of the Study Objectives of the Study Thesis Synopsis	1 6 8 10 11
2	2.1 2.2	LITERATURE REVIEW Introduction 2.1.1 Veterinary Antibiotics and Livestock Usage 2.1.2 Steroid Hormones Generated by CAFOs 2.1.3 Benefits of Antibiotics 2.1.4 Use of Hormones in CAFOs Sources, Transport Pathways and Fate of Veterinary Antibiotics and Hormones in the Environment	13 14 15 17 18
	2.3 2.4	 2.2.1 Sources 2.2.2 Transport, Pathways and Fate Effects of Veterinary Antibiotics and Hormones Enter To The Environment 2.3.1 Antibiotic Resistance 2.3.2 Toxicity to Plants and Animals 2.3.3 Endocrine Disrupting Compounds (EDCs) Analytical Methods To Determine Veterinary Antibiotics and Hormones in Broiler Manure, Soil and Manure Compost 2.4.1 Introduction 2.4.2 Sample Preparation and Extraction 	20 23 26 27 29 30 32
		2.4.3 Instrumental Analysis	35

2.4.3 Instrumental Analysis

	2.5	Waste Management To Control Veterinary Antibiotics and Hormones From CAFOs	
		2.5.2 Regulation and Legislation	41
3		DEVELOPMENT OF HPLC-MS/MS METHOD FOR SIMULTANEOUS DETERMINATION OF VETERINARY ANTIBIOTICS AND HORMONE IN SOIL, CHICKEN MANURE AND MANURE COMPOST	
	3.1	Introduction	44
	3.2	Materials and Methods	
		3.2.1 Selection of Target Analytes	46
		3.2.2 Reference Standards	49
		3.2.3 Reagents and Materials	50
		3.2.4 Preparation of Reference Materials	50
		3.2.5 Cleaning and Silanization of Glassware	51
		3.2.6 Instrumentation	52
		3.2.7 Development and Optimization of HPLC-ESI- MS/MS Method	53
		3.2.8 Identification and Quantification with HPLC- MS/MS	61
		3.2.9 Optimization of Sample Pre-treatment,	62
		Extraction and Solid Phase Extraction	
		3.2.10 Quality Control	67
	3.3	Results and Discussion	
		3.3.1 Selection of Target Analytes	68
		3.3.2 Development and Optimization of HPLC-ESI- MS/MS Method	70
		3.3.3 Optimization of Sample Pre-treatment, Extraction and Solid Phase Extraction	92
	3.4	Conclusion	99
4		INSTRUMENTAL PERFORMANCE AND	
	41	Introduction	103
	4.1	Materials and Methods	105
	1.4	4.2.1 Preparation of Reference Materials	106
		4.2.2 Validation of HPLC-MS/MS Performance	107
		4.2.3 Validation of Method Performance	109
	43	Results and Discussion	107
	1.0	4.3.1 Validation of HPLC-MS/MS Performance	110
		4.3.2 Validation of Method Performance	112

xv

	\sim	1 .
44	Conc	noiniti
-T* T	COIR	.iuoion

5	APPLICATION OF THE DEVELOPED METHOD TO STUDY THE OCCURRENCES OF VETERINARY ANTIBIOTICS AND HORMONE IN BROILER MANURE AND LAND APPLIED MANURE IN SELECTED LOCATIONS IN	
	SELANGOR, NEGERI SEMBILAN AND MELAKA	
5.1	Introduction	131
5.2	Materials and Methods	
	5.2.1 Sampling Locations	133
	5.2.2 Collection of Broiler Manure and Manure Amended Agricultural Soil Samples	136
	5.2.3 Sample Preparation and Extraction	141
	5.2.4 Solid Phase Extraction	142
	5.2.5 HPLC-MS/MS Analysis	143
	5.2.6 Calibration	143
	5.2.4 Statistical Analysis	145
	5.2.5 Quality Control	145
5.3	Results and Discussion	
	5.3.1 Determination of Physico-chemical Properties of Soil and Manure Samples	146
	5.3.2 Occurrence of Antibiotics and Hormone In Broiler Manure Samples	156
	53.3 Occurrence of Antibiotics and Hormone	167
	Residues in Manure Amended Agricultural	10/
	Soil Samples	
5.4	Conclusion	175
6	CONCLUSIONS AND RECOMMENDATIONS	
6.1	Conclusions	177
6.2	Recommendations for Future Work	181
REFERENCES APPENDICES BIODATA OF STUDENT LIST OF PUBLICATIONS		183 196 208 209

LIST OF TABLES

T	able	·	Page
	2.1	Methods to determine veterinary antibiotics in different sample matrices	37
	3.1	Pharmaceuticals under study (therapeutic class, name, CAS-No, molecular structure, molecular weight, $LogP$ and pK_a)	47
	3.2	Gradient of USEPA Method 1694 Group 1 analysis for the separation of seven analytes (enrofloxacin, erythromycin, flumequine, norfloxacin, sulfadiazine, trimethoprim and tylosin) detected in ESI (+) mode	57
	3.3	Gradient of USEPA Method 1694 Group 2 analysis for the separation of one analytes (doxycycline) detected in ESI (+) mode	58
	3.4	Gradient of for the optimized HPLC separation method of ten analytes (doxycycline, enrofloxacin, erythromycin, flumequine, norfloxacin, sulfadiazine, tilmicosin, trimethoprim, tylosin and progesterone) detected in ESI (+) mode	59
	3.5	Internal standard used for each group of analyte	62
	3.6	Malaysia: Drug use in broiler farm	68
	3.7	Common antibiotics used in broiler farms	70
	3.8	Precursor ions, product ions, collision energy, tube lens, retention time and relative retention time for ten native analytes and five internal standards	72
	3.9	Optimal conditions for ESI-MS/MS parameters	90
	3.10	Summary of preliminary optimization of different extraction methods	96
	4.1	Instrumental performance and validation data	114

 \bigcirc

4.2	Intra-day and inter-day instrumental method repeatability	117
4.3	Analytical method performance and validation data	125
4.4	Comparison of current method to others closest method for analyzing veterinary pharmaceuticals in soil and biosolids.	128
5.1	CAFOs as defined by US Environmental Protection Agency	134
5.2	Descriptions of broiler manure sampling locations	137
5.3	Descriptions of manure amended agricultural soil sampling locations	137
5.4	Physico-chemical Properties of soil and manure samples on dry weight basis	150
5.5	Pearson correlation matrix for the physico-chemical properties, heavy metal contents, antibiotics and hormone concentration of the manure samples	152
5.6	Pearson correlation matrix for the physico-chemical properties, heavy metal contents, antibiotics and hormone concentration of the soil samples	154
5.7	Mean concentration (µg/kg dry weight, n=3) of detected antibiotics and hormone in broiler manure samples in each location	158
5.8	Mean concentration (µg/kg dry weight, n=3) of detected antibiotics and hormone in manure amended agricultural soil samples in each location	168

LIST OF FIGURES

Figure		Page
1.1	Population of broiler and value of production of broiler in Malaysia from year 2005 to 2010 (DVS, 2010). The number given in year *2010 represents the estimated value from Department of Veterinary Services Malaysia.	2
2.1	Anticipated sources, fate and effects of veterinary pharmaceuticals to the environment (modified according to (Hamscher & Hartung, 2008)	19
3.1	Solid environmental sample pre-treatment and extraction, a) sample vortex with extraction buffer, b) sample ultrasonic extraction c) centrifuge to separate solid and liquid extract and d) diluted extract filtration	65
3.2	Sample loading during SPE	67
3.3	SRM chromatogram for enrofloxacin standard (250 ng/mL)	73
3.4	SRM chromatogram of IS and native standard mixtures (250 ng/mL) detected in ESI (+) using USEPA Method 1694 Group 1 analysis	79
3.5	SRM chromatogram of IS and native standard mixtures (250 ng/mL) detected in ESI (+) using USEPA Method 1694 Group 2 analysis	80
3.6	SRM chromatograms of standard mixtures (25ng/ml of IS and 250ng/ml of native standards) detected in ESI (+) using optimized HPLC separation method	83
3.7	Effects of the composition of injection solution in terms of peak intensity and peak shape for trimethoprim standard (250 ng/mL). (a) 100% methanol, (b) 100% ACN, (c) 0.1% formic acid in 100% methanol, (d) 95:5 (0.3% formic acid, 0.1% ammonium formate in ultrapure water, ACN:MeOH (1:1)), (e) 50% MeOH in H ₂ O, (f) 25% MeOH in H ₂ O and (g) 0.1% formic acid, 25% MeOH in H ₂ O	87

C

3.8	Optimization of spray voltage for ten analytes detected in ESI(+)	90
3.9	Optimization of capillary temperature for ten analytes detected in ESI (+)	91
3.10	Optimization of vaporizer temperature for ten analytes detected in ESI (+)	91
3.11	Flow chart for determination of PPCPs in soil, sediment, and biosolids by LC-MS/MS in USEPA Method 1694, (USEPA, 2007a) a) sample preparation and extraction and b) solid phase extraction followed by LC-MS/MS analysis	95
3.12	Flowchart of the steps for development of HPLC-MS/MS method for simultaneous determination of veterinary antibiotics and hormone in broiler manure, soil and manure compost	100
3.13	Flowchart of optimized sample preparation and extraction method for simultaneous determination of veterinary antibiotics and hormone in soil and biosolids	101
3.14	Flowchart of optimized solid phase extraction and instrumental analysis for simultaneous determination of veterinary antibiotics and hormone in soil and biosolids	102
4.1	Instrumental detection limit (IDL) and instrumental quantification limit (IQL) for trimethoprim: a) IDL at 0.001 ng/mL; b) IQL at 0.1 ng/mL	113
4.2	Calibration curve for sulfadiazine with $1/x$ weighting using as ${}^{13}C_6$ -Sulfamethazine IS	116
4.3	SRM chromatograms of spiked pre-cleaned reference soil detected in ESI (+)	120
4.4	SRM chromatograms of spiked pre-cleaned reference manure detected in ESI (+)	121
4.5	SRM chromatograms of spiked pre-cleaned reference compost detected in ESI (+)	122

.

G

.

5.1	Location of the sampling sites at Selangor, Negeri Sembilan and Melaka	135
5.2	SRM chromatogram of target antibiotics and hormone in broiler manure samples collected from Broga	159
5.3	Box-plot summarizing the median (+) and the values between the 25th and 75th percentile of the pharmaceuticals detected in broiler manure samples. Values given in brackets represent the number of positive samples out of 10 locations	161
5.4	SRM chromatogram of target antibiotics and hormone in manure amended agricultural soil samples collected from Broga	169
5.5	Box-plot summarizing the median (+) and the values between the 25th and 75th percentile of the pharmaceuticals found in relative manure amended soil samples. Values given in brackets represent the number of positive samples out of 10 locations	171

 \overline{C}

LIST OF ABBREVIATIONS/NOTATION/GLOSSARY OF TERMS

ACN	Acetonitrile
ANADA	Abbreviated new animal drug application
APCI	Atmospheric Pressure Chemical Ionization
API	Atmospheric pressure ionization
ASE	Accelerated Solvent Extraction
C/N	Carbon to nitrogen ratio
Ca	Calcium
CAD	Collisionally activated dissociation
CAFOs	Concentrated animal feeding operations
CAS	Chemical Abstract Service
Cd	Cadmium
C.E.	Collision Energy
CEC	Cation exchange capacity
Cr	Chromium
Cu	Copper
СҮР	Cytochrome P450 aromatase
DCM	Dichloromethane

DMCS	Dimethylchlorosilane
DW	Dry weight
E1	Estrone
EDCs	Endocrine disrupting compounds
EDTA	Ethylenediamine-tetraacetic acid
ESI	Electrospray Ionization
EU	European Union
FDA	US Food and Drug Administration
Fe	Iron
GC	Gas chromatography
GnRH	Gonadotrophin releasing hormones
GPS	Global Positioning System
H ₂ O	water
H ₃ PO ₄	Ortho-phosphoric acid
HCG	Human chorionic gonadothrophin
HCI	Hydrochloric acid
HFBA	Heptafluorobutyric acid
HGPs	Hormonal growth promotants
HLB	Hydrophilic-lipophilic-balanced
HNO ₃	Nitric acid

xxiii

HPLC	High performance liquid chromatography
HPLC-FLD	High performance liquid chromatography-postcolumn fluorescence derivatization
HPLC-MS	High performance liquid chromatography-mass spectrometry
HPLC-MS/MS	High performance liquid chromatography -tandem mass spectrometry
HPLC-UV	performance liquid chromatography-ultraviolet detector
ICH	International Conference on Harmonisation
IDL	Instrumental Detection Limit
IQL	Instrumental Quantification Limit
IS	Internal Standard
IUPAC	International Union for Pure and Applied Chemistry
K _d	Soil partition coefficient
K _{oc}	Organic carbon partition coefficient
LC	Liquid chromatography
LLE	Liquid-liquid extraction
LOD	Limit of detection
Log P	Octanol-water partition coefficient
µg/g	Microgram per gram
µg/kg	Microgram per kg
µg/L	Microgram per liter

m/z	Mass to charge ratio

- MAE Microwave assisted extraction
- MCX Mixed-mode cation exchange
- MDL Method Detection Limit

Methanol

- MeOH Methanol
- MeOH
- mg/kg milligram per kg
- mg/L milligram per liter
- Min Minute
- mM millimolar
- MQL Method quantification limit
- MRLs Maximum residual limits
- MS Mass Spectrometry
- MS/MS Tandem mass spectrometry
- M_w Molecular weight
- N₂ Nitrogen
- Na Sodium
- Na₂HPO₄ Disodium hydrogen phosphate
- NADA New animal drug application

	ng/g	nanogram per gram
	ng/L	nanogram per liter
	ng/mL	nanogram per milliliter
	Ni	Nickel
	NPCB	National Pharmaceutical Control Bureau
	PAHs	Poly aromatic hydrocarbons
	PD	Lead
	PCBs	Polychlorinated biphenyls
-	рК _а	Acid dissociation constant
	PLE	Pressurized liquid extraction
	PPCPs	Pharmaceutical and personal care products
	Q1	First quadrupole
	Q3	Third quadrupole
	QC	Quality control
	QqQ	Triple quadrupole analyzer
	R ²	Coefficient of determination
C	rpm	Revolutions per minute
	RRF	Relative Response Factor
	RRT	Relative retention time
	RSD	Relative standard deviation

RT	Retention time
S/N	Signal to noise ratio
SAX	Strong anion exchange
SCX	Strong cation exchange
SD	Standard Deviation
SIM	Selected Ion Monitoring
SPE	Solid Phase Extraction
SRM	Selected Reaction Monitoring
STPs	Sewage Treatment Plants
ТК	Total potassium
TN	Total nitrogen
TOC	Total organic carbon
TP	Total phosphorous
TrBA	Tributylamine
USDA	United States Department of Agricutural
USEPA	United States Environmental Protection Agency
USP .	United States Pharmacopeia
UV	Ultraviolet
VMP	Veterinary medicinal product
Zn	Zinc

CHAPTER 1

INTRODUCTION

1.1 Background of the Study

The agricultural sector occupies a major portion of Malaysia's economy; the production of broiler chickens raised for human consumption comprises of 48% compared with other livestock (MOA, 2010). To maintain economic viability, large agribusiness companies began contracting with farmers. This arrangement offered a guaranteed price to the farmer and a controlled and stable animal food-producing environment for the agribusiness (Sarmah et al., 2006).

Malaysia's poultry industry has advanced to the integrated production system of broilers, which produce enormous amounts of chicken manure. Broiler chickens, feed, manure, urine, dead animals, and production operations are all gathered on a small land area. According to the Department of Veterinary Services Malaysia, the population of broiler has increased from 121.1 million birds in year 2005 to 158.7 million birds in year 2010. The production of broilers in Malaysia has generated RM 4,369 million income in year 2005 and increase to RM 5,776 million in year 2010 (Figure 1.1). Problems may come to surface with this continuously growing number of broiler chickens. This is a global phenomenon, especially for countries that

rely on agricultural income as their source of economy. Integrated production systems of broilers are classified as concentrated animal feeding operations (CAFOs). CAFOs is a regulatory concept for large animal feeding operations, which take advantage of the cost effectiveness of large facilities (Shore and Pruden, 2009). The United States Environmental Protection Agency (USEPA) defines CAFOs as agricultural operations where animals are kept and raised in confined situations (USEPA, 2007b).



Figure 1.1 Population of broiler and value of production of broiler in Malaysia from year 2005 to 2010 (DVS, 2010). The number given in year *2010 represents the estimated value from Department of Veterinary Services Malaysia.

To date, only conventional "priority" pollutants in effluent discharged from sewages are regulated, such as inorganic compounds, nitrates and heavy metals. Recently, attention has turned to a new group of pollutants released from the CAFOs that cause negative effects to the environment even in very low concentrations (µg/L or ng/L). These micropollutants are known as the "emerging pollutants" and are not included in the legislation. Three groups of compounds discharged from CAFOs are identified as "emerging pollutants", namely, the steroid hormones, antibiotics, pesticides and insecticides (Laurence Shore and Pruden, 2009).

Antibiotics are often used in animal husbandry in large quantities. Antibiotics exhibit antimicrobial activity in animals in four ways: therapy, metaphylaxis, prophylaxis, and growth promotion (Schwarz et al., 2001). Since antibiotics can improve growth and feed efficiency, the number of days broilers need to reach market weight is reduced. Most antibiotics used in the animal food-producing industry are poorly absorbed in the gut of the animal, resulting in as much as 30–90% of the parent compound release into the environment via excetion from animal body (Sarmah et al., 2006). Some see this as a public health concern, believing such drugs could then enter the ground or surface water, or be taken up by plants and thus contribute to antibiotic resistance development and/or produce adverse reactions in those with antibiotic allergies (Venglovsky et al., 2009).

The pathway of veterinary pharmaceuticals entering the environment is different from human pharmaceuticals. Human pharmaceuticals are discharged into the environment mainly through sewage treatment plants (STPs); however, veterinary pharmaceuticals could enter the environment

not only through direct application in aquaculture, wash off from topical treatments and livestock treatment plants, but also through manure (Hamscher and Hartung, 2008). Once a veterinary application pharmaceutical enters the environment, its behaviour is affected by its physical properties including water solubility, lipophilicity, volatility and sorption potential. Therefore, investigation of antibiotic pollution in chicken manure has special importance because they constitute a major source of hazardous unmetabolized antibiotics to the environment via fertilization of agricultural soil. Occurrence of veterinary antibiotics and hormone residuals in the environment is of concern due to the emergence and development of antibiotic resistance in pathogenic bacteria, and the ecotoxicological behaviour of these compounds. Besides, steroid hormones are potential endocrine disrupting compounds (EDCs); when released into the environment they cause the feminization of aquatic organisms (Saaristo et al., 2009).

Veterinary antibiotics and hormone usually occur as a wide range of pollutants from different chemical classes, with different physical and chemical properties. Analysis of this wide range of compounds in complex sample matrices such as soil, animal manure and compost is more challenging than in water samples. The extraction methods of these pharmaceuticals from soil, manure and compost are mainly based on ultrasonication and solid phase extraction (SPE) (Blackwell et al., 2004; Karcı and Balcıoğlu, 2009). Others are based on liquid-liquid extraction (LLE)

(Martínez-Carballo et al., 2007) and pressurized liquid extraction (PLE) or accelerated solvent extraction (ASE) (Jacobsen et al., 2004). High performance liquid chromatography in tandem with mass spectrometry (HPLC-MS/MS) is often employed for the environmental analysis of veterinary antibiotics due to their low limit of detection (LOD) (Martínez-Carballo et al., 2007; Aust et al., 2008). However, high performance liquid chromatography in combination with ultraviolet detector (HPLC-UV) or high performance liquid chromatography with postcolumn fluorescence derivatization (HPLC-FLD) is still used due to their relatively low cost (Blackwell et al., 2004; Malintan and Mohd, 2006; Karcı and Balcıoğlu, 2009).

To date, there is neither a list of veterinary antibiotics and hormone priority pollutants nor a comprehensive method for antibiotics and hormones analysis for soil, animal manure and compost samples. Since broiler manure is a major source of veterinary of antibiotics in agricultural fields, thus the veterinary antibiotics and hormone composition, and their fate in manureamended soil needs to be given special attention. This study is extensively designed to accommodate different classes of antibiotics and hormone for the very first time and the compilation of these compounds is unique for our country.

1.2 Problem Statement

Currently, there is no legislation for the limits of pharmaceuticals in animal manure and soil in any country. Livestock producers have "blindly" used veterinary antibiotics as supplements in animal feed to increase weight gain and prevent disease among their livestock. These chemicals are introduced increasingly without realising the consequences for the environment, as well as direct and indirect effects for human health. Early research and studies have focussed extensively on the aquatic environment and especially on contaminants being released from pharmaceutical and personal care products (PPCPs) (Boyd et al., 2003; Vanderford et al., 2003; Ellis, 2006; Vanderford and Snyder, 2006; Kim et al., 2007; Al-Odaini et al., 2010). Up till now, only a few studies have investigated the occurrence of veterinary antibiotics in animal manure and manure amended agricultural soil (De Liguoro et al., 2003; Blackwell et al., 2004; Martínez-Carballo et al., 2007; Aust et al., 2008; Karci and Balcioğlu, 2009). However, they feature limited classes of pharmaceuticals.

To date, scientists have reported the occurrence of PPCPs in Malaysian waters (Al-Odaini et al., 2010) as well as the veterinary antibiotics in swine wastewater (Malintan and Mohd, 2006). The occurrence and fate of veterinary antibiotics and hormone in Malaysian agricultural soil have not been documented. Previous analytical methods for antibiotics and hormones

analysis in soil and biosolid samples required complex extraction and analysis methods, hence resulted in higher cost. Thus, a multi-residue analytical method was developed in order to fill the gap of knowledge of the occurrences of these compounds in Malaysian agricultural field. The developed method was subsequently applied in chicken manure and manure amended agricultural soil to investigate the level of veterinary antibiotics and hormone pollution in Malaysian terrestrial environment. The same method was applied in manure compost to study the evolution of veterinary antibiotics and hormone in a laboratory composting experiment.

Therefore, the present study is able to answer the following questions:

- i. Is there any available method for the simultaneous determination and quantification of nine veterinary antibiotics and one hormone in broiler chickens manure and manure amended agricultural soil samples?
- ii. What is the level of antibiotics and hormone pollution in manure and agricultural soil in Malaysia?
- iii. Can veterinary antibiotics and hormone lead to the contamination of agricultural soil via fertilization of animal manure?

1.3 Significance of Study

Pollutants such as veterinary antibiotics and hormones that are not covered by environmental legislation have increasingly become important emerging environmental contaminants. By complying with the Food Act 1985, the maximum residual limits (MRLs) in food producing animals does not mean that we are safe from pharmaceutical pollution. There is a growing concern that the antibiotics and steroid hormones discharge from livestock production are contaminating soil, surface and groundwater; contributing to the development of antibiotic resistance in the environment and possesses the potential as EDCs.

To date, there is no data about antibiotics and hormone pollution in the Malaysian agricultural soil. This is the pioneering study on the development of sensitive and selective multi-residue HPLC-MS/MS method for the determination of nine antibiotics and one hormone in chicken manure, land applied manure and manure compost. The occurrence data of antibiotics and hormone in Malaysian agricultural soil in this study is of great importance in awakening the public understanding to the source, transport and effects of these compounds.

Specifically, the significances of the study are listed below:

- A new method is developed for the simultaneous determination and quantification of nine antibiotics and one hormone in broiler manure, manure amended agricultural soil and manure compost.
- The occurrence data in this study can be used to investigate the level of antibiotics and hormone pollution in manure and agricultural soil samples in Malaysia for the very first time.
- iii. The findings obtained in this study can be used to demonstrate that veterinary antibiotics can lead to the contamination of agricultural soils via fertilization with animal manure.
- iv. This study will produce the baseline data for the occurrence of nine antibiotics (doxycycline, enrofloxacin, erythromycin, flumequine, norfloxacin, sulfadiazine, tilmicosin, trimethoprim, tylosin) and one hormone (progesterone) in manure and agricultural soil samples in Malaysia. Data from this study is anticipated to be an important reference for other studies conducted in the future regarding this issue, especially fate and effects of these compounds to environment. These data are especially significant when considering the lack of information on the presence of antibiotic residues in agricultural fields in Malaysia.

1.4 Objectives of the Study

The overall aim of this study is to develop a multi-residue analytical method for quantifying veterinary antibiotics and hormones in soil and biosolid matrices, based on ultrasonication with buffer, solid phase extraction (SPE) followed by HPLC-MS/MS for the simultaneous analysis of veterinary antibiotics and hormone. The method is validated using relative reference matrices, namely, pre-cleaned soil, pre-cleaned chicken manure and precleaned composts. The validated method is subsequently applied on environmental samples to investigate the occurrences of these pollutants in broiler farms and agricultural fields in Malaysia.

This study embarks on the following specific objectives:

- To develop and validate the HPLC-MS/MS method for simultaneous determination of nine antibiotics (doxycycline, enrofloxacin, erythhromycin, flumequine, norfloxacin, sulfadiazine, tilmicosin, trimethoprim and tylosin) and one hormone (progesterone) in broiler chicken manure, soil and manure compost at µg/kg level.
- ii. To apply the developed method to study the occurences of veterinary antibiotics and hormone residues in broiler chicken manure and manure amended soil samples collected from selected locations in Selangor, Negeri Sembilan and Melaka.

1.5 Thesis Synopsis

This thesis focuses on the development of HPLC-MS/MS method for the detection and quantification of veterinary antibiotics and hormone in broiler manure, soil and manure compost. The following section describes the organization of this thesis and its relative chapters.

Chapter 2 gives a full review of veterinary antibiotics and hormone in CAFOs. The sources, transport pathway and fate of veterinary pharmaceuticals in environment are reviewed in detail. In addition, the effects of veterinary pharmaceuticals enter to the environment are summarized. Previous available analytical methods to determine veterinary pharmaceuticals in soil and biosolid are reviewed. Finally, proper waste management to control the pharmaceuticals from CAFOs is also summarized in this chapter.

Chapter 3 describes in detail the development and optimization of the HPLC-MS/MS analytical method for simultaneous detection and quantification of nine antibiotics and one hormone in soil, manure and manure compost. The optimizations of sample preparation and extraction method are discussed comprehensively.

Chapter 4 discusses the validation of the developed method in environmental samples. The instrumental performance and overall analytical method performance are also discussed.

Chapter 5 discusses the application of the developed method on broiler manure samples and its relative manure amended soil samples. The relationship between the physico-chemical properties of manure and soil samples to the concentration of pharmaceuticals in manure and soil were presented in Pearson correlation matrix. Results of analysis of broiler manure and its relative manure amended soil samples are also discussed.

Chapter 6 summaries the research, discusses the overall findings and the contribution of this research. Recommendations for improving the results of the study and some future directions for the study are also discussed.

REFERENCES

- Al-Odaini, N. A. (2010). Development and Application of HPLC-MS/MS Method For Determination of Human Pharmaceuticals and Synthetic Hormones in River Water and Sewage Effluents. Doctoral dissertation, Universiti Putra Malaysia.
- Al-Odaini, N. A., Zakaria, M. P., Yaziz, M. I., & Surif, S. (2010). Multi-residue analytical method for human pharmaceuticals and synthetic hormones in river water and sewage effluents by solid-phase extraction and liquid chromatography tandem mass spectrometry. *Journal of Chromatography A*, 1217(44), 6791-6806.
- Ardrey, R. E. (2003). Applications of High Performance Liquid Chromatography-Mass Spectrometry Liquid Chromatography-Mass Spectrometry: An Introduction (pp. 129-234). West Sussex, England: Wiley.
- Arikan, O. A., Rice, C., & Codling, E. (2008). Occurrence of antibiotics and hormones in a major agricultural watershed. *Desalination*, 226(1-3), 121-133.
- Aust, M.-O., Godlinski, F., Travis, G. R., Hao, X., McAllister, T. A., Leinweber, P., et al. (2008). Distribution of sulfamethazine, chlortetracycline and tylosin in manure and soil of Canadian feedlots after subtherapeutic use in cattle. *Environmental Pollution*, 156(3), 1243-1251.
- Bartelt-Hunt, S., Snow, D. D., Damon-Powell, T., & Miesbach, D. (2011). Occurrence of steroid hormones and antibiotics in shallow groundwater impacted by livestock waste control facilities. *Journal of Contaminant Hydrology*, 123(3-4), 94-103.
- Bevacqua, C. E., Rice, C. P., Torrents, A., & Ramirez, M. (2011). Steroid hormones in biosolids and poultry litter: A comparison of potential environmental inputs. Science of The Total Environment, 409(11), 2120-2126.
- Bialk-Bielinska, A., Kumirska, J., Palavinskas, R., & Stepnowski, P. (2009). Optimization of multiple reaction monitoring mode for the trace analysis of veterinary sulfonamides by LC-MS/MS. *Talanta*, 80(2), 947-953.

- Blackwell, P. A., Holten Lützhøft, H.-C., Ma, H.-P., Halling-Sørensen, B., Boxall, A. B. A., & Kay, P. (2004). Ultrasonic extraction of veterinary antibiotics from soils and pig slurry with SPE clean-up and LC-UV and fluorescence detection. *Talanta*, 64(4), 1058-1064.
- Boxall, A. B. A. (2008a). Fate and Transport of Veterinary Medicines in the Soil Environment. In D. S. Aga (Ed.), *Fate of Pharmaceuticals in the Environment and in Water Treatment Systems* (pp. 123-137). Broken Sound Parkway NW: Taylor and Francis Group.
- Boxall, A. B. A. (2008b). Fate of Veterinary Medicines Applied to Soils. In K. Kümmerer (Ed.), Pharmaceuticals in the Environment-Sources, Fate, Effects and Risks (pp. 103-119). Berlin: Springer.
- Boxall, A. B. A., Johnson, P., Smith, E. J., Sinclair, C. J., Stutt, E., & Levy, L. S. (2006). Uptake of Veterinary Medicines from Soils into Plants. *Journal* of Agricultural and Food Chemistry, 54(6), 2288-2297.
- Boyd, B., Bjork, H., Billing, J., Shimelis, O., Axelsson, S., Leonora, M., et al. (2007). Development of an improved method for trace analysis of chloramphenicol using molecularly imprinted polymers. *Journal of Chromatography A*, 1174(1-2), 63-71.
- Boyd, G. R., Palmeri, J. M., Zhang, S., & Grimm, D. A. (2004). Pharmaceuticals and personal care products (PPCPs) and endocrine disrupting chemicals (EDCs) in stormwater canals and Bayou St. John in New Orleans, Louisiana, USA. Science of The Total Environment, 333(1-3), 137-148.
- Boyd, G. R., Reemtsma, H., Grimm, D. A., & Mitra, S. (2003). Pharmaceuticals and personal care products (PPCPs) in surface and treated waters of Louisiana, USA and Ontario, Canada. *Science of The Total Environment*, 311(1-3), 135-149.
- Bretnall, A. E., Clarke, G. S., Satinder, A., & Stephen, S. (2011). 11 Validation of Analytical Test Methods *Separation Science and Technology* (Vol. Volume 10, pp. 429-457): Academic Press.
- Chang, H., Wu, S., Hu, J., Asami, M., & Kunikane, S. (2008). Trace analysis of androgens and progestogens in environmental waters by ultraperformance liquid chromatography-electrospray tandem mass spectrometry. *Journal of Chromatography A*, 1195(1-2), 44-51.

- Chenxi, W., Spongberg, A. L., & Witter, J. D. (2008). Determination of the persistence of pharmaceuticals in biosolids using liquidchromatography tandem mass spectrometry. *Chemosphere*, 73(4), 511-518.
- De Liguoro, M., Cibin, V., Capolongo, F., Halling-Sørensen, B., & Montesissa, C. (2003). Use of oxytetracycline and tylosin in intensive calf farming: evaluation of transfer to manure and soil. *Chemosphere*, 52(1), 203-212.
- Díaz-Cruz, M. S., & Barceló, D. (2007). Recent advances in LC-MS residue analysis of veterinary medicines in the terrestrial environment. *TrAC Trends in Analytical Chemistry*, 26(6), 637-646.
- Díaz-Cruz, M. S., López de Alda, M. J., & Barceló, D. (2003). Environmental behavior and analysis of veterinary and human drugs in soils, sediments and sludge. *TrAC Trends in Analytical Chemistry*, 22(6), 340-351.
- Dolliver, H. (2007). Fate and transport of veterinary antibiotics in the environment. Unpublished Ph.D., University of Minnesota, United States --Minnesota.
- Dolliver, H., Gupta, S., & Noll, S. (2008). Antibiotic Degradation during Manure Composting. J. Environ. Qual., 37, 1245-1253.
- DrugBank (2011). DrugBank Open Data Drug & Drug Target Database. Retrieved 26-11-2011: http://drugbank.ca/
- DVS (2010). Department of Veterinary Services Malaysia. Drug Used In The Premises.

Ellis, J.B., (2006). Pharmaceutical and personal care products (PPCPs) in urban receiving waters. *Environmental Pollution*, 144(1), 184-189.

Council Directive 81/852/EEC of 28 September 1981 on the approximation of the laws of the Member States relating to analytical, pharmacotoxicological and clinical standards and protocols in respect of the testing of veterinary medicinal products (1981).

- Directive 2001/82/EC of The European Parliament and of The Council of 6 November 2001 on The Community Code Relating to Veterinary Medicinal Products (2001).
- Commission Directive 2009/9/EC of 10 February 2009 amending Directive 2001/82/EC of the European Parliament and of the Council on the Community code relating to medicinal products for veterinary use (2009).
- Eggen, T., Asp, T. N., Grave, K., & Hormazabal, V. (2011). Uptake and translocation of metformin, ciprofloxacin and narasin in forage- and crop plants. *Chemosphere*, 85(1), 26-33.
- EURACHEM (1998). The Fitness for Purpose of Analytical Methods: A Laboratory Guide to Method Validation and Related Topics. Retrieved 12-12-2011: http://www.eurachem.org/guides/pdf/valid.pdf
- Ferrer, I., & Thurman, E. M. (2007). Multi-residue method for the analysis of 101 pesticides and their degradates in food and water samples by liquid chromatography/time-of-flight mass spectrometry. *Journal of Chromatography A*, 1175(1), 24-37.
- García-Galán, M. J., Garrido, T., Fraile, J., Ginebreda, A., Díaz-Cruz, M. S., & Barceló, D. (2010). Simultaneous occurrence of nitrates and sulfonamide antibiotics in two ground water bodies of Catalonia (Spain). Journal of Hydrology, 383(1-2), 93-101.
- Gracia-Lor, E., Sancho, J. V., & Hernandez, F. Multi-class determination of around 50 pharmaceuticals, including 26 antibiotics, in environmental and wastewater samples by ultra-high performance liquid chromatography-tandem mass spectrometry. *Journal of Chromatography A*, 1218(16), 2264-2275.
- Haller, M. Y., Muller, S. R., McArdell, C. S., Alder, A. C., & Suter, M. J. F. (2002). Quantification of veterinary antibiotics (sulfonamides and trimethoprim) in animal manure by liquid chromatography-mass spectrometry. *Journal of Chromatography A*, 952(1-2), 111-120.

Halling-Sørensen, B., Nors Nielsen, S., Lanzky, P. F., Ingerslev, F., Holten Lützhøft, H. C., & Jorgensen, S. E. (1998). Occurrence, fate and effects of pharmaceutical substances in the environment- A review. *Chemosphere*, 36(2), 357-393.

- Hamscher, G., & Hartung, J. (2008). Veterinary antibiotics in dust: sources, environmental concentrations and possible health hazards. In K. Kümmerer (Ed.), *Pharmaceuticals in the Environment: Sources, Fate, Effects, and Risks* (pp. 95-102). Berlin Heidelberg: Springer.
- Hays, V. W. (1986). Benefits and Risks of Antibiotics Use in Agriculture Agricultural Uses of Antibiotics (Vol. 320, pp. 74-87): American Chemical Society.
- Herklotz, P. A., Gurung, P., Vanden Heuvel, B., & Kinney, C. A. (2010). Uptake of human pharmaceuticals by plants grown under hydroponic conditions. *Chemosphere*, 78(11), 1416-1421.
- Heuer, H., Focks, A., Lamshoft, M., Smalla, K., Matthies, M., & Spiteller, M. (2008). Fate of sulfadiazine administered to pigs and its quantitative effect on the dynamics of bacterial resistance genes in manure and manured soil. *Soil Biology and Biochemistry*, 40(7), 1892-1900.
- Hoa, P. T. P., Managaki, S., Nakada, N., Takada, H., Shimizu, A., Anh, D. H., et al. (2011). Antibiotic contamination and occurrence of antibioticresistant bacteria in aquatic environments of northern Vietnam. *Science of The Total Environment*, 409(15), 2894-2901.
- Hu, X.-G., Yi, L., Zhou, Q.-X., & Xu, L. (2008). Determination of Thirteen Antibiotics Residues in Manure by Solid Phase Extraction and High Performance Liquid Chromatography. *Chinese Journal of Analytical Chemistry*, 36(9), 1162-1166.
- Huang, C., Chen, X., Xie, W., Feng, J., Liu, H., & Ding, H. (2009). Detection method for simultaneously measuring residue of tetracyclines (TCs) drugs in royal jelly. China Patent No. CN 101571525 (A).
- ICH (1995). Note For Guidance on Validation of Analytical Procedures: Text and Methodology: European Medicines Agency.
- Jacobsen, A. M., Halling-Sørensen, B., Ingerslev, F., & Honoré Hansen, S. (2004). Simultaneous extraction of tetracycline, macrolide and sulfonamide antibiotics from agricultural soils using pressurised

liquid extraction, followed by solid phase extraction and liquid chromatography-tandem mass spectrometry. *Journal of Chromatography A*, 1038(1-2), 157-170.

- Karcı, A., & Balcıoğlu, I. A. (2009). Investigation of the tetracycline, sulfonamide, and fluoroquinolone antimicrobial compounds in animal manure and agricultural soils in Turkey. Science of The Total Environment, 407(16), 4652-4664.
- Kasprzyk-Hordern, B., Dinsdale, R. M., & Guwy, A. J. (2008). The effect of signal suppression and mobile phase composition on the simultaneous analysis of multiple classes of acidic/neutral pharmaceuticals and personal care products in surface water by solid-phase extraction and ultra performance liquid chromatography-negative electrospray tandem mass spectrometry. *Talanta*, 74(5), 1299-1312.
- Kay, P., Blackwell, P. A., & Boxall, A. B. A. (2005). A lysimeter experiment to investigate the leaching of veterinary antibiotics through a clay soil and comparison with field data. *Environmental Pollution*, 134(2), 333-341.
- Kazeto, Y., Place, A. R., & Trant, J. M. (2004). Effects of endocrine disrupting chemicals on the expression of CYP19 genes in zebrafish (Danio rerio) juveniles. *Aquatic Toxicology*, 69(1), 25-34.
- Kemper, N. (2008). Veterinary antibiotics in the aquatic and terrestrial environment. *Ecological Indicators*, 8(1), 1-13.
- Khan, S. J., Roser, D. J., Davies, C. M., Peters, G. M., Stuetz, R. M., Tucker, R., et al. (2008). Chemical contaminants in feedlot wastes: Concentrations, effects and attenuation. *Environment International*, 34(6), 839-859.
- Kim, S.-C., & Carlson, K. (2005). LC-MS² for quantifying trace amounts of pharmaceutical compounds in soil and sediment matrices. *TrAC Trends in Analytical Chemistry*, 24(7), 635-644.
- Kim, S.-C., & Carlson, K. (2007). Quantification of human and veterinary antibiotics in water and sediment using SPE/LC/MS/MS. Anal. Bioanal. Chem., 387, 1301-1315.

- Kim, S. D., Cho, J., Kim, I. S., Vanderford, B. J., & Snyder, S. A. (2007). Occurrence and removal of pharmaceuticals and endocrine disruptors in South Korean surface, drinking, and waste waters. *Water Research*, 41(5), 1013-1021.
- Kim, Y., Jung, J., Kim, M., Park, J., Boxall, A. B. A., & Choi, K. (2008). Prioritizing veterinary pharmaceuticals for aquatic environment in Korea. Environmental Toxicology and Pharmacology, 26(2), 167-176.
- Kmellar, B., Fodor, P., Pareja, L., Ferrer, C., Martinez-Uroz, M. A., Valverde, A., et al. (2008). Validation and uncertainty study of a comprehensive list of 160 pesticide residues in multi-class vegetables by liquid chromatography-tandem mass spectrometry. *Journal of Chromatography A*, 1215(1-2), 37-50.
- Kong, W. D., Zhu, Y. G., Liang, Y. C., Zhang, J., Smith, F. A., & Yang, M. (2007). Uptake of oxytetracycline and its phytotoxicity to alfalfa (Medicago sativa L.). *Environmental Pollution*, 147(1), 187-193.
- Koschorreck, J., Koch, C., & Ronnefahrt, I. (2002). Environmental risk assessment of veterinary medicinal products in the EU a regulatory perspective. *Toxicology Letters*, 131(1-2), 117-124.
- Kümmerer, K. (2001). Drugs in the environment: emission of drugs, diagnostic aids and disinfectants into wastewater by hospitals in relation to other sources - a review. *Chemosphere*, 45(6-7), 957-969.
- Kümmerer, K. (2008). Antibiotics in the Environment. In K. Kümmerer (Ed.), *Pharmaceuticals in the Environment-Sources, Fate, Effects and Risks* (pp. 75-93). Berlin: Springer.
- Kümmerer, K. (2009). Antibiotics in the aquatic environment A review Part I. Chemosphere, 75(4), 417-434.
- Kumar, K., Gupta, S. C., Baidoo, S. K., Chander, Y., & Rosen, C. J. (2005). Antibiotic Uptake by Plants from Soil Fertilized with Animal Manure. *Journal of Environmental Quality*, 2082-2085.
- Lalumera, G. M., Calamari, D., Galli, P., Castiglioni, S., Crosa, G., & Fanelli, R. (2004). Preliminary investigation on the environmental occurrence

and effects of antibiotics used in aquaculture in Italy. *Chemosphere*, 54(5), 661-668.

- Lange, I. G., Daxenberger, A., Schiffer, B., Witters, H., Ibarreta, D., & Meyer, H. H. D. (2002). Sex hormones originating from different livestock production systems: fate and potential disrupting activity in the environment. *Analytica Chimica Acta*, 473(1-2), 27-37.
- Le, T. X., Munekage, Y., & Kato, S.-i. (2005). Antibiotic resistance in bacteria from shrimp farming in mangrove areas. *Science of The Total Environment*, 349(1-3), 95-105.
- Lin, A. Y.-C., Yu, T.-H., & Lin, C.-F. (2008). Pharmaceutical contamination in residential, industrial, and agricultural waste streams: Risk to aqueous environments in Taiwan. *Chemosphere*, 74(1), 131-141.
- Lindberg, R., Jarnheimer, P.-A., Olsen, B., Johansson, M., & Tysklind, M. (2004). Determination of antibiotic substances in hospital sewage water using solid phase extraction and liquid chromatography/mass spectrometry and group analogue internal standards. *Chemosphere*, 57(10), 1479-1488.
- Lobova, T. I., Barkhatov, Y. V., Salamatina, O. V., & Popova, L. Y. (2008). Multiple antibiotic resistance of heterotrophic bacteria in the littoral zone of Lake Shira as an indicator of human impact on the ecosystem. *Microbiological Research*, 163(2), 152-160.
- Lopez-Serna, R., Petrovic, M., & Barcelo, D. Development of a fast instrumental method for the analysis of pharmaceuticals in environmental and wastewaters based on ultra high performance liquid chromatography (UHPLC)-tandem mass spectrometry (MS/MS). *Chemosphere*, 85(8), 1390-1399.

Mackinnon, J. D. (1993). The proper use and benefits of veterinary antimicrobial agents in swine practice. *Veterinary Microbiology*, 35(3-4), 357-367.

Malintan, N. T., & Mohd, M. A. (2006). Determination of sulfonamides in selected Malaysian swine wastewater by high-performance liquid chromatography. *Journal of Chromatography A*, 1127(1-2), 154-160.

- Managaki, S., Murata, A., Takada, H., Tuyen, B. C., & Chiem, N. H. (2007). Distribution od Macrolides, Sulfonamides, and Trimethoprim in tropical Waters: Ubiquitous Occurence of Veterinary Antibiotics in the Mekong Delta. Environ. Sci. Technol., 41, 8004-8010.
- Martínez-Carballo, E., González-Barreiro, C., Scharf, S., & Gans, O. (2007). Environmental monitoring study of selected veterinary antibiotics in animal manure and soils in Austria. *Environmental Pollution*, 148(2), 570-579.
- Martinez, J. L. (2009). Environmental pollution by antibiotics and by antibiotic resistance determinants. *Environmental Pollution*, 157(11), 2893-2902.
- Marwah, A., Marwah, P., & Lardy, H. (2002). Analysis of ergosteroids: VIII: Enhancement of signal response of neutral steroidal compounds in liquid chromatographic-electrospray ionization mass spectrometric analysis by mobile phase additives. *Journal of Chromatography A*, 964(1-2), 137-151.
- Masahiko, K., Masaki, K., & Takashi, O. (2007). Japan Patent No. WO/2007/046507 W. I. P. Organization.
- Matsui, Y., Ozu, T., Inoue, T., & Matsushita, T. (2008). Occurrence of a veterinary antibiotic in streams in a small catchment area with livestock farms. *Desalination*, 226(1-3), 215-221.
- Melaku, S., Dams, R., & Moens, L. (2005). Determination of trace elements in agricultural soil samples by inductively coupled plasma-mass spectrometry: Microwave acid digestion versus aqua regia extraction. *Analytica Chimica Acta*, 543(1-2), 117-123.
- Messi, P., Guerrieri, E., & Bondi, M. (2005). Antibiotic resistance and antibacterial activity in heterotrophic bacteria of mineral water origin. *Science of The Total Environment*, 346(1-3), 213-219.
- MOA (2010). Ministry of Agricultural Malaysia. *Statistik Asas Sektor Agromakanan* http://www.moa.gov.my/c/document_library/get_file?uuid=fd3b8 983-a68e-4d8c-ae43-021aea365b26&groupId=10136

- Naidong, W., Chen, Y.-L., Shou, W., & Jiang, X. (2001). Importance of injection solution composition for LC-MS-MS methods. *Journal of Pharmaceutical and Biomedical Analysis*, 26(5-6), 753-767.
- NPCB (2009). National Pharmaceutical Control Bureau. Registratrion Guidelines of Veterinary Products (REGOVP). Retrieved 20-12-2011: http://portal.bpfk.gov.my/aeimages/File/Veterinar/REGOVP_ DEC2009(2).pdf.
- Prat, M. D., Ramil, D., Compano, R., Hernandez-Arteseros, J. A., & Granados, M. (2006). Determination of flumequine and oxolinic acid in sediments and soils by microwave-assisted extraction and liquid chromatography-fluorescence. *Analytica Chimica Acta*, 567(2), 229-235.
- Rabolle, M., & Spliid, N. H. (2000). Sorption and mobility of metronidazole, olaquindox, oxytetracycline and tylosin in soil. *Chemosphere*, 40(7), 715-722.
- Ramaswamy, J., Prasher, S. O., Patel, R. M., Hussain, S. A., & Barrington, S. F. (2010). The effect of composting on the degradation of a veterinary pharmaceutical. *Bioresource Technology*, 101(7), 2294-2299.
- Raquel M, W. (2001). Validation of laboratory tests and methods. Seminars in Avian and Exotic Pet Medicine, 10(2), 59-65.
- Redshaw, C. H., Wootton, V. G., & Rowland, S. J. (2008). Uptake of the pharmaceutical Fluoxetine Hydrochloride from growth medium by Brassicaceae. *Phytochemistry*, 69(13), 2510-2516.
- Refsdal, A. O. (2000). To treat or not to treat: a proper use of hormones and antibiotics. *Animal Reproduction Science*, 60-61(0), 109-119.
- Ripp, J. (1996). Analytical Detection Limit Guidance & Laboratory Guide for Determining Method Detection Limits: Wisconsin Department of Natural Resources Laboratory Certification Program.
- Saaristo, M., Craft, J. A., Lehtonen, K. K., & Lindstrom, K. (2009). Sand goby (Pomatoschistus minutus) males exposed to an endocrine disrupting chemical fail in nest and mate competition. *Hormones and Behavior*, 56(3), 315-321.

- Santos, L. H. M. L. M., Araújo, A. N., Fachini, A., Pena, A., Delerue-Matos, C., & Montenegro, M. C. B. S. M. (2010). Ecotoxicological aspects related to the presence of pharmaceuticals in the aquatic environment. *Journal* of Hazardous Materials, 175(1-3), 45-95.
- Sarmah, A. K., Meyer, M. T., & Boxall, A. B. A. (2006). A global perspective on the use, sales, exposure pathways, occurrence, fate and effects of veterinary antibiotics (VAs) in the environment. *Chemosphere*, 65(5), 725-759.
- Schwarz, S., Kehrenberg, C., & Walsh, T. R. (2001). Use of antimicrobial agents in veterinary medicine and food animal production. *International Journal of Antimicrobial Agents*, 17(6), 431-437.
- Scott, L., McGee, P., Walsh, C., Fanning, S., Sweeney, T., Blanco, J., et al. (2009). Detection of numerous verotoxigenic E. coli serotypes, with multiple antibiotic resistance from cattle faeces and soil. *Veterinary Microbiology*, 134(3-4), 288-293.
- Sengeløv, G., Agersø, Y., Halling-Sørensen, B., Baloda, S. B., Andersen, J. S., & Jensen, L. B. (2003). Bacterial antibiotic resistance levels in Danish farmland as a result of treatment with pig manure slurry. *Environment International*, 28(7), 587-595.
- Shao, B., Chen, D., Zhang, J., Wu, Y., & Sun, C. (2009). Determination of 76 pharmaceutical drugs by liquid chromatography-tandem mass spectrometry in slaughterhouse wastewater. *Journal of Chromatography* A, 1216(47), 8312-8318.
- Shore, L., & Pruden, A. (2009). Introduction. In L. S. a. A. Pruden (Ed.), Hormones and Pharmaceuticals Generated by Concentrated Animal Feeding Operations, Transport in Water and Soil (pp. 1-5). New York: Springer Science & Business Media.
- Stoob, K., Singer, H. P., Goetz, C. W., Ruff, M., & Mueller, S. R. (2005). Fully automated online solid phase extraction coupled directly to liquid chromatography-tandem mass spectrometry: Quantification of sulfonamide antibiotics, neutral and acidic pesticides at low concentrations in surface waters. *Journal of Chromatography A*, 1097(1-2), 138-147.

- Syrcause (2011). Syracuse Research Corporation. Retrieved 26-11-2011: http://www.syrres.com
- Teh, C.-H., Murugaiyah, V., & Chan, K.-L. (2011). Developing a validated liquid chromatography-mass spectrometric method for the simultaneous analysis of five bioactive quassinoid markers for the standardization of manufactured batches of Eurycoma longifolia Jack extract as antimalarial medicaments. *Journal of Chromatography A*, 1218(14), 1861-1877.
- Thermo, F. S. (2009). TSQ Series Getting Started Guide Retrieved 05-12-2011, 2011, from http://sjsupport.thermofinnigan.com/techpubs/manuals/TSQ_Serie s_Start.pdf
- Tong, L., Li, P., Wang, Y., & Zhu, K. (2009). Analysis of veterinary antibiotic residues in swine wastewater and environmental water samples using optimized SPE-LC/MS/MS. *Chemosphere*, 74(8), 1090-1097.
- Turiel, E., Bordin, G., & Rodriguez, A. R. (2003). Trace enrichment of (fluoro)quinolone antibiotics in surface waters by solid-phase extraction and their determination by liquid chromatographyultraviolet detection. *Journal of Chromatography A*, 1008(2), 145-155.
- USEPA (1999). Protocol for EPA Approval of New Methods for Organic and Inorganic Analytes in Wastewater and Drinking Water Washington, DC U.S. Environmental Protection Agency.
- USEPA (2007a). Method 1694: Pharmaceuticals and Personal Care Products in Water, Soil, Sediment, and Biosolids by HPLC/MS/MS.
- USEPA (Producer). (2007b, 12 October 2011) Regulatory definitions of large CAFOs, medium CAFO, and small CAFOs. Podcast retrieved from http://www.epa.gov/npdes/pubs/sector_table.pdf.
- Vanderford, B. J., Pearson, R. A., Rexing, D. J., & Snyder, S. A. (2003). Analysis of Endocrine Disruptors, Pharmaceuticals, and Personal Care Products in Water Using Liquid Chromatography/Tandem Mass Spectrometry. Analytical Chemistry, 75(22), 6265-6274.

- Vanderford, B. J., & Snyder, S. A. (2006). Analysis of Pharmaceuticals in Water by Isotope Dilution Liquid Chromatography/Tandem Mass Spectrometry. Environmental Science & Technology, 40(23), 7312-7320.
- Venglovsky, J., Sasakova, N., & Placha, I. (2009). Pathogens and antibiotic residues in animal manures and hygienic and ecological risks related to subsequent land application. *Bioresource Technology*, 100(22), 5386-5391.
- Wang, Z.-J., Wo, S.-K., Wang, L., Lau, C. B. S., Lee, V. H. L., Chow, M. S. S., et al. (2009). Simultaneous quantification of active components in the herbs and products of Si-Wu-Tang by high performance liquid chromatography-mass spectrometry. *Journal of Pharmaceutical and Biomedical Analysis*, 50(2), 232-244.
- Ying, G.-G., Kookana, R. S., & Ru, Y.-J. (2002). Occurrence and fate of hormone steroids in the environment. *Environment International*, 28(6), 545-551.
- Zhang, M.-K., & Fang, L.-p. (2007). Effect of tillage, fertilizer and green manure cropping on soil quality at an abandoned brick making site. *Soil and Tillage Research*, 93(1), 87-93.
- Zimmer, D. (2003). Introduction to Quantitative Liquid Chromatography-Tandem Mass Spectrometry (LC-MS-MS). Chromatographia Supplement, 57, 325-332.
- Zou, S., Xu, W., Zhang, R., Tang, J., Chen, Y., & Zhang, G. (2011). Occurrence and distribution of antibiotics in coastal water of the Bohai Bay, China: Impacts of river discharge and aquaculture activities. *Environmental Pollution*, 159(10), 2913-2920.