

ANTAGONISTIC ACTIVITY OF Xylaria sp. AND Neopestalotiopsis egyptiaca AGAINST THREE TREE PATHOGENS

CHAN THIP A/P BOON SIENG

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CHAN THIP A/P BOON SIENG

FACULTY OF FORESTRY UNIVERSITI PUTRA MALAYSIA

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Ву

CHAN THIP A/P BOON SIENG

A Project Report Submitted in Partial Fulfilment of the Requirements for the Degree of Bachelor of Forestry Science in the Faculty of Forestry Universiti Putra Malaysia

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DEDICATION

Specially dedicated to my:

Parent

Siblings

Prof. Dr. Rozi Mohamed

Faculty of Forestry

Thank you for your encouragement and support

And all the sacrifices that you have given

Thank you for everything. May god bless all of us.

ABSTRACT

Fusarium oxysporum, Ceratocystis fimbriata and Phellinus noxius are pathogenic fungi and causal agent of disease to plantation and forest tree. Chemical fungicides are commonly used to control these pathogen, but they are not environmental friendly. As alternative, biological control agents are being sought to replace chemicals. In this study, the growth performance and morphological characteristics of two endophytic fungal (Xylaria sp. and Neopestalotiopsis egyptiaca) and three fungal pathogens (C. fimbriata, P. noxius and F. oxysporum) were examined. They were inoculated on Potato Dextrose Agar (PDA) : 1) normal PDA (aPDA). and 2) PDA with adjusted pH values between 7.2-7.4 (nPDA). The growth of *N. egyptiaca* was faster than Xylaria sp. growth, while the growth of F. oxysprorum was three times and two times faster than C. fimbriata and of P. noxius respectively. The antagonistic activity was determined through dual culture assay and nonvolatile compound assay. Xylaria sp. was a better pathogen inhibitor compared to *N. egyptiaca*. Based on percentage inhibition of radial growth (PIRG) value Xylaria sp. could inhibit three tree pathogens while N. egyptiaca only could inhibit P. noxius and less effective to control the growth C. fimbriata and F. oxysporum. This shows that between these two endophytic fungi isolates, Xylaria sp. has potential as biological control agents against tested fungal pathogens.

ABSTRAK

Fusarium oxysporum, Ceratocystis fimbriata dan Phellinus noxius adalah kulat patogen dan agen penyebab penyakit kepada pokok ladang dan hutan. Racun kulat kimia biasanya digunakan untuk mengawal patogen ini, tetapi ia tidak mesra alam. Sebagai alternatif, agen kawalan biologi sedang dicari bagi menggantikan bahan kimia. Dalam kajian ini, prestasi pertumbuhan dan ciriciri morfologi dari dua kulat endophitik (Xvlaria sp. dan Neopestalotiopsis egyptiaca) dan tiga patogen kulat (Ceratocystis sp., Phellinus noxius dan Fusarium oxysporum) telah dikaji. Mereka telah diinokulasikan pada Potato Dextrose Agar (PDA): 1) PDA biasa (aPDA). dan 2) PDA dengan nilai pH diselaraskan antara 7.2-7.4 (nPDA). Pertumbuhan N. eqyptiaca lebih cepat daripada pertumbuhan Xylaria sp. sementara pertumbuhan F. oxysprorum adalah tiga kali dan dua kali lebih cepat daripada C. fimbriata dan P. noxius masing-masing. Aktiviti antagonistik ditentukan melalui pengujian kultur dwi dan non-volatile compound. Xylaria sp. adalah perencat patogen yang lebih baik berbanding dengan N. egyptiaca. Berdasarkan perencatan nilai radial (PIRG) nilai Xylaria sp. boleh menghalang tiga patogen pokok manakala N. egyptiaca hanya boleh menghalang P. noxius dan kurang berkesan untuk mengawal pertumbuhan C. fimbriata dan F. oxysporum. Ini menunjukkan bahawa di antara dua kulat endophytic ini, Xylaria sp. mempunyai potensi yang lebih baik sebagai agen kawalan biologi terhadap patogen kulat yang diuji.

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APPROVAL SHEET

I certify that this research project report entitled "Antagonistic activity of *Xylaria* sp. and *Neopestalotiopsis egyptiaca* against three tree pathogen" by Chan Thip a/p Boon Sieng has been examined and approved as a partial fulfillment of the requirements for the Degree of Bachelor of Forestry Science in the Faculty of Forestry, Universiti Putra Malaysia.



Prof. Dr. Rozi Mohamed Faculty of Forestry Universiti Putra Malaysia (Supervisor)

Prof. Dr. Mohamed Zakaria Bin Hussin Dean Faculty of Forestry Universiti Putra Malaysia

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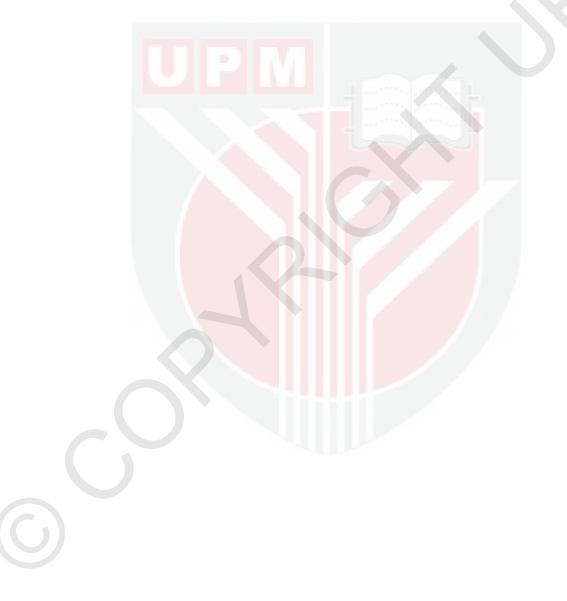
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LIST OF ABBREVIATIONS

aPDA	Potato Dextrose Agar with adjusted pH value 7.2pH -
	7.4pH PDA
nPDA	Potato Dextrose Agar with normal pH value
PIRG	Percentage of inhibition



CHAPTER ONE

INTRODUCTION

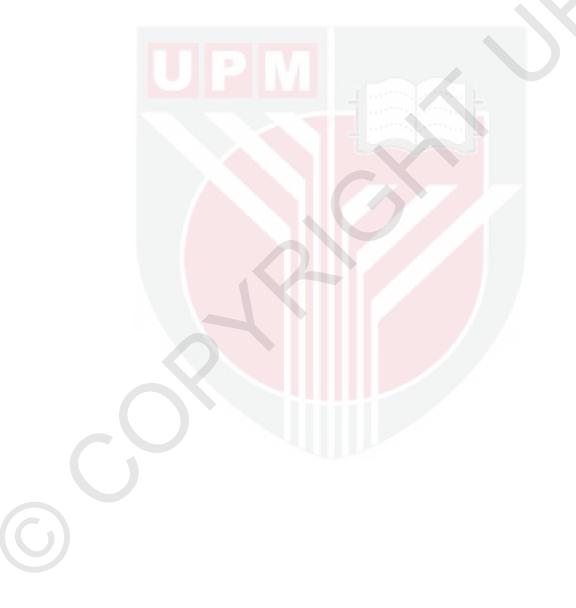
1.1 Background

In Malaysia there are existence of Forest Plantation and Industrial crops plantation. Based on Malaysian Timber Industry Board (MTIB) there are nine tree species recommended by the Ministry to be planted in the forest plantation include Rubber (Hevea brasiliensis), Acacia spp. (Acacia mangium or hybrid), Teak (Tectona grandis), Sentang (Azadirachta excelsa), Khaya (Khaya ivorensis/Khaya senegalensis), Kelempayan or spp. Laran (Neolamarckia cadamba), Batai (Paraserianthes falcataria), Binuang (Octomeles sumatrana) and five selected species of bamboo. Other than forest plantation, industrial crop plantation also important which is planted food crop such as coconut (Cocos nucifera), coffea, tea and etc. There are total 390,000 ha of forest plantation in Peninsular Malaysia, 271,110 ha in 2013 in Sabah and 471,892 ha in 2013 in Sarawak. The total is about 1,133,092 ha and only 133,951.8 ha is industrial crop plantation for the whole Malaysia in 2016. These two kinds of plantation could get infected by diseases that lead to loses of money.

Forest plantations and crop plantation are very important to the country since they provide food and generate income to our country. The forest plantation is an important alternative for the production of wood other than direct felling from natural forest. But in the plantation area the environment is modified environment for the plant compare to the natural forest environment and this will give the impact to the plant in term of their immunity towards disease. The present of human in an area will introducing more disease and pest because of human will bring pathogen from outside.

In Malaysia the pest and disease control is done to avoid the more lost on the yield and income to the plantation. For agriculture sector the pest and disease control is done through their feeding and burrowing activities and the disease that involving microorganism such as fungi, bacteria and virus. The famous disease that affected our crops and forest plantation is *Ganoderma* basal stem rot and *Maramius* bunch rot. The most important thing is uses of environmental friendly control method for pathogenic fungi, but in Malaysia the main weapon to control the disease is chemical fungicides. The used of the fungicides for long period may lead to the fungicide-resistant strains, and the fungicides also able became harmful to the consumer of the crop (Hewajulige and Wijesundera, 2014).

In cocoa plantation there are practices to avoid the pest such as cocoa pod borer that first reported to be used in Sabah in late 1980. There is combination of the control method to increase the degree of success to control the disease. The common methods that been applied is clean and more frequent harvesting, selecting spraying of moth resting and sleeving the young pods to prevent the laying egg of pest. The Vascular Streak Dieback (VSD) is one of the example that occur in the plantation and it been estimated to cause the loss about 25%-30%. The method used to control this disease is by using chemical control, Isolation or barrier, disease avoidance, pruning, disease resistance, rehabilitation and culture or nutrient practices. In oil palm plantation area, a basidiomycete's fungus, *Ganoderma* sp. which is the causal agent for the basal stem rots disease. This disease ruined thousand hectare of oil palm plantation in Southeast Asia especially Indonesia and Malaysia (Azahar et al., 2014).



1.2 Problem Statement

Pathogen can cause many diseases to plant especially crop and this is causing loses in term of the productivity of the crops. Many people solve this problem by using chemical fungicides. The chemical fungicides give effect to the environment such as the soil pollution as well as water pollution. The chemical fungicides also can cause effect to the plant as well. In long time period pathogen can produce the resistance against the fungicides. Chemical fungicides can cause proliferation of resistance in the pathogen populations. (Chet & Indar, 1994).

The biological control is different for different species because every species has different natural enemies. The study about the biological control of each disease that occurred is needed, because the previous finding from study may be not suitable for the disease currently this is because the development of new resistance. Furthermore, the study that has been done in the laboratory may be not suitable to be applied in the real situation, thus further study is needed because we need to apply the result of the research in the field. In order to control the disease there are lack of information about the species that cause the diseases.

1.3 Objectives

- To examine the growth performance of *Xylaria* sp., *Neopestalotiopsis* egyptiaca, *Ceratocystis fimbriata*, *Fusarium oxysporum* and *Phellinus noxius* on normal Potato Dextrose Agar (PDA) and PDA with adjusted pH value (7.2 - 7.4).
- To evaluate the antagonistic effect of two endophytes (*Xylaria* sp. and *N. egyptiaca*) on three tree pathogen (*C. fimbriata*, *F. oxysporum* and *P. noxius*) using dual culture and non-volatile compound assay.

REFERENCES

Amnuaykanjanasin, A., Punya, J., Paungmoung, P., Rungrod, A., Tachaleat, A., Pongpattanakitshote, S., Cheevadhanarak, S., & Tanticharoen, M. (2005). Diversity of type I polyketide synthase genes in the wood-decay fungus Xylaria sp. BCC 1067. *FEMS Microbiology Letters*, 251, 125–136

Aneja, M., Gianfagna, T.J., & Hebbar, P.K. (2006). *Trichoderma* produces nonanoic acid, an inhibitor of spore germination and mycelial growth of two cacao pathogens. *Physiological and Molecular Plant Pathology*, 67, 304–307

Ann, P.J., Chang, T.T., & Ko, W.H. (2002). *Phellinus noxius* Brown Root Rot of Fruit and Ornamental Trees in Taiwan. *Plant Disease*, 86, 820-826

Armstrong, G.M., & Armstrong, J. (1981). Formae speciales and races of *Fusarium oxysporum* causing wilt diseases. *Fusarium : Disease, Biology, and Taxonomy*, 1, 391-399

Arzanlau, M., Jannati, E., Habibzadeh, S., Mohammadi, S., Ahadi, P., Mohammadi, G.B., Dogaheh, H.P., Dibah, S. & Kazemi, E. (2013). Nasal colonization of mecA-positive, oxacillin-susceptible, methicillin-resistant Staphylococcus aureus isolates among nursing staff in an Iranian teaching hospital. *American Journal of Infection Control*, 41, 122-124

Azahar, T., Idris, A., Abu, H., & Norazlin (2014). Assessment of Basal Stem Rot Disease Distribution in Palm Oil Plantation Using Geographical Information System. *Journal of Science and Technology*, 6, 29-60

Bai, Z., Harvey, L.M., & McNeil, B. (2003). Oxidative stress in submerged culture of fungi. *Critical Reviews in Biotechnology*, 23, 267-302

Baker, C.J., Harrington, T.C., Krauss, U., & Alfenas, A.C. (2003) Geneticvariability and host specialization in the Latin American cladeof *Ceratocystis fimbriata. Phytopathology*, 93, 274 –284

Beer, Z.W., Duong, T.A., Bames, I., Wingfield, B.D., & Wingfiled, M.J. (2014). Redefining *Ceratocystis* and allied genera. *Studies in Mycology*, 79, 187–219

Bonthond, G., Denis, M.S., Groenewald, J.Z., & Crous, P.W. (2017). Seiridium (Sporocadaceae): an important genus of plant pathogenic fungi. *Persoonia*, 40, 96–118

Boonphong, S., Kittakoop, P., Isaka, M., Pittayakhajonwut, D., Tanticharoen, & Thebtaranonth, Y. (2001). Multiplolides A and B, New Antifungal 10-Membered Lactones from *Xylaria* multiplex. *Journal of Natural Products,* 64, 65–67

Boyd, L.A., Ridout, C., Sullivan, D.M., Leach, J.E., & Leung, H. (2013). Plantpathogen interactions: disease resistance in modern agriculture. *Trends in Genetics*, 29, 233-240



Brasier, C.M. (1979). Dual origin of recent Dutch elm disease outbreaks in Europe. *Nature*, 281, 78–80

Brasier, C.M. (1983). A cytoplasmically transmitted disease of *Ceratocystis ulmi*. *Nature*, 305, 220–223

Brasier, C.M., & Gibbs, J.N. (1975). Highly fertile form of the aggressive strain of *Ceratocystis ulmi*. *Nature*, 257, 128–131

Brum, M.C.P., Araujo, W.L., Maki, C.S., & Azevedo, J.L. (2012). Endophytic fungi from *Vitis labrusca* L. ('Niagara Rosada') and its potential for the biological control of *Fusarium oxysporum*. *Genetic and Molecular Research*, 11, 187-197

Brunner, F., & Petrini, O. (1992). Taxonomy of some *Xylaria* species and xylariaceous endophytes by isozyme electrophoresis. *Mycolecular Research*, 96, 23-33

Burgess, L.W. (1981). General Ecology of the Fusaria. In: Nelson PE, Toussoun TA, Cook cRJ, eds. Fusarium: diseases, biology and taxonomy. University Park, PA, USA: The Pennsylvania State University Press, 225–235

Camila, M.O., Regasini, L.O., Silva, G.H., Pfenning, L.H., Young, M.C., Berlinck, G.S., Bolzani, V.S., & Araujo, A.R. (2011). Dihydroisocoumarins produced by *Xylaria* sp. and *Penicillium* sp., endophytic fungi associated with Piper aduncum and Alibertia macrophylla. *Phytochemical Letters*, *4*, 93-96

Chakraborty, U., Chakrabort, B., & Basnet, M. (2006). Plant growth promotion and induction of resistance in Camellia sinensis by Bacillus megaterium. *Journal of Basic Microbiology*, 46, 186–195

Chaloupka, J. & Krumphanzl, V. (1987). Extracellular Enzymes of Microorganisms. *Plenum Press*, 1, 216

Chang, T. (1996) Survival of *Phellinus noxius* in soil and in the roots of dead host plants. *Phytopathology*, 86, 272-276

Chapela, I.H., Petrini, O., & Petrini, L.E. (1990). Unusual ascospore germination in Hypoxylon fragiforme: first seps in the establishment of an endophytic symbiosis. *Canadian Journal of Botany*, 68, 571-575

Chaverri, P., Salgado, C., Hirooka, Y., Rossman, A.Y., & Samuels, G.J. (2011) Delimitation of Neonectria and Cylindrocarpon (Nectriaceae, Hypocreales, Ascomycota) and related genera with Cylindrocarpon-like anamorphs. *Studies of Mycology*, 68, 57–78

Chet, I., & Indar, J. (1994). Biological control of fungi. *Biochemistry and biotechnology*, 48, 37-43

Costa, I., Silva, G.H., Camila, M.O., Teles, H.L., Pauletti, P.M., Silva, D.H., Bolzani, V.S., Young, M.C., Claudio, M., Costa, N,. Pfenning. L., Berlinck, R.G., & Araujo, A.R. (2010). Sesquiterpenes from *Xylaria* sp., an endophytic fungus associated with *Piper aduncum* (Piperaceae). *Phytochemistry*, 3, 164–167

Crous, P.W., Wingfield, M.J., & Roux, J.J. (2015). Fungal Planet description sheets: 371–399. *Persoonia*, 35, 264-327

Dai, C., Gao, F., & Liu, X. (2010). Mechanisms of fungal endophytes in plant protection against pathogens. *African Journal of Microbiology Research*, 4, 346-351

Dreyfuss, M.M. (1986). Sydowia. Mycologia, 39, 22-36

Engelbrecht, C.J., Harrington, T.C., & Alfenas, A., (2007). a. Ceratocystiswilt of cacao – a disease of increasing importance. *Phytopathology*, 97, 648–649

Engelbrecht, C.B., & Harrington, T.C. (2017). Intersterility, morphology and taxonomy of *Ceratocystis fimbriata* on sweet potato, cacao and sycamore. *Journal Mycologia*, 97, 57-69

Engelbrecht, C.J., Harrington, T.C., Steimel, J., & Capretti, P. (2004). Genetic variation in eastern North American and putatively introduced populations of *Ceratocystis fimbriata f. platani. Molecular Ecology*, 13, 95–105

Fisher, P.J, & Petrini, O. (1990). A comparative study of fungal endophytes in xylem and bark of Alnus species in England and Switzerland. *Mycology research*, 94, 313-319

Fravel, D., Olivian, C., & Alabouvette, C. (2002). Research review *Fusarium* oxysporum and its biocontrol. *New Phytologist*, 157: 493 – 502

Garrett, D. (1970) *Pathogenic root-infecting fungi*. Cambridge University Press. Cambridge, 294

Gerlach, W., & Nirenberg, H. (1982). The genus Fusarium- A pictoral atlas. *Mitt. Biol. Bundesanst. Land-Forstwirtsch. BerlinDahlem,* 209, 1–406

Gloer, J.B. (1997) Applications of fungal ecology in the search for new bioactive natural products. *Environmental and Microbial Relationships*, 4, 249–268

Guarro, J., Gene, J., Stchigel, A.M., & Figueras, M.J. (2012) – Atlas of soil ascomycetes. *CBS Biodiversity* Series No. 10. Biodiversity Centre, Utrecht CBS-KNAW Fungal

Hamzah, T.N.T., Lee, S.Y., Hidayat, A., Terhem, R., Hanum, F., & Mohamed, R. (2018). Diversity and characterization of endophytic fungi isolated from the

tropical mangrove species, *Rhizophora mucronata*, and identification of potential antagonists against the soil-B. *Front Microbiology*, 9,177

Harper, D.B., Kennedy, J.T., & Hamilton, J.T. (1988). Chloromethane biosynthesis in poroid fungi. *Phytochemistry*, 27, 147-153

Harrington, T.C., & Wingfield, M.J. (1998). The *Ceratocystis* species on conifers. *Canadian Journal of Botany*, 76, 446-447

Hewajulige, I., & Wijesundera, R. (2014). *Thielaviopsis paradoxa, Thielaviopsis basicola* (Black Rot, Black Root Rot). *Control Strategies*, 1, 287-308

Hyde, K.D., & Lee, S.Y. (1995). Ecology of mangrove fungi and their role in nutrient cycling: what gaps occur in our knowledge?. *Hydrobiologia*, 295, 107–118

Ismail, A., Perrone, G., & Crous, (2015). Neopestalotiopsis egyptiaca. *Fungal Planet,* 1, 372 – 374

Jaklitsch, W.M., Gardiennet, A., & Voglmayr, H. (2016). Resolution of morphology-based taxonomic delusions: Acrocordiella, Basiseptospora, Blogiascospora, Clypeosphaeria, Hymenopleella, Lepteutypa, Pseudapiospora, Requienella, Seiridium and Strickeria. *Persoonia*, 37, 82– 105

Jang, Y.W., Lee, I.K., Kim, Y.S., Lee, S., Lee, H.J., Yu, S.H., & Yun, B.S. (2007). Xylarinic Acids A and B, New Antifungal Polypropionates from the Fruiting Body of Xylaria polymorpha. *The Journal of Antibiotics*, 60, 696–699

Jayasuriya, H., Herath, K.B., Ondeyka, J.G., Polishook, J.D., Bills, G.F., Dombrowski, A.W., Springer, M.S., Siciliano, S., Malkowitz, L., Sanchez, M., Guan, Z., Tiwari, S., Stevenson, D.W., Borris, R.P., & Singh, S.B. (2004) – Isolation and structure of antagonists of chemokine receptor (CCR5). *Natural Products*, 67, 36–38

Jayawardena, R.S., Liu, M., & Maharachchikumbura, S.S. (2016). *Neopestalotiopsis vitis* sp. nov. causing grapevine leaf spot in China. *Phytotaxa*, 258, 63-74

Ju, Y.M., & Rogers, J.D. (1990). Astrocystis Reconsidered. *Mycologia*, 82, 342-349

Kile, G.A., Harrington, T.C., Yuan, Z.Q., Dudzinski, M.J., & Old, K.M. (1996). *Ceratocystis eucalypti* sp. nov., a vascular stain fungus from eucalypts in Australia. *Mycoloy Research*, 100, 571–579

Kok, C.J., Haverkamp, W., & VanDerAa, H., (1992). Influence of pH on the growth and leaf-maceration ability of fungi involved in the decomposition of

floating leaves of Nymphaea alba in an acid water. *Microbiology*, 138, 103-108

Kumara, K.L., & Rawal, R.D. (2008). Influence of carbon, nitrogen, temperature and pH on the growth and sporulation of some indian isolates of *Colletotrichum gloeosporioides* causing anthracnose disease of papaya (*Carrica papaya* L). *Tropical Agriculture Research and Extinction*, 11, 7-12

Kusari, K., Pandey, S.P., & Spiteller, M. (2013). Untapped mutualistic radigms linking host plant and endophytic fungal production of similar bioactive secondary metabolites. *Phytochemistry*, 91, 81–87

Larsen, M.J., & Cobb-Poulle, L. (1990). *Phellinus (Hymenochaetaceae)*. *A Survey of the World Taxa. Synopsis Fungorum*, 3, 68-71

Leduc, G., Sankoff, D., Antoine, N., Paquin, B., Lang, B.F., & Cedergren, R. (1992). Gene order comparisons for phylogenetic inference: *Evolution of the mitochondrial genome*, 89, 6575-6579

Lee, S.S. (2014). Guest editorial: Lamenting the state mycology and forest pathology in Malaysia. *Journal of Tropical Forest Science*, 26, 443-445

Lei, H., Lin, X., Han, L., Ma, J., Ma, Q., Zhong, J., Liu, Y., Sun, T., Wang, J., & Huang, X. (2017). New metabolites and bioactive chlorinated benzophenone derivativesproduced by a marine-derived fungus *Pestalotiopsis heterocornis. Marine Drugs*, 15, 69-70

Liew, E., Wingfield, M., Assa, B., Paath, J., Kandowangkossor, D., Sembel, T., Summerell, B., & Burgess, L. (2003). *Ceratocystis fimbriata* associated with clove decline in North Sulawesi. *International Congress of Plant Pathology*, 2,19-40

Liu, X., Dong, M., Chen, X., Jiang, M., Lv, X., & Yan, G. (2007). Antioxidant activity and phenolics of an endophytic *Xylaria* sp. from *Ginkgo biloba*. *Food Chemistry*, 105, 548–554

Lombard, L., Merwe, N.A., Groenewald, J.Z., & Crous, P.W. (2015). Generic concepts in Nectriaceae. *Studies in Mycology* 80: 189-245

Lotter, D.W., Granett, J., & Omer, A.D. (1999). Differences in Grape Phylloxera-related Grapevine Root Damage in Organically and Conventionally Managed Vineyards in California. *HortScience*, 34, 108-111

Lumbsch, H.T. (2000). Phylogeny of filamentous ascomycetes. *Naturwissenschaften*, 87, 335–342

Maharachchikumbura, S.S.N., Hyde, K.D., Groenewald, J.Z., Xu, J., & Crous, P.W. (2014). Pestalotiopsis revisited. *Studies in Mycology*, 79, 121-186

Manion, P.D., (1981). Tree Disease Concepts. *Prentice-Hall, Englewood Cliffs*, 399

Martos. S, Luque. J & Philips. A.J.L. (2005). Botryosphaeria viticola sp. nov. on grapevines: a new species with a Dothiorella anamorph. *Journal Mycologia*, 97, 111-121

Masui, H., Kondo, T., & Kojima, M. (1989). An antifungal compound, 9,12,13-Trihydroxy-(E)- 10-Octadecenoic acid, from colocasia antiquorum inoculated with *Ceratocystis fimbriata*. *Phytochemlstry*, 28, 13-19

Mayers, C.G., Mcnew, D.L., Harrington, T.C., Roeper, R.A., Fraedrich, S.W., Biedermann, P.H.W., Castrillo, L.A., & Reed, S.E. (2015). Three genera in the Ceratocystidaceae are the respective symbionts of three independent lineages of ambrosia beetles with large, complex mycangia. *fungal biology*, 119, 75-92

Migheli, Q., Lauge, R., Daviere, J.M., Gerlinger, C., Kaper, F., Langin, T., & Daboussi, M.J. (1999). Transposition of the autonomous Fot1 element in the filamentous fungus *Fusarium oxysporum*. *Genetics*, 151, 105–113

Mikusova, P., Ritieni, A., Santini, A., & Juhasova, G. (2010). Contamiantion by moulds of grape berries in Slovakia. *Journal Food Additives & Contaminants*, 27, 738-747

Miyashira, C.H., Tanigushi, D.G., Gugliotta, A.M., & Santos, D.Y. (2010). Comparison of radial growth rate of the mutualistic fungus of Atta sexdens rubropilosa forel in two culture media. *Brazilian Journal of Microbiology Print*, 41, 517-382

Mullenborn, C., Steiner, U., Ludwig, M., & Oerke, E.C. (2007). Effect of fungicides on the complex of *Fusarium* species and saprophytic fungi colonizing wheat kernels. *European Journal of Plant Pathology*, 120, 157–166

Nagamani, P., Bhagat, S., Biswas, M.K., & Vismanath, K. (2017). Effect of Volatile and Non Volatile Compounds of Trichoderma spp. against Soil Borne Diseases of Chickpea. *International Journal of Current Microbiology and Applied Sciences*, 6, 486-491

Nakagiri, A. (1998). *Diversity of halophytophthoras in subtropical mangroves and factors affecting their distribution*. Proceedings of the Asia-Pacific Mycological Conference on Biodiversity and Biotechnology. Hua Hin, Prachuapkhirikhan, Thailand, 109-113

Niemela, T., & Kotiranta, H. (1982). Polypore survey of Finland 2. The genus *Phellinus. Karstenia*, 22, 27-42

Nejhad, G.M., & Dai, Y.C. (2007). The genus *Phellinus* s.l. (Basidiomycota) in Iran. *Mycotaxonomy*, 101, 201–222

Nest, M.A., Steenkamp, E.T., McTaggert, A.R., Trollip, C., Godlonton, T., Sauerman, E., Roodt, D., Naidoo, K., Coetzee, M.P.A., Wilken, P.M., Wingfield, M.J., & Wingfield, B.D. (2015). Saprophytic and pathogenic fungi in the Ceratocystidaceae differ in their ability to metabolize plant-derived sucrose. *BMC Evolutionary Biology*, 15, 273

Northolt, M.D., & Bullerman, L.B. (1982). Prevention of mold growth and toxin production through control of environmental condition. Journal of Food Protection, 6, 519-526

Olivain, C., & Alabouvette, C. (1997). Colonization of tomato root by a nonpathogenic strain of Fusarium oxysporum. New Phytologist, 137, 481-494

Oliveira, D., Silva, L.V., Silva, M.J., Goncalves, M.J., Cavaleiro, C., Salgueiro, L., & Pinto, E. (2012). Correlation of the chemical composition of essential oils from Origanum vulgare subsp. virens with their in vitro activity against pathogenic yeasts and filamentous fungi. Journal of Medical Microbiology, 61, 252-260

Packer, A., & Clay, K. (2000). Soil pathogens and spatial patterns of seedling mortality in a temperate tree. Nature, 404, 278-281

Papagianni, M. (2004). Fungal morphology and metabolite production in submerged mycelial processes. Biotechnology Advances, 22, 189 – 259

Park, I.H., Chung, S.K., Lee, K.B., Yoo, Y.C., Kim, S.K., Kim, G.S., & Song, K.S. (2004). An antioxidant hispidin from the mycelial cultures of Phellinus linteus. Archives of Pharmacal Research, 27, 615

Park, J.H., Choi, G.J., Lee, H.B., Kim, K.M., Jung, H.S., Lee, S.W., Jang, K.S., Cho, K.Y., and Kim J.C. (2004). Griseofulvin from Xylaria sp. Strain F0010, an Endophytic Fungus of Abies holophylla and its Antifungal Activity Against Plant Pathogenic Fungi. Journal of Microbiology and Biotechnology, 15, 112-117

Pazouki, M., & Panda, T. (2000). Understanding the morphology of fungi. Bioprocess Engineering, 22, 127-143

Pazzagli, L., Cappugi, G., Manao, G., Camici, G., Santinis, A., & Scala, A. (1999). Purification, Characterization, and Amino Acid Sequence of Ceratoplatanin, a New Phytotoxic Protein from Ceratocystis fimbriata f. sp. Platani. The Journal of Biological Chemistry, 274, 59-64

Pela'ez, F., Marti'nez, M.J., & Marti'nez, A.T. (1995) Screening of 68 species of Basidiomycetes for enzymes involved in lignin degradation. Mycology Research, 99, 37-42

Petrini, L., & Petrini, O. (1985) Xylariaceous fungi as endophytes. Sydowia, 38.216-234

Petrini, O. (1991). Fungal endophytic of tree leaves. Microbial Ecology of Leaves, 1, 179-197

Ponte, L.B., Astolfi, P., Reartes, D.S., Schmale, D.G., Moraes, M.G., & Ponte, E.M. (2009). Trichothecene mycotoxin genotypes of Fusarium graminearum sensu stricto and Fusarium meridionale inwheat from southern Brazil. Plant Pathology, 58, 344-351

Ralph, S., Byther, R., & Paul, H. (1974). Inhibition of sugarcane rooting by Ceratocystis paradoxa. Canadian Journal of Botany, 52, 761-766

69

Rabba, A.S., Vaidya, J.G., & Nanda, M.K. (1994). The genus *Phellinus* from Bhimashankar forest. *Biologia Indica*, 5, 47-56

Raymundo, T., Valenzuela, R., & Esqueda, M. (2009). The family Hymenochaetaceae from México 4. New records from Sierra de Álamos–Río Cuchujaqui biosphere reserve. *Mycotaxonomy*, 110, 387–398

Rodrigues, K.F., & Samuels, G.J. (1989). Studies in the genus *Phylacia* (Xylariaceace). *Memoirs of the new york botanical garden* 49:290-297

Rodrigues, K.F., & Samuel, G.J. (1990) Preliminary study of endophytic fungi in a tropical palm. *Mycological Research*, 94, 827-830

Rogers, J.D. (1979). *Xylaria* magnoliae sp.nov. and comments on several other fruit-inhabiting species. *Canadian Journal of Botany*, 57, 41-45

Rogers, J.D. (2000). Thoughts and musings on tropical Xylariaceae. *Mycology Research*, 104, 412-420

Rossman, A.Y. (1996). Morphological and molecular perspectives on systematics of the Hypocreales. *Mycologia*, 88, 1–19

Rossman, A.Y. (2000). Towards monophyletic genera in the holomorphic Hypocreales. *Studies in mycology*, 45, 27-34

Siameto, E., Okoth, S., Amugune, N., & Chege, N. (2010). Antagonism of Trichoderma farzianum isolates on soil borne plant pathogenic fungi from Embu District, Kenya. *Journal of Yeast and Fungal Research*, 1, 47-54

Sahashi, N., Akiba, M., Ishihara, M., Ota, Y., & Kanzaki, N. (2012). Brown root rot of trees caused by *Phellinus noxius* in the Ryukyu Islands, subtropical areas of Japan. *Forest Pathology*, 42,353-36

Ranadive, K., Japtap, N., & Vaidya, J. (2012). Host diversity of genus Phellinus from world Elixir Appl. *Botany*, 52, 402-408

Ryvarden, L. (2004). Neotropical Polyspores Part 1. Synopsis fungorum, 19, 1-229

Samuels, G.J., & Rossman, S.A. (1992). Thuemenella and Sarawakw. *Mycologla*, 84, 26-40

Santos, M.S., Ghini, R., Fernandes, B.V., & Silva, C.A. (2013). Increased carbon dioxide concentration in the air reduces the severity of *Ceratocystis* wilt in Eucalyptus clonal plantlets. *Australasian Plant Pathology*, 42, 595–599

Schubert, M., Fink, S., & Schwarze, W. (2008). Evaluation of Trichoderma spp. as a biocontrol agent against wood decay fungi in urban trees. *Biological Control*, 45, 111–123

Schulz, B., Boyle, C., Draeger, S., Rommert, A.K., & Krohn, K. (2002). Endophytic fungi: a source of novel biologically active secondary metabolites. *Mycology Research*, 106, 96–104

Schwarze, W. (2007). Wood decay under the microscope. *fungal biology reviews*, 21, 133–170

Schwarze, W., Jauss, F., Spencer, C., Hallam, C., & Schubert, M. (2012). Evaluation of an antagonistic Trichoderma strain for reducing the rate of wood decomposition by the white rot fungus Phellinus noxius. *Biological Control,* 61, 160–168

Sharma, G., & Pandey, R. (2010). Influence of culture media on growth, colony character and sporulation of fungi isolated from decaying vegetable wastes. *Journal of Yeast and Fungal Research*, 1, 157 – 164

Siameto, E., Okoth, S., Amugune, N., & Chege, N. (2010). Antagonism of *Trichoderma farzianum* isolates on soil borne plant pathogenic fungi from Embu District, Kenya. *Journal of Yeast and Fungal Research*, 1, 47-154

Sieber, V.T., & Hugentobler, C. (1987). Endophytische Pilze in Blättern und Ästen gesunder und geschädigter Buchen (Fagus sylvatica L.). *Mycotaxonomy*, 110, 387–398

Smith, S.N. (2007). An Overview of Ecological and Habitat Aspects in the Genus Fusarium with Special Emphasis on the SoilBorne Pathogenic Forms. *Plant Pathology Bulletin*, 16, 97-120

Smith, S., Smith, A., & Jakobsen, I. (2002). Mycorrhizal Fungi Can Dominate Phosphate Supply to Plants Irrespective of Growth Responses. *Plant Physiology*, 133, 16–20

Solarte, F., & Munoz, C.G. (2018). Diversity of *Neopestalotiopsis* and *Pestalotiopsis* spp., Causal Agents of Guava Scab in Colombia. *Plant disease*, 102, 49-59

Stadler, M., Kuhnert, E., Persoh, D., & Fournier, J. (2013). The Xylariaceae as model example for a unified nomenclature following the "One Fungus-One Name" (1F1N) concept. *Mycology An International Journal on Fungal Biology,* 4, 5-21

Takai, S. (1974). Pathogenicity and cerato-ulmin production in *Ceratocystis ulmi. Nature*, 252 124–126

Tan, R.X., & Zou, W.X. (2001) Endophytes: a rich source of functional metabolites. *Natural Products*, 18, 448–459

Thrane, U. (1990). Grouping Fusarium section discolour isolates by statistical analysiss of quantitive high performance liquid chromatographic data on secondary metabolite production. *Journal of Microbiological Methods*, 12, 23 - 39

Torres, U., Adams, P., Kamas, J.R., & Gubler, W. (2009). Identification, Incidence, and Pathogenicity of Fungal Species Associated with Grapevine Dieback in Texas. *American Journal of Enology Viticulture*, 60, 497-507

Tsai, J.N., Ann, P.J., Liou, R.F., Hsieh, W.H., & Ko, W.H (2017). *Phellinus noxius*: molecular diversity among isolates from Taiwan and its phylogenetic

relationship with other species of *Phellinus* based on sequences of the ITS region. *Botanical Studies*, 9, 1-11

Tudor, D., Robinson, S., & Cooper, P.A. (2013). The influence of pH on pigment formation by lignicolous fungi. *International Biodeterioration and Biodegradation*, 80, 22–28

Vaidya, J.G. (1987) - Ecological characteristic of wood decay and cord forming fungi from the campus of Poona University. *Poona University Press*, Pune, India,109

Wasantha, K., & Rawal, R.D. (2008). Influence of carbon, nitrogen, temperature and the pH on the growth and sporulation of some indian isolates of *Colletotrichum gloeosporioides* causing anthracnose disease of papaya (*Carrica papaya* L). *Tropical Agricultural Research and Extension*, 11, 1-6

Wheeler, K.A., & Hocking, A.D. (1993). Interactions among xerophilic fungi associated with dried salted fish. *J. Appl. Microbiology*, 74, 164–169

Wingfield, B., Van, M., Roos, H., & Wingfield, M. (2012). *Ceratocystis*: emergingevidence for discrete generic boundaries. In: Seifert, K.A., De Beer, Wingfield. M (Eds.), Ophiostomatoid Fungi: *Expanding Frontiers*, 1, 57–64

Whalley, A., & Edwards, R. (1987). Xylariaceous fungi: use of secondary metabolites. In: The Evolutionary Biology of Fungi (eds. A.D.M. Rayner, C.M. Brasier and D. Moore). *Cambridge University Press*, 1, 423-434

Whalley, A.J.S., & Edwards, R.L. (1995). Secondary metabolites and systematic arrangement within the Xylariaceae. *Canadian Journal of Botany*, 73, 802-810

Xu, L., Kusakari, S., Hosomi, A., Toyoda, H., & Ouchi, S., (1999). Postharvest diseases of grapes caused by Pestalotiopsis spp. *Annales Phytopathology Society of Japan*, 65, 305-311

Yang, Y., Hu, J.X., Liu, Y.F., Feng, N., Chen, H., Tang, Q.J., Ye, L.B., & Zhang, J.S. (2011). Antioxidant and Cytotoxic Activities of Ethanolic Extracts and Isolated Fractions of Species of the Genus *Phellinus* Quél. (Aphyllophoromycetideae). *International Journal of Medicinal Mushrooms*, 13, 145-152

Zhang, X.M., & Zhuang, W.Y. (2006). Phylogeny of some genera in the Nectriaceae (Hypocreales, Ascomycetes) inferred from 28S nrDNA partial sequences. *Mycosystema*, 25, 15-22

Zhao, S., Chen, S., Wang, B., Niu, S., Wu, W., Guo, L., & Che, Y. (2015). Four new tetramic acid and one new furanone derivatives from the plant endophytic fungus *Neopestalotiopsis* sp. *Fitoterapia*, 103, 106–112

Zivkovic, S., Stojanovic, S., Ivanovic, Z., Gavrilovic, V., Popovic, T., & Balaz, J. (2010). Screening of antagonistic activity of microorganisms against *Colletotrichum acutatum* and *Colletotrichum gloeosporioides*. *Archives of Biological Science Belgrade*, 62, 611-623