

UNIVERSITI PUTRA MALAYSIA

ANTI-TUMOUR PROMOTING ACTIVITY OF SELECTED MALAYSIAN VEGETABLES AND FRUITS, AND IDENTIFICATION OF ANTI-TUMOUR PROMOTING AND ANTIOXIDANT COMPOUNDS FROM COLEUS TUBEROSUS, BENTH (UBI KEMILI)

LIM YANG MOOI

FSMB 2002 26

ANTI-TUMOUR PROMOTING A CTIVITY OF SELECTED MALAYSIAN VEGETABLES AND FRUITS, AND IDENTIFICATION OF ANTI-TUMOUR PROMOTING AND ANTIOXIDANT COMPOUNDS FROM COLEUS TUBEROSUS, BENTH (UBI KEMILI)

LIM YANG MOOI

DOCTOR OF PHILOSOPHY UNIVERSITI PUTRA MALAYSIA

2002

ANTI-TUMOUR PROMOTING ACTIVITY OF SELECTED MALAYSIAN VEGETABLES AND FRUITS, AND IDENTIFICATION OF ANTI-TUMOUR PROMOTING AND ANTIOXIDANT COMPOUNDS FROM COLEUS TUBEROSUS, BENTH (UBI KEMILI)

By LIM YANG MOOI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Doctor of Philosophy

Specially dedicated to my beloved -

Grandmother,

Father, Mother,

Brothers, Sisters,

Husband,

Parents-in-law,

And friends

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Doctor of Philosophy

ANTI-TUMOUR PROMOTING A CTIVITY OF SELECTED MALAYSIAN VEGETABLES AND FRUITS, AND IDENTIFICATION OF ANTI-TUMOUR PROMOTING AND ANTIOXIDANT COMPOUNDS FROM COLEUS TUBEROSUS, BENTH (UBI KEMILI)

By

LIM YANG MOOI

May 2002

Chairman: Professor Dr. Abdul Manaf Ali,

Faculty: Faculty of Food Science and Biotechnology

A convenient short-term in vitro assay, the inhibition of Epstein-Barr virus (EBV) activation induced by phorbol 12-myristate 13-acetate (PMA) and sodium n-butyrate was conducted to detect the naturally occurring anti-tumour promoters of 133 vegetables and fruits. Forty-two plants showed strong inhibitory activity, 33 plants showed moderate inhibitory activity, 21 plants were found to be weakly active and 37 plants were inactive. Coleus tuberosus, Benth was chosen for further study because it showed the strongest activity. Phytosterols (CT 1) and 2α , 3β -dihydroxyl-12-oleanen-28-oic-acid (CT 2) were isolated from Coleus tuberosus, Benth based on bioassay-guided fractionation. CT 1 was identified as a mixture consisting of stigmasterol (32.0%), campesterol (27.7%) and β -sitosterol (40.3%) by gas chromatography. CT 2 was established as 2α , 3β -dihydroxy-12-oleanen-28-oic acid. Five plants that showed the highest anti-tumour promoting activity namely, Carica papaya, Barringtonia macrostachya, Coleus tuberosus, Mangifera indica and Eugenia polyantha also exhibited strong antioxidant activity compared to α -tocopherol in the ferric thiocyanate (FTC) method and showed more than 60%

inhibition rate in the xanthine/xanthine oxidase system. Those plants mentioned above did not exhibit any activity in scavenging stable DPPH (1,1-diphenyl-2picrylhydrazyl) radicals and hydrogen peroxide stimulated in differentiated HL 60 cells by PMA. Isolated compounds CT 1 and CT 2 and commercial standards campesterol, stigmasterol, and β -sitosterol were also tested for their antioxidant activity. Campesterol, CT 1, stigmasterol, and β -sitosterol demonstrated more than 50% inhibition rate in scavenging superoxide anion induced in the xanthine/xanthine oxidase system. CT 2 showed an inhibition rate of 46.62%. Campesterol, CT 1 and β -sitosterol showed more than 50% inhibition rate at 2 μ g/ml in scavenging hydrogen peroxide induced in differentiated HL 60 cells, but CT 2 and stigmasterol, attained an inhibition rate of only 32.97% and 16.37%, respectively. All compounds did not exhibit any activity in scavenging stable DPPH radicals. Campesterol, CT 1, CT 2, stigmasterol and β -sitosterol were found to have very strong anti-tumour promoting activity and their IC₅₀ were determined as 2 µg/ml, 0.7 µg/ml, 0.1 µg/ml, 0.6 µg/ml and 1 µg/ml, respectively. The optimum combination effect of stigmasterol, β -sitosterol and campesterol towards the anti-tumour promoting activity was obtained at 40 µg/ml of each compound analysed by the response surface methodology (RSM). Campesterol was assessed to cause a negative interaction to β sitosterol, while, stigmasterol caused a negative interaction to campesterol at concentrations higher than 20 µg/ml. Immunoblotting analysis was used as a confirmation test for the detection of the EBV early antigen EA-D and EA-R. EA-D was detected as a darker band of about 50-52 kDa while EA-R showed up as a fade band of about 85 kDa. In conclusion, CT 1 and CT 2 were found to show strong anti-tumour promoting activity. This supports the assumption that the consumption

of vegetables and fruits is a highly recommended strategy for cancer chemoprevention and can be a practical approach to control cancer.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia untuk memenuhi keperluan untuk ijazah Doktor Falsafah

AKTIVITI ANTI-PENGGALAK TUMOR BAGI SAYUR-SAYURAN DAN BUAH-BUAHAN MALAYSIA TERPILIH DAN PENGENALPASTIAN SEBATIAN ANTI-PENGGALAK TUMOR DAN ANTIOXIDANT DARIPADA COLEUS TUBEROSUS, BENTH (UBI KEMILI)

Oleh

LIM YANG MOOI

Mei 2002

Pengerusi: Profesor Dr. Abdul Manaf Ali,

Fakulti: Fakulti Sains Makanan dan Bioteknologi

Suatu bio-cerakin in vitro jangka pendek telah dijalankan iaitu perencatan pengaktifan virus Epstein-Barr (EBV) yang diaruh oleh forbol 12-miristat 13-asetat (PMA, 5 μ M) dan 3 mM natrium n-butirat untuk mengesan anti-penggalak tumor semulajadi dalam 133 sayur-sayuran and buah-buahan. Empat puluh dua tumbuhan telah menunjukkan aktiviti perencatan yang tinggi, 33 tumbuhan menunjukkan aktiviti perencatan sederhana, 21 menunjukkan aktiviti perencatan lemah dan 37 tumbuhan pula tidak aktif. Coleus tuberosus, Benth telah dipilih untuk kajian selanjutnya kerana menunjukkan aktiviti perencatan yang tertinggi. Dua sebatian iaitu satu campuran fitosterol (CT 1) dan asid 2α , 3β -dihidroksida-12-oleanen 28-oik (CT 2) telah diperolehi daripada pokok Coleus tuberosus, Benth dengan mengaplikasikan teknik pengasingan berpandukan biocerakinan. CT 1 telah dikenalpasti sebagai campuran sebatian yang terdiri daripada stigmasterol (32.0%), kampesterol (27.7%) dan β -sitosterol (40.3%) dengan kaedah kromatografi gas. CT 2 pula telah dikenalpasti sebagai asid 2α , 3β -dihidroksida-12-oleanen 28-oik. Lima tumbuhan terdiri daripada Carica papaya, Barringtonia macrostachya, Coleus

tuberosus, Mangifera indica dan Eugenia polyantha yang menunjukkan perencatan pengaktifan Epstein-Barr virus yang tinggi juga menunjukkan aktiviti antioxidan yang tinggi berbanding dengan α -tokoferol melalui kaedah Ferik tiosianat (FTC), dan menunjukkan lebih daripada 60 % kadar perencatan dalam sistem xanthina / xanthina oksidase. Lima tumbuhan tersebut tidak memberi sabarang perencatan terhadap aktiviti menghilangkan radikal 1,1-difenil-2-pikrilhidrazil (DPPH) yang stabil dan hidrogen peroksida yang dihasilkan dalam sel-sel HL 60 yang telah distimulasikan untuk melalui proses perbezaan oleh PMA. Ujian aktiviti antioxidant pada sebatian CT 1 and CT2 dan komersial piawai iaitu stigmasterol, kampesterol dan β -sitosterol juga dijalankan. CT 1, stigmasterol, kampesterol dan β -sitosterol telah terbukti menunjukkan kadar perencatan melebihi 50% dalam sistem xanthina / xanthina oksidase. CT 2 menunjukkan kadar perencatan pada 46.62%. CT 1, kampesterol dan β -sitosterol merencat penghasilan hidrogen peroksida dalam sel-sel HL 60 yang telah melalui pembezaan pada kadar perencatan melebihi 50% pada kepekatan 2 μg/ml. CT 2 dan stigmasterol masing masing menunjukkan 32.97% dan 16.37% kadar perencatan pada kepekatan 2 µg/ml. Sebatian ini juga tidak menunjukkan sebarang aktiviti menghilangkan radikal DPPH yang stabil. CT 1, CT 2, stigmasterol, kampesterol dan β -sitosterol telah menunjukkan aktiviti antipenggalak tumor yang tinggi dan memberi IC₅₀ pada 0.7 μg/ml (CT 1), 0.6 μg/ml (stigmasterol), 1 μg/ml (β-sitosterol), 2 μg/ml (kampesterol) dan 0.1 μg/ml (CT 2). Kesan kombinasi optimum stigmasterol, kampesterol dan β -sitosterol bagi aktiviti promoter tumor telah dicapai pada 40 µg/ml bagi setiap sebatian yang dianalisiskan melalui kaedah gerakbalas permukaan (RSM). Kampesterol telah ditaksir untuk menyebabkan interaksi negatif kepada β -sitosterol, sementara, stigmasterol pula menyebabkan interaksi negatif kepada kampesterol pada konsentrasi yang lebih

tinggi daripada 20 µg/ml. Analisis imunokedap dengan menjalankan Western blotting dapat digunakan sebagai ujian pemastian bagi pengesanan antigen EA-D dan EA-R. Antigen EA-D dan EA-R telah dikesan melalui kaedah imunokedap sebagai jalur hitam pada kira-kira 50-52 kDa dan jalur kabur pada kira-kira 85 kDa. Pada kesimpulannya, CT 1 and CT 2 menunjukkan aktiviti promoter anti-tumor yang tinggi. Ini menyokong anggapan iaitu memakan sayur-sayuran and buah-buahan adalah satu strategi yang amat disyorkan bagi rawatan kanser secara kimo-pencegahan dan merupakan pendekatan yang praktikal untuk mengawal kanser.

ACKNOWLEGEMENTS

I wish to express my sincere appreciation and deepest gratitude to my supervisor, Professor Dr. Abdul Manaf Ali, and co-supervisors, Professor Dr. Md. Nordin Haji Lajis, Associate Professor Dr. Norhanom Abdul Wahab, and Associate Professor Dr. Raha Abdul Karim for their invaluable guidance, advice, constructive comments and encouragement during the execution of my project and preparation of this thesis.

Special thanks to Professor Dr. K. Koshimizu and Associate Professor Dr. A. Murakami for arranging an opportunity for me to spend three weeks in their laboratory at Kinki University, Japan. My thanks also goes to Miss Megumi Kadota, Mr. Toyota, Mr. Takao Inoue and Miss Yamamoto for their help during my stay in Japan.

My utmost appreciation to Associate Professor Dr. Khozirah Shaari from Laboratory of Phytomedicine, Institute Bioscience, UPM for her help, ideas and guidance. My deep appreciation is also extended to Dr. Jasbir S. Dhaliwal and Mr. Quek in Immunology Division, IMR Kuala Lumpur, Mr. Ashril in Sports Centre, Miss Uma in Molecular Biology laboratory, University of Malaya, Mr. Dharma Permata and Puan Habsah in Chemistry Laboratory, Universiti Putra Malaysia for their help and advice. Special thanks goes to my friends Dr. Majid, Anthony Ho, Law Sen Yu, Yu Li Ling, Ernie, Kok Yih Yih, Madiha, Tan Boon Keat and Khor Tin Oo for their help and support.

Lastly, I would like to express my utmost gratitude, indebtedness and appreciation to my grandmother, parents, brothers, and sisters, my beloved husband and parents-in-law for their love, support and encouragement that inspired me to accomplish this study.

TABLE OF CONTENTS

		Page
ABS ABS ACK APP DEC	DICATION OTRACT OTRAK CNOWLEDGEMENTS ROVAL CLARATION	2 3 6 9 10 12
LIST LIST	OF TABLES OF FIGURES OF PLATES OF ABBREVIATIONS	16 18 21 22
CH.	APTER	
I	INTRODUCTION	24
П	LITERATURE REVIEW Multistage of Carcinogenesis	29 29
	Initiation Promotion Progression	31 32 33
	Chemoprevention as an Alternative Approach of Cancer Prevention Mechanisms of Chemoprevention Category of Chemopreventive Agents	34 37 40
	Preclinical Models to Assess the Efficacy of Chemopreventive Agents	45
	Human Clinical Trials of Chemopreventive Agents The Focus of Chemoprevention in the New Millennium	49 53
	Bioassays Natural Product	54 58
	Vegetables and Fruits in Relation to Cancer Effects of Food Phytochemicals on Xenobiotic Metabolism and Tumorgenesis	61 64
	Anti-Tumour Promoters from Natural Resources Reactive Oxygen Species and Antioxidants	69 7 2
	Free Radicals Sites of Activated Oxygen Production Antioxidants	72 80 82
Ш	MATERIALS AND METHODS	86
	Cell Culture	86
	Medium Preparation	86
	Cryopreservation of Cells	87
	Maintenance of Cell Culture	87
	Reviving of Raji Cell	88 89
	Inhibitory Assay of Epstein-Barr Virus Activation	67

	Cell Line	89
	Antioxidant Activity Assays	92
	Chemicals	92
	Antioxidant Assay (Ferric thiocynate method)	92
	DPPH Free Radical Scavenging Activity Assay	92
	Xanthine / Xanthine Oxidase Inhibition Activity Assay	94
	Inhibitory Effect on PMA-Induced Hydroperoxide	
	Formation in Differentiated HL 60 Cells	94
	Combination Study of Stigmasterol, β -Sitosterol and	
	Campesterol	95
	Detection of EBV-Early Antigen on PVDF Membrane	96
	Stock Reagent Preparation	96
	Gel Preparation	97
	Sample Preparation for Lysis Treated Raji Cells	98
	Lysis of Cells	99
	SDS-Polyacrylamide Gel Electrophoresis (PAGE)	
	(Laemmli, 1970)	100
	Western Blotting	101
	Preparation of Proteins Transfer onto PVDF Membrane	102
	Blocking Treatment	103
	Binding of Primary Antibody (NPC serum) onto	
	Targeted Protein	104
	Binding of Enzyme-Coupled Secondary Antibody (HP)	104
	Colour Development Solution	105
	Colour Development	106
	Preparation of Marker Staining Solution	106
	Staining of Marker on the PVDF Membrane	106
	Statistical Analysis	107
	Extraction and Isolation of the Active Compounds from	
	Coleus tuberosus, Benth	107
	General Instrumentation	107
	Plant Materials	108
	Extraction and Fractionation of the Extract of Coleus	
	tuberosus, Benth	108
	Isolation of CT 1 (phytosterol) and CT 2 (2α,3β-dihydroxy-12-	
	oleanen-28-oic acid) from the Chloroform Fraction	112
	Gas Chromatography	113
VI	RESULTS	115
	Screening for In Vitro Anti-Tumour Promoting Activity of	
	Malaysian Vegetables and Fruits	115
	Minimum Inhibition Concentration (MIC)	
	Determination of Selected Plants	123
	Bioassay-guided Isolation of Active Anti-Tumour Promoting	
	compounds from Coleus tuberosus, Benth	130
	Bioactivity Determination of the Crude Fractions of	
	Coleus tuberosus, Benth	130
	Identification of CT 1 mixture	133
	Structural Elucidation of CT 2	151

	Antioxidant Activity of Five Selected Plants Crude Extracts	
	and Compounds Isolated from Coleus tuberosus, Benth	167
	Estimation of Peroxide by Ferric Thiocynate Method	167
	Diphenyl picryl hydrazyl (DPPH) radical	
	Scavenging Activity Assay	170
	Inhibition of Xanthine / Xanthine Oxidase Activity Assay	175
	Flow Cytometric Detection of Hydrogen Peroxide Production	
	in Human Promyelocytic Leukaemia Cells (HL 60)	180
	Inhibition of EBV-Early Antigen Expression in Raji Cell:	
	Determination of Activity in Isolated Compounds and Its Combination.	189
	Anti-Tumour Promoting Activity of Isolated Compounds,	
	CT 1 and CT 2	189
	Combination Effect of Campesterol, β -Sitosterol	
	and Stigmasterol	192
	Detection of Anti-Tumour Promoting Activity by	
	Immunoblotting Analysis	202
V	DISCUSSIONS	212
	Screening of Anti-Tumour Promoting Activity of Malaysia	
	Vegetables and Fruits	212
	Antioxidant Activity of Five Selected Plant Crude Extracts	
	and Compounds Isolated from Coleus tuberosus, Benth	215
	Inhibition of EBV-Early Antigen Expression in Raji Cell:	
	Determination of Activity in Isolated Compounds	
	and Its Combination.	225
	Detection of Anti-Tumour Promoting Activity by	
	Immunoblotting Analysis	231
VI	CONCLUSION	233
PERE	RENCES	238
		230
APPE:	NDICES	253
VITA		256

LIST OF TABLES

Table		Page
2.1	Mechanism of actions of chemoprevention: possible molecular targets and promising agents	38
2.2	Inhibitory effects of some phytochemicals in fruits and	
	vegetables on chemically induced carcinogenesis in animal models	63
2.3	List of anti-tumour promoters isolated from natural resources	71
2.4	Active oxygen and related species	74
2.5	Production of active oxygen species	75
2.6	Defence systems in vitro against oxidative damage	83
3.1	Titre determination of NPC sera	91
3.2	Inhibition rate (%) of combined fractions of the chloroform fraction.	111
4.1	In vitro anti-tumour promoting activities of vegetables and fruits	116
4.2	Anti-tumour promoting properties of vegetables and fruits	126
4.3	The composition of stigmasterol, campesterol and β -sitosterol in the mixture if CT 1	134
4.4	Proton NMR chemical shifts of CT 2, augustic acid and eucalyptolic acid	157
4.5	Carbon chemical shifts of CT 2, augustic acid, maslinic acid and eucalyptolic acid	158
4.6	Proton and carbon chemical shifts for CT 2 including 1 <i>J</i> , 2 <i>J</i> , 3 <i>J</i> and 4 <i>J</i> correlations as deduced from HMBC spectrum	159
4.7	¹ HNMR signals of methyl 2,3-dihydroxy-urs-12-en-28-oates and their diacetates	165
4.8	Superoxide suppression in xanthine / xanthine oxidase system by CT 1, CT 2, stigmasterol, β -sitosterol, campesterol and plant crude extracts	176

4.9	Fluorescence intensity of intracellular DCFH oxidation in differentiated HL 60 cells stimulated by PMA	182
4.10	Inhibitory effect of kaempferol towards intracellular DCFH oxidation in differentiated HL 60 cells stimulated by PMA	184
4.11	Inhibitory effect of CT 1 and CT 2 toward intracellular DCFH oxidation in differentiated HL 60 cells stimulated by PMA	187
4.12	Inhibitory effect of β -sitosterol, stigmasterol and campesterol toward intracellular DCFH oxidation in differentiated HL 60 cells stimulated by PMA	187
4.13	Fluorescence intensity of plant crude extracts toward intracellula DCFH oxidation in differentiated HL 60 cells stimulated by PMA	ar 190
4.14	The 50% (IC $_{50}$) inhibition of Epstein-Barr virus activation by CT 1, CT2, β -sitosterol, stigmasterol, campesterol and standard compounds	190
4.15	Combination concentrations of stigmasterol, β -sitosterol and campesterol toward Epstein-Barr virus activation in Raji cells	193
4.16	Inhibition of Epstein-Barr virus activation by the combination compounds of stigmasterol, β -sitosterol and campesterol	193
4.17	Combination concentrations of β -sitosterol and campesterol toward Epstein-Barr virus activation in Raji cells	198
4.18	Combination concentrations of stigmasterol and campesterol toward Epstein-Barr virus activation in Raji cells	198
4.19	Combination concentrations of stigmasterol and β -sitosterol toward Epstein-Barr virus activation in Raji cells	199
4.20	Inhibition of Epstein-Barr virus activation by the combination compounds of β -sitosterol and campesterol	199
4.21	Inhibition of Epstein-Barr virus activation by the combination compounds of stigmasterol and campesterol	200
4.22	Inhibition of Epstein-Barr virus activation by the combination compounds of stigmasterol and β-sitosterol	200

LIST OF FIGURES

Figure		Page
2.1	Multistage carcinogenesis	34
2.2	Carcinogen-blocking activities	37
3.1	Solvent partitioning of crude extract of Coleus tuberosus Benth	110
3.2	Isolation scheme of the active anti-tumour promoting compounds	114
4.1	Proportions of the inhibitory activities toward EBV activation of the extracts from 133 fresh vegetables and fruits	121
4.2 (a-e)	Inhibition of EBV activation by different fractions of crude extract of <i>Coleus tuberosus</i> , Benth. a: petroleum ether b: chloroform fraction, c: ethyl acetate fraction, d: butanol fraction and e: water fraction	132
4.3	Infrared spectrum of CT 1	135
4.4 a & b	Proton NMR spectrum of CT 1	136, 137
4.5 a &b	Carbon-13 NMR spectrum of CT 1	138, 139
4.6 a & b	Proton NMR of mixture of β -sitosterol and stigmasterol	140, 141
4.7 a & b	Carbon-13 NMR of mixture of β -sitosterol and stigmasterol	142, 143
4.8a	Gas chromatography analysis of CT 1, RT (retention time) $12.573 = \text{campesterol}$, RT $12.925 = \text{stigmasterol}$ and RT $13.655 = \beta$ -sitosterol	144
4.8b	Gas chromatography analysis of campesterol	145
4.8c	Gas chromatography analysis of stigmasterol	146
4.8d	Gas chromatography analysis of β -sitosterol	147
4.8e	Gas chromatography analysis of the mixture of commercial standards including campesterol, stigmasterol and β -sitostero	ol 148
4.8f	Gas chromatography analysis of the mixture of CT 1 and commercial standards	149

4.9	Molecular structure of campesterol, β -sitosterol and stigmasterol	150
4.10	Infrared spectrum of 2α , 3β -dihydroxy-12-oleanen-28-oic acid (CT 2)	d 154
4.11	Mass spectrum of 2α , 3β -dihydroxy-12-oleanen-28-oic acid	155
4.12	Proton NMR spectrum of 2α , 3β -dihydroxy-12-oleanen-28-oic acid	156
4.13	HMBC spectrum of 2α , 3β -dihydroxy-12-oleanen-28-oic acid	160
4.14	HMQC spectrum of 2α , 3β -dihydroxy-12-oleanen-28-oic acid	161
4.15	Carbon-13 NMR of 2α , 3β -dihydroxy-12-oleanen-28-oic acid	162
4.16	$^{1}\text{H-}^{1}\text{H COSY spectrum of}$ $2\alpha, 3\beta$ -dihydroxy-12-oleanen-28-oic acid	163
4.17	DEPT 135 spectrum of 2α , 3β -dihydroxy-12-oleanen-28-oic acid	164
4.18	Molecular structure for CT 2 with ¹³ C assignment	166
4.19	Antioxidant activity of five crude plant extracts measured by FTC method	169
4.20a, b, c	DPPH radical scavenging activity of plant crude extracts, commercial standards and isolated compounds (CT 1 and CT 2)	172-174
4.21a, b, c	The inhibition of superoxide generation through enzymatic xanthine / xanthine oxidase system by plant crude extracts, commercial standards and isolated compounds (CT 1 and CT 2)	177-179
4.22	Flow cytometric fluorescence distribution of (C) DCFH-DA non-loaded cells, (B) DCFH-DA loaded cells and (A) DCFH-DA loaded cells treated with 100 nM PMA	181
4.23	Flow cytometric fluorescence distribution of DCFH-DA load cells treated with 100 nM PMA and kaempferol at the concentrations of 20 µg/ml (K1), 2 µg/ml (K2), 0.2 µg/ml (K and 0.02 µg/ml (K4). C: DCFH-DA non-loaded cells, B: DCFH-DA loaded cells and A: DCFH-DA loaded cells treated with 100 nM PMA	

4.24	Flow cytometric fluorescence distributions of DCFH-DA loaded cells treated with 100nM PMA and CT 1 at the concentrations of 20µg/ml (CT 1-1) and 2µg/ml (CT 1-2). C; DCFH-DA non-locells, B: DCFH-DA loaded cells and A: DCFH-DA loaded	
	cells treated with 100nM PMA	185
4.25	Flow cytometric fluorescence distributions of DCFH-DA loaded Treated with 100nM PMA and β-sitosterol at the concentrations of 40μg/ml (B1), 20μg/ml (B2) and 2μg/ml (3). C: DCFH-DA non-loaded cells, B: DCFH-DA loaded cells and A: DCFH-DA loaded cells treated with 100nM PMA	
4.26	Inhibition of Epstein-Barr virus activation by genistein, quercetic CT 1, CT 2, campesterol, β -sitosterol and stigmasterol	in, 191
4.27 (a-c)	Three-dimensional plots demonstrating the combination effect of anti-tumour promoting activity of campesterol, β -sitosterol and stigmasterol	194
4.28 (a-c)	Three-dimensional plots demonstrating the combination effect of anti-tumour promoting activity of campesterol, β -sitosterol and stigmasterol	195
4.29 (a-c)	Three-dimensional plots demonstrating the combination effect of anti-tumour promoting activity of campesterol, β -sitosterol and stigmasterol	196
4.30 (a-c)	Three-dimensional plots demonstrating the combination effect of anti-tumour promoting activity of campesterol, β -sitosterol and stigmasterol	201
4.31(a-i)	Effects of edible plant extracts and isolated compounds on the synthesis o EBV-EA in Raji cells treated with phorbol 12-myristate 13-acetate (PMA, 0.05µM) and sodium <i>n</i> -butyrate (3mM) by immunoblotting analysis	211
5.1	The central role of oxygen free radicals and their affects on multistage carcinogenesis	218
5.2	Reaction of antioxidants with stable 1,1-diphenyl-2-picrylhdraz free radical (DPPH) to 1,1-diphenylhydrazine	yl 220

LIST OF PLATES

Plate		Page
4.1	Imunoflourescence elicited in Raji cells	122
4.2	Pictures a and b show the plant and tuber of Coleus tuberosus, Benth	131

LIST OF ABBREVIATIONS

ACF : Aberrant crypt foci °C : Degree Celsius

CGM : Complete growth medium 4CN : 4-Chloro-1-Naphthol CO₂ : Carbon dioxide

COSY : Correlated spectroscopy
DCF : Dichlorofluorescein
DCFH : Dichlorofluorescin

DCFH-DA: 2',7'-dichlorofluorescin diacetate

DEPT : Distortionless enhancement by polarisation transfer

DMFO: 2-difluoromethylornithine

DMSO : Dimethyl sulfoxide

DPPH: 1,1-diphenyl-2-picrylhydrazyl

DTT : Dithiothreitol EA : Early antigen

EMIS: Electron impact mass spectrometry

FCS: Foetal calf serum
FITC: Flouresceinthiocynate

FT-IR : Fourier transform mass spectroscopy

GST : Gluthathione-S-transferase

HCl : Hydrochloric acid

HL-60 : Human promyelocytic cell

HMBC: Heteronuclear multiple bond correlation
HMQC: Heteronulear multiple quantum correlation

Hp : Horseradish Peroxidase

IR : Infrared

J : Coupling constant m/z : Mass to charge ratio

M : Molar

MDA : Malon-dialdehyde

mg : Milligram
MS : Mass spectrum
ml : Millilitre
min : Minutes
mA : Milliampere

NADPH: Reduced nicotinamide adenine dinucleotide phosphate

NBT : Nitroblue tetrazolium

nm : Nanometer

NMR : Nuclear magnetic resonance NPC : Nasopharyngeal carcinoma

NSAID : Nonsteroidal anti-inflammatory drug

no : Number

ODC : Ornithine decarboxylase PBS : Phosphate buffered saline

PKC: Protein Kinase C

PMA : Phorbol 12-myristate 13-acetate PMSF : Phenylmethylsulfonyl fluoride

QR : Quinone reductase

ROS: Reactive oxygen species

RSM: Response Surface Methodology
RPMI: Rosewell Park Memorial Institute

rpm : Rotation per minute
SDS : Sodium dodecyl sulphate
SOD : Superoxide dismutase

TEMED: N,N,N',N'-tetramethylenediamine
Tris: Tris (hydrosymethl) aminoethane

TMS: Tetramethylsilane

TPA: 12-O-tetradecanol-phorbol-13-acetate

UV : Ultraviolet

XOD : Xanthine Oxidase

 $\begin{array}{cccc} \mu & & : & & \text{Micro} \\ \text{\%} & & : & & \text{Percentage} \\ \delta & & : & & \text{Chemical shift} \end{array}$

 λ_{max} : In UV spectroscopy, the wavelength at which maximum

absorption occurs