



**DEVELOPMENT OF GENE MANIPULATION
SYSTEMS FOR GREEN MICROALGA
(*Ankistrodesmus convolutus* Corda.)**

TRAN THANH

FBSB 2012 2

**DEVELOPMENT OF GENE MANIPULATION
SYSTEMS FOR GREEN MICROALGA
(*Ankistrodesmus convolutus* Corda.)**



By

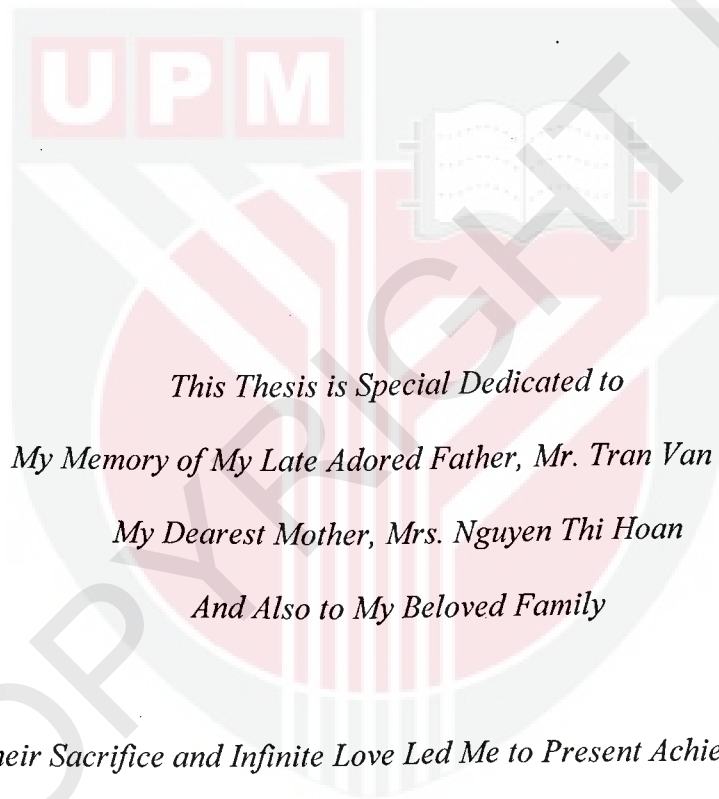
TRAN THANH

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirement for the Degree of Doctor of Philosophy**

January 2012

*Thành quả này xin kính dâng
Hương hồn Ba, Ông Trần Văn Chai
Mẹ Kính Yêu, Bà Nguyễn Thị Hoan
Và Gia Đình Thân Yêu*

Vì Sự Hy Sinh Lớn Lao Cho Cuộc Đời Tôi



*This Thesis is Special Dedicated to
My Memory of My Late Adored Father, Mr. Tran Van Chai
My Dearest Mother, Mrs. Nguyen Thi Hoan
And Also to My Beloved Family*

Their Sacrifice and Infinite Love Led Me to Present Achievements

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

**DEVELOPMENT OF GENE MANIPULATION
SYSTEMS FOR GREEN MICROALGA
(*Ankistrodesmus convolutus* Corda.)**

By

TRAN THANH

January 2012

Chairman: Associate Professor Suhaimi Napis, Ph.D

Faculty : Biotechnology and Biomolecular Sciences

Green microalga *Ankistrodesmus convolutus* is a fast growing alga which produces appreciable amount of carotenoids and polyunsaturated fatty acids. In addition of the ease and cost-effectiveness of culture, the ability of *A. convolutus* to form floating aggregates during its normal growth facilitate harvesting and other beneficiary attributes making it an interesting candidate for many biotechnological applications such as production of natural products or expression of recombinant proteins. However, the lack of efficient genetic transformation systems has been a major limitation in the manipulation of green microalgae. The present study provides, for the first time, information on molecular manipulation in *A. convolutus*, which covered the isolation of nucleic acids, construction of cDNA library and generation of ESTs, cloning and characterization of a highly-expressive cDNA and its promoter towards the establishment of an alternative expression system using *A. convolutus*.

Green microalga *Ankistrodesmus convolutus* was collected from axenic freshwater, grown and maintained in bold's basal medium. *A. convolutus* is typically rich in

lipids and polysaccharides, which make it difficult to get the intact RNA of high quality and quantity suitable for molecular research on this alga. In an effort to develop a suitable RNA extraction procedure for *A. convolutus*, a rapid, relatively non-toxic and effective method was finalized. This procedure was able to produce high quality and intact RNA which was of sufficient quality and suitable for downstream application such as RT-PCR, northern hybridization and cDNA library construction. The developed procedure may also have wider applicability for total RNA isolation from other green microalgae species. In order to provide a robust sequence resource that can be exploited for gene discovery, genome annotation and comparative genomics, a cDNA library of *A. convolutus* was constructed and preliminarily analysed. A total of 415 randomly selected clones were isolated from the primary cDNA library from which 201 high quality ESTs were produced after the sequencing of 337 clones. Among these ESTs, 21.4% of them were found to be related to gene expression, 14.4% to photosynthesis, 10.9% to metabolism, 5.5% to miscellaneous, 2.0% to stress response, and the remaining 45.8% were classified as novel genes. Due to the relatively high abundance, the full-length cDNA of ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) small subunit (*RbcS*) which was classified into photosynthetic cDNA clone category was selected for further studies. Rubisco is currently the target enzyme for improving the efficiency of photosynthesis in the hope of increasing the yield and growth of crops and henceforth green algae. In this study, the full-length of *A. convolutus RbcS* cDNA (*AcRbcS*) contained an open reading frame of 165 amino acids. This cDNA encoded a protein with expected molecular weight of ~21 kD sharing high homology with corresponding protein from other microalgae. Southern hybridization analysis revealed that *AcRbcS* is a member of a small multigene family comprising of two to

six members in *A. convolutus* genome. Under different illumination conditions, RT-PCR analysis showed that *AcRbcS* transcription was reduced in the dark, and drastically recovered in the light condition. As a primary step to investigate the use of *A. convolutus* as an alternative expression system for the production of recombinant proteins, the *AcRbcS* promoter region was isolated through PCR-based methods including ligation-mediated PCR and thermal asymmetric interlaced (TAIL)-PCR. In comparison with the ligation-mediated PCR technique, TAIL-PCR has proven to be an economic, efficient and rapid method to isolate *AcRbcS* promoter region. The *AcRbcS* promoter sequence was then analyzed and later used to construct an expression vector for expression of heterologous genes in *A. convolutus*. The transcription start site (TSS), consensus TATA-box and several putative *cis*-acting elements of representative light-regulatory as well as conserved motifs involved in light responsiveness were found in *AcRbcS* promoter region. The expression vector containing *AcRbcS* promoter and β -glucuronidase (*gusA*) reporter gene were constructed and introduced into *A. convolutus* cells by electroporation. As a result, transgenic *A. convolutus* lines was successfully generated and shown to contain the transgenes by PCR and southern hybridization analysis. It was also demonstrated that the *AcRbcS* promoter could function as a light-regulated promoter in transgenic *A. convolutus* as shown in northern hybridization analysis.

The results presented here represent the initial success on molecular manipulation of green microalgae *A. convolutus*, which has great potentials to become an interesting and excellent candidate for many biotechnological applications. The achievements therefore advance the development of *A. convolutus* as an alternative expression system, and it is now feasible that this alga can be examined as an alternative host for the expression of recombinant proteins.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PEMBANGUNAN SISTEM MANIPULASI
GEN UNTUK MIKROALGA HIJAU
(*Ankistrodesmus convolutus* Corda.)**

Oleh

TRAN THANH

Januari 2012

Pengerusi : Profesor Madya Suhaimi Napis, Ph.D

Fakulti: Bioteknologi and Sains Biomolekul

Mikroalga hijau *Ankistrodesmus convolutus* adalah sejenis alga yang mampu tumbuh dengan cepat dan menghasilkan jumlah karotenoid and asid lemak politatkepu yang tinggi. Tambahan pula ia senang dikultur dan kos pengkulturan yang rendah, keupayaan *A. convolutus* untuk membentuk agregat terapung semasa pertumbuhan yang normal memudahkan penuaian dan waris sifat-sifat yang lain menjadikannya calon yang menarik untuk pelbagai aplikasi bioteknologi seperti penghasilan produk semulajadi atau ekpresi protein rekombinan. Walaubagaimanapun, kekurangan sistem transformasi genetik yang cekap telah menjadi halangan utama dalam manipulasi mikroalga hijau. Kajian ini, untuk kali pertama menyediakan, informasi untuk manipulasi molekul dalam *A. convolutus* yang meliputi pengasingan asid nukleik, pembinaan perpustakaan cDNA dan generasi penanda jujukan terekspres (EST), pengklonan dan pencirian ekpresi cDNA tahap tinggi dan promoternya ke arah pembinaan sistem ekpresi alternatif menggunakan *A. convolutus*.

Mikroalga hijau *Ankistrodesmus convolutus* telah dikutip daripada air tawar axenik, dikultur dan dikekalkan dalam medium bold's basal. *A. convolutus* biasanya kaya dalam lipid dan polisakarida, yang menyebabkan ia sukar mendapatkan RNA utuh yang berkualiti tinggi dan kuantiti yang bersesuaian bagi penyelidikan molekul dalam alga ini. Dalam usaha untuk membangunkan prosedur pengasingan RNA bagi *A. convolutus*, satu kaedah yang, relatifnya tidak beracun dan berkesan telah ditemui dan disahkan. Prosedur ini telah berjaya menghasilkan RNA utuh yang berkualiti tinggi dan sesuai untuk aplikasi hiliran seperti RT-PCR, hibridisasi utara dan pembinaan perpustakaan cDNA. Prosedur yang dimajukan ini mampu diguna pakai secara meluas bagi pengasingan jumlah RNA daripada spesies mikroalga hijau yang lain. Untuk menyediakan sumber urutan yang teguh yang dapat dieksploitasi bagi penemuan gen, penjelasan genom dan genomik perbandingan, perpustakaan cDNA untuk *A. convolutus* telah dibinakan dan dianalisis pada peringkat awal. Sejumlah klon 415 yang dipilih secara rawak telah diasingkan daripada perpustakaan cDNA primer dari mana 201 EST yang bersih telah dihasilkan setelah menjujukan 337 klon. Antara EST ini, 21.4% daripadanya yang ditemui berkaitan dengan ekspresi gen, 14.4% dengan fotosintesis, 10.9% dengan metabolisme, 5.5% dengan kepelbagaian, 2.0% dengan tindak balas tekanan dan yang bakinya 45.8% telah diklasifikasikan sebagai gen yang baru. Disebabkan relatif kelimpahan yang tinggi, rantaian cDNA penuh yang lengkap bagi ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) subunit kecil (*RbcS*) yang telah diklasifikasikan ke dalam kategori cDNA klon fotosintesis telah dipilih untuk pengajian seterusnya. Rubisco kini adalah sasaran enzim untuk memperbaiki kecekapan fotosintesis dengan harapan meningkatkan penghasilan dan penumbuhan tanaman termasuklah alga hijau. Dalam kajian ini, barisan cDNA *A. convolutus* (*AcRbcS*) yang penuh

mengandung bacaan bingkai terbuka sebanyak 165 asid amino. cDNA ini mengkodkan protein dengan jangkaan berat molekul ~21 kD berkongsi homologi yang tinggi dengan protein yang kesamaan daripada mikroalga yang berlainan. Analisa hibridisasi selatan mendedahkan *AcRbcS* adalah ahli keluarga pelbagai gen kecil terdiri daripada dua hingga 5 ahli dalam genome *A. convolutus*. Di dalam keadaan illuminasi yang berbeza, analisa RT-PCR menunjukkan transkrip *AcRbcS* telah berkurang dalam keadaan gelap dan kembali pulih secara ketara dalam keadaan cahaya. Sebagai langkah utama untuk mengkaji kegunaan *A. convolutus* sebagai sistem ekspresi alternatif bagi menghasilkan protein rekombinasi, rantau promoter *AcRbcS* telah diasingkan melalui kaedah PCR termasuk PCR pencantuman pengantaraan dan PCR haba simetri berjalin (TAIL). Berbanding dengan kaedah PCR pencantuman pengantaraan, TAIL-PCR telah menunjukkan sebagai kaedah yang ekonomi, cekap dan pantas dalam pengasingan rantau promoter *AcRbcS*. Jujukan promoter *AcRbcS* kemudian dianalisa dan seterusnya digunakan untuk membina vektor ekspresi bagi pengepresian gen heterologous di *A. convolutus*. Tapak permulaan transkripsi (TSS), konsensus kotak TATA dan beberapa unsur-unsur yang dianggap sebagai cis-bertindak yang mewakili pengawalseliaan cahaya serta motif diperlihat yang terlibat dalam responsif cahaya telah dijumpa dalam rantau promoter *AcRbcS*. Vektor ekspresi yang mengandungi promoter *AcRbcS* dan gen pelapor β -glucuronidase (*gusA*) telah dibina dan diperkenalkan ke dalam sel *A. convolutus* melalui elektroporasi. Keputusan mendapati, talian transgenik *A. convolutus* telah berjaya digenerasikan dan ditunjukkan mengandungi transgen melalui PCR dan analisa hibridisasi selatan. Ia juga menunjukkan bahawa promoter *AcRbcS* dapat berfungsi sebagai promoter pengawalselia cahaya dalam transgenik *A. convolutus* seperti yang ditunjukkan dalam analisa hibridisasi utara.

Keputusan yang dibentangkan ini menunjukkan kejayaan awal dalam memanipulasi molekul untuk mikroalga hijau *A. convolutus*, yang berpotensi besar dan menjadi calon yang menarik dan cemerlang untuk pelbagai aplikasi bioteknologi. Penemuan dan pencapaian ini telah memajukan pembangunan *A. convolutus* sebagai sistem ekspresi alternatif dan kini alga ini dapat dikaji sebagai host alternatif bagi ekspresi protein rekombinan.



ACKNOWLEDGEMENTS

This thesis would not have been done without the financial support of the Vietnam Rubber Group (VRG) and Rubber Research Institute of Vietnam (RRIV). I greatly appreciate their scholarship award. I would like to thank the Ministry of Science, Technology and Innovation (MOSTI), Malaysia for funding this project Grant No. 02-01-04-SF0041.

I would like to express my gratitude to my Supervisory Committee, Assoc. Prof. Dr. Suhaimi Napis (Chairman), Dr. Hishamuddin Omar and Assoc. Prof. Dr. Mohd. Puad Abdullah for their dedicated effort, guidance and support throughout the study. Their experiences are valuable to guide me to the successful research.

I would like to thank Mr. Mai Van Son, former Director of Rubber Research Institute of Vietnam for giving me the opportunity to pursue my study in Universiti Putra Malaysia. Deepest gratitude goes to Dr. Lai Van Lam, Director of RRIV for the approval, invaluable help, support and encouragement during this course. The special thanks also go to Mr. Phan Thanh Dung, Mr. Le Mau Tuy and Dr. Nguyen Anh Nghia for their encouragement and support.

Special thanks again to my main supervisor, Assoc. Prof. Dr. Suhaimi Napis for entrusting me with the project, providing me the valuable support, and for his warm friendship during my academic program.

I appreciate all the support and motivation from my relatives, friends, colleagues, especially my colleagues working in the Breeding Division, Rubber Research Institute of Vietnam during my study in Malaysia.

I also would like to thank all the officers and staffs in the Plant Molecular Biology Lab, Department of Cell and Molecular Biology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia for their help in laboratorial materials and paper works.

My sincere appreciation also goes to Dr. Ky, Dr. Thuc, Dr. Shahanaz and Mrs. Samantha for their technical knowledge, enthusiastic help, sharing and encouragement during my study. Many thanks to Mr. Mostafa, Mr. Faizal, Ms. Hariyanti, Ms. Rosan, Ms. Sim, Ms. Anis, Mr. Yung Chee, Ms. Kalai, Mr. Suresh, Ms. Vasagi, Mrs. Hasna, Ms. Keat Ai, Mr. Amin for their help, friendship and encouragement. My special thanks go to Dr. Yeap Swee Keong for helping me to translate the English abstract into Malay language. Also, my thanks to the Vietnamese friends in Malaysia, Dr. Thanh, Dr. Thach, Mrs. Trang, Mr. Thu, Mr. Thanh, Mr. Dang, Mr. Khanh, Ms. Thai, Ms. Hue, Ms. Tran for their encouragement and support.

I am indebted to my parents, siblings and family for their endless love and constant support throughout my study. Last but not the least, I wish to express my heartfelt gratitude and appreciation to my beloved wife Vu Thi Quynh Chi for her endless love, sacrifice, physical and mental support to make this thesis a reality.

TABLE OF CONTENT

	Page
ABSTRACT	iii
ABSTRAK	vii
ACKNOWLEDGEMENTS	xi
APPROVAL	xiii
DECLARATION	xv
LIST OF TABLES	xxi
LIST OF FIGURES	xxii
LIST OF ABBREVIATIONS	xxiv
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	5
2.1 Green algae	5
2.1.1 Definition	5
2.1.2 Cellular structure	5
2.1.3 Reproduction	6
2.1.4 <i>In vitro</i> culture and maintenance	7
2.1.5 Genomic studies	8
2.1.6 Progress in transformation	9
2.1.7 Selectable markers and reporter genes	13
2.1.8 Promoters used	15
2.1.9 Potential uses of non-transgenic green algae	18
2.1.10 Potential uses of transgenic green algae	19
2.2 <i>Ankistrodesmus convolutus</i> Corda.	22
2.2.1 Taxonomy	22
2.2.2 Morphology	23
2.2.3 Culture conditions	24
2.2.4 Potential uses and application	24
2.3 Molecular approaches	25
2.3.1 RNA extraction	25
2.3.2 cDNA library	27
2.3.3 Expressed sequence tags	28
2.3.4 Regulation of gene expression	31
2.3.4.1 Promoters	31
2.3.4.2 <i>Cis</i> -acting elements	33
2.3.4.3 Promoter analysis	35
2.3.5 Thermal asymmetric interlaced-PCR and its applications	38
2.3.6 Gateway cloning technology	40
3 GENERAL MATERIALS AND METHODS	44
3.1 Alga culture condition	44
3.2 Isolation of total RNA	44
3.2.1 RNA quantification	46
3.2.2 Formaldehyde agarose gel electrophoresis	46

3.3	Isolation of mRNA from the total RNA	47
3.3.1	Isolation of mRNA	47
3.3.2	Quantification of mRNA	48
3.3.3	Ethidium bromide plate assay	48
3.4	Extraction of genomic DNA	49
3.5	Construction of cDNA library	49
3.5.1	First strand cDNA synthesis	50
3.5.2	Second-strand cDNA synthesis	51
3.5.3	Blunting the cDNA termini	51
3.5.4	Ligation with <i>EcoRI</i> adapter	52
3.5.5	Phosphorylating the <i>EcoRI</i> ends	52
3.5.6	Digestion with <i>XhoI</i>	52
3.5.7	Size fractionation of cDNA	53
3.6	cDNA purification	53
3.6.1	Ligation of cDNA into the ZAP Express vector	54
3.6.2	Packaging with Gigapack III extract	54
3.7	Phage plating and titering	54
3.7.1	Preparation of XL1-Blue MRF' cells	54
3.7.2	Plating and titering	55
3.8	Amplification of the cDNA library	55
3.9	Screening of the library	56
3.9.1	Probe preparation	56
3.9.2	Primary screening	57
3.9.3	Hybridization	58
3.9.4	Secondary screening	60
3.9.5	Single clone <i>in vivo</i> excision	60
3.9.6	Plasmid preparation	61
3.9.7	PCR amplification	63
3.9.8	Sequence analysis	63
3.10	Reverse transcription-PCR	64
3.11	Northern and southern hybridization analysis	65
3.11.1	Northern hybridization	65
3.11.2	Southern hybridization	66
3.11.3	Probe preparation	67
3.11.4	Hybridization analysis	68
3.12	Prokaryotic expression	69
3.12.1	Construction of prokaryotic protein expression vector	69
3.12.2	Transformation of prokaryotic protein expression vector into <i>E. coli</i>	70
3.12.3	SDS-PAGE analysis	70
3.13	Ligation-mediated PCR amplification	71
3.14	TAIL-PCR amplification	72
3.15	Promoter analysis	75
3.15.1	Sequence analysis	75
3.15.2	Identification of the transcription start site by 5'-rapid amplification of cDNA ends (5'-RACE)	75
3.15.3	Construction of promoter expression vector	76
3.15.3.1	PCR product of promoter region with flanking <i>attB</i> sites	77

3.15.3.2	BP reaction to create promoter entry clone	77
3.15.3.3	LR reaction to create promoter expression vector	78
3.15.3.4	Verification of promoter expression vector by restriction analysis	79
3.15.4	Nuclear transformation of promoter expression vector into <i>A. convolutus</i>	80
3.15.4.1	Enzymes treated to remove cell wall	80
3.15.4.2	Transformation by electroporation	80
3.15.4.3	Subculture of transformed lines of <i>A. convolutus</i>	81
3.15.4.4	Detection of transgenes using PCR analysis	81
3.15.4.5	Southern hybridization analysis of transgenic <i>A. convolutus</i>	82
3.15.5	Determination of influence of light on the regulation of <i>AcRbcS</i> promoter activity using Northern hybridization	83
4	RAPID AND EFFECTIVE METHOD OF RNA ISOLATION FROM GREEN MICROALGA <i>Ankistrodesmus convolutus</i>	85
	Abstract	85
4.1	Introduction	85
4.2	Materials and methods	87
4.2.1	Sample collection and preparation	87
4.2.2	RNA extraction protocol	87
4.2.3	Total RNA analysis	88
4.2.4	mRNA isolation	89
4.2.5	RT-PCR analysis	89
4.2.6	Northern blot analysis	89
4.2.7	cDNA library construction	90
4.3	Results and discussion	90
4.4	Conclusions	96
5	CONSTRUCTION OF cDNA LIBRARY AND PRELIMINARY ANALYSIS OF EXPRESSED SEQUENCE TAGS FROM GREEN MICROALGA <i>Ankistrodesmus convolutus</i>	98
	Abstract	98
5.1	Introduction	99
5.2	Materials and methods	100
5.2.1	Culture condition	100
5.2.2	Total RNA and mRNA isolation	101
5.2.3	Construction of cDNA library	102
5.2.4	Library screening and generation of expressed sequence tags	102
5.2.5	Sequence analysis	103
5.3	Results	104
5.3.1	Construction of cDNA library	104
5.3.2	Generation of expressed sequence tags and sequence analysis	106
5.4	Discussion	110
5.4.1	Composition of the cDNA library	110
5.4.2	Generation and analysis of ESTs	111
5.5	Conclusions	113

6	CLONING AND CHARACTERIZATION OF RIBULOSE-1,5-BISPHOSPHATE CARBOXYLASE/OXYGENASE SMALL SUBUNIT (<i>RbcS</i>) cDNA FROM GREEN MICROALGA <i>Ankistrodesmus convolutus</i>	114
	Abstract	114
	6.1 Introduction	115
	6.2 Materials and methods	117
	6.2.1 Alga culture	117
	6.2.2 Nucleic acids isolation	117
	6.2.3 Cloning of <i>AcRbcS</i> cDNA	118
	6.2.4 Bioinformatics analysis	118
	6.2.5 Southern blot analysis	119
	6.2.6 Semi-quantitative RT-PCR analysis	120
	6.2.7 Prokaryotic expression of <i>AcRbcS</i>	121
	6.3 Results and discussion	122
	6.3.1 Isolation and sequence analysis of <i>A. convolutus RbcS</i> full-length cDNA	122
	6.3.2 Phylogenetic analysis	127
	6.3.3 Southern hybridization analysis	128
	6.3.4 Expression of <i>AcRbcS</i> in different illumination conditions	130
	6.3.5 Prokaryotic expression of <i>AcRbcS</i> fusion proteins	131
	6.4 Conclusions	132
7	EFFICIENCY OF LIGATION-MEDIATED PCR AND TAIL-PCR METHODS FOR ISOLATION OF <i>RbcS</i> PROMOTER SEQUENCES FROM GREEN MICROALGA <i>Ankistrodesmus convolutus</i>	134
	Abstract	134
	7.1 Introduction	135
	7.2 Materials and methods	136
	7.2.1 Alga culture and genomic DNA isolation	136
	7.2.2 Primers synthesis	137
	7.2.3 Isolation of <i>AcRbcS</i> promoter sequences using ligation-mediated PCR	148
	7.2.4 Isolation of <i>AcRbcS</i> promoter sequences using TAIL-PCR	140
	7.3 Results and discussion	142
	7.4 Conclusions	147
8	SEQUENCE ANALYSIS AND POTENTIALS OF THE NATIVE <i>RbcS</i> PROMOTER IN THE DEVELOPMENT OF AN ALTERNATIVE EUKARYOTIC EXPRESSION SYSTEM USING GREEN MICROALGA <i>Ankistrodesmus convolutus</i>	148
	Abstract	148
	8.1 Introduction	149
	8.2 Materials and methods	152
	8.2.1 Culture conditions	152
	8.2.2 Isolation and sequence analysis of <i>AcRbcS</i> promoter	152
	8.2.3 Identification of the transcription start site by 5'-rapid amplification of cDNA ends (5'-RACE)	153
	8.2.4 Construction of promoter:: <i>gusA</i> fusing vector	154

8.2.5	Nuclear transformation of <i>A. convolutus</i>	156
8.2.6	Analysis of T-DNA integration and transgenes expression	156
8.3	Results and discussion	158
8.3.1	Sequence analysis of <i>AcRbcS</i> promoter	158
8.3.2	Determination of the transcription start site	161
8.3.3	Detection of transgenes using PCR analysis	162
8.3.4	Integration of the transgenes driven by the <i>AcRbcS</i> promoter	164
8.3.5	Influence of light on the regulation of <i>AcRbcS</i> promoter activity	167
8.4	Conclusions	168
9	GENERAL CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	171
9.1	General conclusions	171
9.2	Future research	175
	REFERENCES	177
	APPENDICES	200
	BIODATA OF THE AUTHOR	282

LIST OF TABLES

Tables		Page
2.1	Successful transformation methods used to transform green algae	12
2.2	Summary of transformable green alga species	13
2.3	Selectable marker and reporter genes used in green algae transformation systems	15
2.4	Promoters used in green algae transformation systems	17
2.5	A selection of recently promoter finding and analysis tools	36
2.6	A selection of recently promoter databases	37
2.7	Gateway plant binary destination vectors for promoter study	43
3.1	DNA sequence of arbitrary degenerate primers used in TAIL-PCR	74
3.2	Cycle conditions used for TAIL-PCR	74
5.1	Quality assessment of <i>A. convolutus</i> cDNA library	106
5.2	Summary of EST sequencing	107
5.3	Abundant ESTs found in the <i>A. convolutus</i> cDNA library	109
6.1	Comparison of amino acid sequence homology and biochemical properties of <i>RbcS</i> from different green algae species	124
7.1	DNA sequences of primers used	137
7.2	Cycle settings used for TAIL-PCR	142

LIST OF FIGURES

Figures		Page
2.1	Schematic diagram of TAIL-PCR contrasting the amplification of target with nontarget products (reproduced from Liu and Whittier, 1995)	40
2.2	Two basic steps involved in Gateway cloning technology (Source: Gateway technology instruction manual, Invitrogen)	43
3.1	The cDNA synthesis flow chart (Source: cDNA synthesis kit instruction manual, Stratagene)	50
4.1	Ethidium bromide stained 1.2% agarose formaldehyde gel of total RNA (1 µg) from green alga <i>A. convolutus</i> (lane 1), red seaweed (lane 2), brown seaweed (lane 3), oil palm (roots, lane 4), <i>A. thaliana</i> (leaves, lane 5), <i>E. coli</i> (lane 6), and rice (leaves, lane 7)	93
4.2	An ethidium bromide plate assay was performed by preparing 1% (w/v) agarose spotted with a serial dilutions (e.g., 480, 240, 120, 60, 30 ng/µL) of an RNA standard to confirm the mRNA concentration	93
4.3	Agarose gel electrophoresis analysis of RT-PCR amplification of the 724-bp <i>RbcS</i> cDNA from <i>A. convolutus</i>	94
4.4	Northern blotting results demonstrated the integrity of RNA isolated	95
4.5	Size fractionation of the cDNA was separated on 1% (w/v) agarose gel	95
4.6	Amplification of cDNA inserts from randomly chosen clones of <i>A. convolutus</i> cDNA library using T3 and T7 primers was separated on 1% (w/v) agarose gel	96
5.1	Quantification of size-fractionated cDNA	105
5.2	Amplification of cDNA inserts from randomly chosen phage plaques	106
5.3	Distribution of ESTs from <i>A. convolutus</i> cDNA library according to BLASTX score	107
5.4	The classification of ESTs from <i>A. convolutus</i> cDNA library based on their putative functions	108
6.1	Complete nucleotide and deduced amino acid sequence of the <i>AcRbcS</i> cDNA	125
6.2	Bioinformatics analysis of the amino acid sequences of <i>AcRbcS</i>	126
6.3	Phylogenetic relationship of <i>RbcS</i> among green algae	128
6.4	Southern hybridization analysis of <i>AcRbcS</i>	129

6.5	The expression of <i>AcRbcS</i> was analyzed by RT-PCR using total RNA with or without (–ve) reverse transcriptase	130
6.6	Expression of <i>AcRbcS</i> fusion protein in <i>E. coli</i> BL21(DE3)pLysS strain	132
7.1	Schematic diagram of the ligation-mediated PCR method with use of the TaKaRa LA PCR <i>in vitro</i> Cloning Kit	139
7.2	Schematic diagram of TAIL-PCR showing the amplification of the target (full line) with non-targeted (dashed line) products	141
7.3	Electrophoresis of products of ligation-mediated PCR carried out with use of the TaKaRa LA PCR <i>in vitro</i> Cloning Kit	144
7.4	Triple electrophoresis of the TAIL-PCR products	146
8.1	Schematic diagram showing the construction of pAcRbcS::gusA vector	155
8.2	Nucleotide sequence of the <i>AcRbcS</i> promoter	160
8.3	Determination of the transcription start site using 5'-RACE	161
8.4	PCR analysis of putative transformed lines	163
8.5	Southern hybridization analysis of transformed <i>A. convolutus</i>	166
8.6	Light regulation of the <i>AcRbcS</i> promoter in transformed <i>A. convolutus</i>	168

LIST OF ABBREVIATIONS

α	Alpha
β	Beta
λ	Lambda
g	Gravitational acceleration
μg	Microgram
μl	Microliter
$^{\circ}\text{C}$	Degree centigrade
%	Percentage
AD	Arbitrary degenerate
BBM	Bold's basal medium
BLAST	Basic local alignment search tool
bp	Base pairs
BSA	Bovine serum albumin
Ca	Calcium
CaMV35S	Cauliflower mosaic virus 35S
cDNA	Complementary DNA
CI	Chloroform:isoamyl alcohol
Cl	Chloride
cm	Centimeter
C-terminal	Carboxyl terminal
CTAB	Cetyltrimethylammonium bromide
dATP	2'-deoxy-adenosine-5'-triphosphate
dCTP	2'-deoxy-cytidine-5'-triphosphate

dGTP	2'-deoxy-guanosine-5'-triphosphate
DEPC	Diethyl pyrocarbonate
DMSO	Dimethylsulphonyl oxide
DNA	Deoxyribonucleic acid
DNase	Deoxyribonuclease
dNTPs	Deoxynucleotides
DTT	Dithiothreitol
dTTP	Thymidine-5'-tryphosphate
EDTA	Ethylenediaminetetraacetic acid
EtBr	Ethidium bromide
EST	Expressed sequence tag
g	Gram
gusA	β -glucuronidase
h	Hour(s)
HCl	Hydrochloric acid
HEPES	N-2-hydroxyethylpiperazine-N'-2 ethanesulfonic acid
hpt	Hygromycin phosphotransferase
IPTG	Isopropyl-1-thio- β -D-galactopyranoside
K	Potassium
KOAc	Potassium acetate
kb	Kilo base-pair
kV	Kilo volt
L	Liter
LB	Luria-bertani
LiCl	Lithium chloride

M	Molar
Mb	Mega base-pair
MES	2-(N-Morpholino)ethanesulfonic acid
Mg	Magnesium
MgCl ₂	Magnesium chloride
MgSO ₄	Magnesium sulfate
MOPS	3-(N-morpholino) propane-sulphonic acid
min	Minute(s)
mL	Milliliter
mM	Millimolar
mRNA	Messenger RNA
Na	Sodium
NaCl	Sodium chloride
NaOAc	Sodium acetate
NaOH	Sodium hydroxide
NCBI	National center for biotechnology information
ng	Nanogram
Nos	Nopaline synthase gene terminator
N-terminal	Amino-terminal
OD	Optical density
ORF	Open reading frame
PBS	Phosphate-buffered saline
PCI	Phenol:chloroform:isoamyl alcohol
PCR	Polymerase chain reactions
pfu	Plaque forming units

pI	Isoelectric point
PLACE	Plant <i>cis</i> -acting regulatory DNA elements
PlantCARE	Plant <i>cis</i> -acting regulatory elements
Poly (A) ⁺	Polyadenylated (mRNA)
ppm	Part per million
PVP	Polyvinylpyrrolidone
RACE	Rapid amplification of cDNA ends
RNA	Ribonucleic acid
RNase	Ribonuclease
rpm	Revolution per minute
RT	Reverse transcriptase
RT-PCR	Reverse transcription polymerase chain reaction
s	Seconds
SA-AP	Streptavidine alkaline-AP
SDS	Sodium dodecyl sulphate
SDS-PAGE	Sodium dodecyl sulfate polyacrylamide gel electrophoresis
SSC	Saline sodium citrate
TAE	Tris acetate EDTA
TAIL-PCR	Thermal asymmetric interlaced polymerase chain reaction
T-DNA	Transferred-DNA
TE	Tris-EDTA
TEMED	N, N, N', N'-tetramethylethylene diamine
T _m	Melting temperature
TSS	Transcription start site
U	Unit

UTR	Untranslated region
UV	Ultraviolet
V	Volt
v/v	Volume per volume
w/v	Weight per volume
X	Times
X-gal	5-bromo-4-chloro-3-indolyl- β -D-galactopyranoside



CHAPTER 1

INTRODUCTION

Algae are a remarkably diverse and fascinating group of organisms that are of fundamental ecological importance as primary producers and as basic components of the food chain. They are accountable for the net primary production of ~52 billion tons of organic carbon per year, which is ~50% of the total organic carbon produced on earth each year (Field et al., 1998). Algae are also of commercial importance in the food industry and in aquaculture and they are considered natural sources of high-value products such as carotenoids, long-chain polyunsaturated fatty acids, and phycocolloids (Apt and Behrens, 1999; Tseng, 2001). The diversified traits and living conditions of algae make them extremely attractive for commercial utilization particularly if the desired candidate alga is accessible to genetic manipulation.

The feasibility of green algae to be genetically modified and express heterologous genes opens up the possibility of enhancing the productivity of traditional algae compounds and producing new bioactive products for industrial and pharmaceutical applications through metabolic engineering (León-Bañares et al., 2004). Indeed, as eukaryotes, algae possess the chaperones and cellular machinery required to fold complex human proteins that bacteria and yeast may not process properly (Franklin and Mayfield, 2004). They can be cultured easily, rapidly and economically, and many of them are considered as safe food because they are free from human pathogens and endotoxins (Griesbeck et al., 2006). As a consequence of increasing global demand for recombinant proteins in a variety of industrial, algae have become the interesting species for novel pharmaceuticals, production of large quantities of

proteins particularly in cases where traditional sources are limited due to cost and/or availability (Moore, 1999; Skulberg, 2000; Specht et al., 2010). As photoautotrophs, transgenic algae, particularly green microalgae, can be exploited as cell factories for the production of valuable recombinant products such as vaccines, specialty oils, and novel carotenoids (León-Bañares et al., 2004).

Such fundamental and applied research requires the development of molecular genetic techniques for each algal species that allow the *in vivo* analysis of gene function and regulation, the manipulation of endogenous genes, and the introduction and expression of foreign genes (Walker et al., 2005a). Over the last few years, several genomic studies have been carried out for some important species of green microalgae such as *Chlamydomonas reinhardtii* (Shrager et al., 2003), *Dunaliella salina* (Li et al., 2004), *Acetabularia acetabulum* and *Ostreococcus tauri* (Henry et al., 2004; Derelle et al., 2006). This has resulted in providing a wealth of information and a strong foundation for targeted manipulation. Also, the molecular characterization of these green algae may provide useful DNA elements for genetic engineering of the host alga such as strong promoters which are necessarily used for green algae efficient transformation systems. Moreover, there has been a concerted effort to develop and improve the molecular tools for several important algae species. For instance, several transformation systems have been developed for producing high-value recombinant proteins in *C. reinhardtii* (Mayfield et al., 2003; Sun et al., 2003), *Dunaliella salina* (Geng et al., 2003), and various *Chlorella* species (Hawkins and Nakamura, 1999; Kim et al., 2002; Borovsky, 2003). It has led to the possibility of using such transgenic algae as vectors to deliver either vaccines or toxins to animals that are fed with algae. In short, green algae have proven their utility and tractability as a production system for commercial importance compounds as well as

for therapeutic or industrial proteins and peptides, and algae now seem poised to become the alternative expression system to other protein production systems.

Currently, the increasing importance of molecular research including generation of expressed sequence tags (ESTs), isolation and characterization of interested genes and its promoters, establishment of transformation systems in green algae, can be seen by the recent research reports and patents from numerous laboratories, the funding bodies as well as the establishment of several algal biotechnology companies. Even though the importance of algae biotechnology and genetic engineering has speedily increased, there are still difficulties, inconveniences and problems to be solved. Among these limitations, extensive research on molecular studies as well as optimal transformation constructs are necessary to be implemented. It is reviewed that although some common heterologous promoters (e.g., CaMV35S and the SV40 promoters) have been demonstrated to work in some green algae transformation constructs, inadequate recognition of the heterologous promoter region and lack of adequate regulation are major hurdles (Hallmann, 2007). Therefore, as a major step to optimize the transformation constructs, it is necessary to use a strong homologous promoter, which normally isolated from the high-expressive genes, for the efficient expression of heterologous proteins in green algae transformation.

As a fast growing alga producing appreciable amount of carotenoids and polyunsaturated fatty acids (Chu et al., 1992), freshwater green microalga *Ankistrodesmus convolutus* Corda has great potentials to become an interesting candidate for many biotechnological applications. The ability of *A. convolutus* to form floating aggregates during its normal growth to facilitate harvesting as well as

other beneficiary attributes make it potential as an alternative expression system for production of natural products and expression of therapeutic proteins. At the time the present study was initiated, however, there was no report of any genetic or molecular studies on this green microalga species. Thus, the aims of this study were to isolate and characterize a highly expressed cDNA clone and its promoter for application on the expression of a heterologous gene in green microalga *Ankistrodesmus convolutus* towards the establishment of an alternative expression system using this microalga. The specific objectives of this study were as follows:

1. To develop a suitable procedure for the isolation of high quality and quantity of total RNA from *A. convolutus*.
2. To construct a cDNA library from *A. convolutus*.
3. To isolate and characterize a highly expressed cDNA clone and its promoter region from *A. convolutus*.
4. To use the isolated promoter to drive the expression of a heterologous gene in *A. convolutus*.

REFERENCES

- Adams, M.D., Kelley, J.M., Gocayne, J.D., Dubnick, M., Polymeropoulos, M.H., Xiao, H., Merril, C.R., Wu, A., Olde, B., Moreno, R.F., Kerlavage, A.R., McCombie, W.R. and Venter, J.C. 1991. Complementary DNA sequencing: Expressed sequence tags and human genome project. *Science* 252: 1651-1656.
- Adams, M.D., Kerlavage, A.R., Flieschmann, R.D., Fuldner, R.A., Bult, C.J., Lee, N.H., Kirkness, E.F., Weinstock, K.G., Gocayne, J.D., White, O., Sutton, G., Blake, R.C., Brandon, R.C., Chiu, M.W., Clayton, R.A., Cline, R.T., Cotton, M.D., Earle-Hughes, J., Fine, L.D., Fitzgerald, L.M., Fitzhugh, W.M., Fritchman, J.L., Geoghagen, N.S.M., Glodek, A., Gnehm, C.L. and Venter, C. 1995. Initial assessment of human gene diversity and expression patterns based upon 83 million nucleotides of cDNA sequence. *Nature* 377: 3-174.
- Ahrazem, O., Rubio-Moraga, A., López, R.C. and Gómez-Gómez, L. 2010. The expression of a chromoplast-specific lycopene beta cyclase gene is involved in the high production of saffron's apocarotenoid precursors. *J. Exp. Bot.* 61: 105-119.
- Akiyama, M., Hirose, H., Yamagishi, T. and Hirano, M. 1977. Class Chlorophyceae. In *Illustrations of the Japanese freshwater algae*, ed. H. Hirose, T. Yamagishi, pp. 275-760. Tokyo: Uchidarokakuho-shisha Pub.
- Allona, I., Quinn, M., Shop, E., Swope, K., Cyr, S.S., Carlis, J., Riedl, J., Retzel, E., Campell, M.M., Sederoff, R. and Whetten, R.W. 1998. Analysis of xylem formation in pine by cDNA sequencing. *Proc. Natl. Acad. Sci. USA* 95: 9693-9698.
- Altschul, S.F., Gish, W., Miller, W., Myers, E.W. and Lipman, D.J. 1990. Basic local alignment search tool. *J. Mol. Biol.* 215: 403-410.
- Altschul, S.F., Madden, T.L., Schaffer, A.A., Zhang, J., Miller, W. and Lipman, D.J. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res.* 25: 3389-3402.
- Antal, Z., Rascle, C., Fèvre, M. and Bruel, C. 2004. Single oligonucleotide nested PCR: a rapid method for the isolation of genes and their flanking regions from expressed sequence tags. *Curr. Genet.* 46: 240-246.
- Apt, K.E. and Behrens, P.W. 1999. Commercial developments in microalgal biotechnology. *J. Phycol.* 35: 215-226.
- Apt, K.E. and Grossman, A.R. 1993. Characterization and transcript analysis of the major phycobiliprotein subunit genes from *Aglaothamnion neglectum* (Rhodophyta). *Plant Mol. Biol.* 21: 27-38.
- Apt, K.E., Clendennen, K.S., Powers, D.A. and Grossman, A.R. 1995. The gene

family encoding the fucoxanthin chlorophyll proteins from the brown alga *Macrocystis pyrifera*. *Mol. Genet. Genomics* 246: 455-464.

Arie, T., Christiansen, S.K., Yoder, O.C. and Turgeon, B.G. 1996. Efficient cloning of ascomycete mating type genes by PCR amplification of the conserved MAT HMG Box. *Fungal. Genet. Biol.* 21: 118-130.

Auchincloss, A.H., Loroach, A.I. and Rochaix, J.D. 1999. The argininosuccinate lyase gene of *Chlamydomonas reinhardtii*: cloning of the cDNA and its characterization as a selectable shuttle marker. *Mol. Gen. Genet.* 261: 21-30.

Bailey, T.L. and Elkan, C. 1994. Fitting a mixture model by expectation maximization to discover motifs in biopolymers. *Proc. Int. Conf. Intell. Syst. Mol. Biol.* 2: 28-36.

Banicki, J.J. 2004. An alga a day keeps the doctor away. Engineered algae as a new means to vaccinate fish. *Twine Line* 26: 1-5.

Barlow, J.J., Mathias, A.P., Williamson, R. and Gammack, D.B. 1963. A Simple method for the quantitative isolation of undegraded high molecular weight ribonucleic acid. *Biochem. Biophys. Res. Comm.* 13: 61-66.

Benson, E.E., Fleck, R.A., Bremner, D.A. and Day, J.G. 1998. Assessments of hydroxyl activity and antioxidant status in freeze-recalcitrant and freeze-sensitive algae: implications for cryopreserved culture collections. *In Vitro Cell. Dev. Biol. Plant* 34: P-1039, 52A.

Benton, W.D. and Davis, R.W. 1977. Screening λ gt recombinant clones by hybridization to single plaques in situ. *Science* 196:180-182.

Bernard, P. and Couturier, M. 1992. Cell killing by the F plasmid CcdB protein involves poisoning of DNA-topoisomerase II complexes. *J. Mol. Biol.* 226: 735-745.

Berry-Lowe, S.L., McKnight, T.D., Shah, D.M. and Meagher, R.B. 1982. The nucleotide sequence, expression and evolution of one member of a multigene family encoding the small subunit of ribulose-1,5-bisphosphate carboxylase in soybean. *J. Mol. Appl. Genet.* 1: 483-498.

Blankenship, E. and Kindle, K. 1992. Expression of chimeric genes by the light-regulated cabII-1 promoter in *Chlamydomonas reinhardtii*: a cabII-1/nit1 gene functions as a dominant selectable marker in a nit1 nit2 strain. *Mol. Cell. Biol.* 12: 5268-5279.

Bolle, C., Sopory, S., Lubberstedt, T., Herrmann, R.G. and Oelmüller, R. 1994. Segments encoding 5'-untranslated leaders of genes for thylakoid proteins contain cis-elements essential for transcription. *Plant J.* 6: 513-523.

Borovsky, D. 2003. Trypsin-modulating oostatic factor: a potential new larvicide for mosquito control. *J. Exp. Biol.* 206: 3869-3875.

- Borowitzka, M.A. 1994. Product from algae. In *Proceeding of the conference on algae biotechnology in the Asia-Pacific region*, ed. S.M. Phang, Y.K. Lee, M.A. Borowitzka, B.A. Whitton, pp. 5-15. Kuala Lumpur: University of Malaya.
- Borowitzka, M.A. and Borowitzka, L.J. 1990. Algae biotechnology. In *Biology of marine plants*, ed. M.N. Clayton, R.J. King, pp. 385-399. Melbourne: Longman Cheshire.
- Boynton, J.E., Gillham, N.W., Harris, E.H., Hosler, J.P., Johnson, A.M., Jones, A.R., Randolph-An-Derson, B.L., Robertson, D., Klein, T., Shark, K.B. and Sanford, J.C. 1988. Chloroplast transformation in *Chlamydomonas* with high velocity microprojectiles. *Science* 240: 1534-1538.
- Brown, L.E., Sprecher, S.L. and Keller, L.R. 1991. Introduction of exogenous DNA into *Chlamydomonas reinhardtii* by electroporation. *J. Cell Biol.* 11: 2328-2332.
- Bugos, R.C., Chiang, V.L., Zhang, X.H., Campell, E.R, Podilla, G.K. and Campell, W.H. 1995. RNA isolation from plant tissues recalcitrant to extraction in guanidine. *Biotechniques* 19: 734-737.
- Burrows, E.M. 1991. *Seaweeds of the British Isles, vol 2, Chlorophyta*. London: Natural History Museum.
- Butanaev, A.M. 1994. Hygromycin phosphotransferase gene as a dominant selective marker for transformation of *Chlamydomonas reinhardtii*. *Mol. Biol.* 28: 682-686.
- Butler, J.E.F. and Kadonaga, J.T. 2002. The RNA polymerase II core promoter: a key component in the regulation of gene expression. *Genes Dev.* 16: 2583-2592.
- Cerutti, H., Johnson, A.M., Gillham, N.W. and Boynton, J.E. 1997. A eubacterial gene conferring spectinomycin resistance on *Chlamydomonas reinhardtii*: integration into the nuclear genome and gene expression. *Genetics* 145: 97-110.
- Chan, C.X., Teo, S.S., Ho, C.L., Othman, R.Y. and Phang, S.M. 2004. Optimisation of RNA extraction from *Gracilaria changii* (Gracilariales, Rhodophyta). *J. Appl. Phycol.* 16: 297-301.
- Channeliere, S., Riviere, S., Scalliet, G., Szecsi, J., Jullien, F., Dolle, C., Vergne, P., Dumas, C., Bendahmane, M., Huguene, P. and Cock, J.M. 2002. Analysis of gene expression in rose petals using expressed sequence tags. *FEBS Lett.* 515: 35-38.
- Chapman, V.J. and Chapman, D.J. 1980. *Seaweeds and their uses*, 3rd ed. London: Chapman and Hall.

- Chebolu, S. and Daniell, H. 2009. Chloroplast-derived vaccine antigens and biopharmaceuticals: expression, folding, assembly and functionality. *Curr. Top. Microbiol. Immunol.* 332: 33-54.
- Chen, C., Bai, L.H., Qiao, D.R., Xu, H., Dong, G.L., Ruan, K., Huang, F. and Cao, Y. 2008. Cloning and expression study of a putative carotene biosynthesis related (cbr) gene from the halotolerant green alga *Dunaliella salina*. *Mol. Biol. Rep.* 35: 321-327.
- Chen, Y., Wang, Y., Sun, Y., Zhang, L. and Li, W. 2001. Highly efficient expression of rabbit neutrophil peptide-1 gene in *Chlorella ellipsoidea* cells. *Curr. Genet.* 39: 365-370.
- Chow, K.C. and Tung, W.L. 1999. Electrotransformation of *Chlorella vulgaris*. *Plant Cell Rep.* 18: 778-780.
- Chu, W.L., Phang, S.M. and Goh, S.H. 1995. Influence of carbon source on growth, biochemical composition and pigmentation of *Ankistrodesmus convolutus*. *J. Appl. Phycol.* 7: 59-64.
- Chu, W.L., Phang, S.M., Goh, S.H. and Blakebrough, N. 1992. Promising microalgae for production of useful chemicals. In *Proceeding of the conference on medicinal products from tropical rain forests*, ed. K. Shaari, A.A. Kadir, A.R.M. Ali, pp. 338-345. Kuala Lumpur: Forest Research Institute of Malaysia.
- Cleland, W.W. 1964. Dithiothreitol, a new protective reagent for SH groups. *Biochemistry* 3: 480-482.
- Clifton, K.E. 1997. Mass spawning by green algae on coral reefs. *Science* 275: 1116-1118.
- Coemans, B., Matsumura, H., Terauchi, R., Remy, S., Swennen, R. and Sagi, L. 2005. SuperSAGE combined with PCR walking allows global gene expression profiling of banana (*Musa acuminata*), a non-model organism. *Theor. Appl. Genet.* 111: 1118-1126.
- Cohen, Z. 1986. Products from microalgae. In *CRF Handbook of microalgal culture*, ed. A. Richmond, pp. 421-454. Boca Raton: CRF Press.
- Coll, J.M. 2006. Review. Methodologies for transferring DNA into eukaryotic microalgae. *Spanish J. Agric. Res.* 4: 316-330.
- Cooper, L.D., Marquez-Cedillo, L., Singh, J., Sturbaum, A.K., Zhang, S., Edwards, V., Johnson, K., Kleinhofs, A., Rangel, S., Carollo, V., Bregitzer, P., Lemaux, P.G. and Hayes P.M. 2004. Mapping Ds insertions in barley using a sequence-based approach. *Mol. Gen. Genomics* 272: 181-193.

- Covitz, P.A., Smith, L.S. and Long, S.R. 1998. Expressed sequence tags from a root hair-enriched *Medicago truncatula* cDNA library. *Plant Physiol.* 117: 1325-1332.
- Cowell, I.G. 1998. cDNA libraries. In *Molecular biotechnology handbook*, ed. R. Rapley, J.M. Walker, pp. 131-144. Totowa: Humana Press.
- Crépineau, F., Roscoe, T., Kaas, R., Klaoreg, B. and Boyen, C. 2000. Characterisation of complementary DNAs from the expressed sequence tag analysis of life cycle stages of *Laminaria digitata* (Phaeophyceae). *Plant Mol. Biol.* 43: 503-513.
- Cui, L., Xue, L., Li, J., Zhang, L. and Yan, H. 2010. Characterization of the glucose-6-phosphate isomerase (GPI) gene from the halotolerant alga *Dunaliella salina*. *Mol. Biol. Rep.* 37: 911-916.
- Curtis, M.D. and Grossniklaus, U. 2003. A gateway cloning vector set for high-throughput functional analysis of genes in planta. *Plant Physiol.* 133: 462-469.
- Dawson, H.N., Burlingame, R., Cannons, A.C. 1997. Stable transformation of *Chlorella*: rescue of nitrate reductase-deficient mutants with nitrate reductase gene. *Curr. Microbiol.* 35: 356-362.
- Day J.G., Benson E.E. and Fleck R.A. 1999. In vitro culture and conservation of microalgae: applications for aquaculture, biotechnology and environmental research. *In Vitro Cell. Dev. Biol. Plant* 35: 127-136.
- Day, J.G. 1998. Cryo-conservation of microalgae and cyanobacteria. *Cryo. Lett. Suppl.* 1: 7-14.
- Day, J.G. 1999. Conservation strategies for algae. In *Plant conservation biotechnology*, ed. E.E. Benson, pp. 111-124. London: Taylor and Francis Ltd.
- Day, J.G. and DeVille, M.M. 1995. Cryopreservation of algae. *Methods Mol. Biol.* 38: 81-90.
- de Ruijter, N.C.A., Verhees, J., van Leeuwen, W., van der Krol, A.R. 2003. Evaluation and comparison of the GUS, LUC and GFP reporter system for gene expression studies in plants. *Plant Biol.* 5: 103-115.
- Dean, C., Pichersky, E. and Dunsmuir, P. 1989. Structure, evolution, and regulation of *RbcS* genes in higher plants. *Annu. Rev. Plant Physiol. Plant. Mol. Biol.* 40: 415-439.
- Debuchy, R., Purton, S. and Rochaix, J.D. 1989. The argininosuccinate lyase gene of *Chlamydomonas reinhardtii*: an important tool for nuclear transformation and for correlating the genetic and molecular maps of the ARG7 locus. *EMBO J.* 8: 2803-2809.

- Derelle, E., Ferraz, C., Rombauts, S., Rouze, P., Worden, A.Z., Robbens, S., Partensky, F., Degroeve, S., Echeynie, S., Cooke, R., Saeys, Y., Wuyts, J., Jabbari, K., Bowler, C., Panaud, O., Piegue, B., Ball, S.G., Ral, J.P., Bouget, F.Y., Piganeau, G., de Baets, B., Picard, A., Delseny, M., Demaille, J., van de Peer, Y. and Moreau, H. 2006. Genome analysis of the smallest free-living eukaryote *Ostreococcus tauri* unveils many unique features. *Proc. Natl. Acad. Sci. USA* 103: 11647-11652.
- DeRocher, E.J., Qulgley, F., Meche, R. and Bohnert, H.J. 1993. The six genes of the Rubisco small subunit multigene family from *Mesembqnthemum crystallinum*, a facultative CAM plant. *Mol. Gen. Genet.* 239: 450-462
- Dhanaraj, A.L., Slovin, J.P. and Rowland, L.J. 2004. Analysis of gene expression associated with cold acclimation in blueberry floral buds using expressed sequence tags. *Plant Sci.* 166: 863-872.
- Dhingra, A., Portis, A.R. and Daniell, H. 2004. Enhanced translation of a chloroplast expressed *rbcS* gene restores SSU levels and photosynthesis in nuclear antisense *RbcS* plants. *Proc. Natl. Acad. Sci. USA* 101: 6315-6320.
- Doyle, J.J. and Doyle, J.L. 1990. Isolation of plant DNA from fresh tissue. *Focus* 12: 13-15.
- Dreesen, I.A.J., Charpin-El, H.G. and Fussenegger, M. 2010. Heat-stable oral alga-based vaccine protects mice from *Staphylococcus aureus* infection. *J. Biotechnol.* 145: 273-280.
- Dunahay, T.G. 1993. Transformation of *Chlamydomonas reinhardtii* with silicon carbide whiskers. *Biotechniques* 15: 452-455, 457-458, 460.
- Earley, K.W., Haag, J.R., Pontes, O., Opper, K., Juehne, T., Song, K. and Pikaard, C.S. 2006. Gateway-compatible vectors for plant functional genomics and proteomics. *Plant J.* 45: 616-629.
- El-Sheekh, M.M. 1999. Stable transformation of the intact cells of *Chlorella kessleri* with high velocity microprojectiles. *Biologia Plantarum* 42: 209-216.
- Emanuelsson, O., Nielsen, H. and von Heijne, G. 1999. ChloroP, a neural network-based method for predicting chloroplast transit peptides and their cleavage sites. *Protein Sci.* 8: 978-984.
- Endo, M., Hakozaki, H., Kokubun, T., Masuko, H., Takahata, Y., Tsuchiya, T., Higashitani, A., Tabata, S. and Watanabe, M. 2002. Generation of 919 expressed sequence tags from immature flower buds and gene expression analysis using expressed sequence tags in the model plant *Lotus japonicus*. *Genes Genet. Syst.* 77: 277-282.
- Falcão, V.D.R., Tonon, A.P., Oliveira, M.C. and Colepicolo, P. 2008. RNA Isolation method for polysaccharide rich algae: agar producing *Gracilaria tenuistipitata* (Rhodophyta). *J. Appl. Phycol.* 20: 9-12.

- Feng, S., Xue, L., Liu, H. and Lu, P. 2009. Improvement of efficiency of genetic transformation for *Dunaliella salina* by glass beads method. *Mol. Biol. Rep.* 36: 1433-1439.
- Field, C.B., Behrenfeld, M.J., Randerson, J.T. and Falkowski, P. 1998. Primary production of the biosphere: integrating terrestrial and oceanic components. *Science* 28: 237-240.
- Fischer, N. and Rochaix, J.D. 2001. The flanking regions of *PsaD* drive efficient gene expression in the nucleus of the green alga *Chlamydomonas reinhardtii*. *Mol. Genet. Genomics* 265: 888-894.
- Flieger, K., Wicke, A., Hermann, R.G. and Oelmüller, R. 1994. Promoter and leader sequences of the spinach *PsaD* and *PsaF* genes direct an opposite light response in tobacco cotyledons. *PsaD* sequences downstream of the ATG codon are required for positive light response. *Plant J.* 6: 359-368.
- Franco, G.R., Adams, M.D., Soares, M.B., Simpson, A.J.G., Venter, J.C. and Pena, S.D.J. 1995. Identification of new *Schistosoma mansoni* genes by the EST strategy using a directional cDNA library. *Gene* 152: 141-147.
- Franco, G.R., Rabelo, E.M., Azevedo, V., Pena, H.B., Ortega, J.M., Santos, T.M., Meira, W.S., Rodrigues, N.A., Dias, C.M., Harrop, R., Wilson, A., Saber, M., Abdel-Hamid, H., Faria, M.S., Margutti, M.E., Parra, J.C. and Pena, S.D. 1997. Evaluation of cDNA libraries from different developmental stages of *Schistosoma mansoni* for production of expressed sequence tags (ESTs). *DNA Res.* 4: 231-240.
- Franklin, S., Ngo, B., Efué, E. and Mayfield, S.P. 2002. Development of a GFP reporter gene for *Chlamydomonas reinhardtii* chloroplast. *Plant J.* 30: 733-744.
- Franklin, S.E. and Mayfield, S.P. 2004. Prospects for molecular farming in the green alga *Chlamydomonas reinhardtii*. *Curr. Opin. Plant Biol.* 7: 159-165.
- Frith, M.C., Hansen, U. and Weng, Z. 2001. Detection of *cis*-element clusters in higher eukaryotic DNA. *Bioinformatics* 17: 878-889
- Fuhrmann, M., Oertel, W. and Hegemann, P. 1999. A synthetic gene coding for the green fluorescent protein (GFP) is a versatile reporter in *Chlamydomonas reinhardtii*. *Plant J.* 19: 353-361.
- Gasteiger, E., Gattiker, A., Hoogland, C., Ivanyi, I., Appel, R.D. and Bairoch, A. 2003. ExPASy: the proteomics server for in-depth protein knowledge and analysis. *Nucleic Acids Res.* 31: 3784-3788.
- Gehrig, H.H., Winter, K., Cushman, J., Borland, A. and Taybi, T. 2000. An improved RNA isolation method for succulent plant species rich in polyphenols and polysaccharides. *Plant Mol. Biol. Rep.* 18: 369-376.

- Geng, D., Han, Y., Wang, Y., Wang, P., Zhang, L., Li, W. and Sun, Y. 2004. Construction of a system for the stable expression of foreign genes in *Dunaliella salina*. *Acta Botanica Sinica* 46: 342-346.
- Geng, D., Wang, Y., Wang, P., Li, W. and Sun Y. 2003. Stable expression of hepatitis B surface antigen gene in *Dunaliella salina* (Chlorophyta). *J. Appl. Phycol.* 15: 451-456.
- Geourjon, C. and Deléage, G. 1995. SOPMA: significant improvement in protein secondary structure prediction by consensus prediction from multiple alignments. *Comput. Appl. Biosci.* 11: 681-684.
- Ghosh, D. 2000. Object-oriented Transcription Factors Database (ooTFD). *Nucleic Acids Res.* 28: 308-310.
- Gittins, J.R., Pellny, T.K., Hiles, E.R., Rosa, C., Biricolti, S. and James, D.J. 2000. Transgene expression driven by heterologous ribulose-1,5-bisphosphate carboxylase/oxygenase small-subunit gene promoters in the vegetative tissues of apple (*Malus pumila* Mill.). *Planta* 210: 232-240.
- Giuliano, G., Pechersky, E., Malik, V.S., Timko, M.P., Scolnik, P.A. and Cashmore, A.R. 1988. An evolutionarily conserved protein binding sequence upstream of a plant light-regulated gene. *Proc. Natl. Acad. Sci. USA* 85: 7089-7093.
- Goldschmidt-Clermont, M. and Rahire, M. 1986. Sequence, evolution and differential expression of the two genes encoding variant small subunits of ribulose bisphosphate carboxylase/oxygenase in *Chlamydomonas reinhardtii*. *J. Mol. Biol.* 191: 421-432.
- Griesbeck, C., Kobl, I. and Heitzer, M. 2006. *Chlamydomonas reinhardtii*: a protein expression system for pharmaceutical and biotechnological proteins. *Mol. Biotechnol.* 34: 213-223.
- Grossman, A.R. 2005. Paths toward algal genomics. *Plant Physiol.* 137: 410-427.
- Gruber, A. 2007. Expressed sequence tags. In *Bioinformatics: methods express*, ed. P.H. Dear, pp. 141-167. Bloxham: Scion Publishing Ltd.
- Guillard, R.R.L. and Lorenzen, C.J. 1972. Yellow-green algae with chlorophyllide c. *J. Phycol.* 8: 10-14.
- Hall, L.M., Taylor, K.B. and Jones, D.D. 1993. Expression of a foreign gene in *Chlamydomonas reinhardtii*. *Gene* 124: 75-81.
- Hall, T.A. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucl. Acids Symp. Ser.* 41: 95-98.
- Hallmann, A. 2007. Algal transgenics and biotechnology. *Transgenic Plant J.* 1: 81-98.

- Hanhineva, K.J. and Kärenlampi, S.O. 2007. Production of transgenic strawberries by temporary immersion bioreactor system and verification by TAIL-PCR. *BMC Biotechnol.* 7: 11.
- Hartley, J.L., Temple, G.F. and Brasch, M.A. 2000. DNA cloning using in vitro site-specific recombination. *Genome Res.* 10: 1788-1795.
- Hartman, F.C. and Harpel, M.R. 1994. Structure, function, regulation, and assembly of D-ribulose-1,5-bisphosphate carboxylase/oxygenase. *Annu. Rev. Biochem.* 63: 197-234.
- Hawkins R.L. and Nakamura M. 1999. Expression of human growth hormone by the eukaryotic alga, *Chlorella*. *Curr. Microbiol.* 38: 335-341.
- Hayashi, M., Hirono, M. and Kamiya, R. 2001. Recovery of flagellar dynein function in a *Chlamydomonas* actin/dynein-deficient mutant upon introduction of muscle actin by electroporation. *Cell Motil. Cytoskeleton* 49: 146-153.
- Henry, I.M., Wilkinson, M.D., Hernandez, J.M., Schwarz-Sommer, Z., Grotewold, E. and Mandoli, D.F. 2004. Comparison of ESTs from juvenile and adult phases of the giant unicellular green alga *Acetabularia acetabulum*. *BMC Plant Biol.* 4: 3.
- Hernández, M., Pla, M., Esteve, T., Prat, S., Puigdomènech, P. and Ferrando A. 2003. A specific real-time quantitative PCR detection system for event MON810 in maize YieldGard® based on the 3'-transgene integration sequence. *Transgenic Res.* 12: 179-189.
- Higo, K., Ugawa, Y., Iwamoto, M. and Korenaga, T. 1999. Plant *cis*-acting regulatory DNA elements (PLACE) database. *Nucleic Acids Res.* 27: 297-300.
- Higuchi, R., Dollinger, G., Walsh, P.S. and Griffith, R. 1992. Simultaneous amplification and detection of specific DNA sequences. *Nat. Biotechnol.* 10: 413-417.
- Himmelbach, A., Zierold, U., Hensel, H., Riechen, J., Douchkov, D., Schweizer, P. and Kumlehn, J. 2007. A set of modular binary vectors for transformation of cereals. *Plant Physiol.* 145: 1192-1200.
- Hirooka T., Akiyama, Y., Tsuji, N., Nakamura, T., Nagase, H., Hirata, K. and Miyamoto, K. 2003. Removal of hazardous phenols by microalgae under photoautotrophic conditions. *J. Biosci. Bioeng.* 95: 200-203.
- Hu, C.G., Honda, C., Kita, M., Zhang, Z., Tsuda, T. and Moriguchi, T. 2002. A simple protocol for RNA isolation from fruit trees containing high levels of polysaccharides and polyphenol compounds. *Plant Mol. Biol. Rep.* 20: 69a-69g.

- Huang, X., Weber, J.C., Hinson, T.K., Mathieson, A.C. and Minocha, S.C. 1996. Transient expression of the GUS reporter gene in the protoplasts and partially digested cells of *Ulva lactuca*. *Botanica Marina* 39: 467-474.
- Hudson, M.E. and Quail, P.H. 2003. Identification of promoter motifs involved in the network of phytochrome A-regulated gene expression by combined analysis of genomic sequence and microarray data. *Plant Physiol.* 133: 1605-1616.
- Ichikawa, K., Yamabe, Y., Imamura, O., Kuromitsu, J., Sugawara, K., Suzuki, N., Shimamoto, A., Matsumoto, T., Tokutake, Y., Kitao, S., Kataoka, H., Satoh, M., Sugimoto, M., Goto, M., Sugawara, M. and Furuichi, Y. 1997. Cloning and characterization of a novel gene, WS-3, in human chromosome. *Gene* 189: 277-287.
- Ince, T.A. and Scott, K.W. 1995. A conserved downstream element defines a new class of RNA polymerase II promoter. *J. Biol. Chem.* 270: 30249-30252.
- Jarvis, E.E. and Brown, L.M. 1991. Transient expression of firefly luciferase in protoplasts of the green alga *Chlorella ellipsoidea*. *Curr. Genet.* 19: 317-321.
- Jin, E.S., Polle, J.E., Melis, A. 2001. Involvement of zeaxanthin and of the Cbr protein in the repair of photosystem II from photoinhibition in the green alga *Dunaliella salina*. *Biochem. Biophys. Acta* 1506: 244-259.
- Jin, W.W., Li, Z.Y., Fang, Q., Altosaar, I., Liu, L.H. and Song, Y.C. 2002. Fluorescence *in situ* hybridization analysis of alien genes in *Agrobacterium*-mediated *CryIA(b)*-transformed rice. *Ann. Bot.* 90: 31-36.
- Kadonaga, J.T. 2002. The DPE, a core promoter element for transcription by RNA polymerase II. *Exp. Mol. Med.* 34: 259-264.
- Kakinuma, M., Ikeda, M., Coury, D.A., Tominaga, H., Kobayashi, I. and Amano, H. 2009. Isolation and characterization of the *rbcS* genes from a sterile mutant of *Ulva pertusa* (Ulvales, Chlorophyta) and transient gene expression using the *rbcS* gene promoter. *Fish. Sci.* 75: 1015-1028.
- Kamdar, S. J. and Evans, R. 1992. Modifications of the guanidine hydrochloride procedure for the extraction of RNA: isolation from a variety of tissues and adherent/nonadherent cell types. *Biotechniques* 12: 632-638.
- Karimi, M., Depicker, A. and Hilson, P. 2007. Recombinational cloning with plant Gateway vectors. *Plant Physiol.* 145: 1144-1154.
- Karimi, M., Inze, D. and Depicker, A. 2002. GATEWAY vectors for *Agrobacterium*-mediated plant transformation. *Trends Plant Sci.* 7: 193-195.
- Kathiresan, S., Chandrashekar, A., Ravishankar, G.A. and Sarada, R. 2009. *Agrobacterium*-mediated transformation in the green alga *Haematococcus pluvialis* (Chlorophyceae, Volvocales). *J. Phycol.* 45: 642-649.

- Kel-Margoulis, O.V., Kel, A.E., Reuter, I., Deineko, I.V. and Wingender, E. 2002. TRANSCompel: a database on composite regulatory elements in eukaryotic genes. *Nucleic Acids Res.* 30: 332-334.
- Kim, D.H., Kim, Y.T., Cho, J.J., Bae, J.H., Hur, S.B., Hwang, I. and Choi, T.J. 2002. Stable integration and functional expression of flounder growth hormone gene in transformed microalga, *Chlorella ellipsoidea*. *Mar. Biotechnol.* 4: 63-73.
- Kindle, K.L. 1990. High-frequency nuclear transformation of *Chlamydomonas reinhardtii*. *Proc. Natl. Acad. Sci. USA* 87: 1228-1232.
- Kindle, K.L., Schnell, R.A., Fernandez, E. and Lefebvre, P.A. 1989. Stable nuclear transformation of *Chlamydomonas* using the *Chlamydomonas* gene for nitrate reductase. *J. Cell Biol.* 109: 2589-2601.
- Knudsen, S. 1999. Promoter 2.0: for the recognition of PolIII promoter sequences. *Bioinformatics* 15: 356-361.
- Kolchanov, N.A., Ignatieva, E.V., Ananko, E.A., Podkolodnaya, O.A., Stepanenko, I.L., Merkulova, T.I., Pozdnyakov, M.A., Podkolodny, N.L., Naumochkin, A.N. and Romashchenko, A.G. 2002. Transcription Regulatory Regions Database (TRRD): its status in 2002. *Nucleic Acids Res.* 30: 312-317.
- Kota, R., Holtón, T.A. and Henry, R.J. 1999. Detection of transgenes in crop plants using molecular beacon assays. *Plant Mol. Biol. Rep.* 17: 363-370.
- Kovar, J.L., Zhang, J., Funke, R.P. and Weeks, D.P. 2002. Molecular analysis of the acetolactate synthase gene of *Chlamydomonas reinhardtii* and development of a genetically engineered gene as a dominant selectable marker for genetic transformation. *Plant J.* 29: 109-117.
- Kruse, O., Rupprecht, J., Mussnug, J.H., Dismukes, G.C. and Hankamer, B. 2005. Photosynthesis: a blueprint for solar energy capture and biohydrogen production technologies. *Photochem. Photobiol. Sci.* 4: 957-970
- Kuhlemeier, C. 1992. Transcriptional and post-transcriptional regulation of gene expression in plants. *Plant Mol. Biol.* 19: 1-4.
- Kumar, S.C., Misqitta, R.W., Reddy, V.S., Rao, B.J. and Rajam, M.V. 2004. Genetic transformation of the green alga *Chlamydomonas reinhardtii* by *Agrobacterium tumefaciens*. *Plant Sci.* 166: 731-738.
- Kwak, J.M., Kim, S.A., Hong, S.W. and Nam, H.G. 1997. Evaluation of 515 expressed sequence tags obtained from guard cells of *Brassica campestris*. *Planta* 202: 9-17.
- La Claire, J.W. and Herrin, D.L. 1997. Co-isolation of high quality DNA and RNA from coenocytic green algae. *Plant Mol. Biol. Rep.* 15: 263-272.

- Liu, Y.G. and Whittier, R.F. 1995. Thermal asymmetric interlaced PCR: automatable amplification and sequencing of insert end fragments from P1 and YAC clones for chromosome walking. *Genomics* 25: 674-681.
- Liu, Y.G., Chen, Y. and Zhang, Q. 2005. Amplification of genomic sequences flanking T-DNA insertions by thermal asymmetric interlaced polymerase chain reaction. *Methods Mol. Biol.* 286: 341-348.
- Liu, Y.G., Mitsukawa, N., Oosumi, T. and Whittier, R.F. 1995. Efficient isolation and mapping of *Arabidopsis thaliana* T-DNA insert junctions by thermal asymmetric interlaced PCR. *Plant J.* 8: 457-463.
- Lu, Y., Li, J., Xue, L., Yan, H., Yuan, H. and Wang, C. 2011. A duplicated carbonic anhydrase 1 (DCA1) promoter mediates the nitrate reductase gene switch of *Dunaliella salina*. *J. Appl. Phycol.* 23: 673-680.
- Lumbreras, V. and Purton, S. 1998. Recent advances in *Chlamydomonas* transgenics. *Protist* 149: 23-27.
- Manzara, T. and Gruissem, W. 1988. Organization and expression of the genes encoding ribulose-1,5-bisphosphate carboxylase in higher plants. *Photosynthesis Res.* 16: 117-139.
- Marraccini, P., Courjault, C., Caillet, V., Lausanne, F., Lepage, B., Rogers, W.J., Tessereau, S. and Deshayes, A. 2003. Rubisco small subunit of *Coffea arabica*: cDNA sequence, gene cloning and promoter analysis in transgenic tobacco plants. *Plant Physiol. Biochem.* 41: 17-25.
- Maruyama, M., Hordkovd, I., Honda, H., Xing, X.H., Shiragami, N. and Unno H. 1994. Introduction of foreign DNA into *Chlorella saccharophila* by electroporation. *Biotechnol. Techniques* 8: 821-826.
- Masuda T., Tanaka, A. and Melis, A. 2003. Chlorophyll antenna size adjustments by irradiance in *Dunaliella salina* involve coordinate regulation of chlorophyll a oxygenase (CAO) and *Lhcb* gene expression. *Plant Mol. Biol.* 51: 757-771.
- Mayfield, S.P., Franklin, S.E. and Lerner, R.A. 2003. Expression and assembly of a fully active antibody in algae. *Proc. Natl. Acad. Sci. USA* 100: 438-442.
- Mayfield, S.P., Rahire, M., Frank, G., Zuber, H. and Rochaix, J.D. 1987. Expression of the nuclear gene encoding oxygen-evolving enhancer protein 2 is required for high levels of photosynthetic oxygen evolution in *Chlamydomonas reinhardtii*. *Proc. Natl. Acad. Sci. USA* 84: 749-753.
- Mazars, G.R., Moyret, C., Jeanteur, P. and Theillet, C.G. 1991. Direct sequencing by thermal asymmetric PCR. *Nucleic Acids Res.* 19: 4783.
- Melis, A. and Happe, T. 2001. Hydrogen production. Green algae as a source of energy. *Plant Physiol.* 127: 740-748.

- Melis, A., Zhang, L., Forestier, M., Ghirardi M.L. and Seibert, M. 2000. Sustained photobiological hydrogen gas production upon reversible inactivation of oxygen evolution in the green alga *Chlamydomonas reinhardtii*. *Plant Physiol.* 122: 127-136.
- Meng, S., Qian, K., Su, N., Chang, H., Liu, J. and Chen, G.. 2003. Foot-and-mouth disease virus VP1 protein fused with cholera toxin B subunit expressed in *Chlamydomonas reinhardtii* chloroplast. *Biotechnol. Lett.* 25: 1087-1092.
- Metting, E.B. 1996. Biodiversity and application of microalgae. *J. Ind. Microbiol. Biotechnol.* 17: 477-489.
- Michiels, A., Tucker, M., van Den Ende, W. and van Laere, A. 2003. Chromosomal walking of flanking regions from short known sequences in GC-rich plant genomic DNA. *Plant Mol. Biol. Rep.* 21: 295-302.
- Minge, M.A., Shalchian-Tabrizi, K., Torresen, O.K., Takishita, K., Probert, I., Inagaki, Y., Klaveness, D. and Jakobsen, K.S. 2010. A phylogenetic mosaic plastid proteome and unusual plastid-targeting signals in the green-colored dinoflagellate *Lepidodinium chlorophorum*. *BMC Evol. Biol.* 10: 191
- Moore, B.S. 1999. Biosynthesis of marine natural products: microorganisms and macroalgae. *Nat. Prod. Rep.* 16: 653-674.
- Mori, S., Mori, K., Suzuki, I. and Kasumi, T. 2004. Phylogenetic analysis of *Lactococcus lactis* subspecies based on decoding the sequence of the pepT tripeptidase gene, the pepV dipeptidase gene and 16S rRNA. *Syst. Appl. Microbiol.* 27: 414-422.
- Morris, G.J. 1978. Cryopreservation of 250 strains of Chlorococcales by the method of two step cooling. *Br. Phycot. J.* 13: 15-24.
- Mueller, P.R. and Wold, B. 1989. In vivo footprinting of a muscle specific enhancer by ligation mediated PCR. *Science* 246: 780-786.
- Mukhopadhyay, T. and Roth, J.A. 1998. Isolation of total RNA from tissues or cell lines: visualization in gel. *Methods Mol. Biol.* 86: 55-59.
- Nakayama, T., Soma, M., Takahashi, Y., Rehemudula, D., Sato, M., Uwabo, J., Izumi, Y., and Kanmatsuse, K. 1999. Nucleotide sequence of the 5'-flanking region of the type A human natriuretic peptide receptor gene and association analysis using a novel microsatellite in essential hypertension. *Am. J. Hypertens.* 12: 1144-1148.
- Nelson, J.A. and Lefebvre, P.A. 1995. Targeted disruption of the NIT8 gene in *Chlamydomonas reinhardtii*. *Mol. Cell Biol.* 15: 5762-5769.
- Newberg, L.A., Thompson, W.A., Conlan, S., Smith, T.M., McCue, L.A. and Lawrence, C.E. 2007. A phylogenetic Gibbs sampler that yields centroid solutions for cis-regulatory site prediction. *Bioinformatics* 23: 1718-1727.

- Nguyen, P.D., Ho, C.L., Harikrishna, J.A., Wong, M.C.V.L. and Rahim, R.A. 2006. Generation and analysis of expressed sequence tags from the mangrove plant, *Acanthus ebracteatus* Vahl. *Tree Genet. Genomes* 2: 196-201.
- Nichols, H.W. 1973. Growth media-freshwater. In *Handbook of phycological methods: culture methods and growth measurements*, ed. J.R. Stein, pp. 7-24. Cambridge: Cambridge University Press.
- Nikaido, I., Asamizu, E., Nakajima, M., Nakamura, Y., Saga, N. and Tabata, S. 2000. Generation of 10,154 expressed sequence tags from a leafy gametophyte of a marine red alga, *Porphyra yezoensis*. *DNA Res.* 7: 223-227.
- Nomura, M., Katayama, K., Nishimura, A., Ishida, Y., Ohta, S., Komari, T., Miyao-Tokutomi, M., Tajima, S. and Matsuoka, M. 2000. The promoter of *rbcS* in a C3 plant (rice) directs organ-specific, light-dependent expression in a C4 plant (maize), but does not confer bundle sheath cell-specific expression. *Plant Mol. Biol.* 44: 99-106.
- Novina, C.D. and Roy, A.L. 1996. Core promoters and transcriptional control. *Trends Genet.* 12: 351-355.
- Novo, M.T.M., Soares-Costa, A., de Souza, A.Q.L., Figueira, A.C.M., Molina, G.C., Palacios, C.A., Kull, C.R., Monteiro, I.F., Baldan-Pineda, P.H. and Henriques-Silva, F. 2005. A complete approach for recombinant protein expression training: From gene cloning to assessment of protein functionality. *Biochem. Mol. Biol. Educ.* 33: 34-40.
- Ohler, U., Niemann, H., Lia, G.C. and Rubin, G.M. 2001. Joint modeling of DNA sequence and physical properties to improve eukaryotic promoter recognition. *Bioinformatics* 17: S199-S206.
- Ohresser, M., Matagne, R.F. and Loppes, R. 1997. Expression of the arylsulphatase reporter gene under the control of the *nit1* promoter in *Chlamydomonas reinhardtii*. *Curr. Genet.* 31: 264-271.
- Palmer, J.D. 1985. Comparative organization of chloroplast genomes. *Annu. Rev. Genet.* 19: 325-354.
- Palmer, J.D., Soltis, D.E. and Chase, M.W. 2004. The plant tree of life: an overview and some points of view. *Am. J. Bot.* 91: 1437-1445.
- Parkinson, J. and Blaxter, M. 2009. Expressed sequence tags: an overview. *Methods Mol. Biol.* 533: 1-12.
- Pearson, R.B. and Kemp, B.E. 1991. Protein kinase phosphorylation site sequences and consensus specificity motifs: Tabulations. *Methods Enzymol.* 200: 62-81.
- Prestridge, D.S. 1995. Predicting Pol II promoter sequences using transcription factor binding sites. *J. Mol. Biol.* 249: 923-32.

- Prince, R.C. and Kheshgi, H.S. 2005. The photobiological production of hydrogen: potential efficiency and effectiveness as a renewable fuel. *Crit. Rev. Microbiol.* 31: 19-31.
- Rasala, B.A., Muto, M., Lee, P.A., Jager, M., Cardoso, R.M.F., Behnke, C.A., Kirk, P., Hokanson, C.A., Crea, R., Mendez, M. and Mayfield, S.P. 2010. Production of therapeutic proteins in algae, analysis of expression of seven human proteins in the chloroplast of *Chlamydomonas reinhardtii*. *Plant Biotechnol. J.* 8: 1-15.
- Renaud, S.M., Parry, D.L. and Thinh L.V. 1994. Microalgae for use in tropical aquaculture I: Gross chemical and fatty acid composition of twelve species of microalgae from the Northern Territory, Australia. *J. Appl. Phycol.* 6: 337-345.
- Rosenthal, A. and Jones, D.S. 1990. Genomic walking and sequencing by oligo-cassette mediated polymerase chain reaction. *Nucleic Acids Res.* 18: 3095-3096.
- Rudd, S. 2003. Expressed sequence tags: alternative or complement to whole genome sequence. *Trends Plant Sci.* 8: 321-329.
- Rudin, N. 1997. *Dictionary of modern biology*. New York: Barron's Educational Series, Inc.
- Sahdev, S., Saini, S., Tiwari, P., Saxena, S. and Saini, K.S. 2007. Amplification of GC-rich genes by following a combination strategy of primer design, enhancers and modified PCR cycle conditions. *Mol. Cell Probes* 21: 303-307.
- Salzman, R.A., Fujita, T., Zhu-Salzman, K., Hasegawa, P.M. and Bressan, R.A. 1999. An improved RNA isolation method for plant tissues containing high levels of phenolic compounds or carbohydrates. *Plant Mol. Biol. Rep.* 17: 11-17.
- Sambrook, J. and Russell, D.W. 2001. *Molecular cloning: a laboratory manual*, 3rd ed. New York: Cold Spring Harbor Lab Press.
- Sayre, R.T., Wagner, R.E., Sirporanadulsil, S. and Farias, C. 2001. Transgenic algae for delivery antigens to animals. Int. Patent Number W.O. 01/98335 A2.
- Scala, S., Carels, N., Falciatore, A., Chiusano, M.L. and Bowler, C. 2002. Genome properties of the diatom *Phaeodactylum tricorutum*. *Plant Physiol.* 129: 993-1002.
- Scherf, M., Klingenhoff, A. and Werner, T. 2000. Highly specific localization of promoter regions in large genomic sequences by PromoterInspector: a novel context analysis approach. *J. Mol. Biol.* 297: 599-606.
- Schiedlmeier, B., Schmitt, R., Muller, W., Kirk, M.M., Gruber, H., Mages, W. And Kirk, D.L. 1994. Nuclear transformation of *Volvox carteri*. *Proc. Natl. Acad. Sci. USA:* 91: 5080-5084.

- Schroda, M., Blöcker, D. and Beck, C.F. 2000. The HSP70A promoter as a tool for the improved expression of transgenes in *Chlamydomonas*. *Plant J.* 21: 121-131.
- Schwartz, S., Zhang, Z., Frazer, K.A., Smit, A., Riemer, C., Bouck, J., Gibbs, R., Hardison, R. and Miller, W. 2000. PipMaker - a web server for aligning two genomic DNA sequences. *Genome Res.* 10: 577-586.
- Sessions, A., Burke, E., Presting, G., Aux, G., McElver, J., Patton, D., Dietrich, B., Ho, P., Bacwaden, J., Ko, C., Clarke, J.D., Cotton, D., Bullis, D., Snell, J., Miguel, T., Hutchison, D., Kimmerly, B., Mitzel, T., Katagiri, F., Glazebrook, J., Law, M. and Goff, S.A. 2002. A high-throughput *Arabidopsis* reverse genetics system. *Plant Cell* 14: 2985-2994.
- Settles, A.M., Latshaw, S. and McCarthy, D.R. 2004. Molecular analysis of high-copy insertion sites in maize. *Nucleic Acids Res.* 32: e54.
- Shahmuradov I.A., Solovyev V.V. and Gammerman A.J. 2005. Plant promoter prediction with confidence estimation. *Nucleic Acids Res.* 33: 1069-1076.
- Shahmuradov, I.A., Gammerman, A.J., Hancock, J.M., Bramley, Peter, M. and Solovyev, V.V. 2003. PlantProm: a database of plant promoter sequences. *Nucleic Acids Res.* 31: 114-117.
- Shao, N. and Bock, R. 2008. A codon-optimized luciferase from *Gaussia princeps* facilitates the in vivo monitoring of gene expression in the model alga *Chlamydomonas reinhardtii*. *Curr. Genet.* 53: 381-388.
- Shao, Z.T., Cong, X., Yuan, J.D. Yang, G.W., Chen, Y., Pan, J. and An, L.G. 2009. Construction and characterization of a cDNA library from head kidney of Japanese sea bass (*Lateolabrax japonicus*). *Mol Biol Rep.* 36: 2031-2037.
- Sharma, K., Mishra, A.K. and Misra, R.S. 2009. Identification and characterization of differentially expressed genes in the resistance reaction in taro infected with *Phytophthora colocasiae*. *Mol. Biol. Rep.* 36: 1291-1297.
- Shimogawara, K., Fujiwara, S., Grossman, A. and Usuda, H. 1998. High efficiency transformation of *Chlamydomonas reinhardtii* by electroporation. *Genetics* 148: 1821-1828.
- Shivji, M.S., Rogers, S.O. and Stanhope, M.J. 1992. Rapid isolation of high molecular weight DNA from marine macroalgae. *Mar. Ecol. Prog. Ser.* 84: 197-203.
- Shrager, J., Hause, C., Chang, C.W., Harris, E.H., Davies, J., McDermott, R., Tamse, R., Zhang, Z. and Grossman, A.R. 2003. *Chlamydomonas reinhardtii* genome project: A guide to the generation and use of the cDNA information. *Plant Physiol.* 131: 401-408.

- Siebert, P.D., Chenchik, A., Kellogg, D.E., Lukyanov, K.A. and Lukyanov, S.A. 1995. An improved PCR method for walking in uncloned genomic DNA. *Nucleic Acids Res.* 23: 1087-1088.
- Silflow, C.L. 1998. Organization of the nuclear genome. In *The Molecular Biology of Chloroplasts and Mitochondria in Chlamydomonas*, ed. J.D. Rochaix, M. Goldschmidt-Clermont, S. Merchant, pp. 25-40. Dordrecht: Kluwer Academic Publishers.
- Sims, R.J., Belotserkovskaya, R. and Reinberg, D. 2004. Elongation by RNA polymerase II: the short and long of it. *Genes Dev.* 18: 2437-2468.
- Sizova, I., Fuhrmann, M. and Hegemann, P. 2001. A *Streptomyces rimosus* aphVIII gene coding for a new type phosphotransferase provides stable antibiotic resistance to *Chlamydomonas reinhardtii*. *Gene* 277: 221-229.
- Skulberg, O.M. 2000. Microalgae as a source of bioactive molecules-experience from cyanophyte research. *J. Appl. Phycol.* 12: 341-348.
- Smale, S.T. and Kadonaga, J.T. 2003. The RNA polymerase II core promoter. *Annu. Rev. Biochem.* 72: 449-479.
- Specht, E., Miyake-Stoner, S. and Mayfield S. 2010. Micro-algae come of age as a platform for recombinant protein production. *Biotechnol. Lett.* 32: 1373-1383.
- Spreitzer, R.J. 2003. Role of the small subunit in ribulose 1,5-bisphosphate carboxylase/oxygenase. *Arch. Biochem. Biophys.* 414: 141-149.
- Spreitzer, R.J. and Salvucci, M.E. 2002. Rubisco: structure, regulatory interactions, and possibilities for a better enzyme. *Annu. Rev. Plant. Biol.* 53: 449-475.
- Sreenivasulu, N., Kavi Kishor, P.P., Varshney, R.K. and Altschmied, L. 2002. Mining functional information from cereal genomes the utility of expressed sequence tags. *Curr. Sci.* 83: 965-973.
- Steinbrenner, J. and Sandmann, G. 2006. Transformation of the green alga *Haematococcus pluvialis* with a phytoene desaturase for accelerated astaxanthin biosynthesis. *Appl. Environ. Microbiol.* 72: 7477-7484.
- Stevens, D.R., Rochaix, J.D. and Purton, S. 1996. The bacterial phleomycin resistance gene *ble* as a dominant selectable marker in *Chlamydomonas*. *Mol. Gen. Genet.* 251: 23-30.
- Sugita, M., Manzara, T., Pichersky, E., Cashmore, A. and Grissem, W. 1987. Genomic organization, sequence analysis and expression of all five genes encoding the small subunit of ribulose-1,5-bisphosphate carboxylase/oxygenase from tomato. *Mol. Gen. Genet.* 209: 247-256.

- Sun, M., Qian, K., Su, N., Chang, H., Liu, J. and Shen, G. 2003. Foot-and-mouth disease virus VP1 protein fused with cholera toxin B subunit expressed in *Chlamydomonas reinhardtii* chloroplast. *Biotechnol. Lett.* 25: 1087-1092.
- Sun, Y., Yang, Z.Y., Gao, X.S., Li, Q.Y., Zhang, Q.Q. and Xu, Z.K. 2005. Expression of foreign genes in *Dunaliella* by electroporation. *Mol. Biotechnol.* 30: 185-192.
- Surzycki, S. 2000. General aspects of RNA isolation and purification. In *Basic techniques in molecular biology*, ed. S. Surzycki, pp. 119-144. Berlin, Heidelberg, New York: Springer-Verlag.
- Svejstrup, J.Q. 2004. The RNA polymerase II transcription cycle: cycling through the chromatin. *Biochim. Biophys. Acta* 1677: 64-73.
- Szutorisz, H., Dillon, N. and Tora, L. 2005. The role of enhancers as centres for general transcription factor recruitment. *Trends Biochem. Sci.* 30: 593-599.
- Tabita, F.R. 1995. The biochemistry and metabolic regulation of carbon metabolism and CO₂ fixation in purple bacteria. In *Anoxygenic photosynthetic bacteria*, ed. R.E. Blankenship, M.T. Madigan, C.E. Bauer, pp. 885-914. Dordrecht: Kluwer Academic Publishers.
- Tabita, F.R. 1999. Microbial ribulose 1,5-bisphosphate carboxylase/oxygenase: a different perspective. *Photosynth. Res.* 60: 1-28.
- Tamura, K., Dudley, J., Nei, M. and Kumar, S. 2007. MEGA4: molecular evolutionary genetics analysis (MEGA) software version 4.0. *Mol. Biol. Evol.* 24: 1596-1599.
- Tan, C., Qin, S., Zhang, Q., Jiang, P. and Zhao, F. 2005. Establishment of a microparticle bombardment transformation system for *Dunaliella salina*. *J. Microbiol.* 43: 361-365.
- Tang, D.K.H., Qiao, S.Y. and Wu, M. 1995. Insertion mutagenesis of *Chlamydomonas reinhardtii* by electroporation and heterologous DNA. *Biochem. Mol. Biol. Int.* 36: 1025-1035.
- Teng, C., Qin, S., Liu, J., Yu, D., Liang, C. and Tseng, C. 2002. Transient expression of *lacZ* in bombarded unicellular green alga *Haematococcus pluvialis*. *J. Appl. Phycol.* 14: 495-500.
- Teramoto, H., Nakamori, A., Minagawa, J. and Ono, T. 2002. Light-intensity-dependent expression of Lhc gene family encoding light-harvesting chlorophyll-a/b proteins of photosystem II in *Chlamydomonas reinhardtii*. *Plant Physiol.* 130: 325-333.
- Terauchi, R. and Kahl, G. 2000. Rapid isolation of promoter sequences by TAIL-PCR: the 5' flanking regions of *Pal* and *Pgi* genes from yams (*Dioscorea*). *Mol. Gen. Genet.* 263: 554-560.

- Terzaghi, W.B. and Cashmore, A.R. 1995. Light-regulated transcription. *Annu. Rev. Plant Physiol. Plant. Mol. Biol.* 46: 445-474.
- Thanh, T., Omar, H., Abdullah, M.P., Chi, V.T.Q., Noroozi, M., Ky, H. and Napis, S. 2009. Rapid and effective method of RNA isolation from green microalga *Ankistrodesmus convolutus*. *Mol. Biotechnol.* 43: 148-153.
- Thanh, T., Chi, V.T.Q., Abdullah, M.P., Omar, H., Noroozi, M., Ky, H. and Napis, S. 2011a. Construction of cDNA library and preliminary analysis of expressed sequence tags from green microalga *Ankistrodesmus convolutus* Corda. *Mol. Biol. Rep.* 38: 177-182.
- Thanh, T., Chi, V.T.Q., Abdullah, M.P., Omar, H., Noroozi, M. and Napis, S. 2011b. Cloning and characterization of ribulose-1,5-bisphosphate carboxylase/oxygenase small subunit (*RbcS*) cDNA from green microalga *Ankistrodesmus convolutus*. *Mol. Biol. Rep.* 38: 5297-5305.
- Thanh, T., Chi, V.T.Q., Abdullah, M.P., Omar, H. and Napis, S. 2012. Efficiency of ligation-mediated PCR and TAIL-PCR methods for isolation of *RbcS* promoter sequences from green microalga *Ankistrodesmus convolutus*. *Mol. Biol.* 46: 58-64.
- Thapa, A., Shahnawaz, M., Karki, P., Raj Dahal, G., Golam Sharoar, M., Yub Shin, S., Sup Lee, J., Cho, B. and Park, I.S. 2008. Purification of inclusion body-forming peptides and proteins in soluble form by fusion to *Escherichia coli* thermostable proteins. *Biotechniques* 44: 787-796.
- Thomas, D. 2002. *Seaweeds*. London: Natural History Museum.
- Thomas-Chollier, M., Sand, O., Turatsinze, J. V., Janky, R., Defrance, M., Vervisch, E., Brohee, S. and van Helden, J. 2008. RSAT: regulatory sequence analysis tools. *Nucleic Acids Res.* 36: W119-W127.
- Thomas-Hall, S., Campbell, P.R., Carlens, K., Kawanishi, E., Swennen, R., Sági, L. and Schenk, P.M. 2007. Phylogenetic and molecular analysis of the ribulose-1,5-bisphosphate carboxylase small subunit gene family in banana. *J. Exp. Bot.* 58: 2685-2697.
- Thompson, J.D., Higgin, D.G. and Gibson, T.J. 1994. CLUSTAL W: improving the sensitive of progressive multiple sequence alignment through sequence weighting, position specific gap penalties and weigh matrix choice. *Nucleic Acids Res.* 22: 4673-4680.
- Tran, M., Zhou, B., Pettersson, P.L., Gonzalez, M.J. and Mayfield, S.P. 2009. Synthesis and assembly of a full-length human monoclonal antibody in algal chloroplasts. *Biotechnol. Bioeng.* 104: 663-673.
- Tseng, C.K. 2001. Algal biotechnology industries and research activities in China. *J. Appl. Phycol.* 13: 375-380.

- Valentin, K. and Zetsche, K. 1990. Rubisco genes indicate a close phylogenetic relation between the plastids of Chromophyta and Rhodophyta. *Plant Mol. Biol.* 15: 575-584.
- van den Hoek, C., Mann, D.G. and Jahns, H.M. 1995. *Algae: an introduction to phycology*. Cambridge: Cambridge University Press.
- van Helden, J., André, B. and Collado-Vides, J. 1998. Extracting regulatory sites from the upstream region of yeast genes by computational analysis of oligonucleotide frequencies. *J. Mol. Biol.* 281: 827-842.
- Varela-Álvarez, E., Andreakis, N., Lago-Lestón, A., Pearson, G.A., Serrão, E.A., Procaccini, G., Duarte, C.M. and Marbá, N. 2006. Genomic DNA isolation from green and brown algae (*Caulerpales* and *fucales*) for microsatellite library construction. *J. Phycol.* 42: 741-745.
- Vareli, K. and Frangou-Lazaridis, M. 1996. Modification of the acid guanidinium thiocyanate-phenol-chloroform method for nuclear RNA isolation. *Biotechniques* 21: 236-237.
- Verwoerd, T.C., Dekker, B.M.M. and Hoekema, A. 1989. A small-scale procedure for the rapid isolation of plant RNAs. *Nucleic Acids Res.* 17: 23-62.
- Wahlund, T.M., Hadaegh, A.R., Clark, R., Binh, N., Fanelli, M. and Read, B.A. 2004. Analysis of expressed sequence tags from calcifying cells of marine coccolithophorid (*Emiliana huxleyi*). *Mar. Biotechnol.* 6: 278-290.
- Walker, T.L., Collet, C. and Purton, S. 2005a. Review: Algal transgenics in the genomic era. *J. Phycol.* 41: 1077-1093.
- Walker, T.L., Becker, D.K. and Collet, C. 2005b. Characterization of the *Dunaliella tertiolecta RbcS* genes and their promoter activity in *Chlamydomonas reinhardtii*. *Plant Cell Rep.* 23: 727-735.
- Wang, C., Wang, Y., Su, Q. and Gao, X. 2007. Transient expression of the GUS gene in a unicellular marine green alga, *Chlorella sp.* MACC/C95, via electroporation. *Biotechnol. Bioprocess Eng.* 12: 180-183
- Wang, J. and Oard, J.H. 2003. Rice ubiquitin promoters: deletion analysis and potential usefulness in plant transformation systems. *Plant Cell Rep.* 22: 129-134.
- Wang, P., Sun, Y., Li, X., Zhang, L., Li, W. and Wang, Y. 2004. Rapid isolation and functional analysis of promoter sequences of the nitrate reductase gene from *Chlorella ellipsoidea*. *J. Appl. Phycol.* 16: 11-16.
- Watson, G.M. and Tabita, F.R. 1997. Microbial ribulose-1,5-bisphosphate carboxylase/oxygenase: a molecule for phylogenetic and enzymological investigation. *FEMS Microbiol. Lett.* 146: 13-22.

- Waugh, R. and Brown, J.W. 1991. Plant gene structure and expression. In *Plant genetic engineering*, ed. D. Grierson, pp. 1-31. Glasgow and London: Blakie and Son Ltd.
- Weber, A.P., Oesterhelt, C., Gross, W., Bräutigam, A., Imboden, L.A., Krassovskaya, I., Linka, N., Truchina, J., Schneidereit, J., Voll, H., Voll, L.M., Zimmermann, M., Jamai, A., Riekhof, W.R., Yu, B., Garavito, R.M. and Benning, C. 2004. EST-analysis of the thermo-acidophilic red microalga *Galdieria sulphuraria* reveals potential for lipid A biosynthesis and unveils the pathway of carbon export from rhodoplasts. *Plant Mol. Biol.* 55: 17-32.
- Wilkins, T. and Smart, L. 1996. Isolation of RNA from plant tissue. In *A laboratory guide to RNA: isolation, analysis and synthesis*, ed. P.A. Krieg, pp. 21-42. New York: Wiley-Liss.
- Wilkinson, M. 2000. Purification of RNA. In *Essential molecular biology volume one: a practical approach*, ed. T.A. Brown, pp. 69-88. Oxford: Oxford University Press.
- Williams, M.E., Foster, R. and Chua, N.H. 1992. Sequences flanking the hexameric G-box core CACGTG affect the specificity of protein binding. *Plant Cell* 4: 485-496.
- Wingender, E., Chen, X., Fricke, E., Geffers, R., Hehl, R., Liebich, I., Krull, M., Matys, V., Michael, H., Ohnhäuser, R., Prüess, M., Schacherer, F., Thiele, S. and Urbach, S. 2001. The TRANSFAC system on gene expression regulation. *Nucleic Acids Res.* 29: 281-283.
- Wu, W., Welsh, M.J., Kauffman, P.B. and Zhang, H.H. 1997a. *Methods in gene biotechnology*. New York: CRC Press.
- Wu, L., Enberg, A.W. and Guo, X. 1997b. Effects of elevated selenium and salinity concentration in root zone on selenium and salt secretion in saltgrass (*Distichlis spicata* L.). *Ecotoxicol. Environ. Saf.* 37: 251-258.
- Yamagishi, T. 1998. *Guide book to photomicrographs of the freshwater algae*. Tokyo: Uchidarokakuho-shisha Pub.
- Yamazaki, T., Yamamoto, M., Sakamoto, W. and Kawano, S. 2005. Isolation and molecular characterization of *rbcS* in the unicellular green alga *Nannochloris bacillaris* (Chlorophyta, Trebouxiophyceae). *Phycol. Res.* 53: 67-76.
- Yang, L., Xu, S., Pan, A., Yin, C., Zhang, K., Wang, Z., Zhou, Z. and Zhang, D. 2005. Event specific qualitative and quantitative polymerase chain reaction detection of genetically modified MON863 maize based on the 5'-transgene integration sequence. *J. Agric. Food Chem.* 53: 9312-9318.
- Ye, A.H., Jiang, C.J., Zhu, L., Yu, M., Wang, Z.X., Deng, W.W. and Wei, C.L. 2009. Cloning and sequencing of a full-length cDNA encoding the RuBPCase small subunit (*RbcS*) in tea (*Camellia sinensis*). *Agr. Sci. China* 8: 161-166.

- Ying, S.Y. 2004. Complementary DNA libraries: an overview. *Mol. Biotechnol.* 27: 245-252.
- Zawel, L. and Reinberg, D. 1995. Common themes in assembly and function of eukaryotic transcription complexes. *Ann. Rev. Biochem.* 64: 533-561.
- Zhang, M.Q. 1998. Identification of human gene core promoters *in silico*. *Genome Res.* 8: 319-326.
- Zhang, Q., Liu, H. and Cao, J. 2008. Identification and preliminary analysis of a new PCP promoter from *Brassica rapa ssp. chinensis*. *Mol. Biol. Rep.* 35: 685-691.
- Zhou, J., Huang, H., Meng, K., Shi, P., Wang, Y., Luo, H., Yang, P., Bai, Y. and Yao, B. 2010. Cloning of a new xylanase gene from *Streptomyces sp.* TN119 using a modified thermal asymmetric interlaced-PCR specific for GC-rich genes and biochemical characterization. *Appl. Biochem. Biotechnol.* 160: 1277-1292.
- Zhu, J., Liu, J.S. and Lawrence, C.E. 1998. Bayesian adaptive sequence alignment algorithms. *Bioinformatics* 14: 25-39.
- Zhuang, J., Xiong, A.S., Peng, R.H., Gao, F., Zhu, B., Zhang, J., Fu, X.Y., Jin, X.F., Chen, J.M., Zhang, Z., Qiao, Y.S. and Yao, Q.H. 2009. Analysis of *Brassica rapa* ESTs: gene discovery and expression patterns of AP2/ERF family genes. *Mol. Biol. Rep.* 37: 2485-2492.
- Zweiger, G. 2001. *Transducing the genome: information, anarchy and revolution in the biomedical sciences*, 1st ed. New York: McGraw-Hill Companies.