

UNIVERSITI PUTRA MALAYSIA

EFFECTS OF SUBLETHAL TEMPERATURE STRESS ON THE GROWTH, SURVIVAL AND CULTURABILITY OF LISTERIA MONOCYTOGENES

MOHD NIZAM BIN LANI

FSMB 2002 28

EFFECTS OF SUBLETHAL TEMPERATURE STRESS ON THE GROWTH, SURVIVAL AND CULTURABILITY OF *LISTERIA MONOCYTOGENES*

MOHD NIZAM BIN LANI

MASTER OF SCIENCE UNIVERSITI PUTRA MALAYSIA 2002



EFFECTS OF SUBLETHAL TEMPERATURE STRESS ON THE GROWTH, SURVIVAL AND CULTURABILITY OF *LISTERIA MONOCYTOGENES*

By

MOHD NIZAM BIN LANI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirement for the Degree of Master of Science

November 2002



my ayahanda and bonda my brothers; Daus, Oshin, Afiz, Md Nor and Adik Im my sisters; Angah, Khairiah and Azma my relatives, lecturers and friends

To:

for their love and support who have been inspiring my life



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

EFFECTS OF SUBLETHAL TEMPERATURE STRESS ON THE GROWTH, SURVIVAL AND CULTURABILITY OF *LISTERIA MONOCYTOGENES*

By

MOHD NIZAM BIN LANI

November 2002

| Chairman : | | Abdul | Reezal | bin A | bdul | Latif, | Ph | .D. |
|------------|--|-------|--------|-------|------|--------|----|-----|
|------------|--|-------|--------|-------|------|--------|----|-----|

Faculty : Food Science and Biotechnology

Survival and growth of *Listeria monocytogenes* L56 (IMR isolate) was studied in trypticase soy broth grown at 37°C before being subjected to three selected sublethal stress of temperatures (55°C, 28°C and 4°C) using log and stationary phase as inoculums using two-plating systems; TSA with and without 4% NaCl (TSAS). The influence of morphological changes and listerial motility as affected by sublethal stress of temperatures were also determined using Scanning Electron Microscopy (SEM) and motility media, respectively. A standard growth curve of *L. monocytogenes* at 37°C was established using plate counts showed that the log and stationary phase of the organism were achieved after 12 and 19 hours, respectively. It was observed that viable bacterial population (CFU/ml) after log and stationary phase were 10^8 and 10^9 , respectively. From the growth curve, the generation time of *L. monocytogenes* at 37°C was 60 min.



The bacterial growth rates obtained from culturability on culture plates assessed using two-system media, TSA with and without 4% NaCl concentration (TSAS) were assessed by their generation time. Cells of *L. monocytogenes* grown in exponential phase cultures demonstrated biphasic survival curves at 55°C and 4°C in both media. In contrast, survival curves at 28°C were not biphasic. The growth rates of *L. monocytogenes* grown in stationary phase cultures were also assessed by their generation time. The addition of sodium chloride enhanced heat resistance of microorganism.

It has been proven that biphasic curve and tailing with/without shoulder from thermal inactivation curves in this study were associated with the occurrence of microbial injury. During the exponential phase of *L. monocytogenes*, the percentage injury at 55°C, 28°C and 4°C were ranged between 3.21% to 28.49%, 2.47% to 4.38%, and 4.34% to 8.61%, respectively. Whilst, during the stationary phase of *L. monocytogenes*, the percentage injury at 55°C, 28°C and 4°C were ranged between 2.05% to 4.15%, 1.44% to 3.06%, and 1.07 to 4.25%, respectively.

L. monocytogenes cells were able to survive throughout the sublethal stress of temperatures and undergone morphological changes to adapt to new temperatures. In this study, results from Scanning Electron Microscopy (SEM) revealed three different analyses of temperature-stressed cells which were average mean of cells length, distribution of cells length, and minimum versus maximum cells length. The study demonstrated cells of both log and stationary phase showed a significant variation of morphology. Cells of log phase became elongated only at 55°C, not at 28°C and 4°C whereas cells of stationary phase were shorter and more coccoidal rather than elongated



as in log phase cells. However, cells at 28°C were more intact than cells at other temperatures for most of the observations.

In conjunction with SEM results, the variation of listerial morphology and the effect of listerial motility would be a part of microbial adaptation towards sublethal stress of temperatures. The loss of motility in stationary phase cells strongly suggested that listerial motility play a role in survival of the organism under temperature stress. The stationary phase cells of *L. monocytogenes* were more resistant than exponential phase cells exhibited by increased of generation times, lower percentage injury and most of the cells became coccoid.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KESAN TEKANAN SUHU 'SUBLETHAL' TERHADAP PERTUMBUHAN, KEMANDIRIAN DAN KEUPAYAAN KEKULTURAN LISTERIA MONOCYTOGENES

Oleh

MOHD NIZAM BIN LANI

November 2002

Pengerusi : Abdul Reezal bin Abdul Latif, Ph.D.

Fakulti : Sains Makanan dan Bioteknologi

Kemandirian dan pertumbuhan *Listeria monocytogenes* L56 (sumber organisma daripada IMR) telah dijalankan di dalam trypticase soy broth yang telah dihidupkan pada suhu 37°C sebelum dikenakan tekanan suhu 'sublethal' terhadap tiga suhu yang terpilih (55°C, 28°C and 4°C) daripada fasa eksponensial dan fasa statik sebagai inoculum berbeza menggunakan dua sistem piring agar yang berbeza; TSA dengan dan tanpa 4% NaCl (TSAS). Selain itu, kesan pengaruh morfologi dan pergerakan *Listeria* disebabkan oleh tekanan suhu juga dikaji menggunakan Mikroskopi Pengimejan Elektron dan media pergerakan. Graf pertumbuhan yang seragam pada suhu 37°C telah dilakukan menggunakan pengiraan koloni piring petri, di mana didapati fasa eksponensial dan fasa statik pada suhu 37°C telah diperolehi selepas 12 dan 19 jam dengan pertumbuhan jumlah populasi bakteria dianggarkan sebanyak 10⁸ and 10⁹



(CFU/ml), masing-masing. Daripada graf pertumbuhan bakteria, masa generasi L. monocytogenes pada suhu 37°C ialah 60 minit.

Kadar pertumbuhan bakteria yang diperolehi daripada kekulturan di atas piring petri menggunakan dua sistem media berbeza, iaitu media yang mengandungi dan tanpa mengandungi kepekatan garam sebanyak 4 peratus (TSA and TSAS) dinilai berasaskan masa generasi. Sel *L. monocytogenes* yang dihidupkan dalam fasa eksponential menunjukkan graf yang mempunyai dua keluk pada suhu 55°C dan 4°C dalam keduadua media. Sebaliknya, graf pertumbuhan kesan suhu pada suhu 28°C tidak mempunyai dua keluk. Walaubagaimanapun, kadar pertumbuhan *L. monocytogenes* yang dihidupkan gram telah meningkatkan kerintangan haba mikroorganisma tersebut.

Ia telah dibuktikan bahawa kehadiran dua keluk dan kesan 'tailing' beserta dan tanpa 'shoulder' terhadap rintangan haba dalam kajian ini telah mempunyai kaitan dengan kehadiran mikroorganisma yang tercedera. Semasa pertumbuhan fasa eksponensial *L. monocytogenes*, peratus kecederaan pada suhu 55°C, 28°C dan 4°C di dalam julat 3.21% hingga 28.49%, 2.47% hingga 4.38%, dan 4.34 hingga 8.61%, masing-masing. Manakala, semasa pertumbuhan fasa statik bagi *L. monocytogenes*, peratus kecederaan pada suhu 55°C, 28°C dan 4.15%, 1.44% hingga 3.06%, dan 1.07 hingga 4.25%, masing-masing.

Sel-sel *L. monocytogenes* telah menunjukkan keupayaan merintangi tekanan suhu 'sublethal' dan mengalami perubahan morfologi semasa penyesuaian suhu baru. Dalam



kajian ini, keputusan daripada Mikroskopi Pengimejan Elektron (MPE) telah menunjukkan tiga analisis berbeza terhadap sel-sel tekanan suhu seperti purata kepanjangan sel, taburan kepanjangan sel, serta kepanjangan minimum dan maksimum sel. Kajian ini menunjukkan kedua-dua fasa eksponensial dan fasa statik menunjukkan variasi morfologi yang signifikan. Sel-sel daripada fasa eksponensial telah menjadi panjang hanya pada suhu 55°C, tidak pada suhu 28°C dan 4°C, manakala sel-sel pada fasa statik adalah pendek dan lebih banyak berbentuk 'coccoid' berbanding sel yang panjang pada fasa eksponensial. Walaubagaimanapun, morfologi sel-sel pada suhu 28°C tidak terjejas berbanding dengan suhu-suhu lain pada kebanyakan pemerhatiannya.

Bersama-sama dengan keputusan MPE, variasi terhadap morfologi *Listeria* dan kesan pergerakan *Listeria* merupakan sebahagian daripada penyesuaian mikrobiologi yang dilakukan terhadap tekanan suhu 'sublethal'. Kehilangan keupayaan pergerakan semasa fasa statik dengan tegasnya mencadangkan pergerakan *Listeria* memainkan peranan penting dalam kemandirian organisma terhadap tekanan suhu. Didapati, fasa statik *L. monocytogenes* adalah lebih resistan berbanding sel yang berada pada fasa eksponensial berdasarkan peningkatan nilai masa generasi, peratus kecederaan yang rendah dan kebanyakan sel telah menjadi 'coccoid'.



ACKNOWLEDGEMENTS

The author wishes to thank his Supervisor, Dr. Abdul Reezal Abdul Latif, for his invaluable advice, continuous supervision and guidance, his kindness and his willingness to help throughout the course of this study. The author also wishes to thank his co-supervisors, Dr. Hjh. Zaiton Hassan, for her dynamic help, support and encouragement with golden advises throughout this study, and Associate Professor Dr. Son Radu for his support, advice and encouraging interest in my project.

The author also forwards my special thanks especially to Puan Jamilah Jahari and all lab assistants, En. Zulkifili Nordin, Puan Maimon Zakaria and Puan Kamariah Jaafar in the faculty for their invaluable help. Also, to Institute of Bioscience, UPM and KUSTEM especially Cik Azilah Abd. Jalil, Mr. Ho, Puan Kartini Mohamad and their staffs, to Makmal Biologik, Faculty of Veterninary Medicine, Dr. Abdul Rahman Omar, and Puan Mazlina Othman, Sinarfoto Sdn. Bhd. for their commitment and assistance throughout the project.

Special thanks are also extended to all the members of the Microbiology Laboratory: Siti Zulaiha, Kak Leha, Wai Ling, Kqueen, Apinya, Yousr, Samuel, Zainuri, Haryanti, Noorlis, Zaza, Yuherman, Nasreldin, Ibu Endang, Woo, Yoke Ann, Yin Sze and others. Not forgetting, all my friends, Basri, Khairi, Nazri, Pali, Man Pilot, Nik Rosli, Zam Tuah, Wan Khamis, Ajis, Hamdi, Siti Zulaiha, Salihin, Md Noi, Sazly, Mat Yen, Sofian Hadi, Azlan, Ustaz Ridzuan, Pok Ju, Yanti, Siong, Nisa, Omar Yadi, Aslam, K. Zaid, Fairuz, Pijat and my block and collegemates for helping me in throughout my study.



My sincere thanks is also extended to staffs in Faculty of Food Science and Biotechnology, UPM and to Kolej Universiti Sains Teknologi Malaysia (KUSTEM) for providing the scholarship especially to Professor Dato' Mahyuddin Mohd Dahan, Rector of KUSTEM, Professor Dr. Law Ah Theem as Penyelia Sangkutan SLAB, Associate Professor Dr. Noor Azhar Shazili, Dean of Faculty of Science and Technology, KUSTEM, Puan Wan Sarah Wan Abdullah, Head Department of Food Science, KUSTEM, lecturers in Department of Food Science, KUSTEM such as Associate Professor To' Puan Zakiah Jaafar, Cik Hayati, Cik Norhayati, Dr. Amiza Mat Amin, Associate Professor Dr. Guruprasad Sulebele, and tutors in Department of Food Science, especially to Cik Roshita, Puan Zamazahaila, Cik Norazimah Nazri, En. Fisal, En. Amir, En. Khairi, Cik Norazam and Puan Faridah, not forgetting to Professor Dr. Azmi Ambak and Professor Dr. Loleman Shamsudin, Cik Faridah Mohamad, Cik Hazlina Ahmad Zakeri, Dr. Kamaruzzaman Yunus and Dr. Zaleha Kassim.

Finally, my deepest gratitude and appreciation to both of my parent, Ayahanda Lani bin Hassan and Bonda Kamariah binti Abdullah, my relatives, especially En. Ismail Mohamed and Puan Rohani Hassan for their continuous support since my primary education, and all my brothers and sisters, thank you so much for your support, understanding and encouragement throughout my study.



TABLE OF CONTENTS

Page

| DEDICATION | ii |
|-----------------------|------|
| ABSTRACT | iii |
| ABSTRAK | vi |
| ACKNOWLEDGEMENT | ix |
| APPROVAL SHEETS | xi |
| DECLARATION FORM | xiii |
| LIST OF TABLES | xvii |
| LIST OF FIGURES | xix |
| LIST OF PLATES | xxi |
| LIST OF ABBREVIATIONS | xxii |

CHAPTER

| I | INT | RODUCTION | 1 |
|----|------|--|----|
| II | LIT | ERATURE REVIEW | 6 |
| | 2.1 | Taxonomy and General Characteristics of L. monocytogenes | 6 |
| | 2.2 | Identification of <i>Listeria</i> species | 8 |
| | 2.3 | Isolation and Detection of L. monocytogenes | 9 |
| | 2.4 | Reservoirs of L. monocytogenes | 12 |
| | 2:5 | Occurrence of <i>L. monocytogenes</i> in Food | 13 |
| | 2.6 | Epidemiology of L. monocytogenes | 14 |
| | 2.7 | Disease Associated with L. monocytogenes | 16 |
| | 2.8 | Factors Affecting Growth and Survival of L. monocytogenes | |
| | | in Foods | 17 |
| | | 2.8.1 Temperature | 17 |
| | | 2.8.2 pH. | 18 |
| | | 2.8.3 Acid | 19 |
| | | 2.8.4 Salt | 19 |
| | | 2.8.5 Water Activity | 20 |
| | | 2.8.6 Modified Atmosphere | 20 |
| | 2.9 | Thermal Inactivation of L. monocytogenes | 22 |
| | | 2.9.1 Theory Death by Heating | 22 |
| | | 2.9.2 Heat Resistance of L. monocytogenes in Foods | 28 |
| | | 2.9.2.1 Dairy Products | 28 |
| | | 2.9.2.2 Red Meats and Poultry Products | 30 |
| | | 2.9.2.3 Other Foods | 32 |
| | | 2.9.3 Effect of Sublethal Heating on Thermotolerance | 34 |
| | 2.10 | Sublethal Injury at High and Lower Temperature Stress of | |
| | | L. monocytogenes | 36 |
| | 2.11 | Recovery of Sublethal Injury of L. monocytogenes | 42 |
| | 2.12 | Stress and Effects on Virulence of <i>L. monocytogenes</i> | 46 |



MATERIALS AND METHODS..... Ш 52 3.1 Microorganism..... 52 3.1.1 Source of Culture..... 52 3.1.2 Culture Media 52 3.1.3 Isolate Confirmation of *L. monocytogenes*..... 53 3.2 Development of Standard Growth Curve of *L. monocytogenes*..... 55 3.3 Survival and Culturability of L. monocytogenes Under Sublethal Temperature Stresses..... 55 Determination of Percentage Injury of L. monocytogenes..... 3.4 58 Effects of Morphology and Motility of Temperature-Stressed 3.5 Cells of L. monocytogenes..... 58 3.5.1 SEM's Sample Preparation..... 59 3.5.2 Measurement of Cells Length Using SEMafore..... 59 3.5.3 Effects of Temperature Stress on Motility of L. monocytogenes..... 60 IV RESULTS..... 61 Growth Curve of L. monocytogenes in Trypticase Soy Broth at 4.1 37°C..... 61 Survival and Culturability of L. monocytogenes Subjected to 4.2 Sublethal Temperature Stress Using Media Varying NaCl 4.2.1 Culturability of L. monocytogenes Subjected to Sublethal 4.2.2 Culturability of L. monocytogenes Subjected to Sublethal 4.3 Changes of Morphology and Motility of *L. monocytogenes* Cells 4.4 4.4.1 Mean Cells Length of L. monocytogenes as Affected 4.4.2 Distribution of Cells Length of L. monocytogenes Subjected Distribution of Cells Length (µm) of 4.4.2.1 L. monocytogenes at Log Phase Subjected to Distribution of Cells Length (µm) of 4.4.2.2 L. monocytogenes at Stationary Phase Subjected to Sublethal Temperature Stresses..... 90 4.4.3 Minimum and Maximum Cells Length of L. monocytogenes Subjected to Sublethal Stress of Temperatures..... 95 4.4.4 Effects of Listerial Motility Subjected to Sublethal Stress



| V | DIS | CUSSION | 106 |
|------|------|--|-----|
| | 5.1 | Preliminary Work and Growth Curve of L. monocytogenes | 106 |
| | 5.2 | Relationship between Survival and Culturability of | |
| | | L. monocytogenes | 108 |
| | 5.3 | Relationship between Survival and Sublethal Injury of | |
| | | L. monocytogenes | 116 |
| | 5.4 | Relationship between Survival and Listerial Morphology | |
| | | and Motility | 122 |
| | 5.5 | General Discussion | 127 |
| VI | CO | NCLUSION | 129 |
| REFF | EREN | CES | 132 |
| APPE | | κ | 148 |
| VITA | | | 163 |



LIST OF TABLES

| Table | | Page |
|-------|---|------|
| 2.1 | Differentiation of Listeria from similar genera | 7 |
| 2.2 | Differentiation of L. monocytogenes from other species of Listeria | 10 |
| 2.3 | Media used in in regulatory and other methods for the detection of <i>L. monocytogenes</i> | 12 |
| 2.4 | Limiting conditions for L. monocytogenes growth | 21 |
| 2.5 | Comparison of selected reported growth kinetics for <i>L</i> . <i>monocytogenes</i> versus those predicted by the cubic models | 27 |
| 2.6 | Summary of some findings on the thermal destruction of L. monocytogenes | 29 |
| 2.7 | D and z values for <i>L. monocytogenes</i> heated in thick slurries of chicken or beef | 31 |
| 2.8 | Heat resistance of L. monocytogenes in seafood products | 33 |
| 2.9 | The effect of prior heat shock on the heat resistance of L . monocytogenes in cured meat at 64°C | 35 |
| 2.10 | Similarities in bacterial cell injury by different treatments and its repair. | 39 |
| 2.11 | Types of damage caused to vegetative bacteria by pysical/chemical agents. | 40 |
| 2.12 | Identified virulence determinants in L. monocytogenes. | 50 |
| 4.1 | Estimation of generation time of <i>L. monocytogenes</i> under three selected temperature stresses at log and stationary phases | 67 |
| 4.2 | Percentage injury of <i>L. monocytogenes</i> at log phase under three sublethal temperature stresses using TSA and TSAS | 72 |
| 4.3 | Percentage injury of <i>L. monocytogenes</i> at stationary phase under three sublethal temperature stresses using TSA and TSAS | 73 |
| 4.4 | Comparison of percentage sublethal injury cells of L. <i>monocytogenes</i> at different bacterial phase (log and stationary | 75 |



- 4.5 Comparison of mean cells length (μm) of *L. monocytogenes* on 77 exposure to sublethal temperature stresses at log and stationary phases
- 4.6 Distribution of cells length (μm) of *L. monocytogenes* at log 86 phase on exposure to different sublethal temperature stresses
- 4.7 Distribution of cells length (μm) of *L. monocytogenes* at 92 stationary phase on exposure to different sublethal temperature stresses
- 4.9 Comparison of minimum and maximum length (μm) of L. 94 monocytogenes from log and stationary phases subjected to 55°C, 28°C and 4°C
- 4.10 Motility results of *L. monocytogenes* subjected to sublethal stress 103 of temperatures at log and stationary phases

LIST OF FIGURES

| Figure | | Page |
|--------|---|------|
| 3.1 | The relationship between slope and generation time using semilogarithmic paper | 57 |
| 4.1 | Growth curve of <i>L. monocytogenes</i> using plate counts (CFU/ml) at 37°C | 61 |
| 4.2 | Survival curve of <i>L. monocytogenes</i> (CFU/ml) on exposure to three sublethal temperature stresses at log phase using TSA media | 65 |
| 4.3 | Survival curve of <i>L. monocytogenes</i> (CFU/ml) on exposure to three sublethal temperature stresses at stationary phase using TSA media | 66 |
| 4.4 | Survival curve of <i>L. monocytogenes</i> (CFU/ml) on exposure to three sublethal temperature stresses at log phase using TSAS media | 68 |
| 4.5 | Survival curve of <i>L. monocytogenes</i> (CFU/ml) on exposure to three sublethal temperature stresses at stationary phase using TSAS media | 69 |
| 4.6 | Mean cells lengths of L. monocytogenes subjected to 55° C at log and stationary phases | 83 |
| 4.7 | Mean cells lengths of <i>L. monocytogenes</i> subjected to 28°C at log and stationary phases | 83 |
| 4.8 | Mean cells lengths of <i>L. monocytogenes</i> subjected to $4^{\circ}C$ at log and stationary phases | 84 |
| 4.9 | Distribution of cells length (μ m) of log phase <i>L. monocytogenes</i> subjected to 55°C | 87 |
| 4.10 | Distribution of cells length (μ m) of log phase <i>L. monocytogenes</i> subjected to 28°C | 87 |
| 4.11 | Distribution of cells length (μ m) of log phase <i>L. monocytogenes</i> subjected to 4°C | 88 |
| 4.12 | Distribution of cells length (μ m) of <i>L. monocytogenes</i> subjected to 55°C at stationary phase | 90 |



| 4.13 | Distribution of cells length (μ m) of <i>L. monocytogenes</i> subjected to 28°C at stationary phase | 91 |
|------|---|-----|
| 4.14 | Distribution of cells length (μ m) of <i>L. monocytogenes</i> subjected to 4°C at stationary phase | 91 |
| 4.15 | Minimum and maximum length <i>L. monocytogenes</i> observed using SEM of log phase subjected to 55°C | 96 |
| 4.16 | Minimum and maximum length <i>L. monocytogenes</i> observed using SEM of log phase subjected to 28°C | 96 |
| 4.17 | Minimum and maximum length <i>L. monocytogenes</i> observed using SEM of log phase subjected to 4°C | 97 |
| 4.18 | Minimum and maximum length <i>L. monocytogenes</i> observed using SEM of stationary phase subjected to 55°C | 98 |
| 4.19 | Minimum and maximum length <i>L. monocytogenes</i> observed using SEM of stationary phase subjected to 28°C | 98 |
| 4.20 | Minimum and maximum length <i>L. monocytogenes</i> observed using SEM of stationary phase subjected to 4°C | 99 |
| 4.21 | Comparison of minimum and maximum length L. monocytogenes observed using SEM at log and stationary phases subjected to 55°C | 101 |
| 4.22 | Comparison of minimum and maximum length L. monocytogenes observed using SEM at log and stationary phases subjected to 28°C | 101 |
| 4.23 | Comparison of minimum and maximum length L. | 102 |

4.23 Comparison of minimum and maximum length L. 102 monocytogenes observed using SEM at log and stationary phases subjected to 4°C





LIST OF PLATES

| Plate | | Page |
|-------|---|------|
| 1 | Colonies of L. monocytogenes on PALCAM agar | 54 |
| 2 | SEM's picture shows log phase cultures (12 h) of L. monocytogenes at 37°C | 63 |
| 3 | SEM's picture shows stationary phase cultures (19 h) of L. monocytogenes at $37^{\circ}C$ | 64 |
| 4 | SEM's picture demonstrated a 'fussy phenomenon' observed at temperature stress of 55°C after 4 h at log phase | 79 |
| 5 | SEM's picture demonstrated a 'fussy phenomenon' observed at temperature stress of 4°C after 4 h at log phase | 79 |
| 6 | SEM's picture demonstrated intact cells observed at temperature stress of 28°C after 4 h at log phase | 80 |
| 7 | Cells of <i>L. monocytogenes</i> were observed 'clumping' from SEM during stationary phase after 4 h at 4°C | 80 |
| 8 | Elongated cells of <i>L. monocytogenes</i> observed at temperature stress of 55°C after 4 h during log phase using fluorescent microscopy | 87 |
| 9 | Coccoid cells of <i>L. monocytogenes</i> observed at temperature stress of 55°C after 4 h during stationary phase using fluorescent microscopy | 87 |
| 10 | The right bottle indicates the positive motility result of L . monocytogenes observed after incubation at 25°C for 3 days where the left bottle acts as control | 104 |
| 11 | The same bottles of motility tests at a larger magnification | 104 |
| 12 | Positive results of motility test for log phase cultures of <i>L</i> . <i>monocytogenes</i> representing every hour of six hours | 105 |
| 13 | Negative results of motility test for stationary phase cultures of <i>L. monocytogenes</i> representing every hour of six hours | 105 |



LIST OF ABBREVIATIONS

| CAMP Test | (Christie, Atkins, and Munch-Peterson) Test |
|-----------|--|
| CDC | Centres for Disease Control and Prevention |
| CFU | Colony forming unit |
| D value | Decimal Reduction Time |
| FDA | Food and Drug Administration |
| NaCl | Sodium Chloride |
| RCPA | Royal College of Pathologists of Australasia |
| SEM | Scanning Electron Microscopy |
| TSA | Trypticase Soy Agar |
| TSAS | Trypticase Soy Agar with added 4% Sodium Chloride |
| TSB | Trypticase Soy Broth |
| z value | Reciprocal Slope Value of Thermal Death Time Curve |

CHAPTER I

INTRODUCTION

Listeria monocytogenes was presumably first isolated at the beginning of the twentieth century as a Gram positive rod in tissue specimens of infected patients (Fsihi *et al.*, 2001). In 1919, Hulphers called *Bacillus hepatis* the bacterium he isolated from necrotic foci in rabbit liver. In 1926, *L. monocytogenes* was described by Murray and colleagues as *Bacterium monocytogenes* in connection with an epizootic among laboratory-raised guinea pigs and rabbits (Murray *et al.*, 1926). However, the following year, Pirie described a similar epizootic in wild gerbils termed "Tiger River Disease" and proposed that the species to be called *Listerella hepatolytica*. After finding that *Bacterium monocytogenes*. The current name *Listeria* was suggested in 1940 by Pirie in honour of Lord Lister, an eminent English surgeon and discoverer of antiseptics (Seeliger and Jones, 1986).

L. monocytogenes is appreciably more heat resistant than other non-sporing microorganisms such as Salmonella and Campylobacter, and can grow at refrigeration temperatures (Farber and Peterkin, 1991). The bacterium is ubiquitous in nature and can be isolated from a wide variety of sources, both the environment and food. It also appears to be a common environmental contaminant of food processing plants. The foods that have received most attention have been milk and dairy products, meat, poultry, seafood and vegetables as L. monocytogenes is capable of survival and growth

