



***CHARACTERIZATION OF POTENTIALLY PATHOGENIC *Escherichia coli*  
ISOLATED FROM CHICKEN FARMS IN MALAYSIA***

**ROSELIZA BINTI ROSLEE**

**FPV 2019 9**



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By  
**ROSELIZA BINTI ROSLEE**

Thesis Submitted to School of Graduate Studies, Universiti Putra  
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June 2019

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfillment for the degree of Masters of Science

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ISOLATED FROM CHICKEN FARMS IN MALAYSIA**

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**June 2019**

**Chairman: Assoc. Prof. Siti Khairani Bejo, PhD  
Faculty: Veterinary Medicine**

*Escherichia coli* is a part of normal floral in intestinal tract in various animal species. *E. coli* always considered as non-pathogenic, however certain *E. coli* strain can cause infection when they harbour certain virulent properties. In poultry, *E. coli* is the most predominant and important strains affecting poultry industry due to the significant economic loss as result of high mortality and morbidity. This study was conducted to determine the phenotype and genotype characteristic of 125 *E. coli* isolates available in Veterinary Research Institute (VRI) which are isolated from chicken farms with significant clinical signs and abundant growth upon isolation on growth medium. There was no concrete information regarding *E. coli* isolated from chicken available elsewhere in the country.

This study showed that *E. coli* isolates recovered from chicken farms were found to have diverse biochemical properties, with no single features was specific for *E. coli* identification. All the isolates were further categorized into 12

distinct groups based on their biochemical profiles including haemolysis morphology. Serotyping of the *E. coli* isolates revealed that 69.6% (87/125) isolates in this study cannot be assigned to any serogroups tested. Other serotypes identified were 14.4% O1:K1 (18/125), 10.4% O78:K8O (13/125) and 5.6% O2:K1(7/125). Phylogenetic analysis demonstrated that most of the *E. coli* isolated from chicken farms in this study belonged to group B1 (36.0%) and group D (28.0%), which is associated with non-virulent strain.

Multiplex PCR analysis demonstrated that the most prevalent virulence genes identified were *iss* 52.0% (65/125), followed by *iucD* 36.0% (45/125) *tsh* 32.0% (40/125), *vat* 14.4% (18/125), *astA* 12.0% (15/125), *papC* 12.0% (15/125), *irp2*

9.6% (12/125), and the least is *cva/cvi* gene (0%). None of the isolates harbored more than four virulence genes. Further analysis showed that presence of virulence genes among the isolates were highly diverse regardless their biochemical profiles, serotype and phylogenetic groups.

Antibiogram analysis revealed that 81.6% (102/125) of the *E. coli* isolates showed multidrug resistant profiles to different antibiotics. Most of the *E. coli* isolates were highly resistant to erythromycin 52.8% (66/125), followed with tetracycline 52.0% (65/125), streptomycin 40.0% (50/125), spectinomycin 39.2% (49/125), trimethoprim 38.4% (48/125) and flumequin 37.6% (47/125). These findings also demonstrated that most of the isolates were susceptible to antibiotics commonly used for *E. coli* infections treatment in poultry with lowest resistant score against polymyxin B (92.8%) and colistin (92.0%). There is no association with the multidrug resistant profiles of the isolates with serotypes, phylogenetic groups and virulence genes profiles observed in this study.

Macrorestriction analysis of selected *E. coli* isolates resulted in heterogenous Pulse Field Gel Electrophoresis (PFGE) pattern. Construction of cluster dendrogram of 56 isolates with 60% coefficient similarity showed 41 genotypes consists of various serotypes with different biochemical profiles, serotypes, phylogroups and virulence genes profiles. This finding indicate that *E. coli* isolated from chicken farms in the country derived from different clones which display heterogenous profiles including antimicrobial resistant profiles. In conclusion, this study suggested that *E. coli* strain isolated from chicken farms was potentially pathogenic with highly diverse phenotype and genotypes. They potentially can cause disease in chicken even though initially they are harmless normal floral in gut as they able to inherit virulence genes from other bacterial strains in gut which enable them to cause disease.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**PENCIRIAN BERPOTENSI PATOGENIK *Escherichia coli* YANG DIASINGKAN DARI LADANG AYAM DI MALAYSIA**

Oleh

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**Jun 2019**

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*Escherichia coli* adalah bakteria Gram negatif yang merupakan sebahagian bakteria dari flora normal saluran usus dalam pelbagai spesis haiwan. *E. coli* selalunya dianggap sebagai bukan patogenik, namun begitu terdapat strain *E. coli* tertentu yang boleh menyebabkan jangkitan sekiranya mengandungi ciri virulen tertentu. Pada ayam, *E. coli* merupakan strain yang paling dominan dan penting yang menjelaskan industri unggas kerana kerugian ekonomi yang jelas lantaran dari kematian dan kerosakan yang tinggi. Kajian ini dijalankan untuk menentukan karakter fenotip dan genotip 125 pencilan *E. coli* yang dipencarkan dari ayam dengan tanda klinikal yang signifikan serta pertumbuhan yang banyak atas media pertumbuhan. Tiada maklumat yang jelas berkenaan ciri *E. coli* diasingkan dari ayam yang didapati di negara ini.

Berdasarkan hasil kajian ini, pencilan *E.coli* yang diasingkan dari ayam mempunyai ciri biokimia yang pelbagai di antara satu sama lain, dan tiada ciri yang tunggal dan spesifik yang khusus untuk pengenalpastian *E.coli*. Kesemua pencilan *E.coli* yang diuji dikategorikan kepada 12 kumpulan berbeza berdasarkan profil ujian biokimia yang dijalankan termasuklah ciri hemolisis atas agar darah. Penserotipan semua pencilan tersebut menunjukkan bahawa 69.6% (87/125) pencilan yang diuji tidak dapat dikelaskan dalam mana-mana serotip yang diuji. Serotip lain yang dikenalpasti adalah O1:K1 (14.4%), O78:K8O (10.4%), dan O2:K1 (5.6%).

Analisis filogenetik ke atas pencilan-pencilan tersebut mendedahkan bahawa sebahagian besar *E. coli* yang diasingkan dari ladang ayam dalam kajian ini terdiri dari kumpulan B1 (36.0%) dan kumpulan D (28.0%), yang dikaitkan dengan strain yang tidak virulen atau merbahaya.

Analisis PCR multipleks menunjukkan bahawa gen virulen yang paling banyak dikenalpasti adalah *iss* 52.0% (65/125), diikuti oleh *iucD* 36.0% (45/125), *tsh* 32.0% (40/125), *vat* 14.4% (18/125), *astA* 12.0% (15/125), *papC* 12.0% (15/125), *irp2* 9.6% (12/125), dan yang paling sedikit adalah gen *cva/cvi* (0%). Setiap kumpulan filogenetik diwakili dengan kombinasi gen virulen yang berbeza dan pelbagai, dan tiada satu kombinasi gen virulen yang spesifik dikenalpasti mempunyai kaitan dengan kumpulan filogenetik pencilan *E. coli*.

Analisis antibiogram menunjukkan bahawa 81.6% (102/125) pencilan *E. coli* mempunyai profil kerintangan pelbagai terhadap antibiotik yang berbeza. Kebanyakan *E. coli* menunjukkan kerintangan yang tinggi keatas eritromisin 52.8% (66/125), diikuti dengan tetrasiklin 52.0% (65/125), streptomisin 40.0% (50/125), spektinomisin 39.2% (49/125), trimethoprim 38.4% (48/125) dan flumequin 37.6% (47/125). Penemuan ini juga menunjukkan bahawa kebanyakan *E. coli* berkesan terhadap antibiotik yang seringkali digunakan dalam mengatasi jangkitan *E. coli* pada ayam dengan tahap kerintangan yang paling rendah terhadap polymyxin B (92.8%) dan colistin (92.0%). Tiada kaitan dilihat antara profil kepelbagaiannya kerintangan antibiotik dengan serotip, kumpulan filogenetik dan profil gen virulen dalam kajian ini.

Analisis sekatan makro keatas pencilan *E. coli* yang pilihan menunjukkan corak Pulse Field Gel Electrophoresis (PFGE) yang pelbagai. Pembinaan kumpulan dendrogram keatas 56 pencilan dengan kesan 60% koefisien setara menghasilkan 41 genotip yang terdiri dari pelbagai serotip dengan pelbagai profil biokimia, kumpulan filogenetik dan profil gen virulen. Penemuan ini menyatakan bahawa *E. coli* yang diasingkan dari ladang ayam berpotensi patogenik serta mempunyai ciri fenotip dan genotip yang sangat pelbagai, *E. coli* berpotensi menyebabkan jangkitan pada ayam walaupun pada asalnya tidak merbahaya dan merupakan bakteria normal flora dalam usus memandangkan mereka berupaya untuk mewarisi gen virulen dari bakteria lain dalam usus yang menyebabkan jangkitan.

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## LIST OF ABBREVIATIONS

AFLP	Amplified Fragment Length Polymorphism
AMP	Ampicillin
APEC	Avian Pathogenic <i>E. coli</i>
APR	Apramycin
$\alpha$	Alpha
astA	Enteropathogenic toxin gene
ATCC	American Type Culture Collection
AZM	Azithromycin
bp	Base Pair
$\beta$	Beta
BSA	Bovine Serum Albumin
CDC	Centre of Disease
ChuA	Gene required for heme transport
CLSI	Clinical Laboratory Standard Institute
CN	Gentamicin
CSB	Cell Suspension Buffer
CT	Colistin
cva/cvi	Colicin V Plasmid Operon gene
$^{\circ}\text{C}$	Degree Celcius
DNA	Deoxyribonucleic Acid
DVS	Department of Veterinary Services
E	Erythromycin
ECOR	<i>E. coli</i> Reference Collection
EFT	Ceftiofur

EHEC	Enterohaemorrhagic <i>E. coli</i>
IB	Infectious Bronchitis
<i>lss</i>	Increased serum survival gene
<i>lrp2</i>	Iron repressible protein gene
<i>iucD</i>	Aerobactin gene
ExPec	Extraintestinal Pathogenic <i>E. coli</i>
ESBL	Extended SpectrumBeta Lactamase
<i>gyrA</i>	DNA gyrase gene
K	Kanamycin
Kb	Kilobases
MIC	Minimum Inhibitory Concentration
MLST	Multilocus Sequencing Typing
mm	milimetre
MR	Methyl Red
MRSA	Methicillin Resistant <i>Staphylococcus aureus</i>
MHA	Mueller Hinton Agar
MVLA	Multilocus Variable Number Tandem Repeat Analysis
ND	Newcastle Disease
NMEC	Neonatal Meningitis <i>E. coli</i>
NPCB	National Pharmaceutical Bureau
PAIs	Pathogenicity islands
<i>parC</i>	Topoisomerase IV gene
PB	Polymyxin B
<i>papC</i>	P fimbriae gene
PCR	Polymerase Chain Reaction

PFGE	Pulse Field Gel Electrophoresis
RAPD	Random Amplified of Polymorphic DNA
S	Streptomycin
SH	Spectinomycin
stx	Shiga toxin gene
TBE	Tris-Borate EDTA
TE	Tris- EDTA
TE	Tetracycline
<i>tsh</i>	Temperature sensitive hemagglutinin gene
<i>TSPE4.C2</i>	Gene from published subtractive library of <i>E. coli</i>
µl	Microlitre
UB	Flumequin
UPEC	Uropathogenic <i>E. coli</i>
UPGMA	Unweight Pair Group Method Using Arithmetic Averages
<i>vat</i>	Vacuolating autotransporter toxin gene
VP	Voges Proskur
VRI	Veterinary Research Institute
W	Trimethoprim
WHO	World Health Organization
<i>yjaA</i>	Gene initially identified in genome sequence of <i>E. coli</i> K-12 strain

## CHAPTER 1

### INTRODUCTION

*Escherichia coli* is considered a member of normal microflora of most human beings and other mammals and birds (Ahmed et al., 2013). This bacterium usually considered as commensal bacteria and harmless, but some strains enable to cause fatal diseases in human as well as mammals and birds (Branko et al., 2011). The incidence of *E. coli* infection has increased tremendously over the years (Jafari et al., 2012) and has become an important issues particularly in human due to the formation of superbug multidrug resistance strains arising from those commensal strains of *E. coli*. Despite, *E. coli* has been highlighted as a major health treat due to their ability to cause severe problems in human with the most recent and fatal outbreak in Germany causing high morbidity and mortality (Buccholz et al., 2011).

Since ancient times, *E. coli* has been linked to the cause of diseases in poultry, but has not received any special attention since this strain is often overlooked as contaminants. However, they also have been reported as aetiology agent of various diseases in poultry such as swollen head syndrome, colisepticemia, embryonic mortality, dermatitis, cellulitis, salphingitis and septicemia (Khoo et al., 2010). The most important infection caused by *E. coli* in poultry is colibacillosis. Colibacillosis has become a major problem in poultry industry worldwide due to the significance economic loss through high mortality and morbidity, loss in weight gain and poor carcass quality (Dziva et al, 2008).

Based on Department of Veterinary Services (DVS) annual reports, *E. coli* was isolated from 70% of veterinary samples submitted for diagnosis including chicken samples (Mat Amin et al., 2011; Khoo et al., 2014). Most of these samples exhibit significant clinical signs such as high mortality, respiratory distress and low body weight. Abundant of *E. coli* are isolated from all visceral organs. It's showing that *E. coli* is one of the major problems in the poultry industry that needs to be addressed. Although *E.coli* is often regarded as contaminant and non-pathogenic, but due to a large number of *E.coli* isolates from poultry particularly chickens with significant clinical signs, it has become an alarming signal to the poultry industry in the country that need to be highlighted. However, it is unknown whether *E. coli* is indeed pathogenic and really causative agent causing the disease in the chicken, or simply non-pathogenic bacteria.

In limited information on *E. coli* itself, controlling the infection become a big challenge in many countries. In addition, antibiotic resistance issues in *E. coli* in the poultry also contributed to complicates treatment againts *E. coli* infection, resulting in ineffective disease control (Mooljuntiee *et al.*, 2010; Diarra and Malouin, 2014). Multidrug resistant in *E. coli* strain in food animals particularly in chicken continues to be debated in many countries including Malaysia, with the emergence and spread of multidrug resistant among *E. coli* and other commensal bacteria to newer antimicrobial compounds thus becoming a global threat (Marshall and Levy, 2011; Ong *et al.*, 2014).

Though there are many study on characterization of *E. coli* has been done elsewhere, most of the studies are limited in specific serotypes and involved certain location. Based on the previous information available, *E. coli* characteristics on virulence properties, phylogenetic grouping, antimicrobial resistant and genotyping profiles in different geographical locations were heterogenous (Clermont *et al.*, 2000; Ewers *et al.*, 2005). The diagnosis method and recommended vaccine available were unreliable to be used in the country considering that the features of *E. coli* local strains are very different in different with other countries (Kwon *et al.*, 2008; Ahmed *et al.*, 2013; Zainal Abidin *et al.*, 2013; Schouler *et al.*, 2012). Therefore there are still no effective disease control approach can be apply to resolve the infection. Despite, most of studies on phenotyping and genotyping of *E. coli* conduct in the country involved limited to scope and certain geographical area (Apun *et al.*, 2008; Khoo *et al.*, 2010; Geidam *et al.*, 2012b; Zainal Abidin *et al.* 2013; Khoo *et al.*, 2014). As a result, there is no concrete information regarding phenotyping and genotyping profiles of *E. coli* including in chicken in the country. The information on *E. coli* local isolates will provide detail information about the disease in the country thus useful for future works in establishment of effective diagnosis and disease control programmes. It is hypothesized that *E. coli* strains isolated from chicken in Malaysia is highly diverse and still not fully establish.

Therefore, the objectives of this study were:

- i. To determine the most prevalent *E. coli* serotypes isolated from chicken in Malaysia
- ii. To detect the presence of virulence genes in the *E. coli* isolates by using multiplex PCR
- iii. To determine phylogenetic grouping of *E. coli* isolates by using triplex PCR.
- iv. To determine multidrug resistance profiles of *E. coli* isolates from chicken
- v. To characterize selected *E. coli* isolates using Pulse Field Gel Electrophoresis (PFGE).

## REFERENCES

- Aalipor, F., Mirlohi, M. and Jalali, M. (2014). Determination of antibiotic consumption index for animal originated foods produced in animal husbandry in Iran 2010. *Journal of Environmental Health Sciences and Engineering*, 12(42): 1 – 7.
- Abu Daud N. H. B., Htin N. N., Abba Y., Paan F. H., Kyaw T., Khaing A. T., Jesse F. F. A., Mohammed K., Adamu L. and Tijjani A. (2014). An outbreak of colibacillosis in a broiler farm. *Journal of Animal and Veterinary Advances*, 13(8): 545 – 548.
- Aggad, H., Ammar, Y.H., Hammoudi, A. and Kihal, M. (2010.) Antimicrobial resistance of *Escherichia coli* isolated from chickens with colisepticemia. *Global Veterinaria*, 4(3): 303 – 306.
- Ahmed, A.M., Toshi, S. and Tadashi, S. (2013). Molecular characterization of multidrug resistance Avian Pathogenic *Escherichia coli* (APEC) isolated from septicemic broilers. *International Journal of Medical Microbiology*, 303: 475 – 483.
- Akond, M.A., Alam, S., Hassan, S.M.R. and Shirin, M. (2009). Antibiotic resistance of *Escherichia coli* isolated from poultry and poultry environment of Bangladesh. *Internet Journal of Food Safety*, 11: 19 – 23.
- Al-Ghamdi, M.S., El-Morsy, F., Al-Mustafa, Z.H., Al-Ramadhan, M. and Hanif, M. (1999). Antibiotic resistance of *Escherichia coli* isolated from poultry workers, patients and chickens in the eastern province of Saudi Arabia. *Tropical Medicines and International Health*, 4(4): 278 – 283.
- Allocati, N., Masulli, M., Alexeyev, M.F. and Di Ilio, C.(2013). *Escherichia coli* in Europe: An Overview. *International Journal Environmental Research Public Health*, 10: 6235 – 6254.
- Antao, E.M., Gladde, S., Li, G., Sharifi, R., Homeier, T., Loturnus, C., Diehl, J., Bethe, A., Philip, H.C., Preisinger, R., Wieler, L.H. and Ewers, C. (2008).The chicken as natural model for extra intestinal infections caused by Avian Pathogenic *Escherichia coli*. *Microbiology Pathogenesis*, 45 (5 – 6): 361 – 369.
- Apun, K., Chong, Y.L., Abdullah, M.T. and Micky, V.(2008). Antimicrobial susceptibilities of *Escherichia coli* isolated from food animals and wildlife animals in Sarawak, East Malaysia. *Asian Journal of Animal and Veterinary Advances*, 3(6): 409 – 416.
- Arabi, S., Jafarpour, M., Mirnargesi, M., Behjati Asl, S., Naghshbandi, R. and Shabanpour, M. (2013). Molecular characterization of Avian

- Pathogenic *Escherichia coli* in broilers bred in Northern Iran. *Global Veterinaria*, 10 (4): 382 – 386.
- Arthur, T.M., Ahmed, R. Chase-Topping, M., Kalchayanand, n., Schmidt, J.W. and Bono, J.L.(2015). Characterization of *Escherichia coli* isolated from supershedding cattle. *Applied and Environmental Microbiology*, 79(4): 4294 – 4303.
- Asai, T. Masani, K., Sato, C., Hiki, M., Usui, M., Baba, K., Ozawa, M., Harada, K., Aoki, H. and Sawada, T.(2011). Phylogenetic groups and cephalosporin resistance genes of *Escherichia coli* from diseased food –producing animals in Japan. *Acta Veterinaria Scandinavica*, 53(52).
- Ashraf, A.M., Shimamoto, T. and Shimamoto, T.(2013). Molecular characterization of multidrug-resistant isolated from septicemic broilers. *International Journal of Medical Microbiology*, 303: 473 – 483.
- Aslam, M., Toufeer, M., Bravo, G.N., Lai, V., Rempel, H., Manges, A. and Diarra, M.S.(2014). Characterization of extraintestinal pathogenic *Escherichia coli* isolated from retail poultry meats from Alberta, Canada. *International Journal of Food Microbiology*, 177; 49 – 56.
- Badri, S., Fassouane, A., Filliol, I., Mohammed, H. and Cohen, N.(2009). Clonal analysis of *Escherichia coli* strains isolated from food by Pulse Field Gel Electrophoresis. *Internet Journal of Food Safety*, 11: 44 – 49.
- Bae, I.K., Kim, J., Sun, J.Y.H., Jeong, S.H., Kim, Y.R., Wang, K.K. and L.K.(2014). Comparison of pulse field gel electrophoresis and repetitive sequence based PCR methods for molecular epidemiological studies of *Escherichia coli* clinical isolates. *Indian Journal Medical Research*, 679 – 685.
- Belanger, L., Gareaux, A., Harel, J., Boulianne, M., Nadeau, E. and Dozois, C.M.(2011). *Escherichia coli* from animal reservoirs as a potential source of human extraintestinal pathogenic *E.coli*. *FEMS Immunology Medical Microbiology*, 62: 1 – 10.
- Benamer, Q., Guermour, D., Hammoudi, A., Aoudia, H., Aggad, H., Humblet, M.F. and Saegerman, C.(2014). Antimicrobial resistance *Escherichia coli* isolated from chicken in West Algeria. *International Journal of Science: Basic and Applied Research*, 13(1): 366 – 370.
- Bettelheim, K.A.(1994). Biochemical characteristics of *Escherichia coli*. *Escherichia coli in domestic animals and humans* (pp. 3 -30). UK: Biddles Ltd, Guilford.
- Blanco, J.E., Blanco, M., Mora, A., Jansen, W.H., Garcia, V., Vazquez, M.L. and Blanco, J.(1997). Serotypes of *Escherichia coli* isolated from

- septicaemic chickens in Galicia (Northwest Spain). *Veterinary Microbiology*, 61: 229 – 235.
- Boughattas, S. and Salehi, R. (2014). Molecular approaches for detection and identification of foodborne pathogens. *Journal of Food and Hazard Control*, 1: 1 – 6.
- Branko, V., Vlado, T. and Branka, B. (2011). Recent advances in understanding the virulence of enterohemorrhagic *Escherichia coli* in foods. *Technologija mesa*, 1: 52 – 59.
- Briyne, N.D., Atkinson, J., Pokludová, L. and Borriello, S.P. (2015). Antibiotic used most commonly to treat animal in Europe. *Veterinary Record*, [Doi: 10.1136/vr.102462](https://doi.org/10.1136/vr.102462).
- Buccholz, U., Bernard, H., Werber, D., Bohmer, M.M., Remschmidt, C., Wilking, H., Delere, Y., an der Haiden, M., Adlhoch, C., Dreesman, J., Ehlers, J., Ethelberg, S., Faber, M., Frank, C., Fricke, G., Greiner, M., Hohle, M., Ivarsson, S., Jark, U., Kirchner, M., Koch, J., Krause, G., Luber, P., Rosner, B., Stark, K. and Kuhne, M. (2011). German outbreak of *Escherichia coli* O104:H4 associated with sprouts. *The New England Journal of Medicine* 365,(19): 1763 – 1770.
- Byun, J.W., Jung, B.Y., Kim, H.Y., Fairbrother, J.M., Lee, M.H. and Lee, W.K. (2013). O – serogroups, virulence genes of pathogenic *Escherichia coli* and Pulse field gel electrophoresis (PFGE) patterns of O149 isolates from diarrhoeic piglets in Korea. *Veterinarni Medicina*, 58 (9):468 – 476.
- Cai, H.Y., Caswell, J.L. and Prescott, J.F. (2014). Nonculture molecular techniques for diagnosis of bacterial disease in animal: a diagnostic laboratory perspective. *Veterinary Pathology*, 51(2): 341 – 350.
- Caprioli, A., Scavia, G. and Morabito, S. (2014). Public health microbiology of shiga toxin – producing *Escherichia coli*. *Microbiology Spectrum* 2(6): EHEC 0014-2013. [doi:10.1128/microbiolspec.EHEC-0014-2013](https://doi.org/10.1128/microbiolspec.EHEC-0014-2013).
- Caya, F., Fairbrother, J.M., lessard, L. and Quessy, S. (1999). Characterization of the risk to human health of pathogenic *Escherichia coli* isolates from chicken carcasses. *Journal of Food Protection*, 62(7): 741 – 746.
- Clermont, O., Bonacrosi, S. and Bingen, E. (2000). Rapid and simple determination of the *Escherichia coli* phylogenetic group. *Applied and Environmental Microbiology*, 66 (10): 4555 – 4558.
- Clermont, O., Olier, M., Hoede, C., Diancourt, L., Brisse, S., Kerouden, M., Glodt, J., Picard, B., Oswald, E. and Denamur, E. (2011). Animal and human pathogenic *Escherichia coli* strains common genetic backgrounds. *Infection, Genetics, Evolution*, 11: 654 – 662.

Clinical and Laboratory Standard Institute.(2012).Performance standards for antimicrobial susceptibility testing, 25<sup>th</sup> Informational Supplement. CLSI Document M100 – S25.

Cobbaut, K., Houf, K., Boyen, F., Haesebrouck, F. and De Zulter, L.(2011).Genotyping and antimicrobial resistance patterns of *Escherichia coli* O157 originating from cattle farms. *Foodborne Pathogen and Diseases*, 8(6): 719 – 724.

Collingwood, C., Kemmet, K., Williams, N. and Wigley, P. (2014). Is the concept of avian pathogenic *Escherichia coli* as a single pathotype fundamentally flawed? *Veterinary Infectious Diseases*, 1(5).

Croxen, M.A., Law, R.J., Scholz, R., Keeney, K.M., Wlodarska, M. and Finley, B.B. (2013). Recent advances in understanding Enteric Pathogenic *Escherichia coli*. *Clinical Microbiology Reviews*, 26 (4): 822 – 880.

Dai, L., Lu, M.L., Wu, C.M., Li, B.B., Huang, S.Y., Wang, S.C., Qi, Y.H. and Shen, J.Z.(2008). Characterization of antimicrobial resistance among *Escherichia coli* isolates from chicken in China between year 2001 and 2006. *FEMS Microbiology Letter*, 286: 178 – 183.

Danzeisen, J.L.,Wannemuhler, Y., Nolan, L.K. and Johnson, T.J. (2013).Comparison of Multilocus Sequence Typing (MLST) and virulence genotyping of *Escherichia coli* from live birds, retail poultry meat and human extraintestinal infection. *Veterinary Microbiology and Preventive Medicine*, 57: 104 – 108.

De Carli, S., Ikuta, N., Lehmann, F.K.M., da Silveira, V.P., Predebon, G.M., Fonseca, A.S.K. and Lunge, V.R.(2015). Virulence gene content in *Escherichia coli* isolates from poultry flocks with clinical signs of colibacillosis in Brazil. *Poultry Science*, 94: 2635 – 2640.

Dho Moulin, M. and Fairbrother, J.M.(1999).Avian Pathogenic *Escherichia coli* (APEC). *Veterinary Research*, 30: 299 – 316.

Diarra, M.S.and Malouin, F. (2014).Antibiotics in Canadian poultry productions and anticipated alternatives. *Frontiers in Microbiology*, 5(282): 1 – 15.

Dissanayake, D.R.A., Wijewardana, T.G. and Gunawardana, G.A. (2008). Distribution of lipopolysaccharide core types among Avian Pathogenic *Escherichia coli* (APEC) in relation to the major phylogenetic groups. *Veterinary Microbiology* ,132: 355 – 363.

Dissanayake, D.R., Octavia, S. and Lan, R. (2014). Population structure and virulence content of Avian pathogenic *Escherichia coli* isolated from outbreaks in Sri Lanka. *Veterinary Microbiology*,168: 403 – 412.

- Dou, X., Gong, J., Han, X., Xu, M., Shen, H., Zhang, D., Zhuang, L., Liu, J. and Zou, J.(2015.)Characterization of Avian Pathogenic *Escherichia coli* isolated in Eastern China. *Gene*, 576: 244 – 248.
- Dutil, L., Irwin, R., Finley, R., Ng, L.K., Avery, B., Boerlin, P., Bourgault, A.M., Cole, L., Daignault, D., Desrusseau, A., Demczuk, W., Hoong, L., Harsmen, G.B., Ismail, J., Jamieson, F., Maki, A., Pacagnella, A. and Pillai, D.R. (2010).Certiofur resistance in *Salmonella* enterics serovar Heidelberg from chicken meat and humans, Canada.*Emerging Infectious Disease*, 16(1): 48 – 54.
- Dziva, F and Stevens, M.P. (2008). Colibacillosis in poultry: Unravelling the molecular basis of virulence of avian pathogenic *Escherichia coli* in their natural hosts. *Avian Pathology*, 37(4): 355 – 366.
- Ewers, C., JanBen, T., kießling, S., Philip, H.C. and Wieler, L.H.(2004). Molecular epidemiology of avian pathogenic *Escherichia coli* (APEC) isolated from colisepticemia in poultry. *Veterinary Microbiology*, 104: 91 – 101.
- Ewers, C., JanBen, T., Kiessling, S., Philipp, H.C. and Wieler, L.H. (2005). Rapid detection of virulence associated genes in Avian Pathogenic *Escherichia coli* (APEC) by multiplex Polymerase Chain Reaction (PCR). *Avian Disease*, 49: 269 – 273.
- Ewers, C., Li, G., Wilking, H., Kießling, S., Alt, K., Antao, E.M., Laturnus, C., Diehl, I., Glodde, S., Homeier, T., Bohnke, U., Steinruck, H., Philip, H.C. and Wieler, L.H. (2007). Avian pathogenic, uropathogenic, and newborn meningitis- causing *Escherichia coli*: How closely related they are? *International Journal of Medical Microbiology*, 297(3): 163 – 171.
- Ewers, C., Antao, E.M., Diehl, I., Phillip, H.C. and Wieler, L.H. (2009). Intestine and environment of chicken as reservoirs for extraintestinal pathogenic *Escherichia coli* strains with zoonotic potential. *Applied and Environmental Microbiology*, 75 (1): 184 – 192.
- Fairbrother, J.M. and Nadeau, E. (2006). *Escherichia coli*: On farm contamination of animals.Scientific and Technical Review of the Office International des Epizooties, 25 (2): 555 – 569.
- Frederick, A. (2011). *Escherichia coli*, its Prevalence and Antibiotic Resistant: A Mini Review. *Microbiology Journal*, 1(2): 47 – 53.
- Gales, A.C., Jones, R.N. and Sader, H.S.(2011). Contemporary activity of colistin and polymyxin B against a worldwide collection of Gram negative pathogens: results from SENTRY Antimicrobial Surveillance Program (2006 – 2009). *Journal of Antimicrobial Chemotherapy Advances*, [Doi: 10.1093/jac/dkr 239](https://doi.org/10.1093/jac/dkr239).

- Geidam, G., Ambali, A.G. and Onyeyili, P.A.(2012). Detection and antibiotic sensitivity pattern of avian pathogenic *Escherichia coli* strains among rural chickens in the Arid region of North eastern Nigeria. *Veterinary World*, 5(6): 325 – 329.
- Geidam, G., Zakaria, Z., Bejo, S.K., Abu, J. and Omar, S. (2012b). High prevalence of multidrug resistant bacteria in selected poultry farms in Selangor, Malaysia. *Asia Journal of Animal Veterinary Advances*, 7: 891 – 897.
- Geonaras, I., Hastings, J.W. and van Holly, A. (2001). Genotyping analysis of *Escherichia coli* strains from poultry carcasses and their susceptibilities to antimicrobial agents. *Applied and Environmental Microbiology*, 67(4): 1940 – 1944.
- Graziani, C., Luzzi, I., Corro, M., Tomei, F., Parisi, G., Giufre, M., Marobito, S., Caprioli, A. and Cerquetti, M.(2009). Phylogenetic background and virulence genotype of ciprofloxacin susceptible and ciprofloxacin resistance *Escherichia coli* strains of human and avian origin. *The Journal of Infectious Diseases*, 199: 1209 – 1217.
- Hassan, Q. (2013).Avian Pathogenic *Escherichia coli* (APEC) in Palestine; Characterization of Virulence Factors and Antibiotic Resistance Profile Master Thesis.Palestine Polytechnic University.
- Hiki, M., Usui, M., Akiayama, T., Kawanishi, M., Tsuyuki, M., Imamura, S., Sekiguchi, H., Kojima, A. and Asai, T. (2014). Phylogenetic grouping, epidemiological typing, analysis of virulence genes, and antimicrobial susceptibility of *Escherichia coli* isolated from healthy broilers in Japan. *Irish Veterinary Journal*, 67:14.
- Jafari, A., Aslani, M.M. and Bouzori, S. (2012). *Escherichia coli*: A brief review of diarrheagenic pathotypes and their role in diarrheal diseases in Iran. *Iranian Journal of Microbiology*, 4(3): 102 – 117.
- Jakobsen, L., Spangholm, D.J., Pedersen, K., Jensen, L.B., Emborg, H.B., Agerso, Y., Aarestrup, F.M., Hammerum, A.M. and Moller, N.F. (2010). Broiler chickens, broiler chicken meat, pigs and pork as source of ExPec related virulence genes and resistance in *Escherichia coli* isolates from community- dwelling humans and UTI patients. *International Journal of Food Microbiology*, 264 – 272.
- Jeong, Y.W., Kim, T.E., Kim, J.H. and Kwon, H.J. (2012). Pathotyping avian pathogenic *Escherichia coli* strains in Korea. *Journal Veterinary Science*, 13 92): 145 – 152.
- Johnson, J.R. and Russo, T.A. (2002) Extraintestinal pathogenic *Escherichia coli*: “The other bad *E.coli*”. *Journal Laboratory Clinical Medicine*, 139 (3): 155 – 161.

- Johnson T.J., Kariyawasam, S., Wannemuehler, Y., Mangamele, P., Johnson, S.J., Doetkott, C., Skyberg, J.A., Lynne, A.M., Johnson, J.R. and Nolan, L.K. (2007). The genome sequence of avian pathogenic *Escherichia coli* strain O1:K1:H7 shares strong similarities with human extraintestinal pathogenic *E.coli* genomes. *Journal of Bacteriology*, 189(8): 3228 – 3236.
- Johnson, T.J., Wannemuehler, Y., Johnson, S.J., Stell, A.L., Doetkott, C., Johnson, J.R., Kim, K.S., Spaanjard, L. and Nolan, L.K. (2008). Comparison of Extraintestinal Pathogenic *Escherichia coli* strains from human and avian source reveal a mix subset representing potential zoonotic pathogens. *Applied and Environmental Microbiology*, 74 (22): 7043 – 7050.
- Kabir, S.M.L. (2010). Avian colibacillosis and salmonellosis: A closer look at epidemiology, pathogenesis, diagnosis, control and public health concerns. *International Journal Environmental Research Public Health*, 7: 89 – 114.
- Katouli, M., Kuhn, I. and Mollby, R.(1990).Evaluation of stability of biochemical phenotypes of *Escherichia coli* upon subculturing and storage. *Journal of General Microbiology*, 136: 1681 – 1688.
- Katouli, M.(2010).Population structure of gut *Escherichia coli* and its role in development of extra intestinal infections. *Iranian Journal of Microbiology*, 2(2): 59 – 72.
- Khaton, R., Haider, M., Paul, P.K., das, P.M. and Hossain, M.M.(2008).Colibacillosis in commercial chickens in Bangladesh. *The Bangladesh Veterinarian*, 25(1): 17 – 24.
- Kazemnia, A., Ahmadi, M. and Dilmaghni, M.(2014). Antibiotic resistant pattern of different *Escherichia coli* phylogenetic groups isolated from human Urinary Tract Infection (UTI) and avian colibacillosis. *Iran Biomedical Journal*, 18(4): 219 – 224.
- Kemmet, K., Humphrey, T., Rushton, S., Close, A.Wigley, P. and Williams, N.J. (2013). A longitudinal study simultaneously exploring the carriage of APEC virulence associated genes and the molecular epidemiology of faecal and systemic *E. coli* in commercial broiler chickens. *PLoS ONE* 8, (6): e67749, [Doi: 10.1371/journal.pone.0067749](https://doi.org/10.1371/journal.pone.0067749).
- Khoo, E., Roslee, R., Khoo, L.L., Mohd Noor, N., Mahmud, N., Ramli, S.N., Ramli, S.N.H., Yaacob, H. and Yaakub, R.(2014) Antimicrobial susceptibility of *Escherichia coli* from poultry in VRI for year 2013. *Malaysian Journal of Veterinary Research*, 5(1): 212 – 213.

- Khoo,L.L., Yaacob, H., Yaakub, R., Ramli, S.N., Mat Amin, M. and Mohamed, R. (2010).The prevalence of avian pathogenic *Escherichia coli* in Peninsular Malaysia. *Malaysian Journal of Veterinary Research*, 1(1): 27 – 31.
- Kobayashi, R.K.T., Aquino, I., S. Ferreira, A.L. and Vidotto, M.C. (2011). ECOR phylogenetic analysis and virulence genotyping of Avian Pathogenic *Escherichia coli* strains and *Escherichia coli* isolates from commercial chicken carcasses in Southern Brazil. *Foodborne Pathogen and Disease*, 8(5): 631 – 634.
- Katsunuma, Y., Hanazumi, M., Fujisaki, H., Minato, H., Kataoka, Y., Sawada, T., Hashimoto, Y. and Yonemochi, C. (2008). Comparison of Pulse Field Gel Electrophoresis patterns of antimicrobial resistant *Escherichia coli* and enterococci isolates from the feces of livestock and livestock farmers in Japan. *Journal General Applied Microbiology*, 54: 39 – 50.
- Kwon, S.G., Cha, S.Y., Choi, E.J., Kim, B., Song, H.J. and Jung, H.K. (2008). Epidemiological prevalence of Avian Pathogenic *Escherichia coli* differentiated by multiplex PCR from commercial chickens and hatchery in Korea. *Journal of Bacteriology and Virology*, 38 (4): 179 – 188.
- Landers, T.F., Cohen, B., Wittum, T.E. and Larson, E.L. (2012). A review of antibiotic use in food animals: perspective, policy and potential. *Public Health Reports*, 127: 4 – 22.
- Landman, W.J.M., Buter, G.I., Dijkman, R. and van Eck, J.H.H. (2015). Molecular typing og avian pathogenic *Escherichia coli* colonies originating from outbreaks of *E.coli* peritonitis syndrome in chicken flocks. *Avian Pathology*, 43(4): 345 – 356.
- Lanz, R., Kuhnert, P. and Boerlin, P. (2003). Antimicrobial resistance and resistance gene determinants in clinical *Escherichia coli* from different animal species in Switzerland. *Veterinary Microbiology*, 91: 73 – 84.
- Leverstein- van Hall, M.A., Dierikx, C.M., Stuart, C., Voets, G.M., van den Munckhof, M.P., vam Essen-Zandbergen, A., Platteel, T., Fluit, A.C., van de Sande-Bruinsma, N., Scharinga, T., Bonten, M.J.M. and Mevius, D.J. (2011). Dutch patients, retail chicken meat and poultry share the same ESBL genes, plasmids and traits. *Clinical Microbiology and Infection*, 17(6): 873 – 880.
- Liu, Y.Y., Wang, Y., Walsh, T.R., Yi, L.X., Zhang, R., Spencer, J., Doi, Y., Tian, G., Dong, B., Huang, X., Yu, L.F., Gu, D., Ren, H., Chen, X., Lu, L., He, D., Zhou, H., Liang, Z., Liu, J.H. and Shen, J. (2016). Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *Lancet Infection Disease*, 16: 161– 68.

- López V.H.M., Serrano I.Q., Delgado P.D.P.M., Rodríguez L.E.V, Olague-Marchán, M., Rodriguez, S.H.S., Luna, M.A.L., de la Torre, A.F. and Santoyo, R.M.R. (2017). Genes of Virulence and phylogenetic group in isolates of Avian Pathogenic *Escherichia coli*. *Archives of Medicine*, Vol No:9 Iss No:6:5.
- Lyhs, U., Ikonen, I., Pohjanvirta, T., Raninen, K., Perko-Makela, P. and Pelkonen, S. (2012). Extraintestinal pathogenic *Escherichia coli* in poultry meat products on the Finnish retail market. *Acta Veterinaria Scandinavica*, 54 – 64.
- Manges, A.R. and Johnson, J.R. (2012). Foodborne origin of *Escherichia coli* causing extraintestinal infections. *Clinical Infectious Disease Advances*. Available at <http://cid.oxfordjournals.org>.
- Marshall, B.M. and Levy, S.B. (2011). Food animals and antimicrobials: Impacts on human health. *Clinical Microbiology Reviews*, 24 (4): 718 – 733.
- Mat Amin,M., Khoo, L.L., Yaacob, H., Ramli, S.N. and Hassim, A. (2011). Cases of *E. coli* infection submitted to VRI in 2009 .*Proceeding of 23<sup>d</sup> Veterinary Association Malaysia*, Tower Regency Hotel, Ipoh, 254 – 255.
- Maturana, V.G., Pace, F.D., Carlos, C., Pires, M.M., Campos, T.A.D., Nakazato, G., Stheling, E.G., Logue, C.M. and Nolan, L.K. (2011). Subpathotypes of avian Pathogenic *Escherichia coli* (APEC) exist as defined by their syndromes and virulence traits. *The Open Microbiology Journal*, 5: 55 – 64.
- Mbanga, J. and Nyararai, Y.O. (2015). Virulence gene profiles of Avian Pathogenic *Escherichia coli* (APEC) isolated from chickens with colibacillosis in Bulawayo, Zimbabwe. *Onderstepoort Journal of Veterinary Research*, 82(1): [Doi: 10.4102/ojvr.v82i1.850](https://doi.org/10.4102/ojvr.v82i1.850).
- McNemoy, L.L., Kotetishvili, m., Tigno, J., Keefer-Norris, A., Harris, A.D., Perencevich, E.N., Johnson, J.A., Torpey, D., Sulakvelidza, A., Morris, G. and Stine, O.C. (2005). Multilocus sequence typing versus Pulse Field Gel Electrophoresis for characterization of Extended Spectrum Beta Lactamase Producing *Escherichia coli* isolates. *Journal of Clinical Microbiology*, 43(4): 1786 – 1781.
- Mellata, M. (2013.) Human and avian extraintestinal pathogenic *Escherichia coli*: Infections, zoonotic risks and antibiotic resistance trends. *Foodborne Pathogen and Disease*, 10(11): 916 – 932.
- Micenkova, L., Bosak, J., Vrba, M., Sevcikova, A. and Smajs, D. (2016). Human Extraintestinal Pathogenic *Escherichia coli* strains differ in

- prevalence of virulence factors, pathotype and bacteriocins determinants. *BMC Microbiology*, 16: 218.
- Millman, J.M., Waits, K., Grande, H., Marks, A.R., Marks, J.C., Price, L.B. and Hungate, B.A. (2013). Prevalence of antibiotic resistant *Escherichia coli* in retail chicken: comparing conventional, organic, kosher and raised without antibiotics. *F1000Research*, 2013, 2: 155.
- Mohamed, M.A., Shehata, M.A. and Rafeek, E. (2014). Virulence genes content and antimicrobial resistance in *Escherichia coli* from broiler chickens. [Doi:org/10.1155/2014/195189](https://doi.org/10.1155/2014/195189).
- Moniri, R. and Dastahgoli, K. (2005). Fluroquinolone resistant *Escherichia coli* isolated from healthy broilers with previous exposure to fluroqionolones: Is there a link? *Microbial Ecology in Health and Disease*, 17: 69 – 74.
- Mooljuntree, S., Chansiripornchai, P. and Chansiripornchai, N.(2010). Prevalence of cellular and molecular antimicrobial resistance against *Escherichia coli* isolated from Thai broilers. *Thai Journal of Veterinary Medicine*, 40 (3): 311 – 315.
- Nakazato, G., Campos, T.A., Stehling, E.G., Brocchi, M., Silveira, W.D.(2009). Virulence factors of avian pathogenic *Escherichia coli* (APEC). *Pesquisa. Veterinaria. Brasileira*, 29(7): 479 – 486.
- Nataro, J.P. and Kaper, J.B.(1998). Diarrheagenic *Escherichia coli*. *Clinical Microbiology Reviews*, 11(1): 142 – 201.
- Nataro, J.P., Steiner, T. and Guerrant, R.L.(1998b). Enteropathogenic *Escherichia coli*. *Emerging Infectious Diseases*, 4(2): 251 – 261.
- Ngeleka, M.L., Brereton, G.B. and Fairbrother, M.(2002). Pathotypes of avian *Escherichia coli* as related to *tsh*-, *pap*-, *pil*- and *iuc*- DNA sequences, and antibiotic sensitivity of isolates from internal tissues and the cloacae of broilers. *Avian Disease*, 46: 143 – 152.
- Oh, J.Y., Kang, M.S., Kim, J.M., An, B.K., Song, E.A., Shin, E.G., Kim, M.J., kwon, J.H. and Kwon, Y.K. (2011). Characterization of *Escherichia coli* isolates from laying hens with colibacillosis on 2 commercial egg producing farms in Korea. *Poultry Science*, 90: 1948 – 1954.
- Olarinmoye, A.O., Oladele, O.O., Adejiji, A.A., Ntiwunka, U.G. and Tayo, G.O. (2013). Antibiograms of avian pathogenic *Escherichia coli* isolates from commercial layer with colibacillosis in Southwest Nigeria. *Malaysian Journal of Microbiology*, 9(4): 317 – 325.

- Olsen, R.H., Chadfield, M.S., Christensen, J.P., Scheutz, F., Christensen, H. and Bisgaard, M. (2011). Clonality and virulence traits of *Escherichia coli* associated with haemorrhagic septicemia in turkeys. *Avian Pathology*, 40(6): 587 – 595.
- Oluwasile, B.B., Abgaje, M., Ojo, O.E. and Dipeolu, M.A. (2014). Antibiotic usage pattern in selected poultry farms in Ogun State. *Sokoto Journal of Veterinary Sciences*, 12(1): 45 – 50.
- Omer, M.M., Abusalab, S.M., Gumaa, M.M., Mulla, S.A. and Omer, E.A. (2008). Outbreak of colibacillosis among broiler and layer flocks in intensive and semi intensive poultry farms in Kassala state, Eastern Sudan. *Asian Journal of Poultry Science*, 4(4): 173 – 181.
- Ong, L.P., Muniandy, K., How, S.P., Tang, S.T.P. and Lim, B.K.(2014). A report on antibiotic resistance of *Escherichia coli* isolated from veterinary samples in Malaysia from 2010 to 2013. *Malaysia Journal of Veterinary Research*, 5(1): 70 – 71.
- Paudel, S., Stessl, B., Hess, C., Zloch, A. and Hess, M.(2016).High genetic diversity among extraintestinal *Escherichia coli* isolates in pullets and layers revealed by a longitudinal study. *BMC Veterinary Research*, 12:221, [Doi: 10.1186/s12917-016-0859-5](https://doi.org/10.1186/s12917-016-0859-5).
- Pitout, J.D.D. (2012.) Extraintestinal pathogenic *Escherichia coli*: a combination of virulence with antibiotic resistance. *Frontiers in Microbiology*, [Doi: 10.3389/fmicb.2012.00009](https://doi.org/10.3389/fmicb.2012.00009).
- Platt- Mills, J.A., Operario, D.J. and Houpt, E.R.(2012). Molecular diagnosis of diarrhea: Current status and future potential. *Current. Infectious Disease Report*, 1(1): 41–46. [Doi: 10.007/s11908-011-0223-7](https://doi.org/10.007/s11908-011-0223-7).
- Quinn, P.J., Carter, M.E., Markey, B.K. and Carter, G.R.(1994). Clinical Veterinary Microbiology, First Edition, page 209.
- Rahimi, M.(2013). Antibioresistance profile of avian pathogenic *Escherichia coli* isolates recovered from broiler chicken farms with colibacillosis in Kermanshah Province, Iran. *Global Veterinaria*, 10(4): 447 – 452.
- Rahman, M.A., Samad, M.A., Rahman, M.B. and Kabir, M.L.(2004.) Bacteriological studies on Salmonellosis, colibacillosis and pasteurellosis in natural and experimental infections in chicken. *Bangladesh Journal Veterinary Medicine*, 2(1): 1 – 8.
- Raji, M., Adekeye, J., Kwaga, J., Bale, J. and Henton, M. (2007). Serovars and biochemical characterization of *Escherichia coli* isolated from colibacillosis casesand dead-in-shell embryos in poultry in Zaria-Nigeria. *Veterinarski Archive*, 77 (6), 495 – 505.

- Ranger, J.M., Sparling, P.H., Crowe, C., Griffin, P.M. and Swerdlow, D.L.(2005). Epidemiology of *Escherichia coli* O157:H7 outbreaks, United States 1982 – 2002.*Emerging Infectious Diseases*, 11(4): 603 – 610.
- Rasheed, M.U., Thajuddin, N., Ahamed, P., Teklemariam, Z. and Jamil, K.(2014). *Revista do Instituto de Medicina Tropical de Sao Paolo*, 56 (4): 341 – 346.
- Rodriguez-Siek, K.E., Giddings, C.W., Doetkott, C., Johnson, T.J. and Nolan, L.K.(2005). Characterizing APEC pathotypes. *Veterinary Research*, 36: 241 – 256.
- Rodriguez-Siek, K.E., Giddings, C.W., Doetkott, C., Johnson, T.J., Fakhr, M.K. and Nolan, L.K.(2005b). Comparison of *Escherichia coli* isolates implicated in human urinary tract infection and avian colibacillosis. *Microbiology*, 151: 2097 – 2110.
- Schlackow, I., Stoesser, N., Walker, A.S., Crook, D.W., Peto, T.E., Wyllie, D.H. (2012). Increasing incidence of *Escherichia coli* bacteraemia is driven by an increase in antibiotic resistant isolates: electronic database study in Oxfordshire 1999 – 2011. *Journal Antimicrobial Chemotherapy*, 67(6): 1514 – 1524.
- Schouler, C. Schaeffer, B., Bree, A., Mora, A., Dehbi, G., Blet, F., Oswald, E., Mainil, T., Blanco, J. and Moulin Schouler, M. (2012). Diagnostic strategy for identifying Avian Pathogenic *Escherichia coli* (APEC) based on four patterns of virulence genes. *Clinical Microbiology*, 50 (5), 1673 – 1678.
- Singer, R.S. (2015). Urinary tract infections attributed to diverse ExPec strains in food animals: evidence and data gap. *Frontiers in Microbiology*, 6(28): 1 – 9.
- Smith, J.L., Drum, D.J.V., Dai, Y., Kim, J.M., Sanchez, S., Maurer, J.J., Hofacre, C.L. and Lee, M.D.(2007a). Impact of antimicrobial usage on antimicrobial resistance in commensal *Escherichia coli* strains colonizing broiler chickens.*Applied Environmental Microbiology*, 73(3): 1404 – 1414.
- Smith, J.L., Fratamico, P.M. and Gunther, N.W.(2007b). Extraintestinal Pathogenic *Escherichia coli*. *Foodborne Pathogen and Disease*, 4(2): 134 – 163.
- Sojka, W.J. (1965.). *Escherichia coli* in domestic animals and poultry. Review series No.7 of the Commonwealth Bureau of Animal Health, Weybridge.

- Solà-Ginés, M., Cameron-Veas, K., Badiola, J., Dolz, R., Majó, N., Dahbi, G., Viso, S., Mora, A., Blanco, J., Piadra-Carraasco, N., Gonzales-Lopez, J.J. and Migura-Garcia, L. (2015) Diversity of multi-drug resistant avian pathogenic *Escherichia coli* (APEC) causing outbreaks of colibacillosis in broilers during 2012 in Spain. *PLoS ONE* 10(11):e0143191. [Doi:10.1371/journal.pone.0143191.](https://doi.org/10.1371/journal.pone.0143191)
- Son, R., Samuel, L., Gulam, R. and Zuraini, M.I.(1999). RAPD – PCR analysis, antibiotic resistance and plasmid profiles of *Escherichia coli*. *Malaysia Applied Biology*, 28(1&2): 49 – 57.
- Stenske, K.A., Bemis, D.A., Gillespie, B.E., D'Souza, D.H., Oliver, S.P., Draughon, F.A., Matteson, K.J. and Bartges, J.W.(2009).Comparison of clonal relatedness and antimicrobial susceptibility of fecal *Escherichia coli* from healthy dogs and their owners. *American Journal of Veterinary Research*, 70(9): 1108 – 1116.
- Sultana, R., Siddique, B., Ali, R., Chaudhary, S. and Maqbool, A.(2012). A study on prevalence of respiratory diseases in broiler and layer flocks in Lahore district. *Punjab Univ. Journal Zoology*, 27(1): 13 – 17.
- Szmolka, A. and Nagy, B.(2013).Multidrug resistant commensal *Escherichia coli* in animals and its impact for public health. *Frontiers Microbiology*, 4: 258.
- Talebiyan, R. Kheradmand, M., Khamesipor, F. and Faradonbeh, M.R.(2014). Multiple antimicrobial resistance of *Escherichia coli* isolated from chicken in Iran. *Veterinary Medicine International*, article ID: 491418.
- Timothy, S., Shafi, K., Howard Leatherbarrow, A., Jordan, F.T.W. and Wigley, P.(2008). Molecular epidemiology of a reproductive tract associated colibacillosis outbreak in a layer breeder flock associated with atypical avian pathogenic *Escherichia coli*. *Avian Pathology*, 37:4, pp. 375-378, [Doi: 10.1080/03079450802216579](https://doi.org/10.1080/03079450802216579)
- Tonu, N.S., Sufian, M.A., Sarker, S., Kamal, M.M., Rahman, M.H. and Hossain, M.M.(2011).Pathological study on colibacillosis in chickens and detection of *Escherichia coli* by PCR. *Bangladesh Journal of Veterinary Medicine*, 9(1): 17 – 25.
- Usein, C.R., Chitoiu, D.T., Ciontea, S., Condei, M. and Damian, M. (2009). *Escherichia coli* pathotypes associated with diarrhea in Romanian children younger than five years of age. *Japanese Journal of Infectious Disease*, 62: 289 – 293.
- Usui, M., Ozawa, S., Onozato, H., Kuge, R., Obata, Y., Uemae, T., Ngoc, P.T., Heriyanto, A., Chalemchaikit, T., Makita, K., Muramatsu, Y. and Tamura, Y. (2014). Antimicrobial susceptibility of indicator bacteria isolated from chickens in Southeast Asian countries (Vietnam,

- Indonesia, Thailand). *Japan Veterinary Medical Science*, 76(5): 685 – 692.
- Van der Westhuizen, W.A. and Bragg, R.R. (2012). Multiplex Polymerase Chain Reaction for screening Avian Pathogenic *Escherichia coli* (APEC) for virulence genes. *Avian Pathology*, 41 (1): 33 – 40.
- Vandekerchove, D., Herdt, P.D., Leavens, H., Butaye, P., Muelemans, G. and Pasmans, F. (2004b). Significance of interactions between *Escherichia coli* and respiratory pathogens in layer hen flocks suffering from colibacillosis associated mortality. *Avian Pathology*, 33 (3): 298 – 302.
- Vandekerchove, D., Vandamele, F., Adriaensen, C., Zaleska, M., Hemalsteen, J.P. and de Baets, L. (2005). Virulence associated traits in avian *Escherichia coli* comparison between isolates from colibacillosis affected and clinically healthy layer flocks. *Veterinary Microbiology*, 108: 75 – 87.
- Vincent, C., Boerlin, P., Daignault, D., Dozois, C.M., Dutil, L., Galanakis, C., Reid-Smith, R.J., Telleir, P.P., Tellis, P.A., Ziebell, A. and Manges, A.R. (2010). Food reservoir for *Escherichia coli* causing urinary tract infections. *Emerging Infectious Disease*, 16(1): 88 – 95.
- Wang, X.M., Liao, X.P., Zhang, W.J., Jiang, H.X., San, J., Zhang, M.J., He, X.F., Lao, D.X. and Liu, Y.H. (2010). Prevalence of serogroups, virulence genotypes, antimicrobial resistance of avian pathogenic *Escherichia coli* in South of China. *Foodborne Pathogen Disease*, 7(9): 1099 – 1106.
- Wang, J.Y., Tang, P., Cui, E.H., Wang, L.Q., Liu, W.H., Ren, J.J., Wu, Y., Qiu, Y.H. and Liu, H.J. (2013). Characterization of antimicrobial resistance and related resistance genes in *Escherichia coli* strains isolated from chickens in China during 2007 – 2012. *African Journal of Microbiology Research*, 7 (46): 5238 – 5247.
- Wasteson, Y. (2001). Zoonotic *Escherichia coli*. *Acta Veterinary Scandivanaica*, 95: 79 – 84.
- White, D.G., Wilson, R.A., Emery, D.A., Nagaraja, K.B. and Whittam, T.S. (1993). Clonal diversity among strains of *Escherichia coli* incriminated in turkey colisepticemia. *Veterinary Microbiology*, 34: 19 – 34.
- Wray, C. and Woodward, M.J. (1994). Laboratory Diagnosis of *Escherichia coli* infections. *Escherichia coli in Domestic Animals and Humans* (pp. 595- 620) UK: Biddles Ltd, Guilford.

- Yaguchi, K., Ogitani, T., Osawa, R., Kawano, M., Kokumai, N., Kaneshige, T., Noro, T., Masubuchi, K. and Shimizu, Y. (2010). Virulence factors of Avian Pathogenic *Escherichia coli* strains isolated from chickens with colisepticemia in Japan. *Avian Diseases*, 51(3): 656 – 662.
- Zainal Abidin, N.S., Bejo, M.H., Zakaria, Z. Abdul Rahman, O. and Bejo, S.K. (2013). Molecular Characterization of Avian Pathogenic *Escherichia coli* isolated from commercial broiler chickens with complicated chronic respiratory disease. Paper presented at the Veterinary Association Malaysia Conference, Putrajaya, 2013.
- Zhao, S., Maurer, J.J. Hubert, S.K. and White, D.G.(2005). Antimicrobial susceptibility and molecular characterization of avian pathogenic *Escherichia coli* isolates. *Veterinary Microbiology*, 107(3 – 4): 215 – 224.
- Zhao, L., Gao, S., Huan, H., Xu, X., Zhu, X., Yang, W., Gao, Q. and Liu, X. (2009). Comparison of virulence factors and expression of specific genes between uropathogenic *Escherichia coli* and avian pathogenic *E.coli* in a murine urinary tract infection model and a chicken challenge model. *Microbiology*, 155: 1634 – 1644.