



***ANTIMICROBIAL AND ANTI-QUORUM SENSING ACTIVITIES OF
JAVANESE TURMERIC (*Curcuma xanthorrhiza* Roxb.) ETHANOLIC
EXTRACT AGAINST *Pseudomonas aeruginosa****

AHMAD FIQRI MUSTAQIM BIN OTHMAN

FSTM 2019 15



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By

AHMAD FIQRI MUSTAQIM BIN OTHMAN

**Thesis Submitted to the School of Graduate Studies, Universiti
Putra Malaysia, in Fulfilment of the Requirements for the Degree of
Master of Science**

November 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

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November 2018

Chair : Assoc. Prof. Yaya Rukayadi, PhD
Faculty : Food Science and Technology

Bacteria such as *Pseudomonas aeruginosa* uses quorum sensing (QS) mechanism to regulate the production of virulence factors, swarming motility and biofilm formation. As the synthetic quorum quenching compounds such as halogenated furanones reported to be toxic for human, using medicinal plant as an alternative as an anti-quorum sensing agent have been gaining attention. The objective of this study was to evaluate the antimicrobial and the anti-quorum sensing activities of the *C. xanthorrhiza* Roxb. extract on the *P. aeruginosa* ATCC35554. The rhizome of the *C. xanthorrhiza* Roxb. was extracted using ethanol as the solvent. The crude extract were tested for antibacterial activity against *P. aeruginosa* in terms of well diffusion, minimum inhibitory concentrations (MIC) and minimum bactericidal concentrations (MBC) using the Clinical and Laboratory Standard Institute (CLSI) methods. Additional analysis on the antimicrobial activity of the extract was done on the bacterial growth using the Log₁₀ colony forming unit assay. The quenching of QS mediated swarming was done by measuring the mean diameter of the swarming colonies treated with the *C. xanthorrhiza* Roxb. extract. The pyocyanin inhibition was evaluated colorimetrically by extraction with chloroform and 0.2 M hydrochloric acid (HCl). As for the alkaline protease and the LasB protease, the supernatant of the culture treated with the extract were exposed to the skim milk agar and the casein buffered broth. Biofilm formation prevention was done using the 2, 3-bis (2-methoxy-4-nitro-5-sulfophenyl)-5-[(phenyl-amino) carbonyl]-2H-tetrazolium-hydroxide (XTT) reduction assay on a pre-sterilized 96-wells microtiter plate. The results showed that the extract can inhibit and kill the growth of the *P. aeruginosa* with MIC and MBC values of 200 and 700 mg/mL, respectively. This indicates that the inhibition and killing of this bacterium need a relatively

high concentration. Up to 200 mg/mL of the extract was used in the antibacterial assay as in the quorum quenching assays, 50 mg/mL of the extract did not exhibited significant inhibition. Interestingly, the extract at 200 mg/mL showed 72.12% reduction of swarming motility, 84% inhibition of the pyocyanin production, 50.14% and 40% decrement of alkaline protease and LasB protease secretion, respectively and 78.35% decrease in the biofilm formation. Since production of the QS virulence factors, swarming and biofilm has been known to be regulated by the multiple QS circuits, the quorum quenching activities by the ethanolic extract of the *C. xanthorrhiza* Roxb. suggest that it can interfere on these systems. In conclusion, the *C. xanthorrhiza* Roxb. extract shows a high potential for an alternative natural quorum quenching agent.

Keywords: antimicrobial, Quorum quenching, antibiofilm, *Curcuma xanthorrhiza* Roxb., swarming motility, virulence factors

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**ANTIMIKROBIAL DAN ANTI-PENGESANAN KUORUM AKTIVITI KUNYIT
JAVA (*Curcuma xanthorrhiza* Roxb.) EKSTRAK ETANOL TERHADAP
*Pseudomonas aeruginosa***

Oleh

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Pengerusi : Prof. Madya Yaya Rukayadi, PhD
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Bakteria seperti *Pseudomonas aeruginosa* menggunakan mekanisme pengesanan kuorum (QS) untuk mengawal pengeluaran faktor-faktor virulensi, "swarming" motiliti dan pembentukan "biofilm". Oleh kerana kompaun penghalang quorum sintetik seperti "halogenated furanones" dilaporkan sebagai toksik untuk manusia, penggunaan herba-herba semula jadi sebagai alternatif ejen pengesanan anti-kuorum semakin mendapat perhatian. Objektif kajian ini adalah untuk menilai aktiviti antimikrob dan anti-kuorum ekstrak *C. xanthorrhiza* Roxb. pada *P. aeruginosa* ATCC35554. Rizom *C. xanthorrhiza* Roxb. telah diekstrak menggunakan etanol sebagai pelarut. Etanol ekstrak telah diuji untuk aktiviti antibakteria terhadap *P. aeruginosa* dari segi "well diffusion", konsentrasi penghambatan minimum (MIC) dan kepekatan bakterisida minimum (MBC) berdasarkan kaedah Standard Klinikal dan Makmal Institut (CLSI). Analisis tambahan terhadap aktiviti antimikrob ekstrak juga dilakukan terhadap pertumbuhan bakteria menggunakan ujian unit pertumbuhan koloni pada Log₁₀. Penghalangan pergerakan "swarming" melalui pengesanan kuorum telah dilakukan dengan mengukur diameter koloni "swarming" yang dirawat dengan *C. xanthorrhiza* Roxb. ekstrak. Perencatan pyocyanin dinilai secara kolimetrik selepas pengekstrakan dengan kloroform dan 0.2 M asid hidroklorik (HCl). Cecair supernatan telah diekstrak daripada kultur *P. aeruginosa* telah dirawat dengan ekstrak terhadap agar susu skim untuk ujian alkali "protease" dan "buffer" kasein untuk ujian "LasB protease". Pencegahan pembentukan "biofilm" dilakukan dengan menggunakan perwarna 2, 3-bis (2-methoxy-4-nitro-5-sulfophenyl) -5 - [(phenyl-amino) karboksil] -2H-tetra-zolium-hidroksida (XTT) pada piring microtiter 96 lubang. Hasilnya menunjukkan bahawa ekstrak boleh menghalang dan membunuh pertumbuhan *P. aeruginosa* dengan nilai 200 mg/mL (ujian MIC) dan 700 mg/mL (ujian MBC). Ini menunjukkan bahawa perencatan dan pembunuhan bakteria ini memerlukan kepekatan yang agak tinggi. Ekstrak *C. xanthorrhiza* Roxb. yang digunakan pada ujian antibakteria

telah dilakukan sehingga 200 mg/mL kerana pada ujian penghambatan pengesanan kuorum, konsentrasi ekstrak pada 50 mg/mL tidak menunjukkan pengurangan yang memberansangkan. Menariknya, ekstrak pada 200 mg/mL menunjukkan pengurangan “swarming” berkurang sebanyak 72.12%, 84% daripada produksi pyocyanin, 50.14% terhadap “protease” alkali dan 40% untuk ujian LasB “protease”. Keputusan penghambatan pengesanan daripada ekstrak juga telah menunjukkan 78.35% penurunan terhadap pembentukan biofilm. Oleh kerana produksi faktor virulasi pengesanan kuorum, “swarming” dan “biofilm” telah dikawal oleh pelbagai litar dalam sistem pengesanan kuorum, penghambatan terhadap faktor-faktor virulasi seperti di atas oleh ekstrak etanol *C. xanthorrhiza* Roxb. menandakan bahawa ia boleh mengganggu sistem pengesanan kuorum. Kesimpulannya, ekstrak *C. xanthorrhiza* Roxb. menunjukkan potensi yang tinggi sebagai ejen alternatif penghalang sistem pengesanan kuorum.

Kata-kata kunci: Penghalangan kuorum, anti-“biofilm”, *Curcuma xanthorrhiza* Roxb., antibakteria, faktor-faktor virulasi

ACKNOWLEDGEMENTS

Bismillahirrahmanirrahim,

Alhamdulillah, my profound gratitude goes to Allah the Almighty and the Merciful, whom has blessed me with the strength to overcome every struggle finishing this research project "Antimicrobial and Anti-Quorum Sensing Activities of Javanese Turmeric (*Curcuma xanthorrhiza* Roxb.) Ethanolic Extract against *Pseudomonas aeruginosa*". The deepest of gratitude to my supervisor, Assoc. Prof. Dr. Yaya Rukayadi for his full support, expert guidance, constructive comments, understanding and continuous encouragement throughout my research in which without, my thesis work would have been a frustrating and an overwhelming journey. In addition, I would also like to express my gratefulness to Professor Dr. Son Radu for having served on my committee. Her thoughtful questions and comments were valued greatly. I would also like to thank all of the staffs of the Biochemistry Laboratory and Microbiology Laboratory in Faculty of Food Science and Technology, Universiti Putra Malaysia for their kindness, wisdoms and cooperation during completion of the research project.

Furthermore, I could have not also be able to finish my study without the help of my colleagues whom I could have not traded for anyone else and would like to thank you for the guidance and continuous encouragement; Illi Syuhada, Siti Alyani Mat, Siti Zaharah Rosli, Naziruddin Arrifin, Raihana Abdul Razak, Zulfa Zakuan, Nur Sadrina, Nur Aini Ismail, Nurul Husna Mahazar, Suzita RamLi, Ida Madiha, Raja Asila, Kalaivani, Almizan, and Vivian New. Not to forget, my best friends that continuously listening and supporting me; Haziq Hamzi Hamzah, Farah Harris, Fairuz Hazwani, Muhammad Amir A. Abu Bakar, Mihyoung Lee, Nik Muhamad Hanif, Nawal Amalina, and others, my deepest gratitude for the supports and comfort for all the years growing up with all of you. I will not forget the memories we have made.

I am also deeply indebted to my family especially my beloved parents, Othman Ahmad and my late mother Azlina Zakaria for every supports, encouragement, inspirations, and unconditional love that have been given to me since my birth. My heartfelt gratitude also goes to my siblings Zhorif Othman, Zahin IIman, Ikin Zakaria, Harith Othman and Adibah Hanun that have been through thick and thin during the course of my life and especially during this particular journey but still provide me with endless supports, assistance and consolation. To Ummu Aina Alias, your optimism, confidence, support and love has and will always be a refreshment and inspiration for me, thank you for everything.

As with any work, there are bound to be names I have left out here that have also had an impact at the margin on the following text, however, I am also thankful for their contributions and the help they have provided me along the way.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

UPM	Universiti Putra Malaysia
QS	Quorum sensing
AI	Autoinducer
OdDHL	N-(3-oxododecanoyl)-homoserine lactone
BHL	<i>N</i> -butanoyl-L-homoserine lactone
°C	Degree celcius
%	Percentage
mL	Milimeter
µL	Microliter
nm	Nanometer
g	Gram
mg	Miligram
Rpm	Rotation per minute
N	Normality
mM	MiliMolar
w/v	Weight per volume
XTT	2, 3-bis (2-methoxy-4-nitro-5-sulfo-phenyl)-2H-tetrazolium-5-carbo-xanilide
OD	Optical density



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CHAPTER 1

INTRODUCTION

1.1 Background

Foodborne illnesses and food spoilage has been crucially affecting the global food safety and economy (Fukuda, 2015; Gustavsson *et al.*, 2011). There have been a lot of cases reported due to consumption of foodborne pathogens that has led to mild or serious diarrhea, vomiting, abdominal pain (Adam and Moss, 2000) and even deaths (WHO, 2014; Scallan *et al.*, 2011). Such symptoms affect more on the elderly, children and immune-compromised individuals (FDA, 2018). Spoilage of food products by these organisms have also increase the food wastage and food loss throughout the globe thus affecting the economy (Bhunia, 2008). While well developed countries such as the United States and United Kingdom are able to produce good statistical reports on the foodborne illnesses nor the economic loss due to food wastage (Teisl and Roe, 2010), less urbanized countries are facing difficulties in tackling this issue due to the lack of cases report (Soon *et al.*, 2011). This is also true for Malaysia, a country that holds multiple races with varieties of delicacies that some are produced in an unhygienic environments and lack of awareness regarding food safety protocols (Abdul-Mutalib *et al.*, 2015).

Usage of antibiotics in various of antimicrobial agents such as detergents, preservatives (Gram *et al.*, 2002) are common in fighting and preventing contamination on food products and processing equipment (Wise and Soulsby, 2002). These antimicrobial agents often use the concept of selective toxicity whereby the difference between the microorganism metabolism and the structure and the corresponding features of human cells are exploited (Levinson, 2014), such as the cell wall biosynthesis, nucleic acid metabolism as well as protein synthesis (Sigma Aldrich, 2018). However, the frequent usage of these antimicrobial agents especially antibiotics have been reported to cause an increase of antibiotic resistance strain bacteria (Bhunia, 2008). Moreover, production of extracellular enzymes contributes to the host immune system evasion and protection of the bacterial colonies against antimicrobial agents (Hoge *et al.*, 2010; Hung *et al.*, 2005; Miyoshi *et al.*, 2002). As biofilm is composed of extracellular matrix of polysaccharides, it can help establishing suitable environments for wide spectrum of bacteria as well as providing enough nutrients for growth while protecting bacterial cells against host immune system via intracellular adhesions (Gotz, 2002; Mack *et al.*, 1994; Costerton *et al.*, 1987). This makes cleaning the equipment used during manufacturing of food products even challenging as the surviving bacteria could potentially act as a contamination source (Bai and Rai, 2011; Frank, 2001).

Therefore, numerous alternatives have been researched to reduce the dependencies on the usage of antibiotics in preventing microbial food contamination (Dong *et al.*, 2007). One of them includes quorum sensing (Dong *et al.*, 2007; Zhang and Dong, 2004; Hentzer and Givskov, 2003), a bacterial communication system that is used by various single-celled microorganisms such as bacterial and fungal pathogens in response to the changing environments by regulating its virulence productions (Dong *et al.*, 2007; Fuqua and Winans, 1994). Example of the bacteria that uses quorum sensing in regulating its virulence include *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa* and many more (Waters and Bassler, 2005; Zhang and Dong, 2004; Hornby *et al.*, 2001). These foodborne pathogens have been reported to form biofilm on food equipment and food product surfaces, causing serious health issues and economic losses (Kumar and Anand, 1998). *Pseudomonas aeruginosa* uses quorum sensing to regulate production of virulence factors such as alkaline protease, pyocyanin, swarming motility as well as initiate and maintain biofilm formation (Venturi, 2006; Caballero *et al.*, 2001; Davies *et al.*, 1998).

Therefore, the development of quorum quenching is deeply invested in as it would serve as an alternative method in preventing bacterial growth without killing them (Bacha *et al.*, 2016; Lee and Zhang, 2014, Hong *et al.*, 2012). Numerous non-antibiotic agents such as quorum quenching have been reported to be developed and tested on their efficacy on preventing the quorum mediated virulence production (Chatterjee *et al.*, 2016). For example, synthetic antibiotics macrolide antibiotic azithromycin (AZM) is one of the quorum quenching agents that shows an improvement against quorum sensing inhibitors during the clinical trial (Imperi *et al.*, 2014; van Delden *et al.*, 2012; Høiby, 2011). However, according to Kohler *et al.* (2010), the usage of anti-virulence such as AZM treatments to pose a long term disadvantages, whereby discontinuation from the treatment would lead to more severe infections by *Pseudomonas aeruginosa* (Köhler *et al.*, 2010). Therefore, the usage of traditional medicinal plants as quorum sensing inhibitors have gained much interest as several of them have been reported to shown quorum quenching activities (Jakobsen *et al.*, 2012; Kandasamy *et al.*, 2012; Krishnan *et al.*, 2012).

Traditional medicinal plants have always been widely used by the locals as food as well as medicine due to their highly beneficial active compounds, even now (Choo *et al.*, 2006; Vining, 1990). For example, *Curcuma xanthorrhiza* Roxb. or commonly known as Javanese turmeric or temu lawak is commonly found in Indonesia, Malaysia as well as Thailand (Sylvester *et al.*, 2015). Extensive studies have reported that the beneficial bioactive compounds present in the Javanese turmeric such as terpenoids and curcuminoids among others that show high potential as anti-cancer, anti-bacterial, anti-diuretic, anti-fungal and more (Aznam and Atun, 2016; Rukayadi and Hwang, 2007; Rukayadi *et al.*, 2006; Siagian, 2006; Afifah, 2005; Choi *et al.*, 2004; Hwang *et al.*, 2000; Ozaki, 1990). Recent studies have also shown that these traditional medicinal plants possess anti quorum sensing mechanism (Packiavathy *et al.*, 2012; Wang *et al.*, 2012; Song. *et al.*, 2010). Therefore,

this study aims to explore the effect of the Javanese turmeric extract on inhibiting the quorum sensing mechanism by the *Pseudomonas aeruginosa*.

1.2 Problem Statements

P. aeruginosa is an opportunistic and persistent gram negative bacteria that is commonly associated with foodborne illnesses and food spoilage such as in milk, vegetables and fruits (Motarjemi *et al.*, 2013; Hossain, 2013; Arslan *et al.*, 2011). *Pseudomonas* spp. can cause spoilage in dairy products (Cousin *et al.*, 2001) via the production of the stable extracellular enzymes (Herrera, 2001) including lecithinases (Ray, 2004; Herrera, 2001), protease and lipases (Arslan *et al.*, 2011; Ray, 2004). These enzymes are able to impairs the milk coagulation process causing it to elicit bitter or rancid flavors (Richter and Vedamuthu, 2001). *P. aeruginosa* can also cause food spoilage in blue milk which often is pathogenic towards human and animals as a secondary infection (Collins *et al.*, 1989). *P. aeruginosa* has also been reported to cause high fever, headache and diarrhoea as well as bacteraemia and septicemia in the host blood stream (Motarjemi *et al.*, 2013).

Quorum sensing (QS) is a bacteria cell-to-cell communication mechanism used to determine the bacterial physiology such as local population density as well as applying strategies to adapt to the changes in the environment (Ahmad *et al.*, 2015; Skandamis and Nychas, 2012; Szabó *et al.*, 2010; Choo *et al.*, 2006; Fuqua and Greenberg, 2002). Quorum sensing allows the bacteria to increase their resistance towards antibacterial agents and facilitate bacterial pathogenesis while protecting the growing quorum from the harsh environment (Wei and Ma, 2013). However, usage of synthetic chemical quorum quenchers that focuses on their bactericidal effect have been reported to increase the selection of resistant strains (Ugurlu *et al.*, 2016; Gyawali and Ibrahim, 2014; Lewis, 2008). Synthetic quorum quencher might also have other detrimental effect for example, halogenated furanone have been reported to be chemically reactive, unstable and toxic to human cells (Hentzer and Givskov, 2003).

Therefore, with the increasing emergence of multidrug resistant pathogens, rather than killing or inhibiting bacterial growth, alternative approaches that rely on inhibition of pathogenic traits such as quorum sensing using natural products have gain popularity in the research society (Ahmad *et al.*, 2015; Lew *et al.*, 2015). Javanese turmeric (*Curcuma xanthorrhiza* Roxb.) or locally known as “temu lawak” can be found in tropical countries such as Malaysia and Indonesia (Musfiroh *et al.*, 2013). *C. xanthorrhiza* has been traditionally used for food and medicinal purposes (Lim, 2016; Park *et al.*, 2008). Studies have reported that *C. xanthorrhiza* possesses several of biological activities such as anti-tumor (Itokawa *et al.*, 1985), anti-bacterial (Hentschel *et al.*, 1996; Rukayadi *et al.*, 2006; Hwang *et al.*, 2000) and anti-inflammatory (Lee *et al.*,

2002; Claeson *et al.*, 1996) among others. The Javanese turmeric have been reported to have bioactive compounds including curcuminoids, camphor, geranyl acetate, zerumbone, β -curcumene, zingiberene, ar-curcumene and xanthorrhizol (Jantan *et al.*, 2012). These bioactive compounds such as xanthorrhizol have been proven to elicit biological properties such as antifungal (Lew *et al.*, 2015; Rukayadi *et al.*, 2006), antimicrobial (Lew *et al.*, 2015), anticancer (Choi *et al.*, 2004) activities. Therefore, the aim of this study is to determine the QS inhibition activity of the Javanese turmeric (*C. xanthorrhiza* Roxb.) on the quorum sensing mechanisms of *P. aeruginosa* *in vitro*.

1.3 Objectives

The objectives of this study are:

1. To extract the rhizome of the *C. xanthorrhiza* Roxb. and determine the non-killing or inhibiting concentration of against growth of *P. aeruginosa*
2. To evaluate the inhibitory activity of the *C. xanthorrhiza* Roxb. on the *P. aeruginosa* quorum sensing regulated virulence factors and biofilm formation.

REFERENCES

- Ab Halim, M. R., Mahmud, M., Z., T., M. S., Mahmud, R., and Ismail, S. 2012. Standardization and phytochemical studies of *Curcuma xanthorrhiza* Roxb. *International Journal of Pharmacy and Pharmaceutical Sciences*, 4(3), 606–610.
- Abd Ghafar, S. W. 2017. Food waste in malaysia: Trends, current practices and key challenges.
- Abdul-Mutalib, N. A., Syafinaz, A. N., Sakai, K., and Shirai, Y. 2015. An overview of foodborne illness and food safety in Malaysia. *International Food Research Journal*, 22(3), 896–901.
- Adams, M. S., and Moss, M. O. 2000. *Food Microbiology* (2nd ed.). Cambridge, UK: The Royal Society of Chemistry.
- Addis, M., and Sisay, D. 2015. A review on major food borne bacterial illnesses. *J Trop Dis*, 3(3).
- Ahmad, A., Viljoen, A. M., and Chenia, H. Y. 2015. The impact of plant volatiles on bacterial quorum sensing. *Letters in Applied Microbiology*, 60(1), 8–19.
- Aida, W. 2011. Effect of ethanol concentration, extraction time and extraction temperature on the recovery of phenolic compounds and antioxidant capacity of *Centella asiatica* extracts. *International Food Research Journal*, 18, 571–578.
- Ak, T., and Gülçin, İ. 2008. Antioxidant and radical scavenging properties of curcumin. *Chemico-Biological Interactions*, 174(1), 27–37.
- Al-Rubiay, K. K., Jaber, N. N., Al Mhaawe, B. H., and Alrubaay, L. K. 2008. Antimicrobial of henna extract. *Oman Medical Journal*, 23(4).
- Alipour, M., Suntres, Z. E., Lafrenie, R. M., and Omri, A. 2010. Attenuation of *Pseudomonas aeruginosa* virulence factors and biofilms by co-encapsulation of bismuth–ethanedithiol with tobramycin in liposomes. *Journal of Antimicrobial Chemotherapy*, 65, 684–693.
- Allesen-Holm, M., Barken, K. B., Yang, L., Klausen, M., Webb, J. S., Kjelleberg, S., and Tolker-Nielsen, T. 2006. A characterization of DNA release in *Pseudomonas aeruginosa* cultures and biofilms. *Molecular Microbiology*, 59, 1114–1128.
- Allison, C., and Hughes, C. 1991. Bacterial swarming: an example of prokaryotic differentiation and multicellular behaviour. *Science Progress*, 75(298 Pt 3-4), 403–22.
- Anderson, G. G., and O'Toole, G. A. 2008. Innate and induced resistance mechanisms of bacterial biofilms. *Curr.Top Microbiol. Immunol.*, 322,

- Andrejko, M., Zdybicka-Barabas, A., Janczarek, M., and Cytryńska, M. 2013. Three *Pseudomonas aeruginosa* strains with different protease profiles.
- Anand, P., Thomas, S. G., Kunnumakkara, A. B., Sundaram, C., Harikumar, K. B., Sung, B., and Aggarwal, B. B. 2008. Biological activities of curcumin and its analogues (Congeners) made by man and Mother Nature. *Biochemical Pharmacology*, 76(11), 1590–1611.
- Anggakusuma, Yanti, Lee, M., and Hwang, J.-K. 2009. Estrogenic activity of xanthorrhizol isolated from *Curcuma xanthorrhiza* Roxb. *Biological and Pharmaceutical Bulletin*, 32(11), 1892–7.
- Arslan, S., Eyi, A., and Özdemir, F. 2011. Spoilage potentials and antimicrobial resistance of *Pseudomonas* spp . isolated from cheeses. *Journal of Dairy Science*, 94, 5851–5856.
- Arun Kumar, S., and Muthuselvam, M. 2009. Analysis of phytochemical constituents and antimicrobial activities of *Aloe vera* l. against clinical pathogens. *World J. Agri. Sci.*, 5, 572–576.
- Aybey, A., Demirkan, E., and Aybey, C. A. 2016. Inhibition of quorum sensing-controlled virulence factors in *Pseudomonas aeruginosa* by human serum paraoxonase. *Journal of Medical Microbiology*, 65, 105–113.
- Aznam, N., and Atun, S. 2016. Pharmacological test of herbal products from temulawak (*Curcuma xanthorrhiza*) as antihypercholesterol by *in vivo*. *International Journal of Pharmacognosy and Phytochemical Research*, 8(5), 807–811.
- Azwanida, N. N. 2015. A review on the extraction methods use in medicinal plants , principle, strength and limitation. *Med. Aromat. Plants*, 4,196.
- Bacha, K., Tariku, Y., Gebreyesus, F., Zerihun, S., Mohammed, A., Weiland-Bräuer, N., and Mulat, M. 2016. Antimicrobial and anti-Quorum Sensing activities of selected medicinal plants of Ethiopia: Implication for development of potent antimicrobial agents. *BMC Microbiology*, 16.
- Bahoran, T., Soobrattee, M. A., Luximon-Ramma, V., and Aruoma, O. I. 2007. Free radicals and antioxidants in cardiovascular health and disease. *Internet Journal of Medical Update - Ejournal*, 1(2).
- Bai, A. J., and Rai, V. R. 2011. Bacterial Quorum Sensing and Food Industry. *Comprehensive Reviews in Food Science and Food Safety*, 10(3), 183–193.
- Balaban, N., and Koyfman, N. 2001. Peptides: Bacteria's point of view. *Peptides*, 22(10), 1517–1518.
- Balkwill, F., and Mantovani, A. 2001. Inflammation and cancer: back to Virchow *The Lancet*, 357(9255), 539–545.

- Bardoel, B. W., van der Ent, S., Pel, M. J. C., Tommassen, J., Pieterse, C. M. J., van Kessel, K. P. M., and van Strijp, J. A. G. 2011. *Pseudomonas* evades immune recognition of flagellin in both mammals and plants. *PLoS Pathogens*, 7(8), e1002206.
- Barraud, N., Hassett, D. J., Hwang, S.-H., Rice, S. A., Kjelleberg, S., and Webb, J. S. 2006. Involvement of Nitric Oxide in Biofilm Dispersal of *Pseudomonas aeruginosa*. *Journal of Bacteriology*, 188(21), 7344–7353.
- Barrett, A. J., Rawlings, N. D., and Woessner, J. F. 2004. *Handbook of proteolytic enzymes* (2nd ed.). London: Academic Press.
- Barken, K. B., Pamp, S. J., Yang, L., Gjermansen, M., Bertrand, J. J., Klausen, M., and Tolker-Nielsen, T. 2008. Roles of type IV pili, flagellum-mediated motility and extracellular DNA in the formation of mature multicellular structures in *Pseudomonas aeruginosa* biofilms. *Environmental Microbiology*, 10(9), 2331–2343.
- Basile, V., Ferrari, E., Lazzari, S., Belluti, S., Pignedoli, F., and Imbriano, C. 2009. Curcumin derivatives: Molecular basis of their anti-cancer activity. *Biochemical Pharmacology*, 78(10), 1305–1315.
- Batubara, I., Julita, I., Darusman, L. K., Muddathir, A. M., and Mitsunaga, T. 2015. Flower bracts of temulawak (*Curcuma xanthorrhiza*) for skin care: anti-acne and whitening agents. *Procedia Chemistry*, 14(14), 216–224.
- Baumann, U., Wu, S., Flaherty, K. M., and McKay, D. B. 1993. Three-dimensional structure of the alkaline protease of *Pseudomonas aeruginosa*: a two-domain protein with a calcium binding parallel beta roll motif. *The EMBO Journal*, 12(9), 3357–64.
- Bean, N. H., and Griffins, P. M. 1990. Foodborne disease outbreaks in the United States. *Pathogens, Vehicles and Trends*, 53, 804–817.
- Bever, R. A., and Iglewski, B. H. 1988. Molecular characterization and nucleotide sequence of the *Pseudomonas aeruginosa* elastase structural gene. *Journal of Bacteriology*, 170(9), 4309–14.
- Bhunia, A. K. 2008. *Foodborne microbial pathogens: Mechanisms and pathogenesis*. West Lafayette, IN: Springer.
- Bhunia, A. K. 2008. Introduction of Foodborne Pathogens. In *Foodborne Microbial Pathogens* (pp. 1–16).
- Bijina, B., Chellappan, S., Krishna, J. G., Basheer, S. M., Elyas, K. K., Bahkali, A. H., and Chandrasekaran, M. 2011. Protease inhibitor from *Moringa oleifera* with potential for use as therapeutic drug and as seafood preservative. *Saudi Journal of Biological Sciences*, 18, 273–281.
- Bisswanger, H. 2014. Enzyme assays. *Perspectives in Science*, 1(1–6), 41–55.

- Bjorn, M. J., Sokol, P. A., and Iglewski, B. H. 1979. Influence of iron on yields of extracellular products in *Pseudomonas aeruginosa* cultures. *Journal of Bacteriology*, 138(1), 193–200.
- Bleves, S., Viarre, V., Salacha, R., Michel, G. P. F., Filloux, A., and Voulhoux, R. 2010. Protein secretion systems in *Pseudomonas aeruginosa*: A wealth of pathogenic weapons. *International Journal of Medical Microbiology*, 300(8), 534–543.
- Borges, A., Saavedra, M. J., and Simões, M. 2012. The activity of ferulic and gallic acids in biofilm prevention and control of pathogenic bacteria. *Biofouling*, 28(7), 755–767.
- Borges, A., Serra, S., Cristina Abreu, A., Saavedra, M. J., Salgado, A., and Simões, M. 2014. Evaluation of the effects of selected phytochemicals on quorum sensing inhibition and in vitro cytotoxicity. *Biofouling*, 30(2), 183–195.
- Britigan, B. E., Railsback, M. A., and Cox, C. D. 1999. The *Pseudomonas aeruginosa* secretory product pyocyanin inactivates alpha1 protease inhibitor: implications for the pathogenesis of cystic fibrosis lung disease. *Infection and Immunity*, 67(3), 1207–12.
- Braun, P., de Groot, A., Bitter, W., and Tommassen, J. 1998. Secretion of elastinolytic enzymes and their propeptides by *Pseudomonas aeruginosa*. *Journal of Bacteriology*, 180(13), 3467–9.
- Brencic, A., and Lory, S. 2009. Determination of the regulon and identification of novel mRNA targets of *Pseudomonas aeruginosa* RsmA. *Molecular Microbiology*, 72(3), 612–632.
- Burman, A. R., Savel, R. H., Racine, S., Swanson, B. L., Revadigar, N. S., Fujimoto, J., and Wiener-Kronish, J. P. 2001. Type III protein secretion is associated with death in lower respiratory and systemic *Pseudomonas aeruginosa* infections. *The Journal of Infectious Diseases*, 183(12), 1767–1774.
- Byrd, M. S., Sadovskaya, I., Vinogradov, E., Lu, H., Sprinkle, A. B., Richardson, S. H., and Wozniak, D. J. 2009. Genetic and biochemical analyses of the *Pseudomonas aeruginosa* Psl exopolysaccharide reveal overlapping roles for polysaccharide synthesis enzymes in Psl and LPS production. *Molecular Microbiology*, 73(4), 622–638.
- Caballero, A. R., Moreau, J. M., Engel, L. S., Marquart, M. E., Hill, J. M., and O'Callaghan, R. J. 2001. *Pseudomonas aeruginosa* Protease IV Enzyme Assays and Comparison to Other *Pseudomonas* Proteases. *Analytical Biochemistry*, 290(2), 330–337.
- Caiazza, N. C., and O'toole, G. A. 2004. SadB Is Required for the Transition from Reversible to Irreversible Attachment during Biofilm Formation by *Pseudomonas aeruginosa* PA14. *Journal Of Bacteriology*, 186(14), 4476–4485.

- Caiazza, N. C., Shanks, R. M. Q., and O'toole, G. A. 2005. Rhamnolipids Modulate Swarming Motility Patterns of *Pseudomonas aeruginosa*. *Journal Of Bacteriology*, 187(21), 7351–7361.
- Caiazza, N. C., Merritt, J. H., Brothers, K. M., and O'Toole, G. A. 2007. Inverse regulation of biofilm formation and swarming motility by *Pseudomonas aeruginosa* PA14. *Journal of Bacteriology*, 189(9), 3603–12.
- Casilag, F., Lorenz, A., Krueger, J., Klawonn, F., Weiss, S., and Häussler, S. 2015. LasB elastase of *Pseudomonas aeruginosa* acts in concert with alkaline protease AprA to prevent flagellin-mediated immune recognition. *Infection and Immunity*, 84(1), 162–171.
- Chatterjee, M., Saluja, R., Kanneganti, S., Chinta, S., and Dikshit, M. 2007. Biochemical and molecular evaluation of neutrophil NOS in spontaneously hypertensive rats. *Cellular and molecular biology*. 53(1), 84-93.
- Chatterjee, M., Anju, C. P., Biswas, L., Kumar, V. A., Mohan, C. G., and Biswas, R. 2016. Antibiotic resistance in *Pseudomonas aeruginosa* and alternative therapeutic options. *International Journal of Medical Microbiology*, 306, 48–58.
- Chan, K. G., Atkinson, S., Mathee, K., Sam, C. K., Chhabra, S. R., Cámara, M., and Al, E. 2011. Characterization of N-acylhomoserine lactone-degrading bacteria associated with *Zingiber officinale* (ginger) rhizosphere: Co-existence of quorum quenching and Quorum Sensing in Acinetobacter and Burkholderia. *BMC Microbiology*, 11(51).
- Chen, B. G., Turner, L., and Berg, H. C. 2007. The wetting agent required for swarming in *Salmonella enterica serovar typhimurium* is not a surfactant. *Journal of Bacteriology*, 189(23), 8750–3.
- Choi, M.-A., Kim, S. H., Chung, W.-Y., Hwang, J.-K., and Park, K.-K. 2004. Xanthorrhizol, a natural sesquiterpenoid from *Curcuma xanthorrhiza*, has an anti-metastatic potential in experimental mouse lung metastasis model. *Biochemical and Biophysical Research Communications*, 326(1), 210–217.
- Choo, J. H., Rukayadi, Y., and Hwang, J. K. 2006. Inhibition of bacterial quorum sensing by vanilla extract. *Letters in Applied Microbiology*, 42(6), 637–641.
- Christen, M., Christen, B., Folcher, M., Schauerte, A., and Jenal, U. 2005. Identification and Characterization of a Cyclic di-GMP-specific Phosphodiesterase and Its Allosteric Control by GTP. *Journal of Biological Chemistry*, 280(35), 30829–30837.
- Chien Bong, C. P., Ho, W. S., Hashim, H., Lim, J. S., Ho, C. S., Peng Tan, W. S., and Lee, C. T. 2016. Review on the renewable energy and solid waste management policies towards biogas development in Malaysia. *Renewable and Sustainable ENergy Reviews*.

- Choudhuri, T., Pal, S., Agwarwal, M. L., Das, T., and Sa, G. 2002. Curcumin induces apoptosis in human breast cancer cells through p53-dependent Bax induction. *FEBS Letters*, 512(1–3), 334–40. Retrieved from
- Chun, K.-S., Keum, Y.-S., Han, S. S., Song, Y.-S., Kim, S.-H., and Surh, Y.-J. 2003. Curcumin inhibits phorbol ester-induced expression of cyclooxygenase-2 in mouse skin through suppression of extracellular signal-regulated kinase activity and NF- κ B activation. *Carcinogenesis*, 24(9), 1515–1524.
- Chung, W. Y., Park, J. H., Kim, M. J., Kim, H. O., Hwang, J. K., Lee, S. K., and Park, K. K. 2007. Xanthorrhizol inhibits 12-O-tetradecanoylphorbol-13-acetate-induced acute inflammation and two-stage mouse skin carcinogenesis by blocking the expression of ornithine decarboxylase, cyclooxygenase-2 and inducible nitric oxide synthase through mitogen-activated protein kinases and/or the nuclear factor- κ B. *Carcinogenesis*, 28(6), 1224–1231.
- Chong-Lek, K., Sam, C.-K., Yin, W.-F., Tan, L. Y., Krishnan, T., Chong, Y. M., and Chan, K.-G. 2013. Plant-derived natural products as sources of anti-quorum sensing compounds. *Sensors*, 13, 6217–6228
- Claeson, P., Pongprayoon, U., Sematong, T., Tuchinda, P., Reutrakul, V., Soontornsaratune, P., and Taylor, W. 1996. Non-Phenolic Linear Diarylheptanoids from *Curcuma xanthorrhiza*: A Novel Type of Topical Anti-Inflammatory Agents: Structure-Activity Relationship. *Planta Medica*, 62(3), 236–240.
- Collins, C. H., Lyne P.M., and Grange J. M. 1989. *Collins and Lyne's Microbiological Methods (Pseudomonas, Acinetobacter, Alcaligenes, Flavobacterium, Chromobacterium and Acetobacter)* (6th ed.). Butterworth-Heinemann, Oxford, UK: Oxford University Press Inc.
- Colvin, K. M., Gordon, V. D., Murakami, K., Borlee, B. R., Wozniak, D. J., Wong, G. C. L., and Parsek, M. R. 2011. The Pel Polysaccharide Can Serve a Structural and Protective Role in the Biofilm Matrix of *Pseudomonas aeruginosa*. *PLoS Pathogens*, 7(1).
- Conboy, L., Kaptchuk, T. J., Eisenberg, D. M., Gottlieb, B., and Acevedo-Garcia, D. 2007. The relationship between social factors and attitudes toward conventional and CAM practitioners. *Complement Ther. Clin. Pract.*, 13, 146–157.
- Connelly, M. B., Young, G. M., and Sloma, A. 2004. Extracellular Proteolytic Activity Plays a Central Role in Swarming Motility in *Bacillus subtilis*. *Journal of bacteriology*, 186(13), 4159–4167.
- Cope, W. G., Leidy, R. B., and Hodgson, E. 2004. Classes of Toxicants: Use Classes (chptr). In E. Hodgson (Ed.), *A Textbook of Modern Toxicology* (pp. 50–60). Hoboken, NJ, USA: John Wiley and Sons, Inc.
- Copeland, M. F., and Weibel, D. B. 2009. Bacterial Swarming: A Model

- System for Studying Dynamic Self-assembly. *Soft Matter*, 5(6), 1174–1187.
- Cornelis, P., Dingemans, J., Cellier, M. F., and Wilks, A. 2013. *Pseudomonas aeruginosa* adapts its iron uptake strategies in function of the type of infections.
- Costerton, J. W., Cheng, K. J., Geesey, G. G., Ladd, T. I., Nickel, J. C., Dasgupta, M., and Marrie, T. J. 1987. Bacterial biofilms in nature and disease. *Ann. Rev. Microbiol*, 41, 435–464.
- Cousin, M. A., Vasavada, P. C., and Critzer, F. J. 2001. Psychrotrophic microorganisms. In Downes, F. P. and Ito, K. (Eds.), *Compendium of Methods for the Micro-biological Examination of Foods* (pp. 159–166). Washington, DC: American Public Health Association.
- Coussens, L. M., and Werb, Z. 2002. Inflammation and cancer. *Nature*, 420(6917), 860–867.
- CSPI. Outbreak Alert. 2014. A Review of Foodborne Illness in America from 2002-2011. *Center for Science in the Public Interest*.
- D'Argenio, D. A., and Miller, S. I. 2004. Cyclic di-GMP as a bacterial second messenger. *Microbiology*, 150(8), 2497–2502.
- Dacheux, D., Attree, I., Schneider, C., and Toussaint, B. 1999. Cell death of human polymorphonuclear neutrophils induced by a *Pseudomonas aeruginosa* cystic fibrosis isolate requires a functional type III secretion system. *Infection and Immunity*, 67(11), 6164–7.
- Dai, J., and Mumper, R., J. 2010. Plant phenolics: Extraction, analysis and their antioxidant and anticancer properties. *Molecules*. 15, 7313-52.
- Daniels, R., Vanderleyden, J., and Michiels, J. 2004. Quorum sensing and swarming migration in bacteria. *FEMS Microbiol. Rev.*, 28(3), 261–289.
- Das, T., and Manefield, M. 2012. Pyocyanin Promotes Extracellular DNA Release in *Pseudomonas aeruginosa*.
- Datta, S., Debanjan Jana, B., Tilak Raj Maity, B., Aveek Samanta, B., and Rajarshi Banerjee, B. 2016. Piper betle leaf extract affects the quorum sensing and hence virulence of *Pseudomonas aeruginosa* PAO1. *Biotech*, 6(18).
- Davies, D. G., Parsek, M. R., Pearson, J. P., Iglewski, B. H., Costerton, J. W., and Greenberg, E. P. 1998. The involvement of cell-to-cell signals in the development of a bacterial biofilm. *Science (New York, N. Y.)*, 280(5361), 295–8.
- Defoirdt, T., Miyamoto, C. M., Wood, T. K., Meighen, E. A., Sorgeloos, P., Verstraete, W., and Bossier, P. 2007. The natural furanone (5Z)-4-bromo-5-(bromomethylene)-3-butyl-2(5H)-furanone disrupts quorum

- sensing- regulated gene expression in *Vibrio harveyi* by decreasing the DNA- binding activity of the transcriptional regulator protein luxR. *Environmental Microbiology*, 9, 2486–2495.
- de Kievit, T. R. and Iglewski, B. H. 2000. Bacterial quorum sensing in pathogenic relationships. *Infection and immunity*. 68(9), 4839-4849.
- de Kievit, T. R., Gillis, R., Marx, S., Brown, C., and Iglewski, B. H. 2001. Quorum-Sensing Genes in *Pseudomonas aeruginosa* Biofilms: Their Role and Expression Patterns. *Applied and Environmental Microbiology*, 67(4), 1865–1873.
- de Kievit, T. R. 2009. Quorum sensing in *Pseudomonas aeruginosa* biofilms. *Environmental Microbiology*, 11(2), 279–288.
- de la Fuente-Núñez, C., Korolik, V., Bains, M., Nguyen, U., Breidenstein, E. B. M., Horsman, S., and Hancock, R. E. W. 2012. Inhibition of bacterial biofilm formation and swarming motility by a small synthetic cationic peptide. *Antimicrobial Agents and Chemotherapy*, 56(5), 2696–704.
- Dekimpe, V., and Déziel, E. 2009. Revisiting the quorum-sensing hierarchy in *Pseudomonas aeruginosa*: the transcriptional regulator RhIR regulates LasR-specific factors. *Microbiology*, 155, 712–723.
- Demple, B., and Harrison, L. 1994. Repair of oxidative damage to DNA: Enzymology and biology. *Annual Review of Biochemistry*, 63(1), 915–948.
- Deng, Y., Wu, J., Tao, F., and Zhang, L.-H. 2011. Listening to a new language: DSF-Based quorum sensing in gram-negative bacteria. *Chemical Reviews*, 111(1), 160–173.
- Denning, G. M., Railsback, M. A., Rasmussen, G. T., Cox, C. D., and Britigan, B. E. 1998. *Pseudomonas* pyocyanine alters calcium signaling in human airway epithelial cells. *The American Journal of Physiology*, 274(6 Pt 1), L893-900.
- Deretic, V., Gill, J. F., and Chakrabarty, A. M. 1987. Gene *algD* coding for GDP-mannose dehydrogenase is transcriptionally activated in mucoid *Pseudomonas aeruginosa*. *Journal of Bacteriology*, 169(1), 351–8.
- Deretic, V., and Konyecsni, W. M. 1989. Control of mucoidy in *Pseudomonas aeruginosa*: transcriptional regulation of *algR* and identification of the second regulatory gene, *algQ*. *Journal of Bacteriology*, 171(7), 3680–8.
- Devaraj, S., Esfahani, A. S., Ismail, S., Ramanathan, S., and Yam, M. F. 2010. Evaluation of the antinociceptive activity and acute oral toxicity of standardized ethanolic extract of the rhizome of *Curcuma xanthorrhiza* Roxb. *Molecules*, 15(4), 2925–2934.
- Deziel, E., Lepine, F., Milot, S., He, J., Mindrinos, M. N., Tompkins, R. G., and Rahme, L. G. 2004. Analysis of *Pseudomonas aeruginosa* 4-hydroxy-2-

- alkylquinolines (HAQs) reveals a role for 4-hydroxy-2-heptylquinoline in cell-to-cell communication. *Proceedings of the National Academy of Sciences*, 101, 1339–1344.
- Déziel, E., Lépine, F. O., Milot, S., and Villemur, R. 2003. *rhlA* is required for the production of a novel biosurfactant promoting swarming motility in *Pseudomonas aeruginosa*: 3-(3-hydroxyalkanoyloxy)alkanoic acids (HAAs), the precursors of rhamnolipids. *Microbiology*, 149, 2005–2013.
- Diastuti, H., Syah, Y. M., Juliawaty, L. D., and Singgih, M. 2014. Antibacterial *Curcuma xanthorrhiza* extract and fractions. *Journal of Mathematical and Fundamental Sciences*, 46(3), 224–234.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., and Ju, Y. H. 2014. Effect of extraction solvent on total phenol content, total flavonoid content and antioxidant activity of *Limnophila aromatica*. *Journal of Food and Drug Analysis*. 22(3), 296–302.
- Dong, Y.-H., Xu, J. L., Li, X. Z., and Zhang, L. H. 2000. AiiA, an enzyme that inactivates the acylhomoserine lactone quorum-sensing signal and attenuates the virulence of *Erwinia carotovora*. *Proceedings of the National Academy of Sciences*, 97(7), 3526–3531.
- Dong, Y.-H., Wang, L.-H., Xu, J. L., Zhang, H.-B., Zhang, X.-F., and Zhang, L.-H. 2001. Quenching quorum-sensing-dependent bacterial infection by an N-acyl homoserine lactonase. *Nature*, 411(6839), 813–817.
- Dong, Y., Wang, L., and Zhang, L. H. 2007. Quorum-quenching microbial infections: Mechanisms and implications. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 362(1483), 1201–11.
- Dröge, W. 2002. Free radicals in the physiological control of cell function. *Physiological Reviews*, 82(1), 47–95.
- Duan, K., and Surette, M. G. 2007. Environmental regulation of *Pseudomonas aeruginosa* PAO1 Las and Rhl quorum-sensing systems. *Journal of Bacteriology*, 189(13), 4827–4836.
- Duong, F., Lazdunski, A., and Murgier, M. 1996. Protein secretion by heterologous bacterial ABC-transporters: the C-terminus secretion signal of the secreted protein confers high recognition specificity. *Molecular Microbiology*, 21(3), 459–70.
- Eberl, L., Christiansen, G., Molin, S., and Givskov, M. 1996. Differentiation of *Serratia liquefaciens* into swarm cells is controlled by the expression of the *flhD* master operon. *Journal of Bacteriology*, 178(2), 554–9. Retrieved from
- Engebretson, J. 2002. Culture and complementary therapies. *Complement Ther. Nurs. Midwifery*, 8, 177–184.

- Essar, D. W., Eberly, L., Hadero, A., and Crawford, I. P. 1990. Identification and characterization of genes for a 2nd Anthranilate Synthase in *Pseudomonas aeruginosa*: Interchangeability of the 2 anthranilate synthases and evolutionary implications. *Journal of Bacteriology*, 172(2), 884–900.
- Estrela, A. B., and Abraham, W. R. 2010. Combining biofilm-controlling compounds and antibiotics as a promising new way to control biofilm infections. *Pharmaceuticals*, 3(5), 1374–1393.
- Fagerlind, M. G., Webb, J. S., Barraud, N., McDougald, D., Jansson, A., Nilsson, P. and Rice, S. A. 2011. Dynamic modelling of cell death during biofilm development. *Journal of Theoretical Biology*, 295, 23–36.
- FAO. 2015. Food loss and food waste.
- Farr, S. B., and Kogoma, T. 1991. Oxidative stress responses in *Escherichia coli* and *Salmonella typhimurium*. *Microbiological Reviews*, 55(4), 561–85.
- FDA. 2018. Foodborne Illness: Especially Dangerous for the Vulnerable.
- Filloux, A., and Ramos, J.-L. (Eds.). 2014. *Pseudomonas Methods and Protocols Methods in Molecular Biology* 1149. New York: Humana Press.
- Finkel, S. E., and Kolter, R. 2001. DNA as a nutrient: novel role for bacterial competence gene homologs. *Journal of Bacteriology*, 183(21), 6288–93.
- Flemming, H.-C., Neu, T. R., and Wozniak, D. J. 2007. The EPS matrix: the "house of biofilm cells". *Journal of Bacteriology*, 189(22), 7945–7.
- Flemming, H. C., and Wingender, J. 2010. The biofilm matrix. *Nature Reviews Microbiology*, 8(9), 623–633.
- Frank, D. W. 1997. The exoenzyme S regulon of *Pseudomonas aeruginosa*. *Molecular Microbiology*, 26(4), 621–9.
- Frank, J. F. 2001. Microbial attachment to food and food contact surfaces. *Adv. Food Nutr.*, 43, 319–369.
- Franzetti, L., and Scarpellini, M. 2007. Characterisation of *Pseudomonas* spp . isolated from foods, 57(1), 39–47.
- Friedman, L., and Kolter, R. 2004. Genes involved in matrix formation in *Pseudomonas aeruginosa* PA14 biofilms. *Molecular Microbiology*, 51(3), 675–90.
- Friedman, L., and Kolter, R. 2004. Two genetic loci produce distinct carbohydrate-rich structural components of the *Pseudomonas aeruginosa* biofilm matrix. *Journal of Bacteriology*, 186(14), 4457–4465.

- Fukuda, K. 2015. Food safety in a globalized world. *WHO*, 93, 212.
- Fuqua, W. C., and Winans, S. C. 1994. A LuxR-LuxI type regulatory system activates *Agrobacterium* Ti plasmid conjugal transfer in the presence of a plant tumor metabolite. *Journal of Bacteriology*, 176(10), 2796-2806.
- Fuqua, C., and Greenberg, E. P. 2002. Signalling; Listening in on bacteria: Acyl-homoserine lactone signalling. *Nature Reviews Molecular Cell Biology*, 3(9), 685-695.
- Gambello, M. J., and Iglewski, B. H. 1991. Cloning and characterization of the *Pseudomonas aeruginosa lasR* gene, a transcriptional activator of elastase expression. *Journal of Bacteriology*, 173, 3000-3009.
- Gambello, M. J., Iglewski, B. H., and Kaye, S. 1993. *lasR* of *Pseudomonas aeruginosa* is a transcriptional activator of the alkaline protease gene (*apr*) and an enhancer of exotoxin A expression. *Infection and Immunity*, 61, 1180-1184.
- Gallagher, L. A., McKnight, S. L., Kuznetsova, M. S., Pesci, E. C., and Manoil, C. 2002. Functions required for extracellular quinolone signaling by *Pseudomonas aeruginosa*. *Journal of Bacteriology*, 184, 6472-6480.
- Galloway, D. R. 1991. *Pseudomonas aeruginosa* elastase and elastolysis revisited: Recent developments. *Molecular Microbiology*, a10), 2315-21.
- Ghannoum, M. A., and Rice, L. B. 1999. Antifungal agents: Mode of action, mechanisms of resistance, and correlation of these mechanisms with bacterial resistance, 12(4), 501-517.
- Gloag, E. S., Turnbull, L., Huang, A., Vallotton, P., Wang, H., Nolan, L. M., and Whitchurch, C. B. 2013. Self-organization of bacterial biofilms is facilitated by extracellular DNA. *Proceedings of the National Academy of Sciences*, 110(28), 11541-11546.
- Golberg, K., Pavlov, V., Marks, R. S., and Kushmaro, A. 2013. Coral-associated bacteria, quorum sensing disrupters, and the regulation of biofouling. *Biofouling*, 29, 669-682.
- Gonzalez, J. E., and Keshavan, N. D. 2006. Messing with bacterial quorum sensing. *Microbiology and Molecular Biological Review*, 70, 859-875.
- Goodman, A. L., Kulasekara, B., Rietsch, A., Boyd, D., Smith, R. S., and Lory, S. 2004. A signaling network reciprocally regulates genes associated with acute infection and chronic persistence in *Pseudomonas aeruginosa*. *Developmental Cell*, 7(5), 745-754.
- Goodman, A. L., Merighi, M., Hyodo, M., Ventre, I., Filloux, A., and Lory, S. 2009. Direct interaction between sensor kinase proteins mediates acute and chronic disease phenotypes in a bacterial pathogen. *Genes and Development*, 23(2), 249-259.

- Gotz, F. 2002. *Staphylococcus* and biofilms. *Molecular Microbiology*, 43, 1367–1378.
- Govan, J. R., and Deretic, V. 1996. Microbial pathogenesis in cystic fibrosis: mucoid *Pseudomonas aeruginosa* and *Burkholderia cepacia*. *Microbiological Reviews*, 60(3), 539–74.
- Gram, L., Ravn, L., Rasch, M., Bruhn, J. B., Christensen, A. B., and Givskov, M. 2002. Food spoilage–interactions between food spoilage bacteria. *International Journal of Food Microbiology*, 78(1–2), 79–97.
- Grobe, S., Wingender, J., and Trüper, H. G. 1995. Characterization of mucoid *Pseudomonas aeruginosa* strains isolated from technical water systems. *Journal of Applied Bacteriology*, 79(1), 94–102.
- Guzzo, J., Murgier, M., Filloux, A., and Lazdunski, A. 1990. Cloning of the *Pseudomonas aeruginosa* alkaline protease gene and secretion of the protease into the medium by *Escherichia coli*. *Journal of Bacteriology*, 172(2), 942–8.
- Gyawali, R., and Ibrahim, S. A. 2014. Natural products as antimicrobial agents. *Food Controls*, 46, 412–429.
- Hagerman, A. E., and Butler, L. G. 1981. The specificity of proanthocyanidin-protein interactions. *The Journal of Biological Chemistry*, 256(9), 4494–7.
- Hanouda, T., and Baker, J. R. 2000. Antimicrobial mechanism of action of surfactant lipid preparation in enteric Gram negative bacilli. *Journal of Applied Microbiology*, 89, 397–403.
- Hall-Stoodley, L., Costerton, J. W., and Stoodley, P. 2004. Bacterial biofilms: From the natural environment to infectious diseases. *Nature Reviews Microbiology*, 2(2), 95–108.
- Halliwell, B., and Gutteridge, J. M. C. 2015. *Free Radicals in Biology and Medicine*. Oxford University Press.
- Hancock, R. E. W., and Bell, A. 1988. Antibiotic uptake into Gram-negative bacteria. *Eur. J. Clin. Microbiol. Infect. Dis.*, 7, 713–720.
- Handa, S. S., Khanuja, S. P. S., Longo, G., and Rakesh, D. D. 2008. Extraction technologies for medicinal and aromatic plants. International Centre for Science and High Technology, Trieste, Italy
- Hänsel, W. 1997. *Curcuma xanthorrhiza* Roxb. In *Z Phytotherapy*, 297–306.
- Harborne, J. 1996. *Phytochemical methods. A guide to modern techniques of plants analysis* (2nd ed.). London: Chapman and Hall.
- Harshey, R. M. 1994. Bees aren't the only ones: Swarming in gram-negative bacteria. *Molecular Microbiology*, 13(3), 389–94.

- Harshey, R. M. 2003. Bacterial motility on a surface: Many ways to a common goal. *Annual Review of Microbiology*, 57(1), 249–273.
- Harshey, R. M., and Matsuyama, T. 1994. Dimorphic transition in *Escherichia coli* and *Salmonella typhimurium*: Surface-induced differentiation into hyperflagellate swarmer cells. *Proceedings of the National Academy of Sciences of the United States of America*, 91(18), 8631–5.
- Hartmann, T. 2007. From waste products to ecochemicals: Fifty years research of plant secondary metabolism. *Phytochemical*, 68, 2831–2846.
- Haslam, E. 1996. Natural polyphenols (vegetable tannins) as drugs: Possible modes of action. *Journal of Natural Products*, 59(2), 205–15.
- Hayes, P. R. 1995. *Food Microbiology and Hygiene*. Springer US.
- Hazan, R., Que, Y.-A., Maura, D., and Rahme, L. G. 2012. A method for high throughput determination of viable bacteria cell counts in 96-well plates. *BMC Microbiology*, 12, 1.
- Hector, R. F. 1993. Compounds active against cell walls of medically important fungi. *Clinical Microbiology Reviews*, 6(1), 1–21.
- Heeb, S., Fletcher, M. P., Chhabra, S. R., Diggle, S. P., Williams, P., and Camara, M. 2011. Quinolones: From antibiotics to autoinducers. *FEMS Microbiology Reviews*, 35, 247–274.
- Henrichsen, J. 1972. Bacterial surface translocation: A survey and a classification. *Bacteriological Reviews*, 36(4), 478–503.
- Hentschel, C., Eglau, M. C., and Hahn, E. G. 1996. *Curcuma xanthorrhiza* (Java tumeric) in clinical use. *Fortschritte Der Medizin*, 114(27), 349–50.
- Hentzer, M., Riedel, K., Rasmussen, T., Heydorn, A., Andersen, J., and Al., E. 2002. Inhibition of quorum sensing in *Pseudomonas aeruginosa* biofilm bacteria by a halogenated furanone compound. *Microbiology*, 148, 87–102.
- Hentzer, M., and Givskov, M. 2003. Pharmacological inhibition of quorum sensing for the treatment of chronic bacterial infections. *The Journal of Clinical Investigation*, 112(9), 1300–1307.
- Hentzer, M., Wu, H., Andersen, J. B., Riedel, K., Rasmussen, T. B., Bagge, N., and Givskov, M. 2003. Attenuation of *Pseudomonas aeruginosa* virulence by quorum sensing inhibitors. *The EMBO*, 22(15), 3803–3815.
- Herrera, A. G. 2001. Psychrotrophic Microorganisms. In J. F. T. Spencer and de A. L. R. Spencer (Eds.), *Food Microbiology Protocols* (pp. 3–11). New Jersey: Humana Press.
- Hertiani, T., Palupi, S. I., Sanliferianti, and Nurwindasari, D. H. (2003). In vitro

test on antimicrobial potency against *Staphylococcus aureus*, *Escherichia coli*, *Shigella dysenteriae* and *Candida albicans* of some herbs traditionally used cure infection diseases. *Pharmacol*, 4(2), 89–95.

- Hidayathulla, S., Keshava, C. K., and Chandrashekar, K. R. 2011. Phytochemical evaluation and antibacterial activity of *Pterospermum diversifolium* Blume. *International Journal of Pharmaceutical Sciences*, 3(2), 165–167.
- Hill, D. F., Short, N. J., Perham, R. N., and Petersen, G. B. 1991. DNA sequence of the filamentous bacteriophage Pf1. *Journal of Molecular Biology*, 218(2), 349–364.
- Hirasawa, M., Takada, K., and Otake, S. 2006. Inhibition of acid production in dental plaque bacteria by green tea catechins. *Caries Research*, 40(3), 265–270.
- Hoffman, L. R., D'Argenio, D. A., MacCoss, M. J., Zhang, Z., Jones, R. A., and Al., E. 2005. Aminoglycoside antibiotics induce bacterial biofilm formation. *Nature*, 436, 1171–1175.
- Hoge, R., Pelzer, A., Rosenau, F., and Wilhelm, S. 2010. Weapons of a pathogen: Proteases and their role in virulence of *Pseudomonas aeruginosa*. In A. Mendez-Vilas (Ed.), *Current research, technology and education topics in applied microbiology and microbial biotechnology* (pp. 383–395).
- Hogg, S. 2005. *Essential Microbiology (Microorganisms in Industry)*. West Sussex, UK: John Wiley and Sons.
- Højby, N. 2011. Recent advances in the treatment of *Pseudomonas aeruginosa* infections in cystic fibrosis. *BMC Medicine*, 9(1), 32.
- Hong, Y. Q., and Ghebrehiwet, B. 1992. Effect of *Pseudomonas aeruginosa* elastase and alkaline protease on serum complement and isolated components C1q and C3. *Clinical Immunology and Immunopathology*, 62(2), 133–8.
- Hong, K.-W., Koh, C.-L., Sam, C.-K., Yin, W.-F., and Chan, K.-G. 2012. Quorum quenching revisited—From signal decays to signalling confusion. *Sensors*, 12(4), 4661–4696.
- Hornby, J. M., Jensen, E. C., Lisec, A. D., Tasto, J. J., Jahnke, B., Shoemaker, R., and Nickerson, K. W. 2001. Quorum sensing in the dimorphic fungus *Candida albicans* is mediated by farnesol. *Applied and Environmental Microbiology*, 67(7), 2982–92.
- Hossain, Z. 2013. Bacteria. In Y. Motarjemi, G. Moy, and E. C. D. (Ewen C. D. Todd (Eds.), *Encyclopedia of Food Safety*, 1, 490–491. Academic Press.

- Huber, B., Eberl, L., Feucht, W., and Polster, J. 2003. Influence of polyphenols on bacterial biofilm formation and quorum-sensing. *Z. Naturforsch*, 58, 879–884.
- Hung, C. Y., Seshan, K. R., Yu, J. J., Schaller, R., Xue, J., Basrur, V., and Cole, G. T. 2005. A metalloproteinase of *Coccidioides posadasii* contributes to evasion of host detection. *Infection and Immunity*, 73, 6689–6703.
- Hunter, R. C., Asfour, F., Dingemans, J., Osuna, B. L., Samad, T., Malfroot, A., and Newman, D. K. 2013. Ferrous iron is a significant component of bioavailable iron in cystic fibrosis airways. *MBio*, 4(4), e00557-13-e00557-13.
- Hussain, M. 2016. Food contamination: Major challenges of the future. *Foods*, 5(2), 21.
- Hwang, J. K., Shim, J. S., Baek, N. I., and Pyun, Y. R. 2000. Xanthorrhizol: A potential antibacterial agent from *Curcuma xanthorrhiza* against *Streptococcus mutans*. *Planta Medica*, 66(2), 196–197.
- Imperi, F., Massai, F., Pillai, C. R., Longo, F., Zennaro, E., Rampioni, G., Visca, P., and Leoni, L. 2013. New life for an old drug: the anthelmintic drug niclosamide inhibits *Pseudomonas aeruginosa* quorum sensing. *Antimicrobial agents and chemotherapy*, 57, 2, 996-1005.
- Imperi, F., Leoni, L., and Visca, P. 2014. Antivirulence activity of azithromycin in *Pseudomonas aeruginosa*. *Frontiers in Microbiology*, 5, 178.
- Irie, Y., Starkey, M., Edwards, A. N., Wozniak, D. J., Romeo, T., and Parsek, M. R. 2010. *Pseudomonas aeruginosa* biofilm matrix polysaccharide Psl is regulated transcriptionally by RpoS and post-transcriptionally by RsmA. *Molecular Microbiology*, 78(1), 158–72.
- Irie, Y., Borlee, B. R., O'Connor, J. R., Hill, P. J., Harwood, C. S., Wozniak, D. J., Parsek, M. R. 2012. Self-produced exopolysaccharide is a signal that stimulates biofilm formation in *Pseudomonas aeruginosa*. *Proceedings of the National Academy of Sciences*, 109, 50, 20632-20636.
- Issac, A., Sybiya, V. P., Palani, A., Khadar, S. M., Shunmugiah, K. P. and Arumugam, V. R. 2012. Antibiofilm and quorum sensing inhibitory potential of *Cuminum cyminum* and its secondary metabolite methyl-eugenol against Gram negative bacterial pathogens. *Food Research International*, 45, 85–92.
- Itokawa, H., Hirayama, F., Funakoshi, K., and Takeya, K. 1985. Studies on the antitumor bisabolane sesquiterpenoids isolated from *Curcuma xanthorrhiza*. *Chemical and Pharmaceutical Bulletin*, 33(8), 3488–92.
- Izano, E. A., Amarante, M. A., Kher, W. B., and Kaplan, J. B. 2008. Differential roles of poly-N- acetylglucosamine surface polysaccharide and

- extracellular DNA in *Staphylococcus aureus* and *Staphylococcus epidermidis* biofilms. *Applied and Environmental Microbiology*, 74, 470–476.
- Jackson, K. D., Starkey, M., Kremer, S., Parsek, M. R., and Wozniak, D. J. 2004. Identification of *psl*, a locus encoding a potential exopolysaccharide that is essential for *Pseudomonas aeruginosa* PAO1 biofilm formation. *Journal of Bacteriology*, 186(14), 4466–75.
- Jagannathan, A., Constantinidou, C., and Penn, C. W. 2001. Roles of *rpoN*, *fliA*, and *flgR* in expression of flagella in *Campylobacter jejuni*. *Journal of Bacteriology*, 183(9), 2937–2942.
- Jakobsen, T. H., Bragason, S. K., Phipps, R. K., Christensen, L. D., van Gennip, M., Alhede, M., and Givskov, M. 2012. Food as a source for quorum sensing inhibitors: Iberin from horseradish revealed as a quorum sensing inhibitor of *Pseudomonas aeruginosa*. *Applied and Environmental Microbiology*, 78(7), 2410–2421.
- Jantan, I., Saputri, F. C., Qaisar, M. N., and Buang, F. 2012. Correlation between chemical composition of *Curcuma domestica* and *Curcuma xanthorrhiza* and their antioxidant effect on human low-density lipoprotein oxidation. *Evidence-Based Complementary and Alternative Medicine : eCAM*, 2012, 438356.
- Jarosz, L. M., Ovchinnikova, E. S., Meijler, M. M., and Krom, B. P. 2011. Microbial spy games and host response: Roles of a *Pseudomonas aeruginosa* small molecule in communication with other species. *PLoS Pathogens*, 7, e1002312.
- Jenal, U. 2004. Cyclic di-guanosine-monophosphate comes of age: A novel secondary messenger involved in modulating cell surface structures in bacteria. *Current Opinion in Microbiology*, 7, 185–191.
- Jenke-Kodama, H., Müller, R., and Dittmann, E. 2008. Evolutionary mechanisms underlying secondary metabolite diversity. *Prog Drug Res.*, 121–140.
- Jose, D., Jose, S., Manjusha, K., and Bright Singh, I. 2017. Purification and characterization of highly active LasB protease from *Pseudomonas aeruginosa* MCCB 123. *Indian Journal of Experimental Biology*, 55, 303–310.
- Kahraman, D. Ö. H. 2015. Pyocyanin production in the presence of calcium ion in *Pseudomonas aeruginosa* and recombinant bacteria *Pseudomonas aeruginosa*. *Turkish Journal of Science and Technology*, 10(1), 13–19.
- Kamath, S., Kapatral, V., and Chakrabarty, A. M. 1998. Cellular function of elastase in *Pseudomonas aeruginosa*: Role in the cleavage of nucleoside diphosphate kinase and in alginate synthesis. *Molecular Microbiology*, 30(5), 933–41.

- Kandasamy, S., Khan, W., Evans, F., Critchley, A. T., and Prithiviraj, B. 2012. Tasco®: A product of *Ascophyllum nodosum* enhances immune response of *Caenorhabditis elegans* against *Pseudomonas aeruginosa* infection. *Marine Drugs*, 10(1), 84–105.
- Karatuna, O., and Yagci, A. 2010. Analysis of quorum sensing-dependent virulence factor production and its relationship with antimicrobial susceptibility in *Pseudomonas aeruginosa* respiratory isolates. *Clinical Microbiology and Infection*, 16(12), 1770–1775.
- Karpagam, S., Sudhakar, T., and Lakshmiopathy, M. 2013. Microbicidal response of pyocyanin produced by *Pseudomonas aeruginosa* toward clinical isolates of fungi. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(3), 870–873.
- Kay, E., Humair, B., Dénervaud, V., Riedel, K., Spahr, S., Eberl, L., Haas, D. 2006. Two GacA-dependent small RNAs modulate the quorum-sensing response in *Pseudomonas aeruginosa*. *Journal of Bacteriology*, 188(16), 6026–33.
- Kearns, D. B., and Losick, R. 2003. Swarming motility in undomesticated *Bacillus subtilis*. *Molecular Microbiology*, 49(3), 581–90.
- Kearns, D. B. 2010. A field guide to bacterial swarming motility. *Nature Reviews Microbiology*, 8(9), 634–644.
- Kernacki, K. A., Hobden, J. A., Hazlett, L. D., Fridman, R., and Berk, R. S. 1977. *In vivo* bacterial protease production during *Pseudomonas aeruginosa* corneal infection. *Investigative Ophthalmology and Visual Science*, 36.
- Kessler, E., and Safrin, M. 1983. Comparative effect of ammonium and sodium salts on growth of *Pseudomonas aeruginosa* and on protease (elastase) production. *FEMS Microbiology Letters*, 20(1), 87–90.
- Kessler, E., Safrin, M., Abrams, W R., Rosenbloom, J., and Ohman, D E. 1997. Inhibitors and specificity of *Pseudomonas aeruginosa* LasA. *The Journal of Biological chemistry*, 272(15), 9884-9889.
- Kessler, E., Safrin, M., Gustin, J. K., and Ohman, D. E. 1998. Elastase and the LasA protease of *Pseudomonas aeruginosa* are secreted with their propeptides. *The Journal of Biological Chemistry*, 273(46), 30225–30231.
- Khan, M. S. A., Zahin, M., Hasan, S., Husain, F. M., and Ahmad, I. 2009. Inhibition of quorum sensing regulated bacterial functions by plant essential oils with special reference to clove oil. *Letters in Applied Microbiology*, 49(3), 354–360.
- Khembavi, A. A., Kulharni, A., and Pant, A. A. 1993. Salt-tolerant and thermostable alkaline protease from *Bacillus subtilis*. *Applied Biochemistry of Biotechnology*, 64(38), 83.

- Kim, S. H., Hong, K. O., Chung, W. Y., Hwang, J. K., and Park, K. K. 2004. Abrogation of cisplatin-induced hepatotoxicity in mice by xanthorrhizol is related to its effect on the regulation of gene transcription. *Toxicology and Applied Pharmacology*, 196(3), 346-355.
- Kim, A. J., Kim, Y. O., Shim, J. S., and Hwang, J. K. 2007. Immunostimulating activity of crude polysaccharide extract isolated from *Curcuma xanthorrhiza* Roxb. *Biosci. Biotechnol. Biochem.*, 71, 1428-1438.
- Kim, J. E., Kim, H. E., Hwang, J. K., Lee, H. J., Kwon, H. K., and Kim, B. I. 2008. Antibacterial characteristics of *Curcuma xanthorrhiza* extract on *Streptococcus mutans* biofilm. *Journal of Microbiology*, 46, 228-232.
- Kim, H. S., and Park, H. D. 2013. Ginger extract inhibits biofilm formation by *Pseudomonas aeruginosa* PA14. *PLoS ONE*, 8(9).
- Kim, M. B., Kim, C., Song, Y., and Hwang, J. K. 2014. Antihyperglycemic and anti-inflammatory effects of standardized *Curcuma xanthorrhiza* Roxb. extract and its active compound xanthorrhizol in high-fat diet-induced obese mice. *Evidence-based complementary and alternative medicine: eCAM*,
- King, M., Guragain, M., Sarkisova, S., and Patrauchan, M. 2016. Pyocyanin extraction and quantitative analysis in swarming *Pseudomonas aeruginosa*. *Bio-protocol*, 6(23).
- Kinscherf, T. G., and Willis, D. K. 1999. Swarming by *Pseudomonas syringae* B728a requires *gacS* (*lemA*) and *gacA* but not the acyl-homoserine lactone biosynthetic gene *ahlI*. *Journal of Bacteriology*, 181(13), 4133-6.
- Kiratisin, P., Tucker, K. D., and Passador, L. 2002. LasR, a transcriptional activator of *Pseudomonas aeruginosa* virulence genes, functions as a multimer. *Journal of Bacteriology*, 184, 4912-4919.
- Klausen, M., Aaes-Jørgensen, A., Molin, S., and Tolker-Nielsen, T. 2003. Involvement of bacterial migration in the development of complex multicellular structures in *Pseudomonas aeruginosa* biofilms. *Molecular Microbiology*, 50(1), 61-8.
- Köhler, T., Curty, L. K., Barja, F., van Delden, C., and Pechère, J. C. 2000. Swarming of *Pseudomonas aeruginosa* is dependent on cell-to-cell signaling and requires flagella and pili. *Journal of Bacteriology*, 182(21), 5990-6.
- Köhler, T., Perron, G. G., Buckling, A., van Delden, C. 2010. Quorum sensing inhibition selects for virulence and cooperation in *Pseudomonas aeruginosa*. *PLoS Pathogens*, 6(5), e1000883.
- Kon, Y., Tsukada, H., Hasegawa, T., Igarashi, K., Wada, K., Suzuki, E., Gejyo, F. 1999. The role of *Pseudomonas aeruginosa* elastase as a potent inflammatory factor in a rat air pouch inflammation model. *FEMS*

Immunology and Medical Microbiology, 25(3), 313–21.

- Kong, W., Chen, L., Zhao, J., Shen, T., Surette, M. G., Shen, L., and Duan, K. 2013. Hybrid sensor kinase PA1611 in *Pseudomonas aeruginosa* regulates transitions between acute and chronic infection through direct interaction with RetS. *Molecular Microbiology*, 88(4), 784–797.
- König, B., Jaeger, K. E., Sage, A. E., Vasil, M. L., and König, W. 1996. Role of *Pseudomonas aeruginosa* lipase in inflammatory mediator release from human inflammatory effector cells (platelets, granulocytes, and monocytes). *Infection and Immunity*, 64(8), 3252–8.
- Konola, J. T., Sargent, K. E., and Gow, J. B. 2000. Efficient repair of hydrogen peroxide-induced DNA damage by *Escherichia coli* requires SOS induction of RecA and RuvA proteins. *Mutation Research*, 459(3), 187–94.
- Koshy, P., Wirakarnain, S., Sim, K. S., Saravana, K., Hong, S. L., Lee, G. S., Rahman, S. A. 2009. Antimicrobial activity of some medicinal plants from Malaysia. *American J. Appl. Sci.*, 6, 1613–1617.
- Krishnan, T., Yin, W. F., and Chan, K. G. 2012. Inhibition of quorum sensing-controlled virulence factor production in *Pseudomonas aeruginosa* PAO1 by ayurveda spice clove (*Syzygium aromaticum*) bud extract. *Sensors*, 12(4), 4016–4030.
- Kuchma, S. L., Brothers, K. M., Merritt, J. H., Liberati, N. T., Ausubel, F. M., and O'Toole, G. A. 2007. BifA, a cyclic-di-gmp phosphodiesterase, inversely regulates biofilm formation and swarming motility by *Pseudomonas aeruginosa* PA14. *Journal of Bacteriology*, 189(22), 8165–8178.
- Kudo, C., Yamakoshi, H., Sato, A., Nanjo, H., Ohori, H., Ishioka, C., and Shibata, H. 2011. Synthesis of 86 species of 1,5-diaryl-3-oxo-1,4-pentadienes analogs of curcumin can yield a good lead *in vivo*. *BMC Pharmacology*, 11, 4.
- Kulasakara, H., Lee, V., Brencic, A., Liberati, N., Urbach, J., Miyata, S., and Lory, S. 2006. Analysis of *Pseudomonas aeruginosa* diguanylate cyclases and phosphodiesterases reveals a role for bis-(3'-5')-cyclic-GMP in virulence. *Proceedings of the National Academy of Sciences*, 103(8), 2839–2844.
- Kumar, C. G., and Anand, S. K. 1998. Significance of microbial biofilms in food industry: A review. *International Journal of Food Microbiology*, 42(1–2), 9–27.
- Kuo, M. Y., Yang, M. K., Chen, W. P., and Kuo, T. T. 2000. High-frequency interconversion of turbid and clear plaque strains of bacteriophage f1 and associated host cell death. *Canadian Journal of Microbiology*, 46(9), 841–7.

- Kuo, T. T., Chiang, C. C., Chen, S. Y., Lin, J. H., and Kuo, J. L. 1994. A long lytic cycle in filamentous phage Cf1tv infecting *Xanthomonas campestris* pv. citri. *Archives of Virology*, 135(3–4), 253–64.
- LaBauve, A. E., and Matthew J. Wargo. 2015. Growth and laboratory maintenance of *Pseudomonas aeruginosa*, 1–11.
- LaSarre, B., and Federle, M. J. 2013. Exploiting quorum sensing to confuse bacterial pathogens. *Microbiology and Molecular Biological Review*, 77, 73–111.
- Latifi, A., Foglino, M., Tanaka, K., Williams, P., and Lazdunski, A. 1996. A hierarchical quorum-sensing cascade in *Pseudomonas aeruginosa* links the transcriptional activators LasR and RhlR (VsmR) to expression of the stationary-phase sigma factor RpoS. *Molecular Microbiology*, 21, 1137–1146.
- Lau, G. W., Hassett, D. J., Ran, H., and Kong, F. 2004. The role of pyocyanin in *Pseudomonas aeruginosa* infection. *Trends in Molecular Medicine*, 10(12), 599–606.
- Leadbetter, J. R., and Greenberg, E. P. 2000. Metabolism of acyl-homoserine lactone quorum-sensing signals by *Variovorax paradoxus*. *Journal of Bacteriology*, 182(24), 6921–6.
- Ledgham, F., Soscia, C., Chakrabarty, A., Lazdunski, A., and Foglino, M. 2003. Global regulation in *Pseudomonas aeruginosa*: the regulatory protein AlgR2 (AlgQ) acts as a modulator of quorum sensing. *Research in Microbiology*, 154(3), 207–213.
- Lee, D. G., Urbach, J. M., Wu, G., Liberati, N. T., Feinbaum, R. L., Miyata, S., and Ausubel, F. M. 2006. Genomic analysis reveals that *Pseudomonas aeruginosa* virulence is combinatorial. *Genome Biology*, 7(10), R90.
- Lee, J., Wu, J., Deng, Y., Wang, J., Wang, C., Wang, J., Chang, C., Dong, Y., Williams, P., and Zhang, L. H. 2013. A cell-cell communication signal integrates quorum sensing and stress response. *Nature Chemical Biology*, 9, 339–343
- Lee, J., and Zhang, L. 2014. The hierarchy quorum sensing network in *Pseudomonas aeruginosa*. *Protein and Cell*, 6(1), 26–41.
- Lee, S. K., Hong, C. H., Huh, S. K., Kim, S. S., Oh, O. J., Min, H. Y., and Hwang, J. K. 2002. Suppressive effect of natural sesquiterpenoids on inducible cyclooxygenase (COX-2) and nitric oxide synthase (iNOS) activity in mouse macrophage cells. *Journal of Environmental Pathology, Toxicology and Oncology : Official Organ of the International Society for Environmental Toxicology and Cancer*, 21(2), 141–8.
- Lee, V. T., Matewish, J. M., Kessler, J. L., Hyodo, M., Hayakawa, Y., and Lory, S. 2007. A cyclic-di-GMP receptor required for bacterial exopolysaccharide production. *Molecular Microbiology*, 65(6), 1474–

1484.

- Lesic, B., Lépine, F., Déziel, E., Zhang, J., Zhang, Q., Padfield, K., and Rahme, L. G. 2007. Inhibitors of pathogen intercellular signals as selective anti-infective compounds. *PLoS Pathogens*, 3, e126.
- Levinson, W. 2014. The effects of brief mindfulness intervention on acute pain experience: An examination of individual difference. *Review of Medical Microbiology and Immunology*, 1.
- Lew, K. F., Goh, G. L., Son, R., and Rukayadi, Y. 2015. Effect of Javanese turmeric (*Curcuma xanthorrhiza* Roxb.) extract on natural microflora of oyster mushroom (*Pleurotus sajur-caju*) and its sensory acceptability. *International Food Research Journal*, 22(6), 2446–2451.
- Lewenza, S. 2013. Extracellular DNA-induced antimicrobial peptide resistance mechanisms in *Pseudomonas aeruginosa*. *Frontiers in Microbiology*, 4, 21.
- Lewis, K. 2008. Multidrug tolerance of biofilms and persister cells. *Curr. Top Microbiol. Immunol.*, 322, 107–131.
- Li, J. W. H., and Vederas, J. C. 2009. Drug discovery and natural products: End of an era or an endless frontier? *Science*, 325, 161–165.
- Lim, C. S., Jin, D.-Q., Mok, H., Oh, S. J., Lee, J. U., Hwang, J. K., Han, J.-S. 2005. Antioxidant and antiinflammatory activities of xanthorrhizol in hippocampal neurons and primary cultured microglia. *Journal of Neuroscience Research*, 82(6), 831–838.
- Lim, T. K. 2016. *Curcuma zanthorrhiza*. In *Edible Medicinal and Non-Medicinal Plants*, 384. Cham: Springer International Publishing.
- Lin, S.-C., Teng, C.-W., Lin, C.-C., Lin, Y.-H., and Supriyatna, S. 1996. Protective and therapeutic effect of the Indonesian medicinal herb *Curcuma xanthorrhiza* on β -D-galactosamine-induced liver damage. *Phytotherapy Research*, 10(2), 131–135.
- Linscott, A. J. 2011. Food-borne illnesses. *Clinical Microbiology Newsletter*, 33(6), 41–45.
- Liu, P. V. 1974. Extracellular toxins of *Pseudomonas aeruginosa*. *The Journal of Infectious Diseases*, 130 94-99.
- Lo, Y. L., Shen, L., Chang, C. H., Bhuwan, M., Chiu, C. H., and Chang, H. Y. 2016. Regulation of motility and phenazine pigment production by *flia* is cyclic-di-gmp dependent in *Pseudomonas aeruginosa* PAO1. *PloS ONE*, 11(5).
- Lynch, S. V., and Wiener-Kronisha, J. P. 2008. Novel strategies to combat bacterial virulence. *Current Opinion in Critical Care*, 14, 593–599.

- Ma, L., Jackson, K. D., Landry, R. M., Parsek, M. R., Wozniak, D. J. 2006. Analysis of *Pseudomonas aeruginosa* conditional psl variants reveals roles for the psl polysaccharide in adhesion and maintaining biofilm structure post attachment. *Journal of Bacteriology*, 188(23), 8213-8221.
- Ma, L., Conover, M., Lu, H., Parsek, M. R., Bayles, K., and Wozniak, D. J. 2009. Assembly and development of the *Pseudomonas aeruginosa* biofilm matrix. *PLoS Pathogens*, 5(3), e1000354.
- Mack, D., Nedelmann, M., Krokotsch, A., Schwarzkopf, A., Heesemann, J., and Laufs, R. 1994. Characterization of transposon mutants of biofilm-producing *Staphylococcus epidermidis* impaired in the accumulative phase of biofilm production: genetic identification of a hexosamine-containing polysaccharide intercellular adhesin. *Infection and Immunity*, 62, 3244–3253.
- Malaysia Kini. 2016. Battling Food Waste.
- Mangunwardoyo, W., Deasywaty, and Usia, T. 2012. Antimicrobial and identification of active compound *Curcuma xanthorrhiza* Roxb. *International Journal of Basic and Applied Sciences IJBAS-IJENS*, 12(1), 69–78.
- Mary, H. P., Susheela, G. K., Jayasree, S., Nizzy, A., Rajagopal, B., and Jeeva, S. 2012. Phytochemical characterization and antimicrobial activity of *Curcuma xanthorrhiza* Roxb. *Asian Pacific Journal of Tropical Biomedicine*, 2(2), S637–S640.
- Masuda, T., Isobe, J., Jitoe, A., and Nakatani, N. 1992. Antioxidative curcuminoids from rhizomes of *Curcuma xanthorrhiza*. *Phytochemistry*, 31(10), 3645–3647.
- Matsumoto, M., Minami, T., Sasaki, H., Sobue, S., Hamada, S., and Ooshima, T. 1999. Inhibitory effects of oolong tea extract on caries-inducing properties of mutants *Streptococci*. *Caries Research*, 33(6), 441–445.
- Matsumoto, K. 2004. Role of bacterial proteases in *Pseudomonas* and *Serratia keratitis*. *Biological Chemistry*, 385(11), 1007–16.
- Matsuyama, T., and Nakagawa, Y. 1996. Surface-active exolipids: analysis of absolute chemical structures and biological functions. *Journal of Microbiological Methods*, 25, 165–175.
- Mavrodi, D. V., Bonsall, R. F., Delaney, S. M., Soule, M. J., Phillips, G., and Thomashow, L. S. 2001. Functional analysis of genes for biosynthesis of pyocyanin and phenazine-1-carboxamide from *Pseudomonas aeruginosa* PAO1. *Journal of Bacteriology*, 183, 6454–6465.
- McKnight, S. L., Iglewski, B. H., and Pesci, E. C. 2000. The *Pseudomonas* quinolone signal regulates *rhl* quorum sensing in *Pseudomonas aeruginosa*. *Journal of Bacteriology*, 182, 2702–2708.

- McLean, R. J. C., Whiteley, M., Stickler, D. J., and Fuqua, W. C. 1997. Evidence of autoinducer activity in naturally occurring biofilms. *FEMS Microbiology Letters*, 154, 259–263.
- Menon, V. P., and Sudheer, A. R. 2007. Antioxidant and anti-inflammatory properties of curcumin.
- Merighi, M., Lee, V. T., Hyodo, M., Hayakawa, Y., and Lory, S. 2007. The second messenger bis-(3'-5')-cyclic-GMP and its PilZ domain-containing receptor Alg44 are required for alginate biosynthesis in *Pseudomonas aeruginosa*. *Molecular Microbiology*, 65(4), 876–895.
- Merritt, J. H., Brothers, K. M., Kuchma, S. L., and O'toole, G. A. 2007. SadC reciprocally influences biofilm formation and swarming motility via modulation of exopolysaccharide production and flagellar function. *Journal of Bacteriology*, 189(22), 8154–8164.
- Ministry of Health (MOH). 2014. Majlis anugerah kantin bersih selamat dan sihat peringkat kebangsaan 2014.
- Miyoshi, S., Sonoda, Y., Wakiyama, H., Rahman, M. M., Tomochika, K., Shinoda, S., and Tobe, K. 2002. An exocellular thermolysin-like metalloprotease produced by *Vibrio fluvialis*: purification, characterization, and gene cloning. *Microbial Pathogenesis*, 33, 127–134.
- Mohr, C. D., Rust, L., Albus, A. M., Iglewski, B. H., and Deretic, V. 1990. Expression patterns of genes encoding elastase and controlling mucoidy: co-ordinate regulation of two virulence factors in *Pseudomonas aeruginosa* isolates from cystic fibrosis. *Molecular Microbiology*, 4(12), 2103–10.
- Morihara, K., Tsuzuki, H., Oka, T., Inoue, H., and Ebata, M. 1965. *Pseudomonas aeruginosa* elastase isolation, crystallization, and preliminary characterization. *The Journal of Biological Chemistry*, 240, 3295–304.
- Motarjemi, Y., Moy, G., and Todd, E. (Eds.). 2013. Bacteria- *Pseudomonas*. *Encyclo-paedia of food safety*, 1, 490–491. Academic Press.
- Moure, A., Cruz, J. M., Franco, D., Manuel Domínguez, J. Sineiro, J., Domínguez, H., Núñez, M. J., and Carlos Parajó, J. 2001. Natural antioxidants from residual sources. *Food Chemistry*, 72, 145–171.
- Mulcahy, H., Charron-Mazenod, L., and Lewenza, S. 2008. Extracellular DNA chelates cations and induces antibiotic resistance in *Pseudomonas aeruginosa* biofilms. *PLoS Pathogens*, 4(11), e1000213.
- Mulcahy, H., Charron-Mazenod, L., and Lewenza, S. 2010. *Pseudomonas aeruginosa* produces an extracellular deoxyribonuclease that is required for utilization of DNA as a nutrient source. *Environmental Microbiology*, 12(6), 1621–9.

- Muller, M. 2006. Premature cellular senescence induced by pyocyanin, a redox-active *Pseudomonas aeruginosa* toxin. *Free Radical Biology and Medicine*, 41(11), 1670–1677.
- Murray, T. S., and Kazmierczak, B. I. 2008. *Pseudomonas aeruginosa* exhibits sliding motility in the absence of type IV pili and flagella. *Journal of Bacteriology*, 190, 2700–2708.
- Musfiroh, I., Muchtaridi, M., Muhtadi, A., Diantini, A., Hasanah, A. N., Udin, L. Z., and Ibrahim, S. 2013. Cytotoxicity studies of xanthorrhizol and its mechanism using molecular docking simulation and pharmacophore modelling.
- Mustafa, R. A., Abdul Hamid, A., Mohamed, S., and Bakar, F. A. 2010. Total phenolic compounds, flavonoids, and radical scavenging activity of 21 selected tropical plants. *Journal of Food Science*, 75, C28-35.
- Musthafa, K. S., Balamurugan, K., Pandian, S. K., and Ravi, A. V. 2012. 2,5-Piperazinedione inhibits quorum sensing-dependent factor production in *Pseudomonas aeruginosa* PAO1. *Journal of Basic Microbiology*, 52, 679–686.
- National Center for Biotechnology Information. 2018. Trichloroacetic acid.
- National Centre for Aquatic Animal Health, C. 2001. Purification and characterization of LasB protease from *Pseudomonas aeruginosa*, 138–176.
- Naz, R., Ayub, H., Nawaz, S., Islam, Z. U., Yasmin, T., Bano, A., and Roberts, T. H. 2017. Antimicrobial activity, toxicity and anti-inflammatory potential of methanolic extracts of four ethnomedicinal plant species from Punjab, Pakistan. *BMC Complementary and Alternative Medicine*, 17.
- Nealson, K. H. 1977. Autoinduction of bacterial luciferase. Occurrence, mechanism and significance. *Arch. Microbiol. in Viocrobology*, 112, 73–79.
- Nealson, K., Platt, H., and Hastings, J. W. 1970. Cellular control of the synthesis and activity of the bacterial luminescent system. *Journal of Bacteriology*, 104, 313–322.
- Netotea, S., Bertani, I., Steindler, L., Kerényi, Á., Venturi, V., and Pongor, S. 2009. A simple model for the early events of quorum sensing in *Pseudomonas aeruginosa*: Modeling bacterial swarming as the movement of an “activation zone.” *Biology Direct*, 4.
- Ng, W.-L., and Bassler, B. L. 2009. Bacterial quorum-sensing network architectures. *Annu. Rev. Genet*, 43, 197–222.
- Nohynek, L. J., Alakomi, A., Kähkönen, M. P., Heinonen, M., Helander, I. M., Oksman-Caldentey, K.-M., and Puupponen-Pimiä, R. H. 2006. Berry phenolics: Antimicrobial properties and mechanisms of action against

- severe human pathogens. *Nutrient and Cancer*, 54(1), 18–32.
- Nouwens, A. S., Willcox, M. D. P., Walsh, B. J., and Cordwell, S. J. 2002. Proteomic comparison of membrane and extracellular proteins from invasive (PAO1) and cytotoxic (6206) strains of *Pseudomonas aeruginosa*. *Proteomics*, 2(9), 1325–1346.
- Nouwens, A. S., Beatson, S. A., Whitchurch, C. B., Walsh, B. J., Schweizer, H. P., Mattick, J. S., and Cordwell scordwell, S. J. 2003. Proteome analysis of extracellular proteins regulated by the *las* and *rhl* quorum sensing systems in *Pseudomonas aeruginosa* PAO1. *Microbiology*, 149, 1311–1322.
- Novick, R. P., Projan, S. J., Kornblum, J., Ross, H. F., Ji, G., Kreiswirth, B., and Moghazeh, S. 1995. The *agr* P2 operon: An autocatalytic sensory transduction system in *Staphylococcus aureus*. *Mol Gen Genet*, 248, 446–458.
- Nozawa, Y., and Morita, T. 1986. Molecular mechanisms of antifungal agents associated with membrane ergosterol. Dysfunction of membrane ergosterol and inhibition of ergosterol biosynthesis. In K. Iwata and H. Vanden Bossche (Eds.), *In vitro and in vivo evaluation of antifungal agents*, 111. Amsterdam, The Netherlands: Elsevier Science Publishers.
- Nucleo, E., Steffanoni, L., Fugazza, G., Migliavacca, R., Giacobone, E., and Al., E. 2009. Growth in glucose-based medium and exposure to subinhibitory concentrations of imipenem induce biofilm formation in a multidrug- resistant clinical isolate of *Acinetobacter baumannii*. *BMC Microbiology*, 9(270).
- Nunn, D. N., and Lory, S. 1993. Cleavage, methylation, and localization of the *Pseudomonas aeruginosa* export proteins XcpT, -U, -V, and -W. *Journal of Bacteriology*, 175(14), 4375–82.
- O'Loughlin, C. T., Miller, L. C., Siryaporn, A., Drescher, K., Semmelhack, M. F., and Bassler, B. L. 2013. A quorum-sensing inhibitor blocks *Pseudomonas aeruginosa* virulence and biofilm formation. *Proceedings of the National Academy of Sciences*, 110, 17981–17986.
- O'May, C., and Tufenkji, N. 2011. The swarming motility of *Pseudomonas aeruginosa* is blocked by *Cranberry Proanthocyanidins* and other tannin-containing materials. *Applied And Environmental Microbiology*, 77(9), 3061–3067.
- O'Rear, J., Alberti, L., and Harshey, R. M. 1992. Mutations that impair swarming motility in *Serratia marcescens* 274 include but are not limited to those affecting chemotaxis or flagellar function. *Journal of Bacteriology*, 174(19), 6125–37.
- O'Toole, G. A., and Kolter, R. 1998. Flagellar and twitching motility are necessary for *Pseudomonas aeruginosa* biofilm development. *Molecular*

Microbiology, 30(2), 295–304.

- O'Boyle, G. A. 2008. How *Pseudomonas aeruginosa* regulates surface behaviors. *Microbe*, 3(2), 65–71.
- Ochsner, U. A., Koch, A. K., Fiechter, A., and Reiser, J. 1994. Isolation and characterization of a regulatory gene affecting rhamnolipid biosurfactant synthesis in *Pseudomonas aeruginosa*. *Journal of Bacteriology*, 176(7), 2044–2054.
- Ochsner, U. A., and Reiser, J. 1995. Autoinducer-mediated regulation of rhamnolipid biosurfactant synthesis in *Pseudomonas aeruginosa*. *Proceedings of the National Academy of Sciences*, 92, 6424–6428.
- Okwu, D. E. 2001. Evaluation of the chemical composition of indigenous. Spices and flavouring agents. *Global Journal of Pure Applied Science*, 7, 455–459.
- Oliver S. P., Jayarao B. M. and Almeida R. A. 2005. Foodborne pathogens in milk and the dairy farm environment: food safety and public health implications. *Foodborne Pathog. Dis.*, 2: 115–129
- Olson, J. C., and Ohman, D. E. 1992. Efficient production and processing of elastase and LasA by *Pseudomonas aeruginosa* require zinc and calcium ions. *Journal of Bacteriology*, 174(12), 4140–7.
- Omojasola, P. F., and Awe, S. 2004. The antibacterial of the leaf extract of *Anacardium occidentale* and *Gossypium hirsutum* against some selected microorganism. *Bioscience Research Communication*, 16(1), 25–28.
- Oon, S. F., Nallappan, M., Tee, T. T., Shohaimi, S., Kassim, N. K., Shazrul, M., and Cheah, Y. H. 2015. Xanthorrhizol: A review of its pharmacological activities and anticancer properties. *Cancer Cell Int*, 15.
- Overhage, J., Lewenza, S., Marr, A. K., and Hancock, R. E. W. 2007. Identification of genes involved in swarming motility using a *Pseudomonas aeruginosa* PAO1 mini-Tn5-lux mutant Library. *Journal of Bacteriology*, 189(5), 2164–2169.
- Overhage, J., Bains, M., Brazas, M. D., and Hancock, R. E. W. 2008. Swarming of *Pseudomonas aeruginosa* is a complex adaptation leading to increased production of virulence factors and antibiotic resistance. *Journal of Bacteriology*, 190(8), 2671–2679.
- Ozaki, Y. 1990. Anti-inflammatory effect of *Curcuma xanthorrhiza*. Roxb and its active principles. *Chemical and Pharmaceutical Bulletin*, 38, 1045–1048.
- Parekh, J., and Chanda, S. V. 2007. In vitro antimicrobial activity and phytochemical analysis of some indian medicinal plants. *Turk J Biol.*, 31, 53–58.

- Parks, L. W., and Casey, W. M. 1996. Fungal sterols. In R. Prasad and M. Ghannoum (Eds.), *Lipids of pathogenic fungi*, 63–82. Boca Raton, Fla: CRC Press, Inc.
- Park, J. H., Park, K. K., Kim, M. J., Hwang, J. K., Park, S. K., and Chung, W. Y. 2008. Cancer chemoprotective effects of *Curcuma xanthorrhiza*. *Phytotherapy Research*, 22(5), 695–698.
- Parkins, M. D., Ceri, H., and Storey, D. G. 2001. *Pseudomonas aeruginosa* GacA, a factor in multihost virulence, is also essential for biofilm formation. *Molecular Microbiology*, 40(5), 1215–26.
- Paul, R., Weiser, S., Amiot, N. C., Chan, C., Schirmer, T., Giese, B., and Jenal, U. 2004. Cell cycle-dependent dynamic localization of a bacterial response regulator with a novel di-guanylate cyclase output domain. *Genes and Development*, 18(6), 715–727.
- Pearson, J. P., Feldman, M., Iglewski, B. H., and Prince, A. 2000. *Pseudomonas aeruginosa* cell-to-cell signalling is required for virulence in a model of acute pulmonary infection. *Infection and Immunity*, 68, 4331–4334.
- Pearson, J. P., Pesci, E. C., and Iglewski, B. H. 1997. Roles of *Pseudomonas aeruginosa las* and *rhl* quorum-sensing systems in control of elastase and rhamnolipid biosynthesis genes. *Journal of Bacteriology*, 179, 5756–5767.
- Pereira, C. S., Thompson, J. A., and Xavier, K. B. 2013. AI-2-mediated signalling in bacteria. *FEMS Microbiology Reviews*, 37(2), 156–181.
- Pesci, E. C., Milbank, J. B., Pearson, J. P., McKnight, S. L., Kende, A. S., Greenberg, E. P., and Iglewski, B. H. 1999. Quinolone signaling in the cell-to-cell communication system of *Pseudomonas aeruginosa*. *Proceedings of the National Academy of Sciences*, 96, 11229–11234.
- Peters, J. E., and Galloway, D. R. 1990. Purification and characterization of an active fragment of the LasA protein from *Pseudomonas aeruginosa*: Enhancement of elastase activity. *Journal of Bacteriology*, 172(5), 2236–40.
- Pham-Huy, L. A., He, H., and Pham-Huy, C. 2008. Free Radicals, Antioxidants in Disease and Health. *International Journal of Biomedical Science*, 4(2), 89–96.
- Phillips, R. M., Six, D. A., Dennis, E. A., and Ghosh, P. 2003. *In vivo* phospholipase activity of the *Pseudomonas aeruginosa* cytotoxin ExoU and protection of mammalian cells with phospholipase A₂ inhibitors. *Journal of Biological Chemistry*, 278(42), 41326–41332.
- Pierson, L. S., and Pierson, E. A. 2010. Metabolism and function of phenazines in bacteria: Impacts on the behavior of bacteria in the environment and biotechnological processes. *Applied Microbiology and*

Biotechnology, 86(6), 1659–1670.

- Potempa, J., and Pike, R. N. 2009. Corruption of innate immunity by bacterial proteases. *Journal of Innate Immunity*, 1(2), 70–87.
- Pratt, L. A., and Kolter, R. 1998. Genetic analysis of *Escherichia coli* biofilm formation: Roles of flagella, motility, chemotaxis and type I pili. *Molecular Microbiology*, 30(2), 285–293.
- Prijaya, P., Philip, R., and Singh, I. S. B. 2013. Cloning and overexpression of Phz genes encoding phenazine biosynthetic pathway for the enhanced production of pyocyanin in *Pseudomonas aeruginosa* MCCB117. *Shodhganga*. Cochin University of Science and Technology.
- Pruden, A. R., Pei, T., Storteboom, H., and Carlson, K. H. 2006. Antibiotic resistance genes as emerging contaminants: Studies in Northern Colorado. *Environmental Science and Technology*, 40, 7445–7450.
- Qin, Z., Ou, Y., Yang, L., Zhu, Y., Tolker-Nielsen, T., Molin, S., and Qu, D. 2007. Role of autolysin-mediated DNA release in biofilm formation of *Staphylococcus epidermidis*. *Microbiology*, 153, 2083–2092.
- Rada, B., and Leto, T. 2009. Redox warfare between airway epithelial cells and *Pseudomonas*: Dual oxidase versus pyocyanin. *Immunol. Res.*, 43, 198–209.
- Rada, B., and Leto, T. L. 2013. Pyocyanin effects on respiratory epithelium: Relevance in *Pseudomonas aeruginosa* airway infections. *Trends in Microbiology*, 21(2), 73–81.
- Rahim, R., Ochsner, U. A., Olvera, C., Graninger, M., Messner, P., Lam, J. S., and Soberón-Chávez, G. 2001. Cloning and functional characterization of the *Pseudomonas aeruginosa* *rhIC* gene that encodes Rhamnosyltransferase 2, an enzyme responsible for di-rhamnolipid biosynthesis. *Molecular Microbiology*, 40(3), 708–18.
- Rakoff-Nahoum, S. 2006. Why cancer and inflammation? *The Yale Journal of Biology and Medicine*, 3–4, 123–30.
- Rasamiravaka, T., Labtani, Q., Duez, P., and El Jaziri, M. 2015. The formation of biofilms by *Pseudomonas aeruginosa*: A review of the natural and synthetic compounds interfering with control mechanisms. *BioMed Research International*, 1–17.
- Rasch, M., Buch, C., Austin, B., Larsen, J. L., Johansen, C., Riedel, K., Ekmann, K. S. 2004. An inhibitor of bacterial quorum sensing reduces mortalities caused by Vibriosis in rainbow trout (*Oncorhynchus mykiss*, Walbaum). *Systematic and Applied Microbiology*, 27(3), 350–359.
- Rashid, M. H., and Kornberg, A. 2000. Inorganic polyphosphate is needed for swimming, swarming, and twitching motilities of *Pseudomonas aeruginosa*. *Proceedings of the National Academy of Sciences*, 97(9),

4885–4890.

- Rasmussen, T. B., Bjarnsholt, T., Skindersoe, M. E., Hentzer, M., Kristoffersen, P., Köte, M., and Givskov, M. 2005. Screening for quorum-sensing inhibitors (QSI) by use of a novel genetic system, the QSI selector. *Journal of Bacteriology*, 187(5), 1799–814.
- Rather, P. N. 2005. Swarmer cell differentiation in *Proteus mirabilis*. *Environmental Microbiology*, 7(8), 1065–1073.
- Rawat, S. 2015. Food Spoilage : Microorganisms and their prevention, 5(4), 47–56.
- Ray, B. 2004. *Fundamental food microbiology*. CRC Press.
- Records, A. R., and Gross, D. C. 2010. Sensor kinases RetS and LadS regulate *Pseudomonas syringae* type VI secretion and virulence factors. *Journal of Bacteriology*, 192(14), 3584–96.
- Reimann, C., Beyeler, M., Latifi, A., Winteler, H., Foglino, M., Lazdunski, A., and Haas, D. 1997. The global activator GacA of *Pseudomonas aeruginosa* PAO positively controls the production of the autoinducer N-butyryl-homoserine lactone and the formation of the virulence factors pyocyanin, cyanide, and lipase. *Molecular Microbiology*, 24(2), 309–19.
- Reszka, K. J., O'Malley, Y., McCormick, M. L., Denning, G. M., and Britigan, B. E. 2004. Oxidation of pyocyanin, a cytotoxic product from *Pseudomonas aeruginosa*, by microperoxidase 11 and hydrogen peroxide. *Free Radical Biology and Medicine*, 36(11), 1448–1459.
- Rice, S. A., Tan, C. H., Mikkelsen, P. J., Kung, V., Woo, J., Tay, M., and Kjelleberg, S. 2009. The biofilm life cycle and virulence of *Pseudomonas aeruginosa* are dependent on a filamentous prophage. *The ISME Journal*, 3(3), 271–282.
- Richter, R. L., and Vedomuthu, E. R. 2001. Milk and milk products. In F. P. Downes and K. Ito (Eds.), *Compendium of Methods for the Microbiological Examination of Foods*,. 483–495. Washington DC: American Public Health Association.
- Rishton, G. M. 2008. Natural products as a robust source of new drugs and drug leads: Past successes and present day issues. *Am J Cardiol.*, 101, 43D–49D.
- Römling, U., Gomelsky, M., and Galperin, M. Y. 2005. C-di-GMP: The dawning of a novel bacterial signalling system. *Molecular Microbiology*, 57(3), 629–639.
- Ross, P., Weinhouse, H., Aloni, Y., Michaeli, D., Weinberger-Ohana, P., Mayer, R., and Benziman, M. 1987. Regulation of cellulose synthesis in *Acetobacter xylinum* by cyclic diguanylic acid. *Nature*, 325(6101), 279–81.

- Rukayadi, Y., Yong, D., and Hwang, J. K. 2006. *In vitro* anticandidal activity of xanthorrhizol isolated from *Curcuma xanthorrhiza* Roxb. *Journal of Antimicrobial Chemotherapy*, 57, 1231–1234.
- Rukayadi, Y. and Hwang, J. 2007. *In vitro* antimycotic activity of xanthorrhizol isolated from *Curcuma xanthorrhiza* Roxb. against opportunistic filamentous fungi. *Phytotherapy Research*, 21(5), 434-438.
- Rutherford, S. T., and Bassler, B. L. 2016. Bacterial quorum sensing : Its role in virulence and possibilities for its control, 1–26.
- Ryan, R. P., Fouhy, Y., Lucey, J. F., Crossman, L. C., Spiro, S., He, Y.-W., and Dow, J. M. 2006. Cell-cell signaling in *Xanthomonas campestris* involves an HD-GYP domain protein that functions in cyclic di-GMP turnover. *Proceedings of the National Academy of Sciences*, 103(17), 6712–6717.
- Ryder, C., Byrd, M., and Wozniak, D. J. 2007. Role of polysaccharides in *Pseudomonas aeruginosa* biofilm development. *Current Opinion in Microbiology*, 10(6), 644–8.
- Saifudin, A., Rahayu, H. V. and Teruna, H. Y. 2011. “Standarisasi bahan obat alam”. Graha Ilmu. Yogyakarta. 16.
- Saleem, M., Afaq, F., Adhami, V. M., Mukhtar, H. 2004. Lupeol modulates NF- κ B and PI3K/Akt pathways and inhibits skin cancer in CD-1 mice. *Oncogene*, 23(30), 5203-5214.
- Saleem, A. J. 2012. Relationship study between the alkaline protease production and the growth phases of *Pseudomonas aeruginosa* isolated from patients, 2, 354–357.
- Samanta, S., Thavasi, R., and Jayalakshmi, S. 2008. Phenazine pigments from *Pseudomonas aeruginosa* and their application as antibacterial agent and food colourants. *Research Journal of Microbiology*, 3(3), 122–128.
- Samanta, A., Pal, P., Mandal, A., Sinha, C., Lalee, A., Das, M., and Mitra, D. 2012. Estimation of biosurfactant activity of an alkaline protease producing bacteria isolated from municipal solid waste. *Central European Journal of Experimental Biology*, 1(1), 26–35.
- Sarkisova, S., Patrauchan, M. a, Berglund, D., Nivens, D. E., and Franklin, M. J. 2005. Calcium-induced virulence factors associated with the extracellular matrix of mucoid *Pseudomonas aeruginosa* biofilms. *Society*, 187(13), 4327–4337.
- Sasaki, H., Matsumoto, M., Tanaka, T., Maeda, M., Nakai, M., Hamada, S., and Ooshima, T. 2004. Antibacterial activity of polyphenol components in oolong tea extract against *Streptococcus mutans*. *Caries Research*, 38(1), 2–8.

- Sauer, K., and Camper, A. K. (2001). Characterization of phenotypic changes in *Pseudomonas putida* in response to surface-associated growth. *Journal of Bacteriology*, 183(22), 6579–6589.
- Sauer, K., Camper, A. K., Ehrlich, G. D., Costerton, J. W., and Davies, D. G. 2002. *Pseudomonas aeruginosa* displays multiple phenotypes during development as a biofilm. *Journal of Bacteriology*, 184(4), 1140–54.
- Saulnier, J. M., Curtil, F. M., Duclos, M. C., and Wallach, J. M. 1989. Elastolytic activity of *Pseudomonas aeruginosa* elastase. *Biochimica et Biophysica Acta*, 995(3), 285–90.
- Scallan, E., Hoekstra, R. M., Angulo, F. J., Tauxe, R. V., Widdowson, M. A., Roy, S. L., Griffin, P. M. 2011. Foodborne illness acquired in the United States—major pathogens. *Emerging Infectious Diseases*, 17(1), 7–15.
- Scharff R. L. 2012. Economic burden from health losses due to foodborne illness in the United States. *J. Food Prot.*, 75(1): 123–131.
- Schmidt, B., Ribnicky, D. M., Poulev, A., Logendra, S., Cefalu, W. T., and Raskin, I. 2008. A natural history of botanical therapeutics. *Metabolism*, 57, S3-9.
- Schuster, M., Lostroh, C. P., Ogi, T., and Greenberg, E. P. 2003. Identification, timing, and signal specificity of *Pseudomonas aeruginosa* quorum-controlled genes: a transcriptome analysis. *Journal of Bacteriology*, 185, 2066–2079.
- Schuster, M., Urbanowski, M. L., and Greenberg, E. P. 2004. Promoter specificity in *Pseudomonas aeruginosa* quorum sensing revealed by DNA binding of purified LasR. *Proceedings of the National Academy of Sciences*, 101, 15833–15839.
- Seed, P. C., Passador, L., and Iglewski, B. H. 1995. Activation of the *Pseudomonas aeruginosa lasI* gene by LasR and the *Pseudomonas* autoinducer PAI: An autoinduction regulatory hierarchy. *Journal of Bacteriology*, 177, 654–659.
- Selim, S., Hassan, S., Al Soumaa, K., and EL Anzy, S. 2013. Prevalence, antibiotic resistance and in vitro activity of yogurt against some Gram negative pathogenic bacteria isolated from Arar Hospital, KSA. *Life Science Journal*, 10, 1450–1456.
- Serrano, J., Puupponen-Pimiä, R., Dauer, A., Aura, A.-M., and Saura-Calixto, F. 2009. Tannins: Current knowledge of food sources, intake, bioavailability and biological effects. *Molecular Nutrition and Food Research*, 53(S2), S310–S329.
- Shanks, R. M. Q., Caiazza, N. C., Hinsa, S. M., Toutain, C. M., and O'Toole, G. A. 2006. *Saccharomyces cerevisiae*-based molecular tool kit for manipulation of genes from gram-negative bacteria. *Applied and Environmental Microbiology*, 72(7), 5027–36.

- Shao, Z.-M., Shen, Z.-Z., Liu, C.-H., Sartippour, M. R., Go, V. L., Heber, D., and Nguyen, M. 2002. Curcumin exerts multiple suppressive effects on human breast carcinoma cells. *International Journal of Cancer*, 98(2), 234–40. Retrieved from
- Shieh, G. J., Charng, Y. C., Yang, B. C., Jenn-Tu, Bau, H. J., and Kuo, T. T. 1991. Identification and nucleotide sequence analysis of an open reading frame involved in high-frequency conversion of turbid to clear plaque mutants of filamentous phage Cf1t. *Virology*, 185(1), 316–22.
- Shigematsu, T., Fukushima, J., Oyama, M., Tsuda, M., Kawamoto, S., and Okuda, K. 2001. Iron-mediated regulation of alkaline proteinase production in *Pseudomonas aeruginosa*. *Microbiology and Immunology*, 45(8), 579–90.
- Siagian, M. H. 2006. Temulawak sebagai tanaman obat dan budidayanya secara intensif. *Balitbang Botani*. Bogor: Puslitbang Biologi LIPI.
- Sigma Aldrich. 2018. Mechanism of Action - Antibiotics | Sigma-Aldrich.
- Singh, P. K., Schaefer, A. L., Parsek, M. R., Moninger, T. O., Welsh, M. J., and Greenberg, E. P. 2000. Quorum-sensing signals indicate that cystic fibrosis lungs are infected with bacterial biofilms. *Nature*, 407(6805), 762–764.
- Sixty-third World Health Assembly. Document WHA63/A63/11. Available at http://apps.who.int/gb/ebwha/pdf_files/WHA63/A63_11-en.pdf (Accessed 12th April 2013).
- Skandamis, P. N., and Nychas, G. J. E. 2012. Quorum sensing in the context of food microbiology. *Applied and Environmental Microbiology*, 78(16), 5473–5482.
- Skindersoe, M. E., Alhede, M., Phipps, R., Yang, L., Jensen, P. O., Rasmussen, T. B., and Givskov, M. 2008. Effects of antibiotics on quorum sensing in *Pseudomonas aeruginosa*. *Antimicrobial Agents and Chemotherapy*, 52(10), 3648–3663.
- Solubility and Miscibility. 2015.
- Song, Z., Kong, K. F., Wu, H., Maricic, N., Ramalingam, B., Priestap, H., and Mathee, K. 2010. Panax ginseng has anti-infective activity against opportunistic pathogen *Pseudomonas aeruginosa* by inhibiting quorum sensing, a bacterial communication process critical for establishing infection. *Phytomedicine*, 17(13), 1040–1046.
- Soniya, M., Kuberan, T., Anitha, S., and Sankareswari, P. 2013. In vitro antibacterial activity of plant extracts against Gram positive and Gram negative pathogenic bacteria. *International Journal of Microbiology and Immunology Research*, 2(1), 1–5.
- Soon, J. M., Singh, H., and Baines, R. 2011. Foodborne diseases in Malaysia: A review. *Food Control*, 22, 823–830.

- Sriramulu, D. D., Lünsdorf, H., Lam, J. S., and Römling, U. 2005. Microcolony formation: A novel biofilm model of *Pseudomonas aeruginosa* for the cystic fibrosis lung. *Journal of Medical Microbiology*, 54(7), 667–676.
- Starnbach, M. N., and Lory, S. 1992. The *fliA* (*rpoF*) gene of *Pseudomonas aeruginosa* encodes an alternative sigma factor required for flagellin synthesis. *Molecular Microbiology*, 6(4), 459–69.
- Stover, C. K., Pham, X. Q., Erwin, A. L., Mizoguchi, S. D., Warrener, P., Hickey, M. J., and Olson, M. V. 2000. Complete genome sequence of *Pseudomonas aeruginosa* PAO1, an opportunistic pathogen. *Nature*, 406(6799), 959–964.
- Sundin, G. W., Shankar, S., Chugani, S. A., Chopade, B. A., Kavanaugh-Black, A., and Chakrabarty, A. M. 1996. Nucleoside diphosphate kinase from *Pseudomonas aeruginosa*: Characterization of the gene and its role in cellular growth and exopolysaccharide alginate synthesis. *Molecular Microbiology*, 20(5), 965–79.
- Surh, Y. J., Chun, K. S., Cha, H. H., Han, S. S., Keum, Y. S., Park, K. K., and Lee, S. S. 2001. Molecular mechanisms underlying chemopreventive activities of anti-inflammatory phytochemicals: down-regulation of COX-2 and iNOS through suppression of NF-kappa B activation. *Mutation Research*, 480–481, 243–68.
- Sutherland, I. W. 2001. Biofilm exopolysaccharides: A strong and sticky framework. *Microbiology*, 147(1), 3–9.
- Swift, S., Allan Downie, J., Whitehead, N. A., Barnard, A. M. L., Salmond, G. P. C., and Williams, P. 2001. Quorum sensing as a population-density-dependent determinant of bacterial physiology. *Advances in Microbial Physiology*, 45.
- Packiavathy, I. A. S. V., Agilandeeswari, P., Musthafa, K. S., Karutha Pandian, S., and Veera Ravi, A. 2012. Antibiofilm and quorum sensing inhibitory potential of *Cuminum cyminum* and its secondary metabolite methyl eugenol against Gram negative bacterial pathogens. *Food Research International*, 45(1), 85–92.
- Sylvester, W. S., Son, R., Lew, K. F., and Rukayadi, Y. 2015. Antibacterial activity of Java turmeric (*Curcuma xanthorrhiza* Roxb.) extract against *Klebsiella pneumoniae* isolated from several vegetables. *International Food Research Journal*, 22(5), 1770–1776.
- Szabó, M. Á., Varga, G. Z., Hohmann, J., Schelz, Z., Szegedi, E., Amaral, L., and Molnár, J. 2010. Inhibition of quorum-sensing signals by essential oils. *Phytotherapy Research*, 24(5), 782–786.
- Tamura, Y., Suzuki, S., Kijima, M., Takahashi, T., and Nakamura, M. 1992. Effect of proteolytic enzyme on experimental infection of mice with *Pseudomonas aeruginosa*. *The Journal of Veterinary Medical Science*, 54(3), 597–9.

- Tamura, M., Ajayi, T., Allmond, L., R., Moriyama, K., Wiener-Kronish, J. P., and Sawa, T. 2004. Lysophospholipase A activity of *Pseudomonas aeruginosa* type III secretory toxin ExoU. *Biochemical and Biophysical Research Communications*, 316(2), 323-331.
- Tarigan, J., Zuhra, C. F., and Sihotang, H. 2008. Skrining fitokimia tumbuhan yang digunakan oleh pedagang jamu gendong untuk merawat kulit wajah di Kecamatan Medan Baru. *J. Biologi Sumatera*, 1(3), 1–6.
- Taylor, P. K., Yeung, A. T. Y., and Hancock, R. E. W. 2014. Antibiotic resistance in *Pseudomonas aeruginosa* biofilms: Towards the development of novel anti-biofilm therapies. *Journal of Biotechnology*, 191, 121–130.
- Teisl, M. F., and Roe, B. E. 2010. Consumer willingness-to-pay to reduce the probability of retail foodborne pathogen contamination. *Food Policy*, 35, 521–530.
- Tettmann, B., Niewerth, C., Kirschhöfer, F., Neidig, A., Dötsch, A., Brenner-Weiss, G., Fetzner, S. 2016. Enzyme-mediated quenching of the *Pseudomonas* quinolone signal (PQS) promotes biofilm formation of *Pseudomonas aeruginosa* by increasing iron availability, 7.
- Thayers, M. M., Flahertyq, K. M., and Mckayqv, D. B. 1991. Three-dimensional structure of the elastase of *Pseudomonas aeruginosa* at 1.5-Å Resolution*. *The Journal of Biological Chemistry*, 266(5), 2864–2871.
- Tielen, P., Rosenau, F., Wilhelm, S., Jaeger, K.-E., Flemming, H.-C., Wingender, J., and Petra Tielen, C. 2010. Extracellular enzymes affect biofilm formation of mucoid *Pseudomonas aeruginosa*. *Microbiology*, 156, 2239–2252.
- Tomassen, J., Braun, P., and Bitter, W. 2000. Activation of *Pseudomonas aeruginosa* elastase in *Pseudomonas putida* by triggering dissociation of the propeptide–enzyme complex. *Microbiology*, 146(10), 2565–2572.
- Toutain, C. M., Caizza, N. C., Zegans, M. E., and O'Toole, G. A. 2007. Roles for flagellar stators in biofilm formation by *Pseudomonas aeruginosa*. *Research in Microbiology*, 158(5), 471–477.
- Tremblay, J., Richardson, A.-P., Lépine, F., and Déziel, E. 2007. Self-produced extracellular stimuli modulate the *Pseudomonas aeruginosa* swarming motility behaviour. *Environmental Microbiology*, 9(10), 2622–2630.
- Ugurlu, A., Yagci, A. K., Ulusoy, S., Aksu, B., and Bosgelmez-Tinaz, G. 2016. Phenolic compounds affect production of pyocyanin, swarming motility and biofilm formation of *Pseudomonas aeruginosa*.
- Valko, M., Morris, H., and Cronin, M. T. D. 2005. Metals, toxicity and oxidative stress. *Current Medicinal Chemistry*, 12(10), 1161–208.

- Valko, M., Rhodes, C. J., Moncol, J., Izakovic, M., and Mazur, M. 2006. Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chemico-Biological Interactions*, 160(1), 1–40.
- van Delden, C., and Iglewski, B. H. 1998. Cell-to-cell signaling and *Pseudomonas aeruginosa* infections. *Emerging Infectious Diseases*, 4(4), 551–560.
- van Delden, C., Pesci, E. C., Pearson, J. P., and Iglewski, B. H. 1998. Starvation selection restores elastase and rhamnolipid production in a *Pseudomonas aeruginosa* quorum-sensing mutant. *Infection and Immunity*, 66(9), 4499–502.
- van Delden, C. 2004. Virulence Factors in *Pseudomonas aeruginosa*. In *Pseudomonas* (pp. 3–45). Boston, MA: Springer US.
- van Delden, C., Köhler, T., Brunner-Ferber, F., François, B., Carlet, J., and Pechère, J.-C. 2012. Azithromycin to prevent *Pseudomonas aeruginosa* ventilator-associated pneumonia by inhibition of quorum sensing: a randomized controlled trial. *Intensive Care Medicine*, 38(7), 1118–1125.
- van 't Wout, E. F. A., Van Schadewijk, A., Van Boxtel, R., Dalton, L. E., Clarke, H. J., Tommassen, J., and Hiemstra, P. S. 2015. Virulence factors of *Pseudomonas aeruginosa* induce both the unfolded protein and integrated stress responses in airway epithelial cells. *PLoS Pathogens*, 23.
- Varposhti, M., Abdi Ali, A., Mohammadi, P., and Saboora, A. 2013. Effects of extracts and an essential oil from some medicinal plants against biofilm formation of *Pseudomonas aeruginosa*. *Journal of Medical Microbiology and Infectious Diseases*, 1(1), 36–40.
- Vasavi, H., Arun, A., and Rekha, P. 2016. Anti-quorum sensing activity of flavonoid- rich fraction from *Centella asiatica* L. against *Pseudomonas aeruginosa* PAO1 *Journal of Microbiology Immunology and Infection*, 49, 8–15.
- Vasseur, P., Vallet-Gely, I., Soscia, C., Genin, S., and Filloux, A. (2005). The *pel* genes of the *Pseudomonas aeruginosa* PAK strain are involved at early and late stages of biofilm formation. *Microbiology*, 151(3), 985–997.
- Vattem, D. A., Mihalik, K., Crixell, S. H., and McLean, R. J. C. 2007. Dietary phytochemicals as quorum sensing inhibitors. *Fitoterapia*, 78(4), 302–310.
- Veld*, J. H. J. H. in't. 1996. Microbial and biochemical spoilage of foods : An overview. *International Journal of Food Microbiology*, 33(1), 1–18.
- Ventre, I., Ledgham, F., Prima, V., Lazdunski, A. Foglino, M. 2003. Dimerization of the quorum sensing regulator RhIR: Development of a

- method using EGFP fluorescence anisotropy. *Journal of Bacteriology*, 48,187-198.
- Ventre, I., Goodman, A. L., Vallet-Gely, I., Vasseur, P., Soscia, C., Molin, S., Bleves, S., Lazdunski, A., Lory, S., Filloux, A. 2006. Multiple sensors control reciprocal expression of *Pseudomonas aeruginosa* regulatory RNA and virulence genes. *Proceedings of the National Academy of Science of the United States of America*, 103(1), 171-176.
- Venturi, V. 2006. Regulation of quorum sensing in *Pseudomonas*. *FEMS Microbiology Reviews*, 30(2), 274–291.
- Verstraeten, N., Braeken, K., Debkumari, B., Fauvart, M., Fransaeer, J., Vermant, J., and Michiels, J. 2008. Living on a surface: Swarming and biofilm formation. *Trends in Microbiology*, 16(10), 496–506.
- Vining, L. C. 1990. Functions of secondary metabolites. *Annual Review of Microbiology*, 44(1), 395–427.
- Wade, W. D., Calfee, M. W., Rocha, E. R., Ling, E. A., Engstrom, E., Coleman, J. P., and Pesci, E. C. 2005. Regulation of *Pseudomonas* quinolone signal synthesis in *Pseudomonas aeruginosa*. *Journal of Bacteriology*, 187, 4372–4380.
- Wang, Q., Frye, J. G., McClelland, M., and Harshley, R. M. 2004. Gene expression patterns during swarming in *Salmonella typhimurium*: Genes specific to surface growth and putative new motility and pathogenicity. *Molecular Microbiology*, 52, 169–187.
- Wang, Y., Wilks, J. C., Danhorn, T., Ramos, I., Croal, L., and Newman, D. K. 2011. Phenazine-1-carboxylic acid promotes bacterial biofilm development via ferrous iron acquisition. *Journal of Bacteriology*, 193(14), 3606–3617.
- Wang, R., Starkey, M., Hazan, R., and Rahme, L. G. 2012. Honey's ability to counter bacterial infections arises from both bactericidal compounds and QS inhibition.
- Waters, C. M., and Bassler, B. L. 2005. Quorum sensing: cell-to-cell communication in bacteria. *Annual Review of Cell and Developmental Biology*, 21(1), 319–346.
- Wei, Q., and Ma, L. Z. 2013. Biofilm matrix and its regulation in *Pseudomonas aeruginosa*. *Int. J. Mol. Sci*, 14, 20983–21005.
- Weinstein, R. A., Gaynes, R., and Edwards, J. R. 2005. Overview of Nosocomial Infections Caused by Gram-Negative Bacilli. *Clinical Infectious Diseases*, 41(6), 848–854.
- Werner, E., Roe, F., Bugnicourt, A., Franklin, M. J., Heydorn, A., Molin, S., and Stewart, P. S. 2004. Stratified growth in *Pseudomonas aeruginosa* biofilms. *Applied and Environmental Microbiology*, 70(10), 6188–6196.

- Whitchurch, C. B., Tolker-Nielsen, T., Ragas, P. C., and Mattick, J. S. 2002. Extracellular DNA required for bacterial biofilm formation. *Science*, 295(5559), 1487.
- Whitcombe, T. 2000. Re: Why is ethanol the only relatively safe alcohol for human consumption?
- Whiteley, M., Bangera, M. G., Bumgarner, R. E., Parsek, M. R., Teitzel, G. M., Lory, S., and Greenberg, E. P. 2001. Gene expression in *Pseudomonas aeruginosa* biofilms. *Nature*, 413(6858), 860–864.
- Whitehead, N. A., Barnard, A. M. L., Slater, H., Simpson, N. J. L., and Salmond, G. P. C. 2001. Quorum-sensing in Gram-negative bacteria. *FEMS Microbiology Reviews*, 25(4), 365–404.
- WHO. 2014. Global Health Observatory (GHO) Mortality and Global Health estimate.
- Wilken, R., Veena, M. S., Wang, M. B., and Srivatsan, E. S. 2011. Curcumin: A review of anti-cancer properties and therapeutic activity in head and neck squamous cell carcinoma. *Molecular Cancer*, 10, 12.
- Willcox, M. D., Zhu, H., Conibear, T. C., Hume, E. B., Givskov, M., Kjelleberg, S., and Rice, S. A. 2008. Role of quorum sensing by *Pseudomonas aeruginosa* in microbial keratitis and cystic fibrosis. *Microbiology*, 154, 2184–2194.
- Wilson, R., Pitt, T., Taylor, G., Watson, D., MacDermot, J., Sykes, D., and Cole, P. 1987. Pyocyanin and 1-hydroxyphenazine produced by *Pseudomonas aeruginosa* inhibit the beating of human respiratory cilia *in vitro*. *Journal of Clinical Investigation*, 79(1), 221–229.
- Winzer, K., and Williams, P. 2001. Quorum sensing and the regulation of virulence gene expression in pathogenic bacteria. *International Journal of Medical Microbiology*, 291(2), 131–143.
- Wise, R., and Soulsby, E. J. L. 2002. Antibiotic resistance: An evolving problem. *Vet. Rec.*, 151, 371–372.
- Withers, H., Swift, S., and Williams, P. 2001. Quorum sensing as an integral component of gene regulatory networks in Gram-negative bacteria. *Current Opinion in Microbiology*, 4(2), 186–193.
- Wozniak, D. J., Wyckoff, T. J. O., Starkey, M., Keyser, R., Azadi, P., O'Toole, G. A., and Parsek, M. R. 2003. Alginate is not a significant component of the extracellular polysaccharide matrix of PA14 and PAO1 *Pseudomonas aeruginosa* biofilms. *Proceedings of the National Academy of Sciences*, 100(13), 7907–7912.
- Xiao, G., He, J., and Rahme, L. G. 2006. Mutation analysis of the *Pseudomonas aeruginosa* mvfR and pqsABCDE gene promoters demonstrates complex quorum-sensing circuitry. *Microbiology*, 152,

1679–1686.

- Yahr, T. L., and Parsek, M. R. 2006. *Pseudomonas aeruginosa*. (M. Dworkin, M. Falkow, M. Rosenberg, K. H. Schleifer, and E. Stackebrandt, Eds.) (3rd ed.). New York, NY: Springer.
- Yamarthi, A., and Sarada, J. 2015. Quorum quenchers-past, present and future of this novel therapeutics. *World Journal of Pharmaceutical and Life Sciences*, 1(3), 149–159.
- Yang, L., Barken, K. B., Skindersoe, M. E., Christensen, A. B., Givskov, M., and Tolker-Nielsen, T. 2007. Effects of iron on DNA release and biofilm development by *Pseudomonas aeruginosa*. *Microbiology*, 153(5), 1318–1328.
- Yang, L., Hu, Y., Liu, Y., Zhang, J., Ulstrup, J., and Molin, S. 2011. Distinct roles of extracellular polymeric substances in *Pseudomonas aeruginosa* biofilm development. *Environmental Microbiology*, 13(7), 1705–1717.
- Yang, Z., Koh, S. K., Ng, W. C., Lim, R. C. J., Tan, H. T. W., Tong, Y. W., and Wang, C. H. 2016. Potential application of gasification to recycle food waste and rehabilitate acidic soil from secondary forests on degraded land in Southeast Asia. *Journal of Environmental Management*, 172, 40–48.
- Yasni, S., Imaizumi, K., Sin, K., Sugano, M., Monaka, G., and Sidik. 1994. Identification of an active principle in essential oils and hexane soluble fraction of *Curcuma xanthorriza* Roxb. showing triglyceride lowering action in rats. *Food Chemical Toxicology*, 32, 273–278.
- Yeung, A. T. Y., Torfs, E. C. W., Jamshidi, F., Bains, M., Wiegand, I., Hancock, R. E. W., and Overhage, J. 2009. Swarming of *Pseudomonas aeruginosa* is controlled by a broad spectrum of transcriptional regulators, including MetR. *Journal of Bacteriology*, 191(18), 5592–602.
- Yoon, S. S., Hennigan, R. F., Hilliard, G. M., Ochsner, U. A., Parvatiyar, K., Kamani, M. C., Hassett, D. J. 2002. *Pseudomonas aeruginosa* anaerobic respiration in biofilms: Relationships to cystic fibrosis pathogenesis. *Developmental Cell*, 3(4), 593–603.
- Young, G. M., Smith, M. J., Minnich, S. A., and Miller, V. L. 1999. The *Yersinia enterocolitica* motility master regulatory operon, flhDC, is required for flagellin production, swimming motility, and swarming motility. *Journal of Bacteriology*, 181(9), 2823–33.
- Zhang, L.-H., and Dong, Y.-H. 2004. Quorum sensing and signal interference: Diverse implications. *Molecular Microbiology*, 53(6), 1563–1571.
- Zeng, J., Zhang, N., Huang, B., Cai, R., Wu, B., Fang, C., and Chen, C. 2016. Mechanism of azithromycin inhibition of HSL synthesis in *Pseudomonas aeruginosa*.

Zwaving, J. H., and Bos, R. 1992. Analysis of the essential oils of five curcuma species. *Flavour and Fragrance Journal*, 7(1), 19–22.

