

# **UNIVERSITI PUTRA MALAYSIA**

# BIOLOGICAL PRETREATMENT OF OIL PALM EMPTY FRUIT BUNCH BY A LOCAL ISOLATE OF *Schizophyllum commune* ENN1 FOR PRODUCTION OF FERMENTABLE SUGARS

**ENIS NATASHA NOOR ARBAAIN** 

FBSB 2019 10



## BIOLOGICAL PRETREATMENT OF OIL PALM EMPTY FRUIT BUNCH BY A LOCAL ISOLATE OF *Schizophyllum commune* ENN1 FOR PRODUCTION OF FERMENTABLE SUGARS



By

ENIS NATASHA NOOR BINTI ARBAAIN

Thesis submitted to School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

December 2018

## COPYRIGHT

All materials contained within the thesis, including, without limitation of text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

## BIOLOGICAL PRETREATMENT OF OIL PALM EMPTY FRUIT BUNCH BY A LOCAL ISOLATE OF *Schizophyllum commune* ENN1 FOR PRODUCTION OF FERMENTABLE SUGARS

By

### ENIS NATASHA NOOR BINTI ARBAAIN

December 2018

Chairman : Ezyana Kamal Bahrin, PhD Faculty : Biotechnology and Biomolecular Sciences

Oil palm industry in Malaysia plays a major role in national socio-economic development. In line with the expansion of the industry, oil palm biomass is generated abundantly from the plantations and mills. Oil palm empty fruit bunch (OPEFB) is the most abundant lignocellulosic biomass generated from palm oil mill, composed of 25-44% cellulose, 25-28% hemicellulose and 19-27% lignin. The cellulose and hemicellulose components can be converted into fermentable sugars after being pretreated by either physical, chemical, physicochemical, biological or combination of these pretreatment methods. Currently, physicochemical pretreatment is the most common pretreatment method used to pretreat and convert OPEFB into fermentable sugars. However, this type of pretreatment utilised chemicals that lead to environmental issues and high operational cost. Therefore, biological pretreatment by fungi has been considered as an alternative to pretreat OPEFB as it is environmental friendly and requires low cost for the process. The first objective in this study was to evaluate the feasibility of Schizophyllum commune ENN1 in removing lignin of OPEFB through biological pretreatment. The second objective is to investigate the significant parameters affecting the biological pretreatment of OPEFB by S. commune ENN1 for fermentable sugars production through one-factor-at-a-time (OFAT) method.

The biological pretreatment by locally isolated fungus identified as *Schizophyllum commune* ENN1 was conducted using unwashed OPEFB without supplemented with nutrients or any moistening agents. The lignocellulosic compositional analysis showed that 53.8% of lignin was removed after biological pretreatment using *S. commune* ENN1 compared to 38.6% of lignin removal using *P. chrysosporium* UIA. The determination of

residual oil content showed that S. commune ENN1 was able to reduce the residual oil content by 85.3% while maintaining the moisture content in the range of 51-40%. The effect of incubation time (7-28 days), temperature (25-40°C) and amount of substrate (3-9 g) were analysed in the biological pretreatment. The results also showed the highest lignin removal of 55.2% after 14 days of incubation time. This is followed with significant lignin removal by 66% at temperature 30°C. Meanwhile, the amount of substrate at 5 g gives the highest lignin removal by 71.7%. A maximum lignin removal of 67.9% was achieved at optimum conditions using 5 g of substrate after 14 days of incubation time at temperature 30°C. The highest amount of reducing sugars obtained from biological pretreatment using S. commune ENN1 was 230.4 ± 0.19 mg/g with 54% of hydrolysis yield in 96 h. This amount is 1.8-fold the amount obtained from untreated OPEFB (128.2 ± 0.00 mg/g) with the hydrolysis yield of 35.17%. The finding from this study showed that S. commune ENN1 was feasible to remove the lignin of OPEFB through biological pretreatment for fermentable sugars production.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

### PRA-RAWATAN BIOLOGI TANDAN KOSONG KELAPA SAWIT OLEH Schizophyllum commune ENN1 PENCILAN TEMPATAN UNTUK PENGHASILAN GULA TERFERMENTASI

Oleh

#### ENIS NATASHA NOOR BINTI ARBAAIN

Disember 2018

## Pengerusi : Ezyana Kamal Bahrin, PhD Fakulti : Bioteknologi dan Sains Biomolekul

Industri kelapa sawit di Malaysia memainkan peranan utama dalam pembangunan sosioekonomi kebangsaan. Sejajar dengan perkembangan industri ini, biomas kelapa sawit dijana dengan banyak dari ladang dan kilang kelapa sawit. Tandan kosong kelapa sawit (OPEFB) adalah biomas lignoselulosa paling banyak dihasilkan daripada kilang minyak kelapa sawit yang terdiri daripada 25-44% selulosa, 25-28% hemiselulosa dan 19-27% lignin. Komponen selulosa dan hemiselulosa boleh ditukar kepada gula terfermentasi selepas diprarawat samaada secara fizikal, kimia, fizikokimia, biologi atau gabungan kaedah prarawatan ini. Pada masa kini, prarawatan fizikokimia adalah kaedah prarawatan yang paling biasa digunakan untuk prarawat dan menukar OPEFB menjadi gula terfermentasi. Walau bagaimanapun, jenis prarawatan yang menggunakan bahan kimia menyebabkan isu-isu alam sekitar dan kos operasi yang tinggi. Oleh itu, prarawatan biologi oleh kulat telah dianggap sebagai alternatif untuk prarawat OPEFB kerana ia mesra alam dan memerlukan kos yang rendah untuk proses tersebut. Objektif pertama kajian ini adalah untuk menilai kebolehan Schizophyllum commune ENN1 dalam menyingkirkan lignin OPEFB melalui prarawatan biologi. Objektif kedua adalah untuk mengkaji parameter penting yang memberi kesan dalam prarawatan biologi OPEFB menggunakan S. commune ENN1 untuk penghasilan gula terfermentasi berdasarkan kaedah satu faktor pada satu masa (OFAT).

Prarawatan biologi oleh kulat tempatan yang dipencilkan dikenali sebagai *Schizophyllum commune* ENN1 telah dijalankan menggunakan OPEFB yang tidak dibasuh tanpa ditambah dengan nutrien atau mana-mana agen pelembap. Analisis komposisi lignosellulosik menunjukkan bahawa 53.8%

lignin telah disingkirkan selepas prarawatan biologi menggunakan S. commune ENN1 berbanding 38.6% lignin disingkirkan menggunakan P. Penentuan kandungan sisa minyak menunjukkan chrysosporium UIA. bahawa S. commune ENN1 dapat mengurangkan kandungan sisa minyak sebanyak 85.3% dan pada masa yang sama mengekalkan kandungan lembapan dalam lingkungan 51-40%. Kesan masa inkubasi (7-28 hari), suhu (25-40°C) dan jumlah substrat (3-9 g) telah dianalisis dalam prarawatan biologi. Keputusan menunjukkan penyingkiran lignin tertinggi sebanyak 55.2% selepas 14 hari masa inkubasi. Ini diikuti oleh penyingkiran lignin yang besar sebanyak 66% pada suhu 30°C. Sementara itu, jumlah substrat pada 5 g memberikan penyingkiran lignin tertinggi sebanyak 71.7%. Penyingkiran lignin maksimum sebanyak 67.9% dicapai pada keadaan optimum menggunakan 5 g substrat selepas 14 hari masa inkubasi pada suhu 30°C. Jumlah gula penurun yang diperoleh daripada prarawatan biologi menggunakan S. *commune* ENN1 adalah 230.4 ± 0.19 mg/g dengan 54% hasil hidrolisis dalam 96 h. Jumlah ini adalah 1.8 kali ganda jumlah yang diperolehi daripada OPEFB yang tidak diprarawat (128.2 ± 0.00 mg/g) dengan hasil hidrolisis sebanyak 35.17%. Keputusan dari kajian ini menunjukkan bahawa S. commune ENN1 adalah kulat yang sesuai untuk menyingkirkan lignin OPEFB melalui prarawatan biologi untuk penghasilan gula terfermentasi.

## ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious and Merciful. *Alhamdulillah,* praise to Allah S.W.T. with all His blessing, mercy and guidance I am able to complete my research thesis entitled "Biological pretreatment of oil palm empty fruit bunch by a local isolate of *Schizophyllum commune* ENN1 for production of fermentable sugars" in fulfilment of the requirement for the degree of Master of Science (Environmental Biotechnology).

First of all, I would like to express my highest gratitude and appreciation to my main supervisor, Dr Ezyana Kamal Bahrin for guiding me throughout this study. All the ideas, knowledge and advice from her had been very useful for me to complete this study. This appreciation also goes to my co-supervisors, Prof. Dr Suraini Abd Aziz and Dr Mohamad Faizal Ibrahim for all their suggestions and helps for the entire period of study. I would like to thank Universiti Putra Malaysia and Ministry of Education for the funding and giving me such an opportunity to learn and widen my knowledge in the environmental biotechnology field. My special thanks to all the Environmental Biotechnology Group (EB Group) members for all their help, support and advice during this research.

I would like to convey my greatest gratitude to my family members, especially my parents, Mr Arbaain Rajali and Mrs Rabieah Husin for their never-ending trust and encouragement towards me from the start until the end of this study. Not forgotten to my brothers Faizal Hakimy and Farid Wahidi thank you for all the moral support. Last but not least, a lot of thanks to my friends Farhana, Nursyafiqah, Nabilah, Hazwani, Atiqah and Izzati for their help, advice and encouragement for me to complete this study.

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fullyowned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:

Date:

Name and Matric No.: Enis Natasha Noor Binti Arbaain (GS45781)

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: Name of Chairman of Supervisory Committee:	Dr Ezyana Kamal Bahrin
Signature: Name of Member of Supervisory Committee:	Professor Dr Suraini Abd. Aziz
Signature: Name of Member of Supervisory Committee:	Dr Mohamad Faizal Ibrahim

# TABLE OF CONTENTS

		i age
ABST ABS7 ACKN APPR DECL LIST LIST	TRACT TRAK NOWLEDGEMENTS ROVAL ARATION OF TABLES OF FIGURES OF ABBREVIATIONS	i iii v vi viii xii xii xiv xvi
CHAF 1	TER INTRODUCTION	1
2	<ul> <li>LITERATURE REVIEW</li> <li>2.1 Oil palm industry</li> <li>2.2 Oil palm empty fruit bunch (OPEFB)</li> <li>2.2.1 Physical characteristic of OPEFB</li> <li>2.2.2 Chemical composition of OPEFB</li> <li>2.2.3 Applications of OPEFB</li> <li>2.3 Biomass pretreatment</li> <li>2.3.1 Physical pretreatment</li> <li>2.3.2 Chemical pretreatment</li> <li>2.3.3 Physicochemical pretreatment</li> <li>2.4 Biological pretreatment</li> <li>2.4.1 Enzymatic pretreatment</li> <li>2.4.2 Fungal pretreatment</li> <li>2.5 Ligninolytic enzymes</li> <li>2.6 Fungi used in biological pretreatment</li> <li>2.6.3 Schizophyllum commune</li> <li>2.7 Biomass conversion into fermentable sugars</li> <li>2.8 Concluding remarks</li> </ul>	3 3 5 6 7 8 9 11 12 13 15 16 16 16 16 16 17 19 21 21 21 22 22 23
3	<ul> <li>MATERIALS AND METHODS</li> <li>3.1 General experimental design</li> <li>3.2 Microorganisms</li> <li>3.3 Chemicals and reagents</li> <li>3.4 Oil palm empty fruit bunch (OPEFB)</li> <li>3.5 Fungal isolation and identification <ul> <li>3.5.1 Fungal isolation</li> <li>3.5.2 DNA extraction of isolated fungus</li> <li>3.5.3 Identification of isolated fungus</li> </ul> </li> <li>3.6 Biological pretreatment of OPEFB</li> </ul>	24 24 25 26 26 27 27 27 29 29 30

Х

	3.7	Investigation of parameters for biologically pretreated OPEFB 3.7.1 Effect of incubation time 3.7.2 Effect of temperature	31 31 32
	3.8 3.9 3.10	<ul> <li>3.7.3 Effect of amount of substrate</li> <li>Biological pretreatment of OPEFB at optimum conditions</li> <li>Saccharification</li> <li>Analyses of OPEFB samples</li> <li>3.10.1 Extractive analysis</li> <li>3.10.2 Lignocellulosic composition</li> <li>3.10.3 Ash analysis</li> <li>3.10.4 Determination of residual oil content</li> <li>3.10.5 Estimation of moisture content</li> <li>3.10.6 Fourier transform infrared (FTIR)</li> </ul>	32 32 33 33 34 35 36 36 36
	3.11	3.10.7 Scanning electron microscope (SEM) Determination of enzymes activities 3.11.1 Ligninolytic enzymes assay	37 37 37 29
	3.12 3.13	Determination of reducing sugars Statistical analysis	38 41 41
4	<b>RESU</b> 4.1 4.2 4.3 4.4 4.4 4.5 4.6 <b>CONC</b>	Isolation and identification of isolated fungus Biological pretreatment of OPEFB 4.2.1 Residual oil content 4.2.2 Moisture content Characterisation of biological pretreated OPEFB 4.3.1 Lignocellulosic composition 4.3.2 Morphological observation using SEM 4.3.3 FTIR spectra analysis Investigation of parameters in biological pretreatment by <i>Schizophyllum commune</i> ENN1 4.4.1 Effect of incubation time 4.4.2 Effect of temperature 4.4.3 Effect of amount of substrate Biological pretreatment of OPEFB at optimum conditions Saccharification of biologically pretreated OPEFB	42 46 48 49 50 51 54 56 59 64 69 74 79 83
	5.1 5.2	Conclusions Recommendations	83 83 84
R A B LI	EFERENC PPENDICE IODATA O IST OF PU	ES ES F STUDENT BLICATIONS	85 107 118 119

## LIST OF TABLES

Table		Page
2.1	Annual biomass produced from oil palm industry in Malaysia	5
2.2	The chemical composition of oil palm empty fruit bunch	8
2.3	Substrate preparation process before pretreatment of OPEFB	11
2.4	List of wood decay fungi as lignin degrader for different biomass	s 20
3.1	Composition of M2 medium for Schizophyllum commune ENN1	28
3.2	Composition of nutrient used in biological pretreatment o <i>Phanerochaete crysosporium</i> UIA	f 31
3.3	Furnace temperature ramping program for ash analysis	36
4.1	List of fungal strains for identification of the isolated fungus	43
4.2	Comparison of composition of untreated and pretreated oil palmempty fruit bunch (OPEFB)	ו 53
4.3	Summary of FTIR wavenumber of untreated and pretreated OPEFB	d 58
4.4	Effect of incubation time on biological pretreatment of OPEFB by Schizophyllum commune ENN1	/ 60
4.5	Cellulase activities of biologically pretreated OPEFB by Schizophyllum commune ENN1 at different incubation time	/ 64
4.6	Effect of temperature on biological pretreatment of OPEFB by Schizophyllum commune ENN1	y 66
4.7	Cellulase activities of pretreated OPEFB by Schizophyllun commune ENN1 at different temperatures	n 69
4.8	Effect of amount of substrate on biological pretreatment o OPEFB by <i>Schizophyllum commune</i> ENN1	f 71
4.9	Cellulase activities of pretreated OPEFB by <i>Schizophyllun commune</i> ENN1 at different amount of substrates	n 74
4.10	Comparison of the chemical composition betweenuntreated and biologically pretreated OPEFB by <i>Schizophyllum commune</i> ENN1 at optimum conditions	d 9 75

C

- 4.11 Ligninolytic enzymes and cellulase activities of biologically pretreated OPEFB by *Schizophyllum commune* ENN1 at optimum conditions
- 4.12 Effect of different parameters in biological pretreatment from various lignocellulosic biomass on lignin removal
- 4.13 Enzymatic saccharification of biologically pretreated OPEFB
- 4.14 Comparison of sugar yield of biologically pretreated lignocellulosic biomass after saccharification



76

80

82

# LIST OF FIGURES

Figur	re F	Page
2.1	Biomass generated throughout the year from different industries in Malaysia	4
2.2	OPEFB pile in a palm oil mill	5
2.3	Flowchart of palm oil processing at palm oil mill	6
2.4	Physical structure of OPEFB after removal of fruitlets and its shredded form	7
2.5	Applications of OPEFB into various value-added products	9
2.6	General schematic diagram of the lignocellulosic structure before and after biological pretreatment	9
2.7	Fungal mycelia mat formation and penetration into the lignocellulosic structure	17
2.8	General reaction mechanism of laccase for phenol oxidation	18
2.9	Lignin peroxidase (LiP) catalyzed the oxidation of non-phenolic diarylpropane lignin model dimer	18
2.10	Catalytic cycle of manganese peroxidase	19
2.11	Schematic diagram of bioconversion lignocellulosic to biofuel	23
3.1	General experimental design of biological pretreatment of OPEFB by <i>Schizophyllum commune</i> ENN1	25
3.2	OPEFB preparation after sampling from the palm oil mill	27
3.3	Fungal colonies on the OPEFB piles	28
3.4	Inoculation of fungal agar plugs on OPEFB	30
3.5	Water and solvent extractive analyses using soxhlet extraction method	33
4.1	Morphological structure of Schizophyllum commune ENN1	42
4.2	Phylogenetic analysis based on ITS region 18S ribosomal RNA gene of isolated fungus and related fungi from the gene database of National Center for Biotechnology Information (NCBI)	45

4.3	Side and bottom views of biologically pretreated OPEFB by <i>Schizophyllum commune</i> ENN1	47
4.4	Residual oil content of untreated and biologically pretreated OPEFB by <i>Schizophyllum commune</i> ENN1	48
4.5	Moisture content of untreated and biologically pretreated OPEFB by <i>Schizophyllum commune</i> ENN1	50
4.6	Scanning electron microscope images of OPEFB samples	55
4.7	Fourier transform infrared (FTIR) spectra of OPEFB samples	57
4.8	Lignin peroxidase (LiP) and manganese peroxidase (MnP) activities at different incubation time of biological pretreatment by <i>Schizophyllum commune</i> ENN1	63
4.9	Lignin peroxidase (LiP) and manganese peroxidase (MnP) activities at different temperatures of biological pretreatment by <i>Schizophyllum</i> commune ENN1	68
4.10	Lignin peroxidase (LiP) and manganese peroxidase (MnP) activities at different amount of substrates of biologically pretreated by <i>Schizophyllum commune</i> ENN1	73

 $\bigcirc$ 

## LIST OF ABBREVIATIONS

- OPEFB Oil palm empty fruit bunch
- OPMF Oil palm mesocarp fibre
- PKS Palm kernel shell
- OPF Oil palm frond
- POME Palm oil mill effluent
- FFB Fresh fruit bunch
- GNI Gross national income
- RM Ringgit Malaysia
- CPO Crude palm oil
- OPT Oil palm trunk
- NaOH Sodium hydroxide
- KOH Potassium hydroxide
- CaOH<sub>2</sub> Calcium hydroxide
- NH<sub>4</sub>OH Aqueous ammonia
- AFEX Ammonia fibre explosion
- LWH Liquid hot water
- CO<sub>2</sub> Carbon dioxide
- WO Wet oxidation
- Lac Laccase
- LiP Lignin peroxidase
- MnP Manganese peroxidase
- LME Lignin modifying enzyme
- sp. Species

- NREL National Renewable Energy Laboratory
- PDA Potato dextrose agar
- SEM Scanning electron microscope
- FTIR Fourier transform infrared
- DNA Deoxyribonucleic acid
- FPU Filter paper unit
- DNS Dinitrosalicylic acid
- FPase Filter paper cellulase
- CMCase Carboxymethyl cellulase

### **CHAPTER 1**

#### INTRODUCTION

Malaysia is well known as the second largest palm oil producer and exporter in the world, which supplies 47% of world palm oil after Indonesia (Begum et al., 2013). In the year 2016 to 2020, Malaysian palm oil industry is expected to produce 15.4 million tonnes of palm oil each year (Abdullah & Sulaiman, 2013). The development of the Malaysian palm oil industry has led to the accumulation of biomass in the oil palm plantation and palm oil mill. This biomass includes oil palm empty fruit bunch (OPEFB), oil palm mesocarp fibre (OPMF), palm kernel shell (PKS) and oil palm frond (OPF) (Ying et al., 2014; Kamcharoen et al., 2014; Shariff et al., 2014). In order to address this issue, zero waste strategy has been promoted by utilising the biomass produced from the mill as a raw material for value-added products.

OPEFB is the most abundant biomass produced by palm oil mill with the total amount of 18 million tonnes per year compared to OPMF and PKS (Begum et al., 2013; Abdullah & Sulaiman, 2013). In the palm oil mill, fresh fruit bunch (FFB) undergoes steam sterilisation process and the fruits are stripped off from the bunch leaving behind the OPEFB (Law et al., 2007). Conventionally, the OPEFB was incinerated for steam and electricity generation at the palm oil mill and the ash was used as fertiliser. However, the OPEFB incineration caused incomplete combustion and released a huge amount of white smoke due to high moisture content (60%) of OPEFB (Chang, 2014; Abdullah et al., 2011). Nevertheless, OPEFB is also commonly used as soil mulch and compost in the plantation area (Geng, 2013). OPEFB is categorised as lignocellulosic biomass as it composed of carbohydrate polymers (cellulose, hemicellulose) and aromatic polymer (lignin). OPEFB consists of lignin 19-26%, hemicellulose 25-28% and cellulose 25-44% (Vandenbossche et al., 2014; Nomanbhay et al., 2013).

High cellulose content makes the OPEFB worth to be converted into fermentable sugars and other value-added products (Li et al., 2014). However, due to the intact structure of the lignocellulosic composition, the OPEFB must undergo the pretreatment process such as physical, chemical, biological and/or physicochemical pretreatment in order to alter the lignocellulosic structure (Nor et al., 2016). By combining the physical and chemical pretreatments, the physicochemical pretreatment is considered to have the highest reliability to enhance the digestibility of lignocellulosic biomass. Nonetheless, physicochemical pretreatment involves high cost of equipment and high energy requirement (Brodeur et al., 2011). The use of the chemical in the pretreatment process also leads to environmental concerns within a community (Agbor et al., 2011).



Biological pretreatment is an environmentally friendly method as it offers a mild pretreatment condition compared to another type of biomass pretreatment. Besides, this pretreatment also considered as low-cost and low energy consumption process (Mood et al., 2013). The biological pretreatment involves enzymatic action to partially degrade the lignin and expose the cellulose structure (Isroi et al., 2011). This pretreatment involved the action of fungi that are able to produce ligninolytic enzymes for lignin degradation. The white-rot fungi have been reported to be the most effective fungi for biological pretreatment (Agbor et al., 2011).

Commonly, OPEFB needs to undergo substrate preparation process before any pretreatments to remove oil residue on the surface of untreated OPEFB. The presence of oil residue on the surface of untreated OPEFB may inhibit the growth of non-indigenous fungus. However, the indigenous fungus found on the pile of OPEFB naturally grown on the oily surface of OPEFB. Thus, the biological pretreatment using indigenous fungus (*Schizophyllum commune* ENN1) is a new approach to pretreat the OPEFB by omitting the substrate preparation process and nutrient supplied throughout the biological pretreatment.

To the best of our knowledge, there are limited literature studies on biological pretreatment of OPEFB using *Schizophyllum commune*. The factors such as incubation time, temperature and amount of substrate are crucial factors in biological pretreatment that could influence the efficiency of lignin removal during the pretreatment. Furthermore, a different type of fungi and substrates used in biological pretreatment may have a different performance at favourable conditions. Therefore, optimum conditions for biological pretreatment of OPEFB was investigated in this study in order to obtain pretreated OPEFB that is suitable as a feedstock for fermentable sugars production.

The objectives of this study are:

- 1. To evaluate the feasibility of *Schizophyllum commune* ENN1 for lignin removal of OPEFB through biological pretreatment.
- 2. To investigate the effects of incubation time, temperature and amount of substrate in biological pretreatment of OPEFB using *Schizophyllum commune* ENN1 for fermentable sugars production.

#### REFERENCES

- Abdul, P. M., Jahim, J. M., Harun, S., Markom, M., Lutpi, N. A., Hassan, O., Balan, V., Bale, B. E., Mohd Nor, M. T. (2016). Effects of changes in chemical and structural characteristic of ammonia fibre expansion (AFEX) pretreated oil palm empty fruit bunch fibre on enzymatic saccharification and fermentability for biohydrogen. *Bioresource Technology*, 211, 200–208.
- Abdullah, J. J., & Greetham, D. (2016). Optimizing cellulase production from municipal solid waste (MSW) using solid state fermentation (SSF). *Journal of Fundamentals of Renewable Energy and Applications*, *6*(3).
- Abdullah, N., & Gerhauser, H. (2008). Bio-oil derived from empty fruit bunches. *Fuel*, *87*(12), 2606–2613.
- Abdullah, N., & Sulaiman, F. (2013). The oil palm wastes in Malaysia. In Biomass Now – Sustainable Growth and Use, 75–100.
- Abdullah, N., & Sulaiman, F. (2013). The properties of the washed empty fruit bunches of oil palm. *Journal of Physical Science*, *24*(2), 117–137.
- Abdullah, N., Sulaiman, F., & Gerhauser, H. (2011). Characterisation of oil palm empty fruit bunches for fuel application. *Journal of Physical Science*, 22(1), 1–24.
- Abdullah, R. (2011). World palm oil supply, demand, price and prospects: Focus on Malaysian and Indonesian palm oil industries. *Oil Palm Industry Economic Journal*, 11(2), 13–25.
- Abu Bakar, N. K., Zanirun, Z., Abd-Aziz, S., Ghazali, F. M., & Hassan, M. A. (2012). Production of fermentable sugars from oil palm empty fruit bunch using crude cellulase cocktails with *Trichoderma asperellum* UPM1 and *Aspergillus fumigatus* UPM2 for bioethanol production. *BioResources*, 7(3), 3627–3639.
- Adejoye, O. D., Adebayo-Tayo, B. C., Ogunjobi, A. A., & Afolabi, O. O. (2007). Physicochemical studies on *Schizophyllum commune* (Fries) a Nigerian edible fungus. *World Applied Sciences Journal*, 2(1), 73–76.
- Adejoye, O. D., & Fasidi, I. O. (2010). Effect of cultural conditions on biomass and laccase production in submerged medium by *Schizophyllum commune* (Fries), a Nigerian edible mushroom. *Electronic Journal of Environmental, Agricultural and Food Chemistry*, 9(3), 600–609.
- Adenipekun, C. O., & Lawal, R. (2012). Uses of mushrooms in bioremediation: A review. *Biotechnology and Molecular Biology Reviews*, *7*(3), 62–68.

- Aftab, K., Akhtar, K., Kausar, A., Khaliq, S., Nisar, N., Umbreen, H., & Iqbal, M. (2017). Fungal strains isolation, identification and application for the recovery of Zn(II) ions. *Journal of Photochemistry and Photobiology B: Biology*, *175*(August), 282–290.
- Afzanizam, N., Nazri, M., Jaafar, M., Tung, C., & Jo-han, N. (2015). A review of palm oil biomass as a feedstock for syngas fuel technology. *Jurnal Teknologi (Sciences and Engineering)*, 5, 13–18.
- Agbor, V. B., Cicek, N., Sparling, R., Berlin, A., & Levin, D. B. (2011). Biomass pretreatment: Fundamentals toward application. *Biotechnology Advances*, *29*(6), 675–685.
- Akhtar, J., Idris, A., Teo, C. L., Lai, L. W., Hassan, N., & Khan, M. I. (2014). Comparison of delignification of oil palm empty fruit bunch (EFB) by microwave assisted alkali/acid pretreatment and conventional pretreatment method. *International Journal of Advances in Chemical Engineering & Biological Sciences*, 1(2), 155–157.
- Akindoyo, J. O., Ghazali, S. B., & Beg, M. D. H. (2015). Structural and thermal properties of ultrasound treated oil palm empty fruit bunch (OPEFB) fiber. *Proceedings of Tenth ThellER International Conference*, (February), 60–64.
- Al-mohanna, M. T. (2017). Methods for fungal enumeration, isolation and identification, (May 2016).
- Alam, F. A. S. A., Er, A. C., & Begum, H. (2015). Malaysian oil palm industry: prospect and problem. *Journal of Food, Agriculture and Environment*, *13*(2), 143–148.
- Alam, N., Cha, Y. J., Shim, M. J., Lee, T. S., & Lee, U. Y. (2010). Cultural conditions for mycelial growth and molecular phylogenetic relationship in different wild strains of *Schizophyllum commune*. *Mycobiology*, 38(1), 17.
- Alexandropoulou, M., Antonopoulou, G., Fragkou, E., Ntaikou, I., & Lyberatos, G. (2017). Fungal pretreatment of willow sawdust and its combination with alkaline treatment for enhancing biogas production. *Journal of Environmental Management*, 203, 704–713.
- Altschul, S. F., Madden, T. L., Schäffer, A. A., Zhang, J., Zhang, Z., Miller, W., & Lipman, D. J. (1997). Gapped BLAST and PSI-BLAST: A new generation of protein database search programs. *Nucleic Acids Research*, 25(17), 3389–3402.
- Alvira, P., Tomás-Pejó, E., Ballesteros, M., & Negro, M. J. (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: A review. *Bioresource Technology*, 101(13), 4851–4861.

- Ang, S. K., Shaza, E. M., Adibah, Y. A., Suraini, A. A., & Madihah, M. S. (2013). Production of cellulases and xylanase by *Aspergillus fumigatus* SK1 using untreated oil palm trunk through solid state fermentation. *Process Biochemistry*, 48(9), 1293–1302.
- Annuar, M. S. M., Sammantha, S., Murthy, & Sabanatham, V. (2010). Laccase production from oil palm industry solid waste: Statistical optimization of selected process parameters. *Engineering in Life Sciences*, 10(1), 40– 48.
- Arantes, V., Silva, E. M., & Milagres, A. M. F. (2011). Optimal recovery process conditions for manganese-peroxidase obtained by solid-state fermentation of eucalyptus residue using *Lentinula edodes*. *Biomass and Bioenergy*, 35(9), 4040–4044.
- Arora, A., Priya, S., Sharma, P., Sharma, S., & Nain, L. (2016). Evaluating biological pretreatment as a feasible methodology for ethanol production from paddy straw. *Biocatalysis and Agricultural Biotechnology*, 8, 66–72.
- Arora, D. S., Chander, M., & Gill, P. K. (2002). Involvement of lignin peroxidase, manganese peroxidase and laccase in degradation and selective ligninolysis of wheat straw. *International Biodeterioration and Biodegradation*, 50(2), 115–120.
- Arora, D. S., & Gill, P. K. (2001). Comparison of two assay procedures for lignin peroxidase. *Enzyme and Microbial Technology*, *28*(7–8), 602– 605.
- Asgher, M., Irshad, M., & Iqbal, H. M. N. (2012). Purification and characterization of LiP produced by *Schizophyllum commune* IBL-06 using banana stalk in solid state cultures. *BioResources*, 7(3), 4012– 4021.
- Asgher, M., Khan, S. W., & Bilal, M. (2016). Optimization of lignocellulolytic enzyme production by *Pleurotus eryngii* WC 888 utilizing agro-industrial residues and bio-ethanol production. *Romanian Biotechnological Letters*, *21*(1), 11133–11143.
- Asgher, M., Wahab, A., Bilal, M., & Nasir Iqbal, H. M. (2016). Lignocellulose degradation and production of lignin modifying enzymes by *Schizophyllum commune* IBL-06 in solid-state fermentation. *Biocatalysis and Agricultural Biotechnology*, 6, 195–201.
- Awalludin, M. F., Sulaiman, O., Hashim, R., & Nadhari, W. N. A. W. (2015). An overview of the oil palm industry in Malaysia and its waste utilization through thermochemical conversion, specifically via liquefaction. *Renewable and Sustainable Energy Reviews*, *50*, 1469–1484.

- Badiei, M., Asim, N., Jahim, J. M., & Sopian, K. (2014). Comparison of chemical pretreatment methods for cellulosic biomass. Asia Pacific Chemical Biological Environmental Engineering Society Procedia, 9(2013), 170–174.
- Baharuddin, A. S., Asma, N., Razak, A., Aini, N., & Rahman, A. (2009). Biocoversion of oil palm empty fruit bunch by *Aspergillus niger* EB4 under solid-state fermentation, *32*(12), 143–151.
- Baharuddin, A. S., Sulaiman, A., Kim, D. H., Mokhtar, M. N., Hassan, M. A., Wakisaka, M., Shirai, Y., Nishida, H. (2013). Selective component degradation of oil palm empty fruit bunches (OPEFB) using highpressure steam. *Biomass and Bioenergy*, 55, 268–275.
- Baharuddin, A. S., Yunos, N. S. H., Mahmud, N. A. N., Zakaria, R., & Yunos, K. F. (2012). Effect of high-pressure steam treatment on enzymatic saccharification of oil palm empty fruit bunches. *BioResources*, 7(3), 3525–3538.
- Begerow, D., Nilsson, H., Unterscher, M., & Maier, W. (2010). Current state and perspectives of fungal DNA barcoding and rapid identification procedures. *Applied Microbiology and Biotechnology*, 87(1), 99–108.
- Begum, S., Kumaran, P., & Jayakumar, M. (2013). Use of oil palm waste as a renewable energy source and its impact on reduction of air pollution in context of Malaysia. *IOP Conference Series: Earth and Environmental Science*, *16*, 012026.
- Behera, S., Arora, R., Nandhagopal, N., & Kumar, S. (2014). Importance of chemical pretreatment for bioconversion of lignocellulosic biomass. *Renewable and Sustainable Energy Reviews*, *36*, 91–106.
- Bhutto, A. W., Qureshi, K., Harijan, K., Abro, R., Abbas, T., Bazmi, A. A., Karim, S., Yu, G. (2017). Insight into progress in pre-treatment of lignocellulosic biomass. *Energy*, 122, 724–745.
- Bourbonnais, R., & Paice, M. G. (1988). Veratryl alcohol oxidases from the lignin-degrading basidiomycete *Pleurotus sajor-caju*. *The Biochemical Journal*, *255*(2), 445–450.
- Brodeur, G., Yau, E., Badal, K., Collier, J., Ramachandran, K. B., & Ramakrishnan, S. (2011). Chemical and physicochemical pretreatment of lignocellulosic biomass: A review. *Enzyme Research*, 2011, 787532.
- Bukhari, N. A., Abu Bakar, N., Loh, S. K., & Choo, Y. M. (2014). Bioethanol production by fermentation of oil palm empty fruit bunches pretreated with combined chemicals. *Journal Applied Environmental and Biological Sciences*, *4*(10), 234–242.

- Burhani, D., Mauliva, A., Putri, H., Waluyo, J., Nofiana, Y., & Sudiyani, Y. (2017). The effect of two-stage pretreatment on the physical and chemical characteristic of oil palm empty fruit bunch for bioethanol production. *Proceedings of the 3rd International Symposium on Applied Chemistry*, 1904, 1–14.
- Capolupo, L., & Faraco, V. (2016). Green methods of lignocellulose pretreatment for biorefinery development. *Applied Microbiology and Biotechnology*, *100*(22), 9451–9467.
- Chang, S. H. (2014). An overview of empty fruit bunch from oil palm as feedstock for bio-oil production. *Biomass and Bioenergy*, *62*, 174–181.
- Chaturvedi, V., & Verma, P. (2013). An overview of key pretreatment processes employed for bioconversion of lignocellulosic biomass into biofuels and value added products. *3 Biotech*, *3*(5), 415–431.
- Chen, H., Liu, J., Chang, X., Chen, D., Xue, Y., Liu, P., Lin, H., Han, S. (2017). A review on the pretreatment of lignocellulose for high-value chemicals. *Fuel Processing Technology*, *160*, 196–206.
- Cheng, C. K., & Ismail, K. S. K. (2007). Production of bioethanol from oil palm empty fruit bunch. *International Conference on Sustainable Material*, (September), 69–72.
- Cher, Y., Stevens, M., Holmes, J., & Xu, H. (2013). Understanding the alkaline pretreatments parameters of corn stover enzymatic saccarification. *Biotechnology Biofuels*, 6(1), 8.
- Chiesa, S., & Gnansounou, E. (2014). Use of empty fruit bunches from the oil palm for bioethanol production: A thorough comparison between dilute acid and dilute alkali pretreatment. *Bioresource Technology*, *159*, 355–364.
- Colak, A., Kolcuoglu, Y., Sesli, E., & Dalman, O. (2007). Biochemical composition of some turkish fungi. *Asian Journal of Chemistry*, *19*(3), 2193–2199.
- Corley, R., & Tinker, P. (2003). The products of the oil palm and their extraction. *The Oil Palm*, 445–466.
- Corley, R. H. V, & Thinker, P. B. (2003). The origin and development of the oil palm industry. *The Oil Palm*, *4*(1), 1–26.
- Cruz Ramírez, M. G., Rivera-Ríos, J. M., Téllez-Jurado, A., Maqueda Gálvez, A. P., Mercado-Flores, Y., & Arana-Cuenca, A. (2012). Screening for thermotolerant ligninolytic fungi with laccase, lipase, and protease activity isolated in Mexico. *Journal of Environmental Management*, 95, 256–259.

- Dashtban, M., Schraft, H., Syed, T. A., & Qin, W. (2010). Fungal biodegradation and enzymatic modification of lignin. *International Journal of Biochemistry and Molecular Biology*, *1*(1), 36–50.
- Deswal, D., Gupta, R., Nandal, P., & Kuhad, R. C. (2014). Fungal pretreatment improves amenability of lignocellulosic material for its saccharification to sugars. *Carbohydrate Polymers*, *99*, 264–269.
- Duangwang, S., & Sangwichien, C. (2012). Optimizing alkali pretreatment of oil palm empty fruit bunch for ethanol production by application of response surface methodology. *Advanced Materials Research*, 622– 623(May), 117–121.
- Dungani, R., Jawaid, M., Khalil, H. P. S. A., Jasni, Aprilia, S., Hakeem, K. R., Hartati, S., Islam, M. N. (2013). A review on quality enhancement of oil palm trunk. *BioResources*, 8(2), 3136–3156.
- Duque, A., Manzanares, P., & Ballesteros, M. (2017). Extrusion as a pretreatment for lignocellulosic biomass: fundamentals and applications. *Renewable Energy*, *114*, 1427–1441.
- Elisashvili, V., Kachlishvili, E., & Penninckx, M. (2008). Effect of growth substrate, method of fermentation, and nitrogen source on lignocellulose-degrading enzymes production by white-rot basidiomycetes. *Journal of Industrial Microbiology and Biotechnology*, *35*(11), 1531–1538.
- Fatriasari, W., Syafii, W., Wistara, N. J., Syamsu, K., & Prasetya, B. (2014). The characteristic changes of betung bamboo (*Dendrocalamus asper*) pretreated by fungal pretreatment. *International Journal of Renewable Energy Development (IJRED)*, 3(2), 133–143.
- Fernández-Fueyo, E., Castanera, R., Ruiz-Dueñas, F. J., López-Lucendo, M. F., Ramírez, L., Pisabarro, A. G., & Martínez, A. T. (2014). Ligninolytic peroxidase gene expression by *Pleurotus ostreatus*: Differential regulation in lignocellulose medium and effect of temperature and pH. *Fungal Genetics and Biology*, 72, 150–161.
- Ferrer, A., Vega, A., Ligero, P., & Rodríguez, A. (2011). Pulping of empty fruit bunches (EFB) from the palm oil industry by formic acid. *BioResources*, *6*(4), 4282–4301.
- Fillat, Ú., Ibarra, D., Eugenio, M., Moreno, A., Tomás-Pejó, E., & Martín-Sampedro, R. (2017). Laccases as a potential tool for the efficient conversion of lignocellulosic biomass: A review. *Fermentation*, 3(2), 17.
- Geng, A. (2013). Conversion of oil palm empty fruit bunch to biofuels. In *Liquid, gaseous and solid biofuels* (pp. 479–490).

- Ghaffar, S. H., Fan, M., & McVicar, B. (2015). Bioengineering for utilisation and bioconversion of straw biomass into bio-products. *Industrial Crops and Products*, *77*, 262–274.
- Gomez, J. C., Mokhtar, M. N., Sulaiman, A., Zakaria, R., Baharuddin, A. S., & Busu, Z. (2015a). Study on residual oil recovery from empty fruit bunch by combination of water and steam process. *Journal of Food Process Engineering*, 38(4), 385–394.
- Gonçalves, I., Silva, C., & Cavaco-Paulo, A. (2015). Ultrasound enhanced laccase applications. *Green Chemistry*, *17*(3), 1362–1374.
- Grivell, A. R., & Jackson, J. F. (1969). Microbial culture preservation with silica gel. *Journal of General Microbiology*, *58*(1969), 423–425.
- Gupta, V. K., & Tuohy, M. G. (2014). Progress in physical and chemical pretreatment of lignocellulosic biomass. *Biofuel Technologies: Recent Developments*.
- Haghighi, M. S., Hossein, G. A., Tabatabaei, M., Salehi, J. G., Najafi, G. H., Gholami, M., & Ardjmand, M. (2013). Lignocellulosic biomass to bioethanol, a comprehensive review with a focus on pretreatment. *Renewable and Sustainable Energy Reviews*, 27.
- Hall, T. A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*.
- Hama, S., Nakano, K., Onodera, K., Nakamura, M., Noda, H., & Kondo, A. (2014). Saccharification behavior of cellulose acetate during enzymatic processing for microbial ethanol production. *Bioresource Technology*, *157*, 1–5.
- Hamisan, A. F., Abd-Aziz, S., Kamaruddin, A. H., Md. Shah, U. K., Shahab, N., & Hassan, M. A. (2009). Delignification of oil palm empty fruit bunch using chemical and microbial pretreatment methods.pdf. *International Journal of Agricultural Research*.
- Hanafusa, Y., Hirano, Y., Watabe, H., Hosaka, K., Ikezawa, M., & Shibahara, T. (2016). First isolation of Schizophyllum commune in a harbor seal (*Phoca vitulina*). *International Society for Human and Animal Mycoloy*, 54(March), 492–499.
- Harmsen, P., Huijgen, W., Bermudez, L., & Bakker, R. (2010). Literature review of physical and chemical pretreatment processes for lignocellulosic biomass. *Energy*, (9), 1–49.
- Harun, N. A. F., Baharuddin, A. S., Zainudin, M. H. M., Bahrin, E. K., Naim, M. N., & Zakaria, R. (2013). Cellulase production from treated oil palm empty fruit bunch degradation by locally isolated *Thermobifida fusca*.

*BioResources*, *8*(1), 676–687.

- Hassan, N. S., & Badri, K. (2016). Thermal behaviors of oil palm empty fruit bunch fiber upon exposure to acid-base aqueous solutions. *Malaysian Journal of Analytical Sciences*, *20*(5), 1095–1103.
- Hassan, O., Ling, T. P., Maskat, M. Y., Illias, R. M., Badri, K., Jahim, J., & Mahadi, N. M. (2013). Optimization of pretreatments for the hydrolysis of oil palm empty fruit bunch fiber (EFBF) using enzyme mixtures. *Biomass and Bioenergy*, 56(0), 137–146.
- Hassan, S. S., Williams, G. A., & Jaiswal, A. K. (2018). Emerging technologies for the pretreatment of lignocellulosic biomass. *Bioresource Technology*, 262(March), 310–318.
- Hermiati, E., Anita, S. H., Risanto, L., Styarini, D., Sudiyani, Y., Hanafi, A., & Abimanyu, H. (2013). Biological pretreatment of oil palm frond fiber using white-rot fungi for enzymatic saccharification. *MAKARA Journal* of *Technology Series*, 17(1), 39–43.
- Hofricher, M. (2002). Review: Lignin conversiton by manganese peroxidase (MnP). *Enzyme and Microbial Technology*, *30*, 12.
- Horisawa, S., Ando, H., Ariga, O., & Sakuma, Y. (2015). Direct ethanol production from cellulosic materials by consolidated biological processing using the wood rot fungus Schizophyllum commune. *Bioresource Technology*, 197, 37–41.
- Hosseini, S. E., & Wahid, M. A. (2014). Utilization of palm solid residue as a source of renewable and sustainable energy in Malaysia. *Renewable and Sustainable Energy Reviews*, *40*, 621–632.
- Ibrahim, M. F., Abd-Aziz, S., Yusoff, M. E. M., Phang, L. Y., & Hassan, M. A. (2015). Simultaneous enzymatic saccharification and ABE fermentation using pretreated oil palm empty fruit bunch as substrate to produce butanol and hydrogen as biofuel. *Renewable Energy*, 77, 447–455.
- Idris, J. (2013). Study on biochar production from empty fruit bunch biomass under self-sustained carbonization for the development of yamasen carbonization oven. *Journal of Chemical Information and Modeling*, *53*(9), 1689–1699.
- Iqbal, H. M. N., Asgher, M., & Bhatti, H. N. (2011). Optimization of physical and nutritional factors for synthesis of lignin degrading enzymes by a novel strain *Trametes versicolor*, *6*, 1273–1287.
- Ishmael, U. C., Shah, S. R., Palliah, J. V., Asras, M. F. F., Ahmad, S. S., and Ayodele Bamidele, V. (2016). Statistical modeling and optimization of enzymatic pretreatment of empty fruit bunches with laccase. *BioResources*, *11*(2), 5013–5032.

- Ishmael, U. C., Shah, S. R., Palliah, J. V., Asras, M. F. F., Ahmad, S. S. B. N. W., & Ayodele, V. B. (2016). Statistical modeling and optimization of enzymatic pretreatment of empty fruit bunches with laccase enzyme. *BioResources*, *11*(2), 5013–5032.
- Ishola, M. M., Isroi, & Taherzadeh, M. J. (2014a). Effect of fungal and phosphoric acid pretreatment on ethanol production from oil palm empty fruit bunches (OPEFB). *Bioresource Technology*, 165, 9–12.
- Ishola, M. M., Isroi, & Taherzadeh, M. J. (2014b). Effect of fungal and phosphoric acid pretreatment on ethanol production from oil palm empty fruit bunches (OPEFB). *Bioresource Technology*, *165*(C), 9–12.
- Isroi, Ishola, M. M., Millati, R., Syamsiah, S., Cahyanto, M. N., Niklasson, C., & Taherzadeh, M. J. (2012). Structural changes of oil palm empty fruit bunch (OPEFB) after fungal and phosphoric acid pretreatment. *Molecules*, 17(12), 14995–15012.
- Isroi, Millati, R., Syamsiah, S., Niklasson, C., Cahyanto, M. N., Lundquist, K., & Taherzadeh, M. J. (2011). Biological pretreatment of lignocelluloses with white-rot fungi and its applications: A review. *BioResources*, 6(4), 5224–5259.
- Iwamoto, S., Abe, K., & Yano, H. (2008). The effect of hemicelluloses on wood pulp nanofibrillation and nanofiber network characteristics. *Biomacromolecules*, 9(3), 1022–1026.
- Janusz, G., Kucharzyk, K. H., Pawlik, A., Staszczak, M., & Paszczynski, A. J. (2013). Fungal laccase, manganese peroxidase and lignin peroxidase: Gene expression and regulation. *Enzyme and Microbial Technology*, *52*(1), 1–12.
- Jones, M. P., Jones, M., Huynh, T., Dekiwadia, C., Daver, F., & John, S. (2017). Mycelium Composites : A review of engineering characteristics and growth kinetics, *11*, 241–257.
- Kadimaliev, D. A., Revin, V. V., Atykyan, N. A., & Samuilov, V. D. (2003). Effect of wood modification on lignin consumption and synthesis of lignolytic enzymes by the fungus *Panus (Lentinus) tigrinus*. *Applied Biochemistry and Microbiology*, *39*(5), 488–492.
- Kam, Y. C., Hii, S. L., Sim, C. Y. Y., & Ong, L. G. A. (2016). Schizophyllum commune lipase production on pretreated sugarcane bagasse and its effectiveness. International Journal of Polymer Science, 2016.
- Kamalrudin, M. S., & Abdullah, R. (2014). Malaysian palm oil Moving ahead to sustainable production growth. *Oil Palm Industry Economic Journal*, *14*(1), 24–33.

- Kamcharoen, A., Champreda, V., Eurwilaichitr, L., & Boonsawang, P. (2014). Screening and optimization of parameters affecting fungal pretreatment of oil palm empty fruit bunch (EFB) by experimental design. *International Journal of Energy and Environmental Engineering*, *5*(4), 303–312.
- Kamil, N. N., & Omar, S. F. (2016). Climate variability and its impact on the palm oil industry. *Oil Palm Industry Economic Journal*, *16*(March).
- Kamoldeen, A. A., Lee, C. K., Wan Abdullah, W. N., & Leh, C. P. (2017). Enhanced ethanol production from mild alkali-treated oil-palm empty fruit bunches via co-fermentation of glucose and xylose. *Renewable Energy*, 107, 113–123.
- Karimi, K., & Taherzadeh, M. J. (2016). A critical review of analytical methods in pretreatment of lignocelluloses: Composition, imaging, and crystallinity. *Bioresource Technology*, 200, 1008–1018.
- Karp, S. G., Faraco, V., Amore, A., Alberto, L., Letti, J., Soccol, V. T., & Soccol, C. R. (2015). Statistical optimization of laccase production and delignification of sugarcane bagasse by *Pleurotus ostreatus* in solid-state fermentation. *Biomedical Research International*, 2015, 1–9.
- Khalil, H. P. S. A., Alwani, M. S., Ridzuan, R., Kamarudin, H., & Khairul, A. (2008). Chemical composition, morphological characteristics, and cell wall structure of Malaysian oil palm fibers. *Polymer - Plastics Technology and Engineering*, 47(3), 273–280.
- Kim, C.-H., Kim, D.-S., Sung, Y. J., Hong, H.-E., & Kim, S.-B. (2013). Evaluation of defiberation by organosolv ethanolamine pulping for integral utilization of oil palm EFB. *Journal of Korea Technical Association of The Pulp and Paper Industry*, 45(1), 67–74.
- Kirker, G. T., Blodgett, A. B., Arango, R. A., Lebow, P. K., & Clausen, C. A. (2013). The role of extractives in naturally durable wood species. *International Biodeterioration and Biodegradation*, *82*, 53–58.
- Kong, W., Chen, H., Lyu, S., Ma, F., Yu, H., & Zhang, X. (2016). Characterization of a novel manganese peroxidase from white-rot fungus *Echinodontium taxodii* 2538, and its use for the degradation of lignin-related compounds. *Process Biochemistry*, *51*(11), 1776–1783.
- Konsomboon, S., Pipatmanomai, S., Madhiyanon, T., & Tia, S. (2011). Effect of kaolin addition on ash characteristics of palm empty fruit bunch (EFB) upon combustion. *Applied Energy*, *88*(1), 298–305.
- Krishania, M., Kumar, V., Vijay, V. K., & Malik, A. (2012). Opportunities for improvement of process technology for biomethanation processes. *Green Processing and Synthesis*, *1*(1).

- Kumar, N. M., Ravikumar, R., Thenmozhi, S., & Kirupa Sankar, M. (2017). Development of natural cellulase inhibitor mediated intensified biological pretreatment technology using *Pleurotus florida* for maximum recovery of cellulose from paddy straw under solid state condition. *Bioresource Technology*, 244(May), 353–361.
- Kumar, N. S., & Min, K. (2011). Phenolic compounds biosorption onto Schizophyllum commune fungus: FTIR analysis, kinetics and adsorption isotherms modeling. Chemical Engineering Journal, 168(2), 562–571.
- Kumar, P., Barrett, D. M., Delwiche, M. J., & Stroeve, P. (2009). Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. *Industrial and Engineering Chemistry Research*, 48(8), 3713–3729.
- Lai, L. W., & Idris, A. (2013). Disruption of oil palm trunks and fronds by microwave-alkali pretreatment. *BioResources*, *8*(2), 2792–2804.
- Lalak, J., Kasprzycka, A., Martyniak, D., & Tys, J. (2016). Effect of biological pretreatment of *Agropyron elongatum* "BAMAR" on biogas production by anaerobic digestion. *Bioresource Technology*, *200*, 194–200.
- Law, K.-N., Daud, W. R. W., & Ghazali, A. (2007). Morphological and chemical nature of fiber strands of oil palm empty-fruit-bunch (OPEFB). *BioResources*, 2(3), 351–362.
- Lee, D. H., Back, C. G., Win, N. K. K., Choi, K. H., Kim, K. M., Kang, I. K., ... Jung, H. Y. (2011). Biological characterization of *marssonina coronaria* associated with apple blotch disease. *Mycobiology*, *39*(3), 200–205.
- Lee, H., Lim, W., & Lee, J. (2015). Improvement of ethanol fermentation from lignocellulosic hydrolysates by the removal of inhibitors. *Journal of Industrial and Engineering Chemistry*, *19*(6), 2010–2015.
- Lee, J.-W., Gwak, K.-S., Park, J.-Y., Park, M.-J., Choi, D.-H., Kwon, M., & Choi, I.-G. (2007). Biological pretreatment of softwood *Pinus densiflora* by three white rot fungi. *Journal of Microbiology (Seoul, Korea)*, 45(6), 485–491.
- Li, H., Zhang, R., Tang, L., Zhang, J., & Mao, Z. (2015). Manganese peroxidase production from cassava residue by *Phanerochaete chrysosporium* in solid state fermentation and its decolorization of indigo carmine. *Chinese Journal of Chemical Engineering*, 23(1), 227– 233.
- Li, Q., Ng, W. T., & Wu, J. C. (2014). Isolation, characterization and application of a cellulose-degrading strain *Neurospora crassa* S1 from oil palm empty fruit bunch. *Microbial Cell Factories*, *13*(1), 157.

- Li, X., Jia, R., Li, P., & Ang, S. (2009). Response surface analysis for enzymatic decolorization of Congo red by manganese peroxidase. *Journal of Molecular Catalysis B: Enzymatic*, *56*(1), 1–6.
- Loh, S. K. (2017). The potential of the Malaysian oil palm biomass as a renewable energy source. *Energy Conversion and Management*, 141, 285–298.
- Lordêlo, M., Silva, C., Souza, V. B. De, Santos, S., Kamida, H. M., Vasconcellos-neto, J. R. T. De, & Góes-neto, A. (2014). Production of manganese peroxidase by *Trametes villosa* on unexpensive substrate and its application in the removal of lignin from agricultural wastes. *Advances in Bioscience and Biotechnology*, (December), 1067–1077.
- Low, J. C., Halis, R., Md Shah, U. K., Tahir, P., Abood, F., Tukimin, T., Idris, M. D., Lanika, L., Razali, N. (2015). Enhancing enzymatic digestibility of alkaline pretreated banana pseudostem for sugar production. *BioResources*, 10(1), 1213–1223.
- Machado, C., Souza, A. R. C. de, Baldoni, D. B., Guedes, J. V. C., Marcuz, C., Porto, V., Ferraz, R. C., Lima, J., Mazutti, M. A., Jacques, R. J. S., Kuhn, R. C. (2016). Selection, isolation, and identification of fungi for bioherbicide production. *Brazilian Journal of Microbiology*, 48(1), 101– 108.
- Mahanim, S., Zulkafli, H., & Mori, Y. (2010). Malaysian oil palm biomass. Regional Workshop on UNEP/DTIE/IETC, (March), 1–26.
- Majeau, J. A., Brar, S. K., & Tyagi, R. D. (2010). Laccases for removal of recalcitrant and emerging pollutants. *Bioresource Technology*, *101*(7), 2331–2350.
- Malaysian Palm Oil Board. (2018). Overview of the Malaysian oil palm industry 2017. *Malaysian Palm Oil Board [ MPOB ]*, *1*, 1–6.
- Mallek-Fakhfakh, H., Fakhfakh, J., Walha, K., Hassairi, H., Gargouri, A., & Belghith, H. (2017). Enzymatic hydrolysis of pretreated Alfa fibers (*Stipa tenacissima*) using β-D-glucosidase and xylanase of *Talaromyces thermophilus* from solid-state fermentation. *International Journal of Biological Macromolecules*, *103*, 543–553.

Manan, M. A. (2014). Design aspects of solid state fermentation.

- Manan, M. A., & Webb, C. (2016). Extracted substrate colour as an indicator of fungal growth in solid state fermentation, *12*(6), 445–449.
- Manan, M. A., & Webb, C. (2017). Design aspects of solid state fermentation as applied to microbial bioprocessing. *Journal of Applied Biotechnology* & *Bioengineering*, *4*(1), 511–532.

- Manan, M. A., & Webb, C. (2018). Estimating fungal growth in submerged fermentation in the presence of solid particles based on colour development particles based on colour development. *Biotechnology* and Biotechnological Equipment, 32(3), 618–627.
- Manickam, N. K., Rajarathinam, R., Muthuvelu, K. S., & Senniyappan, T. (2018). New insight into the effect of fungal mycelia present in the biopretreated paddy straw on their enzymatic saccharification and optimization of process parameters. *Bioresource Technology*, 267(July), 291–302.
- Maurya, D. P., Singla, A., & Negi, S. (2013). An overview of key pretreatment processes for biological conversion of lignocellulosic biomass to bioethanol. *3 Biotech*, *3*(5), 415–431.
- McIntosh, S., & Vancov, T. (2011). Optimisation of dilute alkaline pretreatment for enzymatic saccharification of wheat straw. *Biomass and Bioenergy*, 35(7), 3094–3103.
- Md Yunos, N. S. H., Baharuddin, A. S., Md Yunos, K. F., Naim, N., & Nishida, H. (2012). Physicochemical property changes of oil palm mesocarp fibers treated with high-pressure steam. *BioResources*, 7(4), 5983– 5994.
- Medina, J. D., Woiciechowski, A., Zandona Filho, A., Noseda, M. D., Kaur, B. S., & Soccol, C. R. (2015). Lignin preparation from oil palm empty fruit bunches by sequential acid/alkaline treatment A biorefinery approach. *Bioresource Technology*, 194, 172–178.
- Medina, J. D. C., Woiciechowski, A., Filho, A. Z., Nigam, P. S., Ramos, L. P., & Soccol, C. R. (2016). Steam explosion pretreatment of oil palm empty fruit bunches (EFB) using autocatalytic hydrolysis: A biorefinery approach. *Bioresource Technology*, *199*, 173–180.
- Meehnian, H., Jana, A. K., & Jana, M. M. (2016). Effect of particle size, moisture content, and supplements on selective pretreatment of cotton stalks by *Daedalea flavida* and enzymatic saccharification. *3 Biotech*, *6*(2), 1–13.
- Mekala, N. K., Singhania, R. R., Sukumaran, R. K., & Pandey, A. (2008). Cellulase production under solid-state fermentation by *Trichoderma reesei* RUT C30: Statistical optimization of process parameters. *Applied Biochemistry and Biotechnology*, 151(2–3), 122–131.
- Millati, R., Syamsiah, S., Niklasson, C., Cahyanto, M. N., Ludquist, K., & Taherzadeh, M. J. (2011). Biological pretreatment of lignocelluloses with white-rot fungi and its applications: A review. *BioResources*, *6*(4), 5224–5259.

- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*, *31*(3), 426–428.
- Mishra, V., Jana, A. K., Jana, M. M., & Gupta, A. (2017). Enhancement in multiple lignolytic enzymes production for optimized lignin degradation and selectivity in fungal pretreatment of sweet sorghum bagasse. *Bioresource Technology*, 236, 49–59.
- Mishra, V. K., & Singh, B. P. (2015). An overview on fungal diversity in North East India : options for research and development, (October).
- Najafpour, G., Ideris, A., Salmanpour, S., & Norouzi, M. (2007). Acid hydrolysis of pretreated palm oil lignocellulosic wastes. *International Journal of Engineering, Transactions B: Applications*, 20(2), 147–156.
- Nazarpour, F., Abdullah, D. K., Abdullah, N., Motedayen, N., & Zamiri, R. (2013). Biological pretreatment of rubberwood with *Ceriporiopsis* subvermispora for enzymatic hydrolysis and bioethanol production. *BioMed Research International*, 2013, 268349.
- Nazarpour, F., Abdullah, D. K., Abdullah, N., & Zamiri, R. (2013). Evaluation of biological pretreatment of rubberwood with white rot fungi for enzymatic hydrolysis. *Materials*, 6(5), 2059–2073.
- NCIM. (2014). National collection of industrial micro-organisms (NCIM). NCIM Catalogue of Strains, 4, 1–14.
- Nieves, D. C., Karimi, K., & Horvath, I. S. (2011). Improvement of biogas production from oil palm empty fruit bunches (OPEFB). *Industrial Crops and Products*, *34*(1), 1097–1101.
- Nomanbhay, S. M., Hussain, R., & Palanisamy, K. (2013). Microwave-assisted alkaline pretreatment and microwave assisted enzymatic saccharification of oil palm empty fruit bunch fiber for enhanced fermentable sugar yield. *Journal of Sustainable Bioenergy Systems*, *03*(01), 7–17.
- Nor, N. A. M., Mustapha, W. A. W., & Hassan, O. (2016). Deep eutectic solvent (DES) as a pretreatment for oil palm empty fruit bunch (OPEFB) in sugar production. *Procedia Chemistry*, *18*(Mcls 2015), 147–154.
- Ohm, R. A., De Jong, J. F., Lugones, L. G., Aerts, A., Kothe, E., Stajich, J. E., ... Wösten, H. A. B. (2010). Genome sequence of the model mushroom *Schizophyllum commune. Nature Biotechnology*, *28*(9), 957–963.
- Omar, F. N., Hanipah, S. H., Xiang, L. Y., Mohammed, M. A. P., Baharuddin, A. S., & Abdullah, J. (2016). Micromechanical modelling of oil palm empty fruit bunch fibres containing silica bodies. *Journal of the Mechanical Behavior of Biomedical Materials*, 62, 106–118.

- Omar, F. N., Mohammed, M. a. P., & Baharuddin, A. S. (2014). Effect of silica bodies on the mechanical behaviour of oil palm empty fruit bunch fibres. *BioResources*, *9*(4), 7041–7058.
- Padhiar, A., Nagadesi, P. K., Albert, S., & Arya, A. (2009). Morphology, anatomy and cultural characters of two wood decaying fungi Schizophyllum commune and Flavadon flavus, *39*(1), 27–31.
- Palamae, S., Palachum, W., Chisti, Y., & Choorit, W. (2014). Retention of hemicellulose during delignification of oil palm empty fruit bunch (EFB) fiber with peracetic acid and alkaline peroxide. *Biomass and Bioenergy*, 66, 240–248.
- Panapanaan, V. (2009). Sustainability of palm oil production and opportunities for finnish technology and know-how transfer. *Production*, *5*, 1–115.
- Patel, H., & Gupte, A. (2016). Optimization of different culture conditions for enhanced laccase production and its purification from *Tricholoma* giganteum AGHP. *Bioresources and Bioprocessing*, *3*(1), 11.
- Plácido, J., & Capareda, S. (2015). Ligninolytic enzymes: A biotechnological alternative for bioethanol production. *Bioresources and Bioprocessing*, 2(23), 1–12.
- Popescu, C. M., Lisa, G., Manoliu, A., Gradinariu, P., & Vasile, C. (2010). Thermogravimetric analysis of fungus-degraded lime wood. *Carbohydrate Polymers*, *80*(1), 78–83.
- Postemsky, P. D., & Curvetto, N. R. (2015). Solid-state fermentation of cereal grains and sunflower seed hulls by *Grifola gargal* and *Grifola sordulenta*. International Biodeterioration and Biodegradation, 100, 52–61.
- Puri, S., Arora, M., & Sarao, L. (2013). Production and optimization of amylase and glucoamylase using *Aspergillus oryzae* under solid state fermentation. *International Journal of Research in Pure and Applied Microbiology*, *3*(3), 83–88.
- Qian, L.-C., Fu, S.-J., Zhou, H.-M., Sun, J.-Y., & Weng, X.-Y. (2012). Optimization of fermentation parameters for β-glucosidase ptoduction by *Aspergillus niger*. *Journal of Animal and Veterinary Advances*, *11*(5), 583–591.
- Qiu, W., & Chen, H. (2012). Enhanced the enzymatic hydrolysis efficiency of wheat straw after combined steam explosion and laccase pretreatment. *Bioresource Technology*, *118*, 8–12.
- Rabemanolontsoa, H., & Saka, S. (2016). Various pretreatments of lignocellulosics. *Bioresource Technology*, *199*, 83–91.

- Raja, H. A., Miller, A. N., Pearce, C. J., & Oberlies, N. H. (2017). Fungal identification using molecular tools: A primer for the natural products research community. *Journal of Natural Products*, *80*(3), 756–770.
- Ramli, R., Junadi, N., Beg, M. D. H., & Yunus, R. M. (2015). Microcrystalline cellulose (MCC) from oil palm empty fruit bunch (EFB) fiber via simultaneous ultrasonic and alkali treatment. *Chemical, Molecular, Nuclear, Materials and Metallurgical Engineering*, 9(1), 8–11.
- Rasmussen, M. L., Shrestha, P., Khanal, S. K., Pometto, A. L., & (Hans) van Leeuwen, J. (2010). Sequential saccharification of corn fiber and ethanol production by the brown rot fungus *Gloeophyllum trabeum*. *Bioresource Technology*, *101*(10), 3526–3533.
- Remli, N. A. M., Md Shah, U. K., Mohamad, R., & Abd-Aziz, S. (2014). Effects of chemical and thermal pretreatments on the enzymatic saccharification of rice straw for sugars production. *BioResources*, 9(1), 510–522.
- Rouches, E., Herpoël-Gimbert, I., Steyer, J. P., & Carrere, H. (2016). Improvement of anaerobic degradation by white-rot fungi pretreatment of lignocellulosic biomass: A review. *Renewable and Sustainable Energy Reviews*, *59*, 179–198.
- Rozario, M. (2013). National biomass strategy 2020: New wealth creation for Malaysia's palm oil industry. *Agensi Inovasi Malaysia*, (June), 1–32.
- Ruiz-Dueñas, F. J., Morales, M., García, E., Miki, Y., Martínez, M. J., & Martínez, A. T. (2009). Substrate oxidation sites in versatile peroxidase and other basidiomycete peroxidases. *Journal of Experimental Botany*, *60*(2), 441–452.
- Ruqayyah, T. I. D., Jamal, P., Alam, M. Z., Mirghani, M. E. S., Jaswir, I., & Ramli, N. (2014). Application of response surface methodology for protein enrichment of cassava peel as animal feed by the white-rot fungus *Panus tigrinus* M609RQY. *Food Hydrocolloids*, *42*(P2), 298– 303.
- Saha, B. C., Qureshi, N., Kennedy, G. J., & Cotta, M. A. (2016). Biological pretreatment of corn stover with white-rot fungus for improved enzymatic hydrolysis. *International Biodeterioration and Biodegradation*, *109*, 29–35.
- Salvachúa, D., Prieto, A., López-Abelairas, M., Lu-Chau, T., Martínez, Á. T., & Martínez, M. J. (2011). Fungal pretreatment: An alternative in second-generation ethanol from wheat straw. *Bioresource Technology*, *102*(16), 7500–7506.

- Santanaraj, J., Sajab, M. S., Mohammad, A. W., Harun, S., Chia, C. H., Zakari, S., & Kaco, H. (2017). Enhanced delignification of oil palm empty fruit bunch fibers with in situ fenton-oxidation. *BioResources*, 12(3), 5223– 5235.
- Saritha, M., Arora, A., & Lata. (2012). Biological pretreatment of lignocellulosic substrates for enhanced delignification and enzymatic digestibility. *Indian Journal of Microbiology*, *52*(2), 122–130.
- Sasso, M., Barrot, A., Carles, M. J., Griffiths, K., Rispail, P., Crampette, L., Lallemant, B., Lachaud, L. (2017). Direct identification of molds by sequence analysis in fungal chronic rhinosinusitis. *Journal de Mycologie Medicale*, *27*(4), 514–518.
- Schoch, C. L., Seifert, K. a., Huhndorf, S., Robert, V., Spouge, J. L., Levesque, C. A., Schindel, D. (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. *Proceedings of the National Academy of Sciences of the United States of America*, 109(16), 1–6.
- Senthivelan, T., Kanagaraj, J., & Panda, R. C. (2016). Recent trends in fungal laccase for various industrial applications: An eco-friendly approach A review. *Biotechnology and Bioprocess Engineering*, *21*(1), 19–38.
- Sha, M., Kumar, R., & Karimi, K. (2015). Pretreatment of lignocellulosic biomass (Vol. 1).
- Shahriarinour, M., Abdul Wahab, M. N., Mustafa, S., Mohamad, R., & Ariff, A.
   B. (2011). Effect of various pretreatments of oil palm empty on the performance of cellulase production by *Aspergillus terreus*. *BioResources*, 6(1), 291–307.
- Shamsudin, S., Md Shah, U. K., Zainudin, H., Abd-Aziz, S., Mustapa Kamal, S. M., Shirai, Y., & Hassan, M. A. (2012). Effect of steam pretreatment on oil palm empty fruit bunch for the production of sugars. *Biomass and Bioenergy*, *36*, 280–288.
- Shariff, A., Syairah, N., Aziz, M., & Abdullah, N. (2014). Slow pyrolysis of oil palm empty fruit bunches for biochar production and characterisation. *Journal of Physical Science*, *25*(2), 97–112.
- Shi, J., Sharma-Shivappa, R. R., & Chinn, M. S. (2009). Microbial pretreatment of cotton stalks by submerged cultivation of *Phanerochaete chrysosporium*. *Bioresource Technology*, *100*(19), 4388–4395.
- Shirkavand, E., Baroutian, S., Gapes, D. J., & Young, B. R. (2016). Combination of fungal and physicochemical processes for lignocellulosic biomass pretreatment - A review. *Renewable and Sustainable Energy Reviews*, 54, 217–234.

- Shirkavand, E., Baroutian, S., Gapes, D. J., & Young, B. R. (2017). Pretreatment of radiata pine using two white rot fungal strains *Stereum hirsutum* and *Trametes versicolor*. *Energy Conversion and Management*, *142*, 13–19.
- Shrestha, B., Lee, W.-H., Han, S.-K., & Sung, J.-M. (2006). Observations on some of the mycelial growth and pigmentation characteristics of cordyceps militaris isolates. *Mycobiology*, *34*(2), 83–91.
- Shrestha, P., Khanal, S. K., Pomettoiii, A. L., & Van Leeuwen, J. (2009). Enzyme production by wood-rot and soft-rot fungi cultivated on corn fiber followed by simultaneous saccharification and fermentation. *Journal of Agricultural and Food Chemistry*, *57*(10), 4156–4161.
- Sia, C. V., Nakai, Y., Tanaka, H., & Shiozawa, D. (2014). Interfacial fracture toughness evaluation of poly (L-lactide acid)/natural fiber composite by using double shear test method. *Journal of Composite Materials*, (4), 97–105.
- Sidik, D. A. B., Ngadi, N., & Amin, N. A. S. (2013). Optimization of lignin production from empty fruit bunch via liquefaction with ionic liquid. *Bioresource Technology*, *135*, 690–696.
- Sim, C. Y.-Y. (2014). Molecular identification of *Schizophyllum commune* and its production of schizophyllan, (May).
- Simarani, K., Hassan, M. A., Abd-Aziz, S., Wakisaka, M., & Shirai, Y. (2009). Effect of palm oil mill sterilization process on the physicochemical characteristics and enzymatic hydrolysis of empty fruit bunch. *Journal* of the American Medical Association.
- Sindhu, R., Binod, P., & Pandey, A. (2016). Biological pretreatment of lignocellulosic biomass An overview. *Bioresource Technology*, *199*, 76–82.
- Singh, M. K., Singh, J., Kumar, M., & Thakur, I. S. (2014). Novel lipase from basidiomycetes *Schizophyllum commune* ISTL04, produced by solid state fermentation of Leucaena leucocephala seeds. *Journal of Molecular Catalysis B: Enzymatic*, 110, 92–99.
- Sluiter, A., Hames, B., Hyman, D., Payne, C., Ruiz, R., Scarlata, C., Templeton, D., Nrel, J. W. (2008). Determination of total solids in biomass and total dissolved solids in liquid process samples. *National Renewable Energy Laboratory (NREL)*, (March), 9.
- Sluiter, A., Hames, B., Ruiz, R., Scarlata, C., Sluiter, J., & Templeton, D. (2008a). Determination of ash in biomass. *National Renewable Energy Laboratory (NREL)*, (January), 1–5.

- Sluiter, A., Ruiz, R., Scarlata, C., Sluiter, J., & Templeton, D. (2008). Determination of extractives in biomass. *National Renewable Energy Laboratory (NREL)*, (January), 1–9.
- Song, L., Ma, F., Zeng, Y., Zhang, X., & Yu, H. (2013). The promoting effects of manganese on biological pretreatment with *Irpex lacteus* and enzymatic hydrolysis of corn stover. *Bioresource Technology*, 135, 89– 92.
- Sornlake, W., Rattanaphanjak, P., Champreda, V., Eurwilaichitr, L., Kittisenachai, S., Roytrakul, S., Fujii, T., Inoue, H. (2017). Characterization of cellulolytic enzyme system of *Schizophyllum commune* mutant and evaluation of its efficiency on biomass hydrolysis. *Bioscience, Biotechnology and Biochemistry*, *81*(7), 1289–1299.
- Sumathi, S., Chai, S. P., & Mohamed, A. R. (2008). Utilization of oil palm as a source of renewable energy in Malaysia. *Renewable and Sustainable Energy Reviews*, *12*(9), 2404–2421.
- Sun, S., Sun, S., Cao, X., & Sun, R. (2016). The role of pretreatment in improving the enzymatic hydrolysis of lignocellulosic materials. *Bioresource Technology*, *199*, 49–58.
- Sun, Y., & Cheng, J. Y. (2002). Hydrolysis of lignocellulosic materials for ethanol production: A review. *Bioresource Technology*, *83*(1), 1–11.
- Tamura, K., Stecher, G., Peterson, D., Filipski, A., & Kumar, S. (2013). MEGA6: Molecular evolutionary genetics analysis version 6.0. Molecular Biology and Evolution, 30(12), 2725–2729.
- Tan, J. P., Jahim, J. M., Harun, S., Wu, T. Y., & Mumtaz, T. (2016). Utilization of oil palm fronds as a sustainable carbon source in biorefineries. *International Journal of Hydrogen Energy*, *41*(8), 4896–4906.
- Teoh, Y. P., & Don, M. M. (2012). Optimization of parameters for mycelia growth by *Schizophyllum commune* and a kinetic odel study of its growth morphology. *Journal of Applied Sciences*, *12*(11), 1100–1105.
- Teoh, Y. P., & Don, M. M. (2015). Effect of temperature on *Schizophyllum commune* 5-dihydroxy-6-methyl- production using a bubble column bioreactor. *Journal of Science*, *42*(3), 539–548.
- Teoh, Y. P., Don, M. M., & Ujang, S. (2017). Production of biomass by Schizophyllum commune and its antifungal activity towards rubberwood-degrading fungi. *Sains Malaysiana*, *46*(1), 123–128.
- Thiribhuvanamala, G., Kalaiselvi, G., Parthasarathy, S., Adhavan, S. M., & Prakasam, V. (2017). Extracellular secretion of lignocellulolytic enzymes by diverse white rot basidiomycetes fungi, *6*(1), 20–29.

- Tian, X., Fang, Z., & Guo, F. (2012). Impact and prospective of fungal pretreatment of lignocellulosic biomass for enzymatic hydrolysis. *Biofuels, Bioproducts and Biorefining*, *6*(3), 246–256.
- Tien, M., & Kent Kirk, T. (1988). Lignin peroxidase of *Phanerochaete* chrysosporium. Methods in Enzymology, 161, 238–249.
- Ting, A. S. Y., Tay, H., Peh, K. L., Tan, W. S., & Tee, C. S. (2013). Novel isolation of thermophilic *Ureibacillus terrenus* from compost of empty fruit bunches (EFB) of oil palm and its enzymatic activities. *Biocatalysis* and Agricultural Biotechnology, 2(2), 162–164.
- Tsujiyama, S., & Ueno, H. (2011). Production of cellulolytic enzymes containing cinnamic acid esterase from *Schizophyllum commune*. *Journal of General and Applied Microbiology*, *57*(6), 309–317.
- Tullio, V., Mandras, N., Banche, G., Allizond, V., Gaido, E., Roana, J., Cufflni, A. M., Carlone, N. (2008). Schizophyllum commune: an unusual agent of bronchopneumonia in an immunocompromised patient. *Medical Mycology*, 46(7), 735–738.
- Ujor, V. C., Monti, M., Peiris, D. G., Clements, M. O., & Hedger, J. N. (2012). The mycelial response of the white-rot fungus, *Schizophyllum commune* to the biocontrol agent, Trichoderma viride. *Fungal Biology*, *116*(2), 332–341.
- van Kuijk, S. J. A., Sonnenberg, A. S. M., Baars, J. J. P., Hendriks, W. H., & Cone, J. W. (2015). Fungal treated lignocellulosic biomass as ruminant feed ingredient: A review. *Biotechnology Advances*, *33*(1), 191–202.
- Vandenbossche, V., Brault, J., Vilarem, G., Hernandez-Melendez, O., Vivaldo-Lima, E., Hernandez-Luna, M., Barzana, E., Duque, A., Manzanares, P., Ballesteros, M., Mata, J., Castellon, E., Rigal, L. (2014). A new lignocellulosic biomass deconstruction process combining thermomechano chemical action and bio-catalytic enzymatic hydrolysis in a twin-screw extruder. *Industrial Crops and Products*, *55*, 258–266.
- Wagner, A. O., Lackner, N., Mutschlechner, M., Prem, E. M., Markt, R., & Illmer, P. (2018). Biological pretreatment strategies for secondgeneration lignocellulosic resources to enhance biogas production. *Energies*, *11*(7).
- Wan, C., & Li, Y. (2010). Microbial pretreatment of corn stover with *Ceriporiopsis subvermispora* for enzymatic hydrolysis and ethanol production. *Bioresource Technology*, *101*(16), 6398–6403.
- Wan, C., & Li, Y. (2011). Effectiveness of microbial pretreatment by Ceriporiopsis subvermispora on different biomass feedstocks. Bioresource Technology, 102(16), 7507–7512.

- Wan, C., & Li, Y. (2012). Fungal pretreatment of lignocellulosic biomass. *Biotechnology Advances*.
- Wang, S., Dai, G., Yang, H., & Luo, Z. (2017). Lignocellulosic biomass pyrolysis mechanism: A state-of-the-art review. *Progress in Energy and Combustion Science*, 62, 33–86.
- Wang, Z., Zhu, J. Y., Fu, Y., Qin, M., Shao, Z., Jiang, J., & Yang, F. (2013). Lignosulfonate-mediated cellulase adsorption: Enhanced enzymatic saccharification of lignocellulose through weakening nonproductive binding to lignin. *Biotechnology for Biofuels*, 6(1).
- Wong, D. W. S. (2009). Structure and action mechanism of ligninolytic enzymes. *Applied Biochemistry and Biotechnology*, *157*(2), 174–209.
- Wood, T. M., & Bhat, K. M. (1988). Methods for measuring cellulase activities. *Methods in Enzymology*, *160*(C), 87–112.
- Xu, F., Sun, J. X., Geng, Z. C., Liu, C. F., Ren, J. L., Sun, R. C., Fowler, P., Baird, M. S. (2007). Comparative study of water-soluble and alkalisoluble hemicelluloses from perennial ryegrass leaves (*Lolium peree*). *Carbohydrate Polymers*, 67(1), 56–65.
- Yao, W., & Nokes, S. E. (2014). Phanerochaete chrysosporium pretreatment of biomass to enhance solvent production in subsequent bacterial solidsubstrate cultivation. Biomass and Bioenergy, 62, 100–107.
- Ying, T. Y., Teong, L. K., Abdullah, W. N. W., & Peng, L. C. (2014). The effect of various pretreatment methods on oil palm empty fruit bunch (EFB) and kenaf core fibers for sugar production. *Procedia Environmental Sciences*, 20, 328–335.
- Yoon, L. W., Ang, T. N., Ngoh, G. C., & Chua, A. S. M. (2014). Fungal solidstate fermentation and various methods of enhancement in cellulase production. *Biomass and Bioenergy*, 67, 319–338.
- Yu, J., Zhang, J., He, J., Liu, Z., & Yu, Z. (2009). Combinations of mild physical or chemical pretreatment with biological pretreatment for enzymatic hydrolysis of rice hull. *Bioresource Technology*, 100(2), 903–908.
- Yunos, N. S. H. M., Baharuddin, A. S., Md Yunos, K. F., Hafid, H. S., Busu, Z., Mokhtar, M. N., Sulaiman, A., Som, A. M. (2015). The physicochemical characteristics of residual oil and fibers from oil palm empty fruit bunches. *BioResources*, *10*(1), 14–29.
- Zahari, M. A. K. M., Ariffin, H., Mokhtar, M. N., Salihon, J., Shirai, Y., & Hassan, M. A. (2015). Case study for a palm biomass biorefinery utilizing renewable non-food sugars from oil palm frond for the production of poly(3-hydroxybutyrate) bioplastic. *Journal of Cleaner Production*, 87(C), 284–290.

- Zanirun, Z., Bahrin, E. K., Lai-Yee, P., Hassan, M. A., & Abd-Aziz, S. (2015). Enhancement of fermentable sugars production from oil palm empty fruit bunch by ligninolytic enzymes mediator system. *International Biodeterioration and Biodegradation*, *105*, 13–20.
- Zhao, J., Zheng, Y., & Li, Y. (2014). Fungal pretreatment of yard trimmings for enhancement of methane yield from solid-state anaerobic digestion. *Bioresource Technology*, *156*, 176–181.
- Zheng, Y., Zhao, J., Xu, F., & Li, Y. (2014). Pretreatment of lignocellulosic biomass for enhanced biogas production. *Progress in Energy and Combustion Science*, *4*2(1), 35–53.
- Zhu, N., Liu, J., Yang, J., Lin, Y., Yang, Y., Ji, L., Li, M., Yuan, H. (2016). Comparative analysis of the secretomes of *Schizophyllum commune* and other wood-decay basidiomycetes during solid-state fermentation reveals its unique lignocellulose-degrading enzyme system. *Biotechnology for Biofuels*, 9(1), 42.

#### **BIODATA OF STUDENT**



Enis Natasha Noor Binti Arbaain was born in Alor Setar, Kedah Darul Aman on September 23, 1992. She attended her elementary school at Sekolah Kebangsaan Guar Napai for 6 years from 1999 to 2004. She continued her secondary education at Sekolah Menengah Kebangsaan Jitra, Kedah (2005 -2007) and MARA Junior Science College Pendang, Kedah (2008 - 2009). She obtained her pre-university education at Penang Matriculation College, Penang from 2010 to 2011 for one-year matriculation program. In the year of 2011 until 2015, she enrolled as a degree student in Bachelor of Chemical Engineering Technology (Industrial Biotechnology), a four-year program at Universiti Malaysia Perlis (UniMAP), Perlis. She was accepted for six-month internship program (February-July 2015) in Department of Plant Functional Genomics at Sime Darby Technology Centre Sdn. Bhd. During her final year of Bachelor's Degree, she managed to complete a final year project, entitled as "Phase equilibrium studies of biodiesel derivatives based on soybean methyl ester for engine technology". In 2016, she further her study in degree of Master of Science in Environmental Biotechnology. She was attached to Kyushu Institute of Technology starting from August until October of 2017 under sponsorship of Japan Student Services Organization (JASSO).

## LIST OF PUBLICATIONS

### Paper publication:

- Enis Natasha Noor Arbaain, Ezyana Kamal Bahrin\*, Mohamad Faizal Ibrahim, Yoshito Ando and Suraini Abd-Aziz (2019). Biological pretreatment of oil palm empty fruit bunch by *Schizophyllum commune* ENN1 without washing and nutrient addition. *Processes*, 7(7). (Published)
- Enis Natasha Noor Arbaain, Ezyana Kamal Bahrin\*, Nurshakinah Mohd Noor Mohamad Faizal Ibrahim, Norhayati Ramli and Suraini Abd-Aziz. Chemical-free pretreatment of unwashed oil palm empty fruit bunch by using locally isolated fungus (*Schizophyllum commune* ENN1) for delignification. *Food and Bioproducts Processing*. (Accepted)

#### Abstract in conference/symposium:

- Enis Natasha Noor Arbaain, Ezyana Kamal Bahrin, Suraini Abd-Aziz and Mohamad Faizal Ibrahim. In the 5<sup>th</sup> SAES - International Symposium on Applied Engineering and Sciences (SAES2017) UPM - Kyutech, UPM, Selangor, Malaysia. (Poster presenter)
- Enis Natasha Noor Arbaain, Ezyana Kamal Bahrin, Suraini Abd-Aziz and Mohamad Faizal Ibrahim. In the Wood and Biofibre International Conference 2017 (WOBIC2017), Putrajaya, Selangor, Malaysia. (Poster presenter)