

A PLANT PATHOGENIC FUNGUS ELICITING NECROSIS (LEAF SPOT) AGAINST Exbucklandia populnea (R.Br. ex Griff.) R.W.Br. (GEROK) AT FOREST NURSERY TERLA B

TATSURO KIKUCHI

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Ву

TATSURO KIKUCHI



A Project Report Submitted in Partial Fulfilment of the Requirements for the Degree of Bachelor of Forestry Science in the Faculty of Forestry Universiti Putra Malaysia

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DEDICATION

Specially dedicated to my:

Parents

Dr. Razak Bin Terhem

Assoc. Prof. Dr. Mohd Nazre Saleh

Prof. Dr. Arata Momohara

Thank you for your encouragement and support And all the sacrifices that you have given to me Thank you for everything

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ABSTRACT

Exbucklandia populnea is an important species for a restoration programme in Cameron Highlands as it is a common pioneer tree in Cameron Highlands. However, necrotic disease was observed and killed a lot of seedlings of E. populnea at Forest Nursery Terla B, which is the only nursery for the restoration in Cameron Highlands. Further spread of the disease is considered to be able to decrease the supply of seedlings of *E. populnea* because the germination rate of the seeds is low, and the seedlings grow slowly. Therefore, the purpose of this study is to determine the causal agent of the disease and estimate the impact in the nursery. From a survey at the nursery, nearly 30 % of E. populnea seedlings showed necrotic disease, which made it the most dominant disease in the nursery, and the necrotic symptom was categorized into two sub-symptoms: TS (small brown spots) on young leaves and TB (big brown spots) on both old and young leaves. The symptomatic leaves of E. populnea were collected at the nursery, and the necrotic tissues were cut into small pieces. The pieces and the spores on the spots were placed onto potato dextrose agar media and incubated at room temperature (27-30 °C). From both of TS and TB symptomatic leaves a grey colonial fungus named T1 was isolated. T1 had identical spores and hyphae that were confirmed on both TS and TB symptomatic leaves. Thus, T1 was suspected as the causal agent of both of the types of leaf spot diseases. All strains of T1 were identified as Botrytis cinerea by DNA sequencing of ITS region. The fungus is notorious for causing necrotic diseases on plants in nurseries. Mycelial suspension and mycelial plugs were used as inoculums for the pathogenic tests. During the tests. TB symptom was confirmed on the inoculated leaves, but TS symptom did not occur. Thereby, it was not fully confirmed in this study that TS symptom was caused by *B. cinerea*, although this symptom was highly likely caused by *B. cinerea*. TS symptom was considered to be not simply caused by surface inoculation and might involve physiological change of *B. cinerea* from endophytic to necrotrophic life style as previous research shows. Further research about the relationship between E. populnea and B. cinerea would help to find specific measures to control the necrotic diseases.

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APPROVAL SHEET

I certify that this research project report entitled "A Plant Pathogenic Fungi Eliciting Necrosis against *Exbucklandia Populnea* (R.Br. Ex Griff.) R.W.Br. (Gerok) at Forest Nursery Terla B" by Tatsuro Kikuchi has been examined and approved as a partial fulfilment of the requirements for the Degree of Bachelor of Forestry Science in the Faculty of Forestry, Universiti Putra Malaysia.



Dr. Razak Bin Terhem Faculty of Forestry Universiti Putra Malaysia (Supervisor)

Prof. Dr. Mohamed Zakaria Bin Hussin Dean Faculty of Forestry Universiti Putra Malaysia

Date: 24 May 2019

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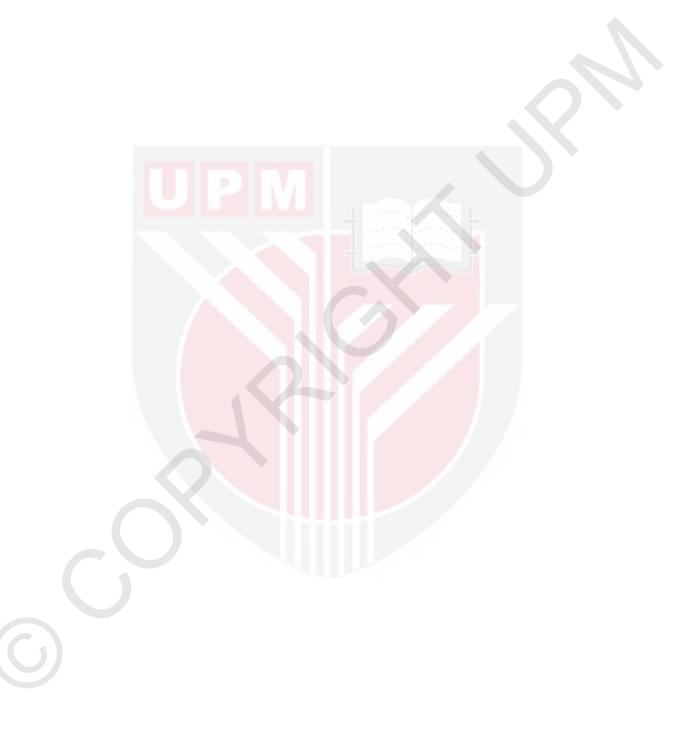
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5.1 Avocado seeds infected with *Botrytis* sp. showing black sclerotia



LIST OF ABBREVIATIONS

FDPM	Forest Department of Peninsular Malaysia
PRF	Permanent Reserved Forest
FNTB	Forest Nursery Terla B
PCR	Polymerase Chain Reaction
ITS	Internal Transcribed Spacer
NCBI	National Centre for Biotechnology Information
BLAST	Basic Local Alignment Search Tool
UPM	Universiti Putra Malaysia
PDA	Potato Dextrose Agarose
RT	Room Temperature
RL	Room Light
UV	Ultraviolet
S	Svedberg
rDNA	ribosomal DNA
FRIM	Forest Research Institute Malaysia
Voltages	V
тѕ	Type Small (small brown spot)
тв	Type Big (big brown spot)
RGR	Radial Growth Rate
bp	Base Pairs

CHAPTER 1

INTRODUCTION

1.1 Background

Cameron Highlands is an extremely important area in the world in biological and ecological aspects of the cloud forest. The cloud forest is limited to high elevation areas and provides habitats of many endemic species that are adapted to high altitude environments (Peh *et al.*, 2011). However, the rich biodiversity of Cameron Highlands has been faced with a lot of human induced problems; unsustainable and unregulated farming, development of residential and resort areas and construction of unsustainable roads. Among them, farming is considered to be one of the major causes of the nature degradation in this area because farming process involves discharge of pesticides and fertilizers, and illegal forest encroachment to expand farmlands for lucrative highland crops. Expansion of farmlands has led to enormous destruction of habitats of the indigenous species and vegetation cover, which promotes landslides and soil erosion on the unstable slopes (Chan, 2006). These landslides have damaged the properties (Pradhan & Lee, 2009).

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Following the catastrophic events, the Cabinet permitted for the Ministry of Water, Land and Natural Resources to implement series of restoration programmes for the degraded forests. In 2015 Forest Department of Peninsular Malaysia (FDPM) was established to carry out the program under the 11th Malaysia Plan with an allocation of RM100 million from 2017 to 2020. This program is planning plantation of 1,025,000 trees in permanent reserved

forest (PRF) areas in Peninsular Malaysia by 2020. In Cameron Highlands, FDPM is going to plant 312,500 trees within about 500 ha land of the PRF by 2020 (FDPM, 2018). As well as FDPM, public groups such as non-governmental organizations and local secondary school students help to plant 40,000 trees within about 40 ha land of the PRF in Cameron Highlands by 2020 (Mohd, 2018).

FDPM selected the following species for the restoration program in Cameron Highlands: *Hopea odorata* (Merawan Siput Jantan: Dipterocarpaceae); *Magnolia elegans* (Cempaka Hutan: Magnoliaceae); *Shorea platyclados* (Meranti Bukit: Dipterocarpaceae); *Casuarina* sp. (Ru: Casuarinaceae); *Parkia speciosa* (Petai: Fabaceae); *Neolamarckia cadamba* (Kelempayan: Rubiaceae); *Exbucklandia populnea* (Gerok: Hamamelidaceae) (Mohd, S. & Mohd, J., personal communication, October 22, 2018). In order to produce high quality seedlings, FDPM established a nursey named Forest Nursery Terla B (FNTB). However, necrotic disease with leaf spot, sub-circular brown lesion, was found on leaves of a lot of *E. populnea* seedlings at FNTB and led many of seedlings to death.

1.2 Problem Statement and Justification

E. populnea is an important species as a common tree for the restoration program in Cameron Highlands (Schmid *et al.*, 1998). However, the necrotic disease damages *E. populnea* seedlings in FNTB. Since FNTB is the only

forest nursery for the restoration in Cameron Highlands and seedlings of *E. populnea* grow very slowly and the germination rate of the seeds is low (Schmid *et al.*, 1998), further spread of the disease might lead to reduction of supply of *E. populnea* seedlings. Therefore, identification of the causal agent was needed to control the disease for management of the forest nursery.

Research on plant pathogenic fungi infecting *E. populnea* has been very limited since Bilgrami and Purohit (1970) described merely one plant pathogenic fungus: *Leptosphaeria* sp.. Thus, this study is expected to play an important role to make a new record of plant pathogenic fungi on *E. populnea*.

1.3 Objectives

To estimate the impact of necrosis (leaf spot) on *E. populnea* in FNTB
 To isolate the causal agent of the necrosis (leaf spot) from *E. populnea*.
 To identify isolates based on morphological and molecular analysis

REFERENCES

Agrios, N. G. (2005). *Plant Pathology 5th Edition*. Burlington, Massachusetts: Elsevier Academic Press.

Aneja, K. R. (2003) *Experiments in Microbiology and Plant Pathology*. Daryaganj, Delhi: New Age International.

Azeman, S. (2016). *Exbucklandia populnea* (R.Br. ex Griff.) R. W. Brown (Hamamelidaceae). Retrieved from http://www.chm.frim.gov.my/Newsletter/Exbucklandia-populnea.aspx. Accessed 13 February 2019.

Barnes, S. E. & Shaw, M. W. (2002). Factors affecting symptom production by latent *Botrytis cinerea* in *Primula* × *polyantha*. *Plant Pathology*, 51, 746-754.

Barnes, S. E. & Shaw, M. W. (2003). Infection of Commercial Hybrid Primula Seed by *Botrytis cinerea* and Latent Disease Spread Through the Plants. *Phytopathology*, 93, 573-578.

Bilgrami, K. S. & Purohit, D. K. (1970). Perithecial stage of *Pestalotia osyridis* Thuem. *Current Science*, 39(1), 7-16.

Bos, L. (1981). Hundred years of Koch's Postulates and the history of etiology in plant virus research. *Netherlands Journal of Plant Pathology*, 87(3), 91-110.

Chan, N. W. (2006). *Cameron Highlands Issues and Challenges in Sustainable Development*. Ipoh, Perak: Bujaya Enterprise.

Chang, K. F., Howard, R. J. & Hwang, S. F. (2007). First Report of Botrytis Blight, Caused by *Botrytis cinerea*, on coneflowers. *Desiase Notes*, 81(12), 1461.

Elad, Y., Williamson, B., Tudzynski, P. & Delen, N. (2007). *Botrytis: Biology, pathology and control*. Dordrecht, South Holland: Springer.

FDPM. (2018). Annual report of restoration program, reclamation and recovery of deteriorated areas in peninsular Malaysia 2017. Kuala Lumpur: Forest Department of Peninsular Malaysia

Fitt, B. D. L., Creighton, N. F. & Bainbridge, A. (1985). Role of wind and rain in dispersal of *Botrytis fabae* conidia. *Transactions of the British Mycological Society*, 85(2), 307-312.

Fredericks, D. N. & Relman, D. A. (1996). Sequence-based identification of microbial pathogens: a reconsideration of Koch's postulates. *Clinical Microbiology Reviews*, 9(1), 18-33.



García, M. M., Denno, B. D., Miller, D. R., Miller, G. L., Ben-Dov, Y. & Hardy, N. B. (2016). ScaleNet: A literature-based model of scale insect biology and systematics. Retrieved from http://scalenet.info/about/. Accessed 12 April 2019.

Hao, F., Ding, T., Wu, M., Zhang, J., Yang, L., Chen, W. & Li, G. (2018). Two novel hypovirulence-associated mycoviruses in the phytopathogenic fungus *Botrytis cinerea*: molecular characterization and suppression of infection cushion formation. *Viruses*, 10(5), 254.

Hou, D. (1958). Hamamelidaceae. Flora Malesiana, 5(1), 374-376.

Hua, C., Kai, K., Wang, X., Shi, W., Zhang, D. & Liu, Y. (2019). Curcumin inhibits gray mold development in kiwifruit by targeting mitogen-activated protein kinase (MAPK) cascades in *Botrytis cinerea*. *Postharvest Biology and Technology*, 151, 152-159.

James, R. L., Dumroese, R. K. & Wenny, D. L. (1995). *Botrytis cinerea* carried by adult fungus gnats (Diptera: Sciaridae) in container nurseries. *The Planter's Notes* 46(2), 48-53.

Jarvis, W. R. (1977). *Botrytinia and Botrytis Species: Taxonomy, Physiology, and Pathogenicity, A guide to the Literature. Monograph No.15.* Ottawa, Ontario: Canada Department of Agriculture.

Jasalavich, C. A. & Schumann, G. L. (2001). Who Done It? Or what's that brown fuzzy stuff on my plum? (Koch's Postulates for Proof of Pathogenicity). Retrieved from https://www.apsnet.org/edcenter/K-12/TeachersGuide/BrownRot/Pages/default.aspx. Accessed 16 January 2019.

Keller, M., Viret, O. & Cole, F. M. (2002). *Botrytis cinerea* Infection in grape defense reaction, latency, and disease expression. *Phytopathology*, 93(3), 316-322

Kimura, M. (1980). A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution* 16, 111-120.

Krik, W., Dudek, T. & Byrne, J. (2007). Managing botrytis blight on Hosta. Retrieved from https://www.canr.msu.edu/news/managing_botrytis_blight_on_hosta. Accessed 21 March 2019.

Kumar, S., Stecher, G. & Tamura, K. (2016). MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. Molecular. *Biology and Evolution* 33, 1870-1874.

Lattore, B. A., Elfar, K., & Ferrada, E. E. (2015). Gray mold caused by *Botrytis cinerea* limits grape production in Chile. *Crop Protection*, 42(3), 305-330.

 \bigcirc

Li, Y., Sun, S., Du, C., Xu, C., Zhang, J., Duan, C. & Zhu, Z. (2016) A new disease of mung bean caused by *Botrytis cinerea*. *Crop Protection*, 85, 52-56.

Lilja, A., Poteri, M., Petäistö, R. L., Rikala, R., Kurkela, T. & Kasanen, R. (2010). Fungal diseases in forest nurseries in Finland. *Silva Fennica*, 44(3), 525-545.

Liu, S., Moayeri, M. & Leppla, S. H. (2014). Anthrax lethal and edema toxins in anthrax pathogenesis. *Trends in Microbiology*, 22(6), 317-325.

Lopes, U. P., Zambolim, L., Costa, H., Pereira, O. L. & Finger, F. L. (2014). Potassium silicate and chitosan application for gray mold management in strawberry during storage. *Crop Protection*, 63, 103-106.

Manners, A. (2016). Scale insects A difficult problem can be managed [Fact Sheet]. Retrieved from https://www.horticulture.com.au/globalassets/hort-innovation/resource-assets/ny15002-scale-insects-pest-mgmt-plan.pdf. Accessed 14 April 2019.

Missouri Botanical Garden. (n.d.). Leaf Spot Diseases of Shade Trees and Ornamentals. Retrieved from http://www.missouribotanicalgarden.org/gardens-gardening/yourgarden/help-for-the-home-gardener/advice-tips-resources/pests-andproblems/diseases/fungal-spots/leaf-spot-shade.aspx. Accessed 14 April 2019.

Mohd, S. (2018). The coordination meeting for the joint action of the rehabilitation of the Cameron Highlands [PowerPoint slides].

Nasehi, A., Kadir, J. B., Zainal, A., Wong, M. Y. & Mahmodi, F. (2012a). First report of *Alternaria tenuissima* causing leaf spot on eggplant in Malaysia. *Disease Notes*, 96(8), 1226.

Nasehi, A., Kadir, J. B., Zainal, A., Wong, M. Y. & Mahmodi, F. (2012b). First report of grey leaf spot on pepper caused by *Stemphylium solani* in Malaysia. *Disease Notes*, 96(8), 1227.

Nasehi, A., Kadir, J. B., Zainal, A., Wong, M. Y. & Mahmodi, F. (2012c). First report of tomato grey leaf spot disease caused by *Stemphylium solani* in Malaysia. *Disease Notes*, 96(8), 1226.

Nasehi, A., Kadir, J., Bin, A. F. A., Nasr-Esfahani, M., Wong, M. Y., Rambe, S. K., Ghadirian, H., Mahmodi, F. & Golkhandan, E. (2014). *Alternaria capsicicola* sp. nov., a new species causing leaf spot of pepper (*Capsicum annuum*) in Malaysia. *Mycological Progress*, 13, 1041-1048.

National Centre for Biotechnology Information. (n.d.). Polymerase Chain Reaction (PCR). Retrieved from https://www.ncbi.nlm.nih.gov/probe/docs/techpcr/. Accessed 16 March 2019.



National Human Genome Research Institute. (n.d.). Polymerase Chain Reaction (PCR) [Fact Sheet]. Retrieved from https://www.genome.gov/10000207/polymerase-chain-reaction-pcr-fact-sheet/. Accessed 16 March 2019.

Pegg, K. & Manners, A. (2017). *Botrytis* An opportunistic pathogen [Fact Sheet]. Retrieved from https://www.ngia.com.au/Attachment?Action=Download&Attachment_id=201 7. Accessed 16 March 2019.

Peh, K. S. H., Soh, M. C. K., Sodhi, N. S., Laurance, W. F., Ong, D. J. & Clements, R. (2011). Up in the clouds: Is sustainable use of tropical montane cloud forests possible in Malaysia? *BioScience*, 61(1), 27-38.

Photita, W., Taylor, P. W. J., Ford, R., Hyde, K. D. & Lumyong, S. (2005). Morphological and molecular characterization of *Colletotrichum* species from herbaceous plants in Thailand, *Fungal Diversity*, 18, 117-133.

Poczai, P. & Hyvönen, J. (2009). Nuclear ribosomal spacer regions in plant phylogenetics: problems and prospects. *Molecular Biology Reports*, 37(4), 1897-1912.

Pradhan, B. & Lee, S. (2009). Regional landslide susceptibility analysis using back-propagation neural network model at Cameron Highland, Malaysia. *Landslides*, 7(1), 13-30.

Rivera, M. C. & Lopez, S. E. (2007). First report of *Botrytis cinerea* on pansy flowers in Buenos Aires. *Disease Notes*, 88(10), 1164.

Romanazzi, G., Nigro, F., Ippolito, A., DiVenere, D. & Salerno, M. (2002). Effects of pre- and postharvest chitosan treatments to control storage grey mold of table grapes. *Journal of Food Science*, 67(5), 1862-1867.

Salleh, B., Safinat, A., Julia, L. & Teo, C. H. (1996). Brown spot caused by *Curvularia* spp., a new disease of asparagus. *Biotropia*, 9, 26-37.

Schmid, R., Sosef, M. S. M., Hong, L. T. & Prawirohatmodjo, S. (1998). Timber trees: lesser-known timbers. *Taxon*, 47(2), 227-229.

Schoch, C. L., Seifert, K. A., Huhndorf, S., Robert, V., Spouge, J. L., Levesque, C. A., Chen, W. & Fungal Barcoding Consortium (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. *Proceedings of the National Academy of Sciences*, 109(16), 6241-6246.

Schumacher, J. (2017). How light affects the life of *Botrytis*. *Fungal Genetics and Biology*, 106, 26-41.



Sutherland, J. R. & Glover, S. G. (1991). *Proceedings of the first meeting of IUFRO Working Party S2.07-09 (Diseases and Insects in Forest Nurseries)*. Victoria, British Columbia: Canadian Forest Service.

Swart, L. & Langenhoven, P. (2007). First report of botrytis blight, caused by *Botrytis cinerea*, on hibiscus in South Africa. *Disease Notes*, 84(4), 487.

Tanovic, B., Hrustic, J., Mihajlovic, M., Grahova, M. & Delibasic, G. (2014). *Pestic Phytomed*, 29(4), 237-247.

Thomas, C. S., Marois, J. J. & English, J. T. (1987). The effect of wind speed, temperature, and relative humidity on development of aerial mycelium and conidia of *Botrytis cinerea* on grape. *Phytopathology*, 78, 260-265.

U.S. Department of Agriculture Forest Service. (n.d.). *Bucklandia populnea* [Fact Sheet]. Retrieved from https://www.fpl.fs.fed.us/documnts/TechSheets/Chudnoff/SEAsian_Oceanic/ new_html_docs/Bucklandia.html. Accessed 20 February 2019.

van Kan, J. A. L., Shaw, M. W. & Grant-Downton, R. T. (2014). *Botrytis* species: relentless necrotrophic thugs or endophytes gone rogue? *Molecular Plant Pathology*, 15(9), 957-961.

Vasilica, M. R., Suciu, L. A. & Puia, C. E. (2012). ProEnvironment, 5, 60-66.

White, T. J., Bruns, T., Lee, S. & Taylor, J. (1990). Amplification and Direct Sequencing of Fungal Ribosomal RNA Genes for Phylogenetics. In N. Innis, D. Gelfand, J. Sninsky, T. White (Eds.), *PCR Protocols, a Guide to Methods and Applications*, 315-322. New York: Academic Press.

Williamson, B., Duncan, G., Harrison, G. J., Harding, A. L., Elad, Y. & Zimand, G. (1995). Effect of humidity on infection of rose petals by dry-inoculated conidia of *B. cinerea*. *Mycological Research*, 99(11), 1303-1310.

Wu, M. D., Zhang, L., Li, G. Q., Jiang, D. H., Hou, M. S. & Huang, H. C. (2007). Hypovirulence and Double-Stranded RNA in *Botrytis cinerea*. *Phytopathology*, 97,1590-1599.

WWF-Malaysia. (2002). Study for the Sustainable Development of the Highlands of Peninsular Malaysia, Final Report Volume II [Governmental Report]. Retrieved from http://repository.wwf.org.my/technical_reports/S/StudyForTheSustainableDev elopmentOfTheHighlandsOfPeninsularMalaysiaFinalReportVolumeIIMainRep ortPart1.pdf. Accessed 17 March 2019.

WWF-Malaysia. (n.d.) The Malaysian Rainforest. Retrieved from http://www.wwf.org.my/about_wwf/what_we_do/forests_main/the_malaysian _rainforest/. Accessed 17 March 2019.



Zimbro, M. J., Power, D. A., Miller, S. M., Wilson, G. E. & Johnson, J. A. (2009). *Difco™ & BBL™ Manual Manual of Microbiological Culture Media, Second Edition.* Sparks, Nevada: Becton Dickinson.

