

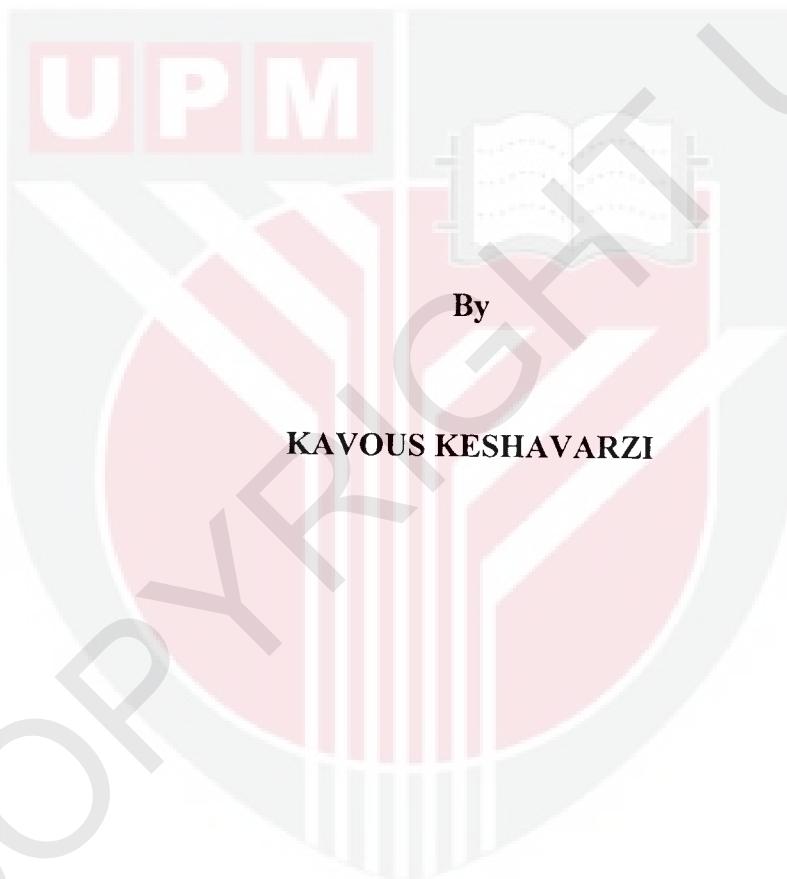


***CHARACTERIZATION AND DETECTION OF AND PHYLOGENETIC  
RELATIONSHIPS AMONG XANTHOMONAS ORYZAE PV. ORYZAE  
ISOLATES FROM SELECTED RICE VARIETIES IN PENINSULAR  
MALAYSIA***

**KAVOUS KESHavarzi**

**FP 2012 25**

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**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

**January 2012**

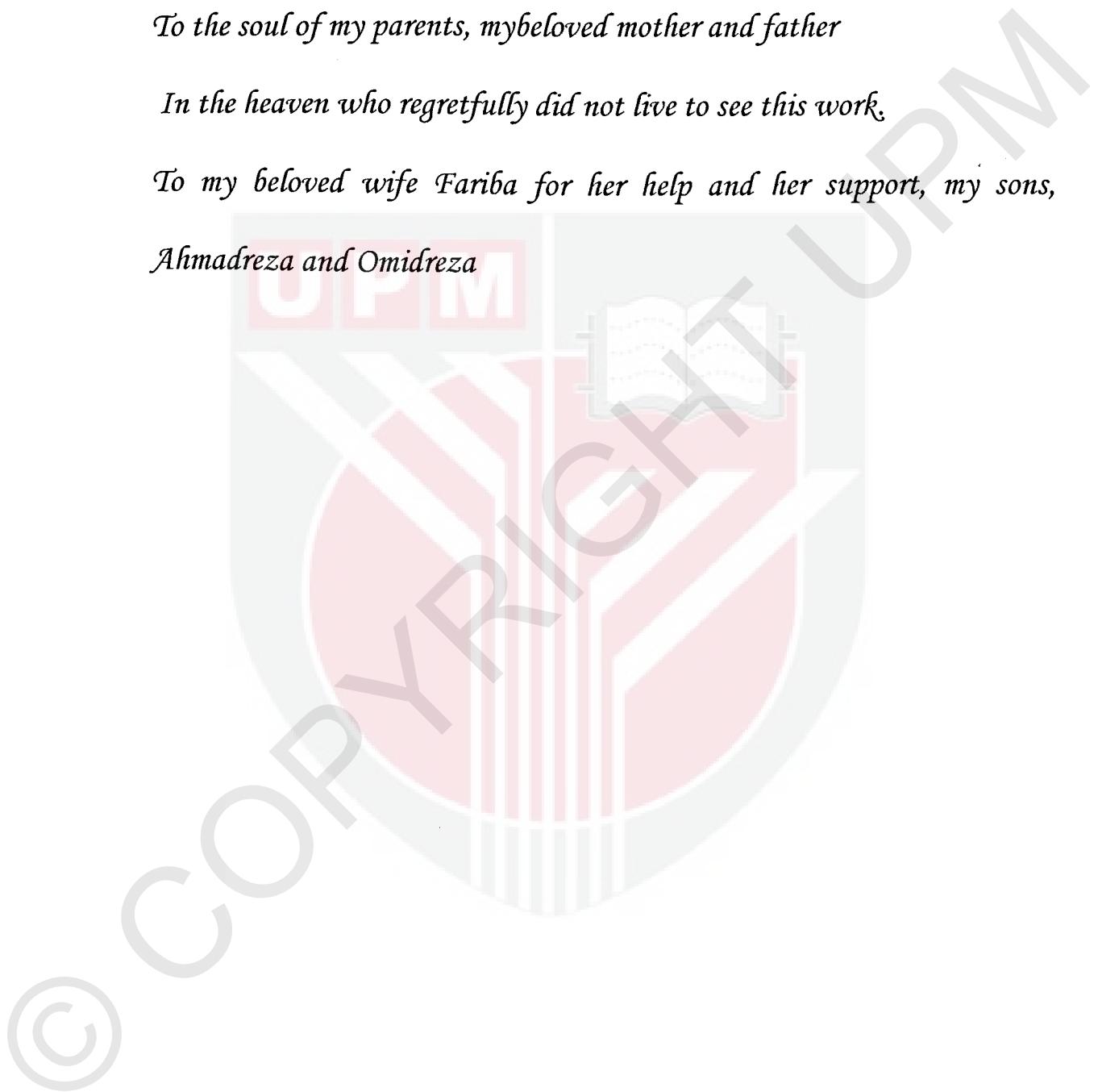
**This work is dedicated to: *All I love Specially***

*To the soul of my parents, my beloved mother and father*

*In the heaven who regrettfully did not live to see this work,*

*To my beloved wife Fariba for her help and her support, my sons,*

*Ahmadreza and Omidreza*



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment  
of the requirement for the degree of Doctor of Philosophy

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RELATIONSHIPS AMONG *XANTHOMONAS ORYZAE* PV. *ORYZAE*  
ISOLATES FROM SELECTED RICE VARIETIES IN PENINSULAR  
MALAYSIA**

By

**KAVOUS KESHAVARZI**

**January 2012**

**Chairman : Associated Professor Dr. Kamaruzaman Sijam, PhD**

**Faculty : Agriculture**

Rice (*Oryza sativa*) is one of the most important food crops in the world and the major crop of Malaysia after oil palm. Bacterial leaf blight (BLB) disease caused by *Xanthomonas oryzae* pv *oryzae* (*Xoo*) is the most important disease in Malaysia. Paddy leaves or stems showing symptoms of disease were collected from rice fields in Penang, Kedah, Selangor, and Melaka in Peninsular Malaysia during the period from 2008 to 2010. The isolated bacteria were found to be oxidase negative, gram negative, anaerobic growth negative and did not produce fluorescent pigment. Thirty collected strains showed leaf blight symptoms on inoculated rice plants, hypersensitive reaction on tobacco, and hydrolysis on gelatin. Pathogenic tests on two rice varieties MR84 and IR8 showed that significant differences were observed between 30 strains. All strains of *Xoo* were classified into four groups, i.e. non-

virulent, slightly virulent, moderately virulent and highly virulent. Seven strains, including MXO1 and SXO8 were highly virulent, five strains (including SXO1 and MXO6) were moderately virulent, while 17 strains represented by PXO43 and KXO182 showed low virulence, and the only non-virulent strain was PXO36. It can be concluded that there is a relation between location and severity of disease that the most and the lowest severity was related to the strains of Melaka and Penang respectively and also the strains from the different rice-growing areas or states were diverse and differed in their degree of virulence. For detection of *Xoo*, infected tissues were crushed and the DNA was extracted using a modified CTAB method. A PCR product of about 470 bp was produced using the XOR-F/XOR-R2 primer pair that amplified the 16S-23S intergenic rDNA sequence region. Comparison indicated that the 30 strains have 98% to 99% nucleotide homology to the sequences of 16S-23S rDNA of *Xoo* accessions ABO26287 and AY251OO4 from Japan and China, respectively. Another PCR product of about 964 bp was generated by the primer pair TXT/TXT4R that amplified the insertion sequence element IS1113 and had 99% homology to the sequences of the *Xoo* accessions GU982970 and AF482989 from India and USA, respectively. According to these results, the causal agent of bacterial leaf blight of rice could be identified in less than three hours. Phylogeny analysis among Malaysian strains indicated genomic diversity of *Xoo* strains exist throughout Malaysia. Additionally, strains of *Xoo* could be divided into two main groups, namely Penang– Kedah and Selangor-Melaka that are neighboring states respectively an indication that geographical area is the predominant factor influencing strain variability. And also in the states the diversities of *Xoo* collected from rice cultivars generally were similar. The generated banding patterns resulting from the amplification of DNA templates of the 30 *Xoo* strains by REP, ERIC and BOX

primers were reproducible as was proven with at least three PCR replications, suggesting the suitability of using these primers as markers for genetic studies of *Xoo*. There was a slight difference between the phylogenetic tree patterns derived from respective REP, ERIC and BOX primer sets. The primers generated four common amplification bands in all strains tested, of the sizes 200, 400, 500 and 800 bp. Cluster analysis based on REP, ERIC and BOX primer sets and the pooled data resulting from all three primers (BER primer) indicated that the 30 *Xoo* strains were divided into two main groups with similarity between about 60% or into four subgroups, representing strain from Penang, Kedah, Selangor and Melaka. Based on these results, there were significant similarities among 30 strains of *Xoo* that have been collected from the same origin in Malaysia, indicating that *Xoo* strains originating from different geographic regions in Peninsular Malaysia were phylogenetically different. Hence, this research indicated that strains of *Xoo* isolated from Malaysia are pathologically, genetically and geographically diverse.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan ijazah Doktor Falsafah

**PENCIRIAN, PENGESANAN DAN HUBUNGAN FILOGENI STRAIN  
*XANTHOMONAS ORYZAE PV. ORYZAE* DARI TANAMAN PADI  
DI SEMENANJUNG MALAYSIA**

Oleh

**KAVOUS KESHAVARZI**

**Januari 2012**

**Pengerusi: Profesor Madaya Dr. Kamaruzaman Sijam, PhD**

**Fakulti: Pertanian**

Padi (*Oryza sativa*) merupakan salah satu tanaman makanan penting dunia dan merupakan juga tanaman makanan utama Malaysia selepas kelapa sawit. Tanaman ini sering dijangkiti beberapa penyakit kulat dan bakteria termasuk penyakit hawar daun bakteria (BLB) yang disebabkan oleh *Xanthomonas oryzae* pv *oryzae* (*Xoo*) atau sebelum ini dikenali sebagai *Xanthomonas campestris* pv *oryzae*. Daun atau batang padi yang menunjukkan simptom penyakit hawar telah dikumpul dari sawah padi di Semenanjung Malaysia dari negeri-negeri Pulau Pinang, Kedah, Selangor dan Melaka dari tahun 2008 hingga 2010. Pengkulturan koloni bakteria yang dipencil telah dilakukan. Bakteria yang dipencil didapati bersifat oxidase negatif, Gram negatif, mempunyai pertumbuhan anaerobik negatif dan tidak menghasilkan pigmen pendarfluor. Tiga puluh isolat *Xoo* yang dikumpul telah memberikan simptom penyakit hawar daun bakteria pada tanaman padi yang diinokulat serta memberikan tindak balas hipersensitif ke atas pokok tembakau dan hidrolisis pada gelatin. Ujian

patogenik pada varieti padi MR84 dan IR8 menunjukkan bahawa strain-strain bakteria tersebut boleh dibahagikan kepada empat kumpulan atau strain berdasarkan kapada tahap kevirulenan mereka. Tujuh strain termasuk MXO1 dan SXO8 menunjukkan tahap kevirulenan yang sangat tinggi. Sebaliknya lima strain (termasuk SXO1 dan MXO6) menunjukkan tahap kevirulenan sederhana, manakala 17 strain yang diwakili oleh PXO43 dan KXO182 menunjukkan tahap kevirulenan rendah, dan PXO36 pula merupakan satu-satunya strain yang tidak virulen. Keputusan juga menunjukkan bahawa strain yang dikumpul dari kawasan geografi yang berlainan adalah berbeza dari segi kevirulenan mereka ke atas tanaman padi. Untuk pengesanan serta-merta jangkitan *Xoo*, sedikit tisu yang dijangkiti telah dihancurkan dan DNA diekstrak dengan menggunakan kaedah CTAB yang diubahsuai. Produk PCR bersaiz 470 bp telah dihasilkan dengan menggunakan pasangan primer XOR-F/XOR-R2 yang mengamplifikasi jujukan intergenik 16S-23S rDNA. Perbandingan menunjukkan bahawa 30 strain berkenaan mempunyai tahap homologi nukleotida antara 98% hingga 99% kepada jujukan intergenik 16S-23S rDNA strain-strain *Xoo* ABO26287 dan AY251OO4 dari Jepun dan China, setiap satunya. Satu lagi produk PCR bersaiz 964 bp telah juga dijana oleh pasangan primer TXT/TXT4R yang mengamplifikasi unsur jujukan IS1113 yang mempunyai tahap homologi setinggi 99% dengan jujukan strain-strain GU982970 dan AF482989 dari India dan Amerika Syarikat, setiap satunya. Menurut keputusan ini, agen penyebab penyakit bakteria hawar daun padi telah dapat dikenalpasti sebagai *Xoo* dalam masa kurang daripada tiga jam penganalisaan. Analisa filogenetik menunjukkan keujudan kepelbagaian genomik diantara strain-strain *Xoo* di Malaysia. Strain-strain *Xoo* dapat dibahagikan kepada dua kumpulan utama negeri berjiran iaitu Penang-Kedah dan Selangor-Melaka, yang menunjukkan bahawa kawasan geografikal merupakan faktor penentu

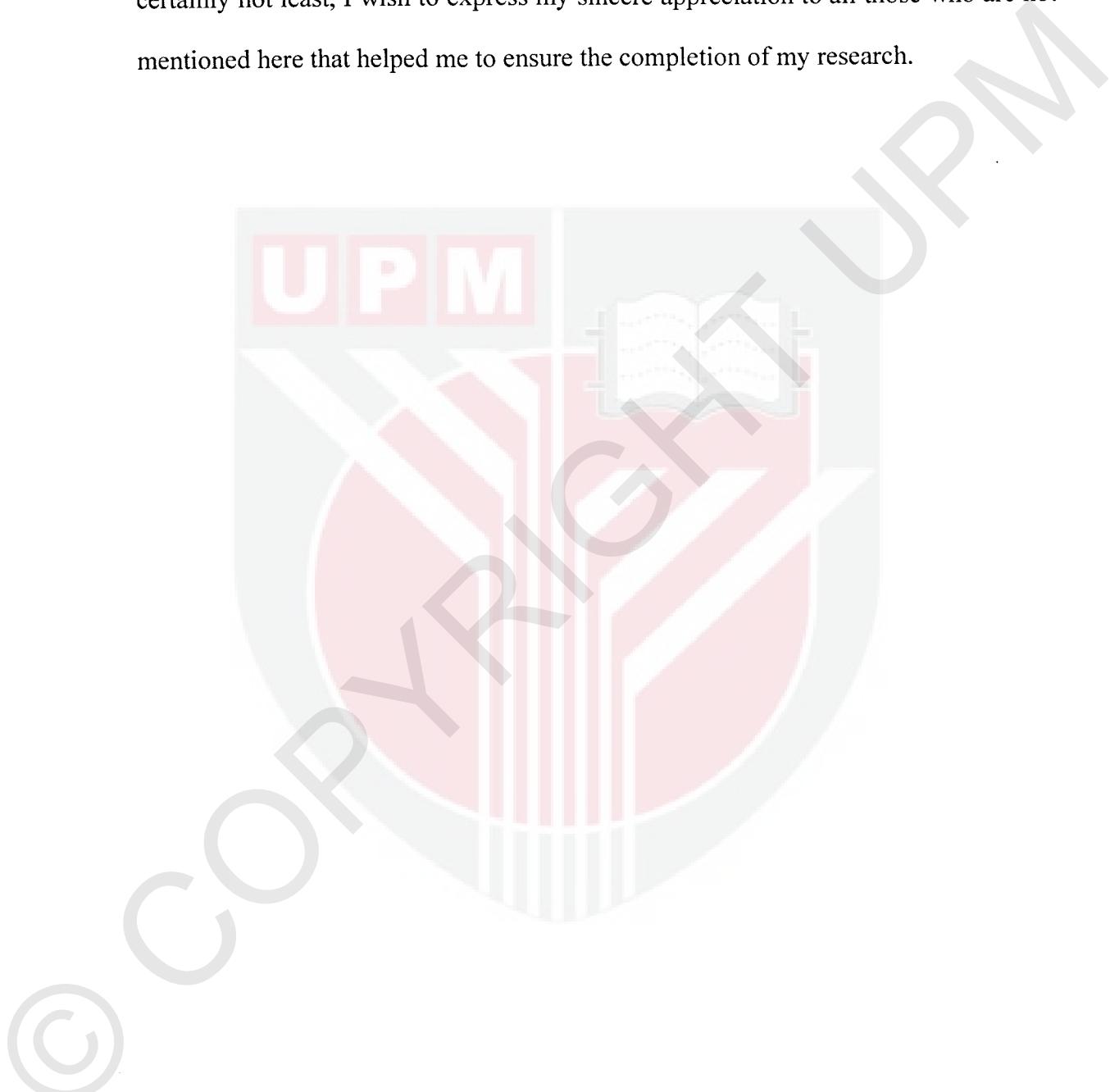
utama dalam kepelbagaian strain. Strain-strain dari sesuatu negeri tidak menunjukkan perbezaan yang ketara dintara mereka. Corak penghasilan “band” dari amplifikasi templat DNA dari 30 strain *Xoo* dengan menggunakan primer-primer REP, ERIC dan BOX adalah konsisten dan boleh dihasil semula. Ini menyarankan kesesuaian penggunaan pasangan-pasangan primer berkenaan dalam kajian genetik *Xoo*. Terdapat sedikit perbezaan antara corak pokok filogenetik yang bersandarkan dari data REP, ERIC atau set primer BOX. Penggunaan primer-primer tersebut telah menghasilkan empat jujukan amplifikasi yang serupa dari semua strain yang dikaji iaitu yang bersaiz 200, 400, 500 dan 800 bp.. Analisis kluster berdasarkan hasilan primer-primer REP, ERIC, BOX dan hasilan data dari ketiga-tiga pasangan primer yang disatukan (BER primer) menunjukkan bahawa 30 strain *Xoo* boleh dibahagikan kepada dua kumpulan utama dengan kesamaan diantara mereka pada tahap 60% atau kapada empat sub-kumpulan mewakili strain dari Pulau Pinang, Kedah, Selangor dan Melaka. Berdasarkan kepada keputusan ini bolehlah disimpulkan bahawa ujud ketaksamaan diantara 30 strain *Xoo* yang dikumpulkan dari lokasi berbeza di Semenanjung Malaysia. Strain dari setiap negeri dikelompokkan bersama. Strain-strain dari negeri jiran mempunyai tahap persamaan yang tinggi jika dibandingkan dengan strain dari negeri yang berjauhan, yang menunjukkan bahawa strain-strain *Xoo* yang berasal dari kawasan geografikal yang berlainan adalah berbeza secara filogenetik. Penyelidikan ini telah berjaya menunjukkan kepelbagaian strain *Xoo* yang dipencarkan dari Malaysia berdasarkan kapada aspek-aspek patologi, genetik dan kawasan geografikal.

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## LIST OF ABBREVIATIONS

BLAST	Basic Local Alignment Search Tool
BLB	Bacterial leaf blight
BOX	Single primer sequence
bp	Base pair
CFU	Colony forming unit
CTAB	Cetyl trimethyl-ammonium bromide
DNA	Deoxyribonucleic acid
DNTP	Deoxyribonucleotides (dATP, dCTP, dGTP, dTTP)
EDTA	Ethylene diaminetetraacetic acid, disodium salt
ELISA	Enzyme linked immunosorbent Assay
ERIC	Enterobacterial repetitive intergenic consensus
NCBI	National Center for Biotechnological Information
PCR	Polymerase chain reaction
PVP	Poly vinyl pyrrolidone
RAPD	Randomly Amplified Polymorphic DNA
rDNA	Ribosomal DNA
REP	Repetitive extragenic palindromic elements sequence
RFLP	Restriction fragment length polymorphism
rpm	Revolution per minute
rRNA	Ribosomal RNA
TBE	Tris Boric acid EDTA
UV	Ultraviolet
W/V	Weight/volume

*Xoo*

*Xanthomonas oryzae* pv *oryzae*



## CHAPTER 1

### INTRODUCTION

Rice (*Oryza sativa* L) is one of the most important food crops in the world, being the staple diet of about half of the world population (Akhtar et al., 2008). This plant belongs to the grass family, and is related to other grass species such as wheat, oats and barley, which together produce food grains known as cereals. Grist, (1965) postulated that the cultivated species may have developed from species of wild rice. It was reported that at least twenty five wild rice species were well distributed in the tropical and subtropical regions in Asia, Africa, America and Northern Australia (Grist, 1965).

The world rice supply had more than doubled from 261 million tonnes in 1950 to 673 million tonnes in 1997. Rice production is expected to increase by 50 percent by the year 2025 (Khush and Virk, 2000). World rice production in 2004 was just less than 610 million tonnes. At least, 114 countries grow rice and more than 50 countries have an annual production of about or more than 100,000 tonnes. Asian farmers produce about 90% of the world's rice due to favorable hot and humid climate in their regions, with two countries, namely China and India, growing more than half of the total crop (International Rice Research Institute, 2005). Interestingly, less than 5% of the total rice production in the world is traded in the international market (Hossain, 1997) and because of that rice is considered as a security crop or commodity in many countries in Asia.

Rice cultivation has a long history in Malaysia. It is believed that rice had spread down from its original centre of origin during the period of 4000-5000BC (Abdullah et al., 1991; Othman et al., 1990) and became established as one of its center of diversity for

the species. Currently, rice has become the second major crop in Malaysia after oil palm. It is an important part of the diet of the people of Malaysia. The total planted area is about 667,310 ha, and the total rice production is about 2,030,000 tonnes (MOA, 2005).

However, the rice crop is frequently infected by several fungal and bacterial diseases that affect its production. One of the important diseases of rice is the bacterial leaf blight (BLB) disease caused by *Xoo* (Gammaproteobacteria, Xanthomonadales, Xanthomonadaceae). It is one of the most important bacterial diseases of rice, which usually infect the crop during the monsoon season (from June to September) in the south-east Asian and Indian subcontinent countries (Mew, 1987; Mew et al., 1993). Bacterial leaf blight was first detected in Japan in 1884, but was only identified as a disease in 1922 (Mew et al., 1993). The disease then spread to all continents of the world, except Europe (Ezuka and Kaku, 2000; Anonymous, 2007; Ghasemie et al., 2008). Rice is the main host of *Xoo*, which is also a seed-borne bacterium. The disease spreads through dispersion of plant straw, wind, rain, hail, wild rice, weeds (Nino-Liu et al., 2006), irrigation water and seeds (Nyvall, 1999). Bacterial inoculum enters the plant tissues either through wounds or water pores in the leaf and then travels systemically throughout the plant xylem (Nino-Liu et al., 2006). Infected plants show leaves wilting and rolls up, turning grayish-green to yellow, until the whole seedling dies. Surviving older plants have dried leaf blades. Lesions begin as water-soaked stripes on the leaf blades and eventually increase in length and width, turning yellow to grayish-white until the whole leaf dries up (Agrios, 1997). Infected plants also show either streaks or leaf blight symptoms (Akhtar et al., 2003). Infection by *Xoo* could cause yield loss of up to 50% in tropical Asia (Anonymous, 2007). In 1988 and 1994,

serious *Xoo* outbreaks in Malaysia were reported in the states of Penang, Kedah, Selangor and Perak, where more than 40% of planted areas were infected with the disease, causing an estimated yield loss of about 10-50% (Saad et al., 2003).

Biochemical tests (Ghasemie et al., 2008), pathogenic races of *Xoo* (Adhikari et al., 1999), fatty acids (Chase et al., 1992) and metabolic profiling (Jones et al., 1993) have been employed as tools for detection of the pathogen. The genome of a *Xanthomonas* strain, KAC10331 has been sequenced (Dharmapuri and Sonti, 1999). Several researchers have used polymerase chain reaction (PCR) technology as a tool to detect and study the variability of the pathogenic bacteria (Li and De Boer, 1995). The 16S ribosomal DNA sequence (16S rDNA) has been used for deducing phylogenetic and evolutionary relationships among bacteria and other prokaryote species (Weisberg et al., 1991). Adachi and Oku (2000) used the XOR-F and XOR-R2 primer pair to amplify a fragment of the intergenic spacer region, 16S-23S rDNA, of *Xanthomonas* species. On the other hand, the TXT and TXT4R primer pair has been used to amplify the insertion sequence element IS1113 of *Xanthomonas oryzae* pathovars (Sakthivel et al., 2001).

Another PCR based technique, known as Rep-PCR has also been used in the identification and classification of bacteria (Versalovic et al., 1991). It is a repetitive sequence based genomic fingerprinting that uses primers that correspond to the endogenous interspersed repetitive sequences of the bacteria. These interspersed sequences are highly conserved elements in the prokaryotic genomes. Sequences of these elements have been extensively characterized in several prokaryotic microorganisms (Versalovic et al., 1991). Families of repetitive DNA sequences, like repetitive extragenic palindromic (REP), enterobacterial repetitive intergenic consensus

(ERIC), and single primer (BOX) elements, which are present in all prokaryotes, could be used for bacterial fingerprinting, and have been used effectively for analysis of several species of bacteria (Louws et al., 1994; Louws et al., 1999). As for the *Xanthomonas* species, rep-PCR was used to assess variations among several pathovars and has been shown to have low levels of intropathovar diversity (Louws et al., 1995; Vauterin et al., 2000).

The occurrence of *Xoo* has been reported in Malaysia (Saad et al., 2003; Saad and Habibuddin, 2010), but information on the diversity of *Xoo* causing pathogen at the molecular level is still lacking. The present research was designed to focus on the detection and confirmation of *Xoo* infection using PCR method on samples of leaves and stems collected from rice fields. The study was also conducted to determine the diversity of Malaysian strains of *Xoo*, based on sequences of the intergenic region of 16S-23S rDNA and the insertion sequence element, IS1113 of the bacterial genome. In addition to that, this study was also designed to assess the phylogenetic relationships between local populations of *Xoo* strains from rice fields in Peninsular Malaysia using REP, ERIC and BOX PCR primers. The specific objectives of the study were to:

1. Characterize phenotypes of *Xanthomonas oryzae* pv *oryzae* from rice plants in Peninsular Malaysia.
2. Detect and identification *Xanthomonas oryzae* pv *oryzae* using universal and specific primers.
3. Investigate the diversity among the strains of *Xanthomonas oryzae* pv *oryzae* using molecular markers.

## REFERENCES

- Abdullah, M.Z., Vaughan, D. A., and Mohammad, O. (1991). Wild relatives of rice in Malaysia: Their characteristics, distribution, ecology and potential in rice breeding. *MARDI Report No, 145*, 28p. Serdang, MARDI.
- Adachi, N., and Oku, T. (2000). PCR-mediated Detection of *Xanthomonas oryzae* pv. *oryzae* by Amplification of the 16S–23S rDNA Spacer Region Sequence. *Journal of General Plant Pathology*, 66(4), 303-309.
- Adhikari, T. B., Cruz, C. M. V., Zhang, Q., Nelson, R. J., Skinner, D. Z., Mew, T. W., and Leach, J. E. (1995). Genetic diversity of *Xanthomonas oryzae* pv. *oryzae* in Asia. *Applied and Environmental Microbiology*, 61(3), 966–971.
- Adhikari, T. B., and Mew, T. W. (1985). Antibiotic sensitivity of *Xanthomonas campestris* pv. *oryzicola* in vitro. *International Rice Research Newsletter*, 10, 19.
- Adhikari, T. B., Mew, T. W., and Leach, J. E. (1999). Genotypic and Pathotypic diversity in *Xanthomonas oryzae* pv. *oryzae* in Nepal. *Phytopathology*, 89(8), 687-694.
- Agarwal, P. C., Mortensen, C. N., and Mathur, S. B. (1989). Seed-borne diseases and seed health testing of rice. *Phytopathological Papers*. CAB International, Wallingford, UK, 30.
- Agrios, G. N. (1997). Plant Pathology. 4th ed. Academic Press. New York. USA. pp: 849
- Ahmed, H. U., Finckh, M. R., Alfonso, R. F., and Mundt, C. C. (1997). Epidemiological effect of gene deployment strategies on bacterial blight of rice. *Phytopathology*, 87, 66 -70.
- Akhtar, M. A., Rafi, A., and Hameed, A. (2008). Comparison of methods of inoculation of *Xanthomonas oryzae* pv *oryzae* in rice cultivar. *Pakistan Journal Botanic*, 40(5), 2171-2175.
- Akhtar, M. A., Zakeri, M., Abassi, F. M., and Masood, M. A. (2003). Incidence of bacterial blight of rice in Pakistan during 2002. *Pakistan Journal Botanic*, 35(5), 993-997.
- Aldrick, S. J., Buddenhagen, I. W., and Reddy, A. P. K. (1973). The occurrence of bacterial leaf blight in wild and cultivated rice in Northern Australia. *Australian Journal of Agricultural research*, 24, 219-227.
- Alias, I. (2010). Breeding for yield improvement in rice. *National Rice conference, Lumut, Malaysia*. Pp 51-62.
- Alias, I., Othman, O., Mohammad, H., Saad, A., Habibuddin, H., Abdulrahman, A.B. and Azlan, S. (2005). Varieti padi baru MR220. *Bulletin Teknologi Tanaman*, 2, 7-13.
- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., and Lipman, D. J. (1990). Basic local alignment search tool. *Journal of Molecular Biology*, 215(3), 403-410.

Amuthan, G., Elumalai, R. P., Ulaganathan, K., Rajini Rani, D. B., and Mahadevan, A. (1992). Variation in plasmid profile of *Xanthomonas* of Indian origin. *Indian Journal of Experimental Biology (New Delhi)*, 30, 808-810.

Anonymous (1990). *Xanthomonas oryzae*. *Organ Eurpian Mediterranean Protection Plant (OEPP/EPPPO)*, Data Sheets on Quarantine Pests, 1-8.

Anonymous (2007). *Xanthomonas oryzae*. Bulletin *Organ Eurpian Mediterranean Protection Plant (OEPP/EPPPO)*, 37, 543-553.

Anuratha, C. S. and Gnanamanickam, S. S. (1987). *Pseudomonas fluorescens* suppresses development of bacterial blight symptoms. *International Rice Research Newsletter*, 12(1), 17.

Ardales, E. Y., Leung, H., Vera Cruz, C. M., Mew, T. W., Leach, J. E., and Nelson, R. J. (1996). Hierarchical analysis of spatial variation of the rice bacterial blight pathogen across diverseAcro ecosystems in the Philippines. *Phytopathology*, 86, 241-252.

Aritua, V., Nanyonjo, A., Kumakech, F., and Tushemereirwe, W. (2007). Rep-PCR reveals a high genetic homogeneity among Ugandan isolates of *Xanthomonas campestris* pv *musacearum*. *African Journalof Biotechnology*, 6 (3), 179-183.

Benedict, A. A., Alvarez, A. M., Berestecky, J., Imanaka, W., Mizumoto, C. Y., Pollard, L. W., Mew, T. W., and Gonzalez,C. F. (1989). Pathovar-specific monoclonal antibodies for *Xanthomonas campestris* pv. *oryzae* and for *Xanthomonas campestris* pv. *oryzicola*. *Phytopathology*, 79(3), 322-328.

Bouzar, H., Jones, J. B., Stall, R. E., Louws, F. J., Schneider, M., Rademaker, J. L. W., De Bruijn, F. J., and Jackson L. E.(1999). Multiphasic analysis of xanthomonads causing bacterial spot disease on tomato and pepper in the Caribbean and Central America: Evidence for common lineages within and between countries. *Phytopathology*, 89(4), 328-335.

Bradbury, J. F. (1984). *Xanthomonas* Dowson 1939, p. 199-210. In N. R. Krieg and J. G. Holt (ed.), *Bergey's manual of systematic bacteriology*, vol 1. The Williams and Wilkins Co., Baltimore.

Chase, A. R., Stall, R. E., Hodge, N. C. and Jones, J. B. (1992). Characterization of *Xanthomonas campestris* strains from aroids using physiological, pathological, and fatty acid analyses. *Phytopathology*, 82(7), 754-759.

Cottyn, B., Bautista, A. T., Nelson, R. J., Swings, J. and Mew, T. W. (1994). Polymerase chain reaction amplification of DNA from bacterial pathogens of rice using specific oligonucleotide primers. International Rice Research Notes (IRRN) 19, 30-32

Cowan, S. T. (1974). . Manual for the identification of Medical bacteria. 2nd Ed. *Cambridge University Press, Great Britain*, 238.

- Cubero, J. and Graham, J. H. (2002). Genetic relationship among worldwide strains of *Xanthomonas* causing canker in citrus species and design of new primers for their identification by PCR. *Applied and Environmental Microbiology*, 68(3), 1257.
- Dardick, C., Goes Da Silva, F., Shen, Y. and Ronald, P. (2003). Antagonistic interactions between strains of *Xanthomonas oryzae* pv *oryzae*. *Phytopathology*, 93, 705-711.
- Dath, A. P. and Devadath, S. (1983). Role of inoculum in irrigation water and soil in the incidence of bacterial blight of rice. *Indian Phytopathology*, 36, 142-144.
- Dath, D. P., Padmanabhan, S. Y. and Devadath, S. (1978). Effect of soil type on the length produced by *Xanthomonas oryzae*. *Scienceand Culture*. 44(9), 417-418.
- De Bruijn, F. J. (1992). Use of repetitive (repetitive extragenic palindromic and enterobacterial repetitive intergeneric consensus) sequences and the polymerase chain reaction to fingerprint the genomes of *Rhizobium meliloti* isolates and other soil bacteria. *Applied and Environmental Microbiology*, 58(7), 2180-2187.
- Devadath, S. (1989). Chemical control of bacterial blight of rice. *International Rice Research Institute, Manila, Philippines*, 89- 98
- De Vos, P., Goor, M. Gillis, M. and De Ley. J.(1985). Ribosomal ribonucleic acid cistron similarities of phytopathogenic *Pseudomonas* species. *International Journal of Systematic Bacteriology*, 35, 169-184.
- Dharmapuri, S. and Sonti, R. V. (1999). A transposon insertion in the gumG homologue of *Xanthomonas oryzae* pv. *oryzae* causes loss of extracellular polysaccharide production and virulence. *FEMS Microbiology Letters (Amsterdam)*, 179(1), 53-59.
- Di, M., Ye, H. Z., Schaad, N. W. and Roth, D. A. (1991). Selective recovery of *Xanthomonas spp* from rice seeds. *Phytopathology*, 81, 1358-1363.
- Dikin, A. (1992). Studies on Bacterial Blight of Rice: Development of Seed Health Testing Methods, Seed Transmission, Survival of Bacterial Pathogens in the Seed. *Danish Government Institute of Seed Pathology for Developing Countries, Heller up (DK)*.
- Exconde, O. R. (1973). Yield losses due to bacterial leaf blight of rice. *Philippines Agriculture*, 57, 128-140.
- Ezuka, A. and Kaku, H. (2000). A historical review of bacterial blight of rice. *Bulletin of the National Institute of Agro biological Resources*, Japan. 15, 1-207.
- Fang, C. R., Lin, C. F. and Chu, C. L. (1956). A preliminary study on the disease cycle of the bacterial leaf blight of rice. *Acta Phytotaxonomica Sinica*, 2, 173-185.
- Food and Agriculture Organisation (FAO). (2005). *Top Paddy Rice Producers*.
- George, M. L. C., Cruz, W. T. Leach, J. E. and Nelson, R. J. (1997). Movement of *Xanthomonas oryzae* pv. *oryzae* in Southeast Asia detected using PCR-based DNA fingerprinting .*Phytopathology* 87: 302-309.

- George, M. L. C., Cruz, W. T. and Nelson, R. J. (1994). DNA fingerprinting of *Xanthomonas oryzae* pv. *oryzae* by ligation-mediated polymerase chain reaction. *International Rice Research Notes*, 19, 29-30.
- Ghasemie, E., Kazempour, M. N. and Padasht, F. (2008). Isolation and identification of *Xanthomonas oryzae* pv. *oryzae* the causal agent of bacterial blight of rice in Iran. *Journal of Plant Protection Research*, 48(1), 53-63.
- Gnanamanickam, S. S., Priyadarisini, V. B., Narayanan, N. N., Vasudevan, P. and Kavitha, S. (1999). An overview of bacterial blight disease of rice and strategies for its management. *Current Science*, 77(11), 1435-1444.
- Gnanamanickam, S. S., Shigaki, T., Medalla, E. S., Mew, T. W. and Alvarez, A. M. (1994). Problems in detection of *Xanthomonas oryzae* pv. *oryzae* in rice seed and potential for improvement using monoclonal antibodies. *Plant Disease*, 78, 173-178.
- Goel, A.K., Rajagopal, L., Nagesh, N. and Sonti, R.V. (2002). Genetic locus encoding functions involved in biosynthesis and outer membrane localization of xanthomonadin in *Xanthomonas oryzae* .pv. *oryzae*. *Journal of Bacteriology*, 184, 3539–3548.
- Gonzalez, C., Szurek, B., Manceau, C., Mathieu, T., Sére, Y. and Verdier, V. (2006). Molecular and pathotypic characterization of new *Xanthomonas oryzae* strains from West Africa. *Molecular Plant-Microbe Interactions*, 20(5), 534-546.
- Goto, M. (1964). Kresek and pale yellow leaf, systematic symptoms of bacterial leaf of rice caused by *Xanthomonas oryzae*. *Plant Disease Reporter*, 48, 858-861.
- Goto, M. (1992). *Fundamentals of Bacterial Plant Pathology*.
- Grist, D. H. (1965). Rice. The Origin and History of Rice. *Longmans, Green and Co LTD*, Pp 3-10.
- Higgins, C. F., McLaren, R. S. and Newbury, S. F. (1988). Repetitive extragenic palindromic sequences, mRNA stability and gene expression evolution by gene conversion. *A Review Gene* 72, 3-14.
- Hossain, M. (1997). Supply, demand and production potential of rice in Asia. *Regional overviews*.[http://www.riceweb.org/\\_overesia.htm](http://www.riceweb.org/_overesia.htm). 10.08.2011
- Hsish, S. P.Y. and Budenhagen,I. W, (1974).suppressing effects of *Erwinia herbicola* on infection by *Xanthomonas oryzae*andon symptom development in rice. *Phytopathology*, 64, 1182-1185.
- <http://tejaratalvand.com/pdf/rice.pdf> 10.08 2011
- Hugh, R. and Leifson, E. (1953). The taxonomic significance of fermentative versus oxidative metabolism of carbohydrates by various gram-negative rods. *Journal of Bacteriology*, 66, 24-26.

Hulton, C. S. J., Higgins, C. F. and Sharp, P. M. (1991). ERIC sequences: a novel family of repetitive elements in the genomes of *Escherichia coli*, *Salmonella typhimurium* and other enterobacteria. *Molecular Microbiology*, 5, 825-834.

International Rice Research Institute (IRRI). (2005). *Rice around the world*. <http://irri.org/science/cnyinfo/index.asp>, 15.08 2011

International Rice Research Institute (IRRI), (1977). *Genetic Evaluation and Utilization Program*.

Jagoueix, S., Bove, J. and Garnier, M. (1994). The phloem-limited bacterium of greening disease of citrus is a member of the alpha subdivision of the Proteobacteria. *International Journal of Systematic and Evolutionary Microbiology*, 44(3), 379.

Jones, J., Chase, A. and Harris, G. (1993). Evaluation of the Biolog GN Micro Plate system for identification of some plant-pathogenic bacteria. *Plant Disease*, 77, 553-558.

Kauffman, H. E. and Reddy, A. P. K., (1975). Seed transmission studies of *Xanthomonas oryzae* in rice. *Phytopathology* 65, 663-666.

Kauffman, H. E., Reddy, A. P. K., Hsieh, S. P. Y. and Merca, S. D. (1973). An improved technique for evaluating resistance of rice varieties to *Xanthomonas oryzae*. *Plant Disease Reporter*, 57, 537-541.

Kaul, M. L. H. and Sharma, K. K. (1987). Bacterial blight of rice. *A review* *Biologisches Zentralblatt*, 106, 141-167.

Keshavarz, K., Sijam, K., Zainal Abidin, M. A., Habibudin, H. and Nazerian, E. (2011). Rapid Identification and Differentiation of *Xanthomonas oryzae* pv *oryzae* Strain with Primer 16S-23S rDNA from the Rice Fields in Peninsular Malaysia. *Asian Journal of Plant Pathology*, 5, 93-99.

Khush, G. S. and Virk, P. S. (2000). Rice breeding: achievements and future strategies. *Crop Improvement Society*. ISSN: 02560933, 27, 115-144.

Kim, H. M. and Song, W. Y. (1996). Characterization of ribosomal RNA intergenic spacer region of several seed borne bacterial pathogens of rice. *Seed Science and Technology*, 24, 571-580.

Kinoshita, T. (1995). Report of committee on gene symbolization, nomenclature and linkage groups. *Rice Genetic Newsletters* 12, 9-115

Klement, Z. and Goodman, R. (1967). The hypersensitive reaction to infection by bacterial plant pathogen. *Annual Review Phytopathology*, 5, 17-44.

Leach, J.E., White, F.F., Rhoads, M.L. and Leung, H. (1990). A repetitive DNA sequence differentiates *Xanthomonas campestris* pv. *oryzae* from other pathovars of *X. campestris*. *Molecular Plant-Microbe Interactions*, 3, 238-246.

- Leach, J. E., Rhoads, M. L., Vera Cruz, C. M., White, F. F., Mew, T. W. and Leung, H, (1992). Assessment of genetic diversity and population structure of *Xanthomonas oryzae* pv. *oryzae* with a repetitive DNA element. *Applied Environment Microbiology*, 58, 2188-2195.
- Lee, B. M., Park, Y. J., Park, D. S., Kang, H. W., Kim, J. G., Song, E. S., Park, I. C., Yoon, U. H., Hahn, J. H., Koo, B. S., Lee, G. B., Kim, H., Park, H. S., Yoon, K. O., Kim, J. H., Jung, C. H., Koh, N. H., Seo, J. S. and Go, S. J. (2005). The genome sequence of *Xanthomonas oryzae* KACC10331, the bacterial blight pathogen of rice. *Nucleic Acids Research*, 33(2), 577.
- Lee, B. M., Young, J. P., Dong, S. P., Jeong, G. K., Hee, W. K., Tae, H. N., Gil, B. L. and Joung, K. A. (2004). PCR-Base sensitive detection and identification of *Xanthomonas oryzae* pv. *oryzae*. *Korean Journal of Microbiology*, 32, 256-264.
- Lee, C. N., Hu, R. M., Chow, T. Y., Lin, J. W., Hui, Y. C., Tseng, Y. H. and Weng, S. F, (2007). Comparison of Genomes of Three *Xanthomonas oryzae* Bacteriophage. *BMC Genomics*, 8, 442
- Lee, S. W., Choi, S. H., Han, S. S., Lee, D. G. and Lee, B. Y. (1999). Distribution of *Xanthomonas oryzae* pv. *oryzae* strains virulent to Xa21 in Korea. *Phytopathology*, 89, 928-933
- Lelliott, R. A. and Stead, D. E. (1987). Methods for the Diagnosis of Bacterial Diseases of Plants, Methods in Plant Pathology (2 ed.): Blackwell Scientific Publications Oxford (GB).
- Leyns, F., De Cleene, M. Swings, J. and De Ley, J. (1984). The host range of the genus *Xanthomonas*. *Botanical Review*, 50, 308-356.
- Li, X. and De Boer, S. H. (1995). Selection of polymerase chain reaction primers from an RNA intergenic spacer region for specific detection of *Clavibacter michiganensis* subsp *sepedonicus*. *Phytopathology*, 85, 837-842.
- Li, Z. Z., Zhao, H. and Ying, X. D., (1985). The weed carriers of bacterial leaf blight of rice. *Acta Phytopathological Sinica*, 15, 246-248.
- Lin, X., Wang, C. and Wen, G. (1998). Progress in map-based cloning of Xa-22(1), a new gene for bacterial blight resistance in rice. *Paper presented at Plant and Animal Genome*, San Diego, CA, USA, Jan 18-22.
- Little, E. L., Bostock, R. M. and Kirkpatrick, B. C. (1998). Genetic characterization of *Pseudomonas syringae* pv. *syringae* strains from stone fruits in California. *Applied and Environmental Microbiology*, 64(10), 3818.
- Louws, F. J., Bell, J., Medina-Mora, C. M., Smart, C. D., Opgenorth, D., Ishimaru, C. A., Hausbeck, M. K., de Bruijn, F. J. and Fulbright, D. W (1999). Rep-PCR-mediated genomic fingerprinting: A rapid and effective method to identify *Clavibacter michiganensis*. *Phytopathology*, 88(8), 862-868.

Louws, F. J. and Cuppels, D. A. (2001). Molecular techniques. *Laboratory Guide for Identification of Plant Pathogenic Bacteria*, 3, 321-333.

Louws, F. J., Fulbright, D. W., Stephens, C. T. and De Bruijn, F. J. (1994). Specific genomic fingerprints of phytopathogenic *Xanthomonas* and *Pseudomonas* pathovars and strains generated with repetitive sequences and PCR. *Applied Environment Microbiology* 60, 2286-2295.

Louws, F. J., Fulbright, D. W., Stephens, C. T. and DeBruijn, F. J. (1995). Differentiation of genomic structure by rep-PCR fingerprinting to rapidly classify *Xanthomonas campestris* pv. *vesicatoria*. *Phytopathology*, 85, 528-536.

Louws, F. J., Rademaker, J. L. W. and De Bruijn, F. J. (1998). The three Ds of PCR-based genomic analysis of phytobacteria: diversity, detection, and disease diagnosis. *Annual Review of Phytopathology*, 37, 81-125.

Louws, F. J., Schneider, M. and De Bruijn, F. J. (1996). Assessing genetic diversity of microbes using repetitive sequence-based PCR (rep-PCR). *Nucleic Acid Amplification Methods for the Analysis of Environmental Samples* (Toranzos, G., Ed.), 63-94.

Lozano, J. C. (1977). Identification of bacterial leaf blight in rice cased by *Xanthomonas oryzae* in America. *Plant Disease*, 61, 644-648.

McGrath, S., and van Sinderen, D., (2007). Bacteriophage: *Genetics and Molecular Biology* (1st ed.). Caister Academic Press. ISBN 978-1-904455-14-1

Mew, T. W. (1987). Current status and future prospects of research on bacterial blight of rice. *Annual Review of Phytopathology*, 25(1), 359-382.

Mew, T. W., Alvarez, A. M., Leach, J. E. and Swings, J. (1993). Focus on bacterial blight of rice. *Plant. Disease*, 77(1), 5-12.

Mew, T. W. and Mistra, J. K. (1994). A Manual of Rice Seed Health Testing. *IRRI, Manila (PH)*.

Mew, T. W. and Vera Cruz, C. M., (1979). Variability of *Xanthomonas oryzae*: specificity in infection of rice differentials. *Phytopathology* 69,152-155.

Ministry of Agriculture and Agro Based Industry (MOA). (2005). Malaysia. [www.malaysia.gov.my/.../IndustryInMalaysia/.../](http://www.malaysia.gov.my/.../IndustryInMalaysia/.../) 15.09 2010

Miyoshi, T., Sawada, H., Tachibana, Y. and Matsuda, I. (1998). Detection of *Xanthomonas campestris* pv. *citri* by PCR using primers from the spacer region between the 16s and 23s rRNA genes. *Annals of the Phytopathological Society of Japan*, 64, 249-254

Mizukami, T. and Wakimoto, S. (1969). Epidemiology and control of bacterial leaf blight of rice. *Annual Review of Phytopathology*, 7, 51-72

- Mohan, S. K. and Rao, Y. P. (1974). Bacterial blight and leaf streak diseases of rice, their epidemiology and control. In Raychaudhuri, S. P. and Verma, J. P. eds. *Current Trends in Plant Pathology. University of Lucknow, India.* 129-133
- Moore, W. E. C., Cato, E. P. and Moore, L. V. H. (1985). Index of the bacterial and yeast nomenclatural changes published in the International Journal of Systematic Bacteriology since the 1980 Approved Lists of Bacterial Names (1 January 1980 to 1 January 1985). *International Journal of Systematic and Evolutionary Microbiology*, 35(3), 382.
- Mortensen, C. N., Manandhar, H. K., Cahyaniati, A. and Haryanti, S. E. (1994). Pathogenic bacteria associated with rice seed samples from Indonesia and Nepal. Paper presented at the Plant Pathogenic Bacteria, Versailles (France), 8th International Conference Les colloquies, and no. 66. INRA, Paris
- Murray, M. G. and Thompson, W. F (1980). Rapid isolation of high molecular weight plant DNA. *Nucleic Acids Reserch*, 8,4321-4325.
- Murty, V. S. T. and Devadath, S. (1981). Studies on the transmission and survival of *Xanthomonas campestris* pv *oryzae* through insects. *Indian Phytopathology*, 34, 162-163
- Murty, V. S. T. and Devadath, S. (1982). Survival of *Xanthomonas campestris* pv *oryzae* and its phage in field water at different temperature. *Indian Phytopathology*, 35, 25-31.
- Nelson, R.J., Baraoidan, M. R., Vera Cruz, C.M., Yap, I.V., Leach, J.E., Mew, T.W. and Leung, H. (1994). Relationship between phylogeny and pathotype for the bacterial blight pathogen of rice. *Applied Environment Microbiology*, 60,3275–3283.
- Nino-Liu, D. O., Ronald, P. C. and Bogdanove, A. J. (2006). *Xanthomonas oryzae* pathovars: model pathogens of a model crop. *Molecular Plant Pathology*, 7(5), 303-324.
- Noda, T., Du, P. V., Dinh, H. D and Kaku, H, (1999).Pathogenesity of *Xanthomonas oryzae* pv *oryzae* strains in Vietnam. *Annual of Phytopathological society Japan*, 65, 293-296
- Nyvall, R. F. (1999). Field crop diseases: Iowa State University Press.
- Ochiai, H., Horino, O., Miyajima, K. and Kaku, H. (2000). Genetic diversity of *Xanthomonas oryzae* pv. *oryzae* strains from Sri Lanka. *Phytopathology*, 90(4), 415-421.
- Ochiai, H., Inoue, Y., Takeya, M., Sasaki, A. and Kaku, H. (2005). Genome sequence of *Xanthomonas oryzae* pv. *oryzae* suggests contribution of large numbers of effector genes and insertion sequences to its race diversity. *Japan Agricultural Research Quarterly*, 39(4), 275-287

On, S. L. W. and Holmes. B. 1995. Classification and identification of *campylobacter*, *helicobacter*, and allied taxa by numerical analysis of phenotypic characters. *Systematic and Applied Microbiology*, 18374–390

Othman, O., Alias, I. and Hadzim, K. (1990). Rice varietal development in Peninsular Malaysia. *MARDI Report No, 109*, 18p, Serdang: MARDI.

Ou, S. H. (1985). Rice Diseases. 2nd edition. Commonwealth Mycological Institute, Kew, Surrey (GB).

Pal, V., Gardan, L. and Charles, M. (1988). Isolation of a plasmid from strain of *Xanthomonas oryzae* pv *oryzae* that cause bacterial blight in rice. *International Rice Research Newsletter*, 13, 2.

Paterson, A. H. Brubaker, C. L. and Wendel, J. F, (1993). A rapid method for extraction of Cotton (*Gossypium* spp) genomic DNA suitable for RFLP or PCR analysis. *Plant Molecular Biology Reporter*, 11,122-127.

Rademaker, J. L. W., Hoste, B., Louws, F. J., Kersters, K., Swings, J., Vauterin, L., Vauterin, P. and De Bruijn, F. J. (2000). Comparison of AFLP and rep-PCR genomic fingerprinting with DNA--DNA homology studies: *Xanthomonas* as a model system. *International Journal of Systematic and Evolutionary Microbiology*, 50(2), 665-677

Rademaker, J. L. W., Louws, F. J., Schultz, M. H., Rossbach, U., Vauterin, L., Swings, J. and De Bruijn, F. J. (2005). A comprehensive species to strain taxonomic framework for *Xanthomonas*. *Phytopathology*, 95(9), 1098-1111.

Rademaker, J. L. W., Louws, F. J., Versalovic, J. and De Bruijn, F. J. (2004). Characterization of the diversity of ecologically important microbes by rep-PCR genomic fingerprinting. *Molecular microbial ecology manual. Volumes 1 and 2*(Ed. 2), 611-643.

Ray,S.K., Rajeshwari,R. and Sonti,R.V, (2000). Mutants of *Xanthomonas oryzae* deficient in general secretory pathway are virulent deficient and unable to secrete xylase. *Molecular Plant-Microbe Interactions*, 13, 394–401.

Reckhaus, P. M. (1983). Occurrence of bacterial blight of rice in Niger, West Afrigha. *Plant Disease*, 67, 1037.

Reddy, A. P. K., Mackenzie, D. R. and Rao, A. V. (1979). Relationship of bacterial leaf blight severity to grain yield of rice. *Phytopathology*, 69, 967-969.

Reddy, P. R. (1983). Evidence of seed transmission of *Xanthomonas campestris* pv. *oryzae*. *Current Science*, 52,265-266.

Reddy, P. R. and Nayak, P. (1984). Progress of bacterial leaf blight disease in mixed population of resistant and susceptible rice cultivars. *Acta Phytopathologica Academiae Scientiarum Hungaricae* 19(3-4), 285- 290

- Saad, A. and Habibuddin, H. (2010). Pathotypes and virulence of *Xanthomonas oryzae* pv *oryzae* causing bacterial blight of rice in Peninsular Malaysia. *Journal Tropical Agriculture and Food Science*, 38(2), 257-266
- Saad, A., Habibuddin, H., Alias, I., Othman, O., Azlan, S. and Zulkifli, R. (2003). Impact and contribution of resistant varieties in rice pest management in Malaysia. Pp. 356-363. In Modern Rice Farming. *Proc.an International Rice Conference, Oct 13-16. Alor Star, Malaysia*.
- Saad, A., Habibuddin, H., Alias, I., Othman, O., Azlan, S. and Zulkifli, R. (2000). Resistance status of released varieties after MR 84 against Bacterial Blight and the incidence on the disease in Muda irrigation scheme. Paper presented at the Conf. *Plant Resource Management, Kuching, and Sarawak, Malaysia*.
- Sakthivel, N., Mortensen, C. N. and Mathur, S. B. (2001). Detection of *Xanthomonas oryzae* pv. *oryzae* in artificially inoculated and naturally infected rice seeds and plants by molecular techniques. *Applied Microbiology Biotechnology*, 56, 435–441
- Salzberg, S. L., Sommer, D. D., Schatz, M. C., Phillippy, A. M., Rabinowicz, P. D., Tsuge, S., Furutani ,A., Ochiai, H., Delcher, A.L., Kelley, D., Madupu, R., Puiu, D., Radune, D., Shumway, M., Trapnell, C., Aparna, G., Jha, G., Pandey, A., Patil, P.B., Ishihara, H., Meyer, D. F., Szurek, B., Verdier, V., Koebnik, R., Dow, J. M., Ryan, R. P., Hirata, H., Tsuyumu, S., Won Lee, S., Seo, Y. S., Sriariyanum, M., Ronald, P.C., Sonti, R. V., Van Sluys, M. A., Leach, J. E., White, F. F. and Bogdanove, A. J.(2008). Genome sequence and rapid evolution of the rice pathogen *Xanthomonas oryzae* pv. *oryzae* PXO99A. *Bio Med Central (BMC) Genomics*, 9, 204.
- Schaad, N. W. (1980). Relationship of incidence of seedborne *Xanthomonas campestris* to black rot of crucifers. *Plant Disease*, 64, 91-92.
- Schaad, N. W., Cheong, S. S., Tamaki, S., Hatziloukas, E. and Panopoulos, N. J. (1995). A combined biological and enzymatic amplification technique to detect *Pseudomonas syringae* pv *phaseolicola* in bean seed extracts. *Phytopathology*, 85, 243–248
- Schaad, N. W., Jones, J. B. and Chun, W. (2001). Laboratory Guide for Identification of Plant Pathogenic Bacteria. *American Phytopathological Society*, APS PRESS.
- Semagn, K., Bjornstad, A. and Ndjidjop, M. N. (2006). An overview of molecular marker methods for plants. *African Journal of Biotechnology*, 5(25), 2540-2568.
- Shekhawat, G. S. and Srivastava, D. N. (1968). Variability in Indian isolates of *Xanthomonas oryzae* (Uyeda and Ishiyama) Dowson, the incitants of bacterial leaf blight of rice. *Annual of the Phytopathological Society of Japan*, 31, 289-297.
- Shen, Y. and Ronald, P. (2002). Molecular determinants of disease and resistance in interactions of *Xanthomonas oryzae* pv. *oryzae* and rice. *Microbes and Infection*, 4(13), 1361-1367.
- Sheng, Z. J., Zhen, L. Y. and Jun, F.-X. (2005). Detection of QTL conferring resistance to bacterial leaf streak in rice chromosome 2 (*Oriza sativa*L. spp. *indica*). *Scientia Agricultura Sinica*, 38, 1923–1925.

- Singh, R. N. (1971). Perpetuation of bacterial blight disease of paddy and preservation of its incitant: I. Survival of *Xanthomonas oryzae* in water. *Indian Phytopathology*, 24:153-154.
- Sinha, S. K. and Nene, Y. L. (1967). Eradication of the seed born inoculum of *Xanthomonas oryzae* by hot water treatment of paddy seeds. *Plant Disease*, 5,882-883.
- Srivastava, D.N. and Rao, Y. P. (1964). Seed transmission and epidemiology of bacterial blight disease of rice in North India. *Indian Phytopathology*, 17,77-78.
- Srivastava, D. N. and Rao, Y. P. (1968). Epidemiology and prevention of bacterial blight disease of rice in India. *International Common Newsletter*, 17(1),27-33.
- Stackebrandt, E., Murrap R. G. E. and Truper, H. G, (1988). Proteobacteria classis Nov. A name for the phylogenetic taxon that includes the “purple bacteria and their relatives.” *International Journal of Systematic Bacteriology*, 38,321-325.
- Stead, D. E (1989). Grouping of *Xanthomonas campestris* pathovars of cereals and grasses by fatty acid profiling. *BulletenOrgan Eurpian Mediterranean Protection Plant* (OEPP/EPPO).
- Stern, M. J., Ames, G. F. L., Smith, N. H., Robinson, E. C. and Higgins, C. F. (1984). Repetitive extragenic palindromic sequences: a major component of the bacterial genome. *Cell* 37, 1015-1026.
- Suslow, T. V., Schroth, M. N. and Isaka, M. (1982). Application of a rapid method for gram differentiation of plant pathogenic and saprophytic bacteria without staining. *Phytopathology*, 72, 917-918.
- Swings, J., Van Den Mooter, M., Vauterin, L., Hoste, B., Gillis, M. and Mew, T. W. (1990). Reclassification of the causal agents of bacterial blight (*Xanthomonas campestris* pv. *oryzae*) and bacterial leaf streak (*Xanthomonas campestris* pv. *oryzicola*) of rice as pathovars of *Xanthomonas oryzae* (ex Ishiyama 1922) sp. nov., nom. rev. *International Journal of Systematic Bacteriology*, 40,309-311.
- Tabei, H. (1967). Anatomical studies of rice plant affected with bacterial leaf blight, with special reference to stomata infection at the coleoptiles and the foliage leaf sheath of rice seedling. *Annual of Phytopathological Society Japan*, 33, 12-16
- Tagami, Y. and Mizukami, T. (1962). Historical review of the researchers on bacterial leaf blight of rice caused by *Xanthomonas oryzae*. *Special report of the plant disease and insect pests forecasting service No. 10*.*Plant Protection Division, Ministry of Agriculture and Forestry, Tokyo, Japan*. 112.
- Tamura, K., Dudley, J., Nei, M. and Kumar, S. (2007). MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. *Molecular Biology and Evolution*, 24, 1596-1599.
- Tang, D., Wu, W., Li, W., Lu, H. and Worland, A.J. (2000). Mapping of QTLs conferring resistance to bacterial leaf streak in rice. *Theoretical and Applied Genetics*, 101, 286-291.

- Thompson, D. and Henry, R. J. (1995). Single step protocol for preparation of plant tissue for analysis by PCR. *Biotechniques*, 19, 394-397.
- Trinh, T. T. (1980). New rice diseases and insects in the Senegal River Basin in 1978/79. *International rice commonNewsletter*, 29(2), pp. 37.
- Ulaganathan, K. and Mahadevan, A. (1991). Plasmids in phytopathogenic bacteria. *Indian Journal Scientific and Resource*, 4, 376.
- Umesh, K. C., Davis, R. M. and Gilbertson, R. L. (1998). Seed contamination thresholds for development of carrot bacterial blight caused by *Xanthomonas campestris* pv. *carotae*. *Plant Disease*, 82(11), 1271-1275.
- Valluvaparadesan, V. and Mariappan, V. (1989). Alternate hosts of rice bacterial blight(BB) pathogen *Xanthomonas campestris* pv.*oryzae*. *International Rice Research Newsletter*, 14(5), 27-28.
- Vandamme, P., Poit, B. M., Gillis, M., De Vos, P., Kersters, K. and Swing, J. (1996). Polyphasic Taxonomy, a Consensus Approach to Bacterial Systematics. *American Society for Microbiology*, 60, 407-438.
- Van den Mooter, M. and J. Swings. (1990). Numerical analysis of 295 phenotypic features of 266 *Xanthomonas* and related strains and an improved taxonomy of the genus. *International Journal Systematic Bacteriology*, 40, 348-369.
- Van den Mooter, M. J., Swings, F., Gosselt, K., Kersters, K. and De Ley, J. (1987). The taxonomy of *Xanthomonas Dowson* 1939, p. 795-796. In E. L. Civerolo, A. Collmer, R. E. Davis, and A. G. Gillaspie (ed.), *Plant pathogenic bacteria*. Martinus Nijhoff Publishers, Dordrecht, the Netherlands.
- Vauterin, L., Rademaker, J. and Swings, J. (2000). Synopsis of the taxonomy of the genus *Xanthomonas*. *Phytopathology*, 90, 677-682.
- Vera Cruz, C. M., Gossele, F., Kersters, K., Segers, P., Van den Mooter, M., Swings, J. and De Ley, J. (1984). Differentiation between *Xanthomonas campestris* pv. *oryzae*, *Xanthomonas campestris* pv. *oryzicola* and the bacterial 'brown blotch' pathogen on rice by numerical analysis of phenotypic features and protein gel electrophoregrams. *Journal of General Microbiology*, 130(11), 2983-2999.
- Vera Cruz, C. M. V., Ardales, E. Y., Skinner, D. Z., Talag, J., Nelson, R. J., Louws, F. J., Leung, H., Mew, T. W. and Leach, J. E. (1996). Measurement of haplotypic variation in *Xanthomonas oryzae* pv. *oryzae* within a single field by rep-PCR and RFLP analyses. *Phytopathology*, 86 (12), 1352-1359.
- Versalovic, J., Koeuth, T. and Lupski, J. R. (1991). Distribution of repetitive DNA sequences in eubacteria and application to fingerprinting of bacterial genomes. *Nucleic Acid Research*, 19, 6823-6831.
- Versalovic, J., Schneider, M., De Bruijn, F. J. and Lupski, J. R. (1994). Genomic fingerprinting of bacteria using repetitive sequence based polymerase chain reaction. *Methods in Molecular and Cellular Biology*, 5, 25-40.

Weisburg, W. G., Barns, S. M., Pelletier, D. A. and Lane, D. J. (1991). 16S ribosomal DNA amplification for phytopathogenic study. *Journal Bacteriology*, 173, 697-703.

Yashitola, J., Krishnaveni, D., Reddy, A. P. K. and Sonti, R. V. (1997). Genetic diversity within the population of *Xanthomonas oryzae* pv. *oryzae* in India. *Phytopathology*, 87(7), 760-765.

Yoshimura, S., Yoshimura, A., Iwata, N., McCouch, S. R., Abenes, M. L., Baraoidan, M. R., Mew, T.W. and Nelson, R. J. (1995). Tagging and combining bacterial blight resistance genes in rice using RAPD and RFLP markers. *Molecular Breeding*, 1(4), 375-387.

Zhang, Y., Uyemoto, J. K. and Kirkpatrick, B. C. (1998). A small-scale procedure for extracting nucleic acids from woody plants infected with various phytopathogens for PCR assay. *Journal of Virological Methods*, 71(1), 45-50.

Zhang, Z., Naqvi, N. and Sim, S. (1997). Fine mapping of blast and bacterial leaf blight resistance genes introgressed from wild species *Oryza minuta*. Abstract presented at the general meeting of the International Program on Rice Biotechnology. Paper presented at the Melaka, Malaysia, and Sep 15-19.

Zhu, W., Magbanua, M.M. and White, F.F. (2000). Identification of two novel *hrp*-associated genes in the *hrp* gene cluster of *Xanthomonas oryzae* pv. *oryzae*. *Journal Bacteriology*, 182, 1844-1853.