

# **UNIVERSITI PUTRA MALAYSIA**

## EFFECTS OF HERBICIDES, HYDROGEN PEROXIDE AND PHYTOHORMONES ON *Ganoderma* INFECTION IN OIL PALM (*Elaeis guineensis* Jacq.) ROOTS

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FBSB 2019 8



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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

November 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

## EFFECTS OF HERBICIDES, HYDROGEN PEROXIDE AND PHYTOHORMONES ON Ganoderma INFECTION IN OIL PALM (Elaeis guineensis Jacq.) ROOTS

By

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Basal Stem Rot (BSR) caused by Ganoderma is the major disease that infects oil palms (Elaeis guineensis Jacq.). Application of herbicides in plantations for weed control might affect BSR development. The effects of hydrogen peroxide  $(H_2O_2)$  and phytohormones which are key signalling agents in plant defense mechanisms on Ganoderma infection are unknown. Therefore, the objectives of this study are to investigate the effects of herbicides, H<sub>2</sub>O<sub>2</sub> and phytohormones on Ganoderma infection in oil palm roots, and to profile the gene expression of transcripts related to hydrogen peroxide production, hormone biosynthesis and signaling during Ganoderma infection in oil palm roots. The effects of three commonly used herbicides in plantations on the growth of Ganoderma spp. and infection progress in G. boninense PER71-inoculated oil palm seedlings were examined. Evaluation on the tolerance of G. boninense (pathogenic), G. miniatocinctum (pathogenic), and G. lucidum (non-pathogenic) to herbicides revealed that glyphosate (Roundup®, GLY) 1800 ppm-2700 ppm can reduce the growth of all three Ganoderma species. Both Basta (GLA at 750-3000 ppm) and Paraguat (PQ at 325-300 ppm) inhibited the growth of all three Ganoderma species studied except for G. lucidum which was able to grow at 650 ppm PQ. GLA and PQ increased the disease severity of oil palm to Ganoderma infection. The fungus may avoid the herbicide contaminated soil environment and then infected the oil palm seedlings that could have been weakened by the herbicides. The open reading frame and conserved functional domains of eight cDNAs encoding  $H_2O_2$ , phytohormone biosynthesis and signalling in oil palm were analyzed. The transcript abundance of EgRBOHB2 in G. boninense-treated oil palm roots increased 2.42 fold at 3 wpi compared to uninoculated oil palm seedlings. Meanwhile, EgRBOHB1, EgRBOHH and EgHIR did not show significant changes in expression at all three time points. The transcript abundance of EgCOI (jasmonic acid, JA-related) increased at 6 and 12 wpi whereas the transcript abundance EgNPR1 (salicyclic acid, SA-related) increased at 3 wpi, reduced at 6 and 12 wpi; suggesting a well-coordinated signal crosstalk between JA and SA. The expression of EqOPR which is related to JA biosynthesis was up-regulated at 6 wpi; coincided with the upregulation of EgCOI (which perceives JA). The EgACO1 (ethylene, ET- related) was also upregulated at the early stage of infection by 3.2 and 2.2 fold at 3 and 6 wpi in the inoculated oil palm seedlings, respectively. The exogenous application of phytohormones did not suppress the BSR in Ganoderma-inoculated oil palm seedlings while the application of their inhibitors, caused an early onset and more severe disease symptoms. DPI pre-treatment was the only treatment that delayed the onset and reduced the severity of disease symptoms. The distilled water-treated Ganoderma inoculated oil palm fresh root samples (control) had a significantly higher H<sub>2</sub>O<sub>2</sub> level at 3 wpi compared with uninoculated oil palm root samples. However, there was no significant difference in H<sub>2</sub>O<sub>2</sub> level between the Ganoderma inoculated and uninoculated oil palm root sample of the other phytohormone-treated or their inhibitor treated oil palm seedlings. The JA-treated and Ganoderma-inoculated oil palm root sample had the lowest H<sub>2</sub>O<sub>2</sub> level among all the other roots samples at 6 wpi while the SA-treated, Ganoderma inoculated oil palm roots had the lowest H<sub>2</sub>O<sub>2</sub> level among all the other root samples at 12 wpi. The H<sub>2</sub>O<sub>2</sub> inhibitor- treated oil palm roots generally had lower H<sub>2</sub>O<sub>2</sub> level when compared with other treatments. Overall, the H<sub>2</sub>O<sub>2</sub> levels did not correspond to the disease symptoms and severity but showed an increase at the onset of disease symptoms. In conclusion, the findings from this study have given an insight on the effects of herbicides, H<sub>2</sub>O<sub>2</sub>, phytohormones and their inhibitors on Ganoderma infection and disease progress of BSR in oil palm seedlings.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

## KESAN RACUN RUMPAI, HIDROGEN PEROKSIDA DAN HORMON TUMBUHAN KE ATAS INFEKSI *Ganoderma* PADA AKAR ANAK BENIH KELAPA SAWIT (*Elaeis guineensis* Jacq.)

Oleh

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Reput pangkal batang (BSR) yang disebabkan oleh kulat patogenik, Ganoderma adalah penyakit utama yang menjangkit kelapa sawit (Elaeis guineensis Jacq.). Penggunaan racun rumpai di ladang kelapa sawit untuk pengawalan rumpai mungkin mempengaruhi perkembangan BSR. Kesan hidrogen peroksida (H<sub>2</sub>O<sub>2</sub>) dan hormon tumbuhan yang merupakan ejen pengisvaratan utama dalam mekanisme pertahanan tumbuhan ketika jangkitan Ganoderma tidak diketahui. Oleh itu, objektif kajian ini adalah untuk mengkaji kesan racun rumpai, H<sub>2</sub>O<sub>2</sub> dan hormon tumbuhan ke atas jangkitan Ganoderma pada akar kelapa sawit, dan untuk memprofil ekspresi gen yang berkaitan penghasilan hidrogen peroksida, biosintesis hormon dengan dan pengisyaratan sistem pertahanan semasa jangkitan Ganoderma pada akar benih kelapa sawit. Kesan tiga racun rumpai yang biasa digunakan untuk mengawal pertumbuhan rumpai ke atas pertumbuhan Ganoderma spp. dan jangkitan G. boninense PER71 pada benih kelapa sawit telah diselidik. Pemeriksaan toleransi G. boninense (patogenik), G. miniatocinctum (patogenik), dan G. lucidum (tidak patogenik) kepada racun rumpai mendedahkan bahawa ® (GLY) pada kadar 1800-2700 ppm merencat pertumbuhan ketiga-tiga species Ganoderma. Basta (GLA, 750-3000 ppm) dan Paraquat (PQ, 325-1300 ppm) menyekat pertumbuhan ketiga-tiga spesis Ganoderma kecuali G. lucidum yang dapat tumbuh di atas media mengandungi 650 ppm PQ. GLA dan PQ mengakibatkan jangkitan Ganoderma yang paling serius pada akar kelapa sawit. Fungus tersubut mungkin terpaksa mengelak dari tanah yang dicemari racun rumpai lalu menjangkiti benih kelapa sawit yang mungkin telah dilemahkan oleh racun rumpai. Domain fungsian dan motif terpelihara lapan cDNA terpilih yang berkaitan dengan H2O2, biosintesis hormon tumbuhan dan pengisyaratan pertahanan dalam kelapa sawit telah dianalisis. Aras transkrip EgRBOHB2 daripada akar kelapa sawit yang diinokulasi G. boninense meningkat 2.42 kali pada 3 minggu selepas inokulasi (wpi) berbanding kelapa sawit yang tidak diinokulasi. Manakala, ekspresi gen EgRBOHB1, EgRBOHH dan EgHIR tidak menunjukkan perubahan signifikan pada tiga titik masa yang dikaji. Tahap transkrip EgCOI (berkait dengan asid jasmonik, JA) meningkat pada 6 wpi dan 12 wpi manakala, tahap transkrip EgNPR1 (berkait dengan asid salisiklik, SA) meningkat pada 3 wpi, berkurang pada 6 dan 12; wpi menunjukkan penyelarasan isyarat hormon JA dan SA yang baik. Ekspresi EgOPR yang berkaitan dengan biosintesis JA meningkat pada 6 wpi selaras dengan peningkatan ekspresi EgCOI yang mentafsir isyarat JA. Ekpresi EgACO1 (biosintesis etilena, ET) juga meningkat secara signifikan sebanyak 3.2 dan 2.2 kali ganda pada 3 dan 6 wpi pada kelapa sawit yang telah diinokulasikan. Aplikasi hormon tumbuhan tidak membataskan BSR pada benih kelapa sawit manakala aplikasi perencat hormone telah menyebabkan permulaan simptom infeksi yang awal dan kesan penyakit yang paling serius. Hanya rawatan dengan perencat H<sub>2</sub>O<sub>2</sub> memperlambatkan simptom penyakit reput pangkal batang. Benih kelapa sawit yang dirawat dengan air-suling dan diinokulasi dengan Ganoderma mempunyai tahap H<sub>2</sub>O<sub>2</sub> yang lebih tinggi berbanding dengan benih yang tidak diinokulasi. Sampel akar kelapa sawit yang dirawat dengan JA dan diinokulasi Ganoderma mempunyai tahap H<sub>2</sub>O<sub>2</sub> paling rendah pada 6 wpi, manakala kelapa sawit yang dirawat dengan SA dan diinokulasi Ganoderma mempunyai tahap H<sub>2</sub>O<sub>2</sub> paling rendah pada 12 wpi. Akar benih kelapa sawit yang dirawat dengan perencat H2O2 menunjukkan tahap H<sub>2</sub>O<sub>2</sub> yang rendah berbanding dengan rawatan lain. Pada keseluruhannya, tahap H<sub>2</sub>O<sub>2</sub> tidak menunjukkan perkaitan dengan symptom penyakit tetapi menunjukkan peningkatan pada permulaan symptom penyakit. Kesimpulannya, penemuan kajian ini telah memberi pengetahuan mengenai kesan racun rumpai, H<sub>2</sub>O<sub>2</sub>, hormon tumbuhan dan perencat hormon ke atas jangkitan Ganoderma dan perkembangan penyakit BSR pada kelapa sawit.

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## TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xv

# CHAPTER

1	INTF	RODUCT	ION	1
2	LITE 2.1 2.2	Oil Palr	erma Economic Importance of Basal Stem Rot (BSR) in Oil Palm	4 4 5 6
	2.3	2.2.2 Plant-P 2.3.1	Mode of <i>Ganoderma</i> Invasion athogen Interaction Reactive Oxygen Species (ROS) in Plants	7 8 10
		2.3.2	Roles of Phytohormones in Plant Defense	11
	2.4	He <mark>rbici</mark> 2.4.1	des in Oil Palm Plantations Roles of Herbicides in Plant	13 15
		2.4.2	Disease Development Herbicide Effects on White Rot Fungi	16
3	ON	Ganod	OF HERBICIDE ADMINISTRATION lerma spp. AND Ganoderma- D OIL PALM SEEDLINGS	18
	3.1			18
	3.2		ils and Methods	19
		3.2.1	Preparation of <i>Ganoderma</i> Fungal Cultures	19
		3.2.2	Growth of <i>Ganoderma</i> on Media with Herbicides	19
		3.2.3	Preparation of <i>Ganoderma</i> boninense PER71 Rubber Wood Block and Artificial Inoculation	20
		3.2.4	Effect of Herbicide Residue to Oil Palm Seedlings-Inoculated with Ganoderma	20

	3.3	Results 3.3.1	and Discussion Effects of Herbicides on the Growth of <i>Ganoderma</i> on Media with Herbicides	21 21
		3.3.2	Effects of Herbicides on Oil Palm Seedlings Inoculated with Ganoderma	26
	3.4	Conclus		32
4	PER TRA OIL PHY Gan	OXIDE, NSCRIP PALM TOHORI	TS IN Ganoderma-INOCULATED ROOTS AND EFFECTS OF MONES AND INHIBITORS ON INOCULATED OIL PALM	33
		Introduc		33
	4.2		Is and Methods	34
		4.2.1	Sequence Analysis of Candidate Transcripts Related to Hydrogen Peroxide, Hormone Biosynthesis and Signalling	34
		4.2.2	Gene Expression Analysis using Real-Time Quantitative Reverse Transcription (RT-qPCR)	35
		4.2.3 4.2.4	Plant Materials and Treatments Quantification of Hydrogen Peroxide	36 40
	4. <mark>3</mark>	Results a	and Discussion	40
		4.3.1	Sequence Analysis	40
		4.3.2		51
		4.3.3	Gene Expression Profiles of Hydrogen Peroxide, Phytohormone Biosynthesis and Signalling Related Genes in Ganoderma Inoculated Oil Palm Root Sample	51
		4.3.4	Disease Severity of Infected Oil Palm Roots	55
		4.3.5	Hydrogen Peroxide Level Assessment	63
	4.4	Conclus	ion	65
5	REC	•	GENERAL CONCLUSION AND DATION FOR FUTURE	66
REFERE				68
APPEND BIODAT PUBLIC	A OF ST	UDENT		85 86 87

## LIST OF TABLES

Table		Page	
2.1	Characteristics of dura, tenera and pisifera fruit forms	7	
3.1	Herbicides used	30	
3.2	Concentration of Herbicides	32	
4.1	List of target sequences	49	
4.2	List of primers used in qRT-PCR	50	
4.3	Treatment and concentration of hormones and inhibitors	54	
4.4	Disease severity scoring in <i>Ganoderma</i> - inoculated oil palm seedlings	55	
4.5	Putative identities and functions of cDNA sequences	58	
4.6	PCR amplification efficiencies for qRT-PCR	71	
4.7	Disease Severity Index (DSI) based on root symptoms in <i>Ganoderma</i> -infected oil palm seedlings	77	
4.8	Root surface of treated <i>Ganoderma</i> - inoculated roots with disease symptoms 3 wpi	78	
4.9	Root surface of treated <i>Ganoderma</i> - inoculated roots with disease symptoms 6 wpi	79	
4.10	Root surface of treated <i>Ganoderma</i> - inoculated roots with disease symptoms 12 wpi	81	

## LIST OF FIGURES

Figure		Page
2.1	Plant recognition of pathogen-associated molecular patterns (PAMPs) and pathogen effectors as signals of invasion.	13
3.1	Experimental layout to study the effect of herbicide on oil palm seedlings	32
3.2	Effect of different concentrations of GLY on growth of <i>Ganoderma</i> spp.	34
3.3	Effect of GLY on the growth of <i>Ganoderma</i> spp. at 3 dpi and 7 dpi.	35
3.4	Effect of different concentrations of GLA on growth of <i>Ganoderma</i> spp.	36
3.5	Effect of GLA on the growth of <i>Ganoderma</i> spp. at 3 dpi and 7 dpi.	36
3.6	Effect of different concentrations of PQ on growth of <i>Ganoderma</i> spp.	37
3.7	Effect of PQ on the growth of <i>Ganoderma</i> spp. at 3 dpi and 7 dpi.	38
3.8	Effects of Herbicides on Oil Palm Seedlings Inoculated with <i>G. boninense</i> PER71 at 22 wpi, scale = 2cm	40
3.9	Mycelia colonization and basidiocarp formation at basal region in two of the GLA treated oil palms seedlings inoculated with <i>G.</i> <i>boninense</i> PER71 at 13, 18, 19 and 21 wpi	41
3.10	Comparison of oil palm seedlings treated with herbicides inoculated with <i>G. boninense</i> PER71 and uninoculated seedlings.	42
3.11	Sectioning of oil palm seedlings treated with herbicides inoculated with <i>G. boninense</i> PER71 and control.	43
4.1	Multiple sequence alignment of EgRBOHB1 with RBOHs from other monocotyledons.	60
4.2	Multiple sequence alignment of EgRBOHB2 with RBOHs from other monocotyledons.	61
4.3	Multiple sequence alignment of EgRBOHH with RBOHs from other monocotyledons.	62
4.4	Multiple sequence alignment of EgHIR1 with HIR1 from other monocotyledons	63
4.5	Multiple sequence alignment of EgACO1 with ACO1 from other monocotyledons.	65
4.6	Multiple sequence alignment of EgOPR5 with OPR from other monocotyledons.	66
4.7	Multiple sequence alignment of EgCOI1 with COI1 from other monocotyledons	68
4.8	Multiple sequence alignment of EgNPR1 with NPR1 from other monocotyledons.	69

4.9	TAE gel electrophoresis of total RNA from oil palm root tissues.	76
4.10	Gene expression profiles in oil palm root	78
	samples upon <i>G. boninense</i> PER71	
	inoculation at 3, 6 and 12 wpi.	
4.11	Comparison of $H_2O_2$ in control (C) and G.	85
	<i>boninense</i> inoculated (G) oil palm seedling	
	treated with phytohormones and inhibitors at	
	3 wpi, 6 wpi and 12 wpi.	



 $\bigcirc$ 

## LIST OF ABBREVIATIONS

°C % A260nm µg µL BLAST bp BSR C:I	Degree celsius Percentage Optical density at wavelength 260 nanometer Microgram Microliter Micromoles Basic local alignment search tool Base pair Basal Stem Rot Chloroform : Isoamylalcohol
CaCl2	Calcium chloride
cDNA	Complementary deoxyribonucleic acid
cm	Centimetre
Ct	Threshold cycle
CTAB	Cetyltrimethylammonium Bromide
CWDE	Cell wall degrading enzyme
DAMP	Damage associated molecular protein
DEPC	Diethylpyrocarbonate
DNA	Deoxyribonucleic acid
DNase	Deoxyribonuclease I
dNTP	deoxynucleoside triphosphate
EDTA	Ethylenediaminetetraacetic acid
E	Efficiency
EST	Expressed sequence tag
ET	Ethylene
ETI	Effector-triggered immunity
ETS	Effector-triggered susceptibility
g	Gram
GLA	Glufosinate ammonium
GLY	Glyphosate
H2O2	Hydrogen Peroxide
HR	Hypersensitive response
JA	Jasmonic Acid
K	Potassium
kDA	Kilodaltons
L	Litre
LiCl2	Lithium chloride
M	Molar
mA	MilliAmps
MAMP	Microbe-associated molecular pattern
mg	Miligram
min	Minute
mm	Milimetre
mM	Milimolar
MPOB	Malaysian Palm Oil Board
NADPH	Nicotinamide adenine dinucleotide phosphate
NCBI	National Centre for Biotechnology Information

ng	Nanogram
OD	Optical density
ORF	Open reading frame
PAL	Phenylalanine ammonia lyase
PAMP	Pathogen-associated molecular pattern
PBZ	Paclobutrazol
P:C:I	phenol:chloroform:isoamylalcohol
PCR	Polymerase chain reaction
PDA	Potato dextrose agar
qRT	Quantitative reverse transcription
RNA	Ribosome inactivating protein
rpm	Revolutions per minute
RT	Reverse transcriptase
s SA SDS TAE U UV v/v w/v wpi × g UPM	second Salicylic acid Systemic acquired resistance Sodium dodecyl sulphate Tris-acetate-EDTA Unit Ultraviolet volume/volume weight/volume week post inoculation times gravity Universiti Putra Malaysia

## CHAPTER 1

### INTRODUCTION

#### 1.1 Introduction

Oil palm (*Elaeis guineensis* Jacq.) is the major agricultural crop grown in Malaysia with 5.81 million hectares of land used for cultivation nationwide (MPOB, 2017). Despite all the efforts to increase the yield of palm oil to meet the increasing demand, basal stem rot (BSR) caused by *Ganoderma* sp. (Hasan *et al.*, 2005) still remains as the major devastating disease that reduces not only the productivity but also the normal economic life span of the palm. Successive replanting on the same land after removing the previously BSR infected stands has increased the disease incidence of BSR as the infective tissues, inoculums and spores of *Ganoderma* spp. still remain at the sites (Turner, 1981; Ariffin *et al.*, 2000). Besides that, *Ganoderma* spp. have shown the ability to adapt to various environment and food sources (Wong *et al.*, 2012) turning this notorious disease a nightmare to the planters. To date, no effective treatment has been reported to be able to curb this disease.

Herbicides are inevitable inputs to control weeds in oil palm plantations. Although herbicides are effective in controlling weed population, administration of these synthetic chemicals may alter the soil microbial community causing potential increase of opportunistic plant pathogens. Moreover, herbicides may also have non-targeted effects on the cultivated crops making them more susceptible to diseases (Johal and Huber, 2009). Previous studies showed that white rot fungi are more tolerant to herbicides as they produce lignin degrading enzymes that are highly oxidative, non-specific and are able to transform a wide range of herbicides (Kersten and Cullen, 2007; Asgher et al., 2008; Moreira et al., 2013). Besides that, this group of fungi can grow on agricultural waste substrates. However, the influence of the herbicide usage on soil microbial ecosystem and interactions of oil palm and pathogenic white rot fungi such as *Ganoderma* in modulating disease development is rarely reported.

Plants naturally activate their defense mechanism in response to biotic and abiotic stresses. Hydrogen peroxide ( $H_2O_2$ ) and other reactive oxygen species (ROS) are the key components of plant defense response (Bhattacharjee, 2012). This rapid accumulation of  $H_2O_2$  or also known as 'oxidative burst' caused by environmental or developmental stresses (abiotic, biotic or physiological) regulates a specific subset of transcription factors that initiate changes in gene expression resulting in activation of various mechanisms including plant defense (Desikan *et al.*, 2000; Neill *et al.*, 2001). Plant immunity is dependent on the interaction between  $H_2O_2$  and other components such as nitric oxide and plant phytohormones as oxidative burst alone is not sufficient to trigger defense pathways (Delledonne *et al.*, 2001).

Previous studies have shown that phytohormones mainly salicylic acid (SA), jasmonic acid (JA) and ethylene (ET) contribute in plant defense through the ROS production triggered by pathogen-associated molecular patterns (PAMPs) and effectors (Kunkel and Brooks, 2002; Mur *et al.*, 2006; Pieterse *et al.*, 2009; Janda, 2015). Although phytohormones play a key role in plant defense mechanism, their roles during various plant-pathogen interaction especially oil palm-*Ganoderma* interaction remains widely uncharacterized (van Loon *et al.*, 2006).

BSR caused by Ganoderma has been studied for decades since its first detection. However, the interaction between oil palm and pathogen, Ganoderma is still not well understood or characterized. In recent years, the availability of oil palm genome sequence, gene expression analysis using expressed sequence tags (ESTs), and studies on differentially expressed genes (DEGs) in Ganoderma-inoculated root tissues have provided insights on the expression of defense related genes. Various pathogenesis and defense related genes such as chitinases, glucanases, isoflavone reductase, metallothioneins, metallothionein-like protein, early methionine-labelled polypeptides, type 2 ribosome inactivating proteins, Bowman Birk serine protease inhibitor, cysteine and nitric oxide associated 1 protein have been reported to be differentially expressed in Ganoderma-inoculated oil palm root tissues (Naher et al., 2011, Yeoh et al., 2012; Tan et al., 2013; Tee et al., 2013, Kwan et al., 2015). Genes encoding signal perception and transduction, phytohormone biosynthesis and signalling, ROS biosynthesis and scavenging, transcription factor, secondary metabolite production, and many others that are involved in defense responses were also reported among the DEGs of Ganoderma-inoculated oil palm roots in comparison with the uninoculated roots (Ho et al., 2016). These up and down-regulated DEGs could either be involved in the oil palm defense mechanism against Ganoderma invasion or a result of host defense suppression caused by the fungal pathogen.

In this study, the effects of herbicides, phytohormones and their inhibitors were monitored in infected oil palm roots by recording the morphological changes and disease severity. The expression of genes related to oxidative burst, SA and JA biosynthesis and signaling were profiled using *Ganoderma*-infected oil palm root samples to investigate the involvement of  $H_2O_2$  and phytohormones during defense response of oil palm seedlings against *Ganoderma boninense* at 3, 6 and 12 week post inoculation (wpi). Analyzing the expression of genes involved during the initial stage of infection such as genes encoding hypersensitive-induced reaction (HIR) protein, proteins related to oxidative burst, phytohormone biosynthesis and signaling proteins following the pathogen challenge may provide information on the role of  $H_2O_2$  and phytohormones during oil palm *Ganoderma* interaction.

## 1.2 Objectives of study

- 1. To investigate the effects of herbicides, phytohormones and their inhibitors on basal stem rot disease progression in oil palm roots;
- 2. To profile the expression of transcripts related to hydrogen peroxide production, hydrogen peroxide-induced cell death, hormone biosynthesis and signaling in Ganoderma infected oil palm roots.



### REFERENCES

- Adams, D. O. and Yang, S. F. 1979. Ethylene biosynthesis: identification of 1aminocyclopropane-1-carboxylic acid as an intermediate in the conversion of methionine to ethylene. *Proceedings of the National Academy of Sciences*, USA 76:170-174.
- Ade, J., DeYoung, B.J., Golstein, C. and Innes, R.W. 2007. Indirect activation of a plant nucleotide binding site-leucine-rich repeat protein by a bacterial protease. Proceedings of the National Academy of Sciences, USA 104: 2531–6.
- Agrios, G., N, 2005. Plant pathology. *Fifth Edition* Elsevier Academic Press, London, United Kingdom.
- Ahn, I. P. 2008. Glufosinate ammonium-induced pathogen inhibition and defense responses culminate in disease protection in bar-transgenic rice. *Plant Physiology* 146: 213-227.
- Alibhai, M. F. and Stallings, W. C. 2001. Closing down on glyphosate inhibition—with a new structure for drug discovery. *Proceedings of the National Academy of Sciences* 98: 2944-2946.
- Al-Momany, B., and Abu-Romman, S. 2016. Homologs of old yellow enzyme in plants. *Australian Journal of Crop Science* 10: 584.
- Altman, J., and Campbell, C. L. 1977. Effect of herbicides on plant diseases. Annual Review of Phytopathology 15: 361-385.
- Alvarez, M.E., Pennell, R.I., Meijer, P.J., Ishikawa, A., Dixon, R.A., and Lamb, C. 1998. Reactive oxygen intermediates mediate a systemic signal network in the establishment of plant immunity. *Cell* 92: 773–784.
- Apel, K., and Hirt, H. 2004. Reactive oxygen species: metabolism, oxidative stress, and signal transduction. *Annual Review in Plant Biology* 55: 373 – 399.
- Apostol, I., Heinstein, P.F., Low, P.S. 1989. Rapid stimulation of an oxidative burst during elicidation of cultured plant cells. Role in defense and signal transduction. *Plant Physiology* 90: 106-16.
- Arif, M. S., Roslan, A., Idris, A. S. and Ramle, M.. Economics of oil palm pests and Ganoderma disease and yield losses. In: *Proceedings of the Third MPOB-IOPRI International Seminar: Integrated Oil Palm Pests and Diseases Management.* Convention Centre, *Kuala Lumpur.* November 2011.
- Ariffin, D., Idris, A. S. and Singh, G. 2000. Status of *Ganoderma* in Oil Palm. In J. Flood, P. D. Bridge and M. Holderness (Eds.), *Ganoderma diseases of perennial crops*, 49-68. CABI, United Kingdom.

- Asai, S., Ohta, K., and Yoshioka, H. 2008. MAPK signaling regulates nitric oxide and NADPH oxidase-dependent oxidative bursts in *Nicotiana benthamiana*. *The Plant Cell* 20:1390-1406.
- Asai, S., and Yoshioka, H. 2009. Nitric oxide as a partner of reactive oxygen species participates in disease resistance to necrotrophic pathogen *Botrytis cinerea* in *Nicotiana benthamiana*. *Molecular Plant-Microbe Interactions* 22:619-629.
- Asgher, M., Bhatti, H. N., Ashraf, M., and Legge, R. L. 2008. Recent developments in biodegradation of industrial pollutants by white rot fungi and their enzyme system. *Biodegradation* 19: 771-783.
- Basiron, Y. 2007. Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology* 109: 289-295.
- Bending, G. D., Friloux, M. and Walker, A. 2002. Degradation of contrasting pesticides by white rot fungi and its relationship with ligninolytic potential. *FEMS Microbiology Letters* 212: 59-63.
- Berova, M and Zlatev, Z. 2000. Physiological response and yield of Paclobutrazol treated tomato plants (*Lycopersicon esculentum* Mill.). *Plant Growth Regulation* 30: 117-123.
- Black, B. D., Russin, J. S., Griffin, J. L., and Snow, J. P. 1996. Herbicide effects on *Rhizoctonia solani* in vitro and *Rhizoctonia* foliar blight of soybean (*Glycine max*). Weed Science 44: 711-716.
- Boller, T. 1995. Chemoperception of microbial signals in plant cells. *Annual Review of Plant Biology* 46: 189-214.
- Borggaard, O. K., and Gimsing, A. L. 2008. Fate of glyphosate in soil and the possibility of leaching to ground and surface waters: a review. *Pest Management Science*, 64: 441-456.
- Coelho, D., J., de Souza, C. G. M., de Oliveira, A. L., Bracht, A., Costa, M. A. F., and Peralta, R. M. 2010. Comparative removal of Bentazon by *Ganoderma lucidum* in liquid and solid state cultures. *Current Microbiology* 60: 350-355.
- Barcelos, E., de Almeida Rios, S., Cunha, R.N., Lopes, R., Motoike, S.Y., Babiychuk, E. and Kushnir, S. 2015. Oil palm natural diversity and the potential for yield improvement. *Frontiers in Plant Science* 6: 190.
- Berrocal-Lobo, M., Stone, S., Yang, X., Antico, J., Callis, J., Ramonell, K. M., and Somerville, S. 2010. ATL9, a RING zinc finger protein with E3 ubiquitin ligase activity implicated in chitin-and NADPH oxidase-mediated defense responses. *PloS one* 5:e14426.
- Bhattacharjee, S. 2012. The language of reactive oxygen species signalling in plants. *Journal of Botany* 1-22.

- Bradley, D. J., Kjellbom, P and Lamb, C. J. 1992. Elicitor and wound induced oxidative cross-linking of a proline rich plant cell wall protein: a novel rapid defense response. *Cell* 70: 21-30.
- Bruinsma, M., Van Broekhoven, S., Poelman, E. H., Posthumus, M. A., Müller, M. J., Van Loon, J. J., and Dicke, M. 2010. Inhibition of lipoxygenase affects induction of both direct and indirect plant defenses against herbivorous insects. *Oecologia* 162: 393-404.
- Bolwell, G.P., Bindschedler, L.V., Blee, K.A., Butt, V.S., Davies, D.R., Gardner, S.L., Gerrish, C., and Minibayeva, F. 2002. The apoplastic oxidative burst in response to biotic stress in plants: a tree component system. *Journal of Experimental Botany* 53:1367-1376
- Carol, R. J., Takeda, S., Linstead, P., Durrant, M. C., Kakesova, H., Derbyshire P., Drea, S., Zarsky, V. and Dolan, L. 2005. A RhoGDP dissociation inhibitor spatially regulates growth in root hair cells. *Nature* 438: 1013-1016.
- Carvalho, F. P. 2017. Pesticides, environment, and food safety. *Food and Energy Security* 6: 48-60.
- Cheng, C., Xu, X., Gao, M., Li, J., Guo, C., Song, J., and Wang, X. 2013. Genome-wide analysis of respiratory burst oxidase homologs in grape (*Vitis vinifera* L.). *International Journal of Molecular Sciences* 14: 24169-24186.
- Cheong, J., Wallwork, H., and Williams, K. J. 2004. Identification of a major QTL for yellow leaf spot resistance in the wheat varieties Brookton and Cranbrook. *Australian Journal of Agriculture Resources* 55:315-319.
- Cooper, R. M. 1984. The role of cell wall degrading enzymes in infection and damage. In Wood, R. K. S., and Jellis, G. J. (Eds.), *Plant Disease: Infection, Damage and Loss*, 13-27. Blackwell Scientific Publication. Oxford, USA.
- Cooper, R. M., Flood, J. and Rees, R. W. 2011. Ganoderma boninense in oil palm plantations: current thinking on epidemiology, resistance and pathology. *Planter* 87: 515-526.
- Cork, D.J. and Krueger, J.P. 1991. Microbial transformation of herbicides and pesticides. *Advances in Applied Microbiology* 36: 1-66.
- Creelman, R. A., and Mullet, J. E. 1997. Biosynthesis and action of jasmonates in plants. *Annual Review of Plant Biology* 48: 355-381.
- Crawford, N. M. 1995. Nitrate: nutrient and signal for plant growth. *The Plant Cell* 7:859.
- Dangl, J. L. and Jones, J. D. 2001. Plant pathogens and integrated defense responses to infection. *Nature* 411: 826-833.

- da Silva Coelho, J., de Oliveira, A. L., de Souza, C. G. M., Bracht, A., and Peralta, R. M. 2010. Effect of the herbicides bentazon and diuron on the production of ligninolytic enzymes by *Ganoderma lucidum*. *International Biodeterioration and Biodegradation* 64:156-161.
- Datnoff, L.E., Elmer, W.H. and Huber, D.M. 2007. *Mineral Nutrition and Plant Disease*. APS Press, St. Paul, Minnesota.
- Davis, E.L., Hussey, R.S. and Baum, T.J. 2004. Getting to the roots of parasitism by nematodes. *Trends in Parasitology* 20: 134–41.
- De Coninck, B., Timmermans, P., Vos, C., Cammue, B. P., and Kazan, K. 2015. What lies beneath: below ground defense strategies in plants. *Trends in Plant Science* 20: 91-101.
- Desikan, R., Neill, S.J. and Hancock, J.T. 2000. Hydrogen peroxide-induced gene expression in *Arabidopsis* thaliana. Free Radical Biology and Medicine 28: 773-778.
- DeYoung, B.J. and Innes, R.W. 2006. Plant NBS-LRR proteins in pathogen sensing and host defense. *Nature Immunology* 7: 1243–9.
- Devine, M., Duke, S. O., and Fedtke, C. 1992. *Physiology of Herbicide Action*. PTR Prentice Hall, Englewood, New Jersey.
- Dill, G.M. 2005. Glyphosate-resistant crops: history, status and future. *Pest Management Science* 61: 219-224.
- Egusa, M., Ozawa, R., Takabayashi, J., Otani, H. and Kodama M. 2009. The jasmonate signalling pathway in tomato regulates susceptibility to a toxin-dependent necrotrophic pathogen. *Planta* 229: 965–976.
- Feng, S., Ma, L., Wang, X., Xie, D., Dinesh-Kumar, S. P., Wei, N., and Deng, X.
  W. 2003. The COP9 signalosome interacts physically with SCFCOI1 and modulates jasmonate responses. *The Plant Cell* 15: 1083-1094.
- Feys, B. J., and Parker, J. E. 2000. Interplay of signaling pathways in plant disease resistance. *Trends in Genetics* 16: 449-455.
- Flor, H.H. 1971. Current Status of the Gene-For-Gene Concept. *Annual Review* of *Phytopathology* 9: 275-296
- Frear, D. S., Swanson, H. R., and Tanaka, F. S. 1969. N-demethylation of substituted 3-(phenyl)-1-methylureas: isolation and characterization of a microsomal mixed function oxidase from cotton. *Phytochemistry* 8: 2157-2169.
- Funderburk Jr, H. H., and Lawrence, J. M. 1964. Mode of action and metabolism of diquat and paraquat. *Weeds* 12: 259-264.

- Gadjev, I., Stone, J. M. and Gechev, T. S. 2008. Programmed cell death in plants: new insights into redox regulation and the role of hydrogen peroxide. *International Review of Cell and Molecular Biology* 270: 87-144
- Galletti, R., Denoux, C., Gambetta, S., Dewdney, J., Ausubel, F. M., De Lorenzo, G., and Ferrari, S. 2008. The Atrohoh-mediated oxidative burst elicited by oligogalacturonides in *Arabidopsis* is dispensable for the activation of defense responses effective against *Botrytis cinerea*. *Plant Physiology* 148:1695-1706.

Gao, Q. M., Zhu, S., Kachroo, P., and Kachroo, A. 2015. Signal regulators of systemic acquired resistance. *Frontiers in Plant Science* 6: 228.

- Gimenez-Ibanez, S., and Solano, R. 2013. Nuclear jasmonate and salicylate signaling and crosstalk in defense against pathogens. *Frontiers in Plant Science* 4: 72.
- Glazebrook, J. 2005. Contrasting mechanisms of defense against biotrophic and necrotrophic pathogens. *Annual Review in Phytopathology* 43: 205-227.
- Greenberg, J. T. 1997. Programmed cell death in plant-pathogen interactions. *Annual Review of Plant Biology* 48: 525-545.
- Gomes, M. P., Smedbol, E., Chalifour, A., Hénault-Ethier, L., Labrecque, M., Lepage, L., Lucotte, M. and Juneau, P. 2014. An overview: Alteration of plant physiology by glyphosate and its by-product aminomethylphosphonic acid. *Journal of Experimental Botany* 65: 4691-4703.
- Govrin, E.M., Levine, A. 2000. The Hypersensitive response facilitates plant infection by the necrotrophic pathogen *Botrytis cinerea*. *Current Biology* 10: 751-757.
- Hasan, Y. and Flood J. 2003. Colonisation of rubberwood and oil palm blocks by monokaryons and dikaryons of *Ganoderma boninense* implications to infection in the field. *The Planter* 79: 31-8.
- Hasan, Y., Foster, H. L., and Flood, J. 2005. Investigations on the causes of upper stem rot (USR) on standing mature oil palms. *Mycopathologia* 159: 109-112.

Hashim, K., and Chong, T. C. 2008. An overview of the current status of *Ganoderma* basal stem rot and its management in a large plantation group in Malaysia. *Planter*, 469-482. Incorporated Society of Planters. Malaysia Hartley, C.W.S. 1977. *The Oil Palm*. Longman. London.

- Halliwell, B. 2006. Oxidative stress and neurodegeneration: where are we now? *Journal of Neurochemistry* 97: 1634–1658
- Hammel, K. E., Kapich, A. N., Jensen, K. A., and Ryan, Z. C. 2002. Reactive oxygen species as agents of wood decay by fungi. *Enzyme and Microbial Technology* 30: 445-453.

- Harikrishnan, R., and Yang, X. B. 2001. Influence of herbicides on growth and sclerotia production in *Rhizoctonia solani*. *Weed Science* 49:241-247.
- Hassall, K. A. 1982. The Chemistry of Pesticides: Their Metabolism, Mode of Action and Uses in Crop Protection. Macmillan. Basingstoke and London.
- Hayat, Q., Hayat, S., Irfan, M., and Ahmad, A. 2010. Effect of exogenous salicylic acid under changing environment: a review. *Environmental and Experimental Botany* 68: 14-25.
- Hepting, G. H. 1971. Diseases of forest and shade trees of the United States. Agriculture Handbook US Department of Agriculture.USA.
- Hiratsuka, N., Wariishi, H., and Tanaka, H. 2001. Degradation of diphenyl ether herbicides by the lignin-degrading basidiomycete *Coriolus versicolor*. *Applied Microbiology and Biotechnology* 57: 563-571.
- Ho, C. L., Tan, Y. C., Yeoh, K. A., Ghazali, A. K., Yee, W. Y., and Hoh, C. C. 2016. *De novo* transcriptome analyses of host-fungal interactions in oil palm (*Elaeis guineensis* Jacq.). *BMC Genomics* 17: 66.
- Hofrichter, M. 2002. Review: lignin conversion by manganese peroxidase (MnP). *Enzyme and Microbial Technology* 30: 454-466.
- Huber, D.M., and Haneklaus, S. 2007. Managing nutrition to control plant disease. *Landbauforschung Volkenrode* 57: 313.
- Idris, S., Arifin, D., Swinburne, R. and Watt, A. A. 2000. The identity of Ganoderma species responsible for BSR disease of oil palm in Malaysia. Morphological Characteristics. *Malaysian Palm Oil Board* 102: 77a.
- Idris, A.S. and Ariffin, D. 2003. *Ganoderma*: Basal stem rot disease and disease control. *Oil Palm Bulletin* 11: 1-10
- Idris, A., Kushairi, A., Ismail, S. and Ariffin, D. 2004. Selection for partial resistance in oil palm progenies to *Ganoderma* basal stem rot. *Journal of Oil Palm Research* 16:12–18.
- Idris, A. S., Kushairi, D., Ariffin, D., and Basri, M. W. 2006. Technique for inoculation of oil palm germinated seeds with *Ganoderma*. MPOB Information Series 314: 1-4.
- Iwamoto, M., and Takano, M. 2011. Phytochrome-regulated EBL1 contributes to ACO1 upregulation in rice. *Biotechnology letters* 33: 173-178.
- Janda, M., and Ruelland, E. 2015. Magical mystery tour: salicylic acid signalling. *Environmental and Experimental Botany* 114: 117-128.

- Jauregui, J., Valderrama, B., Albores, A., and Vazquez-Duhalt, R. 2003. Microsomal transformation of organophosphorus pesticides by white rot fungi. *Biodegradation* 14: 397-406.
- Johal, G. S., and Huber, D. M. 2009. Glyphosate effects on diseases of plants. *European Journal of Agronomy* 31: 144-152.
- Jones, J. D., and Dangl, J. L. 2006. The plant immune system. *Nature* 444: 323-329.
- Kachroo, A., He, Z., Patkar, R., Zhu, Q., Zhong, J., Li, D. and Chattoo, B. B. 2003. Induction of H<sub>2</sub>O<sub>2</sub> in transgenic rice leads to cell death and enhanced resistance to both bacterial and fungal pathogens. *Transgenic Research* 12: 577-586.
- Karrer, E. E., Beachy, R. N., and Holt, C. A. 1998. Cloning of tobacco genes that elicit the hypersensitive response. *Plant Molecular Biology* 36: 681-690.
- Kersten, P., and Cullen, D. 2007. Extracellular oxidative systems of the lignindegrading Basidiomycete *Phanerochaete chrysosporium. Fungal Genetics and Biology* 44: 77-87.
- Keen, N. T. 1990. Gene-for-gene complementarity in plant-pathogen interactions. *Annual Review of Genetics* 24: 447-463.
- Kishore , G. M. and Shah D. M. 1988. Amino acid biosynthesis inhibitors as herbicides. *Annual Review of Biochemistry* 57: 627 663.
- Kobayashi, M., Ohara, I., Kawakita, K., Yokota, N., Fujiwara, M., Shimamoto, K. , Doke, N., and Yoshioka, H. 2007. Calcium-dependent protein kinases regulate the production of reactive oxygen species by potato NADPH oxidase. *The Plant Cell* 19: 1065-1080.
- Koressaar, T. and Remm, M. 2007. Enhancements and modifications of primer design program Primer3. *Bioinformatics* 23:1289-91
- Kuai, X., and C. Després. 2016. Defining *Arabidopsis* NPR1 orthologues in crops for translational plant immunity. *Canadian Journal of Plant Pathology* 38: 25-30.
- Kunkel, B.N., and Brooks, D.M. 2002. Cross talk between signaling pathways in pathogen defense. *Current Opinion in Plant Biology* 5: 325–331.
- Kumar, G.N., Iyer, S. and Knowles, N. R. 2007. StrbohA homologue of NADPH oxidase regulates wound-induced oxidative burst and facilitates wound-healing in potato tubers. *Planta* 227:25-36.
- Kushairi, A. 1992. Prestasi baka kelapa sawit *dura* × *pisifera* di Malaysia.M.Sc. Thesis. Universiti Kebangsaan Malaysia, Selangor, Malaysia.
- Kwan, Y. M., Meon, S., Ho, C. L., and Wong, M. Y. 2015. Cloning of nitric oxide associated 1 (NOA1) transcript from oil palm (*Elaeis guineensis*) and its

expression during *Ganoderma* infection. *Journal of Plant Physiology* 174: 131-136.

- Lam, H.M., Coschigano, K., Schultz, C., Melo-Oliveira, R., Tjaden, G., Oliveira, I., Ngai, N., Hsieh, M.H. and Coruzzi, G. 1995. Use of *Arabidopsis* mutants and genes to study amide amino acid biosynthesis. *The Plant Cell* 7: 887.
- Lamb, C., and Dixon, R. A. 1997. The oxidative burst in plant disease resistance. *Annual Review of Plant Biology* 48: 251-275.
- Lasserre, E., Bouquin, T., Hernandez, J. A., Pech, J. C., Balagué, C., and Bull, J. 1996. Structure and expression of three genes encoding ACC oxidase homologs from melon (*Cucumis melo* L.). *Molecular and General Genetics* 251:81-90.
- Lee, Y., Rubio, M. C., Alassimone, J., and Geldner, N. 2013. A mechanism for localized lignin deposition in the endodermis. *Cell* 153: 402-412.
- Li, C., Williams, M.M., Loh, Y.T., Lee, G.I., Howe, G.A. 2002. Resistance of cultivated tomato to cell content-feeding herbivores is regulated by the octadecanoid-signaling pathway. *Plant Physiol*ogy 130: 494–503
- Li, W., Zhou, F., Liu, B., Feng, D., He, Y., Qi, K., and Wang, J. 2011. Comparative characterization, expression pattern and function analysis of the 12-oxo-phytodienoic acid reductase gene family in rice. *Plant Cell Reports* 30: 981-995
- Liu, Y. H., Offler, C. E., and Ruan, Y. L. 2014. A simple, rapid, and reliable protocol to localize hydrogen peroxide in large plant organs by DAB-mediated tissue printing. *Frontiers in Plant Science* 5: 745.
- Loake, G., and Grant, M. 2007. Salicylic acid in plant defense—the players and protagonists. *Current Opinion in Plant Biology* 10: 466-472.
- López-Berges, M. S., Rispail, N., Prados-Rosales, R. C., and Di Pietro, A. 2010. A nitrogen response pathway regulates virulence functions in *Fusarium oxysporum* via the protein kinase TOR and the bZIP protein MeaB. *The Plant Cell* 22: 2459-2475.
- Lorang, J.M., Sweat, T.A. and Wolpert, T.J. 2007. Plant disease susceptibility conferred by a 'resistance' gene. *Proceedings of the National Academy of Sciences*. USA. 104: 14861-14868.
- Obahiagbon, F. I. 2012. A review: aspects of the African oil palm (*Elaeis guineesis* jacq.) and the implications of its bioactives in human health. *American Journal of Biochemistry and Molecular Biology* 2: 106-119.
- MacDonald, G. E., Gettys, L. A., Ferrell, J. A., and Sellers, B. A. 2013. Herbicides for Natural Area Weed Management. In A.J. Price, J.A. Kelton

(Eds.), *Herbicides- Current Research and Case Studies in Use,* 203–239. InTech Publishers. Croatia.

- Maciel, G. M., Bracht, A., Souza, C. G. M., Costa, A. M., and Peralta, R. M. 2012. Fundamentals, diversity and application of white-rot fungi. In Silva A. P., Sol, M., (Eds.), *Fungi: Types, Environmental Impact and Role in Disease*, 409-457. Nova Science Publishers, New York.
- Maheshwari, R. 2005. *Fungi: Experimental Methods in Biology*. CRC press, United States of America.
- Malaysian Palm Oil Board (MPOB). 2017. Oil Palm Planted Area by States as At December 2017. Economics and Industry Development Division, Malaysia. Retrieved from http://bepi.mpob.gov.my/images/area/2017/Area summary.pdf
- Malaysian Palm Oil Council (MPOC). 2012. The Oil Palm Tree. Retrieved from http://www.mpoc.org.my/The\_Oil\_Palm\_Tree.aspx
- Marmagne, A., Ferro, M., Meinnel, T., Bruley, C., Kuhn, L., Garin, J., Brygoo, H. B., and Ephritikhine, G. 2007. A high content in lipid-modified peripheral proteins and integral receptor kinases features in the *Arabidopsis* plasma membrane proteome. *Molecular and Cellular Proteomics* 6: 1980-1996.
- Matsuzaki, F., and Wariishi, H. 2004. Functional diversity of cytochrome P450s of the white-rot fungus *Phanerochaete chrysosporium*. *Biochemical and Biophysical Research Communications* 324: 387-393.
- McDowell, J. M. and Simon, S. A. 2008. Molecular diversity at the plant– pathogen interface. *Developmental and Comparative Immunology* 32: 736-744.
- Mengiste, T. 2012. Plant immunity to necrotrophs. Annual Review of Phytopathology 50: 267-294.
- Mih, A. M., and Kinge, T. R. 2015. Ecology of Basal Stem Rot Disease of Oil Palm (*Elaeis guineensis* Jacq.) in Cameroon. *American Journal of Agriculture and Forestry* 3: 208-215.
- Mindrinos, M., Katagiri, F., Yu, G. L. and Ausubel, F. M. 1994. The A. thaliana disease resistance gene RPS2 encodes a protein containing a nucleotidebinding site and leucine-rich repeats. *Cell* 78: 1089-1099.
- Mohan, D., Ho, P. Y., Ho, C. L., Namasivayam, P., and Saidi, N. B. 2017. Effects of herbicides on fungal phytopathogens. *Pertanika Journal of Scholarly Research Reviews*, 3: 93-101.
- Mohd, Z., & Faridah, A. 2008. Disease suppression in *Ganoderma*-infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science* 44: 101-107.

- Montanyá, S.I., Zambrana-Quesada, E., and Tenorio-Pasamón, J. L.2013. Weed Management in Cereals in Semi-Arid Environments. In *A Review: Herbicides–Current Research and Case Studies in Use*, 133-152. InTech Publishers, Croatia.
- Montiel, J., Nava, N., Cárdenas, L., Sánchez-López, R., Arthikala, M. K., Santana, O., and Quinto, C. 2012. A *Phaseolus vulgaris* NADPH oxidase gene is required for root infection by Rhizobia. *Plant and Cell Physiology* 53: 1751-1767.
- Moore, H. E. 1973. The major groups of palms and their distribution. *Gentes Herb* 11: 27-141.
- Moreira, D. S., J., Maciel, G. M., Castoldi, R., da Silva Mariano, S., Inácio, F. D., Bracht, A., and Peralta, R. M. 2013. Involvement of lignin-modifying enzymes in the degradation of herbicides, In A.J. Price, J.A. Kelton (Eds.), *Agricultural and Biological Sciences Herbicides-Advances in Research*, 165–187. InTech Publishers. Croatia.
- Mukhtar, M. S., Nishimura, M. T., and Dangl, J. 2009. NPR1 in plant defense: it's not over'til it's turned over. *Cell* **13**7:804-806.
- Mur, L.A., Kenton, P., Lloyd, A.J., Ougham, H. and Prats, E. 2008. The hypersensitive response; the centenary is upon us but how much do we know? *Journal of Experimental Botany* 59: 233-316.
- Mur, L. A., Kenton, P., Atzorn, R., Miersch, O. and Wasternack, C. 2006. The outcomes of concentration-specific interactions between salicylate and jasmonate signaling include synergy, antagonism, and oxidative stress leading to cell death. *Plant Physiology* 140:249-262.
- Naher, L., Yusuf, U. K., Ismail, A., Tan, S. G. and Mondal, M. M. A. 2013. Ecological status of *Ganoderma* and basal stem rot disease of oil palms (*Elaeis guineensis* Jacq.) Australian Journal of Crop Science 7: 1723.
- Nadimpalli, R., Yalpani, N., Johal, G. S., and Simmons, C. R. 2000. Prohibitins, stomatins, and plant disease response genes comprise a protein superfamily that controls cell proliferation, ion channel regulation, and death. *Journal of Biological Chemistry* 275: 29579-29586
- Nandula, V. K. 2010. *Glyphosate Resistance in Crops and Weeds: History, Development, and Management.* John Wiley & Sons, USA.
- Neill, S.J., Desikan, R., Clarke, A., Hurst, R.D., and Hancock, J.T. 2002. Hydrogen peroxide and nitric oxide as signalling molecules in plants. *Journal* of *Experimental Botany* 53: 1237-1247.
- Nelson, K. J., and Parsonage, D. 2011. Measurement of peroxiredoxin activity. *Current Protocols in Toxicology* 49: 7-10.

- Ng, Y. G., Shamsul Bahri, M. T., Irwan Syah, M. Y., Mori, I., and Hashim, Z. 2013. Ergonomics observation: Harvesting tasks at oil palm plantation. *Journal of Occupational Health* 55: 405-414.
- Oerke, E. C. 2006. Crop losses to pests. *The Journal of Agricultural Science* 144: 31-43.
- Ogasawara, Y., Kaya, H., Hiraoka, G., Yumoto, F., Kimura, S., and Kadota, Y. 2008.Synergistic activation of the *Arabidopsis* NADPH oxidase *AtrbohD* by Ca2b and phosphorylation. *Journal of Biological Chemistry* 283: 8885-8892.
- Parween, T., Jan, S., Mahmooduzzafar, S., Fatma, T., and Siddiqui, Z. H. 2016. Selective effect of pesticides on plant—A Review. *Critical Reviews in Food Science and Nutrition* 56: 160-179.
- Perchepied, L., Balagué, C., Riou, C., Claudel-Renard, C., Rivière, N., Grezes-Besset, B., and Roby, D. 2010. Nitric oxide participates in the complex interplay of defense-related signaling pathways controlling disease resistance to *Sclerotinia sclerotiorum* in *Arabidopsis thaliana*. *Molecular Plant-Microbe Interactions* 23:846-860.
- Pieterse, C. M. and Van Loon, L. C. 2004. NPR1: the spider in the web of induced resistance signaling pathways. *Current Opinion in Plant Biology* 7: 456-464.
- Pieterse, C.M.J., Leon-Reyes, A., VanderEnt, S., and Van Wees, S.C.M. 2009. Networking by small-molecule hormones in plant immunity. *Nature Chemical Biology* 5: 308–316.
- Pilotti, C. A. 2005. Stem rots of oil palm caused by *Ganoderma boninense*: Pathogen biology and epidemiology. *Mycopathologia* 159:129-137.
- Pizzul, L., del Pilar Castillo, M., and Stenström, J. 2009. Degradation of glyphosate and other pesticides by ligninolytic enzymes. *Biodegradation* 20: 751-759.
- Pogány, M., von Rad, U., Grün, S., Dongó, A., Pintye, A., Simoneau, P and Durner, J. 2009. Dual roles of reactive oxygen species and NADPH oxidase RBOHD in an *Arabidopsis-Alternaria* pathosystem. *Plant Physiology* 151:1459-1475.
- Pointing, S. 2001. Feasibility of bioremediation by white-rot fungi. *Applied Microbiology and Biotechnology*, 57: 20-33.
- Purseglove, J.W. 1972. *Tropical Crops, Monocotyledons,* 607. Longman, London.
- Quan, L. J., Zhang, B., Shi, W. W and Li, H. Y. 2008. Hydrogen peroxide in plants: a versatile molecule of the reactive oxygen species network. *Journal* of Integrative Plant Biology 50: 2-18.

- Qin, Y. M., Hu, C. Y., Pang, Y., Kastaniotis, A. J., Hiltunen, J. K., and Zhu, Y. X. 2007. Saturated very-long-chain fatty acids promote cotton fiber and *Arabidopsis* cell elongation by activating ethylene biosynthesis. *The Plant Cell* 19: 3692-3704.
- Rahe, J.E., Levesque, C.A. and Johal, G.S. 1990. Synergistic role of soil fungi in the herbicidal efficacy of glyphosate, In Hoagland, R.E. (Ed.), *Biological Weed Control Using Microbes and Microbial Products as Herbicides,* 260– 275. Symposium, April 9–14, 1989. American Chemical Society,Washington, DC.
- Rahman, A., Abdullah, R., Shariff, F. M., and Simeh, M. A. 2008. The Malaysian palm oil supply chain: the role of the independent smallholder. *Oil Palm Industry Economic Journal* 8: 17-27.
- Rashid, M., Choon-Fah, M. R., Bong, J., Khairulmazmi, A., and Idris, A. S. 2014. Genetic and morphological diversity of *Ganoderma* species isolated from infected oil palms (*Elaeis guineensis*). *International Journal of Agriculture and Biology* 16: 691-699.
- Rakib, M. R. M., Bong, C. F. J., Khairulmazmi, A., and Idris, A. S. 2015. Aggressiveness of *Ganoderma boninense* and *G. zonatum* isolated from upper-and basal stem rot of oil palm (*Elaeis guineensis*) in Malaysia. *Journal* of *Oil Palm Research* 27: 229-240.
- Rees, R. 2006. *Ganoderma* stem rot of oil palm (*Elaeis guineensis*): mode of infection, epidemiology and biological control. PhD thesis, University of Bath, United Kingdom.
- Rees, R. W., Flood, J., Hasan, Y., Potter, U. and Cooper, R. M. 2009. Basal stem rot of oil palm (*Elaeis guineensis*); mode of root infection and lower stem invasion by *Ganoderma boninense*. *Plant Pathology* 58: 982-9.
- Reymond, P. and Farmer, E. E. 1998. Jasmonate and salicylate as global signals for defense gene expression. *Current Opinion in Plant Biology* 1: 404-411.
- Rival, A., Beule, T., Barré, P., Hamon, S., Duval, Y., and Noirot, M. 1997. Comparative flow cytometric estimation of nuclear DNA content in oil palm (*Elaeis guineensis* Jacq) tissue cultures and seed-derived plants. *Plant Cell Reports* 16: 884-887.
- Roslan, A., and Idris, A. S. 2012. Economic impact of *Ganoderma* incidence on Malaysian oil palm plantation–a case study in Johor. *Oil Palm Industry Economic Journal* 12: 24-30.
- Santner, A., Calderon-Villalobos, L.I.A., and Estelle, M.2009. Plant hormones are versatile chemical regulators of plant growth. *Nature Chemical Biolog*, 5: 301–307.

- Santner, A., and Estelle, M. 2009. Recent advances and emerging trends in plant hormone signalling. *Nature* 459: 1071–1078.
- Sambanthamurthi, R., Singh, R., Kadir, A. P. G., Abdullah, M. O. and Kushairi, A. 2009. Opportunities for the oil palm via breeding and biotechnology. *Breeding plantation tree crops: Tropical species*, 377-421. Springer New York, USA.
- Schaller, F., Biesgen, C., Müssig, C., Altmann, T., and Weiler, E. W. 2000. 12-Oxophytodienoate reductase 3 (OPR3) is the isoenzyme involved in jasmonate biosynthesis. *Planta* 210: 979-984.
- Schönbrunn, E., Eschenburg, S., Shuttleworth, W. A., Schloss, J. V., Amrhein, N., Evans, J. N., and Kabsch, W. 2001. Interaction of the herbicide glyphosate with its target enzyme 5-enolpyruvylshikimate 3-phosphate synthase in atomic detail. *Proceedings of the National Academy of Sciences* 98: 1376-1380.
- Schwessinger, B. and Ronald, P. C. 2012. Plant innate immunity: perception of conserved microbial signatures. *Annual Review of Plant Biology* 63: 451-482.
- Shapiguzov, A., Vainonen, J., Wrzaczek, M., and Kangasjärvi, J. 2012. ROStalk-how the apoplast, the chloroplast, and the nucleus get the message through. *Frontiers in Plant Science* 3: 292.
- Senseman, S. 2007. Herbicide Handbook. *Weed Science Society of America*, 458. Lawrence, Kentucky State, USA.
- Seo, G. S., Kirk, P. M., Flood, J., Bridge, P. D. and Holderness, M. 2000. Ganodermataceae: nomenclature and classification. In Flood, J. (Ed.), *Ganoderma Diseases of Perennial Crops*, 3-22. Cabi Publishing, USA.
- Shulaev, V., Silvermann, P., and Raskin, I. 1997. Methyl salicylate—an airborne signal in pathogen resistance. *Nature* 385: 718-721.
- Shapiguzov A., Vainonen J. P., Wrzaczek M. and Kangasjarvi J. 2012. ROstalk - how the apoplast, the chloroplast, and the nucleus get the message through. *Frontier in Plant Sciences* 3:292.
- Shelpi, B. J., Swanton, C. J., Mersey, B. G., and Hall, J. C. 1992. Glufosinate (phosphinothricin) inhibition of N metabolism in barley and green foxtail plants. *Journal of Plant Physiology* 139: 605-610.
- Sherwani, S.I., Arif, I.A. and Haseeb, A.K. 2015. Modes of Action of Different Classes of Herbicides, In Andrew Price (Ed.), *Herbicides, Physiology of Action, and Safety*. InTech Publishers. Croatia.
- Smith, C. J. 1996. Accumulation of phytoalexins: defense mechanism and stimulus response system. *New Phytologist* 132: 1-45.

- Snoeijers, S. S., Pérez-García, A., Joosten, M. H., and De Wit, P. J. 2000. The effect of nitrogen on disease development and gene expression in bacterial and fungal plant pathogens. *European Journal of Plant Pathology*,106: 493-506.
- Stakman, E.C. 1915. Relation between *Puccinia graminis* and plants highly resistant to its attack. *Journal of Agricultural Research* 4: 193-201.
- Strassner, J., Schaller, F., Frick, U. B., Howe, G. A., Weiler, E. W., Amrhein, N., and Schaller, A. 2002. Characterization and cDNA-microarray expression analysis of 12-oxophytodienoate reductases reveals differential roles for octadecanoid biosynthesis in the local versus the systemic wound response. *The Plant Journal 32:* 585-601.
- Stintzi, A. 2000. The *Arabidopsis* male-sterile mutant, opr3, lacks the 12oxophytodienoic acid reductase required for jasmonate synthesis. *Proceedings of the National Academy of Sciences USA* 97: 10625-10630.
- Syed, R. A. 1979. Studies on oil palm pollination by insects. *Bulletin of Entomological Research* 69: 213-224.
- Tan, S., Evans, R., and Singh, B. 2006. Herbicidal inhibitors of amino acid biosynthesis and herbicide-tolerant crops. *Amino acids* 30: 195-204.
- Tan, Y. C., Yeoh, K. A., Wong, M. Y., and Ho, C. L. 2013. Expression profiles of putative defence-related proteins in oil palm (*Elaeis guineensis*) colonized by *Ganoderma boninense*. *Journal of Plant Physiology* 170: 1455-1460.
- Tee, S. S., Tan, Y. C., Abdullah, F., Ong-Abdullah, M., and Ho, C. L. 2013. Transcriptome of oil palm (*Elaeis guineensis* Jacq.) roots treated with *Ganoderma boninense. Tree Genetics and Genomes* 9: 377-386.
- Tennakoon, K. M. S., Jaspers, M. V., Ridgway, H. J., and Jones, E. E. 2015. Herbicide injuries on blueberry provide suitable infection sites for *Neofusicoccum ribis*. *New Zealand Plant Protection* 68: 411-414.
- Thatcher, L. F., Manners, J.M., Kazan, K. 2009. *Fusarium oxysporum* hijacks COI1-mediated jasmonate signaling to promote disease development in *Arabidopsis. The Plant Journal* 58: 927–939.
- Thines, B., Katsir, L., Melotto, M., Niu, Y., Mandaokar, A., Liu, G., and Howe, G. A. 2007. JAZ repressor proteins are targets of the SCF COI1 complex during jasmonate signalling. *Nature* 448: 661.
- Torres, M. A., Dangl, J. L., and Jones, J. D. 2002. *Arabidopsis* gp91phox homologues AtrohD and AtrohF are required for accumulation of reactive oxygen intermediates in the plant defense response. *Proceedings of the National Academy of Sciences* USA 99: 517-522.

- Torres, M.A. and Dangl, J.L. 2005. Functions of the respiratory burst oxidase in biotic interaction, abiotic stress and development. *Current Opinion in Plant Biology* 8: 397-403.
- Turner, P.D. 1981. *Oil Palm Diseases and Disorders*. Oxford University Press, Kuala Lumpur.
- Udvardi, M. K., Czechowski, T., and Scheible, W. R. 2008. Eleven golden rules of quantitative RT-PCR. *The Plant Cell* 20:1736-1737.
- Uma, B., Rani, T.S. and Podile A.R. 2011. Warriors at the gate that never sleep: non-host resistance in plants. *Journal of Plant Physiology* 168:2141-52.
- Untergasser, A., Cutcutache, I., Koressaar, T., Ye, J., Faircloth, B. C., Remm, M. and Rozen, S.G. 2012. Primer3-new capabilities and interfaces. *Nucleic Acids Research* 40: e115.
- Van Doorn, W. G., and Woltering, E. J. 2005. Many ways to exit? Cell death categories in plants. *Trends in Plant Science*, 10: 117-122.
- van Wees, S. C., Chang, H. S., Zhu, T., and Glazebrook, J. 2003. Characterization of the early response of *Arabidopsis* to *Alternaria brassicicola* infection using expression profiling. *Plant Physiology* 132: 606-617.
- Vaughn, K. C., and Duke, S. O. 1983. In situ localization of the sites of paraquat action. *Plant, Cell and Environment* 6: 13-20.
- van Loon, L. C., Rep, M., and Pieterse, C. M. 2006. Significance of inducible defense-related proteins in infected plants. *Annual Review of Phytopathology* 44: 135-162.
- Verheye, W. 2010. Growth and production of oil palm. Land use, land cover and soil sciences. Encyclopedia of Life Support Systems (EOLSS). UNESCO-EOLSS Publishers, Oxford, UK. Available at: /Sample-Chapters/C10/E1-05A-27-00.pdf
- Vijayan, P., Shockey, J., Lévesque, C. A., and Cook, R. J. 1998. A role for jasmonate in pathogen defense of *Arabidopsis*. *Proceedings of the National Academy of Sciences* USA 95: 7209-7214.
- Vos, I. A., Moritz, L., Pieterse, C. M. and Van Wees, S. 2015. Impact of hormonal crosstalk on plant resistance and fitness under multi-attacker conditions. *Frontiers in Plant Science* 6: 639
- Wagner G, Nadasy E. 2006. Effect of pre-emergence herbicides on growth parameters of green pea. *Communications in Agricultural and Applied Biological Science* 71: 809–813.

- Wahid, B.M., Abdullah, S.N.A., and Henson, I. 2005. Oil palm-achievements and potential. *Plant Production Science* 8: 288–297.
- Wi, S.J., Ji, N.R., and Park, K.Y. 2012. Synergistic biosynthesis of biphasic ethylene and reactive oxygen species in response to hemibiotrophic *Phytophthora parasitica* in tobacco plants. *Plant Physiology* 159: 251-265.
- Wibawa, W., Mohayidin, M. G., Mohamad, R. B., Juraimi, A. S., and Omar, D. 2010. Efficacy and cost-effectiveness of three broad-spectrum herbicides to control weeds in immature oil palm plantation. *Pertanika Journal of Tropical Agricultural Science* 33: 233-241.
- Wild, A., Sauer, H., and Rühle, W. 1987. The effect of phosphinothricin (glufosinate) on photosynthesis I. inhibition of photosynthesis and accumulation of ammonia. *Zeitschrift für Naturforschung* 42: 263-269.
- Williams, M. E. 2010. Introduction to phytohormones. The Plant Cell 22: 1-9.
- Williams, H. L., Sturrock, R. N., Islam, M. A., Hammett, C., Ekramoddoullah, A. K., and Leal, I. 2014. Gene expression profiling of candidate virulence factors in the laminated root rot pathogen Phellinus sulphurascens. *BMC Genomics* 15: 603.
- Wong, H. L., Pinontoan, R., Hayashi, K., Tabata, R., Yaeno, T., and Hasegawa, K. 2007. Regulation of rice NADPH oxidase by binding of Rac GTPase to its N-terminal extension. *Plant Cell* 19: 4022-4034.
- Wong, L. C., Bong, C. F. J. and Idris, A. S. 2012. *Ganoderma* species associated with basal stem rot disease of oil palm. *American Journal of Applied Sciences* 9:879.
- Woodburn, A.T. 2000. Glyphosate: production, pricing and use worldwide. *Pest Management Science* 56: 309–312.
- Xu, E. 2015. Interaction between Hormone and Apoplastic ROS Signaling in Regulation of Defense Responses and Cell Death. PhD Thesis. University of Helsinki, Finland.
- Yao, Z., Islam, M. R., Badawi, M. A., El-Bebany, A. F., and Daayf, F. 2015. Overexpression of StRbohA in Arabidopsis thaliana enhances defense responses against Verticillium dahliae. Physiological and Molecular Plant Pathology 90:105-114.
- Yeoh, K. A., Othman, A., Meon, S., Abdullah, F., and Ho, C. L. 2012. Sequence analysis and gene expression of putative exo-and endo-glucanases from oil palm (*Elaeis guineensis*) during fungal infection. *Journal of Plant Physiology* 169: 1565-1570.
- Zhao, J.D. 1989. The Ganodermataceae in China. *Bibliotheca Mycologica*. Berlin, Stuttgart.

Zhou, L., Cheung, M. Y., Li, M. W., Fu, Y., Sun, Z., Sun, S. M., and Lam, H. M. 2010. Rice hypersensitive induced reaction protein 1 (OsHIR1) associates with plasma membrane and triggers hypersensitive cell death. *BMC Plant Biology* 10: 290.



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