

UNIVERSITI PUTRA MALAYSIA

IDENTIFICATION OF THERMOSTABLE PEPTIDE MARKERS IN MEATS USING GEL-BASED FRACTIONATION COUPLED WITH MASS SPECTROMETRY

NADIAH BINTI MAT JUNOH

IPPH 2019 8



IDENTIFICATION OF THERMOSTABLE PEPTIDE MARKERS IN MEATS USING GEL-BASED FRACTIONATION COUPLED WITH MASS SPECTROMETRY

By

NADIAH BINTI MAT JUNOH

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

November 2018

COPYRIGHT

All materials contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

G



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

IDENTIFICATION OF THERMOSTABLE PEPTIDE MARKERS IN MEATS USING GEL-BASED FRACTIONATION COUPLED WITH MASS SPECTROMETRY

By

NADIAH BINTI MAT JUNOH

November 2018

Chairman: Dhilia Udie Lamasudin, PhDInstitute: Halal Products Research Institute

Sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) followed by mass spectrometry is widely used in proteomic study mainly in meats due to their effectiveness and efficiency in generating reliable data. An alternative method such as enzyme-linked immunosorbent assay (ELISA), although known to be highly-specific but it bears the risk of cross-reactivity. The genomic approach is another commonly used method but suffers from potential deoxyribonucleic acid (DNA) contamination from other organism and due to its structure, DNA molecule is less resistant to heat treatment. The limitation of these methods may lead to a false positive result in detection analysis. First, the purpose of this study was to compare the electrophoretic pattern of proteins in one-dimensional (1DE) and two-dimensional (2DE) gel electrophoresis in meat derived from cow, water buffalo, pig and wild boar upon different heat treatments. Then, the next aim was to screen the species-specific thermostable protein markers in all species using principal component analysis (PCA) and partial least square-discriminant analysis (PLS-DA). The third aim was to identify the thermostable protein and peptide markers obtained from electrospray ionization liquid chromatography-mass spectrometry (ESI-LC-MS) and matrix-assisted laser desorption ionization time-of-flight/time-of-flight tandem mass spectrometry (MALDI-TOF/TOF-MS/MS). The meats were subjected into several heat treatments which were (1) chilled at 4°C for 30 min, (2) roasted at 150°C for 20 min, and (3) fried at 160°C for 6 min, before subjecting to two different combinations of proteomic approaches i.e. SDS-PAGE coupled with ESI-LC-MS and 2DE coupled with MALDI-TOF/TOF-MS/MS. The extracted proteins were fractionated using 1DE gel electrophoresis coupled with combined multivariate analysis of PCA and PLS-DA for grouping and discriminative analysis. The pattern of electrophoretic proteins in all species was similar but differences appeared between the raw and cooked meats. The potential thermostable protein markers derived from all species were determined using ESI-LC-MS. At the molecular weight of 55.06 kDa, proteins that have been identified from cow samples were tropomyosin, moesin, cadherin, and septin. As for water buffalo, 5-oxoprolinase has been identified at the molecular weight of 181.22 kDa. In the pig, serum albumin, calpain-7, and ATP synthase subunit alpha, mitochondrial were identified with the molecular weight of 77.02 kDa while wild boar has shown to have

cathepsin K and calcium/calmodulin-dependent protein kinase type II subunit delta at the molecular weight of 48.61 kDa. These approaches were successful in providing preliminary data analysis for the screening of thermostable protein markers with speciesspecificity. For the second approach, tropomyosin was selected and analyzed using 2DE gel electrophoresis followed by MALDI-TOF/TOF-MS/MS. Minor differences were observed in the amino acids sequences in both tropomyosin (TPM) isoforms i.e. TPM2 and TPM1 for each species. Moreover, several thermostable peptides that were found to belong to the Bovidae family were HIAEDSDR and LDKENAIDR and Suidae family, RIQLVEEELDR. These potential peptides could be used as markers to discriminate between Bovidae and Suidae families. Thus, the result indicated that these isoforms have the potential to be selected as thermostable species-specific protein and peptide markers in meat authentication.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

MENGENAL PASTI PENANDA PEPTIDA YANG TAHAN HABA DI DALAM DAGING DENGAN MENGGUNAKAN PEMISAHAN BERDASARKAN GEL DAN SPEKTROMETRI JISIM

Oleh

NADIAH BINTI MAT JUNOH

November 2018

Pengerusi: Dhilia Udie Lamasudin,PhDInstitut: Institut Penyelidikan Produk Halal

Kaedah natrium dodesil sulfat-elektroforesis gel poliakrilamida (SDS-PAGE) berserta dengan spektrometri jisim digunakan secara meluas dalam bidang proteomik bagi kajian daging kerana ia berkesan dan cekap dalam menghasilkan data yang dipercayai. Antara kaedah lain yang diguna pakai adalah antibodi menangkap enzim-esei imuno serapan (ELISA) yang mempunyai kespesifikan yang tinggi tetapi berisiko untuk berlakunya reaksi silang. Pendekatan genetik adalah antara kaedah lain yang digunakan, namun berpotensi untuk berlakunya pencemaran antara asid deoksiribonukleik (DNA) dengan organisma lain dan strukturnya sendiri yang tidak tahan dengan rawatan haba. Kelemahan dalam kaedah-kaedah ini boleh mengakibatkan penghasilan keputusan pengesanan analisis yang kurang tepat. Tujuan kajian ini dijalankan, pertama adalah untuk membandingkan corak elektroforetik protin dalam satu dimensi (1DE) dan dua dimensi (2DE) gel elecktoforesis dalam daging daripada lembu, kerbau, babi dan babi hutan setelah dikenakan rawatan haba yang berbeza. Kemudian, tujuan kedua adalah untuk melakukan pemeriksaan saringan bagi penanda protin yang stabil haba dan spesifik spesis bagi setiap spesis dengan menggunakan analisis komponen utama (PCA) dan analisis diskriminasi sebahagian kecil persegi (PLS-DA). Tujuan ketiga adalah untuk mengenal pasti penanda protin dan peptida yang stabil haba yang diperoleh daripada elektrospray pengionan kromatografi cecair spektrometri jisim (ESI-LC-MS) dan matrik dibantu laser desorpsi ionisasi-masa penerbangan/masa penerbangan spektrometri jisim (MALDI-TOF/TOF-MS/MS). Setiap daging dikenakan kepada beberapa rawatan haba iaitu (1) penyejukkan pada suhu 4°C selama 30 min, (2) panggang pada suhu 150°C selama 20 min, dan (3) goreng pada suhu 160°C selama 6 min sebelum melalui dua kaedah gabungan proteomik yang berbeza, SDS-PAGE berserta dengan ESI-LC-MS dan 2DE berserta dengan MALDI-TOF/TOF-MS/MS. Protin yang telah diekstrak, dipisahkan dengan menggunakan 1DE gel elektroforesis berserta dengan gabungan analisis multivariat, PCA untuk pembahagian kepada kumpulan dan PLS-DA untuk diskriminasi. Corak bagi elektroforetik protin bagi setiap spesis adalah sama tetapi berbeza antara daging yang tidak dimasak dan dimasak. Beberapa protin yang stabil haba telah dikenal pasti dengan menggunakan ESI-LC-MS dan berpotensi untuk dijadikan penanda bagi setiap spesis. Lembu mempunyai tropomyosin, moesin, kaderin dan septin pada berat molekular, 55.06 kDa. Kerbau mempunyai protin 5-oxoprolinase dengan berat molekular, 181.22 kDa. Babi pula memiliki serum albumin, calpain-7 dan ATP sintase subunit alpha, mitokondria pada berat molekular, 77.02 kDa manakala babi hutan memiliki cathepsin K dan calcium/calmodulin-dependent protein kinase type II subunit delta pada berat molekular, 48.61 kDa. Kaedah ini menyediakan data analisis awalan bagi penanda protin yang stabil haba untuk spesifik spesis. Bagi pendekatan kedua, tropomyosin telah dipilih dan dianalisis dengan menggunakan 2DE gel elektroforesis diikuti dengan MALDI-TOF/TOF-MS/MS. Perubahan kecil dalam urutan asid amino dapat dilihat dalam kedua-dua isoform tropomyosin (TPM), TPM2 and TPM1 bagi setiap spesis. Tambahan pula, beberapa peptida yang stabil haba yang dijumpai milik keluarga Bovidae adalah HIAEDSDR dan LDKENAIDR manakala keluarga Suidae adalah RIQLVEEELDR. Kesemua peptida ini berpotensi untuk dijadikan penanda bagi mendiskriminasikan antara keluarga Bovidae and Suidae. Keputusan menunjukkan kedua-dua isoform mempunyai potensi untuk dipilih sebagai penanda protin dan peptida yang tahan haba bagi spesifik spesis dalam pengesahan daging.

ACKNOWLEDGEMENTS

In the name of Allah, The Most Gracious and The Most Merciful. Alhamdulillah. First and foremost, I would like to express my highest gratitude to Allah S.W.T for His blessing by giving me a great opportunity and strength to complete this project successfully after several hardships.

I would like to extend my special appreciation to my outstanding supervisor, Dr. Dhilia Udie Lamasudin, may Allah bless her and her family for her remarkable patience in guiding me throughout my research study. I appreciate her willingness in providing me the valuable advice and supervision for the whole period of my study.

My sincere appreciation is also extended to my co-supervisors, Prof. Dr. Shuhaimi Mustafa, Dr. Amalia Mohd Hashim, and Assoc. Prof. Dr. Nazia Abdul Majid for their encouragement, motivation, and guidance. Their commitment and kindness are highly appreciated.

I also extend my sincere gratitude towards Dr. Siti Aimi Sarah, my teammates; Muhammad Hafis Yuswan Mohd Yusoff, Ahmad Afifullah Abdul Rahman and Farah Wahida Mohd Sabri. In addition, thanks to all my labmates; Ummu Nasuha Mohd Asri, Mohd Shirwan Abdullah Sani, Norsahida Azri, Suhana Mustafa, Nur Royhaila Mohamad, Farrah Fayyadah Borhan and Nur Izzah Azmi. Thanks also to Sarah Syazwani Shaifuddin, Yeni Mardhiah Syafrizal Shah and others for all their support and encouragement throughout these tough times until the end of this project.

Thanks to Halal Products Research Institute for providing me with all the necessary instruments that I need as well as to all their committed staff. Thank you to Universiti Putra Malaysia for funding my study and research until the end. May Allah bless all of you.

Last but not least, a world of thanks to my beloved parents, Mat Junoh Awang Kechik and Fauziah Kamarudin for their prayers, understanding and supporting me. Thanks also to my lovely siblings; Muhammad Safwan, Munirah, Solehah, Syahidah, Abdul Rahman, Nurul Husna and Muhammad Aiman for their encouragement and wisdom towards everything I did. Thank you so much. This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Dhilia Udie Lamasudin, PhD

Senior Lecturer Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Shuhaimi Bin Mustafa, PhD

Professor Halal Product Research Institute Universiti Putra Malaysia (Member)

Amalia Binti Mohd Hashim, PhD

Senior Lecturer Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

Nazia Binti Abdul Majid, PhD

Associate Professor Faculty of Science University of Malaya (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:
Name and Matric No.:	Nadiah binti Mat Junoh (GS 44326)

Declaration by Members of Supervisory Committee

This is to confirm that:

C

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:	
Name of Chairman of Supervisory Committee: <u>Dr. Dhilia Udie Lamasudin</u>	
Signature:	
Name of Member of Supervisory Committee: <u>Prof. Dr. Shuhaimi Bin Mustafa</u>	
Signature:	
Name of Chairman of	
Supervisory Committee: Dr. Amalia Binti Mohd Hashim	
Signature:	
Name of Member of	
	:4
Supervisory Committee: Assoc. Prof. Dr. Nazia Binti Abdul Maj	<u>10</u>

TABLE OF CONTENTS

			Page
ABSTR	ACT		i
ABSTR			iii
		GEMENTS	v
APPRO			vi
	RATIO		viii
	F TABL		xii
	F FIGU		xiii
LIST O	F ABBR	EVIATIONS	XV
СНАРТ			
1		RODUCTION	1
	1.1	Research Objectives	3
2		ERATURE REVIEW	4
	2.1	Halal Meat	4
	2.2	Meat	7
		2.2.1 Meat Definition	7
		2.2.2 Chemical Composition of The Meat	7
		2.2.3 The Structure of Meat	8
	2.3	2.2.4 Conversion of Muscle to Meat	11 12
	2.5 2.4	Cooking-induced Protein Denaturation Species Identification	12
	2.4	Biomarkers	15
	2.5	Multivariate Analysis	16
	2.0	Proteomic Approaches	10
	2.7	2.7.1 Electrophoretic Gel	17
		2.7.1.1 Sodium Dodecyl Sulfate-Polyacrylamide	17
		Gel Electrophoresis (SDS-PAGE)	17
		2.7.1.2 Isoelectric Focusing (IEF)	18
		2.7.2 Mass Spectrometry	19
		2.7.2.1 Liquid Chromatography-Mass Spectrometry	
		(LC-MS)	22
		2.7.2.2 Matrix-assisted Laser Desorption/Ionization	
		Time of Flight Mass Spectrometry	
		(MALDI-TOF-MS)	23
	2.8	Proteomic Application	24
3	MFT	THODOLOGY	27
5	3.1	Materials	27
	3.2	Heat Treatments of The Sample	28
	3.3	Meat Protein Extraction	28
	3.4	Protein Concentration Determination	29
	3.5	Protein Fractionation	29
	0.0	3.5.1 One-Dimensional Gel Electrophoresis (1DE)	29
		3.5.1.1 SDS-PAGE Gel Preparation	29
		3.5.1.2 Sample Loading	29
		=	

PUBLIC	ATION		87
BIODAT		TUDENT	86
APPEND			84
REFERE	NCES		71
5	CONC	LUSION AND RECOMMENDATION	70
		using MALDI-TOF/TOF-MS/MS	61
	4.7	Identification of The Potential Thermostable Peptide Markers	<u></u>
	4.6	Tropomyosin as The Potential Thermostable Protein Marker	52
	4.5	Protein Identification by using ESI-LC-MS	47
	4.4	Partial Least Square-Discriminant Analysis (PLS-DA)	45
	4.3	Principal Component Analysis (PCA)	41
	7.4	Species	37
	4.1 4.2	Optimization of The Extraction Buffers in SDS-PAGE Comparison of The Electrophoretic Pattern of Proteins among	34
4		LTS AND DISCUSSION	34
Λ	DECT		
		3.8.2 MASCOT Analysis for MALDI-TOF/TOF-MS/MS	33
	5.0	3.8.1 MS-FIT Analysis for ESI-LC-MS	32
	3.8	Database Search and Sequence Analysis	32
		3.7.1 ESI-LC-MS Analysis3.7.2 MALDI-TOF/TOF-MS/MS Analysis	32 32
	3.7	Protein Identification	32 32
	3.6	In Gel Digestion	31
		3.5.2.2 ImageAnalysis	31
		3.5.2.1 2DE Gel Preparation	30
		3.5.2 Two-Dimensional Gel Electrophoresis (2DE)	30
		3.5.1.3 Multivariate Analysis	30

 \bigcirc

LIST OF TABLES

Table		Page
2.1	Summary of Proteomic Approaches for Species-Specific Peptide Markers Identification	26
4.1	Identification of The Thermostable Protein Markers from Every Species using ESI-LC-MS	49
4.2	The Number of The Total Protein Spots Present in 2DE Images from Every Species across The Heat Treatments	53
4.3	The Molecular Weights (kDa) of TPM2 and TPM1 Present in 2DE Images from Every Species across The Heat Treatments	54
4.4	The Spot Volumes (OD*mm ²) of TPM2 and TPM1 Present in 2DE Images from Every Species	56
4.5	Identification of Protein Spots of TPM2 and TPM1 from Every Species using Peptide Finger Printing (MALDI-TOF/TOF-MS)	62
4.6	Identification of Peptides of TPM2 and TPM1 from Every Species using Tandem Mass Spectrometry (MALDI-TOF/TOF-MS/MS)	64
4.7	The Potential Thermostable Peptide Markers from Every Species	68

G

LIST OF FIGURES

Figur	e Pa	age
2.1	The Recognized Foreign Halal Logos Certified by JAKIM	6
2.2	Transverse Section of Skeletal Muscle	9
2.3	Striated Banding Pattern of Skeletal-Muscle Fibers	10
2.4	Family of Suidae from (a) Domesticated Pig and (b) Wild Boar	13
2.5	Family of Bovidae from (a) Cow and (b) Water Buffalo	14
2.6	Diagram of Two-Dimensional Gel Electrophoresis using IEF Followed by SDS-PAGE for Protein Fractionation	19
2.7	Basic Components of Mass Spectrometry	20
2.8	Combination of Quadrupole and Time of Flight in Mass Spectrometry	21
2.9	Diagram of Linear Ion Trap using ESI	23
2.10	Diagram of MALDI-TOF-MS	24
3.1	Workflow for Species-Specific Protein Identification	27
4.1	Electrophoretic Pattern of The Proteins Extracted from The First Supernatan of Raw and Cooked Beef Samples using Two Types of Extraction Buffers	t 35
4.2	Electrophoretic Pattern of The Proteins Extracted from The Second Supernat of Raw and Cooked Beef Samples using Two Types of Extraction Buffers	tant 36
4.3	The Optimization of The First Supernatant using Tris-HCl for Four Species in All Treatments	38
4.4	The Optimization of The Second Supernatant using Tris-HCl, Urea and Thiourea for Four Species in All Treatments	40
4.5	Score Plot of PC1 and PC2	42
4.6	Loading Plot of PC1 and PC2	44
4.7	The Correlation of PC1 and PC2 According to Species	46
4.8	Images of 2DE Gels from Chilled Pork	55

6

4.9	Images of 2DE Gels Represent Beef Derived from Chilled, Roasted and Fried Treatments	57
4.10	Images of 2DE Gels Represent Water Buffalo Meats Derived from Chilled, Roasted and Fried Treatments	58
4.11	Images of 2DE Gels Represent Pork Derived from Chilled, Roasted and Fried Treatments	59
4.12	Images of 2DE Gels Represent Wild Boar Meats Derived from Chilled, Roasted and Fried Treatments	60
4.13	Pairwise Sequence Alignment of TPM2 for Pig and Bovine (using Cow Database for Bovine)	66
4.14	Pairwise Sequence Alignment of TPM1 for Pig and Bovine (using Cow Database for Bovine)	67

 \bigcirc

LIST OF ABBREVIATIONS

	1DE	One-dimensional
	2DE	Two-dimensional
	ACN	Acetonitrile
	ADP	Adenosine diphosphate
	APCI	Atmospheric pressure chemical ionization
	APS	Ammonium persulfate
	ATP	Adenosine triphosphate
	BSA	Bovine serum albumin
	CaMKII	Ca ²⁺ calmodulin (CaM)-dependent protein kinase
	CHAPS	3-[3-cholamidopropyl)dimethylammonio]-1- propanesulfate
	CI	Chemical ionisation
	CID	Collision-induced dissociation
	СР	Creatine phosphate
	CWF	Compassion in World Farming
	DHAP	2,5-dihydroxyacetophenone
	DHB	2,5-dihydroxybenzoic acid
	DNA	Deoxyribonucleic acid
	DTT	Dithiothreitol
	EI	Electron impact
	ELISA	Enzyme-linked immunosorbent assay
	ERM	Ezrin/radixin/moesin
	ESI	Electrospray ionization
	ESI-LC-MS	Electrospray ionization liquid chromatography- mass spectrometry
\bigcirc	FAB	Fast atom bombardment
	FAO	Food and Agriculture Organization
	FD/FI	Field desorption/Field ionization
	GC	Gas chromatography

	HCBs	Halal Certification Bodies
	HCCA	4-hydroxy-α-cyanocinnamic acid
	HCL	Hydrochloric acid
	HPA	3-hydroxypicolinic acid
	IAA	Iodoacetamide
	IDA	Information Dependent Acquisition
	IEF	Isoelectric focusing
	IPG	Immobilized pH gradient
	IUPAC	International Union of Pure and Applied Chemistry
	JAKIM	Jabatan Kemajuan Islam Malaysia
	K ₂ HPO ₄	Dipotassium hydrogenphosphate
	KCl	Potassium chloride
	kDa	Kilodalton
	KH ₂ PO ₄	Potassium dihydrogen phosphate
	LC	Liquid chromatography
	LC-MS	Liquid chromatography-mass spectrometry
	m/z	mass to charge ratio
	MALDI	Matrix-assisted laser desorption ionisation
	MALDI-TOF-MS/MS	Matrix-assisted laser desorption/ionization time of flight tandem mass spectrometry
	MLC1f	Myosin light chain 1 isoform
	MLC2f	Myosin light chain 2 isoform
	MLC3f	Myosin light chain 3 isoform
	MRM	Multiple monitoring reaction
	MUI	Majelis Ulama Indonesia
	MUIS	Majlis Ugama Islam Singapura
(\mathbf{C})	OPLS-DA	Orthogonal projections to latent structure discriminant analysis
	PC1	Principal component 1
	PC2	Principal component 2
	PCA	Principal component analysis
		xvi

PCR	Polymerase chain reaction
PCR-RFLP	Polymerase chain reaction restriction fragment length polymorphism
PLS-DA	Partial least square-discriminant analysis
PMF	Peptide finger printing
Q	Quadrupole
Q-TOF	Quadrupole time of flight
RAPD	Random amplified polymorphic deoxyribonucleic acid
SA	Sinapinic acid
SDS-PAGE	Sodium dodecyl sulphate-polyacrylamide gel electrophoresis
SES	Socio-economic status
S-S	Sulphur-sulphur
TEMED	N,N,N',N'-tetramethylethylenediamine
TI	Thermospray ionization
TPM	Tropomyosin
TPM1	Tropomyosin alpha-1 chain
TPM2	Tropomyosin beta chain
UAE	United Arab Emirates
UK	United Kingdom
WHO	World Health Organization

 \bigcirc

CHAPTER 1

INTRODUCTION

In recent decades, livestock production has shown to be increasing dynamically due to the high demand for meat. The situation is a result of global population's modern lifestyle where food preparation has to be done quickly and easily, hence consumers incline to include in their diet with fast food which contains mainly vegetable oils, sugar and animal products over the carbohydrate-rich staple food and this transitional phenomenon is known as the substitution stage (Vranken, Avermaete, Petalios, & Mathijs, 2014). This transition in the diet which mainly consists of fast food and frozen food, as well as the increasing purchasing power and urbanization process has directly influenced the demand for meat supply. In general, livestock production in developing countries is considered to be growing rapidly. However, the situation is contradicting in developed countries where the production of livestock is slow or stagnant even at the high-income levels (Thornton, 2010; Vranken et al., 2014).

Referring to a report released by World Bank 2009, the total number of meat production in the developing countries was tripled from 45 to 134 million tons since the year 1980 to 2002 (Thornton, 2010). Its number is expected to be increased in 2030 by achieving 37 kg from a modest average annual per capita consumption of 10 kg in the 1960s (Heinz & Hautzinger, 2007). In general, the pattern of meat consumption is differed geographically, due to diet regulation, religion or the economic situation. In developed countries such as the United State and the United Kingdom, the vital meat sources are mostly obtained from pig, sheep, and cattle. Moreover, almost half of all meat being consumed is derived from meat products (Kearney, 2010). In the developing country, the greatest livestock production in East Asia region is revolved between poultry and pig (Thornton, 2010). In other regions such as Malaysia, poultry is the major proportion of meat consumption followed by pig and cattle (Sheng, Shamsudin, Mohamed, Abdullah, & Radam, 2010).

Recent years, consumers' awareness of the authenticity of meat being sold in the market has been evolving ever since the horsemeat scandals have been publicized in the media and followed by other authentication issues. According to BBC News (2013b), halal chicken sausages which were served in several primary schools in London was claimed to contain pork DNA. Moreover, The Local (2013) has also reported that pork has been found in halal-marked salami in Swedish markets with surprisingly more than 10% of the meat content came from pork. In Jakarta, bakso, which is the local traditional delicacy, which it serves together with meatballs, has been publicized in the media that the meatballs contained beef contaminated with pork. It was reported that such cases emerged as a result of the increasing price of beef being sold in Indonesia (ABC Rural, 2012). The price of pork is comparatively cheaper than other livestock animals, as result, pork is commonly used as an alternative to replace other meats such as chicken, goat, and beef (Mutalib et al., 2012; Von Bargen, Dojahn, Waidelich, Humpf, & Brockmeyer, 2013). In Malaysia, the issue has been raised on the way of the storage of lamb and pork in one container, although the meats had been packed individually according to their species in different boxes (BH Online, 2017).

The awareness is emerged due to cultural practice, religious beliefs, gender, health and socio-economic status (SES) (Kearney, 2010; Vranken et al., 2014). For example, pork and beef are forbidden in Hinduism and Buddhism while pork is forbidden in Islam and Judaism (Bonne & Verbeke, 2008). The global population of Muslims is estimated to be around 1.6 to 1.8 billion and the figure is expected to be growing every year. In 2030, it is expected that the population to be increased up to 27% of the total global population. The growing population of Muslims will have a direct impact on the growth of the Halal industry (Farouk, 2012; 2013). A large number of Muslim population predetermined the importance of meat authenticity among consumers as they do not consume pork and its derivative in their diet. These issues must be taken seriously and to be addressed accordingly to regain consumers' trust in local meat supply and products. In addition, consumers nowadays are well-aware of their rights and it is their right to demand that the food that they consumed must be accurately labelled and the correct ingredients must be declared on the packaging of the products.

The development of an efficient and sensitive method for tracing the origin of the food products is of utmost importance as the total negligence on the issue has a big impact on many aspects. Various studies have been performed to find the most efficient, highly sensitive, time-saving and cost-effective methods for food authentication purposes. The desirable methods should be applicable to both heat-treated and nonheat-treated meat products. It is difficult to differentiate the species through naked eyes such as cow, pig, sheep, and poultry when they are mixed together. Meat authentication can be performed by using genomic and proteomic approaches such as polymerase chain reaction (PCR), ELISA and mass spectrometry.

The genomic approach is commonly practiced by using polymerase chain reactionrestriction fragment length polymorphism (PCR-RFLP), real-time PCR, species-specific PCR and random amplified polymorphic DNA (RAPD) to determine the species identification (Nakyinsige, Man, & Sazili, 2012). It is selected due to stable molecules and the ability to withstand heat processing (Chisholm, Conyers, Booth, Lawley, & Hird, 2005). However, the extraction method for PCR is time-consuming and the potential for DNA contamination with other organisms (Montowska, Alexander, Tucker, & Barrett, 2014). Moreover, DNA analysis also may provide low-reliability result towards processed food as its structure will be degraded at the temperature above 100°C and provide the opportunity for having nonspecific fragments (Ebbehoj & Thomsen, 1991; Sentandreu, Fraser, Halket, Patel, & Bramley, 2010).

Limitations in the genomic approach have encouraged researchers to choose the proteomic approach due to the stability of the primary amino acid sequence (Sentandreu et al., 2010). Several common methods used in proteomic approach are ELISA, electrophoretic and chromatographic. ELISA, although possess a high-specify, it has the risk for cross-reactivity (Chen & Hsieh, 2015). Furthermore, target proteins may denature during food processing and subsequently destroyed the target protein epitope which is the binding domain for the antibodies (Asensio, González, García, & Martín, 2008). Most of the studies are preferred electrophoretic approach using SDS-PAGE followed by mass spectrometry because they provide more extensive, thorough and comprehensive data and information for meat authentication analysis (Montowska & Pospiech, 2012; 2013b; Sarah, Karsani, Amin, Mokhtar, & Sazili, 2014). In order to improvise the previous method, multivariate analysis was integrated into this study. It

2

provides preliminary screening data which further facilitate the downstream analysis (Nur Azira, Che Man, Raja Mohd Hafidz, Aina, & Amin, 2014).

This study proved the hypothesis the types of proteins that have been present in 1DE and 2DE in meats derived from cow, water buffalo, pig and wild boar upon heat treatments, although between same species, were varied due to the different chemical solutions and mechanical treatments have been used in the extraction and fractionation steps in both electrophoresis and the mass spectrometries. This study also demonstrated that multivariate analysis using PCA and PLS-DA have facilitated the screening of species-specific thermostable protein markers prior to the mass spectrometry analysis. Two types of mass spectrometries i.e. ESI-LC-MS and MALDI-TOF/TOF-MS/MS were identified the thermostable protein and peptide markers from these species.

1.1 Research Objectives

General objective:

• To determine the potential thermostable protein and peptide markers from meats via two different approaches i.e. (1) SDS-PAGE coupled with ESI-LC-MS and (2) 2DE coupled with MALDI-TOF/TOF tandem mass spectrometry.

Specific objectives:

- 1. To compare the electrophoretic pattern of proteins in 1DE and 2DE gel electrophoresis in meats derived from cow, water buffalo, pig and wild boar upon different heat treatments.
- 2. To screen the species-specific thermostable protein markers in all species using PCA and PLS-DA.
- 3. To identify the thermostable protein and peptide markers obtained from two mass spectrometries:
 - a. ESI-LC-MS.
 - b. MALDI-TOF/TOF-MS/MS.

REFERENCES

- ABC Rural (2012, 21 December). People arrested in Indonesia for allegedly sneaking pork in meatballs. Retrieved from http://www.abc.net.au/site-archive/rural/news/content/201212/s3659232.htm
- Abdi, H., & Lynne, J. W. (2010). Principle Component Analysis. Wiley Interdisciplinary Reviews: Computational Statistics, 2(4), 433–459.
- Adeyemi, K. D., Mislan, N., Aghwan, Z. A., Sarah, S. A., & Sazili, A. Q. (2014). Myofibrillar protein profile of Pectoralis major muscle in broiler chickens subjected to different freezing and thawing methods. *International Food Research Journal*, 21(3), 1125–1129.
- Agilent Technologies (2001). Basics of LC/MS. Retrieved from http://ccc.chem.pitt.edu/wipf/Agilent%20LC-MS%20primer.pdf
- Agilent Technologies (2007). Considerations for Selecting GC/MS or LC/MS for Metabolomics. Retrieved from https://pdfs.semanticscholar.org/dbe6/b9e6ef4f73d40efc95e828b08103c02c57e7. pdf
- Aravena, P., & Skewes, O. (2007). European Wild Boar Purebred and Sus scrofa Intercrosses. Discrimination Proposals. A Review. Agro-Ciencia, 23(3), 133–147.
- Ardrey, R. E. (2003). Liquid Chromatography Mass Spectrometry: An Introduction (Vol. 1). John Wiley & Sons, Ltd. pp 1-233.
- Asensio, L., González, I., García, T., & Martín, R. (2008). Determination of food authenticity by enzyme-linked immunosorbent assay (ELISA). *Food Control*, 19(1), 1–8.
- Ashkenazy, H., Erez, E., Martz, E., Pupko, T., & Ben-Tal, N. (2010). ConSurf 2010: Calculating evolutionary conservation in sequence and structure of proteins and nucleic acids. *Nucleic Acids Research*, 38(SUPPL. 2), 529–533.
- Ayaz, Y., Ayaz, N. D., & Erol, I. (2005). Detection of species in meat and meat products using enzyme-linked immunosorbent assay. *Journal of Muscle Foods*, *17*(2006), 214–220.
- BBC News (2013a, 1 March). Horsemeat scandal: Four new products test positive. Retrieved from http://www.bbc.com/news/uk-21631961
- BBC News (2013b, 14 March). Pork DNA in halal sausages at Westminster primary school. Retrieved from http://www.bbc.com/news/uk-england-london-21791513
- BH Online (2017, 17 July). 4 kontena daging kambing campur babi dirampas. Retrieved from https://www.bharian.com.my/berita/nasional/2017/07/303117/4-kontena-daging-kambing-campur-babi-dirampas

- Becker, W.M., Kleinsmith, L.J., Hardin, J., & Bertoni, G.P. (2009). *The World of the Cell*. 7th Edition. Pearson International Edition.
- Bonne, K., & Verbeke, W. (2008). Religious values informing halal meat production and the control and delivery of halal credence quality. *Agriculture and Human Values*, 25, 35–47.
- Borghese, A. (Ed.)(2005). Buffalo meat and meat industry. In *Buffalo Production and Research* (pp.197-218). Italy.
- Bouley, J., Chambon, C., & Picard, B. (2004). Mapping of bovine skeletal muscle proteins using two-dimensional gel electrophoresis and mass spectrometry. *Proteomics*, 4(6), 1811–1824.
- Boye, J.I., Ma, C.Y., & Harwalkar, V.R. (1997). Thermal denaturation and coagulation of proteins. *A.Food Sci Techno*, 80, 25-6.
- Breidenstein, B. C., Kinsman, D. M., & Kotula, A. W. (1994). *Muscle Foods Meat Poultry and Seafood Technology*. Chapman and Hall. Boston, MA.
- Brodmann, P. D., & Moor, D. (2003). Sensitive and semi-quantitative TaqMan TM realtime polymerase chain reaction systems for the detection of beef (*Bos taurus*) and the detection of the family Mammalia in food and feed. *Meat Science*, 65, 599– 607.
- Buckley, M., Melton, N. D., & Montgomery, J. (2013). Proteomics analysis of ancient food vessel stitching reveals>4000-year-old milk protein. *Rapid Communications* in Mass Spectrometry, 27(4), 531–538.
- Burns E. R. & Cave, M. D. (2002). Histology and Cell Biology. Mosby.
- Cai, Z., & Liu, S. (Eds) (2013). Applications of MALDI-TOF Spectroscopy. Topics in Current Chemistry. Springer. pp.1-210.
- Cañeque, V., Pérez, C., Velasco, S., Diaz, M. T., Lauzurica, S., Alvarez, I., ... De la Fuente, J. (2004). Carcass and meat quality of light lambs using principal component analysis. *Meat Science*, 67(4), 595–605.
- Chan, X.C.Y., McDermott, J. C., & Michael Siu, K.W. (2007). Identification of Secreted Proteins during Skeletal Muscle Development. *Journal of Proteome Research*, 6(2), 698–710.
- Chen, F. C., & Hsieh, Y. H. (2015). Detection of pork in heat-processed meat products by monoclonal antibody-based ELISA. *Journal of AOAC International*, 83(1), 79–85.
- Chen, F.C., & Hsieh, Y.H.P. (2002). Porcine troponin I : a thermostable species marker protein. *Meat Science*, *61*, 55–60.
- Chisholm, J., Conyers, C., Booth, C., Lawley, W., & Hird, H. (2005). The detection of horse and donkey using real-time PCR. *Meat Science*, 70, 727–732.

- Chiter, A., Forbes, J. M., & Blair, G. E. (2000). DNA stability in plant tissues: Implications for the possible transfer of genes from genetically modified food. *FEBS Letters*, 481(2), 164–168.
- Clark, B. N., & Gutstein, H. B. (2008). The myth of automated, high-throughput twodimensional gel analysis. *Proteomics*, 8(6), 1197–1203.
- Compassion in World Farming (2013, 20 May). The Life of-Pigs. Retrieved from https://www.ciwf.org.uk/media/5235118/The-life-of-Pigs.pdf
- Creative Proteomics (2018). 2D Electrophoresis. Retrieved from https://www.creativeproteomics.com/services/2d-electrophoresis.htm
- Cruz-Romero, M. C., & Kerry, J. P. (2011). Packaging of cooked meats and musclebased, convenience-style processed foods. In J.P. Kerry & J.F. Kerry (Eds), *Processed Meats* (pp. 666–705). The avi Publishing Company, Connecticut.
- Da Silva Campos, N., De Sá Oliveira, K., Almeida, M. R., Stephani, R., & De Oliveira, L. F. C. (2014). Classification of Frankfurters by FT-Raman spectroscopy and chemometric methods. *Molecules*, 19(11), 18980–18992.
- Dahlan, H. M., Karsani, S. A., Abdul Rahman, M., Abdul Hamid, N. A., Mat Top, A. G., & Wan Ngah, W. Z. (2012). Proteomic analysis reveals that treatment with tocotrienols reverses the effect of H 2O 2 exposure on peroxiredoxin expression in human lymphocytes from young and old individuals. *Journal of Nutritional Biochemistry*, 23(7), 741–751.
- Dass, C. (2006). Fundamentals of Contemporary Mass Spectrometry. Wiley-Interscience.
- Davis, P. J., & Williams, S. C. (1998). Protein modification by thermal processing. *Allergy*, 53, 102–105.
- De Liu, X., Jayasena, D. D., Jung, Y., Jung, S., Kang, B. S., Heo, K. N., ... Jo, C. (2012). Differential Proteome Analysis of Breast and Thigh Muscles between Korean Native Chickens and Commercial Broilers. *Asian-Australasian Journal of Animal Sciences*, 25(6), 895–902.
- Destefanis, G., Barge, M. T., Brugiapaglia, A., & Tassone, S. (2000). The use of principal component analysis (PCA) to characterize beef. *Meat Science*, *56*, 255-259.
- Doecke, J. D. E. A. (2012). Blood-based protein biomarkers for diagnosis of Alzheimer disease. Archives of Neurology, 69(10), 1318–1325.
- Doherty, M. K., McLean, L., Hayter, J. R., Pratt, J. M., Robertson, D. H. L., El-Shafei, A., ... Beynon, R. J. (2004). The proteome of chicken skeletal muscle: changes in soluble protein expression during growth in a layer strain. *Proteomics*, 4(7), 2082–2093.
- Ebbehoj, K. F., & Thomsen, P. D. (1991). Species Differentiation of Heated Meat Products by D N A Hybridization. *Meat Science*, *30*, 221–234.

- Encyclopedia of Life (n.d). Mammalia. Retrieved from http://eol.org/pages/1642/media?page=8&sort_by=status&status%5B%5D=all&t ype%5B%5D=image&utf8=%E2%9C%93
- Ekvall, K. (2012). Development of LC-QTOF method for analysis of extracts from urinary catheters. Uppsala University, Sweden.
- Faila, S., Barone, C. M. A., & Antonio, B. (2012). Carcass, meat quality and products. In A. Borghese (Ed.), *Buffalo livestock and products* (pp. 260-283). Food and Agriculture Organization of the United Nations.
- Farouk, M. M. (2013). Advances in the industrial production of halal and kosher red meat. MESC, 95(4), 805–820.
- Farouk, M. M. (2012). New Zealand meat industry must optimise halal market opportunities. NZ Food Technology, 47(8), 9.
- Feiner, G. (2006). *Meat products handbook practical science and technology*. Woodhead Publishing Limited, Cambridge, England.
- FAO (2014, 25 November). Sources of Meat. Retrieved from http://www.fao.org/ag/againfo/themes/en/meat/backgr_sources.html
- FAO (1994). A manual for the primary animal health care worker. Retrieved from http://www.fao.org/docrep/t0690e/t0690e00.htm#Contents
- Fuseini, A., Wotton, S. B., Knowles, T. G., & Hadley, P. J. (2017). Halal Meat Fraud and Safety Issues in the UK: a Review in the Context of the European Union. *Food Ethics*. Doi: 10.1007/s41055-017-0009-1.
- Giordano, G., Guarini, P., Ferrari, P., Schiavone, B., & Giordano, A. (2010). Beneficial impact on cardiovascular risk profile of water buffalo meat consumption. *European Journal of Clinical Nutrition*, 64(9), 1000–1006.
- Giovannacci, I., Guizard, C., Carlier, M., Duval, V., Martin, J.L., & Demeulemester, C. (2004). Species identification of meat products by ELISA. *International Journal* of Food Science and Technology, 39, 863–867.
- Giuffra, E., Kijas, J. M. H., Amarger, V., Carlborg, Ö., Jeon, J. T., & Andersson, L. (2000). The origin of the domestic pig: Independent domestication and subsequent introgression. *Genetics*, 154(4), 1785–1791.
- Guntarti, A., Martono, S., Yuswanto, A., & Rohman, A. (2015). FTIR Spectroscopy in Combination with Chemometrics for Analysis of Wild Boar Meat in Meatball Formulation. *Asian Journal of Biochemistry*, *10*(4), 165–172.
- Halal Hub Division JAKIM (2018). The Recognised Foreign Halal Certification Bodies & Authorities as at February 8th, 2018, 1–54. Retrieved from http://www.halal.gov.my/ckfinder/userfiles/files/cb/SENARAI%20CB%20TERK INI%20SETAKAT%20FEBRUARI%20%202018.pdf

- Heinz, G., & Hautzinger, P. (2007). *Meat processing technology for small-to medium scale producers*. Rap publication 2007/20, Food and agriculture organization of the united nations regional office for asia and the pacific Bangkok, 2007. Retrieved from http://www.fao.org/docrep/010/ai407e/AI407E00.htm#Contents
- Holle, A., Haase, A., Kayser, M., & Höhndorf, J. (2006). Optimizing UV laser focus profiles for improved MALDI performance. *Journal of Mass Spectrometry*, 41(6), 705–716.
- Huang, M., & Ochiai, Y. (2005). Fish fast skeletal muscle tropomyosins show speciesspecific thermal stability. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 141(4), 461–471.
- Hudmon, A., & Schulman, H. (2002). Structure-function of the multifunctional Ca2+/calmodulin-dependent protein kinase II. *The Biochemical Journal*, 364, 593–611.
- Huff-Lonergan, E., & Lonergan, S. M. (2005). Mechanisms of water-holding capacity of meat: The role of postmortem biochemical and structural changes. *Meat Science*, *71*(1), 194–204.
- Issaq, H. J., Veenstra, T. D., Conrads, T. P., & Felschow, D. (2002). The SELDI-TOF MS approach to proteomics: protein profiling and biomarker identification. *Biochemical and Biophysical Research Communications*, 292(3), 587–592.
- Ivetic, A., & Ridley, A. J. (2004). Ezrin/radixin/moesin proteins and Rho GTPase signalling in leucocytes. *Immunology*, 112(2), 165–176.
- Johnston, D.E., Knight, M.K., & Ledward, D.A. (1992). *The Chemistry of muscle-based foods*. Royal Society of Chemistry, Cambridge.
- Kara, D. (2009). Evaluation of trace metal concentrations in some herbs and herbal teas by principal component analysis. *Food Chemistry*, *114*, 347–354.
- Kearney, J. (2010). Food consumption trends and drivers. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 365(1554), 2793–2807.
- Kerry, J. F. (2011). Effects of novel thermal processing technologies on the sensory quality of meat and meat products. In J.P. Kerry & J.F. Kerry (Eds), *Processed Meats Improving Safety, Nutrition and Quality* (pp. 617–665). The avi Publishing Company, Connecticut.
- Kesmen, Z., Sahin, F., & Yetim, H. (2007). PCR assay for the identification of animal species in cooked sausages. *Meat Science*, 77, 649–653.
- Kierszenbaum, A. L. & Tres, L.L. (2007). *Histology and cell biology: an introduction to pathology*. Elesivier Saunders.
- Kim, G. D., Seo, J. K., Yum, H. W., Jeong, J. Y., & Yang, H. S. (2017). Protein markers for discrimination of meat species in raw beef, pork and poultry and their mixtures. *Food Chemistry*, 217, 163–170.

- Kinoshita, M. (2003). Assembly of Mammalian Septins. The Japanese Biochemical Society, 134(4), 491–496.
- Kopuzlu, S., Onenc, A., Bilgin, O. C., & Esenbuga, N. (2011). Determination of meat quality through principal components analysis. *The Journal of Animal & Plant Sciences*, 21(2), 151–156.
- Kumar, N., Bansal, A., Sarma, G. S., & Rawal, R. K. (2014). Talanta Chemometrics tools used in analytical chemistry : An overview. *Talanta*, 123, 186–199.
- La Neve, F., Civera, T., Mucci, N., & Bottero, M. T. (2008). Authentication of meat from game and domestic species by SNaPshot minisequencing analysis. *Meat Science*, 80(2), 216–224.
- Larson, G., Dobney, K., Albarella, U., Fang, M., Matisoo-smith, E., Robins, J., ... Cooper, A. (2005). Worldwide Phylogeography of Wild Boar Reveals Multiple Centers of Pig Domestication. *Science*, 30, 1618-1621.
- Lawrie, R. A., & Ledward, D. A. (2006). *Lawrie's meat science* (7th Edition). Woodhead Publishing Limited.
- Lee, S. *Modern Mass Spectrometry*. Paper presented in at the meeting of the MacMillan Group Meeting, 2005.
- Live Science (2015, 24 April). Pigs, Hogs & Boars: Facts About Swine. Retrieved from https://www.livescience.com/50623-pigs-facts.html
- Linseisen, J., Kesse, E., Slimani, N., Bueno-de-Mesquita, H., Ocké, M., Skeie, G., ... Riboli, E. (2002). Meat consumption in the European Prospective Investigation into Cancer and Nutrition (EPIC) cohorts: results from 24-hour dietary recalls. *Public Health Nutrition*, 5(6b), 1243-1258.
- Liu, Y., Lyon, B. G., Windham, W. R., Lyon, C. E., & Savage, E. M. (2004). Principal component analysis of physical, color, and sensory characteristics of chicken breasts deboned at two, four, six, and twenty-four hours postmortem. *Poultry Science*, 83, 101–108.
- López, J. L., Marina, A., Alvarez, G., & Vázquez, J. (2002). Application of proteomics for fast identification of species-specific peptides from marine species. *Proteomics*, 2(12), 1658–1665.

Magdeldin, S. (Ed) (2012). Gel Electrophoresis - Principles and Basics. Intech.

- Marina, A. M., Man, Y. B. C., & Amin, I. (2010). Use of the SAW sensor electronic nose for detecting the adulteration of virgin coconut oil with RBD palm kernel olein. *JAOCS, Journal of the American Oil Chemists' Society*, 87(3), 263–270.
- Marques, A. Y. C., Maróstica, M. R., & Pastore, G. M. (2010). Some nutritional, technological and environmental advances in the use of enzymes in meat products. *Enzyme Research*, 2010, 480923. Doi: 10.4061/2010/480923.

- Martinez, I., & Friis, T. J. (2004). Application of proteome analysis to seafood authentication. *Proteomics*, 4(2), 347–354.
- Marvin, L. F., Roberts, M. A., & Fay, L. B. (2003). Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry in clinical chemistry. *Clinica Chimica Acta*, 337(1–2), 11–21.
- Mazzeo, M. F., De Giulio, B., Guerriero, G., Ciarcia, G., Malorni, A., Russo, G. L., & Siciliano, R. A. (2008). Fish authentication by MALDI-TOF mass spectrometry. *Journal of Agricultural and Food Chemistry*, 56(23), 11071–11076.
- Meijaard, E., d'Huart, J.P., & Oliver, W.R. (2016). Family Suidae (Pigs). In W. Mittermeier, *Handbook of the Mammals of the World* (pp. 248-291). Lynx Edicions.
- Meister, A. (1988). Glutathione metabolism and its selective modification. *The Journal* of *Biological Chemistry*, 263(33), 17205–17208.
- Melville, J. (2014). *Matrix-Assisted Laser Desorption / Ionization (MALDI)*. University of California.
- Montowska, M., Alexander, M. R., Tucker, G. A., & Barrett, D. A. (2014). Rapid Detection of Peptide Markers for Authentication Purposes in Raw and Cooked Meat Using Ambient Liquid Extraction Surface Analysis Mass Spectrometry. *Analytical Chemistry*, 86(20), 10257–10265.
- Montowska, M., Alexander, M. R., Tucker, G. A., & Barrett, D. A. (2015). Authentication of processed meat products by peptidomic analysis using rapid ambient mass spectrometry. *Food Chemistry*, 187, 297–304.
- Montowska, M., & Pospiech, E. (2011). Differences in two-dimensional gel electrophoresis patterns of skeletal muscle myosin light chain between *Bos taurus*, *Sus scrofa* and selected poultry species. *Journal of the Science of Food and Agriculture*, 91(13), 2449–2456.
- Montowska, M., & Pospiech, E. (2012). Myosin light chain retain their species-specific electrophoretic mobility after processing, which enables differentiation between six species: 2DE analysis of minced meat and meat products made from beef, pork and poultry. *Proteomics*, *12*(18), 2879–2889.
- Montowska, M., & Pospiech, E. (2013a). Species-specific expression of various proteins in meat tissue : Proteomic analysis of raw and cooked meat and meat products made from beef, pork and selected poultry species. *Food Chemistry*, *136*(3–4), 1461–1469.
- Montowska, M., & Pospiech, E. (2013b). Species-specific expression of various proteins in meat tissue: Proteomic analysis of raw and cooked meat and meat products made from beef, pork and selected poultry species. *Food Chemistry*, *136*(3–4), 1461–1469.

- Montowska, M., & Pospiech, E. (2016). Processed Meat Protein and Heat-Stable Peptide Marker Identification Using Microwave-Assisted Tryptic Digestion. *Food Technol. Biotechnol*, 54(4), 482–488.
- Montowska, M., Rao, W., Alexander, M. R., Tucker, G. A., & Barrett, D. A. (2014). Tryptic Digestion Coupled with Ambient Desorption Electrospray Ionization and Liquid Extraction Surface Analysis Mass Spectrometry Enabling Identification of Skeletal Muscle Proteins in Mixtures and Distinguishing between Beef, Pork, Horse, Chicken, and T. Analytical Chemistry, 86(9), 4479–4487.
- Mutalib, S. A., Wan Nazri, W. S., Shahimi, S., Yaakob, N., Abdullah Sani, N., Abdullah, A., ... Abd Ghani, M. (2012). Comparison between pork and wild boar meat (*Sus scrofa*) by polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP). *Sains Malaysiana*, 41(2), 199–204.
- Nakyinsige, K., Man, Y. B. C., & Sazili, A. Q. (2012). Halal authenticity issues in meat and meat products. *Meat Science*, 91(3), 207–214.
- Naveena, B. M., Jagadeesh, D. S., Kamuni, V., Muthukumar, M., Kulkarni, V. V, Kiran, M., & Rapole, S. (2018). In-gel and OFFGEL-based proteomic approach for authentication of meat species from minced meat and meat products. *Journal of the Science of Food and Agriculture*, 98(3), 1188–1196.
- Naylor, S. (2003). Biomarkers: current perspectives and future prospects. *Expert Review* of Molecular Diagnostics, 3, 525–529.
- Nemati, M., Oveisi, M. R., Abdollahi, H., & Sabzevari, O. (2004). Differentiation of bovine and porcine gelatins using principal component analysis. *Journal of Pharmaceutical and Biomedical Analysis*, 34(3), 485–492.
- Noh, H., Kim, G., Kim, Y., Kim, S., & Baek, N. (2013). Quality Evaluation of Panax ginseng Roots Using a Rapid Resolution LC-QTOF/MS-Based Metabolomics Approach. *Molecules*, 18, 14849–14861.
- Nur Azira, T., Che Man, Y. B., Raja Mohd Hafidz, R. N., Aina, M. A., & Amin, I. (2014). Use of principal component analysis for differentiation of gelatine sources based on protein molecular weights. *Food Chemistry*, 151, 286–292.
- Nur Azira, T., Amin, I., & Che Man, Y. B. (2012). Differentiation of bovine and porcine gelatins in processed products via Sodium Dodecyl Sulphate-Polyacrylamide Gel Electrophoresis (SDS-PAGE) and principal component analysis (PCA) techniques. *International Food Research Journal*, 19(3), 1175–1180.
- O'Brien, E. P., Dima, R. I., Brooks, B., & Thirumalai, D. (2007). Interactions between hydrophobic and ionic solutes in aqueous guanidinium chloride and urea solutions: Lessons for protein denaturation mechanism. *Journal of the American Chemical Society*, 129(23), 7346–7353.
- O'Gorman, A., Gibbons, H., & Brennan, L. (2013). Metabolomics in the Identification of Biomarkers of Dietary Intake. *Computational and Structural Biotechnology Journal*, 4(5), e201301004. Doi: org/10.5936/csbj.201301004

- Oe, M., Ohnishi-Kameyama, M., Nakajima, I., Muroya, S., & Chikuni, K. (2007). Muscle type specific expression of tropomyosin in bovine skeletal muscles. *Meat Science*, 75(4), 558–563.
- Olsen, J. V, Mann, M., Shevchenko, A., Tomas, H., & Havlis, J. (2007). In-gel digestion for mass spectrometric characterization of proteins and proteomes. *Nature Protocols*, 1(6), 2856–2860.
- Oluwaniyi, O. O., Dosumu, O. O., & Awolola, G. V. (2010). Effect of local processing methods (boiling, frying and roasting) on the amino acid composition of four marine fishes commonly consumed in Nigeria. *Food Chemistry*, *123*(4), 1000–1006.
- Ortea, I., Connor, G. O., & Maquet, A. (2016). Review on proteomics for food authentication. *Journal of Proteomics*, 147, 212–225.
- Parasuraman, S., R, A., Balamurugan, S., Muralidharan, S., Kumar, K. J., & Vijayan, V. (2014). An Overview of Liquid Chromatography-Mass Spectroscopy Instrumentation. *Pharmaceutical Methods*, 5(2), 47–55.
- Paredi, G., Sentandreu, M. A., Mozzarelli, A., Fadda, S., Hollung, K., & de Almeida, A. M. (2013). Muscle and meat: New horizons and applications for proteomics on a farm to fork perspective. *Journal of Proteomics*, 88, 58–82.
- Perry, S. V. (2001). Vertebrate tropomyosin: Distribution, properties and function. *Journal of Muscle Research and Cell Motility*, 22(1), 5–49.
- Pew Research Center (2017, 9 August). Muslims and Islam: Key findings in the U.S. and around the world. Retrieved from http://www.pewresearch.org/facttank/2017/08/09/muslims-and-islam-key-findings-in-the-u-s-and-around-theworld/
- Pitt, J. J. (2009). Principles and Applications of Liquid Chromatography- Mass Spectrometry in Clinical Biochemistry. *Clin Biochem Rev*, 30, 19–34.
- Porzio, M. A., & Pearson, A. M. (1977). Improved Resolution of Myofibrillar Proteins With Sodium. *Biochimica et Biophysica Acta*, 490, 27–34.
- Prensner, J. R., Rubin, M. A., Wei, J. T., & Chinnaiyan, A. M. (2012). Beyond PSA: the next generation of prostate cancer biomarkers. *Science Translational Medicine*, 4(127).
- Price, M. (2010). Meat Science. In J. Hudson, Robert (Ed.), *Animal and Plant Productivity* (pp. 177–210). Encyclopedia of Life Support Systems (EOLSS).
- Rohsenow, W. M., Hartnett, J. P., & Ganić, E. N. (Eds.) (1985). *Handbook of heat transfer applications* (Third Edition). MCGRAW-HILL.
- Rosemberg, I. M. (2006). *Protein Analysis and Purification*. Boston, MA: Birkhäuser Boston.

- Ruiz Orduna, A., Husby, E., Yang, C. T., Ghosh, D., & Beaudry, F. (2015). Assessment of meat authenticity using bioinformatics, targeted peptide biomarkers and highresolution mass spectrometry. *Food Additives and Contaminants - Part A Chemistry, Analysis, Control, Exposure and Risk Assessment, 32*(10), 1709–1717.
- Saez, R., Sanz, Y., & Toldra, F. (2004). PCR-based fingerprinting techniques for rapid detection of animal species in meat products. *Meat Science*, 66, 659–665.
- Sales, J., & Kotrba, R. (2013). Meat from wild boar (Sus scrofa L.): A review. Meat Science, 94(2), 187–201.
- Samaraweera, M., & Himali, S. (2012). Development of molecular tools to differentiate Sri Lankan wild boar (*Sus scrofa affinis*) meat from exotic and village pig (*Sus scrofa domestica*) meat. *Tropical Agricultural Research*, 23(1), 11–20.
- Sarah, S. A., Faradalila, W. N., Salwani, M. S., Amin, I., Karsani, S. A., & Sazili, A. Q. (2016). LC – QTOF-MS identification of porcine-specific peptide in heat treated pork identifies candidate markers for meat species determination. *Food Chemistry*, 199, 157–164.
- Sarah, S. A., Karsani, S.A., Amin, I., Mokhtar, N. F. K., & Sazili, A. Q. (2013). Differences in thermostable actin profile of goat meat as observed in twodimensional gel electrophoresis (2DE). *International Food Research Journal*, 20(2), 897–901.
- Sarah, S. A., Karsani, S. A., Amin, I., Mokhtar, N. F. K., & Sazili, A. Q. (2014). A proteomic based assessment on changes in myofibrillar proteins of goat Longissimus muscle as affected by heat treatments. *Journal of Animal and Plant Sciences*, 24(2), 406–412.
- Sentandreu, M. A., Fraser, P. D., Halket, J., Patel, R., & Bramley, P. M. (2010). A Proteomic-Based Approach for Detection of Chicken in Meat Mixes. *Journal of Proteome Research*, 9(7), 3374–3383.
- Seriramalu, R., Pang, W. W., Jayapalan, J. J., Mohamed, E., Abdul-Rahman, P. S., Bustam, A. Z., ... Hashim, O. H. (2010). Application of champedak mannosebinding lectin in the glycoproteomic profiling of serum samples unmasks reduced expression of alpha-2 macroglobulin and complement factor B in patients with nasopharyngeal carcinoma. *Electrophoresis*, 31(14), 2388–2395.
- Sheng, T. Y., Shamsudin, M. N., Mohamed, Z., Abdullah, A. M., & Radam, A. (2010). Demand analysis of meat in Malaysia. *Journal of Food Products Marketing*, *16*(2), 199–211.
- Singhal, N., Kumar, M., Kanaujia, P. K., & Virdi, J. S. (2015). MALDI-TOF mass spectrometry: An emerging technology for microbial identification and diagnosis. *Frontiers in Microbiology*, *6*, 1–16.
- Snyder, L. R., & Dolan, J. W. (2006). High-Performance Gradient Elution The Practical Application of the Linear-Solvent-Strength Model. Wiley-Interscience A John Wiley & Sons, Inc., Publication.

- Soares, S., Amaral, J. S., Oliveira, M. B. P. P., & Mafra, I. (2013). A SYBR Green realtime PCR assay to detect and quantify pork meat in processed poultry meat products. *MESC*, 94(1), 115–120.
- Stillwell, W. (2016). An Introduction to Biological Membranes: Composition, Structure, and Function (Second Edition). Academic Press.
- Stolba, A., & Wood-Gush, D. G. M. (1989). The behaviour of pigs in a semi-natural environment. Animal Production, 48(2), 419–425.
- Strimbu, K., & Tavel, J. A. (2010). What are biomarkers? *Current Opinion in HIV and AIDS*, 5(6), 463–466.
- Suckau, D., Resemann, A., Schuerenberg, M., Hufnagel, P., Franzen, J., & Holle, A. (2003). A novel MALDI LIFT-TOF/TOF mass spectrometer for proteomics. *Analytical and Bioanalytical Chemistry*, 376(7), 952–965.
- Takeichi, M. (1991). Cadherin cell adhesion receptors as a morphogenetic. *Science*, 251(5000), 1451–1455.
- The Local (2013, 10 April). Swedish halal sausages laced with pork. Retrieved from https://www.thelocal.se/20130410/47260
- Thermos Fisher Scientific (2015). Protein gel electrophoresis technical handbook. Retrieved from https://www.thermofisher.com/content/dam/LifeTech/global/Forms/PDF/proteingel-electrophoresis-technical-handbook.pdf
- The Star Online (2016, 15 May). Import of buffalo meat necessary to meet local beef demand. Retrieved from https://www.thestar.com.my/news/nation/2016/05/15/ import-of-buffalo-meat-necessary-to-meet-local-beef-demand/
- Thornton, P. K. (2010). Livestock production: recent trends, future prospects. *Philosophical Transactions of the Royal Society of London.Series B, Biological Sciences*, 365(1554), 2853–2867.
- Toldrá, F. (2017). Advances in Food and Nutrition Research. Academic Press.
- Trivedi, D. K., Hollywood, K. A., Rattray, N. J. W., Ward, H., Trivedi, D. K., Greenwood, J., ... Goodacre, R. (2016). Meat, the metabolites: an integrated metabolite profiling and lipidomics approach for the detection of the adulteration of beef with pork. *The Analyst*, *141*(7), 2155–2164.
- Valin, C., Pinkas, A., Dragnev, H., Boikovski, S., & Polikronov, D. (1984). Comparative study of buffalo meat and beef. *Meat Science*, 10(1), 69–84.
- Vander, A. J., Sherman, J. H., & Luciono, D. S. (2001). Human Physiology: The Mechanism of Body Function (Eight Edition). The McGraw-Hill.
- Van Bramer, S. E. (1997). An Introduction to Mass Spectrometry. Widener University, Pennsylvania, USA

- Von Bargen, C., Brockmeyer, J., & Humpf, H. (2014). Meat Authentication: A New HPLC–MS/MS Based Method for the Fast and Sensitive Detection of Horse and Pork in Highly Processed Food. *Journal of Agricultural and Food Chemistry*, 62(39), 9428–9435.
- Von Bargen, C., Dojahn, J., Waidelich, D., Humpf, H., & Brockmeyer, J. (2013). New Sensitive High-Performance Liquid Chromatography–Tandem Mass Spectrometry Method for the Detection of Horse and Pork in Halal Beef. *Journal* of Agricultural and Food Chemistry, 61(49), 11986–11994.
- Vranken, L., Avermaete, T., Petalios, D., & Mathijs, E. (2014). Curbing global meat consumption: Emerging evidence of a second nutrition transition. *Environmental Science and Policy*, 39, 1–12.
- Walker, J. E., Saraste, M., Runswick, M., & Gay, N. J. (1982). Distantly related sequences in the alpha- and beta-subunits of ATP synthase, myosin, kinases and other ATP-requiring enzymes and a common nucleotide binding fold. *The EMBO Journal*, 1(8), 945–951.
- Walker, J. M. (1996). The Protein Protocols Handbook. Humana Press, New Jersey.
- Warriss, P.D. (2000). Meat Science an introductory text. CABI Publishing.
- WHO International Programme on Chemical Safety Biomarkers and Risk Assessment: Concepts and Principles. 1993. Retrieved from http://www.inchem.org/documents/ehc/ehc/ehc155.htm.
- Williams, D. L., & Swenson, C. A. (1981). Tropomyosin stability: assignment of thermally induced conformational transitions to separate regions of the molecule. *Biochemistry*, 20(13), 3856–3864.
- Williams, P. G. (2007). Nutritional composition of red meat. *Nutrition & Dietetics*, 64(Suppl 4), S113-S119.
- XLSTAT (2017, 20 October). Partial least squares discriminant analysis PLSDA tutorial. Retrieved from https://help.xlstat.com/customer/en/portal/articles/2062368partial-least-squares-discriminant-analysis-plsda-tutorial?b_id=9283
- Xu, Y. J., Jin, M. L., Wang, L. J., Zhang, A. D., Zuo, B., Xu, D. Q., ... Xiong, Y. Z. (2009). Differential proteome analysis of porcine skeletal muscles between Meishan and Large White. *Journal of Animal Science*, 87(8), 2519–2527.
- Yu, T. Y., Morton, J. D., Clerens, S., & Dyer, J. M. (2017). Cooking-Induced Protein Modifications in Meat. *Comprehensive Reviews in Food Science and Food Safety*, 16(1), 141–159.
- Yuan, K., Kong, H., Guan, Y., Yang, J., & Xu, G. (2007). A GC-based metabonomics investigation of type 2 diabetes by organic acids metabolic profile. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 850(1–2), 236–240.

- Zenobi, R., & Knochenmuss, R. (1998). Ion formation in MALDI mass spectrometry. Mass Spectrometry Reviews, 17(5), 337–366.
- Zhan, X., & Desiderio, D. M. (2003). Spot volume vs. amount of protein loaded onto a gel: A detailed, statistical comparison of two gel electrophoresis systems. *Electrophoresis*, 24(11), 1818–1833.
- Zvereva, E. A., Kovalev, L. I., Ivanov, A. V, Kovaleva, M. A., Zherdev, A. V, Shishkin, S. S., ... Dzantiev, B. B. (2015). Enzyme immunoassay and proteomic characterization of troponin I as a marker of mammalian muscle compounds in raw meat and some meat products. *MESC*, 105, 46–52.



BIODATA OF STUDENT

Nadiah binti Mat Junoh was born on 18th February 1991 at Hospital Changkat Melintang, Perak. She got her primary education at Sekolah Rendah Islam Nurul Ehsan, Titi Gantung, Perak. She continued her secondary education at Sekolah Raja Perempuan Ta'ayah, Ipoh, Perak. Next, she further her study for higher education at International Islamic University Malaysia (IIUM), Kuantan, Pahang in Bachelor of Biotechnology after completed her foundation study at Center of Foundation Study, International Islamic University Malaysia (CFSIIUM), Petaling Jaya, Kuala Lumpur. In 2015, she enrolled as a master student at Halal Products Research Institute, Universiti Putra Malaysia (UPM) under Halal Product Science.



PUBLICATION

Journal:

Nadiah Mat Junoh., Mohd Hafis Yuswan., Shuhaimi Mustafa., Amalia Mohd Hashim., Rozi Mohamed., Shiou Yih Lee., Nazia Abdul Majid., Siti Aimi Sarah., and Dhilia Udie Lamasudin (2019). Comparing The Effect of Heat on Tropomyosin Isoforms Patterns from Water Buffalo and Wild Boar Meat by Two-Dimensional Gel Electrophoresis. *Malays. Appl. Biol, 48*(2), 131-139.





UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : FIRST SEMESTER 2015/2016

TITLE OF THESIS / PROJECT REPORT :

IDENTIFICATION OF THERMOSTABLE PEPTIDE MARKERS IN MEATS USING GEL-BASED

FRACTIONATION COUPLED WITH MASS SPECTROMETRY

NAME OF STUDENT : NADIAH BINTI MATJUNOH

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (√)



CONFIDENTIAL



OPEN ACCESS

(Contain confidential information under Official Secret Act 1972).

(Contains restricted information as specified by the organization/institution where research was done).

I agree that my thesis/project report to be published as hard copy or online open access.

(date)

This thesis is submitted for :



Embargo from____

(date)

Approved by:

(Signature of Student) New IC No/ Passport No.:910218-08-5266 (Signature of Chairman of Supervisory Committee) Name: Dr. Dhilia Udie Lamasudin

until

Date :

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]